

Renoprotective Effects of Melatonin against Vancomycin-Related Acute Kidney Injury in Hospitalized Patients: a Retrospective Cohort Study

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ABSTRACT Vancomycin is associated with nephrotoxicity, and the mechanism may in part be related to oxidative stress. In vitro and preclinical studies suggest that melatonin supplementation decreases oxidative stress. The objective of this study was to evaluate concomitant use of melatonin and vancomycin and the incidence of acute kidney injury (AKI). We performed a retrospective cohort study at a large community medical center. All consecutive patients admitted to the medical center between January 2016 and September 2020 who received vancomycin therapy alone or concomitantly with melatonin as part of ordinary care were considered for inclusion. The primary endpoint was the development of AKI, defined as an absolute increase in serum creatinine of \geq 0.3 mg/dl or $a \geq$ 50% increase in serum creatinine. All data were analyzed using descriptive statistics. A multivariable logistic regression was constructed to account for potential confounding variables. We identified a total of 303 adult patients meeting inclusion and exclusion criteria treated with vancomycin, 101 of which received melatonin concomitantly. Overall baseline characteristics were similar between the two groups except for the incidence of bacteremia/sepsis. After controlling for the vancomycin area under the curve, baseline creatinine clearance, and intensive care unit admission in a multivariable logistic regression analysis, melatonin use was associated with a 63% decrease in AKI (odds ratio [OR], 0.37; 95% confidence interval [CI], 0.14 to 0.96; $P = 0.041$). Melatonin use was associated with a significant reduction in vancomycin-related AKI. Although this was a retrospective study with a small sample size, given the magnitude of the difference seen, further large prospective studies are warranted.

KEYWORDS acute kidney injury, antioxidant, melatonin, vancomycin

Vancomycin was introduced into clinical practice in 1956 as a first line of invasive treatment for Gram-positive infections. Vancomycin is a glycopeptide antibiotic effective against Gram-positive infections, including methicillin-resistant Staphylococcus aureus (MRSA). In many severe MRSA infections, vancomycin dose adjustment is recommended to achieve an area under the curve (AUC)/MIC ratio for better response. However, vancomycin causes acute kidney injury (AKI) with prolonged and higher doses [\(1](#page-6-0)[–](#page-6-1)[3](#page-6-2)). Initially, the nephrotoxicity and ototoxicity were attributed to the earlier formulations of vancomycin, which contained product impurities and were termed "Mississippi mud" [\(4](#page-6-3)[–](#page-6-4)[6\)](#page-6-5). Despite formulation improvements, the reported incidence of AKI with vancomycin use ranges between 5% and 43%, based on the dose and duration of therapy [\(7](#page-6-6)–[10\)](#page-6-7). In a recent study, approximately 9% of hospitalized patients with acute bacterial skin and skin structure infections, a population with a relatively

Citation Hong TS, Briscese K, Yuan M, Deshpande K, Aleksunes LM, Brunetti L. 2021. Renoprotective effects of melatonin against vancomycin-related acute kidney injury in hospitalized patients: a retrospective cohort study. Antimicrob Agents Chemother 65:e00462- 21. <https://doi.org/10.1128/AAC.00462-21>.

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Received 6 March 2021 Returned for modification 2 May 2021 Accepted 20 June 2021

Accepted manuscript posted online 21 June 2021 Published 17 August 2021

low risk of AKI, treated with vancomycin developed AKI ([11](#page-7-0)). Furthermore, each additional day of vancomycin therapy beyond 72 h was associated with a 14%-increased risk of AKI, with a $>$ 2-fold-higher risk with treatment durations exceeding 7 days ([11](#page-7-0)).

Although the exact mechanism of vancomycin-related AKI is not clearly defined, reported preclinical data suggest that vancomycin may cause renal damage by inducing oxidative stress in proximal tubules [\(12](#page-7-1)–[17](#page-7-2)). There are some preclinical studies conducted in mice that have indicated that vancomycin is associated with the downregulation of antioxidant genes and, subsequently, a negative impact on mitochondrial bioenergetics ([18](#page-7-3)[–](#page-7-4)[20\)](#page-7-5).

A variety of antioxidants have been investigated for protection against vancomycin-related AKI, primarily in preclinical studies. These have included vitamin E [\(21\)](#page-7-6), gingko biloba, vitamin C, N-acetylcysteine, curcumin, and melatonin [\(22,](#page-7-7) [23\)](#page-7-8). First isolated in 1958, N-acetyl-5-methoxytryptamine, commonly known as melatonin, is a synthetic product of the vertebrate pineal gland. Melatonin was initially discovered as a mediator of circadian rhythm, but it has since been studied as a direct scavenger of free radicals and a general antioxidant ([24](#page-7-9)[–](#page-7-10)[26\)](#page-7-11). Melatonin has a receptor-mediated mechanism, and its structure itself helps to reinforce the inhibition of oxidatively mediated stress. The main advantage of using melatonin as an antioxidant is that it is selectively taken up by the mitochondrial membrane, which is not applicable to other antioxidants [\(27](#page-7-12)). Melatonin is now well recognized as an antioxidant, and several preclinical studies demonstrate its renoprotective properties against oxidative-stressinduced nephrotoxicity, including that caused by vancomycin [\(12](#page-7-1), [28](#page-7-13), [29\)](#page-7-14). The consideration of melatonin for renoprotection is an interesting option, since it is commonly used as a sleep aid in hospitalized patients due to an efficacy similar to and a safety profile better than that of non-benzodiazepine hypnotics, as we have previously reported [\(30\)](#page-7-15). In this study, we evaluated the association between melatonin use and the development of AKI in hospitalized adult patients receiving vancomycin.

