



LncRNAs as Therapeutic Targets and Potential Biomarkers for Lipid-Related Diseases

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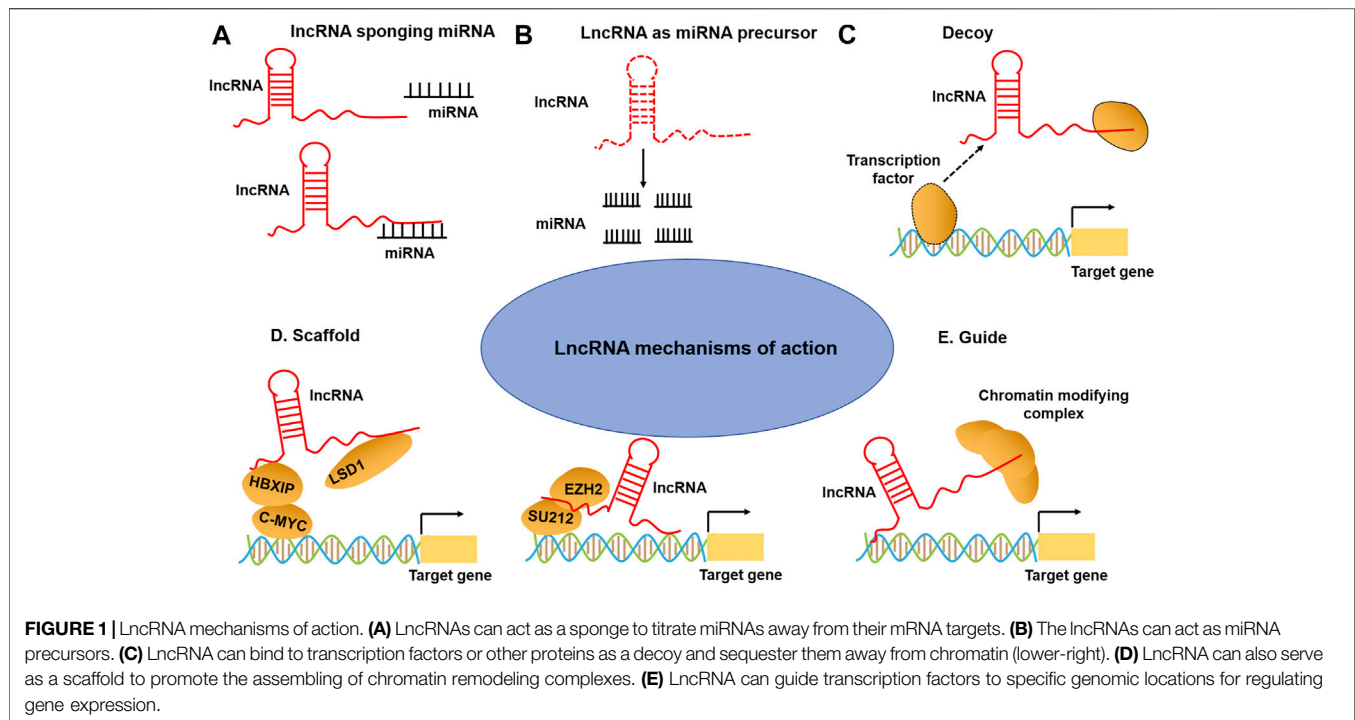
Lipid metabolism is an essential biological process involved in nutrient adjustment, hormone regulation, and lipid homeostasis. An irregular lifestyle and long-term nutrient overload can cause lipid-related diseases, including atherosclerosis, myocardial infarction (MI), obesity, and fatty liver diseases. Thus, novel tools for efficient diagnosis and treatment of dysfunctional lipid metabolism are urgently required. Furthermore, it is known that lncRNAs based regulation like sponging microRNAs (miRNAs) or serving as a reservoir for microRNAs play an essential role in the progression of lipid-related diseases. Accordingly, a better understanding of the regulatory roles of lncRNAs in lipid-related diseases would provide the basis for identifying potential biomarkers and therapeutic targets for lipid-related diseases. This review highlighted the latest advances on the potential biomarkers of lncRNAs in lipid-related diseases and summarised current knowledge on dysregulated lncRNAs and their potential molecular mechanisms. We have also provided novel insights into the underlying mechanisms of lncRNAs which might serve as potential biomarkers and therapeutic targets for lipid-related diseases. The information presented here may be useful for designing future studies and advancing investigations of lncRNAs as biomarkers for diagnosis, prognosis, and therapy of lipid-related diseases.

Keywords: lncRNAs, therapeutic targets, lipid metabolism, lipid-related diseases, biomarkers

INTRODUCTION

Lipid metabolism is an intricate and complex physiological process that is involved in the progression of lipid-related diseases (Li et al., 2017). Importantly, since modern society is associated with irregular lifestyle patterns and long-term nutrient overload, severe lipid metabolism disorders and lipid accumulation have become commonplace (Liu and Ding, 2017; Dłubek et al., 2021). Abnormal lipid metabolism is the primary feature of several refractory chronic diseases (Yang et al., 2016), such as atherosclerotic disease (Michos et al., 2019), obesity (Wang et al., 2014), fatty liver disease (Vernon et al., 2011), and diabetes mellitus (Garde et al., 2019). Thus, developing novel tools and strategies for maintaining cholesterol homeostasis is urgently required to prevent and treat these diseases.

