



Deep grey matter injury in multiple sclerosis: a NAIMS consensus statement

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Although multiple sclerosis has traditionally been considered a white matter disease, extensive research documents the presence and importance of grey matter injury including cortical and deep regions. The deep grey matter exhibits a broad range of pathology and is uniquely suited to study the mechanisms and clinical relevance of tissue injury in multiple sclerosis using magnetic resonance techniques. Deep grey matter injury has been associated with clinical and cognitive disability.

Recently, MRI characterization of deep grey matter properties, such as thalamic volume, have been tested as potential clinical trial end points associated with neurodegenerative aspects of multiple sclerosis. Given this emerging area of interest and its potential clinical trial relevance, the North American Imaging in Multiple Sclerosis (NAIMS) Cooperative held a workshop and reached consensus on imaging topics related to deep grey matter.

Herein, we review current knowledge regarding deep grey matter injury in multiple sclerosis from an imaging perspective, including insights from histopathology, image acquisition and post-processing for deep grey matter. We discuss the clinical relevance of deep grey matter injury and specific regions of interest within the deep grey matter. We highlight unanswered questions and propose future directions, with the aim of focusing research priorities towards better methods, analysis, and interpretation of results.

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Abbreviations: DGM = deep grey matter; MR = magnetic resonance

Introduction

Multiple sclerosis has traditionally been viewed as a white matter disease, with focal lesions resulting in neuronal injury and tissue destruction.^{1,2} However, detailed neuropathological examination also reveals extensive pathology in cortical and deep grey matter (DGM).³ While the mechanisms of DGM injury vary across structures, the net effect of this pathology can be examined *in vivo* using MRI. Volume loss/atrophy of the thalamus and other DGM structures has been well-documented in multiple sclerosis using MRI and is clinically relevant.^{4–9} The precise mechanisms of DGM loss remain to be fully elucidated and likely represent a complex interplay between the various aspects of pathology in multiple sclerosis.

Conventional MRI can be used to measure DGM volume and lesions. However, DGM lesions are best visualized at 7 T,^{10,11} are more difficult to visualize at conventional field strengths, and have even been considered a red flag for a diagnosis of multiple sclerosis.¹² Advanced techniques, such as functional MRI (fMRI), diffusion tensor MRI, relaxometry and magnetic resonance (MR) spectroscopy, can quantify additional changes in DGM structures.^{13–15} With recent advances in imaging, there is an opportunity to satisfy an unmet need to fully characterize DGM injury in multiple sclerosis and develop measures for use in clinical care and research. A better understanding of the mechanisms of DGM injury will provide fundamental knowledge about the pathophysiology of multiple sclerosis and may enable more efficient clinical trial design for neuroprotective therapies.

Materials and methods

The North American Imaging in Multiple Sclerosis Cooperative (NAIMS) initially met to discuss DGM injury in San Diego, California in February 2018. Topics included: pathological mechanisms of DGM injury in multiple sclerosis, measurement of DGM volume,

clinical relevance and regions of interest, and DGM metrics as clinical trial outcomes. Consensus points were drafted after completion of each topic presentations with input from all attendees. Consensus opinions were presented to the group at the completion of the meeting, and these were further refined by meeting presenters during teleconferences conducted after the meeting. Finally, all authors approved the final version of the manuscript. The goals of this paper are to summarize the proceedings, update and review the current understanding of DGM imaging with a focus on lesions and atrophy based on conventional MRI sequences, and provide recommendations for future research.

Pathological mechanisms of deep grey matter injury in multiple sclerosis

Deep grey matter lesions

Focal DGM lesions have been well-documented in multiple sclerosis and differ quantitatively and qualitatively from white matter lesions.⁵ Focal lesions occur in all DGM nuclei but are more common in the caudate, thalamus, and hypothalamus (Table 1).¹⁶ DGM lesions are characterized by demyelination with varying degrees of inflammatory changes.¹⁶ Similar to white matter lesions, DGM lesions can be classified into 'active', 'chronic active', and 'chronic inactive'.^{16,33,34} Active DGM lesions show increased perivascular cuffs and lymphocytic infiltrates.¹⁶ Chronic active lesions are characterized by a rim of activated microglia/macrophages, and are common in DGM.¹⁶ When compared to white matter lesions (more inflammation) and cortical lesions (less inflammation), DGM lesions have an intermediate inflammatory pattern.¹⁶

The most extensively studied DGM structure in multiple sclerosis is the thalamus. Focal thalamic lesions can be identified in over two-thirds of multiple sclerosis cases using microscopy and high-field MRI.^{10,11} Pathologically, focal thalamic lesions exhibit

Table 1 Histopathological and MRI findings in DGM and related grey matter regions in multiple sclerosis

DGM Structure	Histopathological findings						MRI findings		
	Lesions	Normal-appearing tissue					Early volume loss ^{19–21}	DGM lesions MRI-visible ^{22–24}	Iron accumulation ^{25–27}
		Focal lesions present ^{16–18}	Diffuse inflammatory infiltrate ^{16,17}	Activated microglia ^{16–18}	Iron deposition ^{16,*}	Neuronal loss ^{16,17}			
Thalamus ^{16–20,22}	+++	+	+	+	+	+	+++	+++	+
Caudate ^{16,17,20,23}	+++	+	+	+	+	+	++	+	+
Putamen ^{16,17,19,20,23}	++	+	+	+	+	+	+++	+	+
Globus pallidus ^{16,17}	++	+	+	+	+	+	NA	NA	+
Substantia nigra ¹⁷	+	+	NA	NA	NA	NA	NA	NA	NA
Amygdala ¹⁷	+	+	NA	NA	NA	NA	NA	NA	NA
Hypothalamus ^{17,28}	+++	+	+	NA	+	NA	NA	NA	NA
Hippocampus ^{29–32}	+++	NA	+	NA	+	NA	+	+++	NA

NA = not available.

*Did not reach statistical significance.

neuronal loss,¹⁷ demyelination, and axonal transection.^{35,36} MRI and histopathological studies have identified two thalamic lesion types: perivascular lesions, which are typically ovoid in shape, and subependymal lesions, which are thin bands of demyelination that line the third ventricle¹⁸ (Fig. 1). The presence of ovoid, perivascular, thalamic lesions shows conflicting results regarding correlation to cortical lesions.^{10,11} The appearance of subependymal thalamic lesions resembles that of subpial cortical lesions, and close proximity to the ventricle suggest a soluble or diffusible factor present in the CSF, perhaps with a role of inflammatory cells.^{37,38} The exact reasons for the variability in lesion frequency across DGM structures is unknown, but may relate to myelin content, contact with CSF spaces, and/or vascular distribution.

Non-lesional pathology in deep grey matter

DGM volume loss occurs early in multiple sclerosis and precedes measurable whole brain volume loss.^{8,19,39–41} The histopathological changes observed in non-lesional DGM may help explain this early susceptibility. Neuronal density in normal appearing DGM is reduced by as much as 33%¹⁷ compared to controls and could be explained by several mechanisms. Retrograde and anterograde degeneration due to the focal axonal transection that occurs within white matter lesions are probably major mechanisms of DGM neuronal loss.^{42–46} Trans-synaptic degeneration, which has been well-documented with hippocampal demyelination but has also been described in the visual system (which runs through the thalamus),^{47,48} may also play a role. Finally, deafferentation-induced neuronal loss, as described in the cortex, may also occur in DGM.⁴⁹ The cascade of white matter pathology resulting in axonal, neuronal, and synaptic changes suggests that early neurodegeneration in DGM may be a secondary phenomenon; however, a primary mechanism cannot be excluded (Fig. 2).

