



Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.



## Review Article

# An overview of SARS-COV-2 epidemiology, mutant variants, vaccines, and management strategies



Tahmeena Farooqi<sup>a,1</sup>, Jonaid Ahmad Malik<sup>b,c,1</sup>, Almas Hanif Mulla<sup>d</sup>, Turki Al Hagbani<sup>e</sup>, Khaled Almansour<sup>e</sup>, Mohammed Abrar Ubaid<sup>f</sup>, Saleh Alghamdi<sup>g</sup>, Sirajudheen Anwar<sup>h,\*</sup>

<sup>a</sup> Department of Pharmacology and Toxicology, National Institute of Pharmaceutical Education and Research, Hyderabad, Telangana, India

<sup>b</sup> Department of Pharmacology and Toxicology, National Institute of Pharmaceutical Education and Research, Guwahati, India

<sup>c</sup> Department of Biomedical Engineering, Indian Institute of Technology (IIT), Ropar, Punjab, India

<sup>d</sup> Department of Pharmacology, Goa College of Pharmacy, Goa, India

<sup>e</sup> Department of Pharmaceutics, College of Pharmacy, University of Hail, Hail, Saudi Arabia

<sup>f</sup> Department of Pharmaceutics, JSS College of Pharmacy, Ooty, Tamil Nadu, India

<sup>g</sup> Department of Clinical Pharmacy, Faculty of Clinical Pharmacy, Albaha University, Albaha, Saudi Arabia

<sup>h</sup> Department of Pharmacology and Toxicology, College of Pharmacy, University of Hail, Hail, Saudi Arabia

## ARTICLE INFO

## Article history:

Received 24 May 2021

Received in revised form 7 August 2021

Accepted 10 August 2021

## Keywords:

SARS-COV-2

Epidemiology

Variant strains

Vaccine candidates

WHO landscape

## ABSTRACT

**Background:** Over the last two decades, humanity has observed the extraordinary anomaly caused by novel, weird coronavirus strains, such as severe acute respiratory syndrome (SARS) and Middle East respiratory syndrome (MERS). As the severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) virus has made its entry into the world, it has dramatically affected life in every domain by continuously producing new variants. The vaccine development is an ongoing process, although some vaccines got marketed. The big challenge is now whether the vaccine candidates can provide long-lasting protection or prevention against mutant variants.

**Methods:** The information was gathered from various journals, electronic searches via Internet-based information such as PubMed, Google Scholar, Science Direct, online electronic journals, WHO landscape, world meters, WHO website, and News.

**Results:** This review will present and discuss some coronavirus disease 19 (COVID-19) related aspects including: the pathophysiology, epidemiology, mutant variants vaccine candidates, vaccine efficacy, and

**Abbreviations:** ACE2, angiotensin-converting enzyme-2; ADRP, ADP ribose-1'-phosphatase; ARTES, Germany: based biotechnology company specialized in recombinant protein production and process development from microbial expression systems; BIOCAD, BIO computer aided design RNA; CanVirex AG, Swiss Biotech Association; CARDS, Covid 19 acute respiratory distress syndrome; CBC, complete blood count; CCHFV, Crimean-Congo hemorrhagic fever virus; CDC, center for disease control and prevention; CEA, carcinoembryonic antigen; CHIKV, chikungunya virus; CMV, cytomegalovirus; CNBG, China National Biotec Group; CNRS, centre national de la recherche scientifique; COPD, chronic obstructive pulmonary disease; COVID-19, corona virus disease 2019; CPAP, continuous positive airway pressure; CRP, C-reactive protein; DMVs, double-membrane vesicles; DWRAIR, Diseases/Walter Reed Army Institute of Research; DZIF, German Center for Infection Research; EBOV, Ebola virus; ECDC, European Centre for Disease Prevention and Control; ERGIC, endoplasmic reticulum- golgi intermediate compartment; ExoN, exoribonuclease; FiO<sub>2</sub>, fraction of inspired oxygen; GCIR, German Center for Infection Research; GMV, glycine mosaic virus; GLA, glucopyranosyl Lipid A; GPO, Government Pharmaceutical Organization; HeV, hepatitis virus E; HBV, hepatitis B virus; HFNC, high-flow nasal cannula; HLC, high lung compliance; IAVI, international AIDS vaccine initiative; IDIBAPS, Pi i Sunyer Biomedical Research Institute; IEM, Institute For Engineering in Medicine; InfA, influenza virus-A; INRAE, National Research Institute for Agriculture, Food and Environment; IPV, inactivated polio virus; IMV, Instruments de Médecine Vétérinaire; LASSA, lassa virus; LASV, lassa mammarenavirus; LinKinVax, French biotechnology startup that focuses on speeding up vaccine; LiteVax BV, spike-based (epitope screening); LLC, low lung compliance; LVVV, live viral vectored vaccine; MARV, Marburg virus; MDA5, melanoma differentiation associated protein; MERS-CoV, Middle East Respiratory Syndrome coronavirus; MIGAL, Galilee Research Institute Ltd; MMR, measles mumps rubella; MVA, modified vaccinia virus Ankara; NERVTAG, new and emerging respiratory virus Threats advisory group; NiV, Nippa virus; NIV, non-invasive ventilation; NLC, nanostructured Lipid Carriers; NORV, norovirus; NSCLC, non-small cell lung cancer; NSP, non-Structural proteins; O-MT, O-methyl transferase-2; OMV, outer membrane vesicle; ORFs, open reading frames; Osivax, clinical stage biotechnology company; P.C., preclinical; PaCO<sub>2</sub>, partial pressure of carbon dioxide; PaO<sub>2</sub>, partial pressure of oxygen; PEEP, positive end-expiratory pressure; PPI, proton pump inhibitors; RBD, receptor-binding domain; RVF, Rift Valley fever; RdRp, RNA dependent RNA polymerase; RTC, replication transcription complex; RTI, respiratory tract infections; SARS-CoV-2, severe acute respiratory syndrome coronavirus-2; SIG, SARS-COV-2 Interagency Group; SpO<sub>2</sub>, oxygen saturation; SRC VB VECTOR, State Research Centre of Virology and Biotechnology; TMPRSS2, transmembrane protein serine 2; TRSs, transcriptional regulatory sequences; USAMRIID/WARIAR, United States Army Medical Research Institute of Infectious; VEE, Venezuelan equine encephalitis; VLP, virus like particle; VOC, variant of concern; VOHC, variant of high consequences; VOI, variant of interest; VRI, Vaccine Research Institute; VSV, vesicular stomatitis virus.

\* Corresponding author.

E-mail address: [si.anwar@uoh.edu.sa](mailto:si.anwar@uoh.edu.sa) (S. Anwar).

<sup>1</sup> These authors have equal contribution.

<https://doi.org/10.1016/j.jiph.2021.08.014>

1876-0341/© 2021 The Authors. Published by Elsevier Ltd on behalf of King Saud Bin Abdulaziz University for Health Sciences. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

management strategies. Due to the high death rate, continuous spread, an inadequate workforce, lack of required therapeutics, and incomplete understanding of the viral strain, it becomes crucial to build the knowledge of its biological characteristics and make available the rapid diagnostic and vital therapeutic machinery for the combat and management of an infection.

**Conclusion:** The data summarizes current research on the COVID 19 infection and therapeutic interventions, which will direct future decision-making on the effort-worthy phases of the COVID 19 and the development of critical therapeutics. The only possible solution is the vaccine development targeting against all variant strains to halt its progress; the identified theoretical and practical knowledge can eliminate the gaps to improve a better understanding of the novel coronavirus structure and its design of a vaccine. In addition, to that the long-lasting protection is another challenging objective that need to be looked into.

© 2021 The Authors. Published by Elsevier Ltd on behalf of King Saud Bin Abdulaziz University for Health Sciences. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

## Contents

Introduction .....	1300
Pathophysiology .....	1301
Other NSPs protein functions are listed below in <a href="#">Table 1</a> .....	1301
Epidemiology .....	1301
Origin .....	1301
Topological dispersal .....	1301
Global response .....	1301
Italy .....	1301
France and Germany .....	1301
Japan .....	1301
United States .....	1301
Brazil .....	1302
Russia .....	1302
India .....	1303
Etiology in different groups .....	1303
Mutations and severity of infection .....	1303
VOHC's .....	1304
VOC's .....	1304
VOI's .....	1304
Variants under monitoring .....	1304
Management of COVID 19 .....	1306
Cough management .....	1310
Fever management .....	1310
GI disturbances .....	1310
Vaccines .....	1310
COVID 19 vaccine efficacy .....	1310
Conclusion .....	1310
Author's contribution .....	1311
Funding .....	1311
Competing interests .....	1311
Ethical approval .....	1311
Acknowledgement .....	1311
References .....	1311

## Introduction

Andrews and Gledhill in 1951 screened a hepatitis virus from mice which are now known as a single-stranded RNA coronavirus. Its diseases and cause have been recognized in animals and humans for over 50 years. 229E and OC43 were the first coronaviruses to cause very mild infections like common colds in humans. Later on, severe acute respiratory syndrome (SARS) and Middle East respiratory syndrome (MERS) emerged from civet cats, camels, bats [1]. The family of coronavirus is called as Coronaviridae which comprises of two subfamilies, Coronavirinae and Letovirinae [2,3]. Pneumonia's initial diagnosis and unknown cause made the cluster of patients in China admitted into hospitals in December 2019. The reports confirmed the potential coronavirus outbreak, of coronavirus disease 19 (COVID-19), and WHO gave it a name on February 11, 2020 [4]. In response to the outbreak, the center for disease control and prevention (CDC) conducted epidemiological and etio-

logical investigations via Wuhan city's Health authorities. Nearly 1 billion cases per year and economic losses of hundreds of billions of dollars occur due to this zoonotic illness, demonstrating the importance of developing vaccine design strategies for virus families with pools of causing extensive zoonotic diseases. The transcriptional regulatory sequences (TRSs) mediates discontinuous transcription and transcribes sub genomic RNAs, which make up the structure of a virus particle [5,6]. Advances in understanding viral machinery, the role of various viral proteins, their genetic structure, host immune responses, and the ability to confer protection switch its way to developing a vaccine successfully. An additional challenge is conducting clinical trials in this pandemic loss by utilizing the current evidence and critical knowledge gaps to better understand the virus strategy to safeguard public health. This review, has focused and emphasized points on the COVID 19 pathophysiology, epidemi-

ology, mutant variants, vaccine candidates, vaccine efficacy and strategies for disease management.