Long non-coding RNAs (lncRNAs) are a class of RNA that do not encode proteins (Kim et al., 2009). Instead, they are involved in complex biological processes and pathophysiological conditions, including lipid metabolism disorders (Zeng et al., 2018; Simion et al., 2019). Recently, numerous clinical studies have shown that lncRNAs impair cholesterol homeostasis and play a critical role in the progression of lipid-related diseases (Han et al., 2019; Ou et al., 2020). For example, a primate-specific lncRNA (*CHROME*)



was found to be elevated in the plasma and atherosclerotic plaques of patients with coronary heart disease (CHD) (Hennessy et al., 2019). Similarly, highly up-regulated in liver cancer (*HULC*) lncRNA was discovered to modulate the deregulation of lipid metabolism in hepatoma cells and result in malignant development (Cui et al., 2015). These findings suggest that lncRNAs regulate lipid metabolism and promote the development of lipid-related diseases. LncRNAs might also function as the miRNAs sponges and affect lipid metabolism and related diseases (Lan et al., 2019). Importantly, lncRNAs also play an essential in the progression of some other diseases, such as cancer (Hahne and Valeri, 2018). Much research has been conducted on the specific functions of lncRNAs in these diseases.

The emerging role of lncRNAs as potential biomarkers and therapeutic targets for lipid-related diseases has not explicitly been summarised, and the present review aims to fill this gap in the literature. LncRNAs have been increasingly recognized as potential biomarkers for various human diseases, including atherosclerosis (Simion et al., 2020), MI (Spiroski et al., 2021), liver disease (Yang et al., 2021), and cancer (Xing et al., 2021). Here, we mainly reviewed the recent investigations of the role of lncRNAs as potential biomarkers and therapeutic targets in lipid-related diseases. Findings from this review would summarize the mechanisms by which lncRNAs act as biomarkers and therapeutic targets for lipid-related diseases.

LNCRNAS MECHANISMS OF ACTION

Recent studies have illustrated that lncRNAs can bind to the proteins, RNA, DNA, or a combination of them to exert their

functions (Fasolo et al., 2019; Hu Y. et al., 2019). As regulators of gene expression, lncRNAs involve in various biological processes (Fernandes et al., 2019; Mumtaz and Online, 2017), acting as miRNA sponge, decoys, scaffolds, guides, and post-translation regulation (Rinn and Chang, 2012) (Figure 1). For instance, many lncRNAs act as a miRNA sponge to regulate miRNAs and their targets. For example, small nucleolar RNA host gene 16 (*SNHG16*) facilitated the development and progression of neuroblastoma by upregulating homeobox A7 (*HOXA7*) expression via sponging miR-128-3p (Bao et al., 2020). Decoying lncRNAs mediated transcriptional repression by guiding chromatin modifiers such as m⁶A formation and recognition to genomic targets, such as *XIST* (Patil et al., 2016), *HOTAIR* (Loewen et al., 2014), and *GAS5* (Sun et al., 2017). LncRNAs can be used as scaffolds to form enhancer loops or as structural components of ribonucleoprotein complexes (Stackhouse et al., 2020). Nuclear paraspeckle assembly transcript 1 (*NEAT1*) scaffolds broadly interacts with NONO/PSF and other RNA-binding proteins (RBPs) and that globally enhance pri-miRNA processing (Jiang et al., 2017). Additionally, many lncRNAs exert their functions by sequestering regulatory factors in the nucleus or cytoplasm: for example, colon cancer-associated transcript-2 (*CCAT2*) can block miR-145 maturation by inhibiting pre-miR-145 export to cytoplasm (Yu Y. et al., 2017); whereas cytoplasmic lncRNAs, such as *lincRNA-p21*, interact with RNA-binding protein HuR to recruit let-7/Ago2 to inhibit their repression of *lincRNA-p21* stability (Yoon et al., 2012). Finally, lncRNAs can act as enhancers or co-activators of target gene activation, such as *H19* and *GAS5*. LncRNA may

TABLE 1 | Summary of the act of lncRNAs as therapeutic targets and potential biomarkers for lipid-related diseases.