RESULTS

We identified a total of 303 adult patients who were treated with vancomycin during the specified time period, 101 of which were on melatonin in combination with vancomycin. The mean number of melatonin doses was 6.5 ± 8.1 doses per patient in the treatment group. More than half of patients (53.5%) received melatonin before or on the same day as vancomycin therapy initiation. [Table 1](#page-2-0) provides a summary of the patient demographics and vancomycin dosing characteristics. Overall, baseline characteristics were similar between the two groups except for the incidence of bacteremia/ sepsis (25.2% for the control group versus 37.6% for the treatment group $[P = 0.026]$).

Patients frequently received other antibiotics ([Table 2\)](#page-3-0), with the most common agent being piperacillin-tazobactam (49.5% for the control versus 46.5% for the treatment group $[P = 0.626]$). Most patients also received other nephrotoxins (73.3% for the control versus 78.2% for the treatment group $[P = 0.349]$, such as nonsteroidal anti-inflammatory drugs (NSAIDs), loop diuretics, and angiotensin-converting enzyme inhibitors (ACEIs) or angiotensin receptor blockers (ARBs), all of which were similar between groups. The most common vancomycin dosing frequency was every 8 h (Q8h) for the control group (46.0%) and Q12h for the treatment group (57.4%). The total daily dose of vancomycin was higher in the control group (2.8 \pm 1.2 g/day for the control versus 2.6 \pm 1.1 g/day for the treatment group $[P = 0.186]$), but this difference was not statistically significant. The total number of days of vancomycin therapy was longer for the treatment group (4.2 \pm 2.5 days for the control versus 4.8 \pm 3.1 days for the treatment group [P = 0.068]), but this difference was also not statistically significant. The initial mean trough concentrations at steady state were similar between the two groups (16.2 \pm 6.6 μ g/ml for the control versus 16.7 \pm 6.6 μ g/ml for the treatment group $[P = 0.632]$). The mean AUCs, estimated with a Bayesian model, were also similar between the two groups $(487.4 \pm 215.9 \text{ mg} \cdot \text{h/liter}$ for the control versus 479.2 \pm 256.7 mg \cdot h/liter for the treatment group [P = 0.770]). The median changes in creatinine clearance (CL_{CR}) from the start of vancomycin therapy to discontinuation of therapy

TABLE 1 Baseline comparison between groups^{a}

^aBUN, blood urea nitrogen; NA, not available.

were significantly different between the two groups (3.5 ml/min for the control versus 11.0 ml/min for the treatment group $[P = 0.047]$). However, the mean change in serum creatinine (sCr) from the start of vancomycin therapy to discontinuation of therapy was lower in the treatment group (0.1 \pm 0.5 mg/dl for the control versus -0.1 ± 0.4 mg/dl for the treatment group $[P = 0.007]$) ([Table 3\)](#page-4-0). Furthermore, patients who experienced AKI had significantly longer median lengths of stay (LOS) than the patients who did not experience AKI (6.0 days for patients without AKI versus 11.0 days for patients with AKI [P < 0.001]). For patients who developed AKI in the treatment group, there was no statistically significant

Clindamycin 8, 4.0 2, 2.0 0.505 Other 49, 24.3 25, 25.3 0.925 Trimethoprim-sulfamethoxazole $7, 3.5$ 5, 5.1 0.532 Daptomycin 10, 5.0 4, 4.0 0.781 Linezolid 5, 2.5 1, 1.0 0.667 Metronidazole 33, 16.3 15, 15.2 0.739 Nitrofurantoin 1, 0.1 2, 2.0 0.259 Other nephrotoxins^a 148, 73.3 79, 78.2 0.349 NSAIDs 99, 49.0 47, 46.5 0.684 ACEI/ARB 51, 25.2 29, 28.7 0.519 Loop diuretics 69, 34.1 46, 45.5 0.054 Contrast media 87, 43.1 36, 35.6 0.264

TABLE 2 Comparison of other prescribed antibiotics and nephrotoxins between patients receiving concomitant vancomycin and melatonin and those receiving vancomycin alone

^aNSAIDs, nonsteroidal anti-inflammatory drugs; ACEI, angiotensin-converting enzyme inhibitors; ARB, angiotensin receptor blockers.

difference between the patients who started melatonin before or on the same day as vancomycin therapy initiation and the patients who started melatonin after vancomycin therapy initiation (2/54 versus $6/47$ [$P = 0.141$]).

