Hindawi Journal of Healthcare Engineering Volume 2021, Article ID 2257066, 12 pages https://doi.org/10.1155/2021/2257066

Research Article

Gene Characteristics and Prognostic Values of m⁶A RNA Methylation Regulators in Nonsmall Cell Lung Cancer

Na Li, 1 Xiaojuan Chen, 2 Yanhong Liu, 1 Tieming Zhou 10, 1 and Wei Li 103

Correspondence should be addressed to Tieming Zhou; zhoutieming2008@126.com and Wei Li; xylw2015@csu.edu.cn

Received 28 June 2021; Accepted 16 July 2021; Published 30 July 2021

Academic Editor: Dr. Dilbag Singh

Copyright © 2021 Na Li et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Background. N⁶-methyladenosine (m⁶A) is the most common internal modification present in mRNAs and long noncoding RNAs (lncRNAs), associated with tumorigenesis and cancer progression. However, little is known about the roles of m⁶A and its regulatory genes in nonsmall cell lung cancer (NSCLC). Here, we systematically explored the roles and prognostic significance of m⁶A-associated regulatory genes in NSCLC. Methods. The copy number variation (CNV), mutation, mRNA expression data, and corresponding clinical pathology information of 1057 NSCLC patients were downloaded from the cancer genome atlas (TCGA) database. The gain and loss levels of CNVs were determined by utilizing segmentation analysis and GISTIC algorithm. The GSEA was conducted to explore the functions related to different levels of m⁶A regulatory genes. Logrank test was utilized to assess the prognostic significance of m⁶A-related gene's CNV. Results. The genetic alterations of ten m⁶A-associated regulators were identified in 102 independent NSCLC samples and significantly related to advanced tumor stage. Deletions or shallow deletions corresponded to lower mRNA expression while copy number gains or amplifications were related to increased mRNA expression of m⁶A regulatory genes. Survival analysis showed the patients with copy number loss of FTO with worse disease-free survival (DFS) or overall survival (OS). Besides, copy number loss of YTHDC2 was also with poor OS for NSCLC patients. Moreover, high FTO expression was significantly associated with oxidative phosphorylation, translation, and metabolism of mRNA. Conclusion. Our findings provide novel insight for better understanding of the roles of m⁶A regulators and RNA epigenetic modification in the pathogenesis of NSCLC.

1. Introduction

Lung cancer is one of the most prevalent malignant tumors and also the most lethal cancer with an approximate 5-year survival rate of 16% all over the world [1]. The latest cancer statistic data indicate that there will be 19,300,000 new tumor patients and more than 10,000,000 deaths in 2020 [1, 2]. Nonsmall cell lung cancer (NSCLC) is the most prevalent type of lung tumor which accounts for 80% of all cases. In the past decades, a series of therapeutics including chemotherapy, surgery, radiotherapy, and immunotherapy were applied to lung tumor patients; however, the prognosis of patients is still unfavorable and is especially poor in advanced NSCLC [3]. NSCLC has become a serious health problem worldwide. Hence, to

further explore the molecular pathogenesis underlying NSCLC to develop effective diagnostics and therapies is urgently needed.

The genetic and epigenetic alterations of nucleotides were involved in various regular bioprocesses such as regulation of gene expression, variable splicing, and protein translation, which play essential roles in the occurrence and progression of various diseases [4–6]. RNAs nucleotides modification is a common epigenetic alteration and more than 100 chemically modified nucleotides in different RNAs have been identified [7, 8]. Among these, methylation of N⁶-methyladenosine (m⁶A) is the most prevalent internal form of messenger RNAs (mRNAs) and long noncoding RNAs (lncRNAs) modification in eukaryotes [9]. Previous studies have shown that m⁶A modification is present in more than

¹Department of Pathology, The Second Affiliated Hospital of Hunan University of Chinese Medicine, Changsha, China

²Department of Clinical Laboratory, Maternal and Child Care Hospital, Changsha, Hunan, China

³Department of Geriatrics, Clinical Laboratory, Xiangya Hospital of Central South University, Changsha, China

7,600 mRNAs and in over 300 noncoding RNAs [10]. It is known as m⁶A modification closely related to RNA splicing, localization, stability, export, RNA-protein interactions, alternative polyadenylation, and translation [11–13]. The cellular m⁶A methylation is mediated by a group of regulatory enzymes including "writers" Wilms' tumor 1-associated protein (WTAP), methyltransferase-like 3 (METTL3) and METTL14, "erasers" fat mass and obesity-associated protein (*FTO*) and alkB homolog 5 (ALKBH5), and "readers" YTH domain containing 1 (YTHDC1/2) and YTH N⁶-methyladenosine RNA binding protein 1/2/3 (YTHDF1/2/3) [14–16]. Generally, m⁶A methylation is increased by writers, erased by *FTO* or ALKBH5, and deciphered by YTHDF1/2/3 or YTHDC1/2.