LncRNAs	Dys-regulation	Human samples	Targets	Molecular mechanisms	Diseases	References
<i>HOXC-AS1</i>	Up	Carotid atherosclerosis	HOXC6	Facilitates HOXC6 expression	Atherosclerosis	Huang et al. (2016)
<i>GAS5</i>	Down	Atherosclerotic plaque	—	—	Atherosclerosis	Chen et al. (2017)
<i>RAPIA</i>	Up	Atherosclerotic plaque	miR-183-5p, ITGB1	Promotes ITGB1 expression by targeting miR-183-5p	Atherosclerosis	Sun et al. (2020)
<i>MIAT</i>	Up	Serum	miR-149-5p, CD47	Promotes CD47 expression by targeting miR-149-5p	Atherosclerosis	Ye et al. (2019)
<i>LncRNA-ATB</i>	Up	Serum	Caspase-3	Promotes the expression of caspase-3	Atherosclerosis	Yu et al. (2019)
<i>CHROME</i>	Up	Plasma	miR-27b, miR-33a, miR-33b, miR-128 and ABCA1	Regulates cholesterol efflux and nascent HDL particle formation by miRNAs/ABCA1 pathway	Atherosclerosis	Hennessey et al. (2019)
<i>RP11-714G18.1</i>	Down	Atherosclerotic plaques	LRP2BP, MMP1	Display athero-protective role via LRP2BP/MMP1 pathway	—	—
<i>CASC11</i>	Down	Plasma	IL-9	Improve atherosclerosis by inhibiting IL-9 expression	Atherosclerosis	Tao et al. (2019)
<i>NEXN-AS1</i>	Down	Atherosclerotic plaques, blood	NEXN	Mitigates atherosclerosis by regulating NEXN	CAD	Hu et al. (2019a)
<i>ENST00000416361</i>	Up	Plasma	SREBP1, SREBP2	Promotes SREBP1 and SREBP2 expression	CAD	Li et al. (2020a)
<i>MEG3</i>	Up	Tissues	miR-26a, Smad1	Promotes Smad1 expression by targeting miR-26a	CAD	Bai et al. (2019)
<i>ANRIL</i>	Up	Tissue	EZR, CXCL11 or TMEM106B	Exerts opposing effects on endothelial cell activities associated with coronary artery disease	CAD	Cho et al. (2020)
<i>Ang362</i>	Up	Plasma	—	—	CHD	Wang et al. (2020a)
<i>KCNQ1OT1</i>	Up	Serum	miR-26a-5p, ATG12	Promotes cardiomyocyte autophagy and aggravates MI by miR-26a-5p/ATG12 axis	MI	Li et al. (2021a)
<i>LINC00261</i>	Up	Tissues	miR-522-3p, TNRC6A	Promotes MI through the miR-522-3p/TNRC6A axis	MI	Jiang et al. (2021)
<i>NRF</i>	Up	Blood	—	—	MI patients with HF	Yan et al. (2020)
<i>NEAT1</i>	Up	Blood	miR-378a-3p, ATG12	Promotes cardiomyocytes injury by targeting miR-378a-3p	MI	Zhao et al. (2020)
<i>CHAST</i>	Up	Blood	—	—	MI	Wang et al. (2020b)
<i>MALAT1</i>	Up	Tissue	miR-144-3p	Promotes cardiomyocyte apoptosis after MI via targeting miR-144-3p	MI	Gong et al. (2019)
<i>TTTY15</i>	Up	Blood	miR-455-5p, JDP2	Promotes hypoxia-induced cardiomyocytes injury by targeting miR-455-5p	MI	Huang et al. (2019a)
<i>CAIF</i>	Down	Tissues and serum	—	—	MI	Wu et al. (2019)
<i>MALAT1</i>	Up	Serum	miR-200a-3p, PDCD4	Regulates cardiomyocytes apoptosis after via modulating miR-200a-3p/PDCD4 axis	MI	Sun and Zhang, (2019)
<i>TUG1</i>	Up	Aortic valves	miR-204-5p, Runx2	Promotes osteoblast differentiation by miR-204-5p/Runx2 axis	CAVD	Yu et al. (2018)
<i>LncARSR</i>	Up	Serum	SREBP-2, HMGCR	Increases SREBP-2 expression and HMGCR.	Hypercholesterolemia	Huang et al. (2018)
<i>HULC</i>	Up	HCC tissues	ASCL1, PPARA	miR-9/PPARA/ACSL1/cholesterol/RXRA/HULC signalling	Hepatocellular carcinoma	Cui et al. (2015)
<i>NEAT1</i>	Up	Serum	miR-129-5p, SOCS2	Promotes liver fibrosis by miR-129-5p/SOCS2	ASH	Ye et al. (2020)
<i>MALAT1</i>	Up	Liver biopsy	miR-20b-5p, TXNIP	Promotes TXNIP expression by targeting mR-20b-5p	NAFLD	Li et al. (2021b)
<i>LeXis</i>	Up	Liver biopsy	—	—	NAFLD	Park et al. (2020)
<i>B4GALT1-AS1</i>	Down	Liver tissues	hnRNPA1	Recruits hnRNPA1 to suppress hepatic lipogenesis and gluconeogenesis	NAFLD	Wang et al. (2018)
<i>GAS5</i>	Up	Plasma	—	—	NAFLD	Han et al. (2020)
<i>LncARSR</i>	Up	Liver tissues	Akt, SREBP-1c	—	NAFLD	—

(Continued on following page)

TABLE 1 | (Continued) Summary of the act of lncRNAs as therapeutic targets and potential biomarkers for lipid-related diseases.

LncRNAs	Dys-regulation	Human samples	Targets	Molecular mechanisms	Diseases	References
<i>Lnc18q22.2</i>	Up	Liver tissues	—	Promotes hepatic lipogenesis via Akt/SREBP-1c pathway	NAFLD	Zhang et al. (2018)
<i>RP11-142A22.4</i>	Up	Visceral adipose tissue	miR-587, Wnt5β	Promotes adipogenesis by sponging miR-587 to modulate Wnt5β expression	Obesity	Atanasovska et al. (2017) Zhang et al. (2020c)
<i>LINC00473</i>	Down	Adipose tissue	—	—	Obesity and type-2 diabetes	Tran et al. (2020)
<i>E330013P06</i>	Up	Blood	—	—	Breast cancer patient with type-2 diabetes	Chen et al. (2020)
<i>SNHG8</i>	Up	Blood	SOCS3, ICAM1	Promotes SOCS3 or ICAM1 expression by sponging miR-411-5p	AMI	Zhuo et al. (2019)