### Pathophysiology

Coronavirus spike glycoprotein (S) with the expression of their subunits S1 and S2 binds to the host cell receptor's surface. S1 determines the cellular tropism and host range and helps attach the virus to the target cell. In contrast, the S2 subunit helps infusion with the host cells (mainly alveolar epithelial cells and small intestine enterocytes) with the aid of Angiotensin-Converting Enzyme-2 (ACE2) claw-like structure of the receptor by endocytosis [7,8]. When this S protein binds to the ACE-2 receptor, the transmembrane protein serine 2 (TMPRSS2), which is associated with the cell surface, mediates cleavage of S protein trimer, and the fusion mechanism is activated via endosomal acid proteases (cathepsin L) [9–11]. ACE2 is an 805 amino acid containing integral membrane protein consists of 3 domains that are a domain at the transmembrane side, a domain at the extracellular side and a domain at the cytoplasmic side, and the N-terminal peptide. The enzyme sheddase helps release soluble protein into the bloodstream by cleavage of the active carboxypeptidase domain from the transmembrane domain [12]. When there is a lysosome-mediated drop in pH, the endosome membrane fuses with the virus's envelope and releases a nucleocapsid in the cytoplasm. The cellular proteases degrade the capsid, and the positive sense RNA viral genome ranges between 27–32 kbp is left free. This viral genomic RNA consists of at least 6 open reading frames (ORFs). ORF1a and ORF1b are present at the 5' end, constitute a common fraction of the whole genome length RNAs, and are translated into pp1a and pp1ab proteins. Protease helps in cleavage of those proteins in to 16 non-structural proteins (NSP1–NSP16) [13–15]. Some of these NSPs, like NSP3, encode for papain-like proteases (PLP) and NSP5 codes for 3CL-proteases. Both of these proteins form a polypeptide “Replication transcription complex” (RTC) and helps in innate immune response blockage via genomic transcription and translation process. NSP15 encodes for RNA helicase, whereas NSP12 for RNA dependent RNA polymerase (RdRp) and synthesis of subgenomic RNAs (sgRNAs) stand genomic RNAs occurs [16] (Fig. 1).

Other NSPs protein functions are listed below in Table 1

Four Structural proteins are coded by ORFs located on 3' end:

- 1) Membrane (M) shape the virions [17].
- 2) Spike (S) recognizes the ACE2 receptor on the host cell surface [18].
- 3) Envelope (E) helps in assembly and release of virions [19].
- 4) Nucleocapsid (N) packages the genome in the virions, provides pathogenicity.

There are many other structural and accessory proteins that are specific to the different species. Membrane exocytosis helps budding of completed assembled SARS-COV-2 particles via endoplasmic reticulum in the endoplasmic reticulum-golgi intermediate compartment (ERGIC). It is currently known that interaction of mainly M protein with different structural proteins of virus E, S aids in the generation of the virion scaffold promoting assembly, budding, and release of mature virus particle by exocytosis. After the final phase of maturation, all the components of the virus fit together, the particle is infectious and ready to begin a new cycle [20–23].

### Epidemiology

Coronaviruses which mainly infect birds and mammals, comprise a large family (Coronaviridae) of giant enveloped

positive-strand RNA viruses, which can be a cause of upper and lower respiratory tract infections (RTI) manifest as pneumonia, bronchitis, MERS, SARS, COVID 19. These three new coronaviruses caused respiratory disease outbreaks with their unique features, but SARS and MERS are less infectious and have significantly higher fatality rates than SARS-COV-2. The following Table 2 demonstrates the fundamental difference between the three of them [57–59].

#### Origin

Disease spread globally over eight million after the emergence of many pneumonia-like cases in Wuhan, Hubei province China (Early December 2019).

#### Topological dispersal

On December 31, 2019, the Chinese government first confess the numerous cases of pneumonia of an unknown cause presumed to be a zoonotic illness that would later be called COVID 19. The first case of death was reported on January 11. The Wuhan city was locked down with over 11 million populations on January 23. A week later, thousands of cases were reported, and WHO immediately declared a global health emergency [60]. With the start of July 2020, there were approximately 8.6 million cases and 450,000 deaths due to COVID 19 [61].

### Global response

#### Italy

Italy was the following unfortunate country after China experienced a significant outbreak and resulted in the highest death rate, 7.2%. Milan, the most dynamic city alone, represents 10% of the Italian economy, “The country's economic engine” was drastically slowed after Italy experienced a surge in coronavirus cases. Italy went from discovering the first official COVID 19 case to the prohibition of all movements and non-essential business activities in a matter of weeks (February 21- March 22). Around 3,949,517 cases and 119,021 deaths took place until 25th of April 2021 [61].

#### France and Germany

The “quick and dirty Sunday morning” analysis confirmed the trend of viral proliferation in France and Germany exceeds more than in Italy, South Korea, and Japan with a similar temperature. The study also suggests that intense containment measures as in Italy, South Korea, and Japan may help to slowdown proliferation [62]. France is 4th country affected and around 5,473,579 cases and 102,713 deaths took place until 25th of April 2021. Germany 10th leading country in COVID 19 cases and around 3,277,661 cases and 82,117 deaths took place until 25th of April 2021 [61].

#### Japan

Japan, though with a more significant percentage of the population, experienced a low mortality rate compared to Italy mainly because of their cluster-based testing approach and adoption of the “3C” method (avoidance of closed spaces, crowded spaces, close contact) (Bloomberg, 2020). Around 556,999 cases and 9854 deaths were reported in Japan until 25th of April 2021 [61].

#### United States

U.S reached the highest count of reported cases worldwide from the first known patient in late January to August 28th where there are 6 million cases and nearly 2 lakh deaths. About 32,766,119 cases

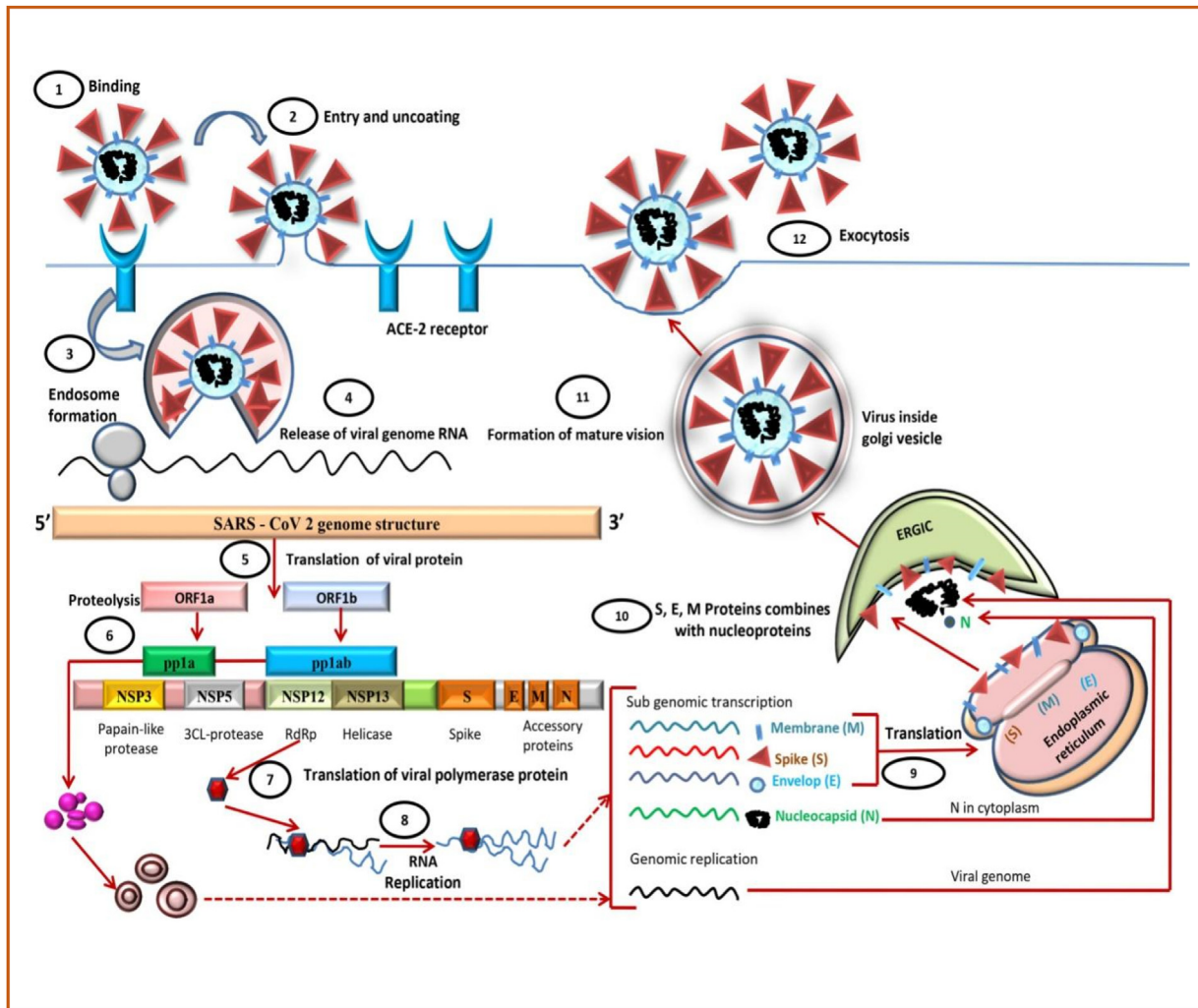


Fig. 1. The life cycle of SARS-CoV-2 in the host cells.

Table 1  
Functions of non-structural proteins (NSP1-16).