In recent years, growing evidences indicated that m⁶A dysregulation plays critical roles in tumorigenesis and cancer progression through diverse molecular mechanisms [14-18]. Meanwhile, the expression level of m⁶A-related regulatory proteins has been shown to be critically involved in tumorigenesis [19-21]. Knockout of m⁶A methyltransferase can regulate cancer occurrence by affecting the activity of p53 signaling pathway [22]. More recently, downregulation of FTO reduced lung cancer cell proliferation and invasion and promoted cell apoptosis [23, 24]. Another study also revealed that METTL14 by regulating its mRNA targets promotes leukemogenesis through mRNA m⁶A modification [25]. All these results show that m⁶A modification factors play essential roles in the occurrence of a variety of cancers. However, the connection between m⁶Arelated regulatory factors and NSCLC remained not very clear. In the present work, we systematically explored the expression pattern of m⁶A regulators in NSCLC based on the data from TCGA database. We also analyzed the association between m⁶A-related genetic alterations and clinical features including age, sex, pathological stage, disease-free survival (DFS), and overall survival (OS).

2. Materials and Methods

- 2.1. Acquisition of NSCLC Data. The CNV, mutation, mRNA expression information, and corresponding clinicopathological information of 1057 NSCLC patients were obtained from the TCGA database (GDC data portal) (https://cancergenome.nih.gov/).
- 2.2. Data Preprocessing and Copy Number Variations Identifying. The gain and loss levels of copy number variations (CNVs) were determined by utilizing segmentation analysis and GISTIC algorithm.

The NSCLC samples were grouped into two classes: without CNVs and mutation of ten m⁶A regulators and with CNVs and/or mutation. The expression of mRNA in each CNV groups was calculated using R package "DESeq2."

2.3. Gene Set Enrichment Analysis (GSEA). The GSEA was implemented to explore the functions related to different levels of m⁶A regulatory genes.

The JAVA program with MSigDB v6.1 was used to execute GSEA. All samples were sorted into low- and high-FTO level groups. Then, significant enrichment of gene sets was calculated with a false discovery rate (FDR) value less than 0.25 and a normalized P value less than 0.05.

- 2.4. Survival Analysis. All NSCLC samples were grouped by with or without deletion/gain of each m⁶A regulator gene and then a survival analysis was conducted by utilizing R packages "survival" (https://cran.r-project.org/web/packages/survival/index.html) and R package "survminer" (https://cran.r-project.org/web/packages/survminer/index. html). The prognosis value of the CNV of m⁶A regulatory gene was assessed by logrank test. Moreover, the Kaplan–Meier plotter (https://kmplot.com/analysis/) was used to analyze the prognosis value of each m⁶A regulator.
- 2.5. Statistical Analysis. All data were processed by utilizing R (4.0). The relationship between the CNV of m⁶A regulators and clinical-pathological features was investigated with chisquare test or Kruskal–Wallis rank sum test. All *P* values less than 0.05 were treated to be statistically significant.

3. Results

- 3.1. Mutations and CNV Events of m⁶A Regulators in NSCLC Samples. In total, 1057 NSCLC specimens with sequencing data were included in the present research. Among these, the genetic alterations of m⁶A moderators were identified in 102 independent samples (Figure 1). In detail, the m⁶A "reader" genes YTHDF3 (8.8%, 93/1057), YTHDF1 (7.85%, 83/1057), and YTHDC2 (5.58%, 59/1057) are the top three genes in the CNVs frequency (Table 1, Figure 2(a)). Moreover, the CNVs of NSCLC-driven genes EGFR, KRAS, and TP53 were assessed and the results were 13.43%, 10.69%, and 6.53% (Table 1, Figure 2(a)), respectively. Subsequently, we counted all CNV patterns in NSCLC samples and found that the copy number loss events are the most of all CNVs (264/ 495) (Table 1, Figure 2(b)). The shallow deletions of YTHDC2 are the most frequent copy number loss of these ten m⁶A regulators, while the gain of YTHDF3 DNA copy number is the most common change in the CNVs of ten m⁶A regulatory genes (Table 1, Figure 2), suggesting a key significance of m⁶A reader genes in RNA m⁶A methylation in NSCLC patients.
- 3.2. Association between the Alterations of m^6A Regulators and Clinical-Pathological Characteristics. To investigate the connection between genetic variations of m^6A regulators and the clinicopathological characteristics of NSCLC patients, we implemented a correlation analysis. The results showed that genetic alterations of m^6A -related regulators were obviously correlated to advanced tumor stage (P < 0.05) (Table 2). Given that EGFR, TP53, and KRAS play crucial roles in tumorigenesis and progression of lung cancer, we examined the connection between m^6A -related regulators and the variations of the above three tumor-

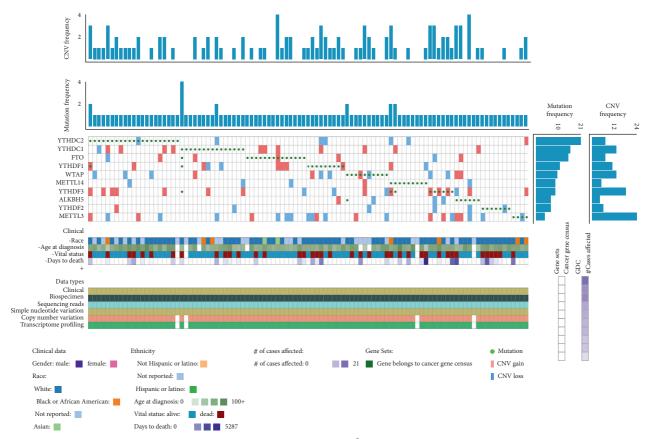


Figure 1: CNVs and mutations of top ten m⁶A regulators in NSCLC patients.