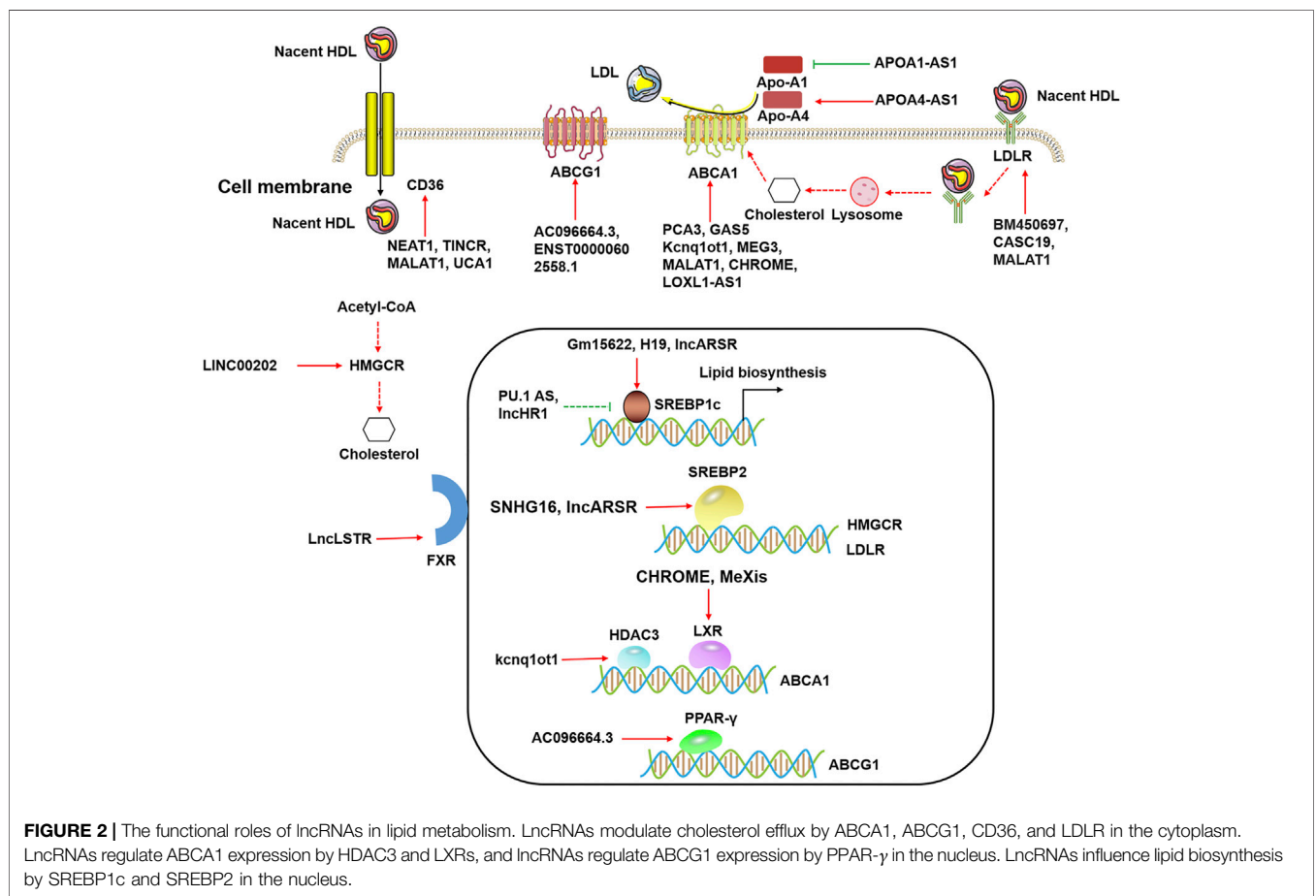


FIGURE 2 | The functional roles of lncRNAs in lipid metabolism. LncRNAs modulate cholesterol efflux by ABCA1, ABCG1, CD36, and LDLR in the cytoplasm. LncRNAs regulate ABCA1 expression by HDAC3 and LXRs, and lncRNAs regulate ABCG1 expression by PPAR-γ in the nucleus. LncRNAs influence lipid biosynthesis by SREBP1c and SREBP2 in the nucleus.

have more than one function, varying by subcellular localization, stimuli, and/or cell types. With the continuous increase of lncRNA-mediated functions, it has become clear that they are important regulators of multiple biological and cellular processes and can be used as candidate diagnostic and prognostic biomarkers for human diseases.

LNCRNAs PARTICIPATE IN THE DEVELOPMENT OF LIPID-RELATED DISEASES

Lipid metabolism is the biosynthesis and biodegradation of lipids in cells (Santos and Schulze, 2012). It involves the breakdown and

storage of fats for energy and the synthesis of structural and functional lipids (de Carvalho and Caramujo, 2018). Lipid biosynthesis is a part of metabolic abnormalities in cells, which require large quantities of lipids to synthesize cytomembranes, organelles, and signaling molecules during cell proliferation (Xu et al., 2020). Importantly, fatty acid oxidation (FAO) can provide abundant ATP for cells (Jeon et al., 2012), and fatty acids are a major source of ATP molecules (Fhu and Ali, 2020). In addition, lncRNAs affect gene expression that is involved in lipid metabolism (Table 1). Numerous studies have shown that lncRNAs participate in lipid metabolism by influencing the expression of key genes, networks, and pathways involved in lipid biosynthesis, cholesterol transport, lipid uptake, and cholesterol efflux (Figure 2).