Results of the univariate and multivariable analyses on the risk factors associated with the onset of AKI are shown in [Table 4.](#page-4-1) The incidence of AKI was lower in patients treated with melatonin than in the control group (odds ratio [OR], 0.44; 95% confidence interval [CI], 0.20 to 0.99; P value = 0.048). In the univariate analysis, concomitant piperacillin-tazobactam use during the vancomycin therapy was associated with a significantly increased risk of developing AKI (OR, 3.90; 95% CI, 1.83 to 8.29; P value $<$ 0.001). The use of a loop diuretic, which was one of the common nephrotoxic agents used in both groups, was also a major risk factor for developing AKI (OR, 3.37; 95% CI, 1.70 to 6.7; P value = 0.001). We have also

TABLE 3 Comparison of clinical characteristics and outcomes

^aIQR, interquartile range.

identified other risk factors in univariate analysis, such as an AUC of >600 mg \cdot h/liter, a Charlson-Deyo comorbidity index (CCI) of >3 , and admission to an intensive care unit (ICU). However, only covariates for which the P was \leq 0.1 were considered for further evaluation in the multivariable analysis. After controlling for an AUC of >600 mg \cdot h/liter, a CL_{CR} of \geq 60 ml/min, and admission to the ICU in multivariable logistic regression, melatonin use was associated with a 63% decrease in AKI (OR, 0.37; 95% CI, 0.14 to 0.96; P value = 0.041).

DISCUSSION

To our knowledge, this is the first human study assessing the association between melatonin and vancomycin-related nephrotoxicity in hospitalized patients. Although the impact of melatonin on drug-related nephrotoxicity in in vitro models has been previously reported, its clinical applicability has not been evaluated in hospitalized patients. In this study, the use of melatonin during the vancomycin therapy was associated with a 63% reduction in AKI.

In our univariate analysis, an age of \geq 65 years, AUC values of $>$ 600 mg \cdot h/liter, a CCI score of >3 , loop diuretics, ICU admission, and concomitant piperacillin-tazobactam use were associated with an increased risk of developing AKI. Previous studies have reported on the increased risk of nephrotoxicity with the concomitant use of piperacillin-tazobactam and vancomycin therapy [\(31](#page-7-16)[–](#page-7-17)[33](#page-7-18)).

Preclinical studies have demonstrated that vancomycin may induce kidney injury through oxidative stress and mitochondrial damage [\(18,](#page-7-3) [34](#page-7-19)). As melatonin can restore the renal mitochondrial dynamic balance by inhibiting fission and promoting fusion activities ([35](#page-7-20)), melatonin may prevent vancomycin-related nephrotoxicity by improving renal mitochondrial function. Melatonin also increases the anti-inflammatory cytokines, thus reducing oxidative stress [\(19](#page-7-4), [36\)](#page-7-21). Furthermore, since a CL_{CR} of ≥ 60 ml/min was identified in our multivariable analysis to significantly lower the risk of developing AKI (OR, 0.45; 95% CI, 0.21 to 0.95; P value = 0.036), improving CL_{CR} should be prioritized to prevent hospitalized patients from developing vancomycin-related AKI.

^aAbbreviations: AUC, area under the curve; CCI, Charlson-Deyo comorbidity index; CL_{ce}, creatinine clearance; ICU, intensive care unit. b Only covariates with a P of \leq 0.1 were considered for further testing in the multivariable model.

AKI is commonly observed in hospitalized patients and is associated with an increased likelihood of mortality, hospital LOS, and cost [\(37](#page-7-22)[–](#page-7-23)[39\)](#page-7-24). Similarly, in our study, patients who experienced AKI had a significantly longer median LOS than the patients who did not experience AKI (6.0 days for patients without AKI versus 11.0 days for patients with AKI [$P < 0.001$]). Since each case of AKI increases the hospitalization costs by \$11,016 to \$42,077 per case ([40](#page-7-25)), this additional cost related to AKI indicates that melatonin may not only attenuate vancomycin-related AKI but also reduce the overall hospital LOS, further driving down the total cost for inpatient stay.