Deep deletion Gene Diploid Shallow deletion Copy number gain Amplification CNV sum Percentage ALKBH5 5.49 Eraser FTO 5.01 METTL14 2.18 Writer METTL3 2.37 WTAP 5.01 YTHDF1 7.85 YTHDF2 2.84 Reader YTHDC1 1.70 YTHDC2 5.58 YTHDF3 8.80 **EGFR** 13.43 Others **KRAS** 10.69 **TP53** 6.53

Table 1: CNV patterns in NSCLC patients (N = 1057).

related genes. We found that the variations of m^6A regulatory genes were obviously linked to *EGFR* and *TP53* alteration (P < 0.05) (Table 3). In detail, only 54 samples were missing from alterations of m^6A regulators among the 316 patients with *EGFR* alteration and 57 samples were absent in 321 patients with *TP53* alteration (Table 3). However, variations of m^6A regulators did not correlate significantly with *KRAS* mutation (P > 0.05) (Table 3).

Subsequently, we further explored whether alterations in m⁶A-related genes affect mRNA expression. The results suggested that the expression of mRNA was significantly related to different CNV types in NSCLC patients. Deletions

or shallow deletions were corresponding to lower mRNA expression while copy number gains or amplifications were linked to increased mRNA expression of the ten m⁶A regulators (Figure 3).

3.3. Prognostic Significance of CNVs in m⁶A Regulatory Genes for NSCLC Patients. The prognostic significance of CNVs in the m⁶A regulators for DFS and OS among NSCLC patients was investigated, there was no significant correlation between patients with/without CNVs of m⁶A-related regulators and OS or DFS (Figures 4(a)-4(b)). However, further

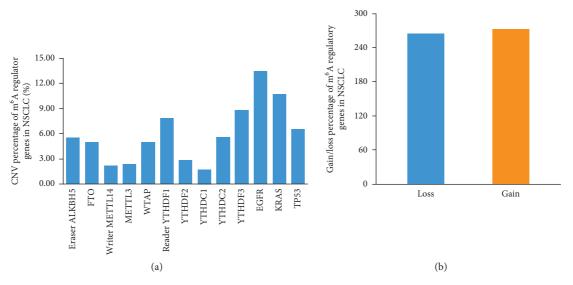


FIGURE 2: CNVs of ten m⁶A regulators in NSCLC. (a) CNV ratio of m⁶A regulators in NSCLC samples. (b) Number of gain or loss of DNA copy number of m⁶A-related regulators in NSCLC patients.

TABLE 2: Clinical features of patients with NSCLC who are with or without genetic alterations of m⁶A regulators.

Parameters		With mutation and/or CNVs	Without mutation and/or CNVs	P value
A.	≤60	90	176	0.217
Age	>60	223	524	
Gender	Female	123	283	0.787
	Male	190	417	
Primary diagnosis	Acinar cell carcinoma	6	16	0.07
	Adenocarcinoma, NOS	102	210	
	Adenocarcinoma with mixed subtypes	37	71	
	Basaloid squamous cell carcinoma	5	9	
	Bronchiolo-alveolar carcinoma, mucinous	2	3	
	Bronchiolo-alveolar adenocarcinoma, NOS	1	2	
	Bronchiolo-alveolar carcinoma, nonmucinous	6	12	
	Clear cell adenocarcinoma, NOS	1	1	
	Micropapillary carcinoma, NOS	0	3	
	Mucinous adenocarcinoma	1	12	
	Papillary adenocarcinoma, NOS	6	16	
	Papillary squamous cell carcinoma	1	3	
	Signet ring cell carcinoma	0	1	
	Solid carcinoma, NOS	2	4	
	Squamous cell carcinoma, keratinizing, NOS	3	9	
	Squamous cell carcinoma, small cell, nonkeratinizing	1	3	
	Squamous cell carcinoma, NOS	139	325	
Tumor stage	Not reported	3	8	0.001*
	Stage I-II	242	559	
	Stage III-IV	68	133	
Tissue or organ of origin	Lower lobe, lung	99	251	0.137
	Lung, NOS	14	37	
	Main bronchus	4	5	
	Middle lobe, lung	14	23	
	Overlapping lesion of lung	6	5	
	Upper lobe, lung	176	379	

NOS, not otherwise specified.

TABLE 3: Relationship between EGFR/KRAS/TP53 and m⁶A genes.

Gene		With alteration in 10 m ⁶ A genes	Without alteration in 10 m ⁶ A genes	χ^2	P value
EGFR	WT	262	629	6.598	0.010
n = 1023	Alteration	54	78		
TP53	WT	264	694	96.505	< 0.001
n = 1024	Alteration	57	9		
KRAS	WT	283	647	2.039	0.153
n = 1036	Alteration	40	66		

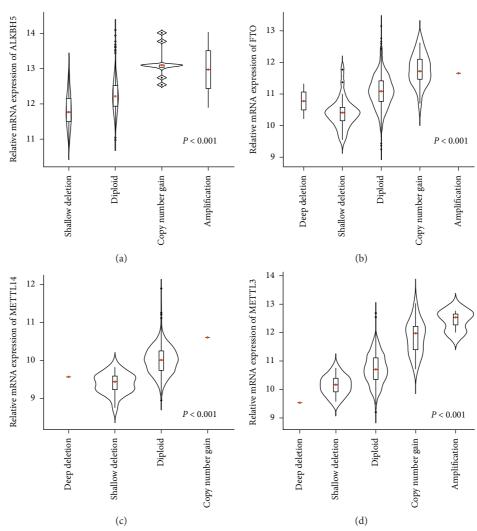


FIGURE 3: Continued.

