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Biomolecular QM/MM Simulations: What are some of the “Burning Issues”?

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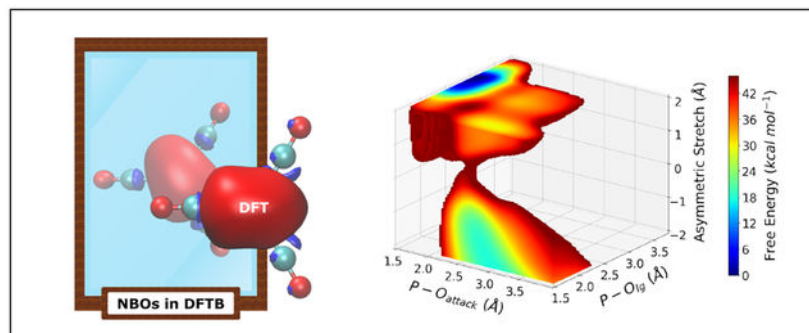
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Abstract

QM/MM simulations have become an indispensable tool in many chemical and biochemical investigations. Considering the tremendous degree of success, including the recognition by a 2013 Nobel Prize in Chemistry, are there still “burning challenges” in QM/MM methods, especially for biomolecular systems? In this short Perspective, we discuss several issues that we believe greatly impact the robustness and quantitative applicability of QM/MM simulations to many, if not all, biomolecules. We highlight these issues with observations and relevant advances from recent studies in our group and others in the field. Despite such limited scope, we hope the discussions are of general interest and will stimulate additional developments that help push the field forward in meaningful directions.

Graphical Abstract



1 Introduction

Following many years of development and calibration, hybrid quantum mechanical/molecular mechanical (QM/MM) methods^{1–4} have become an essential tool^{5–8} in chemical

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Supporting Information Available

Additional data for the second virial coefficients for small molecules and 2-/3-body energies for water at different levels of NDDO and tight-binding methods, and calculation details for the 3-dimensional free energy surface of methyl phosphate hydrolysis are included. This material is available free of charge via the Internet at <http://pubs.acs.org/>.

and biochemical research. Applications of QM/MM simulations to enzymes include not only dissection of reaction mechanisms, elucidation of various factors that contribute to catalysis, but also design of enzyme inhibitors, analysis and improvement of designed enzymes as well as understanding evolutionary relations between enzymes.^{9–16} Indeed, a search using the keyword “QM/MM” or “ONIOM”¹⁷ in the Web of Science points to >800 articles published in 2019–2020 alone. Numerous molecular simulation packages such as CHARMM, AMBER, NAMD, OpenMM and Gromacs all have QM/MM functionalities, some with build-in QM modules, while others providing interfaces with popular QM packages such as Gaussian, Q-Chem, Turbomole, Psi4 and ORCA; similarly, many quantum chemistry packages such as Gaussian, ORCA, C2PK and DFTB+ offer coupling with external MM models; there are also platforms that focus entirely on QM/MM calculations, such as the celebrated Chemshell environment¹⁸ pioneered by Sherwood and the late Walter Thiel,¹⁹ who has made numerous important contributions to the field in terms of both method developments and systematic analyses of factors that impact the accuracy of QM/MM simulations.

In light of such tremendous success, including the recognition by the 2013 Nobel Prize in Chemistry, one is tempted to ask: are there still “burning issues” for QM/MM methods, especially for biomolecular systems? If so, are the corresponding challenges fundamental in nature, or should they be better thought as establishing the best practice and enabling most efficient execution for realistic applications? The answers to such questions are clearly highly subjective, and in this short Perspective article, we discuss our views based on the recent research interest and developments in the group. Despite such limited scope, we hope the discussions are of general interest and will stimulate additional developments that help push the field forward in meaningful directions.

Before turning to these discussions, it is worth noting that application of QM/MM methods to solid/liquid interfaces,^{20,21} for example in the important context of electrochemistry²² and heterogeneous catalysis,²³ has been much more limited compared to biomolecular studies. This is perhaps due mainly to the less obvious scheme to divide the solid into QM and MM regions without considerable perturbation to the electronic structure of the solid, especially for metallic systems (e.g., the electrode). Therefore, efforts have focused primarily on pure QM based embedding approaches, such as those pioneered by Carter and co-workers based on orbital-free DFT,²⁴ which have been successfully applied to many materials problems. Nevertheless, these pure QM based embedding approaches remain computationally expensive for solid/liquid interfaces, for which an adequate sampling of the interfacial solvent and ions (i.e., the electric double layer²⁵) is expected to be essential. Therefore, further development, implementation and calibration of robust QM/MM methods for solid/liquid interfaces, especially under the condition of constant potential,^{26,27} ought to be considered an important area of research. Similar to the studies of reactions in solution, QM/MM simulations with explicit solvent²⁸ and advanced implicit solvent models^{29,30} are expected to be complementary to each other for the mechanistic analysis of electrochemical reactions.

2 Several “Burning Issues”

In the following, we discuss several “burning issues” that we believe greatly impact the robustness and quantitative applicability of QM/MM simulations to many, if not all, biomolecules. We group these issues into several general topics related to QM/MM simulations, and we highlight them with observations and relevant advances from recent studies in our group and others in the field.

2.1 Potential Function: QM and MM Selections

The choices for QM/MM partitioning and the relevant QM and MM methods are largely system-dependent. Nevertheless, given a considerable number of recent studies and sometimes passionate debates^{31–40} regarding these choices, it is worth discussing several issues explicitly.

2.1.1 The value of semi-empirical and machine learning potentials—Since adequate sampling is important to biomolecular applications, the need of an efficient QM potential is compelling, especially when the proper QM region contains at least hundreds of atoms. Along this line, approximate QM approaches such as semi-empirical QM methods^{41,42} and the empirical valence bond (EVB) method⁴³ remain particularly attractive in many applications. They need to be carefully calibrated to generate meaningful results; once calibrated (e.g., using spline fits to high-level reference reactions⁴⁴), they are uniquely powerful for the analysis of systems and/or experimental observables that demand extensive sampling. For example, as discussed further below, DFTB3/MM simulations enable the sampling of multi-dimensional free energy surfaces, which are required to analyze complex reaction pathways and to characterize the coupling between different processes (e.g., mechanochemical coupling). On the other hand, EVB based simulations have been instrumental to the computation of experimental observables that require extensive sampling to achieve the necessary numerical precision, such as activation entropy⁴⁵ and linear free energy relations,⁴⁶ which are difficult to compute otherwise. These examples include >150 ns of DFTB3/MM simulations⁴⁷ and thousands of independent EVB free energy profile simulations,⁴⁸ which are indeed very difficult, if not currently impossible, to reach with *ab initio* QM/MM simulations, despite impressive progress in computational algorithm as well as hardware.^{49,50}

Motivated by these considerations, a “burning issue” is to improve the accuracy and applicability of semi-empirical QM methods; work along this line has been reviewed fairly recently⁵¹ and will not be repeated here. A notable advance since then is the development of the extended tight binding model of Grimme et al. (xTB),⁵² which has been parameterized for the entire periodic table. While the model was developed mainly for capturing structures and vibrational frequencies, it gives encouraging results also for energetics, especially for non-covalent interactions.⁵³ In Fig. 1a, we compare the performance of several popular semi-empirical methods for computing the second virial coefficients of small molecules of different levels of polarity; in Fig. 1b, we compare the two-body and three-body interactions of water molecules by DFTB3 and xTB using the dataset of Paesani and co-workers.⁵⁴ Evidently, despite the minimal basis nature of these methods, the performance

is quite respectable even for three-body interactions of water, and these tight binding methods outperforms several NDDO methods, even those including empirical corrections for hydrogen-bonding interactions (see Supporting Information). On the other hand, there is clearly room for improvements, which would also require analysis in the condensed phase environment,⁵⁵ such as the computation of solvation free energy and binding affinity.

A major remaining challenge for the semi-empirical QM methods (and *ab initio* QM methods) is the treatment of transition metal ions, which are common in enzymes. The highly localized nature of the *d* and *f* electrons requires a reliable treatment of electron correlation, both dynamic and static. For structural properties, both DFTB3 and xTB appear to be able to provide adequate equilibrium parameters,^{51,53} although the energy landscape has not been thoroughly analyzed. Analysis of the electronic structure using Natural Bonding Orbital analysis^{57,58} found that DFTB3 provides physically sound descriptions for different bonding scenarios, including those exhibiting pseudo Jahn-Teller effects (see Fig.2a as an example). The degree of ligand to metal charge transfer and ionic character of certain bonds are overestimated, likely reflecting the minimal basis nature of DFTB3, and certain orbital interactions, such as geminal interactions, were observed to be grossly overestimated by DFTB3 for several transition metal compounds. Energetics are generally less satisfying, especially when considering different spin states, since interactions among *d/f* electrons are treated in an averaged fashion in the current tight-binding models. One possible improvement is to treat the *d/f* electrons separately, along the line of the DFT+U model in materials science⁵⁹ or the ligand-field model in the inorganic chemistry literature.^{60,61} In a recent explorative study,⁶² together with collaborators we have implemented a preliminary version of the DFTB3+U model,

$$E^{DFTB3+U} = E^{DFTB3} + \sum_{\alpha, \beta, \alpha', \beta' \in d/f} \left[P_{\alpha\beta}^U P_{\alpha'\beta'}^U \langle \alpha\alpha' || \beta\beta' \rangle - P_{\alpha\beta}^I P_{\alpha'\beta'}^I \langle \alpha\beta' | \beta'\alpha \rangle \right], \quad (1)$$

in which $P^U(P^I)$ are the spin unpolarized (polarized) density matrix elements involving the *d/f* electrons and the usual two-electron integrals are parameterized in terms of the Slater integrals or the Racah parameters.⁶⁰ It was found⁶² that the additional *U* contribution indeed improved the splitting between the low-spin and high-spin states in a series of Ni(II) and Ni(III) compounds as well as the populations of the 3*d* orbitals.⁶³ Further self-consistent tuning of the electronic parameters in the model will be informative regarding the degree of transferability and expected accuracy for the +U model, and whether a multi-determinant formulation of the DFTB3 model (e.g., in the framework of ensemble⁶⁴ or multi-state⁶⁵ DFT) is warranted.

Another promising direction for improving approximate (tight-binding and DFT) methods is to employ machine learning (ML) techniques,⁶⁶ which have seen explosive progress in recent years for developing potential energy functions,⁶⁷ predicting molecular/materials properties⁶⁸ as well as enhancing conformational sampling.⁶⁹ Specific for metal ions, for example, ML approaches have been used to analyze spin state properties⁷⁰ as well as diagnostics for static electron correlations.⁷¹ ML models have also been used to improve the accuracy of mean-field models such as tight-binding and Hartree-Fock, with

features ranging from simply atomic coordinates to Fock matrix elements.^{72,73} For realistic condensed phase QM/MM applications, the judicious combination of a physical reference QM level and ML is required to ensure the optimal balance of accuracy, robustness and transferability. Further developments along this line, so as to enable stable, energy-conserving MD simulations,^{74,75} will be exciting and transformative in terms of the type of problems that QM/MM simulations are able to tackle.