Nonstructural Proteins	Function	Reference
NSP1	Innate immune response blockage by promotion of mRNA degradation	[24–27]
NSP2	Unknown function, binds to prohibiting proteins	[28,29]
NSP3	Papain lyase, cytokine expression promotion, blocking host innate immune response, activity of ADRP	[30–35]
NSP4	Potential transmembrane scaffold protein, maintains DMVs proper structure	[36,37]
NSP5	Main protease (Mpro), viral polyprotein cleavage	[38]
NSP6	Potential transmembrane scaffold protein	[39]
NSP7	NSP7 forms a hexadecameric complex with NSP8. For RNA polymerase, it acts as a processivity clamp	[40]
NSP8	For RNA polymerase, it acts as a processivity clamp and also plays a role as a primase	[41,42]
NSP9	Binding protein for RNA	[43]
NSP10	It acts as a cofactor for NSP14 and 16 by forming a heterodimer with both of them and stimulating the activity of N viral exoribonuclease (ExoN) and O-methyl transferase	[44,45]
NSP12	RdRp)	[46]
NSP13	RNA Helicase, 5' Triphosphatase	[47,48]
NSP14	Shows ExoN activity which is important for viral genome proofreading, addition of 5' cap to viral RNAs by MTase.	[49–52]
NSP15	Endoribonuclease of virus	[53,54]
NSP16	O-methyl transferase (2'-O-MT); viral RNA shielding from MDA5 recognition	[55,56]

have been reported and 585,449 deaths had taken place on 25th of April 2021 and US is at the forefront [61].

**Brazil**

The second highest number of cases is found in Brazil, especially the Amazon state country's northwest, which has the highest mortality rate. Manaus, the state capital, and the bustling city make the

potential hotspots for transmitting the virus (CDC, 2020). Brazil is third affected country and around 14,238,110 cases and 386,623 deaths took place until 25th of April [61].

**Russia**

Russia was centered in Moscow's city, accounted for the highest number of cases measuring 257.7 thousand, followed by Moscow



**Table 2**  
Differences of SARS-CoV, MERS CoV, SARS-CoV-2.

	SARS-CoV	MERS-CoV	SARS-CoV-2
First notified	November 2002 in China's Guangdong province	September 2012, in Saudi Arabia	December 2019 in Wuhan, China
Mode of transmission	Infected civets, droplets produced by sneezing, coughing, breathing, talking	Droplets from person to person (unclear from camels to humans)	Droplets by coughing, sneezing, talking
Mean incubation period	4–5 days	6–7 days	1–14 days
Key symptoms	Dry cough at first, fever, malaise, body aches and pains, diarrhea (in the first or second week)	Fever, cough, shortness of breath, nausea/vomiting, diarrhea	Fever, dry cough, shortness of breath, fatigue
Treatment	No specific treatment	No specific treatment	No specific treatment
Mortality rate	11%	34%	3–4%
Vaccine	No vaccine	No vaccine	No vaccine

**Table 3**  
Variants of concern (VOC).

Additional mutations and lineage	First detected	Substitutions on spike protein	Impact on immunity evidence
B.1.1.7 (20I/501Y.V1)	United Kingdom (Sept 2020)	Δ69/70, Δ144, (E484K*), (S494P*), N501Y, A570D, D614G, P681H, T716I, S982A, D1118H (K1191N*)	Unclear [2]
B.1.1.7 + E484K	United Kingdom (Dec 2020)	E484K, N501Y, D614G	Neutralisation (v) [2,5]
B.1.351 (20H/501.V2)	South Africa (Sep 2020)	D80A, D215G, Δ241/242/243, K417N, E484K, N501Y, D614G, A701V	Escape (v) [7,8]
B.1.427 (20C/S:452R)	U.S.A (California)	L452R, D614G	Neutralisation (v)
B.1.429 (20C/S:452R)	U.S.A (California)	S13I, W152C, L452R, D614G	Neutralisation (v)
P.1 (20J/501Y.V3)	Japan/Brazil	L18F, T20N, P26S, D138Y, R190S, K417T, E484K, N501Y, D614G, H655Y, T1027I	Neutralisation (v) [11]

\* = Aminoacid substitution on the variants.

Oblast with 67 thousand cases by August 24, 2020. It has registered ten folds of low mortality than Spain, Britain, Italy, and France despite having many cases. The country's well-funded healthcare system is to be appreciated for managing better than those in the U.S and Western European countries. Mass testing (has nearly 6 million tests carried out so far) helps people identify and isolate more people affected by the virus. They quickly convert hospitals and clinics to virus treatment centers. Russia is also at top leading in COVID 19 cases. Around 4,753,789 cases and 107,900 deaths took place until 25th of April 2021 [61].

### India

India, among all other countries, has the lowest fatality rate (2.41%) as of July 23 in the World even though it stands third after the United States and Brazil for having a large number of cases in Asia. The first case of COVID 19 was reported in Kerala on January 30. By mid of May 2020, 6 major cities like Delhi, Mumbai, Kolkata, Chennai, Pune, and Ahmedabad accounted for around 50% of all cases reported in the country [65]. In India, there is progressive rise of COVID 19 and about 16,951,621 cases and 192,309 deaths are reported till 25th of April 2021 [61].

### Etiology in different groups

The virus is transmitted via major routes such as droplets, contact, and aerosols. It has also been detected in the faecal samples of patients in the United States and China. Major risk factors include people more than 60 years of age, and even people underlying non-communicable diseases (NCDs): diabetes, hypertension, chronic lung disease, cerebrovascular disease, cardiac disease, chronic kidney failure, immune-suppression and cancer patient. Due to immunosuppressive state and many other physiological adaptive changes pregnant women are more susceptible to RTI but currently evidence of SARS-CoV-2 transmission through the placenta to the new-born has not been observed. In one of the studies, the samples of amniotic fluid, blood obtained from the umbilical cord, throat

swab of neonates, and maternal milk were collected from newborn babies whose mothers were SARS-CoV-2 positive. Still, none of the neonates were found to be positive [66].

### Mutations and severity of infection

Before SARS-CoV-2 other viruses also mutate themselves. There are different mutant variants of SARS-CoV-2 that have reported and increased the severity of infections. In UK, new and emerging respiratory virus threats advisory group (NERVTAG) published a paper with the result outcome from many preliminary analyses of B.1.1.7 [67]. One of the variants was detected in England and was highly transmissible and got dispersed to several other countries. This variant contains seventeen mutations in the genome in which 8 are on S protein which is main antigenic target of 3 SARS-CoV-2 vaccines that have been licensed in England [67]. The NERVTAG proposed that infections by B.1.1.7 have high chances of death, as compared to parent virus. The other highly infectious variant P.1 in Brazil was reported in the mid-2020. This variant has increased the rate of infections, severity of the disease and the Manaus city, in Brazil where the health department is totally in a collapsed position. Today Brazil is third leading country in the SARS-CoV-2 infection globally because of these variants [67].

The B.1.351 variant was first identified in South Africa in 2020. The vaccines manufactured by Moderna claimed on Jan 25, 2021 that our vaccine is effective against both B.1.1.7 and B.1.351 variants. However, the claim was based on an *in vitro* study. In addition to that, a South African variant was having decreased levels of neutralizing antibody titers. The company Pfizer has claimed that our vaccine will work against B.1.1.7 variant because of their investigations in laboratory, although these studies have not been peer reviewed [67]. As long as the variants emerged, the more havoc it will create, vaccines will not work and the severity of the disease will be more. In India, more than three hundred thousand cases hit each day as on 25th of April 2021 because of the double and triple mutant variants [61]. There should be big focus on the drug development against SARS-CoV-2 rather than whole focus

**Table 4**  
Variants of interest (VOI) [2].

Additional mutations and lineage	First detected	Substitutions on spike protein	Impact on immunity evidence
B.1.525	Nigeria (Dec 2020)	E484K, D614G, Q677H	Neutralisation
B.1.427/B.1.429	United States (Sep 2020)	L452R, D614G	Neutralisation
P.3	The Philippines (Jan 2021)	E484K, N501Y, D614G	Neutralisation
B.1.616	France (Feb 2021)	V483A, D614G, H655Y, G669S	
B.1.617 (20A)	India (Feb 2020)	L452R, E484Q, D614G	
B.1.617.1 (20A/S:154K)	India (Dec 2020)	(T95I), G142D, E154K, L452R, E484Q, D614G, P681R, Q1071H	Neutralisation
B.1.617.2 (20A/S:478K)	India (Dec 2020)	T19R, (G142D), Δ156, Δ157, R158G, L452R, T478K, D614G, P681R, D950N	Neutralisation
B.1.617.3 (20A)	India (Oct 2020)/(Feb 2021)	T19R, G142D, L452R, E484Q, D614G, P681R, D950N	Neutralisation
B.1.620	Place not clear (Feb 2021)	S477N, E484K, D614G	Neutralisation
B.1.621	Colombia (Jan 2020)	R346K, E484K, N501Y, D614G	Neutralisation

**Table 5**  
Variants of monitoring.

Additional mutations and lineage	First detected	Substitutions on spike protein	Impact on immunity evidence
B.1.214.2	Place not clear (Dec 2020)	Q414K, N450K, ins214TDR, D614G	
A.23.1 + E484K	United Kingdom (Dec 2020)	E484K, Q613H	Neutralisation
A.27	Place not clear (Dec 2020)	L452R, N501Y, H655Y	Neutralisation
A.28	Place not clear (Dec 2020)	E484K, N501T, H655Y	Neutralisation
C.16	Place not clear (Oct 2020)	L452R, D614G	Neutralisation
C.37	Peru (Dec 2020)	L452Q, F490S, D614G	
B.1.351 + P384L	South Africa (Dec 2020)	P384L, K417N, E484K, N501Y, D614G	Escape
B.1.351 + E516Q	Place not clear (Jan 2021)	K417N, E484K, N501Y, E516Q, D614G	Escape
B.1.1.7 + L452R	United Kingdom (Jan 2021)	L452R, N501Y, D614G	Neutralisation
B.1.1.7 + S494P	United Kingdom (Jan 2021)	S494P, N501Y, D614G	Neutralisation
C.36 + L452R	Egypt (Dec 2020)	L452R, D614G	Neutralisation
AT.1	Russia (Jan 2021)	E484K, D614G	Neutralisation
B.1.526 (20C/S:484K)	United States New York (Nov/Dec 2020)	(L5F*), T95I, D253G, (S477N*), (E484K*), D614G, (A701V*)	Neutralisation
B.1.526.1 (20C)	United States New York (Oct/Nov 2020)	D80G, Δ144, F157S, L452R, D614G, (T791I*), (T859N*), D950H	Neutralisation
B.1.526.2	United States (Dec 2020)	S477N, D614G	
B.1.1.318	Place not clear (Jan 2021)	E484K, D614G	Neutralisation
P.2	Brazil (Jan 2021)	E484K, D614G	Neutralisation

\* Detected in few sequences but not all.

on the vaccines. Such as in cases of other viral diseases which have been controlled by drugs not by vaccines like HIV. Vaccines may be effective against a single variant but drugs might work against many variants as is evidenced by many antiviral drugs. There are three CDC established classifications for multiple variants of the virus in collaboration with SIG. They are: variant of concern (VOC), variant of interest (VOI), and variant of high consequences (VOHC).