There are some limitations to our study that should be considered. First, the melatonin serum levels were not collected due to the lack of data availability. The oral bioavailability of melatonin in humans has a wide range, with melatonin serum levels differing up to 28-fold ([41](#page-7-26)). Therefore, the melatonin serum levels for each patient may vary substantially based on the patient's other clinical factors. Second, the doses and frequency of melatonin administration were not consistent among the patients in the treatment group. The mean number of melatonin doses was 6.5 ± 8.1 doses per patient in the treatment group. However, there were some variations in the dosage of melatonin administered. Most patients received 5 mg of melatonin, while some patients received 3 mg of melatonin (68.3% for 5 mg, 31.7% for 3 mg). Several preclinical studies in mice ([42\)](#page-7-27), rats [\(29,](#page-7-14) [43](#page-7-28), [44](#page-7-29)), and rabbits [\(45](#page-7-30)) have investigated the benefits of melatonin for protection against nephrotoxicity secondary to various insults. Dosing has varied among the studies; however, rats dosed with melatonin at 500 μ g/kg of body weight orally [\(43\)](#page-7-28) and 1 mg/kg ([44](#page-7-29)) appreciated a reduction in markers of kidney injury. These doses translate to approximately 5 to 10 mg in a 70-kg human using allometric scaling [\(46](#page-7-31)). Moreover, melatonin concentrations in highly perfused organs, such as the kidney, may be higher than those quantitated in plasma based on previous physiologically based pharmacokinetic modeling [\(47\)](#page-7-32). As such, the dose used in the current retrospective cohort reasonably recapitulates what has been appreciated in animal models. Nonetheless, the optimal dose and frequency of melatonin for its renoprotective effects still need further investigation. Similarly, although the timing of the melatonin administered did not significantly impact the incidence of vancomycinrelated AKI in the treatment group, there may be a greater benefit if the melatonin is started prior to the vancomycin therapy initiation. The race and gender of our study population were mostly white and male. This limits applicability to other patient populations, as the average rate of kidney function decline differs among different racial and ethnic groups [\(48](#page-7-33)). Moreover, our study included patients who received at least three doses of vancomycin regardless of their treatment indications. Therefore, the levels of severity of a disease may have differed among the patient populations, which may directly or indirectly impact their kidney functions. Even though most of the patients were treated with vancomycin empirically, the treatment group had a significantly greater number of patients treated for bacteremia/sepsis than the control group (25.2% for the control versus 37.6% for the treatment group $[P = 0.026]$). Nevertheless, we did account for overall comorbidity by adjusting our primary outcome for the CCI. We utilized sCr as a biomarker for AKI in our study. sCr is not an ideal biomarker because it may be influenced by other factors. Nonetheless, the current standard of care for assessing renal function is sCr. Finally, since this was a retrospective cohort study, there are inherent limitations with this study design. Confounding factors are always a concern in observational studies. We considered propensity score matching using covariates known to influence the incidence of AKI; however, the conclusions were unchanged (see Table S1 in the supplemental material). Despite these limitations, our findings provide valuable information about the impact of melatonin on hospitalized patients treated with vancomycin.

MATERIALS AND METHODS

Data source and patient selection. We performed a single-center, retrospective, institution review board (IRB)-approved cohort study using data from electronic patient medical records at a large community medical center. All consecutive patients (aged 18 years or older) discharged from the hospital between January 2016 and September 2020 who received at least three doses of vancomycin alone or with melatonin (GenDose Pharmaceuticals, Davidson, NC) were screened for inclusion. Patients were required to have a creatinine clearance (CL_{CP}) greater than or equal to 30 ml/min at baseline (first vancomycin dose). Patients who were on hemodialysis, peritoneal dialysis, or had severe kidney dysfunction, defined as a CL_{CR} of \leq 30 ml/min at baseline, were excluded.

Data extraction and collection. All data were extracted from the electronic health record (Allscripts Sunrise Clinical Manager). Patient age, sex, race/ethnicity, concomitant nephrotoxic agents, concomitant antibiotics, vancomycin indications, height, weight, and other relevant laboratory data were extracted from the medical record. Serum creatinine (sCr) concentrations were collected and used as a marker of vancomycin-related AKI. Baseline sCr concentrations were collected prior to the vancomycin therapy initiation, and the highest sCr concentrations during the vancomycin therapy were collected after vancomycin therapy initiation and until 24 h after the last dose. Patient comorbidities were identified using International Classification of Diseases, tenth revision, clinical modification (ICD-10-CM) codes, and subsequently, the Charlson-Deyo comorbidity index (CCI) was calculated to capture patient overall comorbidity. CL_{CR} was calculated with the Cockcroft-Gault equation using actual body weight. Area under the curve (AUC) data were estimated with DoseMe software (Tabula Rasa Healthcare, Moorsetown, NJ), which utilizes the Bayesian approach based on a published model ([49](#page-8-0)).

Outcome. Patients were divided into all those who received vancomycin concomitantly with melatonin and a random sample of those who did not receive melatonin using a 1-to-2 ratio. The primary endpoint of AKI was defined as an absolute increase in serum creatinine of ≥ 0.3 mg/dl or a $\geq 50\%$ increase in serum creatinine from the start of vancomycin therapy [\(50](#page-8-1)[–](#page-8-2)[52](#page-8-3)). Patients were evaluated for vancomycin-related AKI at any point after the first dose of vancomycin was initiated and until 24 h after the last dose. Two consecutive elevated sCr measurements were required to establish an AKI. Secondary endpoints included a change in CL_{CR} and sCr from baseline levels to completion of vancomycin therapy.

Statistical analysis. Patient demographics were summarized using descriptive statistics. Continuous data are reported as means and standard deviations, and nominal data are reported as percentages. The differences in the baseline characteristics, vancomycin dosing variables, and clinical outcomes were tested using the t test for continuous data and the chi-square test or Fisher's exact test for categorical data. Normality testing was performed using visual inspection of histograms and the Mann-Whitney U test. For data that were not normally distributed, results are reported as medians and interquartile ranges and were compared using the Kruskal-Wallis test. The paired t test was used to assess significance in covariates from baseline to posttreatment. A multivariable logistic regression was constructed using melatonin as the exposure variable and AKI as the outcome variable. Covariates were selected for inclusion based on previous literature and results of univariate analysis. Variables were retained in the final model if P was <0.05. All analyses were performed in SPSS v26 (IBM Corporation, Armonk, NY).