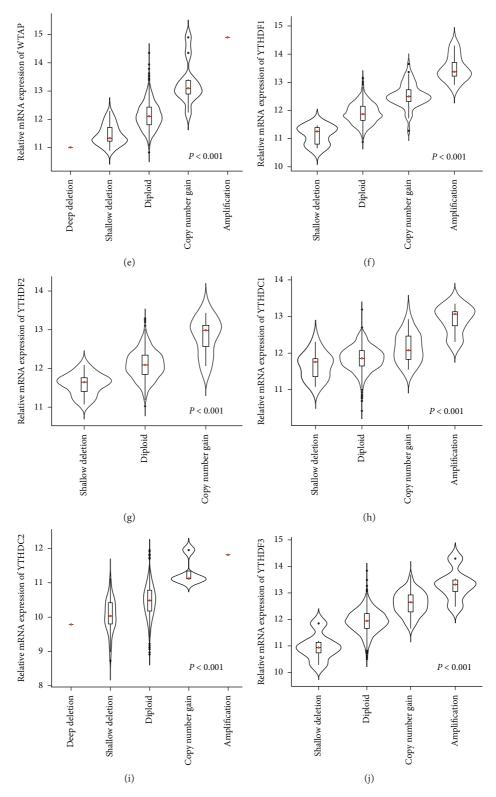


FIGURE 3: The relation between CNV types and m⁶A regulator expression.

analysis showed copy number loss of FTO with worse DFS and OS (Figures 4(c)-4(d)). Besides, copy number loss of YTHDC2 is also with poor OS for NSCLC patients

(Figure 4(e)). Furthermore, we further found low FTO expression was closely related to poor OS of lung cancer patients by using Kaplan–Meier plotter (Figure 4(f)).

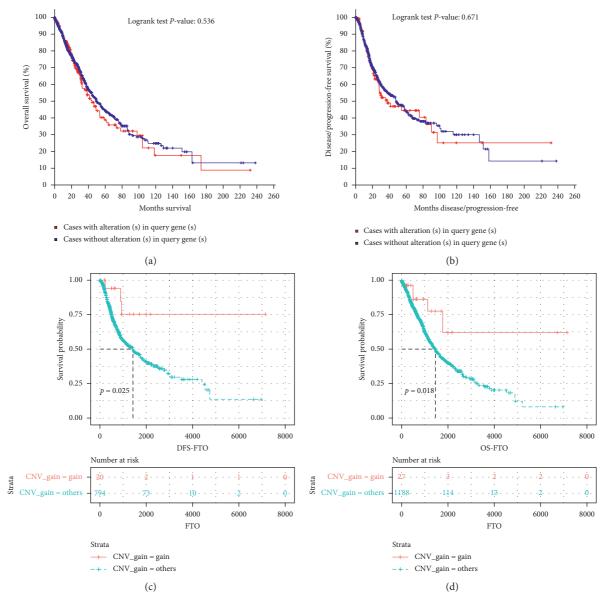


FIGURE 4: Continued.

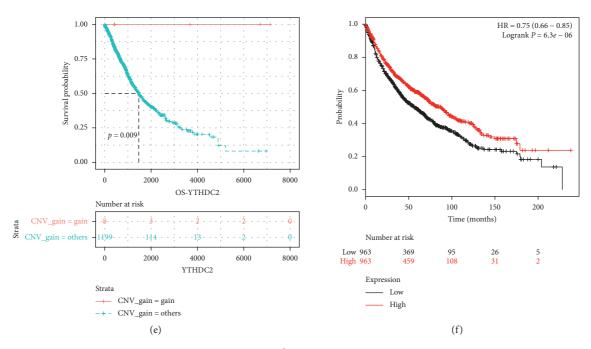


FIGURE 4: Survival analysis of NSCLC patients with CNVs of m⁶A-related regulators. ((a)-(b)) OS and DFS for NSCLC patients who have any alteration of m⁶A-related regulators, ((c)-(d)) DFS and OS for patients with NSCLC who have different CNV types of *FTO*, (e) OS for patients with NSCLC who have different CNV types of *YTHDC2*, and (f) OS for patients with *FTO* mRNA expression levels by Kaplan–Meier plotter.

3.4. Enrichment Analysis of FTO Gains of Function. Given that there is close correlation between CNVs and m⁶A regulator FTO and NSCLC prognosis, a GSEA was executed to investigate the gene enrichment in patients with different FTO mRNA expression. The results showed that high FTO expression was significantly associated with multiple key biological processes, including oxidative phosphorylation, ribosome, translation, 3'-UTR-mediated translational regulation, metabolism of mRNA, and influenza life cycle (Figure 5), which provides new clues for understanding the pathogenesis of NSCLC.

4. Discussion

RNA modification is an emerging branch of epigenetics that is increasingly attracting the interest of related researchers. Currently, RNA modification is thought to be widespread in almost all forms of RNA, including mRNAs, lncRNAs, rRNAs, tRNAs, micro-RNAs, and small nucleolar RNAs [26-28]. Previous studies have shown that RNA modifications include multiple forms, such as pseudouridine, N7methyladenosine, 2'-O-methylation, N1-methyladenosine, N6,2-O-dimethyladenosine (m⁶A), and 5-methylcytosine [29, 30]. Among them, m⁶A methylation is the most common mRNA modification form and it participates in the regulation of numerous biological processes in eukaryotes [29–31]. The cellular m⁶A status is dynamically regulated by methyltransferases, binding proteins, and demethylases. The alterations of these regulatory factors lead to dysregulation of m⁶A methylation and hence play an essential role in the progression of various diseases [31-33].