**VOHC's**

These variants have clear evidence that medical counter measures (MCMs) or measures for prevention reduced the effectiveness significantly compared to the previous variants. Fortunately, there are no SARS-CoV-2 variants observed so far at this high level of consequences [68].

**VOC's**

There is a clear evidence of transmissibility, severity, and immunity which require efforts to control the virus spread, CDC reporting, health actions of public, test and research to evaluate the vaccine effectiveness [68]. List of variants are given in Table 3.

**VOI's**

These variants have genetic markers which are specific which changes the binding of receptor, treatment efficacy reduction, has a clear evidence of transmissibility, severity, neutralization reduction by previously generated infection or vaccination. These variants require health actions of public, increased laboratory

**Table 6**  
Effect and management of corona in different phenotypes.

L-Phenotype	H-Phenotype
Low elastance (HLC)	High elastance (LLC)
Low ventilation perfusion ratio	High shunt from right to left
The low weight of lung	The high weight of lung
Low recruitability	High recruitability
PaO <sub>2</sub> /FiO <sub>2</sub> = 95 mmHg	PaO <sub>2</sub> /FiO <sub>2</sub> = 84 mmHg
Non-intubated patients	Severe ARDS treated patients
-HFNC, CPAP, NIV	-PEEP is higher,
Intubated patients:	-Prone position lying
-Volumes greater than 6 mg/kg (up to 8–9 mg/kg) can be ventilated for hypercapnic patients	-Extracorporeal support
-PEEP reduced 8–10 cm H <sub>2</sub> O	
-Prone positioning only as a rescue maneuver	

characterization, assessing how quickly the virus spreads by epidemiological investigations. B.1.617 and B.1.617.2 are classified as VOC by WHO and UK on 7th May 2021 respectively [2]. List of variants of interest are given in Table 4.

**Variants under monitoring**

Through genomic variant rules-based screening, preliminary scientific evidence, epidemic intelligence these variants are detected as signals. There is a weak evidence which is not been assessed by European centre for disease prevention and control (ECDC) that they show similar properties as of VOC (Table 5).

Biomarkers helpful in assessing the clinical trend are HFNC (high-flow nasal cannula), CPAP (continuous positive airway pres-

**Table 7**  
Overview of efficacy of various marketed COVID 19 vaccine in human subjects.

	Dose regimen	Formulation	Effective against (Phase III trails)	Post implementation effectiveness	Response in humans for antibodies	Response in humans for T cells	References
mRNA Pfizer/BioNtech (BNT162b2)	30 g Mrna, 2 doses, 21 days apart	To lock protein in the pre – fusion state “S” subunit is modified by two mutations of proline by forming lipid nanoparticle	After 2 doses – 95% After 1 dose – 52% Data review suggestion 14 days after 1st dose – 93% 6 months post 2nd dose – 91%	Symptomatic infection - 1 dose – 94–96% 2 doses – 46–80% Asymptomatic infection : 1 dose – 79% 2 doses 90% Hospitalization: 1 dose – 71–85% 2 doses - 87% Any infection: 1 dose – 46–72% 2 doses 86–92%	After 2nd dose S1-binding antibody increases. Nab was present in significant amount after 2nd dose	After 2nd dose increase in antigen-specific IFN $\gamma$ <sup>+</sup> CD4 <sup>+</sup> and CD8 <sup>+</sup> T cells, more IFN $\gamma$ and IL-2 secretion than IL- 4, T <sub>H</sub> 1 cell polarization	[80]
Moderna (Mrna – 1273)	100 g Mrna, 2 doses, 28 days apart	To lock protein in the pre – fusion state “S” subunit is modified by two mutations of proline by forming lipid nanoparticle	After 1 dose – 95% After 2 doses – 92%	Symptomatic infection - 1 dose – 80% 2 doses – 90%	After 14 days S - binding antibody detected and its levels increases slightly by 28 days and marked increase after 2nd dose. Nab levels are minimum after 1st dose reaches at peak after 2nd dose 14 days	After 1st dose small increases in TNF and IL-2-secreting cells. After 2nd dose Significant increases in CD4 <sup>+</sup> T cells secreting T <sub>H</sub> 1 type cytokines (TNF > IL-2 > IFN $\gamma$ ). Minimum change in T <sub>H</sub> 2 cell, CD8 <sup>+</sup> responses	
Viral vector Oxford University/Astra-Zeneca (ChAdOx1 nCoV-19)	2.5–5 × 10 <sup>10</sup> viral particles, 2 doses, ≥28 days apart	Simian adenovirus vector recombinant – replication – deficient full – length S protein with a Tpa leader sequence	After 1 dose – 76% After 2 doses – 62–67% Low dose followed by high dose – 90% ≥12-week interval, (81%), <6-week interval (55%)	Hospitalization : After 1 <sup>st</sup> dose 80–94%	After 14 days S - binding antibody detected and its levels increases slightly by 28 days and marked increase after 2nd dose 14 days. More IgG3 and IgG1. Nab detected after 1st dose and increases after 14 days of 2nd dose. After 28–56 days of single dose and peak IgM and IgA responses at day 14 or 28	After 1 <sup>st</sup> dose 14 days Peak T cell responses which is higher after 28 days 2nd dose. At 14 day there is increase in TNF and IFN $\gamma$ production by CD4 <sup>+</sup> T cells	
Gamaleya Research Institute (Gam-COVID-Vac)	10 <sup>11</sup> viral particles, 2 doses, 21 days apart	Dose 1 human adenovirus 26 replication-deficient, recombinant	After dose 1 74% After dose 2 91%	–	After 14 days of 1st dose - Nab (61)% S – binding antibody (85–89%) are detected. After 14 days of 2nd dose- binding antibody (98%) and neutralizing antibody (95%)	After 1st dose 14 days CD4 <sup>+</sup> and CD8 <sup>+</sup> T cells are observed. After 2nd dose 7 days S-specific IFN $\gamma$ responses are observed	
Janssen (Ad26.COV2.S)	5 × 10 <sup>10</sup> viral particles, 1 dose	S protein two amino acid changes at S1/S2 junction that remove cleavage of furin and 2 proline substitutions for replication-deficient recombinant Human adenovirus 26	After 1st dose 67%	–	After 28 days of vaccination S-binding and neutralizing antibody are present and their levels remain after 84 days of post vaccination.	At 14 and 28 days of post vaccination CD4 <sup>+</sup> and CD8 <sup>+</sup> T cells are present IFN $\gamma$ and/or IL-2 secretion suggesting T <sub>H</sub> 1 cell	
CanSino Biologics (Ad5-nCoV)	1 dose 5 × 10 <sup>10</sup> viral particles	Simian adenovirus vector recombinant replication – deficient full – length S protein with a Tpa leader sequence	After 1st dose 66% Decreases to 50% at 5–6 months	After 1st dose Hospitalization (80–94%)	RBD binding antibodies are observed after 14 days of vaccination (44%). Anti RBD binding antibodies after 28 days of vaccination, Nab's (47–50%)	After 28 days 78–88% had T cell response based on IFN $\gamma$ ELISpot After 14 day peak T cell responses were observed	



Table 7 (Continued)

	Dose regimen	Formulation	Effective against (Phase III trails)	Post implementation effectiveness	Response in humans for antibodies	Response in humans for T cells	References
Protein subunit Novavax (NVX – CoV2373)	(5 µg protein, 2 doses, 21 days apart	Full-length S protein with mutations at S1/S2 cleavage	After 2nd dose 7 days 90%		After 1st dose 21 days S-binding antibody and Nab is detected After 2nd dose- 7 days significant increase is observed	After 2nd dose 7 days CD4 <sup>+</sup> T cell responses are observed. Based on IL-2 and TNF production T <sub>H</sub> 1 cell phenotype; minimal T <sub>H</sub> 2 cell responses are measured	
Whole - cell inactivated virus Bharatth Biotech (BBV152)	6 µg protein, 2 doses, 28 days apart	Grow SARS – CoV-2 in Vero cells adsorbed on to aluminium hydroxide and imidazoquinoline molecule after inactivation with β-propiolactone	After 2 doses 78%		After 1st dose anti – S binding antibodies are observed (65%), NABs (48%) After 2nd dose 14 day (98%) and 97% respectively	Strong bias towards T <sub>H</sub> 1 cell phenotype (IFNγ and TNF), with minimal T <sub>H</sub> 2 cell responses After 2nd dose 76 day CD4 <sup>+</sup> CD45RO <sup>+</sup> memory T cells increases	

sure), NIV (noninvasive ventilation), PEEP (positive end-expiratory pressure), FiO2 (fraction of inspired oxygen), PaO2 (partial pressure of arterial blood), ARDS (acute respiratory distress syndrome), HLC (High Lung Compliance), LLC (Low lung Compliance) [68].