SUPPLEMENTAL MATERIAL

Supplemental material is available online only. SUPPLEMENTAL FILE 1, PDF file, 0.03 MB.

ACKNOWLEDGMENTS

This study was presented, in part, at the American Society of Health-System Pharmacists (ASHP) Midyear Clinical Meeting and Exhibition, 2020 (SP-363).

L.M.A. is supported by the National Institute of Environmental Health Sciences (grant P30ES005022), the National Institute of General Medical Science (grant R01GM123330), and the National Center for Advancing Translational Sciences (grant UL1TR003017).

We have no conflicts of interest to disclose.

REFERENCES

- 1. Gupta A, Biyani M, Khaira A. 2011. Vancomycin nephrotoxicity: myths and facts. Neth J Med 69:379–383.
- 2. Du H, Li Z, Yang Y, Li X, Wei Y, Lin Y, Zhuang X. 2020. New insights into the vancomycin-induced nephrotoxicity using in vitro metabolomics combined with physiologically based pharmacokinetic modeling. J Appl Toxicol 40:897–907. <https://doi.org/10.1002/jat.3951>.
- 3. Nolin TD. 2016. Vancomycin and the risk of AKI: now clearer than Mississippi mud. Clin J Am Soc Nephrol 11:2101–2103. [https://doi.org/10.2215/](https://doi.org/10.2215/CJN.11011016) [CJN.11011016](https://doi.org/10.2215/CJN.11011016).
- 4. Hodoshima N, Masuda S, Inui K. 2007. Decreased renal accumulation and toxicity of a new VCM formulation in rats with chronic renal failure. Drug Metab Pharmacokinet 22:419–427. [https://doi.org/10.2133/dmpk.22.419.](https://doi.org/10.2133/dmpk.22.419)
- 5. Moellering RC, Jr. 2006. Vancomycin: a 50-year reassessment. Clin Infect Dis 42(Suppl 1):S3–S4. [https://doi.org/10.1086/491708.](https://doi.org/10.1086/491708)
- 6. Filippone EJ, Kraft WK, Farber JL. 2017. The nephrotoxicity of vancomycin. Clin Pharmacol Ther 102:459–469. <https://doi.org/10.1002/cpt.726>.
- 7. Elyasi S, Khalili H, Dashti-Khavidaki S, Mohammadpour A. 2012. Vancomycin-induced nephrotoxicity: mechanism, incidence, risk factors and special populations. A literature review. Eur J Clin Pharmacol 68:1243–1255. <https://doi.org/10.1007/s00228-012-1259-9>.
- 8. Lodise TP, Patel N, Lomaestro BM, Rodvold KA, Drusano GL. 2009. Relationship between initial vancomycin concentration-time profile and nephrotoxicity among hospitalized patients. Clin Infect Dis 49:507–514. [https://doi.org/10](https://doi.org/10.1086/600884) [.1086/600884.](https://doi.org/10.1086/600884)
- 9. Pritchard L, Baker C, Leggett J, Sehdev P, Brown A, Bayley KB. 2010. Increasing vancomycin serum trough concentrations and incidence of nephrotoxicity. Am J Med 123:1143–1149. [https://doi.org/10.1016/j.amjmed.2010.07.025.](https://doi.org/10.1016/j.amjmed.2010.07.025)
- 10. van Hal SJ, Paterson DL, Lodise TP. 2013. Systematic review and metaanalysis of vancomycin-induced nephrotoxicity associated with dosing schedules that maintain troughs between 15 and 20 milligrams per liter. Antimicrob Agents Chemother 57:734–744. [https://doi.org/10.1128/AAC](https://doi.org/10.1128/AAC.01568-12) [.01568-12](https://doi.org/10.1128/AAC.01568-12).
- 11. Jorgensen SCJ, Murray KP, Lagnf AM, Melvin S, Bhatia S, Shamim M-D, Smith JR, Brade KD, Simon SP, Nagel J, Williams KS, Ortwine JK, Veve MP, Truong J, Huang DB, Davis SL, Rybak MJ. 2020. A multicenter evaluation of vancomycin-associated acute kidney injury in hospitalized patients with acute bacterial skin and skin structure infections. Infect Dis Ther 9:89–106. <https://doi.org/10.1007/s40121-019-00278-1>.
- 12. Celik I, Cihangiroglu M, Ilhan N, Akpolat N, Akbulut HH. 2005. Protective effects of different antioxidants and amrinone on vancomycin-induced nephrotoxicity. Basic Clin Pharmacol Toxicol 97:325–332. [https://doi.org/](https://doi.org/10.1111/j.1742-7843.2005.pto_153.x) [10.1111/j.1742-7843.2005.pto_153.x.](https://doi.org/10.1111/j.1742-7843.2005.pto_153.x)