Recent studies have reported that lncRNAs participate in the regulation of various genes expression in lipid metabolism that was induced by hormones (Fu et al., 2020), environmental stress (Wen et al., 2020), lipid/cholesterol (Ma et al., 2018), and obesity/type 2 diabetes (Hu et al., 2020). A single lncRNA often targets multiple mRNAs, and these mRNAs are linked to the different metabolic pathways (Huang, 2018). It is important to note that each mRNA is typically targeted by several lncRNAs, enabling coordinated gene expression. Many molecules are involved in lipid metabolism, including nuclear transcription factors such as LXR, FXR, SREBP, and the scavenger receptor CD36 (Figure 2) (Shimano and Sato, 2017; Yan et al., 2018; Piccinin et al., 2021). These regulatory molecules, along with lncRNAs, are implicated in the regulation of lipid metabolism.

Given the fact that lipid metabolism is distributed different cellular organelles also transport of the intermediates between the different organelles is an important point in lipid metabolism (Khor et al., 2013; Xu and Taubert, 2021). For example, lipid metabolism is located in the endoplasmic reticulum (ER) for lipid biosynthesis (Jacquemyn et al., 2017), mitochondria and peroxisomes for β -oxidation (Zhou et al., 2018), lipid droplets (LDs) for storage and transport (Freyre et al., 2019), and lysosomes for lipid hydrolysis and recycling (Go et al., 2012). Lipid metabolism includes processes such as lipid uptake, biosynthesis, catabolism, and secretion. LncRNAs can affect biological functions in many ways, such as the miRNA sponge, guide or decoy, scaffold, and chromatin remodeling. Currently, numerous lncRNAs have been identified to be involved in the regulation of lipid metabolism. However, many lncRNAs with lipid metabolism functions do not directly target genes involved in lipid metabolism pathways (He et al., 2019; Lan et al., 2019), such as triglyceride and cholesterol biosynthesis and fatty acid oxidation. Instead, they target the lncRNA-miRNA-mRNA and lncRNA-mRNA axes. For example, the lncRNA *HULC* has been shown to regulate abnormal lipid metabolism by decreasing miR-9 expression, leading to the upregulation of RXRA expression (Cui et al., 2015). RXRA, a member of the RXR family that can be activated by sterol (Costet et al., 2000), modulates the lipid metabolism disorders by activating acyl-CoA synthetase long-chain family member 1 (ACSL1) (Cui et al., 2015). Similarly, lncRNA *PU.1 AS* regulates lipid metabolism via the sterol regulatory element-binding protein-1c (SREBP-1c) pathway, resulting in reduced triglyceride

synthesis (Dong et al., 2019). Transcription factors of the SREBP family, including SREBP-1a, SREBP-1c, and SREBP-2, are central to transcriptional control of genes related to lipid and fatty acid metabolism (Brown and Goldstein, 1999). Interestingly, overexpression of SREBP-1c is known to facilitate fatty acid and triglyceride synthesis and lead to lipid accumulation in the liver (Yan et al., 2016). On the other hand, the inhibition of SREBP-1c is shown to alleviate lipid accumulation and lipotoxicity (Jin et al., 2020). The involvement of a lncRNA derived from hepatocytes (*lnc-HC*) in lipid metabolism has been extensively reported. For example, *lnc-HC* was found to regulate PPAR γ -mediated lipid metabolism and triglyceride (TG) concentration via miR-130b-3p, where *lnc-HC* expression was positively correlated with the miR-130b-3p expression (Lan et al., 2019). Furthermore, it has been illustrated that *lnc-HC* forms a complex with hnRNPA2B1 and negatively regulates Cyp7a1 and Abca1 expressions; both are implicated in hepatocytic cholesterol metabolism (Lan et al., 2016). Another lncRNA and hnRNP complex has also been identified with LeXis and RALY hnRNP, which are involved in lipid metabolism and influence metabolic gene expression (Sallam et al., 2016).

DISEASES ASSOCIATED WITH LNCRNA-RELATED LIPID DYSREGULATION

Several diseases, including atherosclerosis, MI, liver disease, and hypercholesterolemia, are caused by or associated with lipid dysregulation (Butt et al., 2017; Gluchowski et al., 2017; Michos et al., 2019). Importantly, studies focused on these diseases were performed using patient specimens, animal models (ApoE $^{-/-}$ and LDL $^{-/-}$), and atherosclerosis model cell lines, such as human umbilical vein endothelial cells (HUVECs) (Chen L. et al., 2019), human peripheral blood monocytes (THP-1) (Choi et al., 2021), human vascular smooth muscle cells (HVSMCs) (Li X. et al., 2021). Therefore, we only summarized several representative studies that mainly focused on lncRNA functions in lipid-related disease processes.

Disruption of lipid metabolism has been confirmed as a significant factor in the pathogenesis of atherosclerosis (Sukhorukov et al., 2020). The progression of atherosclerosis is known to be regulated by disturbances of lipid metabolism (Lovren et al., 2015), which impairs endothelial cells' function. Recent studies have identified *H19* as a well-known lncRNA associated with atherosclerosis (Huang Y. et al., 2019). *H19* expression has been reported to be up-regulated in patients with atherosclerosis and may be a potential therapeutic target for atherosclerosis (Yang Y. et al., 2019). Knockdown of *H19* inhibits hyperlipidemia and alleviates atherosclerotic lesions in HFD-treated ApoE $^{-/-}$ mice (Pan and sciences, 2017; Shi et al., 2020), while lentivirus-mediated *H19*-forced expression increase the plaque area size (Huang Y. et al., 2019). Technically, *H19* acts as a molecular sponge for miR-148b-3p and activates its expression of ELF5 (E74 like ETS transcription factor 5), resulting in the restoration of ELF5 that inhibit the cell migration in ox-LDL-stimulated HUVECs (Liu S. et al., 2021).