**Management of COVID 19**

Breathing movement: oxygen support: 5–15% of patients with COVID 19 require intensive care and ventilatory support as it pri-

**Table 8**  
Draft of COVID 19 vaccine candidates- WHO landscape 2020.

Description of vaccine	Type of vaccine candidate	The target for coronavirus	Non-corona virus candidate's same platform	Developers
A vaccine based on DNA	Vaccine inserts compatible with multiple delivery systems are engineered DNA Vaccine Plasmid DNA vaccine N and RBD  DNA with electroporation DNA with electroporation DNA Needle-free delivery for DNA with plasmid  S, S1, S2, RBD and N, DNA plasmid vaccine DNA vaccine ms DNA vaccine  DNA Vaccine S-gene containing DNA plasmids Plasmid vaccine with DNA Nanostructured RBD with plasmid DNA	SARS-CoV-2 and Sarbeco-CoV SARS-CoV2	SARS	University of Cambridge + DIOSynVax Ltd University of Ege Nottingham/Nottingham Trent University/Scancell Cobra Biologics/Karolinska Institute Vaccine Research Center Chula Evvivax/Applied DNA Sciences/Takis Pharmajet/Immunomic Therapeutics, Inc./EpiVax, Inc. Egypt, National Research Centre BioNet Asia Waterloo University/MediphageBioceuticals Santos Pharmaceuticals Biosun Pharmed Bangladesh, Globe Biotech Limited Slovenia, National Institute of Chemistry Norway, Vaccibody, Oslo Research Park Inserm PATH/Dynavax/Institute of Vaccines and Medical Biologicals (IVAC; Vietnam)
Inactivated virus (IV)	Vaccine encoding RBD with plasmid DNA Immunostimulatory DNA sequences CpG 1018 + Inactivated, egg-based, Membrane expressing whole Chimeric Newcastle Disease Virus (NDV) - SARS - CoV- 2, S protein (Lexapro) anchored Pre-fusion-stabilized trimeric CpG 1018 + Inactivated, egg-based, Membrane expressing whole Chimeric Newcastle Disease Virus (NDV) - SARS - CoV- 2, S protein (Lexapro) anchored Pre-fusion-stabilized trimeric CpG 1018 + Inactivated, egg-based, Membrane expressing whole Chimeric Newcastle Disease Virus (NDV) - SARS - CoV- 2, S protein (Lexapro) anchored Pre-fusion-stabilized trimeric Alum + Inactivated Inactivated  CpG 1018 + Inactivated CpG 1018 + Inactivated		J.E., ZIKA	PATH/Dynavax/GPO; Thailand)  PATH/Dynavax/Institute Butantan (Brazil)

Table 8 (Continued)

Description of vaccine	Type of vaccine candidate	The target for coronavirus	Non-corona virus candidate's same platform	Developers
Live attenuated virus (LAV)	The whole virus Inactivated	Pan-Corona SARS-CoV2	MMR, IPV MMR, IPV	Egypt, National Research Centre Kocak Farma Ilac ve Kimya San. A. S Shifa Pharmed
	Inactivated			Milad Pharmaceuticals Co.
	Alum + Inactivated			Zista Kian Azuma Co.
	Inactivated			AcibademLabmed Health Services
	Inactivated			A.S./Mehmet Ali Aydinlar University
	Live attenuated vaccines, which are codon deoptimized			University of Griffith/Indian
	Live attenuated vaccines, which are codon deoptimized			Immunologicals Ltd
	Live attenuated vaccine bacterial (pertussis)			Institut Pasteur Lille
	Live attenuated bacterial vector			TheRex, AltraBio
	Viral vector (non-replicating)			Sendai virus vector
Live attenuated bacterial vector (LABV)	Adenovirus-based	Pan-Corona SARS-CoV2	MARV, HIV, EBOV, LASV Multiple candidates	University of Ankara
	(AAV SARS-COV-2) Adeno-associated virus vector			AveXis/Massachusetts General Hospital/Massachusetts Eye and Ear
	VLP encoded by MVA			BravoVax/GeoVax
	MVA-S encoded			IDT Biologika GmbH/GCIR-DZIF
	MVA-S			Spain, IDIBAPS-Hospital Clinic
	Adeno5-based			University of Erciyes
	(GREVAX™ platform) Ad5 S			Greffex
	Oral Ad5 S			Stabilitech Biopharma Ltd
	HLA-matched peptides + adenovirus-based			Valo Therapeutics Ltd
	Structural proteins expressing MVA			Spain, Centro Nacional Biotecnología
Viral vector (non-replicating)	Spike protein expressing vaccine Parainfluenza virus 5 (PIV5)-based	Pan-Corona SARS-CoV2	Multiple candidates MERS	Lowa University/Georgia University
	S1 containing Recombinant deactivated rabies virus			CCHFV, LASSA, EBOV, MERS, NiV, HeV
	H1N1 vector H1N1 vector			Thomas Jefferson University/Bharat Biotech
	S expressing Newcastle disease virus			Egypt, National Research Centre
	Lentiviral Vector			Mount Sinai, Icahn School of Medicine
	Lentiviral Vector			Institut Pasteur -Theravectys
	Retro-VLP Particles Lentiviral Vector			AIOVA
	intranasal administration Ad 5 vector			University of Sorbonne
	TBD			Eastern Finland University and Helsinki University
	Vaxart			
Protein subunit (P.S.)	Mannose-conjugated chitosan nanoparticle delivered via RBD protein	Pan-Corona SARS-CoV2	VEE, HBV, RVF, EBOV, LASV, CHIKV, NORV, InFA	University of Kazakh National Agrarian/University of Ohio State
	Essai O/W 1,849,101 adjuvant with recombinant spike protein			University of Kazakh National Agrarian
	Peptides			Neo7Logic
	Essai O/W 1,849,101 adjuvant with recombinant spike protein			National Scientific Center for Especially Dangerous Infections/Kazakhstan, University of Kazakh National Agrarian
	Recombinant spike protein			Colloids and Interfaces
	FAR-Squalene adjuvant + RBD protein (baculovirus production)			Max-Planck-Institute
	Protein Subunit			Universidad Peruana Cayetano Heredia (UPCH)/FarmacológicosVeterinarios SAC (FARVET SAC)
	RBD-protein			Rep of Kazakhstan, Research Institute for Biological Safety Problems
	Recombinant S protein			Mynvax
	Novel adjuvant + Peptide			Biomedicine and Genome Center Izmir
Viral vector (non-replicating)	3M052 adjs./S subunit intranasal liposomal formulation with GLA	Pan-Corona SARS-CoV2	HIV, Malaria, Zika, NSCLC	University of Bogazici
	Adjuvant, E coli-based Expression + S-Protein (Subunit)			University of Virginia
	S, N, M and S1 protein subunit			Nigeria, Oyo State, Ogbomoso, Trinity
	Protein Subunit			Immonoefficient and Ogbomoso Laboratory, Helix Biogen Consult
	Adj. + RBD protein fused with Fc of IgG			Egypt, National Research Centre
	Capsid-like Particle			Argentina, San Martin and CONICET
	VLPs Drosophila S2 insect cell expression system			University
	LNP formulated peptide antigens			Thailand, GPO/University of Chulalongkorn
	S Protein			AdaptVac (PREVENT - n CoV consortium)
				ExpreS2ion

Table 8 (Continued)

Description of vaccine	Type of vaccine candidate	The target for coronavirus	Non-corona virus candidate's same platform	Developers
	Adjuvant + Sprotein		Influenza	UMN Pharma/Shionogi/National Institute of Infectious Disease, Japan
	Adjuvant + VLP-recombinant protein			BIKEN/University of Osaka/National Institutes of Biomedical Innovation, Japan
	S1 subunit microneedle arrays		MERS	Pittsburgh University
	Peptide			Vaxil Bio
	Adjuvanted protein subunit (RBD)		Breast CA vaccine, HPV therapeutic vaccine, ZIKA, Ebola, Marburg, HIV, Influenza	Biological E Ltd
	Peptide			Flow Pharma Inc
	S protein			A.J. Vaccines
	li-Key peptide		SARS-CoV, Influenza, HIV	EpiVax/Generex
	S protein		H7N9	Georgia University/EpiVax
	EPV-CoV-19 protein subunit			EpiVax
	gp-96 backbone		HIV, Malaria, Zika, NSCLC	Miami University/Heat Biologics
	vaccine subunit			Koltsovo, Rospotrebnadzor, FBRI SRC
	RBD protein or S1		SARS	VB VECTOR
	Plant produced protein subunit			Baylor College of Medicine
	Nanoparticles (based on S-protein and other epitopes), recombinant protein			CC-Pharming/iBio
	Truncated S (spike) proteins SARS-COV-2		HPV	Saint-Petersburg scientific research institute of vaccines
	XWG-03			GSK/University of Xiamen/Innovax
	Microsphere adjuvanted peptide			VIDO-Intervac, Saskatchewan University
	S and M proteins synthetic Long Peptide			OncoGen
	Vaccine candidate			
	S and N proteins Oral <i>E. coli</i> -based protein expression system			MIGAL Galilee Research Institute
	Nanoparticle vaccine			Lake Pharma, Inc.
	(RBD-Fc + Adjuvant) plant-based vaccine			Chula Vaccine Research Center/BaiyaPhytopharm
	vaccine based on OMV		Flu A, Plague	Quadram Institute Biosciences
	vaccine based on OMV			Trento University/BiOMViSSrI
	tobacco mosaic virus (TMV) structurally modified spherical particles		Rotavirus, Rubella	University of Lomonosov Moscow State
	Spike-based		Hepatitis C	Alberta University
	S1-Fc fusion recombinant protein			AnyGo Technology
	Recombinant protein			Yisheng Biopharma
	(Insect cell line baculovirus expression system)			Bristol University U.K. and Vietnam,
	Recombinant S protein in IC-BEVS			Vabiotech
	Heat stable, orally delivered subunit			Applied Biotechnology Institute, Inc.
	Spike protein peptides			Axon Neuroscience S.E.
	Protein Subunit			G.C. Pharma, MOGAM Institute for Biomedical Research
	RBD-based			Tel Aviv University/Neovii
	OMVsubunit			Epivax/Intravacc
	(Epitope screening) Spike-based			LiteVax BV ImmunoPrecise
	Spike-based			University of Ankara, Middle East Technical University, Nanografi Nano Technology
	Adjuvant with a recombinant spike			Iran
	BEVS produced recombinant S protein			University of Tampere
	Nanoformulated protein subunit			INRAE, CEA, Vaxinano
	Adenoviral Carrier protein subunit			CNRS, CEA
	DC-targeted epitopes			VRI, LinkinVax
Bacterial vector replicating	Protein expression system of RBD based on Oral <i>Salmonella enteritidis</i> (3934Vac)			Universidad Peruana Cayetano Heredia (UPCH)/FarmacológicosVeterinarios SAC (FARVET SAC)
Viral vector Replicating	YF17D Vector			K.U. Leuven
	Measles Vector			Cadila Healthcare Limited
	Measles Vector			Koltsovo, Rospotrebnadzor, FBRI SRC
	S, N targets measles virus		CHIKV, H7N9, Zika	VB VECTOR
	S protein expressing horsepox vector			CanVirex AG/DZIF - German Center for Infection Research
	(Intranasal) Attenuated influenza virus backbone LVVV		Monkeypox, Smallpox	Southern Research/Tonix Pharma
	(Intranasal) Influenza A virus, recombinant vaccine		Influenza	IEM and BIOCAD
				Koltsovo, Rospotrebnadzor, FBRI SRC
				VB VECTOR

Table 8 (Continued)