2.1.2 Going beyond popular additive MM models—Most QM/MM applications to biomolecules use popular fixed-charge MM force fields. The general success of these applications suggests that this combination is adequate for many problems, an observation also made in several comparisons of QM/MM results using both fixed-charge and polarizable force fields.⁷⁶ Nevertheless, the need to include explicit electronic polarization for certain problems has also been well documented, such as for the prediction of absorption spectra,^{77,78} reduction potential⁷⁹ and pK_a values;⁸⁰ i.e., for situations where there is a very large change in the electrostatic properties (e.g., either the net charge or dipole moment) of the QM region. While more *ad hoc* models such as charge scaling⁸⁰ or atomic polarizability models⁸¹ were used in earlier studies, more systematic efforts that integrate QM with well calibrated polarizable force fields such as AMOEBA, Effective Fragment Potential and MB-pol have been reported.^{79,82–84}

A systematic comparison between QM/MM simulations with different MM models will be informative for better identifying problems for which an explicit treatment of electronic polarization makes a qualitative difference; for example, it was suggested that to properly capture the electric field in enzyme active site for the purpose of guiding design, including electronic polarization is essential.⁸⁵ Along this line, we highlight that a meaningful comparison will require extensive sampling. In part, this is because electrostatic interactions between groups in a biomolecule are screened quite effectively;^{86,87} this is especially the case for many enzyme active sites, which are buried but not far from the protein/solvent interface. To properly capture the screening effect, it is important to consider reorientation of the protein and solvent dipoles during a chemical reaction (this contrasts to the situation of ultrafast spectroscopy,^{78,83} for which electronic dielectric response of the environment tends to be particularly important due to the limited degree of dipolar reorientation during the short time scale); without adequate sampling of such dipolar reorientations, the effect of distant groups can be overestimated. A somewhat extreme illustration is that contributions from distant charged residues to the pK_a of a titratable group are significantly overestimated (>10 kcal/mol) when analyzed in a perturbative fashion,^{88,89} i.e., by computing the change of QM/MM interaction energies without re-sampling the MD trajectory after the MM partial charges on a charged residue are turned off. For a similar reason, without adequate sampling, the effect of altering the description (MM vs. QM) of distal residues can be exaggerated, thus the issue of sampling is also highly pertinent to the discussion of QM region size in QM/MM simulations.^{31–39}

The sampling challenge is particularly significant for the prediction of properties that involve a net change of total charge, such as reduction potential and pK_a values, since the corresponding “charging” process is likely coupled with non-trivial changes in the protein structure and local hydration levels.^{90,91} Along this line, recent studies suggested that the

lack of explicit electronic polarization leads to errors more than 1 eV for the reduction potential of flavins in proteins;⁷⁹ this important observation is somewhat surprising as fixed-charge force field models have been quite successful at capturing *relative* reduction potential and pK_a values based on alchemical free energy simulations,^{92–94} provided that adequate sampling is done; one possible explanation is that the average polarization effect is empirically included in the fixed-charge force fields. Therefore, thorough comparison of different QM/MM models for both *absolute* and *relative* free energies with sufficient sampling is of great value. To enable such comparisons, advanced sampling techniques, including grand canonical Monte Carlo approaches for efficiently sampling local hydration level, are expected to be essential; subtle technical issues such as the effect of net charge change in Ewald summation, which are well understood,⁹⁵ should be considered when comparing results from different set ups, as many *ab initio* QM/MM simulations, especially those employ advanced MM models, still do not use periodic boundary condition while many semi-empirical QM/MM simulations do.^{96,97}

Another deviation from standard QM/MM setups is to employ coarse-grained (CG) models for at least part of the MM environment,^{98–100} which was motivated by applications that involve slow processes such as large-scale structural transitions of the biomolecule¹⁰¹ or protein design. Considering that QM/MM interaction terms are crucial to the accuracy of QM/MM simulations in general, it is challenging to construct reliable QM/CG models that are transferable among distinct conformational states, although solid progress have been reported.⁹⁸

2.2 Local Sampling: Multi-dimensional and Multi-level free energy simulations

For effective sampling, depending on the nature of the bottleneck, different strategies are required. Thus we separately discuss several “burning issues” related to local sampling and remote (allosteric) effects in this and subsequent subsections, respectively. We focus on issues most relevant to biomolecular QM/MM applications, since excellent reviews are available on the general topic of enhanced sampling for biomolecules.^{102,103}

2.2.1 Multi-dimensional free energy simulations: competitive pathways and causal relations among different processes—One could argue that to capture the key features of the chemical step(s) in most enzymes, standard QM/MM free energy simulations (e.g., umbrella sampling and metadynamics) using one or two collective variables (CVs) are likely adequate. For more complex transformations that explicitly involve multiple catalytic groups, such as ATP hydrolysis in molecular motors, finite temperature string methods¹⁰⁴ that parameterize the minimum free energy path with a large number of CVs can be very effective.^{105,106} A direction of major interest is to develop the finite temperature analog of automated and exhaustive reaction path searches^{107,108} on potential energy surfaces such that catalytic mechanism analysis for multi-step reactions in enzymes can be done in an automated fashion without prior bias by human intuition.

On the other hand, some problems require alternative approaches. For example, one hallmark that distinguishes biomolecules from artificial catalysts is that the chemical activities in the former are often tightly coupled to other processes. As a result, chemical reactions

can be used in biology to drive other events such as conformational transitions or pumping ions across the cell membrane, forming the basis of energy and signal transductions in cells.¹⁰⁹ As the other side of the coin, the level of chemical activity in biomolecules can be significantly perturbed by processes that occur either proximal or distal to the active site, giving rise to a multitude of regulatory mechanisms of enzyme catalysis. Therefore, understanding the physical principles that govern the *coupling* between the chemical step and other events, such as penetration of water molecules into the active site,^{110–112} recruitment of transient metal ions,^{113,114} or conformational rearrangements near and afar,^{115–119} is of great fundamental and practical importance.

For such problems, the most revealing approach that can clearly elucidate the causality among different processes is to conduct multi-dimensional free energy simulations. This is computationally demanding and therefore particularly requires a balance between computational accuracy and efficiency; semi-empirical QM/MM methods, perhaps augmented with ML corrections, are uniquely appropriate in this context. Even with inexpensive QM/MM methods, it is essential to ensure optimal efficiency in building up the free energy surface and allocating computational resources for the sampling of the relevant CV space; the selection of the appropriate set of CVs is not unique to QM/MM simulations and has been discussed extensively in the literature.^{69,103}

Along this line, ML approaches can again offer simple and effective solutions. For example, Zhang et al.¹²¹ developed a reinforcement learning approach in which a neural network ensemble is used to learn the multi-dimensional free energy surface. The approach takes advantage of the numerical flexibility of neural networks to parameterize high-dimensional functions; in addition, the use of an ensemble of neural networks makes it straightforward to identify regions of the CV space that have been undersampled, so that computational resources are automatically re-allocated accordingly. The approach can be straightforwardly applied to QM/MM simulations (Fig. 3) to analyze competition of multiple reaction pathways as well as coupling between the chemical step and other local processes, such as metal ion trafficking. In the current form, the approach does not actively enhance the sampling of degrees of freedom not included in the CV space, which again is not a unique challenge to QM/MM simulations.

2.2.2 Multi-level free energy simulations: bridging the distributions at different levels of theory—In quantum chemistry, it is routine to combine different levels of theory to focus on structure and energetics, respectively.¹²² It is therefore natural to pursue “multi-level” free energy simulations^{123,124} in which an inexpensive QM/MM potential is used to explore the conformational space and a more accurate QM/MM potential is used to improve the energetics. The fundamental challenge for condensed phase simulations at a finite temperature is that the configurations with high Boltzmann weights may differ significantly at different levels of theory, thus merely re-weighting of samples collected from a low-level simulation with high-level energies¹²⁵ is unlikely to lead to proper convergence,¹²⁶ especially when the QM region is large (for example, 100–250 atoms, as in many semi-empirical QM/MM simulations¹²⁷).

As discussed extensively in the literature, the success of “multi-level” free energy simulations depends critically on the overlap of the configurational distributions; specifically in the framework of free energy perturbation upon changing potential energy, one useful metric is the overlap in the energy gap (i.e., $U^{LH}(\mathbf{X}) = U^H(\mathbf{X}) - U^L(\mathbf{X})$) distribution at the two levels (L, H) of theory ($\rho_L(U^{LH})$ vs. $\rho_H(U^{LH})$), as reflected by the well-established identity,¹²⁸

$$e^{-\beta\Delta U^{LH}} \rho_L(\Delta U^{LH}) = e^{-\beta\Delta A^{LH}} \rho_H(\Delta U^{LH}), \quad (2)$$

in which A^{LH} is the free energy difference between the two levels of theory; more quantitative criteria for the degree of overlap have been proposed in the literature,^{129–131} many of which are based on the work of Kofke and co-workers for free energy perturbation in general.¹³² Therefore, the key challenge is to establish protocols that ensure adequate overlap during the L to H transformation while maintaining computational efficiency, i.e., to minimize the amount of computations at the H level.

One natural approach is to improve the L level towards the H level, by either system-specific reparameterization of L,¹³³ learning the difference between L and H on the fly via ML,⁷² or switching from L to H via many short, non-equilibrium simulations¹³⁴ in which L and H potentials are mixed explicitly. Alternatively, one can systematically identify the degrees of freedom that lead to poor energy gap overlaps at L/H levels, and treat these “problematic” degrees of freedom separately. Several studies of systems that included relatively small QM regions suggested that the bonded, stiff degrees of freedom tend to be problematic,^{131,135–137} as even small structural differences between L and H can have significant energetic consequences and thus leading to a poor overlap in energy gap distributions. Different approaches have been developed accordingly, which involve either refitting these degrees of freedom via, for example, force-matching,^{131,138} or ignoring them so as to focus on interaction energies between the QM and MM degrees of freedom.^{139,140}

In the “staged transformation” approach explored by us recently,¹⁴¹ these problematic degrees of freedom are subject to constraints or restraints, such that the conversion from L to H models is done through a thermodynamic route that ensures favorable distribution overlaps along the way. The free energy components associated with different steps are mostly evaluated explicitly, thus the final result can be compared to the rigorous free energy difference between the two levels of theory with limited and well-defined approximations. Importantly, the additional free energy component calculations involve simulations at the low level of theory and therefore do not incur high computational costs. The approach has been illustrated with solution systems with encouraging results, although further tests and improvements are required for treating more complex systems.

For systems that involve larger QM regions, however, it is not clear at all that only bonded degrees of freedom are problematic, as small errors associated with non-bonded degree of freedom can accumulate quickly, leading to poor energy gap overlaps. Therefore, additional developments are sorely needed to automatically identify the “problematic” degrees of freedom and mitigate their impact on the computed free energies. Along this line, we note that broadening (as opposed to narrowing, as commonly done in previous work) the

distribution at the L level and/or introducing intermediate distributions that bridge the L/H distributions with generalized ensemble based sampling^{102,142} or generative ML methods¹⁴³ are interesting alternative directions to pursue.

2.3 Remote Effects: Allostery, Enzyme Evolution and Design

As alluded to above, one fascinating aspect about biomolecules is the existence of coupling, or co-operativity, among distant sites. Although the mechanism of allostery in biomolecules has been studied for decades,^{144–149} it remains difficult to precisely predict residues that dictate the long-range co-operativity; this incomplete level of understanding limits our ability to engineer allostery into biomolecules, although sporadic successful examples have been reported in the literature.^{149–151} Specifically in the context of enzyme catalysis, it has been well documented that remote mutations may have a significant impact on catalysis,¹¹⁸ leading to hypotheses and debates about the roles of enzyme motions in catalysis. In directed evolution studies of designed enzymes, it is often observed that improvement of catalytic proficiency or expansion of substrate scope involves residues not in the active site.^{118,152–154} In enzyme evolution studies,¹⁵⁵ fitness landscape of several enzymes has been shown to be determined by residues distributed throughout the enzyme structure,¹⁵⁴ again highlighting the holistic nature of protein function.