- 13. Oktem F, Arslan MK, Ozguner F, Candir O, Yilmaz HR, Ciris M, Uz E. 2005. In vivo evidences suggesting the role of oxidative stress in pathogenesis of vancomycin-induced nephrotoxicity: protection by erdosteine. Toxicology 215:227–233. <https://doi.org/10.1016/j.tox.2005.07.009>.
- 14. Qu S, Dai C, Lang F, Hu L, Tang Q, Wang H, Zhang Y, Hao Z. 2019. Rutin attenuates vancomycin-induced nephrotoxicity by ameliorating oxidative stress, apoptosis, and inflammation in rats. Antimicrob Agents Chemother 63:e01545-18. [https://doi.org/10.1128/AAC.01545-18.](https://doi.org/10.1128/AAC.01545-18)
- 15. Sakamoto Y, Yano T, Hanada Y, Takeshita A, Inagaki F, Masuda S, Matsunaga N, Koyanagi S, Ohdo S. 2017. Vancomycin induces reactive oxygen speciesdependent apoptosis via mitochondrial cardiolipin peroxidation in renal tubular epithelial cells. Eur J Pharmacol 800:48–56. [https://doi.org/10.1016/j](https://doi.org/10.1016/j.ejphar.2017.02.025) [.ejphar.2017.02.025](https://doi.org/10.1016/j.ejphar.2017.02.025).
- 16. Shi H, Zou J, Zhang T, Che H, Gao X, Wang C, Wang Y, Xue C. 2018. Protective effects of DHA-PC against vancomycin-induced nephrotoxicity through the inhibition of oxidative stress and apoptosis in BALB/c mice. J Agric Food Chem 66:475–484. <https://doi.org/10.1021/acs.jafc.7b04565>.
- 17. Nishino Y, Takemura S, Minamiyama Y, Hirohashi K, Ogino T, Inoue M, Okada S, Kinoshita H. 2003. Targeting superoxide dismutase to renal proximal tubule cells attenuates vancomycin-induced nephrotoxicity in rats. Free Radic Res 37:373–379. [https://doi.org/10.1080/1071576031000061002.](https://doi.org/10.1080/1071576031000061002)
- 18. Dieterich C, Puey A, Lin S, Lyn S, Swezey R, Furimsky A, Fairchild D, Mirsalis JC, Ng HH. 2009. Gene expression analysis reveals new possible mechanisms of vancomycin-induced nephrotoxicity and identifies gene markers candidates. Toxicol Sci 107:258-269. [https://doi.org/10.1093/toxsci/kfn203.](https://doi.org/10.1093/toxsci/kfn203)
- 19. Raza Z, Naureen Z. 2020. Melatonin ameliorates the drug induced nephrotoxicity: molecular insights. Nefrologia (Engl Ed) 40:12-25. [https://doi](https://doi.org/10.1016/j.nefro.2019.06.009) [.org/10.1016/j.nefro.2019.06.009.](https://doi.org/10.1016/j.nefro.2019.06.009)
- 20. Arimura Y, Yano T, Hirano M, Sakamoto Y, Egashira N, Oishi R. 2012. Mitochondrial superoxide production contributes to vancomycin-induced renal tubular cell apoptosis. Free Radic Biol Med 52:1865–1873. [https://doi](https://doi.org/10.1016/j.freeradbiomed.2012.02.038) [.org/10.1016/j.freeradbiomed.2012.02.038](https://doi.org/10.1016/j.freeradbiomed.2012.02.038).
- 21. Soltani R, Khorvash F, Meidani M, Badri S, Alaei S, Taheri S. 2020. Vitamin E in the prevention of vancomycin-induced nephrotoxicity. Res Pharma Sci 15:137–143. [https://doi.org/10.4103/1735-5362.283813.](https://doi.org/10.4103/1735-5362.283813)
- 22. Ocak S, Gorur S, Hakverdi S, Celik S, Erdogan S. 2007. Protective effects of caffeic acid phenethyl ester, vitamin C, vitamin E and N-acetylcysteine on vancomycin-induced nephrotoxicity in rats. Basic Clin Pharmacol Toxicol 100:328–333. [https://doi.org/10.1111/j.1742-7843.2007.00051.x.](https://doi.org/10.1111/j.1742-7843.2007.00051.x)
- 23. Ahmida MH. 2012. Protective role of curcumin in nephrotoxic oxidative damage induced by vancomycin in rats. Exp Toxicol Pathol 64:149–153. [https://doi.org/10.1016/j.etp.2010.07.010.](https://doi.org/10.1016/j.etp.2010.07.010)
- 24. Reiter RJ, Tan D-X, Mayo JC, Sainz RM, Leon J, Czarnocki Z. 2003. Melatonin as an antioxidant: biochemical mechanisms and pathophysiological implications in humans. Acta Biochim Pol 50:1129–1146. [https://doi.org/](https://doi.org/10.18388/abp.2003_3637) [10.18388/abp.2003_3637.](https://doi.org/10.18388/abp.2003_3637)
- 25. Rodriguez C, Mayo JC, Sainz RM, Antolín I, Herrera F, Martín V, Reiter RJ. 2004. Regulation of antioxidant enzymes: a significant role for melatonin. J Pineal Res 36:1–9. [https://doi.org/10.1046/j.1600-079x.2003.00092.x.](https://doi.org/10.1046/j.1600-079x.2003.00092.x)