Description of vaccine	Type of vaccine candidate	The target for coronavirus	Non-corona virus candidate's same platform	Developers	
Vaccine based on RNA	Expressing antigenic portion of the Spike protein attenuated influenza RBD expressing influenza vector	SARS-CoV2	Influenza	Fundação Oswaldo Cruz and Instituto Buntantan	
	SARS-CoV-2 Spike (S) glycoprotein delivered by Replication-competent VSV chimeric virus technology (VSVΔG)		Lassa, Marburg, Ebola	Hong Kong University Merck/IAVI	
	DC-targeting replicating VSV vector		MERS, HIV	Influenza	Manitoba University
	VSV-S				Western Ontario University
	VSV-S				Aurobindo
	VSV vector				FBRI SRC VB VECTOR, Rospotrebnadzor, Koltsovo
	Influenza vector M2-deficient single replication (M2SR)				Bharat Biotech/FluGen/UW-Madison
	(NDV-SARS-CoV-2/Spike) Newcastle disease virus vector				Utrecht University/Intravacc/Wageningen Bio veterinary Research
	(APMV) Avian paramyxovirus vector		SARS-CoV3		University of Lancaster, UK
	RBD expressing Intranasal Newcastle disease virus vector (rNDV-FARVET)				Universidad Peruana Cayetano Heredia (UPCH)/FarmacológicosVeterinarios SAC (FARVET SAC)
NLC saRNA formulated			Amyris, Inc. Infectious Disease Research Institute		
S encoding LNP-encapsulated Mrna			Max-Planck-Institute of Colloids and Interfaces		
Self-amplifying RNA mRNA		Live, attenuated virus (LAV)	Gennova University of Selcuk		
LNP-mRNA			Sanofi Pasteur/Translate Bio Precision Nanosystems/CanSino Biologics		
LNP-mRNA		Live, attenuated bacterial vector (LABV)			
Cocktail encoding VLP LNP-encapsulated Mrna			RNA cure Biopharma/Shanghai Jiao Tong University/Fudan University		
RBD encoding LNP-encapsulated mRNA			RNA cure Biopharma/Shanghai Jiao Tong University/Fudan University		
SARS-CoV-2 derived replicating Defective RNAs		MERS	Centro Nacional Biotecnología (CNB-CSIC), Spain		
mRNA encapsulated LNP			Daiichi-Sankyo/Tokyo University		
mRNA encapsulated Liposome			BIOCAD		
mRNA candidates			RNAimmune, Inc.		
mRNA			Koltsovo, Rospotrebnadzor, FBRI SRC VB VECTOR		
Intranasal delivery system mRNA			Stermina/University of Tongji/China CDC		
mRNA			eTheRNA		
mRNA			Greenlight Biosciences		
mRNA			IDIBAPS-Hospital Clinic, Spain		
mRNA			Providence Therapeutics		
mRNA			Cell Tech Pharmed		
LNP-encapsulated mRNA D614G variant			ReNAP Co		
Encapsulated mRNA			Globe Biotech Ltd		
SARS-CoV-2 S, M, N and NSPs targets to induce T cell responses (CD8)			CEA		
VLP			OSE immunotherapeutics		
Protein subunit	Virus-like particle-based Dendritic Cell (D.C.)-targeting vaccine			Max Planck Institute for Dynamics of Complex Technical Systems	
	VLP			Manitoba University	
sVirus like particle	VLP			University of Bezmialemlakif	
	(eVLP) Enveloped Virus-Like Particle	SARS-CoV-2, SARS-CoV, & MERS-CoV	CMV, GBM, Zika	Middle East Technical University VBI Vaccines Inc.	
	HIV VLPs integrated by S protein	SARS-CoV2		Grifols/Barcelona Supercomputing/IRTA-CReSA/IrsiCaixa	
	Adjuvant + VLP			AIDS Research GPO/Siriraj Hospital/University of Mahidol	
	Baculovirus vehicles, Virus-like particles and lentivirus		Malaria	Oncoimmunology group, Navarrabiomed	
RBD displayed on virus-like particles			Saiba GmbH		
Multipeptide display ADDomer™			Bristol University's Max Planck Centre and Imophoron Ltd		
Unknown			Doherty Institute		
VLP	SARS-CoV1, SARS-CoV2		OSIVAX		

Table 8 (Continued)

Description of vaccine	Type of vaccine candidate	The target for coronavirus	Non-corona virus candidate's same platform	Developers
	eVLP	SARS-CoV2	Malaria	ARTES
	Whole virus/VLPs peptides VLPs produced in BEVS VLP derived by plant			BiotechnologARTESBiotechnolog Sao Paulo University University of Tampere University of Shiraz

marily injures the vascular endothelium as it is a systemic disease, and a patient with COVID 19 ARDS that is covid 19 acute respiratory distress syndrome (CARDS) can develop multiorgan failure if not managed expertly. There are two types of SARS-CoV-2 phenotypes for Respiratory support management. Target SpO<sub>2</sub>: 92–96% (82–92% in COPD patients), PaO<sub>2</sub> ≥ 8 kPa = 60 mm Hg, PaCO<sub>2</sub> < 6 kPa or pH > 7.3, FiO<sub>2</sub> ≤ 0.4, Morphine sulphate, Midazolam, Benzodiazepine is given as per the severity of disease. Awake proning, monitoring C-reactive protein (CRP), CBC examination, chest x-ray [69]. Table 6, shown, demonstrates the differences between different phenotypes [70,71].

#### Cough management

Sixty percent of COVID 19 positive cases report dry cough with zero phlegm production due to inflammation or irritation in your respiratory tract, which can get better with steam, humidifiers, honey consumption, cough suppressants, saltwater gargle, codeine phosphate, morphine sulfate as per the severity of infection [72,73].

#### Fever management

Fever after five days of infection is the most common in 99% of cases as your body's normal immune response tries to kill a virus. Advising the patients to consume a large number of fluids and to take paracetamol or ibuprofen can help to manage fever. Still, the use of ibuprofen use has marked a question of concern as the adaptive immune response will interfere with the release of prostaglandins, suppressing the fever response also leads to an increase in activity of lymphocyte, hyperemic response, and organ tissues oxygenation, causing failure of multiple organs [74,75].

#### GI disturbances

Pain in the abdomen (3.6%), looseness of the bowels (10.1%), Emesis (3.6%) is joint in COVID 19 patients due to its ACE2 receptor, which is expressed highly in gastrointestinal intestinal epithelial cells. Also, its viral RNA has been found in stool specimens of patients. A cohort endoscopy study of 95 COVID 19 patients reported six additional cases and identified viral RNA in the stomach, esophagus, intestinal duodenum, and rectum from 2 severe cases. Treatment relies on supportive care, antiemetics, proton pump inhibitors (PPI), antidiarrheals, promethazine, ondansetron, metoclopramide, adequate hydration, fresh ginger boiled in water added with honey can also treat nausea and weakness can reduce vomiting. If loose motions persist, stop the diet and consume coconut water (John Wiley, 2020), (Lipi Roy, 2020).

#### Vaccines

Acute viral infections remain a leading cause of morbidity and mortality. This novel coronavirus pandemic has triggered unprecedented global health researchers and scientists to find a safe, effective vaccine against this virus. Extensive research can be done by gaining the knowledge from SARS and MERS vaccines development strategies and knowing the key targets such as

receptor-binding domain (RBD) of spike protein, nucleotide identification, immunization route, suitable animal model utilization, production facility scalability, are some of the essential parameters to be considered [78]. There are seven COVID 19 vaccines in the third phase of clinical trials. Out of which four are from China, two of the candidates are from China National Biotech Group (CNBG) [79]. On August 27, Sputnik V advanced Russia vaccine trials of Sputnik V announced COVID 19 vaccine trials for over six months in forty thousand volunteers (Table 8).

#### COVID 19 vaccine efficacy

The vaccines were marketed in short span of time after the deadly pandemic. The most important thing was whether these vaccines have good efficacy in neutralising the SARS-CoV-2 virus or there is long term protection by generating memory B-cells and T-cells. In Table 7 we have discussed various marketed vaccines and their efficacy like the dosage regimen, antibody response, T cell response, and effectiveness. There are many vaccines which have received emergency approval in many countries. As of April 2021, 28 vaccines which have entered phase III clinical trials, and other 5 reported showed efficacy in the submitted reports to the peer-reviewed regulatory authorities for their emergency use through literature and/or through detailed publicly reports are available. For minimal protection 2 doses of vaccines are required for most of the them. Only 2 mRNA vaccines have shown efficacy at first dose after the detection of moderate TH1 cells and non neutralizing antibodies (Nabs). Induction of antibody dependent effector mechanisms, T-cell response, virus neutralization suggests that T cells, innate immune mechanisms, NABs low levels and other immune effector mechanisms are involved which helps in easy identification of protection mechanism and further understanding of immune system involvement for further development of vaccines.

#### Conclusion

SARS-CoV-2 is a novel strain of coronavirus that is liable for causing the global pandemic. This has challenged all the crucial factors like the global economy, medical infrastructure, and public work life, particularly the variant strains are causing havoc. The impact of this pandemic is so severe that it has shaken most countries' economies. Since the tremendous advancement has been achieved in understanding the condition, shortly there seems a strong possibility of some of the therapeutic interventions to combat the SARS-CoV-2 pandemic; till then, the only trusted intervention which is currently viable and proven to control is the following of strict quarantine measures, but to reach that intervention to all the affected groups there is a need of a more extensive set of randomized trials and fast testing of the condition to combat the disease effectively. However, this comprehensive review can provide some of the references for the follow-up medical studies. The spread of coronavirus trajectory across the World is difficult to predict as one country's problems will become global. The only possible solution is the vaccine development targeting against all variant strains to halt its progress, the identified theoretical and practical knowledge, current evidence, international alliances, ini-



tiatives, and ideas based on the values of cooperation, inclusiveness, and equity can eliminate the gaps to improve better understanding of the novel coronavirus structure and its design of a vaccine.

### Author's contribution

TF and JAM contributed equally in data collection and drafting the article. AHM, TA and KA contributed in data analysis and interpretation, and drafting the article. SA supervision and critical revision. All authors approved the final version of the article.

### Funding

No funding sources.

### Competing interests

None declared.

### Ethical approval

Not required.

### Acknowledgement

The authors would like to thank National Institute of Pharmaceutical Education and Research (NIPER), Guwahati and Hyderabad for fellowships.