Bioinformatics analysis plays a fundamental role in disease diagnosis and pathogenesis research [34-36]. In this study, we used bioinformatics approaches to systematically identify the roles and prognostic values of m⁶A-related regulatory factors in NSCLC. A total of 1057 NSCLC samples with CNV information and clinical data from TCGA were included. Of these, 102 independent samples were identified as having the genetic variations of m⁶Arelated regulators. Compared with clear cell renal cell carcinoma (ccRCC) and acute myeloid leukemia, the frequency of alterations in ten m⁶A-related genes in lung cancer is less [37, 38]. Among all CNV patterns in NSCLC, the copy number loss was the most important part of all CNV events, which was similar as the CNV patterns in ccRCC [37] and acute myeloid leukemia [38]. For all m⁶A regulators, deletions are the most important part of CNVs in "eraser" and "writer" genes, but the number of gain of CNV events in "eraser" genes is higher than those of "writer" genes, which eventually decreased the m⁶A level in NSCLC cells. Previous studies have shown that m⁶A levels were downregulated in various tumors, such as glioblastoma and breast cancer [39–41]. This may also be explained by the opposite effect on m⁶A status for "eraser" and "writer" genes.

Furthermore, we found that copy number gains or amplifications were positively correlated with mRNA levels of the 10 m⁶A-related regulators while deletions or shallow deletions were negatively related to mRNA levels, implying that alterations in CNVs affect m⁶A-related genes expression in NSCLC. Further analysis revealed that alterations of m⁶A-related regulators were positively linked to cancer stage of NSCLC. These results suggested that CNVs of m⁶A

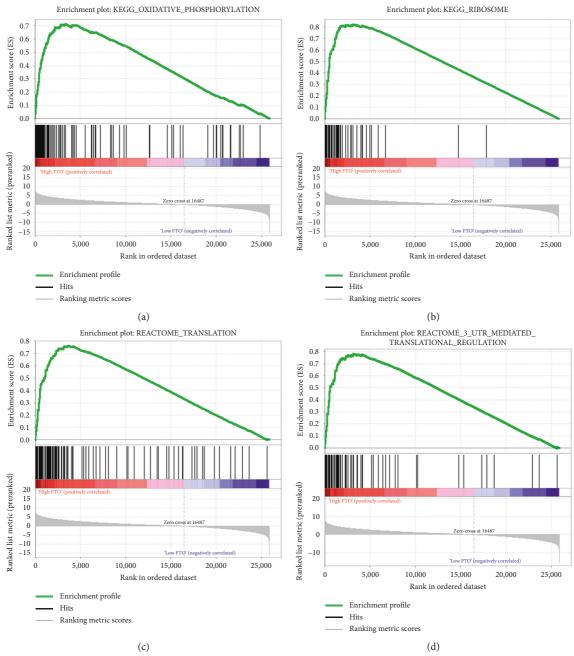


FIGURE 5: Continued.

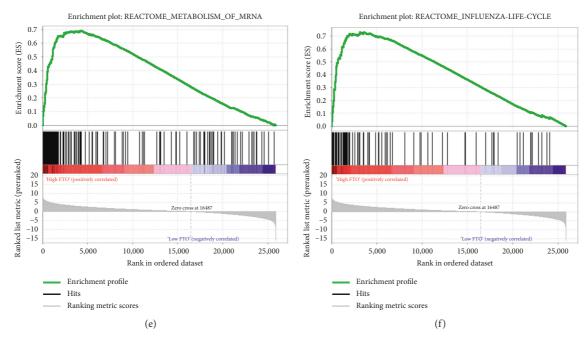


FIGURE 5: GSEA outcomes of different expression level of FTO. (a) Oxidative phosphorylation, (b) ribosome, (c) translation, (d) 3'-UTR-mediated translational regulation, (e) metabolism of mRNA, and (f) influenza life cycle.

regulatory genes are involved in the regulation of tumor progression by affecting methylation modification of RNAs. A recent study found that overexpression of m⁶A methyltransferase METTL3 facilitates tumor development through AFF4/NF- κ B/MYC signal pathway in bladder cancer [42]. Another study also indicated changes in the expression of m⁶A regulatory genes by regulating corresponding genes to promote breast cancer progression [41]. We also found that the changes of m⁶A modulators were significantly associated with EGFR and TP53 alteration. EGFR is an important oncogene, and its mediated pathways play crucial roles in tumor occurrence and development [43-45], while TP53 is a classic tumor suppressor gene [46]. Based on these findings, we speculate that dysregulated m⁶A status induced EGFR activation and TP53 inactivation to facilitate the tumorigenesis and progress of NSCLC.

Moreover, we explored the prognostic value of m⁶A regulator alterations in NSCLC. For all ten m⁶A-related genes, only copy number loss of eraser gene FTO and reader gene YTHDC2 was significantly associated with poor survival status for NSCLC patients. Besides, the results of Kaplan-Meier plotter analysis also indicate that low FTO expression is related to worse OS of lung cancer patients. Taking together, the present results showed that the FTO levels are inversely related to the survival time of patients with NSCLC. However, studies have revealed that FTO as an m⁶A demethylase participates in promoting the growth of lung cancer cells in vitro [23, 24]. These findings revealed that m⁶A regulatory genes are a "double-edged sword" in tumorigenesis, which could lead to not only tumor suppression but also tumor progression. Thus, restoring the balanced state of RNA methylation in tumor cells is a new anticancer strategy.