Additionally, *lncARSR*, a lncRNA regulator of Akt signaling associated with HCC and RCC, has recently been studied as a potential therapeutic target for cholesterol disorder, and its downstream target SREBP-2 was identified. SREBP-2 has been found to bind to HMG-CoA reductase (HMGCR) to promote hepatic cholesterol biosynthesis, resulting in aberrant regulation of cholesterol metabolism (Huang et al., 2018). Collectively, *lncARSR*-SREBP-2-HMGCR plays a pivotal role in regulating lipid metabolism and the development of atherosclerosis (Xiao and Song, 2013).

Dysregulated lipid metabolism is a hallmark of non-alcoholic steatohepatitis (NASH), a very common liver disorder (Musso et al., 2013). Recently, growing evidence has suggested that dysregulated lncRNA expression is associated with inflammation and fibrosis in NASH (Leti et al., 2017). Whole transcriptome analysis and identified differentially expressed lncRNAs (*RP11-128N14.5* and *TGFB2-OT1*) in patients with non-alcoholic fatty liver disease (NAFLD) (Di Mauro et al., 2019). Several lncRNAs, including hepatocellular carcinoma up-regulated lncRNA, *NEAT1*, and metastasis-associated lung adenocarcinoma transcript 1 (*MALAT1*), were highly expressed in liver biopsies from NAFLD patients (Leti et al., 2017). Furthermore, expression of *MALAT1* was upregulated in livers of ob/ob mice and hepatocytes exposed to palmitate (Yan et al., 2016). Another lncRNA, Alu-mediated p21 transcriptional regulator (APTR), was discovered to be significantly increased in human cirrhosis and activate hepatic stellate cells (Yu et al., 2015). Hepatic *LeXis* expression is a mediator of cholesterol biosynthesis (Sallam et al., 2016). Thus, raising or lowering *LeXis* levels influence the expression of genes involved in cholesterol biosynthesis and alter liver and plasma cholesterol levels (Sallam et al., 2016). Brown fat-enriched lncRNA 1 (*Blncl*) was strongly elevated in obesity and NAFLD in mice (Zhao et al., 2018). Hepatic *Blncl* deficiency is suggested to abrogate high-fat diet-induced hepatic steatosis and insulin resistance and ameliorate NASH pathogenesis (Zhao et al., 2018). These findings provide a further rationale for analyzing global changes in lncRNA expression in NAFLD and NASH.

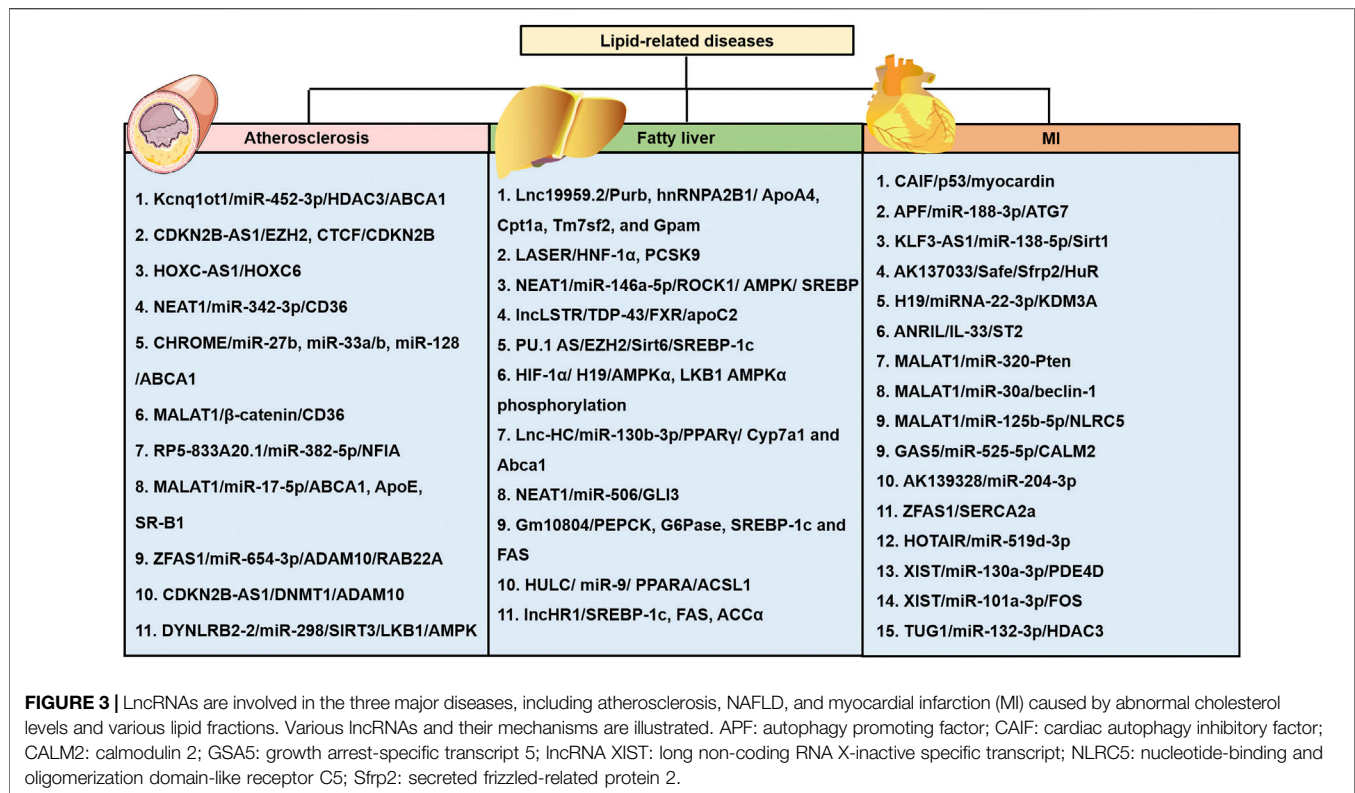
Recent bioinformatics and high-throughput sequencing studies have revealed that lncRNAs are differentially expressed in patients with hypoalphalipoproteinemia and MI caused by abnormal lipid metabolism (Wang et al., 2019). Differently expressed lncRNAs and mRNAs in atherosclerosis by analyzing dataset GSE28829 (Wang et al., 2019). A total of 654 lncRNAs and 5,784 mRNAs were significantly dysregulated in the progression of atherosclerosis (Wang et al., 2019). Moreover, six lncRNAs, *ZFAS1* (ZNF1 antisense RNA 1), *LOC100506730*, *LOC100506691*, *DOCK9-AS2*, *RP11-612.3*, and *LOC100130219*, were confirmed as potential novel therapeutic and prognostic targets for atherosclerosis (Wang et al., 2019). lncRNA *ENST00000416361* was higher in the plasma of 50 patients with coronary artery disease (CAD) than the 50 healthy volunteers (Li P. et al., 2020). SREBP1 and SREBP2 were also up-regulated in CAD patients and showed positive correlations with *ENST00000416361* (Li P. et al., 2020). Single nucleotide polymorphisms (SNPs) on the cyclin-dependent kinase inhibitor 2B antisense RNA (*ANRIL*) and *MALAT1*,