### References

- Malik JA, Maqbool M. COVID-19: an overview of current scenario n.d. <https://doi.org/10.5667/CellMed.2020.0021>.
- Isaacs A, Gledhill AW, Andrewes CH. Influenza A viruses; laboratory studies, with special reference to European outbreak of 1950-1. *Bull World Health Organ* 1952;6:287–315.
- Vassilara F, Spyridaki A, Pothitos G, Deliveliotou A, Papadopoulos A. A rare case of human coronavirus 229E associated with acute respiratory distress syndrome in a healthy adult. *Case Rep Infect Dis* 2018;2018:1–4. <http://dx.doi.org/10.1155/2018/6796839>.
- World Health Organization. Coronavirus press conference 11 February, 2020 Speaker. *J Chem Inf Model* 2019;53:1689–99. <http://dx.doi.org/10.1017/CBO9781107415324.004>.
- Alonso S, Izeta A, Sola I, Enjuanes L. Transcription regulatory sequences and mRNA expression levels in the coronavirus transmissible gastroenteritis virus. *J Virol* 2002;76:1293–308. <http://dx.doi.org/10.1128/jvi.76.3.1293-1308.2002>.
- Di H, McIntyre AA, Brinton MA. New insights about the regulation of Nidovirus subgenomic mRNA synthesis. *Virology* 2018;517:38–43. <http://dx.doi.org/10.1016/j.virol.2018.01.026>.
- Kirchdoerfer RN, Cottrell CA, Wang N, Pallesen J, Yassine HM, Turner HL, et al. Pre-fusion structure of a human coronavirus spike protein. *Nature* 2016. <http://dx.doi.org/10.1038/nature17200>.
- Du L, He Y, Zhou Y, Liu S, Zheng BJ, Jiang S. The spike protein of SARS-CoV-A target for vaccine and therapeutic development. *Nat Rev Microbiol* 2009. <http://dx.doi.org/10.1038/nrmicro2090>.
- Alanagreh L, Alzoughool F, Atoum M. The human coronavirus disease covid-19: its origin, characteristics, and insights into potential drugs and its mechanisms. *Pathogens* 2020. <http://dx.doi.org/10.3390/pathogens9050331>.
- Inhibitor P, Hoffmann M, Kleine-weber H, Schroeder S, Mu MA, Drosten C, et al. SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven article SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. *Cell* 2020:271–80. <http://dx.doi.org/10.1016/j.cell.2020.02.052>.
- Zhou P, Yang X, Wang X, Hu B, Zhang L, Zhang W, et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature* 2020;2019. <http://dx.doi.org/10.1038/s41586-020-2012-7>.
- Liu M, Wang T, Zhou Y, Zhao Y, Zhang Y. Potential role of ACE2 in coronavirus disease 2019 (COVID-19) prevention and management. *J Transl Int Med* 2020;8. <http://dx.doi.org/10.2478/jtim-2020-0003>.
- Cui J. Origin and evolution of pathogenic coronaviruses. *Nat Rev Microbiol* 2019;17:181–92. <http://dx.doi.org/10.1038/s41579-018-0118-9>.
- Chen Y, Liu Q, Guo D. Emerging coronaviruses: genome structure, replication, and pathogenesis. *J Med Virol* 2020:418–23. <http://dx.doi.org/10.1002/jmv.25681>.
- Masters PS. The molecular biology of coronaviruses n.d.;66. [https://doi.org/10.1016/S0065-3527\(06\)66005-3](https://doi.org/10.1016/S0065-3527(06)66005-3).
- Neuman BW, Kiss G, Kunding AH, Bhella D, Baksh MF, Connelly S, et al. A structural analysis of M protein in coronavirus assembly and morphology. *J Struct Biol* 2011. <http://dx.doi.org/10.1016/j.jsb.2010.11.021>.
- Demogines A, Farzan M, Sawyer SL, Demogines A, Farzan M, Sawyer SL. Evidence for ACE2-utilizing coronaviruses (CoVs) related to severe acute respiratory syndrome CoV in bats. *J Virol* 2012;86. <http://dx.doi.org/10.1128/JVI.00311-12>.
- Almaza F, Dediago ML, Enrique A, Rejas T, Lamirande E, Roberts A, et al. A severe acute respiratory syndrome coronavirus that lacks the E gene is attenuated in vitro and in vivo. *J Virol* 2007;81:1701–13. <http://dx.doi.org/10.1128/JVI.01467-06>.
- Fehr AR, Perlman S. Chapter 1 coronaviruses: an overview of their replication and pathogenesis. *Coronaviruses 2015*;1282. <http://dx.doi.org/10.1007/978-1-4939-2438-7>.
- Cui L, Wang H, Ji Y, Yang J, Xu S, Huang X, Wang Z, Qin L, Tien P, Zhou X, Guo D, Chen Y. The nucleocapsid protein of coronaviruses acts as a viral suppressor of RNA silencing in mammalian cells. *J Virol* 2015;89(17):9029–43. <http://dx.doi.org/10.1128/JVI.01331-15>. Epub 2015 Jun 17. PMID: 26085159; PMCID: PMC4524063.
- Wit E De, Doremalen N Van, Falzarano D, Munster VJ. REVIEWS SARS and MERS: recent insights into emerging coronaviruses. *Nat Rev Microbiol* 2016;14:523–34. <http://dx.doi.org/10.1038/nrmicro.2016.81>. Nature Publishing Group.
- Lim YX, Ng YL, Tam JP, Liu DX. Human coronaviruses: a review of virus – host interactions. *Diseases* 2016. <http://dx.doi.org/10.3390/diseases4030026>.
- Huang C, Lokugamage KG, Rozovics JM, Narayanan K, Semler BL, Makino S, et al. Alphacoronavirus transmissible gastroenteritis virus nsp1 protein suppresses protein translation in mammalian cells and in cell-free HeLa cell extracts but not in rabbit reticulocyte lysate alphacoronavirus transmissible gastroenteritis virus nsp1 protein. *J Virol* 2011. <http://dx.doi.org/10.1128/JVI.01806-10>.
- Kamitani W, Huang C, Narayanan K, Lokugamage KG, Makino S. A two-pronged strategy to suppress host protein synthesis by SARS coronavirus Nsp1 protein. *Nat Struct Mol Biol* 2009;16:1134–40. <http://dx.doi.org/10.1038/nsmb.1680>.
- Kamitani W, Narayanan K, Huang C, Lokugamage K, Ikegami T, Ito N, et al. Severe acute respiratory syndrome coronavirus nsp1 protein suppresses host gene expression by promoting host mRNA degradation. *Proc Natl Acad Sci U S A* 2006.
- Tanaka T, Kamitani W, Dediago ML, Enjuanes L, Matsuura Y. Severe acute respiratory syndrome coronavirus nsp1 facilitates efficient propagation in cells through a specific translational shutoff of host mRNA. *J Virol* 2012;86:11128–37. <http://dx.doi.org/10.1128/JVI.01700-12>.
- Graham RL, Sims AC, Brockway SM, Baric RS, Denison MR. The nsp2 replicase proteins of murine hepatitis virus and severe acute respiratory syndrome coronavirus are dispensable for viral replication. *J Virol* 2005;79:13399–411. <http://dx.doi.org/10.1128/JVI.79.21.13399>.
- Cornillez-ty CT, Liao L, Iii JRY, Kuhn P, Buchmeier MJ. Severe acute respiratory syndrome coronavirus nonstructural protein 2 interacts with a host protein complex involved in mitochondrial biogenesis and intracellular signaling. *J Virol* 2009;83:10314–8. <http://dx.doi.org/10.1128/JVI.100842-09>.
- Chatterjee A, Johnson MA, Serrano P, Pedrini B, Joseph JS, Neuman BW, et al. Nuclear magnetic resonance structure shows that the severe acute respiratory syndrome coronavirus-unique domain contains a macrodomain fold. *J Virol* 2009;83:1823–36. <http://dx.doi.org/10.1128/JVI.01781-08>.
- Heinonen M, Eglhoff M, Frangeul A, Gruetz A, Cambillau C, Ziebuhr J, et al. Structural and functional basis for ADP-ribose and poly (ADP-ribose) binding by viral macro domains. *J Virol* 2006;80:8493–502. <http://dx.doi.org/10.1128/JVI.00713-06>.
- Eriksson KK, Cervantes-barraga L, Ludewig B, Thiel V. Mouse hepatitis virus liver pathology is dependent on ADP-ribose-1'-phosphatase, a viral function conserved in the alpha-like supergroup. *J Virol* 2008;82:12325–34. <http://dx.doi.org/10.1128/JVI.02082-08>.
- Signaling N-B, Frieman M, Ratia K, Johnston RE, Mesecar AD, Baric RS. Severe acute respiratory syndrome coronavirus papain-like protease ubiquitin-like domain and catalytic domain regulate antagonism of IRF3 and NF-kappaB signaling. *J Virol* 2009;83:6689–705. <http://dx.doi.org/10.1128/JVI.02220-08>.
- Neuman BW, Joseph JS, Saikatendu KS, Serrano P, Chatterjee A, Johnson MA, et al. Proteomics analysis unravels the functional repertoire of coronavirus nonstructural protein 3. *J Virol* 2008;82:5279–94. <http://dx.doi.org/10.1128/JVI.02631-07>.
- Serrano P, Johnson MA, Almeida MS, Horst R, Herrmann T, Joseph JS, et al. Nuclear magnetic resonance structure of the N-terminal domain of nonstructural protein 3 from the severe acute respiratory syndrome coronavirus. *J Virol* 2007;81:12049–60. <http://dx.doi.org/10.1128/jvi.00969-07>.
- Clementz MA, Kanjanahaluethai A, O'Brien TE, Baker SC. Mutation in murine coronavirus replication protein nsp4 alters assembly of double membrane vesicles. *Virology* 2008;375:118–29. <http://dx.doi.org/10.1016/j.virol.2008.01.018>.
- Gadlage MJ, Sparks JS, Beachboard DC, Cox RG, Doyle JD, Stobart CC, et al. Murine hepatitis virus nonstructural protein 4 regulates virus-induced membrane modifications and replication complex function. *J Virol* 2010;84:280–90. <http://dx.doi.org/10.1128/jvi.01772-09>.
- Lu Y, Lu X, Denison MR. Identification and characterization of a serine-like proteinase of the murine coronavirus MHV-A59. *J Virol* 1995;69:3554–9. <http://dx.doi.org/10.1128/jvi.69.6.3554-3559.1995>.