Iron accumulation is a proposed mechanism for DGM injury. DGM contains high iron content, which increases with age in controls.⁵⁰ In multiple sclerosis DGM, iron accumulation has been demonstrated better *in vivo* using iron-sensitive MRI sequences⁵¹ than on pathology.¹⁶ Iron accumulation may lead to oxidative stress, cellular damage, and neuronal injury.⁵² Iron release from injured oligodendrocytes results in higher turnover to microglia, which may be heightened in DGM structures and may drive more chronic inflammatory responses.⁴³ Susceptibility measures from the thalamus are associated with disability⁵³ and may differentiate

relapsing remitting multiple sclerosis from primary progressive multiple sclerosis.⁵⁴ Increased iron concentration may be the result of atrophy in the setting of a fixed iron content.⁵⁵ In thalamus, some studies have shown increased T₂*-weighted signal, which may represent loss of iron^{27,56} but may also be explained by loss of myelin or inflammation. Overall, the exact role of iron in thalamus is not clear and quantitative susceptibility mapping (QSM) may help resolve these questions. QSM has already been explored in DGM in multiple sclerosis.^{43,57,58}

Future directions

Larger and more detailed pathologic studies are needed to elucidate the exact cellular and subcellular substrates of DGM tissue loss in multiple sclerosis. Single cell genomics to further characterize DGM cell types may shed further light on changes secondary to afferent/efferent tracts with lesions or selective vulnerability of various thalamic subregions.

Measurement of deep grey matter injury

MRI acquisition

Recent advances in pulse sequences, increased magnetic field strength, and post-processing algorithms have improved the accuracy of MRI-based DGM volume measurements.⁵⁹ Several MRI contrasts can be used to identify DGM structures (Table 2), including T₁-weighted imaging,⁶⁰ T₂-weighted imaging,⁶² diffusion-weighted imaging,⁷⁰ magnetization transfer imaging,^{71,73} R₂* relaxation, and T₂*-weighted imaging/QSM.^{11,56–58} Gadolinium accumulation may preferentially affect DGM structures^{74,75} and may affect analysis of pixel intensities on T₁/T₂-weighted images and values of T₂*, R₂*, quantitative susceptibility mapping.

MRI analysis

DGM segmentation methods can be either atlas-based, algorithm-based, learning-based, or hybrid.⁷⁶ Atlas-based approaches generated using histology or MRI have been the most widely used and include the creation of anatomical labels, which are non-linearly warped to new data.⁷⁷ Atlases from several MR contrasts with specific sensitivity to different DGM structures are available. Atlas-based packages using probabilistic algorithms for segmentation of DGM structures are widely used and include FMRIB's

A Thalamic Lesions

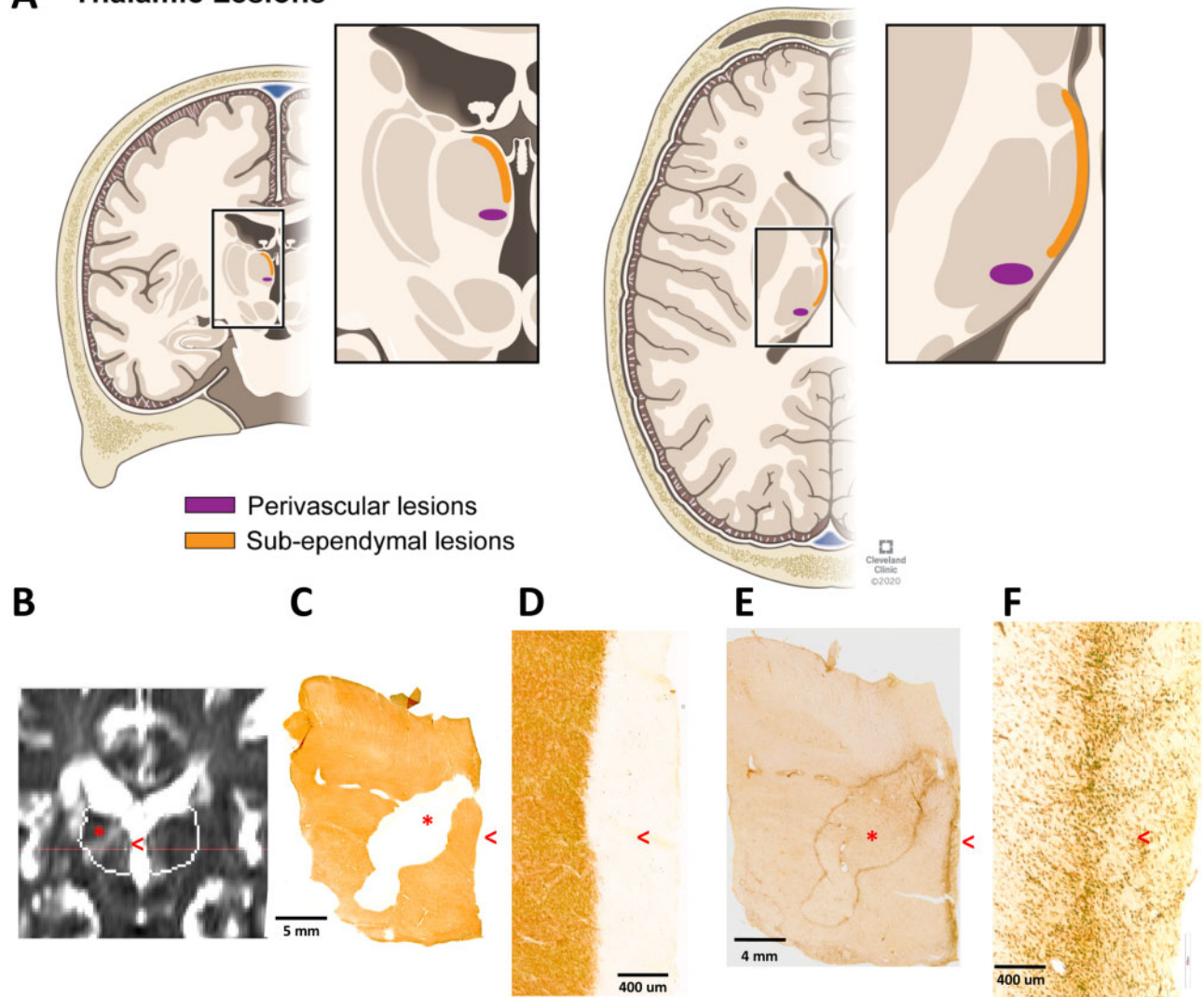


Figure 1 Demonstration of thalamic demyelinating lesion types. (A) Thalamic lesions are represented in coronal (left) and axial (right) planes. Lesions can either border the lateral and third ventricles (termed subependymal lesions, in orange) or have an ovoid appearance around blood vessels (termed perivascular, in purple). (B) Thalamic lesions (perivascular labelled with an asterisk and subependymal with a red open arrowhead) on T₂-weighted coronal 3 T MRI from a post-mortem multiple sclerosis case, and matching histological images from the same case (C–F), highlighting that subependymal lesions are more difficult to visualize. Myelin proteolipid protein immunohistochemistry demonstrates demyelination in perivascular and subependymal lesions (low magnification in C; higher magnification of subependymal lesion in D). Activated microglia/macrophages (MHC class II) immunohistochemistry demonstrating a chronic-active perivascular lesion and a rim at the border of the subependymal lesion (low magnification in E, higher magnification in F). Panel A is reprinted with permission, Cleveland Clinic Center for Medical Art & Photography ©2020. All Rights Reserved.