The present study displayed that m⁶A-related regulatory genes were also associated with multiple signaling

pathways and biological processes of NSCLC occurrence and development. The results of the GSEA suggested that the expression of FTO was significantly related to oxidative phosphorylation, ribosome, translation, 3'-UTR-mediated translational regulation, and metabolism of mRNA. Similar to our results, a previous study reported that FTO expression was positively related to dextrose oxidation rates and levels of genes related to oxidative phosphorylation in skeletal muscle [47]. It has been shown that oxidative phosphorylation plays significant roles in lung cancer proliferation, invasion, metastasis, and drug resistance [48-50]. Therefore, it is likely that genetic alterations of FTO regulate the progression of NSCLC by affecting cellular oxidative phosphorylation levels. The specific molecular mechanism deserves to be explored through further work. In addition, several biological processes regulated by the m⁶A regulators have been identified, including RNA metabolism, translational regulation, and protein translation [23, 51], which are consistent with our GSEA results.

In conclusion, our work systematically displayed the genetic alterations, expression patterns, potential roles, and prognostic significance of m⁶A-related regulators in NSCLC and found that the alterations of m⁶A regulators are highly related to the malignant clinicopathological characteristics including survival. These results help us to find out the functions of m⁶A RNA methylation in the pathogenesis of NSCLC. However, these findings need to be validated with further clinical and molecular biology experiments.

Data Availability

Publicly available datasets were analyzed in this study. These data can be found at https://www.cancer.gov/.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Acknowledgments

This project was supported by the Natural Science Foundation of Hunan Province (2019JJ50425 and 2020JJ5909) and General Project of Hunan Provincial Education Department (18C1129).

References

- [1] H. Sung, J. Ferlay, and R. L. Siegel, "Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries," *CA: A Cancer Journal for Clinicians*, vol. 71, pp. 209–249, 2021.
- [2] R. L. Siegel, K. D. Miller, H. E. Fuchs, and A. Jemal, "Cancer statistics," *CA: A Cancer Journal for Clinicians*, vol. 71, pp. 7–33, 2021.
- [3] F. Bray, J. Ferlay, I. Soerjomataram, R. L. Siegel, L. A. Torre, and A. Jemal, "Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries," CA: A Cancer Journal for Clinicians, vol. 68, pp. 394–424, 2018.
- [4] W. M. Grady, "Epigenetic alterations in the gastrointestinal tract: current and emerging use for biomarkers of cancer," *Advances in Cancer Research*, vol. 151, pp. 425–468, 2021.
- [5] M. A. Koldobskiy, G. Jenkinson, J. Abante, V. A. R. DiBlasi, W. Zhou, and E. Pujadas, "Converging genetic and epigenetic drivers of paediatric acute lymphoblastic leukaemia identified by an information-theoretic analysis," *Nature Biomedical Engineering*, vol. 5, pp. 360–376, 2021.
- [6] A. S. Rathore, H. Birla, S. S. Singh, W. Zahra, H. Dilnashin, and R. Singh, "Epigenetic modulation in Parkinson's disease and potential treatment therapies," *Neurochemical Research*, vol. 46, pp. 1618–1626, 2021.
- [7] W. A. Cantara, P. F. Crain, J. Rozenski et al., "The RNA modification database, RNAMDB: 2011 update," *Nucleic Acids Research*, vol. 39, pp. D195–D201, 2011.
- [8] I. Manfredonia and D. Incarnato, "RNA post-transcriptional modification mapping data analysis using RNA framework," *Methods in Molecular Biology*, vol. 2298, pp. 3–13, 2021.
- [9] J. Luo, T. Xu, and K. Sun, "N6-Methyladenosine RNA modification in inflammation: roles, mechanisms, and applications," Frontiers in Cell and Developmental Biology, vol. 9, Article ID 670711, 2021.
- [10] K. D. Meyer, Y. Saletore, P. Zumbo, O. Elemento, C. E. Mason, and S. R. Jaffrey, "Comprehensive analysis of mRNA methylation reveals enrichment in 3' UTRs and near stop codons," *Cell*, vol. 149, pp. 1635–1646, 2012.
- [11] X. Han, J. Guo, and Z. Fan, "Interactions between m6A modification and miRNAs in malignant tumors," *Cell Death & Disease*, vol. 12, Article ID 598, 2021.
- [12] R. Vasic, Y. Gao, C. Liu, and S. Halene, "The role of RNA epigenetic modification in normal and malignant hematopoiesis," *Current Stem Cell Reports*, vol. 6, pp. 144–155, 2020.
- [13] R. Fang, L. Ye, and H. Shi, "Understanding the roles of N(6)-methyladenosine writers, readers and erasers in breast cancer," *Neoplasia*, vol. 23, pp. 551–560, 2021.
- [14] Q. Lan, P. Y. Liu, J. Haase, J. L. Bell, S. Huttelmaier, and T. Liu, "The critical role of RNA m(6)A methylation in cancer," *Cancer Research*, vol. 79, pp. 1285–1292, 2019.