In some cases, the roles of remote residues can be intuitively understood at a structural level;¹¹⁹ i.e., through a domino or Goldberg machine fashion, remote mutation(s) perturb the dominant conformation of active site residues^{156,157} and therefore the chemical activity therein, including both catalytic proficiency and substrate scope. The challenge for any computational analysis is thus to capture such “population shifts” due to distal mutations. Several compelling examples have been reported recently using enhanced sampling techniques, such as path-variable based metadynamics simulations¹⁰³ for variants of Tryptophan synthase subunit B (TrpB);¹⁵⁸ the simulations showed that the populations of several conformational states of TrpB were modulated by distal mutations, providing a molecular level rationale for the emergence of these mutation in directed evolution studies^{159,160} that engineered TrpB into a standalone enzyme in the absence of the neighboring *α* subunit.

However, in other cases, the effects of distal changes appear more indirect. For example, in our recent analysis of ATP hydrolysis of myosin,¹⁰⁶ we studied two models for the motor domain that have almost identical conformations for residues in the nucleotide binding site but differ in distal regions and hydration of the critical R238-E459 salt bridge (Fig. 5a). String based DFTB3/MM free energy simulations found that activation free energy and exergonicity in the two models differ by as much as 9 kcal/mol, highlighting that the average ground state configurations of catalytic residues are not the only determinant. Indeed, the distal residues can instead modify the fluctuations of active site residues and water molecules, and thus the reorganization energy or entropic contributions to the activation free energy.⁴⁵ In a different context, elegant EVB analysis of Aqvist and co-workers have clearly shown that flexibility of surface protein residues plays a major role in modulating the enthalpic and entropic components of activation free energy,¹⁶¹ leading to different temperature adaptations for mesophilic and psychrophilic enzymes.⁴⁸

The temperature dependence of enzymes has led to many fascinating discussions regarding the mechanisms that govern enzyme stability and catalysis.^{48,162–165} A particularly intriguing phenomenon noted in recent literature concerns non-linearity in the Arrhenius plot at high temperatures,^{165–167} for which quantum mechanical tunneling of nuclei^{11,168,169} is unlikely the cause. Two different models have been proposed to explain the non-linear Arrhenius plot: conformational redistribution^{170,171} vs. activation heat capacity.^{166,172} In the former model, the equilibrium between catalytically active and inactive conformations is perturbed by temperature, leading to different populations of the active conformation and therefore different apparent rate constants. In the latter case, a non-vanishing heat capacity difference between the ground and transition states (ΔC_p^\ddagger) makes the activation free energy temperature-dependent, leading also to deviation of the Arrhenius plot from linearity. Both thermodynamic models can be used to fit available experimental kinetic data, and for different systems, both models have been supported with microscopic analysis based on extensive molecular dynamics simulations,^{167,171–173} respectively. As discussed by Aqvist and co-workers,¹⁷¹ the two models do lead to significantly different predictions about activation enthalpy and entropy for broader temperatures, for which no experimental data are yet available. On the other hand, it has been suggested that the different proposals consistently point toward a general picture of more than one reactant state before passing the transition state.¹⁶⁵

A particularly interesting case concerns the de novo designed enzyme for catalyzing the Kemp elimination reaction.¹⁷⁴ While the designed enzyme, which has low catalytic proficiency, features a linear Arrhenius plot and therefore zero activation heat capacity, its more efficient variant following several generations of directed evolution exhibits a non-linear Arrhenius plot, which can be fitted with a negative ΔC_p^\ddagger . Mulholland and co-workers¹⁷³ showed that extensive ($\sim 5 \mu s$) classical MD simulations with a MM model for the transition state indeed recapitulated a negative ΔC_p^\ddagger , which was estimated based on the fluctuation of the protein energy in the MD simulations. Further analysis of the trajectories found that the negative ΔC_p^\ddagger in the more active enzyme variant was likely the result of a more compact active site in the transition state due to a higher population of an active site loop in its closed conformation. The more compact structure is also congruent with extensive correlated motions throughout the protein (Fig.5b). Therefore, an interesting notion from the analysis is that to further enhance catalytic proficiency of designed enzymes, one strategy might be to enhance correlated motions that implicate both active site and distal regions; this point was also made in the aforementioned study of TrpB,¹⁵⁸ in which the distal mutation sites observed in the directed evolution study were also identified with a motional network analysis for the wild type enzyme.

Evidently, the causal relation between correlated motion, negative ΔC_p^\ddagger and catalytic proficiency remains to be further analyzed. Nevertheless, these recent studies further highlight the significance of better understanding collective motions of enzymes, their thermodynamic signature and factors that control the nature and functional implication of such motions. Along this line, close integration of extensive classical and QM/MM studies with recent breakthrough in continuous evolution techniques,^{175–177} which makes

it straightforward to evolve enzymes for many (>100) generations in independent trajectories,¹⁷⁸ is poised to lead to significant insights into new strategies that can be used to enhance the catalytic proficiency of designed enzymes.

3 Concluding Remarks

In this short perspective, we discussed a number of “burning issues” related to biomolecular QM/MM simulations. Although most of these discussions are motivated primarily by our own research interest, we believe many of the discussions are relevant to biomolecular QM/MM applications in general. Many aspects of the discussed issues are related to establishing “best practice” for realistic applications, such as choices of the QM region size, QM and MM potential functions, and methodologies that best integrate physical methods and established machine learning techniques for efficient free energy simulations. There are, nevertheless, in our opinion, fundamental conceptual issues, such as developing efficient methods for better treating transition metal ions,^{179,180} pursuing generally applicable machine learning models for both short-range and long-range interactions,^{67,181} identifying problems for which the explicit inclusion of electronic polarization makes a qualitative difference, and clearly dissecting and defining the roles of distal residues and specific motions in enzyme catalysis.

In terms of connections with experiments, while it remains important to compute multiple observables that characterize physical properties of biomolecules, such as kinetic isotope effects, free energy relations and various spectroscopies,¹⁸² an emerging opportunity is to integrate computations with high-throughput experiments. For the analysis of protein allostery, for example, deep mutation studies¹⁸³ have revealed a rather different picture for the distribution of “hotspot” residues that are essential to co-operativity.¹⁸⁴ Similarly, continuous evolution studies are starting to generate large datasets on mutation effects on catalytic properties.¹⁷⁸ Providing a molecular level understanding for these rich datasets and making testable predictions with multi-faceted computational analysis is a formidable yet exciting challenge. To paraphrase one of the ten big ideas from the National Science Foundation, developing novel methodologies that can truly embrace “the data revolution” is of great urgency. With further development and integration with data science/machine learning techniques, QM/MM methods will play increasingly essential roles in the mechanistic analysis and rational design of novel biomolecular functions.^{119,185}

Before closing, we note that exciting developments in *ab initio* quantum chemistry methodologies continue to take place at a pace as rapid as ever.¹⁸⁶ We also can't resist mentioning that major strides are being made in the areas of quantum Monte Carlo¹⁸⁷ and related ML methods¹⁸⁸ as well as quantum computing,^{189,190} in terms of both fundamental algorithms and hardware. While these more advanced and expensive methodologies are yet to directly impact biophysical and biochemical applications, it is clear that the list of powerful tools available to multi-scale computations will continue to expand rapidly and therefore the future is undoubtedly bright.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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Biography

Qiang Cui

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References

- (1). Warshel A; Levitt M Theoretical Studies of Enzymic Reactions - Dielectric, Electrostatic and Steric Stabilization of Carbonium-Ion in Reaction of Lysozyme. *J. Mol. Biol* 1976, 103, 227–249. [PubMed: 985660]
- (2). Field MJ; Bash PA; Karplus MA Combined Quantum-Mechanical and Molecular Mechanical Potential for Molecular-Dynamics Simulations. *J. Comput. Chem* 1990, 11, 700–733.

- (3). Lipkowitz KB, Boyd DB, Eds. Gao J, In Reviews in Computational Chemistry VII; VCH: New York, 1995; p 119.
- (4). Senn HM; Thiel WQM/MM methods for biomolecular systems. *Angew. Chem. Int. Ed*2009, 48, 1198–1229.
- (5). Friesner RA; Guallar VAb initio QM and QM/MM methods for studying enzyme catalysis. *Annu. Rev. Phys. Chem*2005, 56, 389–427. [PubMed: 15796706]
- (6). Hu H; Yang WFree Energies of Chemical Reactions in Solution and in Enzymes with Ab Initio Quantum Mechanics/Molecular Mechanics Methods. *Annu. Rev. Phys. Chem*2008, 59, 573–601. [PubMed: 18393679]
- (7). Kamerlin SCL; Haranczyk M; Warshel AProgress in Ab Initio QM/MM Free-Energy Simulations of Electrostatic Energies in Proteins: Accelerated QM/MM Studies of pK(a), Redox Reactions and Solvation Free Energies. *J. Phys. Chem. B*2009, 113, 1253–1272. [PubMed: 19055405]
- (8). Lu X; Fang D; Ito S; Okamoto Y; Ovchinnikov V; Cui QQM/MM Free Energy Simulations: Recent Progress and Challenges. *Mol. Simul. (Special Issue on Free Energy Simulations)*2016, 42, 1056–1078.
- (9). Gao JL; Ma SH; Major DT; Nam K; Pu JZ; Truhlar DGMechanisms and free energies of enzymatic reactions. *Chem. Rev*2006, 106, 3188–3209. [PubMed: 16895324]
- (10). Alexandrova AN; Rothlisberger D; Baker D; Jorgensen WLCatalytic Mechanism and Performance of Computationally Designed Enzymes for Kemp Elimination. *J. Am. Chem. Soc*2008, 130, 15907–15915. [PubMed: 18975945]
- (11). Gao JL; Truhlar DGQuantum mechanical methods for enzyme kinetics. *Annu. Rev. Phys. Chem*2002, 53, 467–505. [PubMed: 11972016]
- (12). Merz KM Jr., Using Quantum Mechanical Approaches to Study Biological Systems. *Acc. Chem. Res*2014, 47, 2804–2811. [PubMed: 25099338]
- (13). Shaik S; Cohen S; Wang Y; Chen H; Kumar D; Thiel WP450 Enzymes: Their Structure, Reactivity, and Selectivity-Modeled by QM/MM Calculations. *Chem. Rev*2010, 110, 949–1017. [PubMed: 19813749]
- (14). Lonsdale R; Mulholland AQM/MM Modelling of Drug-Metabolizing Enzymes. *Curr. Top. Med. Chem*2014, 14, 1339–1347. [PubMed: 24805066]
- (15). Brunk E; Rothlisberger UMixed Quantum Mechanical/Molecular Mechanical Molecular Dynamics Simulations of Biological Systems in Ground and Electronically Excited States. *Chem. Rev*2015, 115, 6217–6263. [PubMed: 25880693]
- (16). Amaro RE; Mulholland AJMultiscale methods in drug design bridge chemical and biological complexity in the search for cures. *Nat. Rev. Chem*2018, 2, 0148. [PubMed: 30949587]
- (17). Chung LW; Sameera WMC; Ramozzi R; Page AJ; Hatanaka M; Petrova GP; Harris TV; Li X; Ke ZF; Liu FYet al., The ONIOM Method and Its Applications. *Chem. Rev*2015, 115, 5678–5769. [PubMed: 25853797]
- (18). Metz S; Kästner J; Sokol AA; Keal TW; Sherwood PChemShell—a modular software package for QM/MM simulations. *WIREs Comput. Mol. Sci*2013, 4, 101–110.
- (19). Fürstner A; List B; Ritter T; Schüth F; Neese FWalter Thiel (1949–2019). *Angew. Chem. Int. Ed*2020, 59, 1382–1383.
- (20). Sokol AA; Bromley ST; French SA; Richard C; Catlow A; Sherwood PHybrid QM/MM Embedding Approach for the Treatment of Localized Surface States in Ionic Materials. *Int. J. Quant. Chem*2004, 99, 695–712.
- (21). Golze D; Iannuzzi M; Nguyen MT; Passerone D; Hutter JSimulation of Adsorption Processes at Metallic Interfaces: An Image Charge Augmented QM/MM Approach. *J. Chem. Theory Comput*2013, 9, 5086–5097. [PubMed: 26583423]
- (22). Magnussen OM; Gross AToward an atomic-scale understanding of electrochemical interface structure and dynamics. *J. Am. Chem. Soc*2019, 141, 4777–4790. [PubMed: 30768905]
- (23). Bruix A; Margraf JT; Andersen M; Reuter KFirst-principles-based multiscale modelling of heterogeneous catalysis. *Nat. Cata*2019, 2, 659–670.
- (24). Huang P; Carter EAAdvances in correlated electronic structure methods for solids, surfaces, and nanostructures. *Annu. Rev. Phys. Chem*2008, 59, 261–290. [PubMed: 18031211]