Integrated Registration and Segmentation Tool (FIRST),⁷⁸ FreeSurfer,⁷⁹ and statistical parametric mapping (SPM).⁸⁰ These are primarily based on T₁-weighted images, but can be improved by multiple contrasts and MR modalities used simultaneously.⁵⁸ Segmentation of lesional multiple sclerosis tissue is an important step to avoid misclassification of gray matter.⁸¹ Integrated probabilistic methods with simultaneous lesion masking are available.⁸²

Several comparisons of FSL-FIRST, SPM, and FreeSurfer have been conducted. FSL-FIRST shows the highest correlations with cognitive measures.⁸³ A recent systematic review compared several automated segmentation techniques⁷⁶ in multiple sclerosis and showed that learning-based approaches achieved the highest DGM segmentation accuracy. Multi-atlas approaches and use of additional MRI contrasts (QSM, T₂^{*}, R₂^{*}) improved results. These

techniques can be used in learning-based approaches for training datasets to establish improved measures of ground truth.

Segmentation of thalamic subregions has been achieved using ultra-high field MRI in combination with multicontrast atlases. Data thus far suggest that certain subregions may show greater volume loss, including the antero-ventral, pulvinar, and habenular regions.⁶¹ Using probabilistic connectivity at 3 T, differences in thalamic subregions have been demonstrated between multiple sclerosis and neuromyelitis optica,⁸⁴ with subregion 5 (premotor connection) mean diffusivity being the best discriminator. Selective vulnerability of thalamic subregions may relate to lesion location, susceptibility of specific pathways, and neuronal factors. Lesions in projecting/receiving tracts may be preferentially affected,⁴⁶ as well as neurons located closer to the ventricular surface.⁸⁵

Mechanisms of Thalamic Neuronal Degeneration Pathology in Multiple Sclerosis

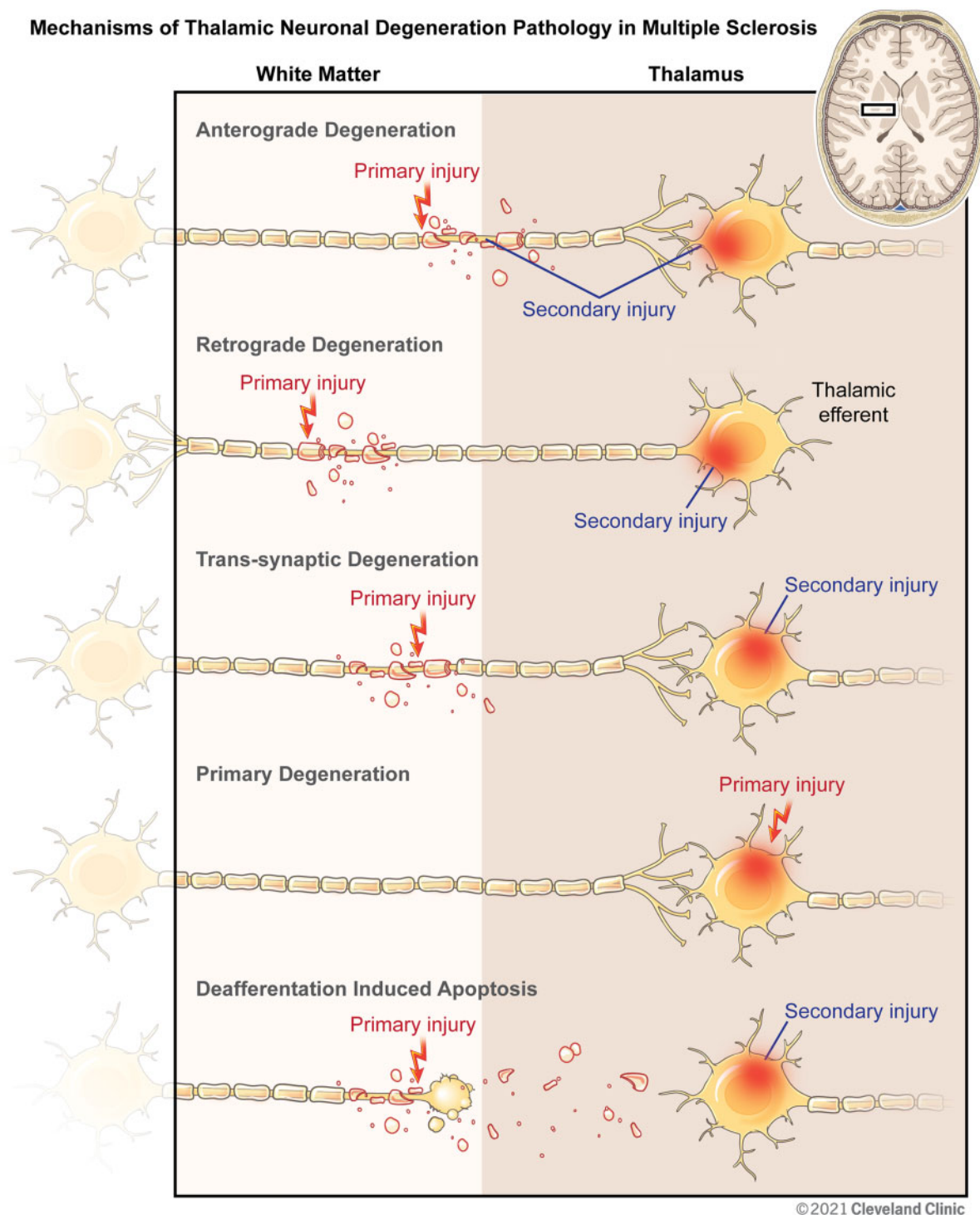


Figure 2 Mechanisms of thalamic neuronal degeneration pathology in multiple sclerosis. The figure shows representations of afferent/efferent neuronal cell bodies and axons in the thalamus and outflow/inflow tracts. Under 'Anterograde Degeneration', a focal white matter lesion in a thalamic afferent with secondary anterograde degeneration in the axon is depicted. Under 'Retrograde Degeneration', a focal white matter lesion in a thalamic efferent is depicted. Under 'Trans-synaptic Degeneration', secondary injury in the neuron as a result of degeneration of the axon with which it forms a synapse is demonstrated. Under 'Primary Neurodegeneration', neuronal injury independent of a direct connection to an axon is illustrated. Under 'Deafferentation Induced Apoptosis' we illustrate loss of neurons resulting as a consequence of axonal transection. Reprinted with permission, Cleveland Clinic Center for Medical Art & Photography ©2020. All Rights Reserved.