- [15] F. V. Hagen, L. Gundert, A. Strick, N. Klumper, D. Schmidt, and G. Kristiansen, "N(6) -Methyladenosine (m(6) A) readers are dysregulated in renal cell carcinoma," *Molecular Carcinogenesis*, vol. 60, pp. 354–362, 2021.
- [16] Z. Zou, S. Zhou, G. Liang, Z. Tang, K. Li, and S. Tan, "The pancancer analysis of the two types of uterine cancer uncovered clinical and prognostic associations with m6A RNA methylation regulators," *Molecular Omics*, vol. 17, pp. 438–453, 2021
- [17] B. Chen, Y. Li, R. Song, C. Xue, and F. Xu, "Functions of RNA N6-methyladenosine modification in cancer progression," *Molecular Biology Reports*, vol. 46, pp. 1383–1391, 2019.
- [18] X. Lin, G. Chai, Y. Wu, J. Li, F. Chen, and J. Liu, "RNA m(6)A methylation regulates the epithelial mesenchymal transition of cancer cells and translation of Snail," *Nature Communications*, vol. 10, Article ID 2065, 2019.
- [19] J. Yu, W. Mao, S. Sun, Q. Hu, C. Wang, and Z. Xu, "Identification of an m6A-related lncRNA signature for predicting the prognosis in patients with kidney renal clear cell carcinoma," *Frontiers in Oncology*, vol. 11, Article ID 663263, 2021.
- [20] C. Gu, X. Shi, W. Qiu, Z. Huang, Y. Yu, and F. Shen, "Comprehensive analysis of the prognostic role and mutational characteristics of m6A-related genes in lung squamous cell carcinoma," Frontiers in Cell and Developmental Biology, vol. 9, Article ID 661792, 2021.
- [21] S. Ma, J. Yan, T. Barr, J. Zhang, Z. Chen, and L. S. Wang, "The RNA m6A reader YTHDF2 controls NK cell antitumor and antiviral immunity," *Journal of Experimental Medicine*, vol. 218, Article ID e20210279, 2021.
- [22] D. Dominissini, S. M. Moshkovitz, S. Schwartz, M. Salmon-Divon, L. Ungar, and S. Osenberg, "Topology of the human and mouse m6A RNA methylomes revealed by m6A-seq," *Nature*, vol. 485, pp. 201–206, 2012.
- [23] J. Li, Y. Han, H. Zhang, Z. Qian, W. Jia, and Y. Gao, "The m6A demethylase FTO promotes the growth of lung cancer cells by regulating the m6A level of USP7 mRNA," *Biochemical and Biophysical Research Communications*, vol. 512, pp. 479–485, 2019
- [24] J. Liu, D. Ren, Z. Du, H. Wang, H. Zhang, and Y. Jin, "m(6)A demethylase FTO facilitates tumor progression in lung squamous cell carcinoma by regulating MZF1 expression," *Biochemical and Biophysical Research Communications*, vol. 502, pp. 456–464, 2018.
- [25] H. Weng, H. Huang, H. Wu, X. Qin, B. S. Zhao, and L. Dong, "METTL14 inhibits hematopoietic stem/progenitor differentiation and promotes leukemogenesis via mRNA m(6)A modification," *Cell Stem Cell*, vol. 22, pp. 191–205, 2018.
- [26] I. A. Roundtree, M. E. Evans, T. Pan, and C. He, "Dynamic RNA modifications in gene expression regulation," *Cell*, vol. 169, pp. 1187–1200, 2017.
- [27] Y. Yang, P. J. Hsu, Y. S. Chen, and Y. G. Yang, "Dynamic transcriptomic m(6)A decoration: writers, erasers, readers and functions in RNA metabolism," *Cell Research*, vol. 28, pp. 616–624, 2018.
- [28] R. Janostiak and N. Wajapeyee, "RNA modification regulatory genes in DNA damage," *Methods in Molecular Biology*, vol. 1870, pp. 249–262, 2019.
- [29] J. Liu, B. T. Harada, and C. He, "Regulation of gene expression by N(6)-methyladenosine in cancer," *Trends in Cell Biology*, vol. 29, pp. 487–499, 2019.
- [30] Y. Wang, X. Su, M. Zhao, M. Xu, Y. Chen, and Z. Li, "Importance of N(6)-methyladenosine RNA modification in lung cancer (Review)," *Molecular and Clinical Oncology*, vol. 14, Article ID 128, 2021.