- (25). Sakong S; Gross A The electric double layer at metal-water interfaces revisited based on a charge polarization scheme. *J. Chem. Phys* 2018, 149, 084705. [PubMed: 30193475]
- (26). Sakong S; Gross A Water structures on a Pt(111) electrode from ab initio molecular dynamic simulations for a variety of electrochemical conditions. *Phys. Chem. Chem. Phys* 2020, 22, 10431. [PubMed: 31976502]
- (27). Le JB; Cheng J Modeling electrochemical interfaces from ab initio molecular dynamics: water adsorption on metal surfaces at potential of zero charge. *Curr. Opin. Electrochem* 2020, 19, 129–136.
- (28). Liang DY; Hong JW; Dong F; Bennett JW; Mason SE; Hamers RJ; Cui Q Analysis of Conformational Properties of Amine Ligands at the Gold/Water Interface with QM, MM and QM/MM simulations. *Phys. Chem. Chem. Phys* 2018, 20, 3349–3362. [PubMed: 29226924]
- (29). Heenen HH; Gauthier JA; Kristoffersen HH; Ludwig T; Chan K Solvation at metal/water interfaces: An ab initio molecular dynamics benchmark of common computational approaches. *J. Chem. Phys* 2020, 152, 144703. [PubMed: 32295363]
- (30). Ringe S; Oberhofer H; Hille C; Matera S; Reuter K Function-Space-Based Solution Scheme for the Size-Modified Poisson-Boltzmann Equation in Full-Potential DFT. *J. Chem. Theory Comput* 2016, 12, 4052–4066. [PubMed: 27323006]
- (31). Sumowski CV; Ochsenfeld CA Convergence Study of QM/MM Isomerization Energies with the Selected Size of the QM Region for Peptidic Systems. *J. Phys. Chem. A* 2009, 113, 11734–11741. [PubMed: 19585981]
- (32). Flaig D; Beer M; Ochsenfeld C Convergence of Electronic Structure with the Size of the QM Region: Example of QM/MM NMR Shieldings. *J. Chem. Theory Comput* 2012, 8, 2260–2271. [PubMed: 26588959]
- (33). Hu L; Söderhjelm P; Ryde U On the Convergence of QM/MM Energies. *J. Chem. Theory Comput* 2011, 7, 761–777. [PubMed: 26596307]
- (34). Roßbach S; Ochsenfeld C Influence of Coupling and Embedding Schemes on QM Size Convergence in QM/MM Approaches for the Example of a Proton Transfer in DNA. *J. Chem. Theory Comput* 2017, 13, 1102–1107. [PubMed: 28195707]
- (35). Kulik HJ; Zhang J; Klinman JP; Martínez TJ How Large Should the QM Region Be in QM/MM Calculations? The Case of Catechol O-Methyltransferase. *J. Phys. Chem. B* 2016, 120, 11381–11394. [PubMed: 27704827]
- (36). Jindal G; Warshel A Exploring the Dependence of QM/MM Calculations of Enzyme Catalysis on the Size of the QM Region. *J. Phys. Chem. B* 2016, 120, 9913–9921. [PubMed: 27552257]
- (37). Kulik HJ Large-scale QM/MM free energy simulations of enzyme catalysis reveal the influence of charge transfer. *Phys. Chem. Chem. Phys* 2018, 20, 20650–20660. [PubMed: 30059109]
- (38). Das S; Nam K; Major D Rapid Convergence of Energy and Free Energy Profiles with Quantum Mechanical Size in Quantum Mechanical–Molecular Mechanical Simulations of Proton Transfer in DNA. *J. Chem. Theory Comput* 2018, 14, 1695–1705. [PubMed: 29446946]
- (39). Mehmood R; Kulik HJ Both Configuration and QM Region Size Matter: Zinc Stability in QM/MM Models of DNA Methyltransferase. *J. Chem. Theory Comput* 2020, 16, 3121–3134. [PubMed: 32243149]
- (40). Watanabe H; Cui Q Quantitative analysis and correction of QM/MM boundary artifacts in adaptive QM/MM methods. *J. Chem. Theory Comput* 2019, 15, 3917–3928. [PubMed: 31095912]
- (41). Thiel W Perspectives on semiempirical molecular orbital theory. *Adv. Chem. Phys* 1996, 93, 703–757.
- (42). Gaus M; Cui Q; Elstner M Density Functional Tight Binding (DFTB): Application to organic and biological molecules. *WIREs Comput. Mol. Sci* 2014, 4, 49–61.
- (43). Warshel A Computer Modeling of Chemical Reactions in Enzymes and Solution; Wiley, New York, 1991.
- (44). Ruiz-Perinà JJ; Silla E; Tuñón I; Martí S Hybrid Quantum Mechanics/Molecular Mechanics Simulations with Two-Dimensional Interpolated Corrections: Application to Enzymatic Processes. *J. Phys. Chem. B* 2006, 110, 17663–17670. [PubMed: 16942112]

- (45). Aqvist J; Kazemi M; Isaksen GV; Brandsdal BO Entropy and Enzyme Catalysis. *Acc. Chem. Res*2017, 50, 199–207. [PubMed: 28169522]
- (46). Kulkarni YS; Amyes TL; Richard J; Kamerlin SCL Uncovering the Role of Key Active-Site Side Chains in Catalysis: An Extended Bronsted Relationship for Substrate Deprotonation Catalyzed by Wild-Type and Variants of Triosephosphate Isomerase. *J. Am. Chem. Soc*2019, 141, 16139–16150. [PubMed: 31508957]
- (47). Nomura Y; Roston D; Montemayor EJ; Cui Q; Butcher SE Structural and Mechanistic Basis for Preferential Deadenylation of U6 snRNA by Usb1. *Nuc. Acids Res*2018, 46, 11488–11501.
- (48). Aqvist J; Isaksen GV; Brandsdal BO Computation of enzyme cold adaptation. *Nat. Rev. Chem*2017, 1, 0051.
- (49). Kussmann J; Beer M; Ochsenfeld CL Linear-scaling self-consistent field methods for large molecules. *WIREs Comput. Mol. Sci*2013, 3, 614–636.
- (50). Seritan S; Bannwarth C; Fales BS; Hohenstein EG; Kokkila-Schumacher SIL; Leuhr N; Snyder JW; Song CC; Titov AV; Ufimtsev I Set al., TeraChem: Accelerating electronic structure and ab initio molecular dynamics with graphical processing units. *J. Chem. Phys*2020, 152, 224110. [PubMed: 32534542]
- (51). Christensen AS; Kubar T; Cui Q; Elstner M, Semi-empirical Quantum Mechanical Methods for Non-covalent Interactions for Chemical and Biochemical Applications. *Chem. Rev*2016, 116, 5301–5337. [PubMed: 27074247]
- (52). Bannwarth C; Ehlert S; Grimme SG FN2-xTB-An Accurate and Broadly Parametrized Self-Consistent Tight-Binding Quantum Chemical Method with Multipole Electrostatics and Density-Dependent Dispersion Contributions. *J. Chem. Theory Comput*2019, 15, 1652–1671. [PubMed: 30741547]
- (53). Bannwarth C; Caldeweyher E; Ehlert S; Hansen A; Pracht P; Seibert J; Spicher S; Grimme SE Extended tight-binding quantum chemistry methods. *WIREs Comput. Mol. Sci*2020, e01493.
- (54). Redders GR; Babin V; Paesani FA Critical Assessment of Two-Body and Three-Body Interactions in Water. *J. Chem. Theory Comput*2013, 9, 1103–1114. [PubMed: 26588754]
- (55). Christensen AS; Kromann JC; Jensen JH; Cui Q Intermolecular Interactions in the Condensed Phase: Evaluation of Semi-empirical Quantum Mechanical Methods. *J. Chem. Phys*2017, 147, 161704. [PubMed: 29096452]
- (56). Vanommeslaeghe K; Hatcher E; Acharya C; Kundu S; Zhong S; Shim J; Darian E; Guvench O; Lopes P; Vorobyov I et al., CHARMM General Force Field (CGenFF): A force field for drug-like molecules compatible with the CHARMM allatom additive biological force fields. *J. Comput. Chem*2010, 31, 671–690. [PubMed: 19575467]
- (57). Weinhold F; Landis CR Valency and Bonding; Cambridge University Press, 2005.
- (58). Lu X; Duchimaza-Heredia J; Cui Q Analysis of Density Functional Tight Binding (DFTB) with Natural Bonding Orbitals (NBOs). *J. Phys. Chem. A*2019, 123, 7439–7453. [PubMed: 31373822]
- (59). Kulik HJ Perspective: Treating electron over-delocalization with the DFT+U method. *J. Chem. Phys*2016, 142, 240901.
- (60). Griffith JS The Theory of Transition-Metal Ions; Cambridge University Press, 1963.
- (61). Singh SK; Eng J; Atanasov M; Neese FC Covalency and chemical bonding in transition metal complexes: An ab initio based ligand field perspective. *Coord. Chem. Rev*2017, 344, 2–25.
- (62). Stepanovic S; Lai R; Elstner M; Gruden M; Garcia-Fernandez P; Cui Q Improvement of d-d interactions in Density Functional Tight Binding for Transition Metal Ions with a Ligand Field Model: Assessment of a DFTB3+U model on Nickel coordination compounds. *Phys. Chem. Chem. Phys*2020, In press (10.1039/D0CP04694A).
- (63). Vujovic M; Huynh M; Steiner S; Garcia-Fernandez P; Elstner M.; Cui Q; Gruden M Exploring the Applicability of Density Functional Tight Binding to Transition Metal Ions: Parametrization for Nickel with the Spin-polarized DFTB3 model. *J. Comput. Chem*2019, 40, 400–413. [PubMed: 30299559]
- (64). Filatov M Spin-restricted ensemble-referenced Kohn-Sham method: basic principles and application to strongly correlated ground and excited states of molecules. *WIREs Comput. Mol. Sci*2015, 5, 146–167.