Machine learning approaches

Machine learning may offer several distinct advantages over current analysis and processing techniques. Machine learning and deep learning can incorporate several MRI contrasts and

measures to more efficiently explore tissue composition, structure, and function. When presented with labelled training data, machine learning algorithms can identify complex patterns from large datasets with a high number of variables, make generalizations from these learned patterns, rank the importance of

Table 2 MRI contrasts in DGM

Contrast	Utility	Acquisition comments
T ₁ -weighted ^{10,60,61}	Anatomical identification of boundaries and segmentation for volumetric assessments; potentially useful for lesion identification	High resolution images can be obtained with relatively short acquisition time; substantial experience in multicentre applications; thalamus may be difficult to segment due to high myelin content
T ₂ -weighted ⁶² or T ₂ -FLAIR	Identification of focal demyelinating lesions	Quick acquisition; substantial experience in multicentre applications
T ₁ relaxation ^{63,64}	Intrinsic changes in T ₂ signal, some sensitivity to iron Quantitative measure, can differentiate different DGM structures and subregions	Requires multiple acquisitions hence multicentre applications have been limited, but newer single-sequences approaches (e.g. MP2RAGE) may alleviate these drawbacks
T ₂ relaxation ^{64,65}	Quantitative measure, has some sensitivity to iron content in DGM, short component is particularly sensitive to myelin	Time consuming, with limited success in multicentre applications
T ₂ ^{*11,56,66,67}	Sensitive to iron and myelin content and blood vessels in DGM, also ideal to identify boundaries of iron-containing deep grey matter structures	High resolution can be attained with relatively quick acquisition times; multicentre application is relatively straightforward; less useful at 1.5 T
Double inversion recovery ⁶⁸	Sensitive to detection of DGM lesions	Feasible, and may also allow identification of few DGM lesions but sensitivity unknown
Quantitative susceptibility mapping ⁶⁹	Sensitive to iron, myelin and blood vessels; can differentiate diamagnetic and paramagnetic changes	Feasible, but post-processing required; less useful at 1.5 T
Diffusion weighted imaging ⁷⁰	Useful sensitive to microstructural	Typically lower resolution than other sequences; acquisition times are long for more complex biophysical models, which are not routinely available on commercial scanners; limited success in multicentre studies
Magnetization transfer imaging ^{71,72}	Sensitive to deep grey matter demyelination and useful for thalamic segmentation	Feasible clinical acquisition times; needs careful normalization for multicentre application

variables, and then use this information to make predictions on new data.

From an image-processing standpoint, deep learning may improve the accuracy of image segmentation in multiple sclerosis,^{86,87} and importantly, can reduce image heterogeneity across studies. Deep learning methods can normalize image intensity, resolution, and contrast across time points and across subjects.⁸⁸ Supervised machine learning methods using random forest classification of volume, thickness, surface area of cortical grey matter regions, and volumes of DGM nuclei, distinguished relapsing remitting multiple sclerosis from neuromyelitis optica with an accuracy of 74%.⁸⁹ Further development of supervised machine learning and unsupervised deep learning approaches may improve analysis of DGM structures to identify optimal regions of interest.

Future directions

Significant progress has been made in DGM segmentation. Use of multimodal approaches, multi-atlas approaches, subregional approaches, and machine learning may significantly improve DGM segmentation and its utility. Pulse sequences that generate quantitative data should take into consideration the local contrast generated for each specific structure and the resolution needed. Creation of data repositories and imaging toolboxes as shared resources will advance discovery. To enhance real-world utility, segmentation would ideally use pulse sequences that are widely available or postprocessing algorithms that are robust to different input sequences. Finally, the long-term goal of these measures is clinical translation, which requires multicentre validation and determination of clinical meaningfulness.

Clinical relevance of deep grey matter injury

Thalamus

Thalamic volume loss is observed on MRI in the earliest identifiable phases of multiple sclerosis, including pediatric multiple sclerosis,^{90,91} clinically isolated syndrome (CIS),^{40,41} and radiologically isolated syndrome,¹⁹ whereas whole brain volume and total grey matter volume may still be preserved. Thalamic volume declines consistently as a function of disease⁷ and, correlates with clinical end points, including cognition,^{6,7,92,93} and provides feasible sample sizes as a primary end point for clinical trials.⁷ Thalamic volume was shown to be modifiable in several recent randomized, placebo-controlled trials (see below). Given its high sensitivity in early disease and correlation with cognition, thalamic volume may be particularly useful in studies targeting cognition, or younger patients early in the disease.

Impact of normal ageing on deep grey matter measurements

Limited data exist regarding the impact of normal ageing on grey matter structures in multiple sclerosis. Whole and regional brain volumes can follow a linear or non-linear trajectory of decline,^{94–96} suggesting that the contribution of normal ageing to brain atrophy in multiple sclerosis may not be constant.

In a recent analysis, normal ageing was shown to contribute to whole brain and thalamic atrophy, whereas it did not in the putamen and caudate.²⁰ Most atrophy observed in whole brain and thalamus in early adulthood was multiple sclerosis-related, and by age 60, most of the atrophy was primarily attributable to normal ageing. The lack of ageing effect in the caudate and putamen

suggests DGM atrophy may differ across structures, an area of high interest for future studies.

Deep grey matter as an outcome in clinical trials

While the effects of disease-modifying therapies on whole brain volume in multiple sclerosis⁹⁷ have been extensively described, effects of disease-modifying therapies on DGM structures are only beginning to be investigated. Thalamic volume loss was reduced by ozanimod and is the first example where thalamic volume was reported in the primary analysis of a phase 3 study in multiple sclerosis.^{98,99} Thalamic volume loss was slower in laquinomod,¹⁰⁰ fingolimod,¹⁰¹ and ibudilast¹⁰² treated patients compared to placebo. In a multivariate retrospective analysis of a small, non-randomized cohort, natalizumab and rituximab were associated with a slower rate of thalamic and putaminal atrophy compared to interferon-beta and glatiramer acetate, whereas whole-brain volume decline did not differ between the two groups.¹⁰³ Exploring the differential effect of treatments on DGM may provide insight into specific pathways of injury related to these structures¹⁰¹; however, the feasibility of advanced MRI sequences must be considered for trial application (Table 2). It is likely that DGM MRI end points will be incorporated in future clinical trials. Tailoring the MRI end point to the patient population or clinical end point being studied could reduce sample sizes and improve trial efficiency.

Incorporating deep grey matter measurements into clinical practice

Incorporating brain volumetrics, including DGM, into clinical practice has proven challenging for several reasons, which have been reviewed.¹⁰⁴ Developing image-processing algorithms that are robust to image heterogeneity would address many barriers. A gold standard for DGM measurements is needed, and the variability in each structure needs to be known and accounted for. Differences in acquisition, gradient distortion, intrascanner variability, movement, and scanner upgrades have been cited as sources of variability for brain volume.¹⁰⁵ Whether DGM measures are susceptible to these same confounders as well as the effects of pseudoatrophy (initial apparent brain volume loss associated with anti-inflammatory effect),^{106,107} in measures of total versus individual DGM structures, are outstanding questions. Statistical techniques to translate DGM measurements into clinically relevant outcomes at the individual level are needed, and will need to account for age, treatment status, and measurement error at a minimum.

Hippocampus

Although not considered a DGM region, many studies examining DGM include the hippocampus in the analysis and classification of DGM structures,¹⁰⁸ and also because imaging methods sensitive to DGM damage are used to assess the hippocampus. The hippocampus is composed of several subregions, each of which has a distinct function and susceptibility to pathology.¹⁰⁹ It plays an important role in memory, regulation of mood, and emotional response.¹¹⁰ Histopathological studies show extensive involvement of the hippocampus in multiple sclerosis, including widespread demyelination, neuronal loss, and synaptic loss.^{29–32} Using MRI, total hippocampal volume loss has been associated with cognitive impairment in multiple sclerosis.¹¹¹ Atrophy in the CA1 subregion is associated with deficits in verbal memory in early relapsing remitting multiple sclerosis,²¹ and atrophy in the CA3 subregion is associated with depression.¹¹² CA4 subregion/dentate gyrus may be the earliest subfield with volume loss compared to

healthy controls, and that CA4/dentate volume predicts atrophy in the CA1 subregion 1 year later,¹¹³ suggesting a ‘dynamic spread’ of pathology.