two lncRNAs, affect the prognosis of MI (Li Y. et al., 2020). *ANRIL* rs9632884 and *MALAT1* rs3200401 were significantly associated with the lipid levels of both controls and MI patients (Li Y. et al., 2020). *KCNQ1* overlapping transcript 1 (*KCNQ1OT1*) was found to be increased in the serum of myocardial infarction (MI) patients, ischemia/reperfusion (I/R) mouse and hypoxia/reoxygenation (H/R)-induced cell model (Li J. et al., 2021). Moreover, several SNPs interacted with sex and age and modified the total cholesterol (rs9632884), LDL-C (rs1537373), and creatinine levels, affecting the risk of MI (Li Y. et al., 2020). These studies using clinical specimens and *in vitro* disease models have suggested that lncRNAs are involved in lipid-related diseases. However, the results should be further validated via *in vitro* and *in vivo* systems. Further research is required to analyze potential biomarkers and therapeutic targets in various lipid-related diseases (see Figure 3). This review provides a comprehensive insight into the current knowledge regarding the involvement of lncRNAs in regulating lipid metabolism, which may unveil the potential biomarkers and therapeutic targets for treating lipid-related diseases (Table 1).

LNCRNAs ARE IDEAL DIAGNOSTIC BIOMARKERS AND THERAPEUTIC TARGETS

Diagnosis of several lipid-related diseases and their associated disease risks are mainly accomplished by analyzing the concentrations of lipid components such as total cholesterol, HDL, LDL, and triglycerides in the blood (Gotto, 2011; Paredes et al., 2019). This method only obtains accurate results when patients are fasted for at least 9–12 h. However, it provides limited information on cholesterol levels. Thus, it is necessary to search for better diagnostics and novel biomarkers for lipid-related diseases to overcome these disadvantages. lncRNAs are present in body fluids and are as stable as mRNA. Due to their tissue-specific properties, lncRNAs can be used as clinical indicators for diagnosis and are expected to become a new target for disease treatment (Table 1). Therefore, the application of lncRNAs as diagnostic biomarkers can result in a timely collection of more accurate and detailed disease information and risk factor data.

Previous attempts to use lncRNAs as biomarkers for disease diagnosis have been demonstrated in several cancer studies (Ratti et al., 2020) (Table 2). They revealed the functional roles of lncRNAs during cancer progression, including tumorigenesis, metastasis, and resistance to cancer treatment (Shen et al., 2015; Bin et al., 2018). Interestingly, some lipid-related lncRNAs mentioned in this review have also been emphasized in some cancer studies and proposed as potential diagnostic biomarkers (Peng et al., 2020). For example, the *CHROME*, which is mainly involved in cholesterol efflux and HDL biogenesis, was elevated in the plasma and atherosclerotic plaques of individuals and identified as a novel biomarker for the progression of CAD (Hennessy et al., 2019). On the other hand, plasma *LeXis*, which participates in cholesterol metabolism and the development of hepatic steatosis, was found to act as a



non-invasive diagnostic biomarker for NASH (Park et al., 2020). *NEAT1* and *ANRIL*, which are associated with cholesterol synthesis and MI, respectively, were suggested to be biomarkers that identify non-small cell lung carcinoma (NSCLC) (Yu X. et al., 2017; Osielska and Jagodziński, 2018). Furthermore, elevated plasma levels of *HULC*, which is involved in cholesterol synthesis, were identified as a biomarker for liver cancer (Xie et al., 2013). Additionally, the correlation between *MALAT1*, known to participate in cholesterol efflux, and lung cancer has been suggested as a diagnostic indicator (Lin et al., 2018). Moreover, the role of *TUG1*, an atherosclerosis-associated lncRNA, in various cancers has been previously studied (Niu et al., 2017; Guo et al., 2019). *TUG1* was found to recruit specific RNA-binding proteins to facilitate cancer progression (Duan et al., 2019). These results suggest that lncRNAs play multiple functional roles in various disease processes and, as has frequently been reported in recent studies, cholesterol homeostasis is closely related to cancer occurrence. Collectively, these reports on lncRNAs in cancer indicate that the development of lncRNA biomarkers for diagnosing lipid-related diseases is very promising.