- (65). Grofe A; Chen X; Liu WJ; Gao JL Spin-Multiplet Components and Energy Splittings by Multistate Density Functional Theory. *J. Phys. Chem. Lett* 2017, 8, 4838–4845. [PubMed: 28914545]
- (66). Noe F; Tkatchenko A; Müller KR; Clementi C Machine Learning for Molecular Simulation. *Annu. Rev. Phys. Chem* 2020, 71, 361–390. [PubMed: 32092281]
- (67). Unke OT; Chmiela S; Sauceda HE; Gastegger M; Poltavsky I; Schütt KT; Tkatchenko A; Müller K R Machine Learning Force Fields. *arXiv:2010.07067v1* 2020,
- (68). von Lilienfeld OA; Müller K-R; Tkatchenko A Exploring chemical compound space with quantum-based machine learning. *Nat. Rev. Chem* 2020, 4, 347–358.
- (69). Wang YH; Ribeiro JML; Tiwary P Machine learning approaches for analyzing and enhancing molecular dynamics simulations. *Curr. Opin. Struct. Biol* 2020, 61, 139–145. [PubMed: 31972477]
- (70). Janet JP; Kulik HJ Predicting electronic structure properties of transition metal complexes with neural networks. *Chem. Sci* 2017, 8, 5137–5152. [PubMed: 30155224]
- (71). Duan CR; Liu F; Nandy A; Kulik HJ Semi-supervised Machine Learning Enables the Robust Detection of Multireference Character at Low Cost. *J. Phys. Chem. Lett* 2020, 11, 6640–6648. [PubMed: 32692570]
- (72). Shen L; Yang W T Molecular Dynamics Simulations with Quantum Mechanics/Molecular Mechanics and Adaptive Neural Networks. *J. Chem. Theory. Comput* 2018, 14, 1442–1455. [PubMed: 29438614]
- (73). Qiao Z; Welborn M; Anandkumar A; Manby FR; Miller III TF, OrbNet: Deep Learning for Quantum Chemistry Using Symmetry-Adapted Atomic-Orbital Features. *J. Chem. Phys* 2020, 153, 124111. [PubMed: 33003742]
- (74). Zhang L; Han J; Wang H; Car R; E W Deep Potential Molecular Dynamics: A Scalable Model with the Accuracy of Quantum Mechanics. *Phys. Rev. Lett* 2018, 120, 143001. [PubMed: 29694129]
- (75). Wang H; Zhang L; Han J; E W DeePMD-kit: A deep learning package for many-body potential energy representation and molecular dynamics. *Comput. Phys. Comm* 2018, 228, 178–184.
- (76). Ganguly A; Boulanger E; Thiel W Importance of MM Polarization in QM/MM Studies of Enzymatic Reactions: Assessment of the QM/MM Drude Oscillator Model. *J. Chem. Theory Comput* 2017, 13, 2954–2961. [PubMed: 28437096]
- (77). Wanko M; Hoffmann M; Strodel P; Koslowski A; Thiel W; Neese F; Frauenheim T; Elstner M Calculating absorption shifts for retinal proteins: Computational challenges. *J. Phys. Chem. B* 2005, 109, 3606–3615. [PubMed: 16851399]
- (78). Mennucci B Modeling environment effects on spectroscopies through QM/classical models. *Phys. Chem. Chem. Phys* 2013, 15, 6583–6594. [PubMed: 23385350]
- (79). Tazhigulov RN; Gurunathan PK; Kim Y; Slipchenko LV; Bravaya KB Polarizable embedding for simulating redox potentials of biomolecules. *Phys. Chem. Chem. Phys* 2019, 21, 11642–11650. [PubMed: 31116217]
- (80). Goyal P; Lu J; Yang S; Gunner MR; Cui Q Changing hydration level in an internal cavity modulates the proton affinity of a key glutamate in Cytochrome c Oxidase. *Proc. Natl. Acad. Sci. U.S.A.* 2013, 110, 18886–18891. [PubMed: 24198332]
- (81). Wanko M; Hoffmann M; Frahmcke J; Frauenheim T; Elstner M Effect of polarization on the opsin shift in rhodopsins. 2. empirical polarization models for proteins. *J. Phys. Chem. B* 2008, 112, 11468–11478. [PubMed: 18729405]
- (82). Loco D; Lagardere L; Cisneros GA; Scalmani G; Frisch M; Lipparini F; Mennucci B; Piquemal J P Towards large scale hybrid QM/MM dynamics of complex systems with advanced point dipole polarizable embeddings. *Chem. Sci* 2019, 10, 7200–7211. [PubMed: 31588288]
- (83). Bondanza M; Nottoli M; Cupellini L; Lipparini F; Mennucci B Polarizable embedding QM/MM: the future gold standard for complex (bio)systems? *Phys. Chem. Chem. Phys* 2020, 22, 14433–14448. [PubMed: 32588851]
- (84). Lambros E; Lipparini F; Cisneros GA; Paesani FA Many-Body, Fully Polarizable Approach to QM/MM Simulations. *J. Chem. Theory Comput* 2020, In press, (10.1021/acs.jctc.0c00932).

- (85). Welborn VV; Pestana LR; Head-Gordon T Computational optimization of electric fields for better catalysis design. *Nat. Catal* 2018, 1, 649–655.
- (86). Simonson T; Perahia D Internal and interfacial dielectric properties of cytochrome c from molecular dynamics in aqueous solution. *Proc. Natl. Acad. Sci. U.S.A* 1995, 92, 1082–1086. [PubMed: 7862638]
- (87). Warshel A Computer simulations of enzyme catalysis: Methods, Progress and Insights. *Annu. Rev. Biophys. Biomol. Struct* 2003, 32, 425–443. [PubMed: 12574064]
- (88). Riccardi D; Cui Q pK_a analysis for the zinc-bound water in Human Carbonic Anhydrase II: benchmark for "multi-scale" QM/MM simulations and mechanistic implications. *J. Phys. Chem. A* 2007, 111, 5703–5711. [PubMed: 17506534]
- (89). Ghosh N; Cui Q pK_a of residue 66 in *Staphylococcal nuclease*: insights from QM/MM simulations with conventional sampling. *J. Phys. Chem. B* 2008, 112, 8387–8397. [PubMed: 18540669]
- (90). Chimenti MS; Castaneda CA; Majumdar A; Garcia-Moreno EB Structural Origins of High Apparent Dielectric Constants Experienced by Ionizable Groups in the Hydrophobic Core of a Protein. *J. Mol. Biol* 2011, 405, 361–377. [PubMed: 21059359]
- (91). Zheng YQ; Cui Q Microscopic Mechanisms that Govern the Titration Response and pK_a Values of Buried Residues in Staphylococcal Nuclease Mutants. *Proteins: Struct., Funct., & Bioinf* 2017, 85, 268–281.
- (92). Simonson T; Carlsson J; Case D Proton binding to proteins: $pK(a)$ calculations with explicit and implicit solvent models. *J. Am. Chem. Soc* 2004, 126, 4167–4180. [PubMed: 15053606]
- (93). Goh GB; Knight JL; Brooks III CL, Constant pH Molecular Dynamics Simulations of Nucleic Acids in Explicit Solvent. *J. Chem. Theo. Comp* 2012, 8, 36–46.
- (94). Huang YD; Chen W; Wallace JA; Shen J All-Atom Continuous Constant pH Molecular Dynamics With Particle Mesh Ewald and Titratable Water. *J. Chem. Theory Comput* 2016, 12, 5411–5421. [PubMed: 27709966]
- (95). Lin YL; Aleksandrov A; Simonson T; Roux B An Overview of Electrostatic Free Energy Computations for Solutions and Proteins. *J. Chem. Theory Comput* 2014, 10, 2690–2709. [PubMed: 26586504]
- (96). Riccardi D; Schaefer P; Cui Q pK_a calculations in solution and proteins with QM/MM free energy perturbation simulations. *J. Phys. Chem. B* 2005, 109, 17715–17733. [PubMed: 16853267]
- (97). Nam K; Gao JL; York D An efficient linear-scaling Ewald method for long-range electrostatic interactions in combined QM/MM calculations. *J. Chem. Theo. Comp* 2005, 1, 2–13.
- (98). Sinitskiy AV; Voth GA Quantum mechanics/coarse-grained molecular mechanics (QM/CG-MM). *J. Chem. Phys* 2018, 148, 014102. [PubMed: 29306280]
- (99). Valdez CE; Morgenstern A; Eberhart ME; Alexandrova AN Predictive methods for computational metalloenzyme redesign - a test case with carboxypeptidase A. *Phys. Chem. Chem. Phys* 2016, 18, 31744–31756. [PubMed: 27841396]
- (100). Sokkar P; Boulanger E; Thiel W; Sanchez-Garcia E Hybrid Quantum Mechanics/Molecular Mechanics/Coarse Grained Modeling: A Triple-Resolution Approach for Biomolecular Systems. *J. Chem. Theory Comput* 2015, 11, 1809–1818. [PubMed: 26574388]
- (101). Hocky GM; Dannenhoffer-Lafage T; Voth GA Coarse-Grained Directed Simulation. *J. Chem. Theory Comput* 2017, 13, 4593–4603. [PubMed: 28800392]
- (102). Zuckerman DMEquilibrium Sampling in Biomolecular Simulations. *Annu. Rev. Biophys* 2011, 40, 41–62. [PubMed: 21370970]
- (103). Valsson O; Tiwary P; Parrinello M Enhancing Important Fluctuations: Rare Events and Metadynamics from a Conceptual Viewpoint. *Annu. Rev. Phys. Chem* 2016, 67, 159–184. [PubMed: 26980304]
- (104). E W; Vanden-Eijnden E Transition-Path Theory and Path-Finding Algorithms for the Study of Rare Events. *Annu. Rev. Phys. Chem* 2010, 61, 391–420. [PubMed: 18999998]
- (105). Rosta E; Nowotny M; Yang W; Hummer G Catalytic Mechanism of RNA Back-bone Cleavage by Ribonuclease H from Quantum Mechanics/Molecular Mechanics Simulations. *J. Am. Chem. Soc* 2011, 133, 8934–8941. [PubMed: 21539371]