Total hippocampal volume, or CA1 and/or CA3 subregional volumes, could be useful to study cognitive dysfunction and depression in multiple sclerosis, which lack sensitive and specific MRI correlates. Moreover, because of the potential for structural and functional plasticity within the hippocampal network,¹¹⁴ the hippocampus may be an interesting target for cognitive rehabilitation and neuroprotection interventions. The effects of exercise¹¹⁵ and cognitive rehabilitation¹¹⁶ on the hippocampus have been studied in several populations, including multiple sclerosis. Exercise interventions preserve or even increase hippocampal volume or activity, potentially leading to improvements in memory.^{116,117}

Because of low MR contrast with surrounding tissues, the hippocampus remains a technically challenging region to segment, particularly at the subregion level. Multiple automated segmentation algorithms exist, with variable levels of agreement with manual segmentation.¹¹⁸ Areas of interest for future hippocampal work in multiple sclerosis include improving longitudinal volume measurements, validating automated subregion techniques, pursuing hippocampal connectivity studies, and exploring the potential to preserve or reverse hippocampal damage, memory loss, and mood impairments using rehabilitative strategies.

Deep grey matter atrophy to understand disease biology better

Most of the current literature in multiple sclerosis has examined whole-brain and DGM volumes as predictors of clinical worsening. However, brain atrophy represents irreversible tissue loss, which is precisely what disease-modifying therapies aim to prevent. Future studies may use atrophy as the outcome rather than the predictor, to study biologic mechanisms of volume loss. The DGM is particularly well-suited for such studies because of the breadth of multiple sclerosis pathology present, mixed white and GM structures, early disease impact, and the highly connected nature of several regions. Different structures may provide insights into specific pathogenic mechanisms. For example, the mechanism of thalamic atrophy likely differs from that of hippocampal or basal ganglia atrophy, with different contributions from white matter lesions, intrinsic DGM lesions, iron deposition, oxidative stress, primary degeneration, etc., within each structure.^{10,11,18,45,46} A long-term goal is to understand the contribution of mechanisms of injury within each structure, leading to more specific targets using imaging endpoints for specific biological pathways.

Conclusions

DGM injury in multiple sclerosis is common and results from a combination of focal lesions and changes in normal-appearing DGM, as well as upstream/downstream injury secondary to distant lesions. Significant improvement in DGM segmentation has resulted in ample evidence demonstrating the clinical relevance of DGM in multiple sclerosis. Improvements in acquisition, segmentation (including subregions), and analysis methods promise to refine DGM measures further (DGM lesional volume, thalamic volume, caudate volume, hippocampal volume) and enable future use in clinical trials, potentially even as primary outcome for phase 2 studies focusing on neuroprotection. DGM clinical diagnostic tools and biomarkers of therapeutic response will be

accelerated by shared resources including pulse sequences, imaging datasets, and analysis tools.

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References

1. Trapp BD, Peterson J, Ransohoff RM, Rudick R, Mörk S, Bö L. Axonal transection in the lesions of multiple sclerosis. *N Engl J Med*. 1998;338(5):278–285.
2. Kornek B, Lassmann H. Axonal pathology in multiple sclerosis. A historical note. *Brain Pathol*. 1999;9(4):651–656.
3. Kutzelnigg A, Lassmann H. Cortical lesions and brain atrophy in MS. *J Neurologic Sci*. 2005;233(1-2):55–59.
4. Bakshi R, Czarnecki D, Shaikh ZA. Brain MRI lesions and atrophy are related to depression in multiple sclerosis. *Neuroreport*. 2000;11(6):1153–1158.
5. Cifelli A, Arridge M, Jezard P, Esiri MM, Palace J, Matthews PM. Thalamic neurodegeneration in multiple sclerosis. *Ann Neurol*. 2002;52(5):650–653.
6. Houtchens MK, Benedict RHB, Killiany R, et al. Thalamic atrophy and cognition in multiple sclerosis. *Neurology*. 2007;69(12):1213–1223.
7. Azevedo CJ, Cen SY, Khadka S, et al. Thalamic atrophy in multiple sclerosis: A magnetic resonance imaging marker of neurodegeneration throughout disease. *Ann Neurol*. 2018;83(2):223–234.
8. Zivadinov R, Havrdova E, Bergsland N, et al. Thalamic atrophy is associated with development of clinically definite multiple sclerosis. *Radiology*. 2013;268(3):831–841.
9. Shiee N, Bazin PL, Zackowski KM, et al. Revisiting brain atrophy and its relationship to disability in multiple sclerosis. *PLoS One*. 2012;7(5):e37049.
10. Harrison DM, Oh J, Roy S, et al. Thalamic lesions in multiple sclerosis by 7T MRI: Clinical implications and relationship to cortical pathology. *Mult Scler*. 2015;21(9):1139–1150.
11. Mehndiratta A, Treaba CA, Barletta V, et al. Characterization of thalamic lesions and their correlates in multiple sclerosis by ultra-high-field MRI. *Mult Scler J*. 2021;27(5):674–683.
12. Miller DH, Weinshenker BG, Filippi M, et al. Differential diagnosis of suspected multiple sclerosis: A consensus approach. *Mult Scler*. 2008;14(9):1157–1174.
13. Cercignani M, Bozzali M, Iannucci G, Comi G, Filippi M. Magnetisation transfer ratio and mean diffusivity of normal