- [39] Oostra M, Hagemerijer MC, van Gent M, Bekker CPJ, te Lintelo EG, Rottier PJM, et al. Topology and membrane anchoring of the coronavirus replication complex: not all hydrophobic domains of nsp3 and nsp6 are membrane spanning. *J Virol* 2008;82:12392–405, <http://dx.doi.org/10.1128/jvi.01219-08>.
- [40] Zhai Y, Sun F, Li X, Pang H, Xu X, Bartlam M, et al. Insights into SARS-CoV transcription and replication from the structure of the nsp7–nsp8 hexadecamer. *Nat Struct Mol Biol* 2005;12:980–6, <http://dx.doi.org/10.1038/nsmb999>.
- [41] Zhai Y, Sun F, Li X, Pang H, Xu X, Bartlam M, et al. Insights into SARS-CoV transcription and replication from the structure of the nsp7–nsp8 hexadecamer. *Nat Struct Mol Biol* 2005;12:980–6, <http://dx.doi.org/10.1038/nsmb999>.
- [42] Imbert I, Guillemot JC, Bourhis JM, Bussetta C, Coutard B, Egloff MP, et al. A second, non-canonical RNA-dependent RNA polymerase in SARS coronavirus. *EMBO J* 2006;25:4933–42, <http://dx.doi.org/10.1038/sj.emboj.7601368>.
- [43] Egloff MP, Ferron F, Campanacci V, Longhi S, Rancurel C, Dutartre H, et al. The severe acute respiratory syndrome-coronavirus replicative protein nsp9 is a single-stranded RNA-binding subunit unique in the RNA virus world. *Proc Natl Acad Sci U S A* 2004;101:3792–6, <http://dx.doi.org/10.1073/pnas.0307877101>.
- [44] Bouvet M, Debarnot C, Imbert I, Selisko B, Snijder EJ, Canard B, et al. In vitro reconstitution of sars-coronavirus mRNA cap methylation. *PLoS Pathog* 2010;6:1–13, <http://dx.doi.org/10.1371/journal.ppat.1000863>.
- [45] Decroly E, Debarnot C, Ferron F, Bouvet M, Coutard B, Imbert I, et al. Crystal structure and functional analysis of the SARS-coronavirus RNA cap 2'-O-methyltransferase nsp10/nsp16 complex. *PLoS Pathog* 2011;7, <http://dx.doi.org/10.1371/journal.ppat.1002059>.
- [46] Xu X, Liu Y, Weiss S, Arnold E, Sara SG. Molecular model of SARS coronavirus polymerase: implications for biochemical functions and drug design. *Nucleic Acids Res* 2003;31, <http://dx.doi.org/10.1093/nar/gkg916>.
- [47] Ivanov KA, Thiel V, Dobbe JC, Meer Y Van Der, Snijder EJ, Ziebuhr J. Multiple enzymatic activities associated with severe acute respiratory syndrome coronavirus helicase. *J Virol* 2004;78:5619–32, <http://dx.doi.org/10.1128/JVI.78.11.5619>.
- [48] Ivanov KA, Ziebuhr J. Human coronavirus 229E nonstructural protein 13: characterization of duplex-unwinding, nucleoside triphosphatase, and RNA 5'-J-triphosphatase activities. *J Virol* 2004;78:7833–8, <http://dx.doi.org/10.1128/JVI.78.14.7833>.
- [49] Eckerle LD, Becker MM, Halpin RA, Li K, Venter E, Lu X, et al. Infidelity of SARS-CoV Nsp14-exonuclease mutant virus replication is revealed by complete genome sequencing. *PLoS Pathog* 2010;6, <http://dx.doi.org/10.1371/journal.ppat.1000896>.
- [50] Eckerle LD, Lu X, Sperry SM, Choi L, Denison MR. High fidelity of murine hepatitis virus replication is decreased in nsp14 exonuclease mutants. *J Virol* 2007;81:12135–44, <http://dx.doi.org/10.1128/JVI.01296-07>.
- [51] Cambillau C, Minskaia E, Hertzog T, Gorbalenya AE, Canard B, Ziebuhr J. Discovery of an RNA virus 3'→5' exonuclease that is critically involved in coronavirus RNA synthesis. *Proc Natl Acad Sci U S A* 2006:10–5.
- [52] Chen Y, Cai H, Xiang N, Tien P, Ahola T, Guo D. Functional screen reveals SARS coronavirus nonstructural protein nsp14 as a novel cap N7 methyltransferase. *Proc Natl Acad Sci U S A* 2009;106:3–8.
- [53] Bhardwaj K, Sun J, Holzenburg A, Guarino LA, Kao CC. RNA recognition and cleavage by the SARS coronavirus endoribonuclease. *J Mol Biol* 2020, <http://dx.doi.org/10.1016/j.jmb.2006.06.021>.
- [54] Ivanov KA, Hertzog T, Rozanov M, Bayer S, Thiel V, Gorbalenya AE, et al. Major genetic marker of nidoviruses encodes a replicative endoribonuclease. *Proc Natl Acad Sci U S A* 2004.
- [55] Decroly E, Imbert I, Coutard B, Selisko B, Alvarez K, Gorbalenya AE, et al. Coronavirus nonstructural protein 16 is a Cap-0 binding enzyme possessing (Nucleoside-2'J O)-methyltransferase activity. *J Virol* 2008;82:8071–84, <http://dx.doi.org/10.1128/JVI.00407-08>.
- [56] Züst R, Cervantes-barragan L, Habjan M, Maier R, Neuman BW, Ziebuhr J, et al. Ribose 2'-O-methylation provides a molecular signature for the distinction of self and non-self mRNA dependent on the RNA sensor Mda5. *Nat Immunol* 2011;12:137–44, <http://dx.doi.org/10.1038/ni.1979>. Nature Publishing Group.
- [57] Gilbert GL. Commentary: SARS, MERS and COVID-19—new threats; old lessons. *Int J Epidemiol* 2020:1–3, <http://dx.doi.org/10.1093/ije/dyaa061>.
- [58] Petrosillo N, Viceconte G, Ergonul O, Ippolito G, Petersen E. COVID-19, SARS and MERS: are they closely related? *Clin Microbiol Infect* 2020;26:729–34, <http://dx.doi.org/10.1016/j.cmi.2020.03.026>.
- [59] Rockx B, Kuiken T, Herfst S, Bestebroer T, Lamers MM, Munnink BBO, et al. Comparative pathogenesis of COVID-19, MERS, and SARS in a nonhuman primate model. *Science* 2020;7314:1–10.
- [60] Jennifer M. Characteristics of and important lessons from the coronavirus disease 2019 (COVID-19) outbreak in China summary of a report of 72 314 cases from the Chinese Center for Disease Control and Prevention. *JAMA* 2020.
- [61] COVID Live Update: 146,881,796 Cases and 3,107,317 Deaths from the Coronavirus - Worldometer n.d. <https://www.worldometers.info/coronavirus/> (accessed April 25, 2021).
- [62] Puhani PA. France and Germany exceed Italy, South Korea and Japan in temperature-adjusted corona proliferation a quick and dirty Sunday morning analysis; 2020.
- [65] March I. COVID-19 pandemic in India; 2020, 2019.
- [66] Kohler PF, Farr RS. Elevation of Cord over maternal IgG immunoglobulin: evidence for an active placental IgG transport. *Nature* 1966;210:1070–1, <http://dx.doi.org/10.1038/2101070a0>.
- [67] Burki T. Understanding variants of SARS-CoV-2. *Lancet* 2021, [http://dx.doi.org/10.1016/S0140-6736\(21\)00298-1](http://dx.doi.org/10.1016/S0140-6736(21)00298-1).
- [68] Who W. Clinical management of severe acute respiratory infection (SARI) when COVID-19 disease is suspected; 2020, 2019.
- [69] Möhlenkamp S, Thiele H. Ventilation of COVID-19 patients in intensive care units. *Herz* 2020;45:329–31, <http://dx.doi.org/10.1007/s00059-020-04923-1>.
- [70] Gattinoni L, Chiumello D, Caironi P, Busana M, Romitti F, Brazzi L, et al. COVID-19 pneumonia: different respiratory treatments for different phenotypes? *Intensive Care Med* 2020;46:1099–102, <http://dx.doi.org/10.1007/s00134-020-06033-2>.
- [71] Marini JJ, Hotchkiss JR, Broccard AF. Bench-to bedside review: microvascular and airspace linkage in ventilator-induced lung injury. *Crit Care* 2003;7:435–44, <http://dx.doi.org/10.1186/cc2392>.
- [72] Zhao Y, Cao Y, Wang S, Cai K, Xu K. COVID-19 and gastrointestinal symptoms. *Br J Surg* 2020;107(September (10)):e382–3, <http://dx.doi.org/10.1002/bjs.11821>. Epub 2020 Aug 5. PMID: 32757447; PMCID: PMC7436706.
- [73] Who W. Clinical management of severe acute respiratory infection (SARI) when COVID-19 disease is suspected; 2020, 2019.
- [74] Zhao Y, Cao Y, Wang S, Cai K, Xu K. COVID-19 and gastrointestinal symptoms. *Br J Surg* 2020;2019:2019–20, <http://dx.doi.org/10.1002/bjs.11821>.
- [75] Moore N, Carleton B, Blin P, Bosco P, Cecile L. Does ibuprofen worsen COVID 19? *Drug Saf* 2020:19–22, <http://dx.doi.org/10.1007/s40264-020-00953-0>.
- [78] Malik JA, Mulla AH, Farooqi T, Potttoo FH, Anwar S, Rengasamy KRR. Targets and strategies for vaccine development against SARS-CoV-2. *Biomed Pharmacother* 2021;137:111254, <http://dx.doi.org/10.1016/j.biopha.2021.111254>.
- [79] Dutta NK, Mazumdar K, Gordy JT. The nucleocapsid protein of SARS-CoV-2: a target for vaccine development. *J Virol* 2020;94:1–2, <http://dx.doi.org/10.1128/jvi.00647-20>.
- [80] Sadarangani M, Marchant A, Kollmann TR. Immunological mechanisms of vaccine-induced protection against COVID-19 in humans. *Nat Rev Immunol* 2021;21:1–10, <http://dx.doi.org/10.1038/s41577-021-00578-z>.