- (106). Lu X; Ovchinnikov V; Roston DR; Demapan D; Cui Q Regulation and Plasticity of Catalysis in Enzymes: Insights from Analysis of Mechanochemical Coupling in Myosin. *Biochem.* 2017, 56, 1482–1497. [PubMed: 28225609]
- (107). Maeda S; Ohno K; Morokuma K Systematic exploration of the mechanism of chemical reactions: the global reaction route mapping (GRRM) strategy using the ADDF and AFIR methods. *Phys. Chem. Chem. Phys.* 2013, 15, 3683–3701. [PubMed: 23389653]
- (108). Dewyer AL; Arguelles AJ; Zimmerman PM Methods for exploring reaction space in molecular systems. *WIREs Comput. Mol. Sci.* 2018, 8, e1354.
- (109). Nicholls DG; Ferguson SJ *Bioenergetics*, 3rd ed.; Academic Press: New York, 2002.
- (110). Son CY; Yethiraj A; Cui Q Cavity Hydration Dynamics in Cytochrome c Oxidase and Functional Implications. *Proc. Natl. Acad. Sci. USA* 2017, 114, E8830–E8836. [PubMed: 28973914]
- (111). Liang RB; Swanson MJ; Peng YX; Wikström M; Voth GA Multiscale simulations reveal key features of the proton-pumping mechanism in cytochrome c oxidase. *Proc. Natl. Acad. Sci. USA* 2016, 113, 7420–7425. [PubMed: 27339133]
- (112). Chakrabarty S; Warshel A Capturing the energetics of water insertion in biological systems: The water flooding approach. *Proteins: Struct., Funct., & Bioinf.* 2013, 81, 93–106.
- (113). Yang W; Weng PJ; Gao YA new paradigm of DNA synthesis: three-metal-ion catalysis. *Cell Biosci.* 2016, 6, 51. [PubMed: 27602203]
- (114). Samara NL; Yang W Cation trafficking propels RNA hydrolysis. *Nat. Struct. Mol. Biol.* 2018, 25, 715–721. [PubMed: 30076410]
- (115). Benkovic SJ; Hammes GG; Hammes-Schiffer S Free energy landscape of enzyme catalysis. *Biochem.* 2008, 47, 3317–3321. [PubMed: 18298083]
- (116). Nashine VC; Hammes-Schiffer S; Benkovic S Coupled motions in enzyme catalysis. *Curr. Opin. Chem. Biol.* 2010, 14, 644–651. [PubMed: 20729130]
- (117). Tokuriki N; Tawfik D S Protein Dynamics and Evolvability. *Science* 2009, 324, 203–207. [PubMed: 19359577]
- (118). Lee J; Goodey N M Catalytic Contributions from Remote Regions of Enzyme Structure. *Chem. Rev.* 2011, 111, 7595–7624. [PubMed: 21923192]
- (119). Crean RM; Gardner JM; Kamerlin S C L Harnessing Conformational Plasticity to Generate Designer Enzymes. *J. Am. Chem. Soc.* 2020, 142, 11324–11342. [PubMed: 32496764]
- (120). Duarte F; Åqvist J; Williams NH; Kamerlin S C L Resolving Apparent Conflicts between Theoretical and Experimental Models of Phosphate Monoester Hydrolysis. *J. Am. Chem. Soc.* 2015, 137, 1081–1093. [PubMed: 25423607]
- (121). Zhang LF; Wang H; E W N Reinforced dynamics for enhanced sampling in large atomic and molecular systems. *J. Chem. Phys.* 2018, 148, 124113. [PubMed: 29604808]
- (122). Helgaker T; Jorgensen P; Olsen J M *Molecular Electronic Structure Theory*; J. Wiley & Sons: West Sussex, England, 2000.
- (123). Gao J Absolute Free Energy of Solvation from Monte Carlo Simulations Using Combined Quantum and Molecular Mechanical Potentials. *J. Phys. Chem.* 1992, 96, 537–540.
- (124). Luzhkov V; Warshel A Microscopic Models for Quantum Mechanical Calculations of Chemical Processes in Solutions: LD/AMPAC and SCAAS/AMPAC Calculations of Solvation Energies. *J. Comp. Chem.* 1992, 13, 199–213.
- (125). König G; Hudson PS; Boresch S; Woodcock H L Multiscale free energy simulations: An efficient method for connecting classical MD simulations to QM or QM/MM free energies using Non-Boltzmann Bennett reweighting schemes. *J. Chem. Theo. Comp.* 2014, 10, 1406–1419.
- (126). Ryde U How Many Conformations Need To Be Sampled To Obtain Converged QM/MM Energies? The Curse of Exponential Averaging. *J. Chem. Theory Comput.* 2017, 13, 5745–5752. [PubMed: 29024586]
- (127). Roston D; Demapan D; Cui Q Extensive Free Energy Simulations Identify Water as the Base in Nucleotide Addition by DNA Polymerase. *Proc. Natl. Acad. Sci. USA* 2019, 116, 25048–25056. [PubMed: 31757846]
- (128). Pohorille A; Jarzynski C; Chipot C Good Practices in Free Energy Calculations. *J. Phys. Chem. B* 2010, 114, 10235–10253. [PubMed: 20701361]

- (129). Boresch S; Woodcock HL Convergence of single-step free energy perturbation. *Mol. Phys*2017, 115, 1200–1213.
- (130). König G; Brooks BR; Thiel W; York DM On the convergence of multi-scale free energy simulations. *Mol. Simulat*2018, 44, 1062–1081.
- (131). Giese TJ; York DM Development of a Robust Indirect Approach for MM → QM Free Energy Calculations That Combines Force-Matched Reference Potential and Bennett’s Acceptance Ratio Methods. *J. Chem. Theory Comput*2019, 15, 5543–5562. [PubMed: 31507179]
- (132). Lu N; Kofke DA Accuracy of free-energy perturbation calculations in molecular simulation. II. Heuristics. *J. Chem. Phys*2001, 115, 6866–6875.
- (133). Zhou Y; Ojeda-May P; Nagaraju M; Pu J Toward Determining ATPase Mechanism in ABC Transporters: Development of the Reaction Path-Force Matching QM/MM Method. *Methods in Enzymol.* 2016, 577, 185–212. [PubMed: 27498639]
- (134). Kearns FL; Hudson PS; Woodcock HL; Boresch S Computing Converged Free Energy Differences between Levels of Theory via Nonequilibrium Work Methods: Challenges and Opportunities. *J. Comput. Chem*2017, 38, 1376–1388. [PubMed: 28272811]
- (135). Kearns FL; Warrensford L; Boresch S; Woodcock HL The Good, the Bad, and the Ugly: “HiPen”, a New Dataset for Validating (S)QM/MM Free Energy Simulations. *Molecules*2019, 24, 681.
- (136). König G; Brooks BR Correcting for the free energy costs of bond or angle constraints in molecular dynamics simulations. *Biochim. Biophys. Acta*2015, 1850, 932–943. [PubMed: 25218695]
- (137). Heimdal J; Ryde U Convergence of QM/MM free-energy perturbations based on molecular-mechanics or semiempirical simulations. *Phys. Chem. Chem. Phys*2012, 14, 12592. [PubMed: 22797613]
- (138). Hudson PS; Boresch S; Rogers DM; Woodcock HL Accelerating QM/MM Free Energy Computations via Intramolecular Force Matching. *J. Chem. Theory Comput*2018, 14, 6327–6335. [PubMed: 30300543]
- (139). Hudson PS; Woodcock HL; Boresch S Use of Interaction Energies in QM/MM Free Energy Simulations. *J. Chem. Theory Comput*2019, 15, 4632–4645. [PubMed: 31142113]
- (140). Olsson MA; Soderhjelm P; Ryde U Converging Ligand-Binding Free Energies Obtained with Free-Energy Perturbations at the Quantum Mechanical Level. *J. Comp. Chem*2016, 37, 1589–1600. [PubMed: 27117350]
- (141). Ito S; Cui Q Multi-level Free Energy Simulation with a Staged Transformation Approach. *J. Chem. Phys*2020, 153, 044115. [PubMed: 32752685]
- (142). Andricioaei I; Straub J E Generalized simulated annealing algorithms using Tsallis statistics: Application to conformational optimization of a tetrapeptide. *Phys. Rev. E*1996, 53, R3055–R3058.
- (143). Noé F; Olsson S; Köhler J; Wu H Boltzmann generators: Sampling equilibrium states of many-body systems with deep learning. *Science*2019, 365, eaaw1147. [PubMed: 31488660]
- (144). Cui Q; Karplus M Allosterity and cooperativity revisited. *Protein Science*2008, 17, 1295–1307. [PubMed: 18560010]
- (145). Nussinov R; Tsai C J Allosterity in Disease and in Drug Discovery. *Cell*2013, 153, 293–305. [PubMed: 23582321]
- (146). Changeux J P Allosterity and the Monod-Wyman-Changeux Model After 50 Years. *Annu. Rev. Biophys*2012, 41, 103–133. [PubMed: 22224598]
- (147). Motlagh HN; Wrabl JO; Li J; Hilser V J The ensemble nature of allosterity. *Nature*2014, 508, 331–339. [PubMed: 24740064]
- (148). Marzen S; Garcia H G; Phillips R Statistical Mechanics of Monod-Wyman-Changeux (MWC) Models. *J. Mol. Biol*2013, 425, 1433–1460. [PubMed: 23499654]
- (149). Raman A S; White K I; Ranganathan R Origins of allosterity and evolvability in proteins: a case study. *Cell*2016, 166, 468–480. [PubMed: 27321669]
- (150). Taylor N D; Garruss A S; Moretti R; Chan S; Arbing M A; Cascio D; Rogers J K; Isaacs F J; Kosuri S; Baker D et al., Engineering an allosteric transcription factor to respond to new ligands. *Nat. Methods*2016, 13, 177+. [PubMed: 26689263]

- (151). Kuhlman B; Bradley PAdvances in protein structure prediction and design. *Nat. Rev. Mol. Cell Biol*2019, 20, 681–697. [PubMed: 31417196]
- (152). Yang G; Hong N; Baier F; Jackson CJ; Tokuriki NConformational tinkering drives evolution of a promiscuous activity through indirect mutational effects. *Biochem.* 2016, 55, 4583–4593. [PubMed: 27444875]
- (153). Oue S; Okamoto A; Yano T; Kagamiyama HRedesigning the substrate specificity of an enzyme by cumulative effects of the mutations of non-active site residues. *J. Biol. Chem*1999, 274, 2344–2349. [PubMed: 9891001]
- (154). Wrenbeck EE; Azouz LR; Whitehead TASingle-mutation fitness landscapes for an enzyme on multiple substrates reveal specificity is globally encoded. *Nat. Comm*2016, 8, 15695.
- (155). Romero A; Arnold FHEXploring protein fitness landscapes by directed evolution. *Nat. Rev. Mol. Cell Biol*2009, 10, 866–876. [PubMed: 19935669]
- (156). Hong NS; Petrovic D; Lee R; Gryn'ova G; Purg M; Saunders J; Bauer P; Carr PD; Lin CY; Mabbitt PDet al., The evolution of multiple active site configurations in a designed enzyme. *Nat. Comm*2018, 9, 3900.
- (157). Campbell E; Kaltenbach M; Correy GJ; Carr PD; Porebski BT; Living-stone EK; Afriat-Jurnou L; Buckle AM; Weik M; Hollfelder Fet al., The role of protein dynamics in the evolution of new enzyme function. *Nat. Chem. Biol*2016, 12, 944. [PubMed: 27618189]
- (158). Maria-Solano MA; Iglesias-Fernandez J; Osuna SDeciphering the Allosterically Driven Conformational Ensemble in Tryptophan Synthase Evolution. *J. Am. Chem. Soc*2019, 141, 13049–13056. [PubMed: 31356074]
- (159). Buller AR; Brinkmann-Chen S; Romney DK; Herger M; Murciano-Calles J; Arnold FHDirected evolution of the tryptophan synthase β -subunit for standalone function recapitulates allosteric activation. *Proc. Natl. Acad. Sci. USA*2015, 112, 14599–14604. [PubMed: 26553994]
- (160). Buller AR; van Roye P; Cahn JKB; Scheele RA; Herger M; Arnold FHDirected Evolution Mimics Allosteric Activation by Stepwise Tuning of the Conformational Ensemble. *J. Am. Chem. Soc*2018, 140, 7256–7266. [PubMed: 29712420]
- (161). Isaksen GV; Aqvist J; Brandsdal BOEnzyme surface rigidity tunes the temperature dependence of catalytic rates. *Proc. Natl. Acad. Sci. USA*2016, 113, 7822–7827. [PubMed: 27354533]
- (162). Bae E; Phillips GN Jr., Roles of static and dynamic domains in stability and catalysis of adenylate kinase. *Proc. Natl. Acad. Sci. USA*2006, 103, 2132–2137. [PubMed: 16452168]
- (163). Daily MD; Phillips GN; Cui QInterconversion of Functional Motions between Mesophilic and Thermophilic Adenylate Kinases. *PLoS Comput. Biol*2011, 7, e1002103. [PubMed: 21779157]
- (164). Nguyen V; Wilson C; Hemberger M; Stiller JB; Agafonov RV; Kutter S; English J; Theobald DL; Kern DEvolutionary drivers of thermoadaptation in enzyme catalysis. *Science*2017, 355, 289–294. [PubMed: 28008087]
- (165). Arcus VL; Mulholland AJTemperature, Dynamics, and Enzyme-Catalyzed Reaction Rates. *Annu. Rev. Biophys*2020, 49, 163–180. [PubMed: 32040931]
- (166). Arcus VL; Prentice EJ; Hobbs JK; Mulholland AJ; van der Kamp MW.; Pudney CR; Parker EJ; Schipper LAOn the Temperature Dependence of Enzyme-Catalyzed Rates. *Biochem.* 2016, 55, 1681–1688. [PubMed: 26881922]
- (167). Socan J; Purg M; Aqvist JComputer simulations explain the anomalous temperature optimum in a cold-adapted enzyme. *Nat. Comm*2020, 11, 2644.
- (168). Klinman JP; Kohen AHydrogen Tunneling Links Protein Dynamics to Enzyme Catalysis. *Annu. Rev. Biochem*2013, 82, 471–496. [PubMed: 23746260]
- (169). Benkovic SJ; Hammes-Schiffer SA perspective on enzyme catalysis. *Science*2003, 301, 1196–1202. [PubMed: 12947189]
- (170). Daniel RM; Danson MJA new understanding of how temperature affects the catalytic activity of enzymes. *Trends Biochem. Sci*2010, 35, 584–591. [PubMed: 20554446]
- (171). Aqvist J; Socan J; Purg MHidden Conformational States and Strange Temperature Optima in Enzyme Catalysis. *Biochem.* 2020, 59, 3844–3855. [PubMed: 32975950]
- (172). van der Kamp MW.; Prentice EJ; Kraakman KL; Connolly M; Mulholland AJ; Arcus VLDynamical origins of heat capacity changes in enzymecatalysed reactions. *Nat. Comm*2018, 9, 1177.