- appearing white and grey matter from patients with multiple sclerosis. *J Neurol Neurosurg Psychiatry*. 2001;70(3):311–317.
14. Reddy H, Narayanan S, Arnoutelis R, et al. Evidence for adaptive functional changes in the cerebral cortex with axonal injury from multiple sclerosis. *Brain*. 2000;123(11):2314–2320.
 15. Sarchielli P, Presciutti O, Tarducci R, et al. Localized (1)H magnetic resonance spectroscopy in mainly cortical grey matter of patients with multiple sclerosis. *J Neurol*. 2002;249(7):902–910.
 16. Haider L, Simeonidou C, Steinberger G, et al. Multiple sclerosis deep grey matter: The relation between demyelination, neurodegeneration, inflammation and iron. *J Neurol Neurosurg Psychiatry*. 2014;85(12):1386–1395.
 17. Vercellino M, Masera S, Lorenzatti M, et al. Demyelination, inflammation, and neurodegeneration in multiple sclerosis deep grey matter. *J Neuropathol Exp Neurol*. 2009;68(5):489–502.
 18. Mahajan KR, Nakamura K, Cohen JA, Trapp BD, Ontaneda D. Intrinsic and extrinsic mechanisms of thalamic pathology in multiple sclerosis. *Ann Neurol*. 2020;88(1):81–92.
 19. Azevedo CJ, Overton E, Khadka S, et al. Early CNS neurodegeneration in radiologically isolated syndrome. *Neurol Neuroimmunol Neuroinflamm*. 2015;2(3):e102.
 20. Azevedo CJ, Cen SY, Jaberzadeh A, Zheng L, Hauser SL, Pelletier D. Contribution of normal aging to brain atrophy in MS. *Neurol Neuroimmunol Neuroinflamm*. 2019;6(6):e616.
 21. Sicotte NL, Kern KC, Giesser BS, et al. Regional hippocampal atrophy in multiple sclerosis. *Brain*. 2008;131(Pt 4):1134–1141.
 22. Harrison DM, Roy S, Oh J, et al. Association of cortical lesion burden on 7-T magnetic resonance imaging with cognition and disability in multiple sclerosis. *JAMA Neurol*. 2015;72(9):1004–1012.
 23. Ruggieri S, Petracca M, Miller A, et al. Association of deep gray matter damage with cortical and spinal cord degeneration in primary progressive multiple sclerosis. *JAMA Neurol*. 2015;72(12):1466.
 24. Roosendaal SD, Moraal B, Pouwels PJW, et al. Accumulation of cortical lesions in MS: Relation with cognitive impairment. *Mult Scler*. 2009;15(6):708–714.
 25. Hopp K, Popescu BFG, McCrea RPE, et al. Brain iron detected by SWI high pass filtered phase calibrated with synchrotron X-ray fluorescence. *J Magn Reson Imaging*. 2010;31(6):1346–1354.
 26. Al-Radaideh AM, Wharton SJ, Lim S-Y, et al. Increased iron accumulation occurs in the earliest stages of demyelinating disease: An ultra-high field susceptibility mapping study in Clinically Isolated Syndrome. *Mult Scler*. 2013;19(7):896–903.
 27. Khalil M, Langkammer C, Pichler A, et al. Dynamics of brain iron levels in multiple sclerosis: A longitudinal 3T MRI study. *Neurology*. 2015;84(24):2396–2402.
 28. Huitinga I, De Groot CJ, Van der Valk P, Kamphorst W, Tilders FJ, Swaab DF. Hypothalamic lesions in multiple sclerosis. *J Neuropathol Exp Neurol*. 2001;60(12):1208–1218.
 29. Vercellino M, Plano F, Votta B, Mutani R, Giordana MT, Cavalla P. Grey matter pathology in multiple sclerosis. *J Neuropathol Exp Neurol*. 2005;64(12):1101–1107.
 30. Geurts JJ, Bo L, Roosendaal SD, et al. Extensive hippocampal demyelination in multiple sclerosis. *J Neuropathol Exp Neurol*. 2007;66(9):819–827.
 31. Papadopoulos D, Dukes S, Patel R, Nicholas R, Vora A, Reynolds R. Substantial archaeocortical atrophy and neuronal loss in multiple sclerosis. *Brain Pathol*. 2009;19(2):238–253.
 32. Dutta R, Chang A, Doud MK, et al. Demyelination causes synaptic alterations in hippocampi from multiple sclerosis patients. *Ann Neurol*. 2011;69(3):445–454.
 33. Dal Bianco A, Bradl M, Frischer J, Kutzelnigg A, Jellinger K, Lassmann H. Multiple sclerosis and Alzheimer's disease. *Ann Neurol*. 2008;63(2):174–183.
 34. Peterson JW, Bo L, Mork S, Chang A, Trapp BD. Transected neurites, apoptotic neurons, and reduced inflammation in cortical multiple sclerosis lesions. *Ann Neurol*. 2001;50(3):389–400.
 35. Kipp M, Wagenknecht N, Beyer C, Samer S, Wuerfel J, Nikoubashman O. Thalamus pathology in multiple sclerosis: From biology to clinical application. *Cell Mol Life Sci*. 2015;72(6):1127–1147.
 36. Minagar A, Barnett MH, Benedict RH, et al. The thalamus and multiple sclerosis: Modern views on pathologic, imaging, and clinical aspects. *Neurology*. 2013;80(2):210–219.
 37. Gilmore CP, Donaldson I, Bö L, et al. Regional variations in the extent and pattern of grey matter demyelination in multiple sclerosis: A comparison between the cerebral cortex, cerebellar cortex, deep grey matter nuclei and the spinal cord. *J Neurol Neurosurg Psychiatry*. 2009;80(2):182–187.
 38. Adams CWM, Abdulla YH, Torres EM, Poston RN. Periventricular lesions in multiple sclerosis: Their perivenous origin and relationship to granular ependymitis. *Neuropathol Appl Neurobiol*. 1987;13(2):141–152.
 39. Eshaghi A, Marinescu RV, Young AL, et al. Progression of regional grey matter atrophy in multiple sclerosis. *Brain*. 2018;141(6):1665–1677.
 40. Steckova T, Hlustik P, Sladkova V, Odstřil F, Mares J, Kanovsky P. Thalamic atrophy and cognitive impairment in clinically isolated syndrome and multiple sclerosis. *J Neurol Sci*. 2014;342(1-2):62–68.
 41. Henry RG, Shieh M, Okuda DT, Evangelista A, Gorno-Tempini ML, Pelletier D. Regional grey matter atrophy in clinically isolated syndromes at presentation. *J Neurol Neurosurg Psychiatry*. 2008;79(11):1236–1244.
 42. Nikić I, Merkle D, Sorbara C, et al. A reversible form of axon damage in experimental autoimmune encephalomyelitis and multiple sclerosis. *Nat Med*. 2011;17(4):495–499.
 43. Schweser F, Raffaini Duarte Martins AL, Hagemeyer J, et al. Mapping of thalamic magnetic susceptibility in multiple sclerosis indicates decreasing iron with disease duration: A proposed mechanistic relationship between inflammation and oligodendrocyte vitality. *Neuroimage*. 2018;167:438–452.
 44. Papadopoulou A, Gaetano L, Pfister A, et al. Damage of the lateral geniculate nucleus in MS. *Neurology*. 2019;92(19):e2240–e2249.
 45. Kolasinski J, Stagg CJ, Chance SA, et al. A combined post-mortem magnetic resonance imaging and quantitative histological study of multiple sclerosis pathology. *Brain*. 2012;135(Pt 10):2938–2951.
 46. Henry RG, Shieh M, Amirbekian B, Chung S, Okuda DT, Pelletier D. Connecting white matter injury and thalamic atrophy in clinically isolated syndromes. *J Neurol Sci*. 2009;282(1-2):61–66.
 47. Reich DS, Smith SA, Gordon-Lipkin EM, et al. Damage to the optic radiation in multiple sclerosis is associated with retinal injury and visual disability. *Arch Neurol*. 2009;66(8):998–1006.
 48. Al-Louzi O, Button J, Newsome SD, Calabresi PA, Saidha S. Retrograde trans-synaptic visual pathway degeneration in multiple sclerosis: A case series. *Mult Scler*. 2017;23(7):1035–1039.
 49. Koliatsos VE, Dawson TM, Kecojevic A, Zhou Y, Wang YF, Huang KX. Cortical interneurons become activated by deafferentation and instruct the apoptosis of pyramidal neurons. *Proc Natl Acad Sci U S A*. 2004;101(39):14264–14269.
 50. Hallgren B, Sourander P. The effect of age on the non-haemin iron in the human brain. *J Neurochem*. 1958;3(1):41–51.