- 26. Tan D-X, Reiter RJ, Manchester LC, Yan M-T, El-Sawi M, Sainz RM, Mayo JC, Kohen R, Allegra M, Hardeland R. 2002. Chemical and physical properties and potential mechanisms: melatonin as a broad spectrum antioxidant and free radical scavenger. Curr Top Med Chem 2:181–197. [https://doi](https://doi.org/10.2174/1568026023394443) [.org/10.2174/1568026023394443](https://doi.org/10.2174/1568026023394443).
- 27. Srinivasan V, Spence DW, Pandi-Perumal SR, Brown GM, Cardinali DP. 2011. Melatonin in mitochondrial dysfunction and related disorders. Int J Alzheimers Dis 2011:326320. [https://doi.org/10.4061/2011/326320.](https://doi.org/10.4061/2011/326320)
- 28. Hara M, Yoshida M, Nishijima H, Yokosuka M, Iigo M, Ohtani-Kaneko R, Shimada A, Hasegawa T, Akama Y, Hirata K. 2001. Melatonin, a pineal secretory product with antioxidant properties, protects against cisplatininduced nephrotoxicity in rats. J Pineal Res 30:129-138. [https://doi.org/](https://doi.org/10.1034/j.1600-079x.2001.300301.x) [10.1034/j.1600-079x.2001.300301.x](https://doi.org/10.1034/j.1600-079x.2001.300301.x).
- 29. Cheng M-C, Wu T-H, Huang L-T, Tain Y-L. 2014. Renoprotective effects of melatonin in young spontaneously hypertensive rats with L-NAME. Pediatr Neonatol 55:189–195. [https://doi.org/10.1016/j.pedneo.2013.09.005.](https://doi.org/10.1016/j.pedneo.2013.09.005)
- 30. Stoianovici R, Brunetti L, Adams CD. 2019. Comparison of melatonin and zolpidem for sleep in an academic community hospital: an analysis of patient perception and inpatient outcomes. J Pharm Pract 34:44–50. <https://doi.org/10.1177/0897190019851888>.
- 31. Navalkele B, Pogue JM, Karino S, Nishan B, Salim M, Solanki S, Pervaiz A, Tashtoush N, Shaikh H, Koppula S, Koons J, Hussain T, Perry W, Evans R, Martin ET, Mynatt RP, Murray KP, Rybak MJ, Kaye KS. 2017. Risk of acute kidney injury in patients on concomitant vancomycin and piperacillintazobactam compared to those on vancomycin and cefepime. Clin Infect Dis 64:116–123. <https://doi.org/10.1093/cid/ciw709>.
- 32. Rutter WC, Cox JN, Martin CA, Burgess DR, Burgess DS. 2017. Nephrotoxicity during vancomycin therapy in combination with piperacillin-tazobactam or cefepime. Antimicrob Agents Chemother 61:e00314-17. [https://](https://doi.org/10.1128/AAC.00314-17) [doi.org/10.1128/AAC.00314-17.](https://doi.org/10.1128/AAC.00314-17)
- 33. Luther MK, Timbrook TT, Caffrey AR, Dosa D, Lodise TP, LaPlante KL. 2018. Vancomycin plus piperacillin-tazobactam and acute kidney injury in adults: a systematic review and meta-analysis. Crit Care Med 46:12–20. <https://doi.org/10.1097/CCM.0000000000002769>.
- 34. Qu S, Dai C, Guo H, Wang C, Hao Z, Tang Q, Wang H, Zhang Y. 2019. Rutin attenuates vancomycin-induced renal tubular cell apoptosis via suppression of apoptosis, mitochondrial dysfunction, and oxidative stress. Phytother Res 33:2056–2063. [https://doi.org/10.1002/ptr.6391.](https://doi.org/10.1002/ptr.6391)
- 35. Agil A, Chayah M, Visiedo L, Navarro-Alarcon M, Rodríguez Ferrer JM, Tassi M, Reiter RJ, Fernández-Vázquez G. 2020. Melatonin improves mitochondrial dynamics and function in the kidney of Zücker diabetic fatty rats. J Clin Med 9:2916. [https://doi.org/10.3390/jcm9092916.](https://doi.org/10.3390/jcm9092916)
- 36. Ghaznavi H, Mehrzadi S, Dormanesh B, Tabatabaei SMTH, Vahedi H, Hosseinzadeh A, Pazoki-Toroudi H, Rashidian A. 2016. Comparison of the protective effects of melatonin and silymarin against gentamicin-induced nephrotoxicity in rats. J Evid Based Complementary Altern Med 21: NP49–NP55. <https://doi.org/10.1177/2156587215621672>.
- 37. Wang HE, Muntner P, Chertow GM, Warnock DG. 2012. Acute kidney injury and mortality in hospitalized patients. Am J Nephrol 35:349–355. <https://doi.org/10.1159/000337487>.