- (173). Bunzel HA; Anderson JLR; Hilvert D; Arcus VL; van der Kamp MW.; Mulholland AJ Evolution of dynamical networks enhances catalysis in a designer enzyme. *bioRxiv*2020, 10.1101/2020.08.21.260885.
- (174). Bunzel HA; Kries H; Marchetti L; Zeymer C; Mittl PRE; Mulholland AJ; Hilvert D Emergence of a Negative Activation Heat Capacity during Evolution of a Designed Enzyme. *J. Am. Chem. Soc*2019, 141, 11745–11748. [PubMed: 31282667]
- (175). Packer MS; Liu DR Methods for the directed evolution of proteins. *Nat. Rev. Genet*2015, 16, 379–394. [PubMed: 26055155]
- (176). Chen K; Arnold FH Engineering new catalytic activities in enzymes. *Nat. Cata*2020, 3, 203–213.
- (177). Ravikumar A; Arzumanyan GA; Obadi MKA; Javanpour AA; Liu CC Scalable, Continuous Evolution of Genes at Mutation Rates Above Genomic Error Thresholds. *Cell*2018, 175, 1946–1957. [PubMed: 30415839]
- (178). Rix G; Watkins-Dulaney EJ; Almhjell PJ; Boville CE; Arnold FH; Liu CC Scalable, continuous evolution for the generation of diverse enzyme variants encompassing promiscuous activities. *Nat. Comm*2020, 11, 5644.
- (179). Gagliardi L; Truhlar DG; Li Manni G.; Carlson RK; Hoyer CE; Bao JW LMulticonfiguration Pair-Density Functional Theory: A New Way To Treat Strongly Correlated Systems. *Acc. Chem. Res*2017, 50, 66–73. [PubMed: 28001359]
- (180). Chan GKL; Sharma S The Density Matrix Renormalization Group in Quantum Chemistry. *Annu. Rev. Phys. Chem*2011, 62, 465–481. [PubMed: 21219144]
- (181). Grisafi A; Ceriotti M Incorporating long-range physics in atomic-scale machine learning featured. *J. Chem. Phys*2019, 151, 204105. [PubMed: 31779318]
- (182). Cui Q Quantum Mechanical Methods in Biochemistry and Biophysics. *J. Chem. Phys*2016, 145, 140901. [PubMed: 27782516]
- (183). Raman S Systems Approaches to Understanding and Designing Allosteric Proteins. *Biochem.* 2018, 57, 376–382. [PubMed: 29235352]
- (184). Leander M; Yuan YC; Mager A; Cui Q; Raman S Functional Plasticity and Evolutionary Adaptation of Allosteric Regulation. *Proc. Natl. Acad. Sci. USA*2020, 117, 25445–25454. [PubMed: 32999067]
- (185). Swiderek K; Tuñón I; Moliner V; Bertran J Computational strategies for the design of new enzymatic functions. *Arch. Biochem. Biophys*2015, 582, 68–79. [PubMed: 25797438]
- (186). Sherrill CD; Manolopoulos DE; Martinez TJ; Michaelides A Electronic structure software. *J. Chem. Phys*2020, 153, 070401. [PubMed: 32828107]
- (187). Zen A; Brandenburg JG; Klimes J; Tkatchenko A; Alfe D; Michaelides A Fast and accurate quantum Monte Carlo for molecular crystals. *Proc. Natl. Acad. Sci. USA*2018, 115, 1724–1729. [PubMed: 29432177]
- (188). Hermann J; Schatzle Z; Noe F Deep-neural-network solution of the electronic Schrodinger equation. *Nat. Chem*2020, 12, 891. [PubMed: 32968231]
- (189). Google AI Quantum and Collaborators, Hartree-Fock on a superconducting qubit quantum computer. *Science*2020, 369, 1084–1089. [PubMed: 32855334]
- (190). McArdle S; Endo S; Aspuru-Guzik A; Benjamin SC; Yuan X Quantum computational chemistry. *Rev. Mod. Phys*2020, 92, 015003.

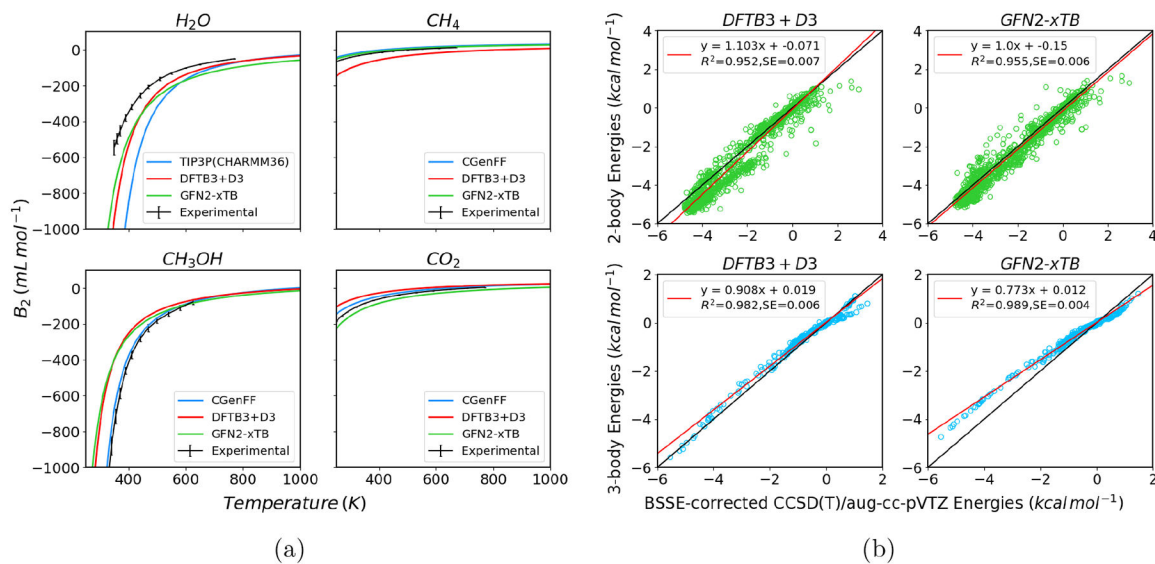


Figure 1: Non-covalent interaction with two density functional tight binding methods. (a) Second virial coefficients (B_2) for several small molecules, in comparison with CHARMM-CGenFF⁵⁶ and experimental results; (b) 2,3-body interactions of water in comparison with the BSSE corrected CCSD(T)/aug-cc-pVTZ database of Paesani and co-workers.⁵⁴ For additional results, see Supporting Information.

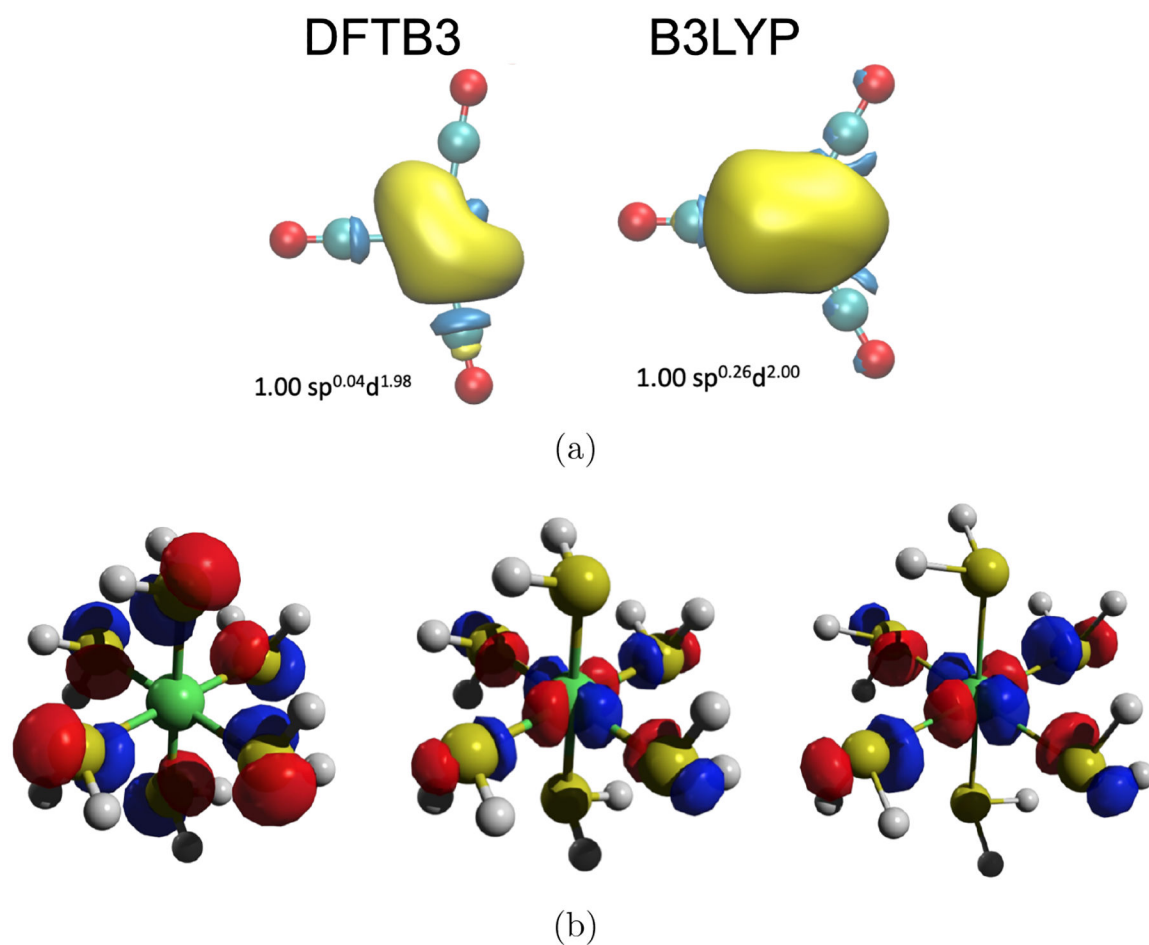


Figure 2:

Description of Ni compounds with DFTB3. (a) Comparison of the nickel lone valence hybrid in $[\text{Ni}(\text{CO})_3]^{2+}$ with DFTB3/3OB and B3LYP based Natural Bonding Orbital analyses;^{57,58} DFTB3 and B3LYP favor D_{3h} and C_{2v} symmetry, respectively. (b) Examples of frontier orbital comparisons between DFTB3, DFTB3+U and PBE calculations for high-spin $[\text{Ni}(\text{H}_2\text{S})_6]^{2+}$. Including the +U correction in DFTB3/3OB⁶² improves various properties such as d orbital populations, nature of frontier orbitals, ligand field splitting and energy difference between low/high-spin states.