51. Hammond KE, Metcalf M, Carvajal L, et al. Quantitative in vivo magnetic resonance imaging of multiple sclerosis at 7 Tesla with sensitivity to iron. *Ann Neurol*. 2008;64(6):707–713.
52. Zecca L, Youdim MBH, Riederer P, Connor JR, Crichton RR. Iron, brain ageing and neurodegenerative disorders. *Nat Rev Neurosci*. 2004;5(11):863–873.
53. Zivadinov R, Tavazzi E, Bergsland N, et al. Brain iron at quantitative MRI is associated with disability in multiple sclerosis. *Radiology*. 2018;289(2):487–496.
54. Burgetova A, Dusek P, Vaneckova M, et al. Thalamic iron differentiates primary-progressive and relapsing-remitting multiple sclerosis. *Am J Neuroradiol*. 2017;38(6):1079–1086.
55. Hernández-Torres E, Wiggermann V, Machan L, et al. Increased mean R2 in the deep gray matter of multiple sclerosis patients: Have we been measuring atrophy? *J Magn Reson Imaging*. 2019;50(1):201–208.
56. Louapre C, Govindarajan ST, Gianni C, et al. Heterogeneous pathological processes account for thalamic degeneration in multiple sclerosis: Insights from 7 T imaging. *Mult Scler J*. 2018;24(11):1433–1444.
57. Cobzas D, Sun H, Walsh AJ, Lebel RM, Blevins G, Wilman AH. Subcortical gray matter segmentation and voxel-based analysis using transverse relaxation and quantitative susceptibility mapping with application to multiple sclerosis. *J Magn Reson Imaging*. 2015;42(6):1601–1610.
58. Feng X, Deistung A, Dwyer MG, et al. An improved FSL-FIRST pipeline for subcortical gray matter segmentation to study abnormal brain anatomy using quantitative susceptibility mapping (QSM). *Magn Reson Imaging*. 2017;39:110–122.
59. Keller SS, Gerdes JS, Mohammadi S, et al. Volume estimation of the thalamus using freesurfer and stereology: Consistency between methods. *Neuroinformatics*. 2012;10(4):341–350.
60. Rocca MA, Comi G, Filippi M. The role of T1-weighted derived measures of neurodegeneration for assessing disability progression in multiple sclerosis. *Front Neurol*. 2017;8:433.
61. Planche V, Su JH, Mourmet S, et al. White-matter-nulled MPRAGE at 7T reveals thalamic lesions and atrophy of specific thalamic nuclei in multiple sclerosis. *Mult Scler J*. 2020;26(8):987–992.
62. Bakshi R, Benedict RH, Bermel RA, et al. T2 hypointensity in the deep gray matter of patients with multiple sclerosis: A quantitative magnetic resonance imaging study. *Arch Neurol*. 2002;59(1):62–68.
63. Steenwijk MD, Vrenken H, Jonkman LE, et al. High-resolution T1-relaxation time mapping displays subtle, clinically relevant, gray matter damage in long-standing multiple sclerosis. *Mult Scler*. 2016;22(10):1279–1288.
64. Traynor CR, Barker GJ, Crum WR, Williams SCR, Richardson MP. Segmentation of the thalamus in MRI based on T1 and T2. *Neuroimage*. 2011;56(3):939–950.
65. Hasan KM, Walimuni IS, Abid H, et al. Multimodal quantitative magnetic resonance imaging of thalamic development and aging across the human lifespan: Implications to neurodegeneration in multiple sclerosis. *J Neurosci*. 2011;31(46):16826–16832.
66. Khalil M, Langkammer C, Ropele S, et al. Determinants of brain iron in multiple sclerosis: A quantitative 3T MRI study. *Neurology*. 2011;77(18):1691–1697.
67. Yarnykh VL, Krutenkova EP, Aitmagambetova G, et al. Iron-insensitive quantitative assessment of subcortical gray matter demyelination in multiple sclerosis using the macromolecular proton fraction. *Am J Neuroradiol*. 2018;39(4):618–625.
68. Van De Pavert SHP, Muhlert N, Sethi V, et al. DIR-visible grey matter lesions and atrophy in multiple sclerosis: Partners in crime? *J Neurol Neurosurg Psychiatry*. 2016;87(5):461–467.
69. Fujiwara E, Kmech JA, Cobzas D, et al. Cognitive implications of deep gray matter iron in multiple sclerosis. *Am J Neuroradiol*. 2017;38(5):942–948.
70. Pfefferbaum A, Adalsteinsson E, Rohlfing T, Sullivan EV. Diffusion tensor imaging of deep gray matter brain structures: Effects of age and iron concentration. *Neurobiol Aging*. 2010;31(3):482–493.
71. Helms G, Draganski B, Frackowiak R, Ashburner J, Weiskopf N. Improved segmentation of deep brain grey matter structures using magnetization transfer (MT) parameter maps. *Neuroimage*. 2009;47(1):194–198.
72. Mallik S, Muhlert N, Samson RS, et al. Regional patterns of grey matter atrophy and magnetisation transfer ratio abnormalities in multiple sclerosis clinical subgroups: A voxel-based analysis study. *Mult Scler J*. 2015;21(4):423–432.
73. Gringel T, Schulz-Schaeffer W, Eloff E, Frölich A, Dechent P, Helms G. Optimized high-resolution mapping of magnetization transfer (MT) at 3 Tesla for direct visualization of substructures of the human thalamus in clinically feasible measurement time. *J Magn Reson Imaging*. 2009;29(6):1285–1292.
74. Kanda T, Ishii K, Kawaguchi H, Kitajima K, Takenaka D. High signal intensity in the dentate nucleus and globus pallidus on unenhanced T1-weighted MR images: Relationship with increasing cumulative dose of a gadolinium-based contrast material. *Radiology*. 2014;270(3):834–841.
75. McDonald RJ, Levine D, Weinreb J, et al. Gadolinium retention: A research roadmap from the 2018 NIH/ACR/RSNA workshop on gadolinium chelates. *Radiology*. 2018;289(2):517–534.
76. Pagnozzi AM, Fripp J, Rose SE. Quantifying deep grey matter atrophy using automated segmentation approaches: A systematic review of structural MRI studies. *Neuroimage*. 2019;201:116018.
77. Fischl B, Salat DH, Busa E, et al. Whole brain segmentation: Automated labeling of neuroanatomical structures in the human brain. *Neuron*. 2002;33(3):341–355.
78. Patenaude B, Smith SM, Kennedy DN, Jenkinson M. A Bayesian model of shape and appearance for subcortical brain segmentation. *Neuroimage*. 2011;56(3):907–922.
79. Fischl B. FreeSurfer. *Neuroimage*. 2012;62(2):774–781.
80. Ashburner J, Friston KJ. Unified segmentation. *Neuroimage*. 2005;26(3):839–851.
81. González-Villà S, Oliver A, Huo Y, Lladó X, Landman BA. Brain structure segmentation in the presence of multiple sclerosis lesions. *NeuroImage Clin*. 2019;22:101709.
82. Cerri S, Hoopes A, Greve D, Mühlau M, Van Leemput K. A Longitudinal Method for Simultaneous Whole-Brain and Lesion Segmentation in Multiple Sclerosis. In: Kia SM, Mohyud-Din H, Abdulkadir A, et al, eds. *Machine Learning in Clinical Neuroimaging and Radiogenomics in Neuro-oncology*. MLCN 2020, RNO-AI 2020. Lecture Notes in Computer Science; vol 12449. Springer, Cham; 2020:119–128.
83. Popescu V, Schoonheim MM, Versteeg A, et al. Grey matter atrophy in multiple sclerosis: Clinical interpretation depends on choice of analysis method. *PLoS One*. 2016;11(1):e0143942.
84. Liu Y, Duan Y, Huang J, et al. Multimodal quantitative MR imaging of the thalamus in multiple sclerosis and neuromyelitis optica1. *Radiology*. 2015;277(3):784–792.
85. Fadda G, Brown RA, Magliozzi R, et al.; Canadian Pediatric Demyelinating Disease Network. A surface-in gradient of thalamic damage evolves in pediatric multiple sclerosis. *Ann Neurol*. 2019;85(3):340–351.
86. Sweeney EM, Vogelstein JT, Cuzzocreo JL, et al. A comparison of supervised machine learning algorithms and feature