- 38. Bedford M, Stevens PE, Wheeler TWK, Farmer CKT. 2014. What is the real impact of acute kidney injury? BMC Nephrol 15:95. [https://doi.org/10](https://doi.org/10.1186/1471-2369-15-95) [.1186/1471-2369-15-95.](https://doi.org/10.1186/1471-2369-15-95)
- 39. Chertow GM, Burdick E, Honour M, Bonventre JV, Bates DW. 2005. Acute kidney injury, mortality, length of stay, and costs in hospitalized patients. J Am Soc Nephrol 16:3365–3370. [https://doi.org/10.1681/ASN.2004090740.](https://doi.org/10.1681/ASN.2004090740)
- 40. Silver SA, Chertow GM. 2017. The economic consequences of acute kidney injury. Nephron 137:297–301. <https://doi.org/10.1159/000475607>.
- 41. Kovács J, Brodner W, Kirchlechner V, Arif T, Waldhauser F. 2000. Measurement of urinary melatonin: a useful tool for monitoring serum melatonin after its oral administration. J Clin Endocrinol Metab 85:666–670. [https://](https://doi.org/10.1210/jcem.85.2.6349) doi.org/10.1210/jcem.85.2.6349.
- 42. Stacchiotti A, Favero G, Giugno L, Lavazza A, Reiter RJ, Rodella LF, Rezzani R. 2014. Mitochondrial and metabolic dysfunction in renal convoluted tubules of obese mice: protective role of melatonin. PLoS One 9:e111141. <https://doi.org/10.1371/journal.pone.0111141>.
- 43. Kumar KV, Naidu MU, Shifow AA, Prayag A, Ratnakar KS. 1999. Melatonin: an antioxidant protects against cyclosporine-induced nephrotoxicity. Transplantation 67:1065–1068. <https://doi.org/10.1097/00007890-199904150-00022>.
- 44. Stacchiotti A, Lavazza A, Rezzani R, Bianchi R. 2002. Cyclosporine Ainduced kidney alterations are limited by melatonin in rats: an electron microscope study. Ultrastruct Pathol 26:81–87. [https://doi.org/10.1080/](https://doi.org/10.1080/01913120252959254) [01913120252959254.](https://doi.org/10.1080/01913120252959254)
- 45. Ali S, Qaisarani M, Farhat K, Waheed A. 2018. Study of preventive effect of melatonin on high dose vancomycin induced nephrotoxicity in rabbits. Pak Armed Forces Med J 68:1625–1629.
- 46. Nair AB, Jacob S. 2016. A simple practice guide for dose conversion between animals and human. J Basic Clin Pharm 7:27–31. [https://doi.org/](https://doi.org/10.4103/0976-0105.177703) [10.4103/0976-0105.177703](https://doi.org/10.4103/0976-0105.177703).
- 47. Savoca A, Manca D. 2019. Physiologically-based pharmacokinetic simulations in pharmacotherapy: selection of the optimal administration route for exogenous melatonin. ADMET DMPK 7:44–59. <https://doi.org/10.5599/admet.625>.
- 48. Peralta CA, Katz R, DeBoer I, Ix J, Sarnak M, Kramer H, Siscovick D, Shea S, Szklo M, Shlipak M. 2011. Racial and ethnic differences in kidney function decline among persons without chronic kidney disease. J Am Soc Nephrol 22:1327–1334. [https://doi.org/10.1681/ASN.2010090960.](https://doi.org/10.1681/ASN.2010090960)
- 49. Buelga DS, del Mar Fernandez de Gatta M, Herrera EV, Dominguez-Gil A, García MJ. 2005. Population pharmacokinetic analysis of vancomycin in patients with hematological malignancies. Antimicrob Agents Chemother 49:4934–4941. [https://doi.org/10.1128/AAC.49.12.4934-4941](https://doi.org/10.1128/AAC.49.12.4934-4941.2005) [.2005.](https://doi.org/10.1128/AAC.49.12.4934-4941.2005)
- 50. Lopes JA, Jorge S. 2013. The RIFLE and AKIN classifications for acute kidney injury: a critical and comprehensive review. Clin Kidney J 6:8–14. [https://doi.org/10.1093/ckj/sfs160.](https://doi.org/10.1093/ckj/sfs160)
- 51. Minejima E, Choi J, Beringer P, Lou M, Tse E, Wong-Beringer A. 2011. Applying new diagnostic criteria for acute kidney injury to facilitate early identification of nephrotoxicity in vancomycin-treated patients. Antimicrob Agents Chemother 55:3278–3283. [https://doi.org/10.1128/AAC.00173-11.](https://doi.org/10.1128/AAC.00173-11)
- 52. Mehta RL, Kellum JA, Shah SV, Molitoris BA, Ronco C, Warnock DG, Levin A, Acute Kidney Injury Network. 2007. Acute Kidney Injury Network: report of an initiative to improve outcomes in acute kidney injury. Crit Care 11:R31. [https://doi.org/10.1186/cc5713.](https://doi.org/10.1186/cc5713)