- [31] Y. Gu, X. Wu, J. Zhang, Y. Fang, Y. Pan, and Y. Shu, "The evolving landscape of N(6)-methyladenosine modification in the tumor microenvironment," *Molecular Therapy*, vol. 29, pp. 1703–1715, 2021.
- [32] X. Chen, C. Yu, M. Guo, X. Zheng, S. Ali, and H. Huang, "Down-regulation of m6A mRNA methylation is involved in dopaminergic neuronal death," *ACS Chemical Neuroscience*, vol. 10, pp. 2355–2363, 2019.
- [33] K. Du, L. Zhang, T. Lee, and T. Sun, "m(6)A RNA methylation controls neural development and is involved in human diseases," *Molecular Neurobiology*, vol. 56, pp. 1596–1606, 2019.
- [34] S. Gao and Z. Wang, "Comprehensive analysis of regulatory network for LINC00472 in clear cell renal cell carcinoma," *Journal of Healthcare Engineering*, vol. 2021, Article ID 3533608, 20 pages, 2021.
- [35] E. M. Senan, M. H. A. Adhaileh, F. W. Alsaade, T. H. H. Aldhyani, A. A. Alqarni, and N. Alsharif, "Diagnosis of chronic kidney disease using effective classification algorithms and recursive feature elimination techniques," *Journal* of *Healthcare Engineering*, vol. 2021, Article ID 1004767, 10 pages, 2021.
- [36] S. Lang, Y. Xu, L. Li, B. Wang, Y. Yang, and Y. Xue, "Joint detection of tap and CEA based on deep learning medical image segmentation: risk prediction of thyroid cancer," *Journal of Healthcare Engineering*, vol. 2021, Article ID 5920035, 9 pages, 2021.
- [37] J. Zhou, J. Wang, B. Hong, K. Ma, H. Xie, and L. Li, "Gene signatures and prognostic values of m6A regulators in clear cell renal cell carcinoma-a retrospective study using TCGA database," *Aging*, vol. 11, pp. 1633–1647, 2019.
- [38] C. T. Kwok, A. D. Marshall, J. E. Rasko, and J. J. Wong, "Genetic alterations of m(6)A regulators predict poorer survival in acute myeloid leukemia," *Journal of Hematology & Oncology*, vol. 10, Article ID 39, 2017.
- [39] Q. Cui, H. Shi, P. Ye, L. Li, Q. Qu, and G. Sun, "m(6)A RNA methylation regulates the self-renewal and tumorigenesis of glioblastoma stem cells," *Cell Reports*, vol. 18, pp. 2622–2634, 2017.
- [40] N. J. Fry, B. A. Law, O. R. Ilkayeva, K. R. Carraway, C. L. Holley, and K. D. Mansfield, "N(6)-methyladenosine contributes to cellular phenotype in a genetically-defined model of breast cancer progression," *Oncotarget*, vol. 9, Article ID 31231, 2018.
- [41] L. Wu, D. Wu, J. Ning, W. Liu, and D. Zhang, "Changes of N6-methyladenosine modulators promote breast cancer progression," *BMC Cancer*, vol. 19, Article ID 326, 2019.
- [42] M. Cheng, L. Sheng, Q. Gao, Q. Xiong, H. Zhang, and M. Wu, "The m(6)A methyltransferase METTL3 promotes bladder cancer progression via AFF4/NF-kappaB/MYC signaling network," Oncogene, vol. 38, pp. 3667–3680, 2019.
- [43] H. Wang, F. Yao, S. Luo, K. Ma, M. Liu, and L. Bai, "A mutual activation loop between the Ca(2+)-activated chloride channel TMEM16A and EGFR/STAT3 signaling promotes breast cancer tumorigenesis," *Cancer Letters*, vol. 455, pp. 48–59, 2019.
- [44] Z. Ding, J. Zhu, Y. Zeng, W. Du, Y. Zhang, and H. Tang, "The regulation of Neuropilin 1 expression by miR-338-3p promotes non-small cell lung cancer via changes in EGFR signaling," *Molecular Carcinogenesis*, vol. 58, pp. 1019–1032, 2019.
- [45] L. T. H. Phi, I. N. Sari, Y. T. Wijaya, K. S. Kim, K. Park, and A. E. Cho, "Ginsenoside rd inhibits the metastasis of colorectal cancer via epidermal growth factor receptor signaling Axis," *IUBMB Life*, vol. 71, pp. 601–610, 2019.

- [46] H. Ko and M. M. Kim, "TP53 tumor-suppressor gene plays a key role in IGF1 signaling pathway related to the aging of human melanocytes," *Anticancer Research*, vol. 39, pp. 2447–2451, 2019.
- [47] L. G. Grunnet, E. Nilsson, C. Ling, T. Hansen, O. Pedersen, and L. Groop, "Regulation and function of FTO mRNA expression in human skeletal muscle and subcutaneous adipose tissue," *Diabetes*, vol. 58, pp. 2402–2408, 2009.
- [48] R. Kikuchi, Y. Iwai, T. Tsuji, Y. Watanabe, N. Koyama, and K. Yamaguchi, "Hypercapnic tumor microenvironment confers chemoresistance to lung cancer cells by reprogramming mitochondrial metabolism in vitro," Free Radical Biology and Medicine, vol. 134, pp. 200–214, 2019.
- [49] S. E. Dyrstad, M. L. Lotsberg, T. Z. Tan, I. K. N. Pettersen, S. Hjellbrekke, and D. Tusubira, "Blocking aerobic glycolysis by targeting pyruvate dehydrogenase kinase in combination with EGFR TKI and ionizing radiation increases therapeutic effect in non-small cell lung cancer cells," *Cancers*, vol. 13, Article ID 2065, 2021.
- [50] C. C. Cheng, J. Wooten, Z. A. Gibbs, K. McGlynn, P. Mishra, and A. W. Whitehurst, "Sperm-specific COX6B2 enhances oxidative phosphorylation, proliferation, and survival in human lung adenocarcinoma," *Elife*, vol. 9, Article ID e58108, 2020.
- [51] Y. Wang, Z. Cheng, J. Xu, M. Lai, L. Liu, and M. Zuo, "Fat mass and obesity-associated protein (FTO) mediates signal transducer and activator of transcription 3 (STAT3)-drived resistance of breast cancer to doxorubicin," *Bioengineered*, vol. 12, pp. 1874–1889, 2021.