- vectors for MS lesion segmentation using multimodal structural MRI. *PLoS One*. 2014;9(4):e95753.
87. Brosch T, Tang LYW, Yoo Y, Li DKB, Traboulsee A, Tam R. Deep 3D convolutional encoder networks with shortcuts for multiscale feature integration applied to multiple sclerosis lesion segmentation. *IEEE Trans Med Imaging*. 2016;35(5):1229–1239.
 88. Dewey BE, Zhao C, Reinhold JC, et al. DeepHarmony: A deep learning approach to contrast harmonization across scanner changes. *Magn Reson Imaging*. 2019;64:160–170.
 89. Eshaghi A, Wottschel V, Cortese R, et al. Gray matter MRI differentiates neuromyelitis optica from multiple sclerosis using random forest. *Neurology*. 2016;87(23):2463–2470.
 90. Aubert-Broche B, Fonov V, Ghassemi R, et al. Regional brain atrophy in children with multiple sclerosis. *Neuroimage*. 2011;58(2):409–415.
 91. Mesaros S, Rocca MA, Absinta M, et al. Evidence of thalamic gray matter loss in pediatric multiple sclerosis. *Neurology*. 2008;70(13 Part 2):1107–1112.
 92. Zivadinov R, Bergsland N, Dolezal O, et al. Evolution of cortical and thalamus atrophy and disability progression in early relapsing-remitting MS during 5 years. *Am J Neuroradiol*. 2013;34(10):1931–1939.
 93. Eshaghi A, Prados F, Brownlee WJ, et al.; on behalf of the MAGNIMS study group. Deep gray matter volume loss drives disability worsening in multiple sclerosis. *Ann Neurol*. 2018;83(2):210–222.
 94. Fjell AM, Walhovd KB, Westlye LT, et al. When does brain aging accelerate? Dangers of quadratic fits in cross-sectional studies. *Neuroimage*. 2010;50(4):1376–1383.
 95. Schippling S, Ostwaldt AC, Suppa P, et al. Global and regional annual brain volume loss rates in physiological aging. *J Neurol*. 2017;264(3):520–528.
 96. Coupé P, Catheline G, Lanuza E, Manjón JV.; Alzheimer's Disease Neuroimaging Initiative. Towards a unified analysis of brain maturation and aging across the entire lifespan: A MRI analysis. *Hum Brain Mapp*. 2017;38(11):5501–5518.
 97. De Stefano N, Stromillo ML, Giorgio A, et al. Establishing pathological cut-offs of brain atrophy rates in multiple sclerosis. *J Neurol Neurosurg Psychiatry*. 2016;87(1):93–99.
 98. Comi G, Kappos L, Selmaj KW, et al. Safety and efficacy of ozanimod versus interferon beta-1a in relapsing multiple sclerosis (SUNBEAM): A multicentre, randomised, minimum 12-month, phase 3 trial. *Lancet Neurol*. 2019;18(11):1009–1020.
 99. Cohen JA, Comi G, Selmaj KW, et al. Safety and efficacy of ozanimod versus interferon beta-1a in relapsing multiple sclerosis (RADIANCE): A multicentre, randomised, 24-month, phase 3 trial. *Lancet Neurol*. 2019;18(11):1021–1033.
 100. Filippini G, Del Giovane C, Clerico M, et al. Treatment with disease-modifying drugs for people with a first clinical attack suggestive of multiple sclerosis. *Cochrane Database Syst Rev*. 2017;4:CD012200.
 101. Gaetano L, Häring DA, Radue EW, et al. Fingolimod effect on gray matter, thalamus, and white matter in patients with multiple sclerosis. *Neurology*. 2018;90(15):e1324–e1332.
 102. Fox RJ, Coffey CS, Cudkovic ME, et al. Design, rationale, and baseline characteristics of the randomized double-blind phase II clinical trial of ibudilast in progressive multiple sclerosis. *Contemp Clin Trials*. 2016;50:166–177.
 103. Sotirchos ES, Gonzalez-Caldito N, Dewey BE, et al. Effect of disease-modifying therapies on subcortical gray matter atrophy in multiple sclerosis. *Mult Scler J*. 2020;26(3):312–321.
 104. Azevedo CJ, Pelletier D. Whole-brain atrophy: Ready for implementation into clinical decision-making in multiple sclerosis? *Curr Opin Neurol*. 2016;29(3):237–242.
 105. Sastre-Garriga J, Pareto D, Battaglini M, et al.; MAGNIMS study group. MAGNIMS consensus recommendations on the use of brain and spinal cord atrophy measures in clinical practice. *Nat Rev Neurol*. 2020;16(3):171–182.
 106. Dwyer MG, Zivadinov R, Tao Y, et al. Immunological and short-term brain volume changes in relapsing forms of multiple sclerosis treated with interferon beta-1a subcutaneously three times weekly: an open-label two-arm trial. *BMC Neurol*. 2015;15:232.
 107. Fisher E, Nakamura K, Lee JC, You X, Sperling B, Rudick RA. Effect of intramuscular interferon beta-1a on gray matter atrophy in relapsing-remitting multiple sclerosis: a retrospective analysis. *Mult Scler*. 2016;22(5):668–676.
 108. Narvacan K, Treit S, Camicioli R, Martin W, Beaulieu C. Evolution of deep gray matter volume across the human lifespan. *Hum Brain Mapp*. 2017;38(8):3771–3790.
 109. Nunn J, Hodges H. Cognitive deficits induced by global cerebral ischaemia: Relationship to brain damage and reversal by transplants. *Behav Brain Res*. 1994;65(1):1–31.
 110. McDonald AJ, Mott DD. Functional neuroanatomy of amygdalohippocampal interconnections and their role in learning and memory. *J Neurosci Res*. 2017;95(3):797–820.
 111. Damjanovic D, Valsasina P, Rocca MA, et al. Hippocampal and deep gray matter nuclei atrophy is relevant for explaining cognitive impairment in MS: A multicenter study. *Am J Neuroradiol*. 2017;38(1):18–24.
 112. Gold SM, Kern KC, O'Connor MF, et al. Smaller cornu ammonis 2-3/dentate gyrus volumes and elevated cortisol in multiple sclerosis patients with depressive symptoms. *Biol Psychiatry*. 2010;68(6):553–559.
 113. Planche V, Koubiyr I, Romero JE, et al. Regional hippocampal vulnerability in early multiple sclerosis: Dynamic pathological spreading from dentate gyrus to CA1. *Hum Brain Mapp*. 2018;39(4):1814–1824.
 114. Rocca MA, Barkhof F, De Luca J, et al. The hippocampus in multiple sclerosis. *Lancet Neurol*. 2018;17(10):918–926.
 115. Firth J, Stubbs B, Vancampfort D, et al. Effect of aerobic exercise on hippocampal volume in humans: A systematic review and meta-analysis. *Neuroimage*. 2018;166:230–238.
 116. Hampstead BM, Stringer AY, Stilla RF, Giddens M, Sathian K. Mnemonic strategy training partially restores hippocampal activity in patients with mild cognitive impairment. *Hippocampus*. 2012;22(8):1652–1658.
 117. Erickson KI, Voss MW, Prakash RS, et al. Exercise training increases size of hippocampus and improves memory. *Proc Natl Acad Sci U S A*. 2011;108(7):3017–3022.
 118. Dill V, Franco AR, Pinho MS. Automated methods for hippocampus segmentation: The evolution and a review of the state of the art. *Neuroinformatics*. 2015;13(2):133–150.