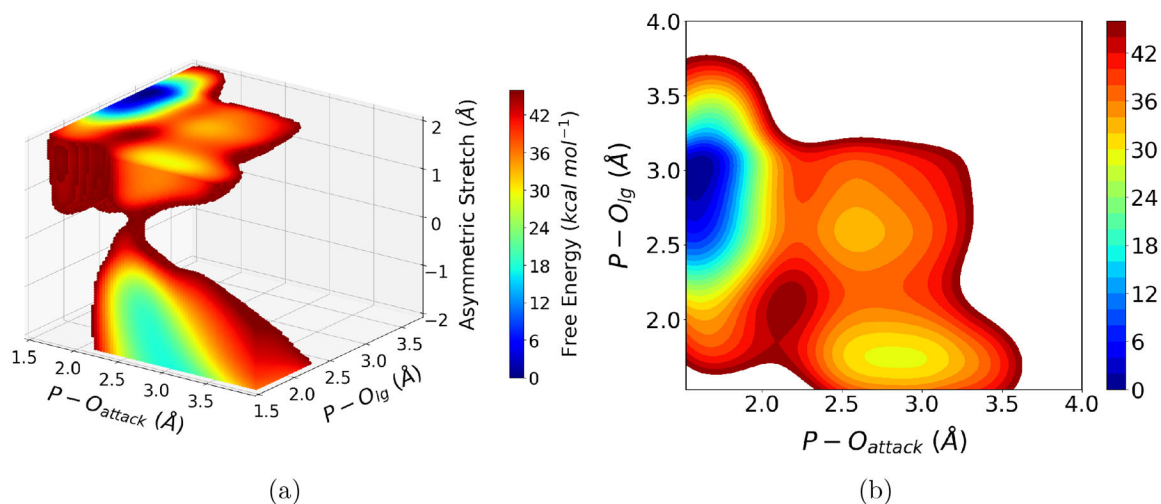
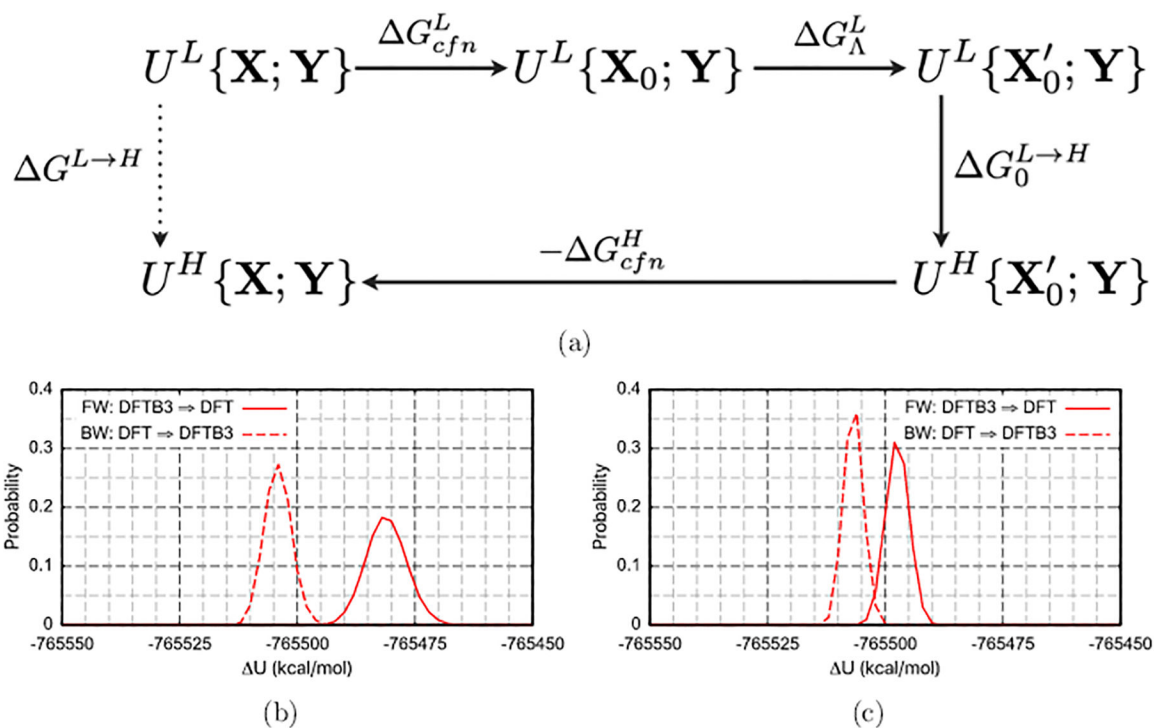


Figure 3: Results from an automated reinforcement learning driven free energy simulation of methyl phosphate hydrolysis in solution. The three coordinates are the nucleophilic attack P-O distance, the leaving group P-O distance and the antisymmetric O-H-O stretch that describes the proton transfer from the nucleophile (water) to the phosphate oxygen. (a) The three-dimensional PMF converges after 42 iterations of automated restrained MD-reinforcement learning cycles; the results indicate that with the current DFTB3/MM model, the solvent-assisted pathway¹²⁰ is not the dominant mechanism. (b) The two-dimensional PMF cut after the proton transfer is complete indicates a dissociative pathway that involves a loosely bound metaphosphate species. Further refinement of the QM/MM energetics will provide insights into this prototypical phosphoryl transfer reaction at an unprecedented level of detail.

**Figure 4:**

A staged transformation approach¹⁴¹ for computing the free energy difference at two levels (L/H) of theory, $G^{L \rightarrow H}$. (A) The staged thermodynamic path treats selected degrees of freedom (\mathbf{X}) separately from the rest (\mathbf{Y}); \mathbf{X} represents the degrees of freedom that lead to a large gap in the U^{LH} distribution. Assuming that the free energy costs for confining \mathbf{X} to values at (or near) the free energy minima are similar at the L and H levels, $G^{L \rightarrow H}$ is given by the sum of $\Delta G_0^{L \rightarrow H}$, which converges readily since the sampling involves only \mathbf{Y} , and the “reorganization free energy”, ΔG_{Λ}^L , which is the free energy cost of changing \mathbf{X}_0 to \mathbf{X}'_0 at the low level of theory. (b-c) Illustration of the impact of bond and angle restraints on the U^{LH} distribution for a methyl diphosphate, which is treated with either DFTB3 (L) or B3LYP (H), solvated by TIP3P water.

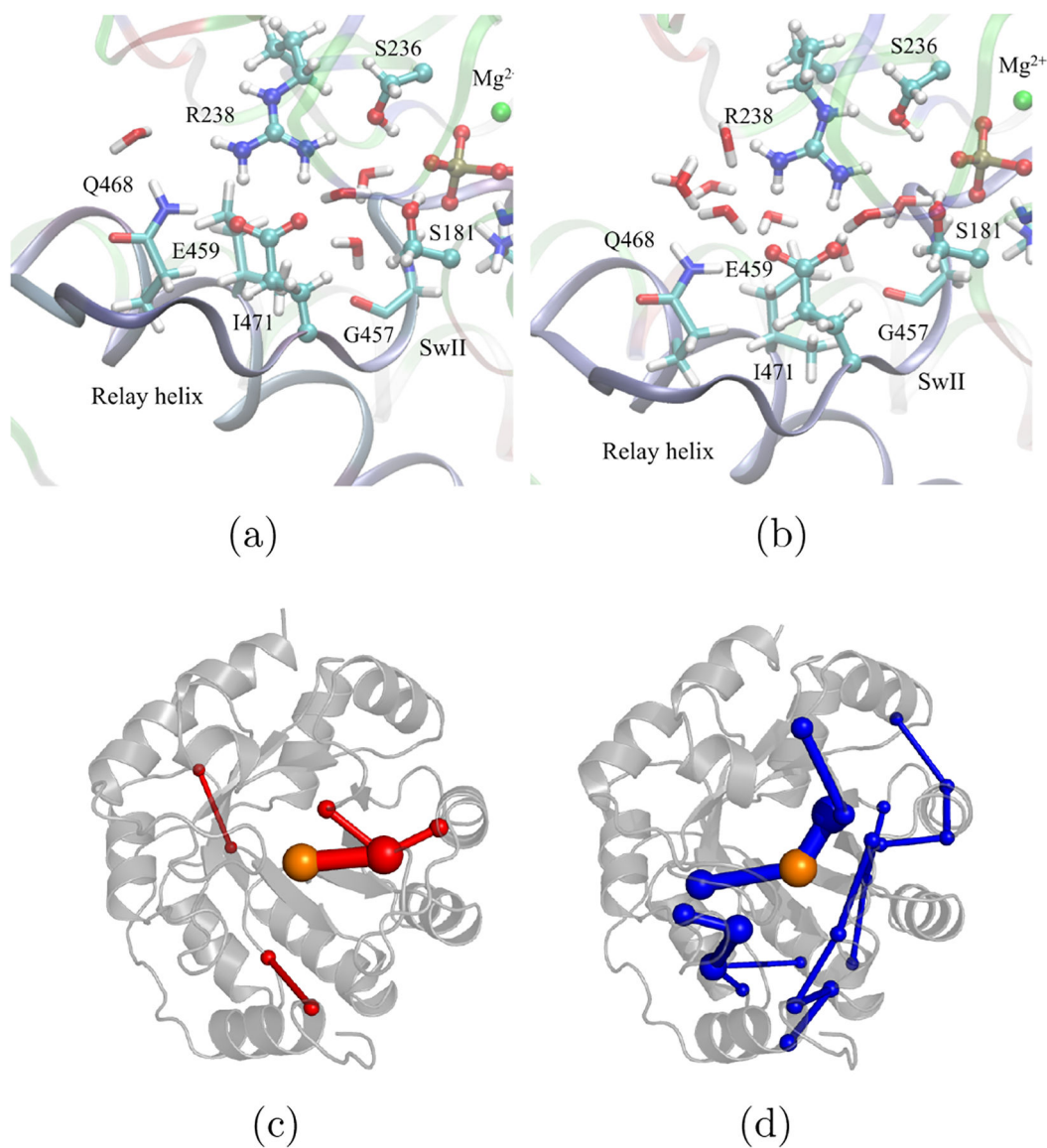


Figure 5: Examples of distal contributions to enzyme catalysis from computational analyses. (a-b) ATP hydrolysis in two models of the myosin motor domain may differ by ~ 9 kcal/mol in activation free energy although the nucleotide binding site residues have almost comparable average configurations;¹⁰⁶ (c-d) A de novo designed enzyme for Kemp elimination and its more efficient variant following directed evolution feature rather different collective motions;¹⁷³ the latter also features a negative activation heat capacity.¹⁷⁴ Panels c-d have been provided by Drs. A. Bunzel and A. Mulholland.