Importantly, from a therapeutic perspective, the best approach to prevent and treat lipid-related diseases is to make certain lifestyle modifications, such as exercising more and consuming a healthy diet (Mannu et al., 2013). However, if high lipid levels persist, medication must be taken to lower them. As mentioned earlier, the diagnosis criteria for lipid-related diseases are based on detecting cholesterol levels present in plasma (Płaczkowska et al., 2014). Thus, the primary purpose of treatment is to reduce cholesterol to appropriate levels.

However, it is essential to note that the relationship between cholesterol and lipid-related diseases is ever-changing, which means that treatments also vary depending on the type and condition of the related disease. For instance, statin-based drugs, bile acid sequestrants, and cholesterol absorption inhibitors (Ezetimibe) are used clinically for different conditions. Specifically, statins decrease substances required for liver cholesterol production, bile oxides or bile acid sequestrants facilitate bile acid production from cholesterol, and cholesterol absorption inhibitors reduce cholesterol and limit cholesterol absorption from the small intestine (Taoufiq et al., 2011). In addition, drugs that only increase the absorption of LDL cholesterol have also been increasingly used recently (Lee et al., 2020). Due to their specific actions and side effects, these drugs are commonly used in combination in clinical and surgical treatments.

Importantly, lncRNAs involved in lipid metabolism can also be used as potential therapeutic targets to maintain cholesterol levels in the normal range. In general, RNA interference (RNAi), using shRNA, siRNA, or anti-sense oligonucleotide (ASO), is the most promising approach to target lncRNA silencing (Chi et al., 2017). This approach has been proven effective at the whole animal and cellular levels through various research (Liu et al., 2017; Zhang L. et al., 2020). For instance, the lentiviral shRNA targeting of lncRNA myocardial infarction associated transcript (*MIAT*) significantly attenuates atherosclerosis progression and increases plaque stability *in vivo* (Ye et al., 2019). Thus, a novel method for achieving safe and efficient RNAi delivery should be investigated and developed by further research. Furthermore, ASO-based methods are also studied for more stable and less off-

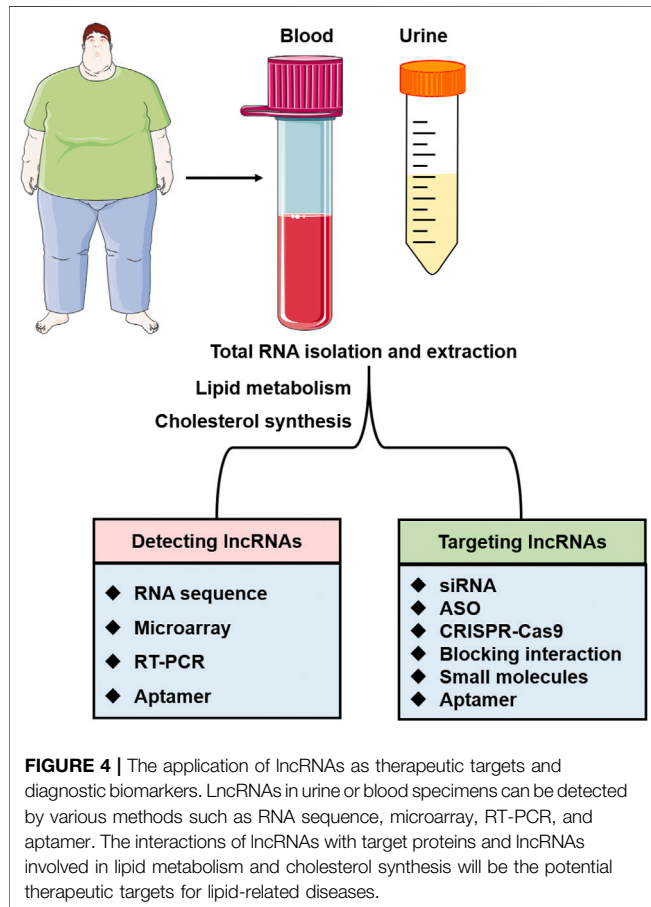
TABLE 2 | Summary of data from relevant lncRNAs-based biomarkers in human multiple tumors.

Biomarkers	Dys-regulation	Tumors	Sample type	Sample size	Technological approach	Application	Comments	References
<i>LncRNA-ATB</i> <i>FAM83H-AS1</i>	Up	Breast cancer	Serum	90 breast cancer patients	RT-PCR	Prognosis; disease monitoring	Serum lncRNA-ATB and FAM83H-AS1 could be used as a non-invasive diagnostic marker for early stages of breast cancer	El-Ashmawy et al. (2020)
<i>LINC00114</i> , <i>LINC00261</i> , <i>HOTAIR</i>	HOTAIR (Up), <i>LINC00114</i> and <i>LINC00261</i> (Down)	CRC	Tissues	459 nonmetastatic CRC samples and 87 metastatic CRC samples	RT-PCR	Prognosis; disease monitoring	3-lncRNA signature that includes <i>LINC00114</i> , <i>LINC00261</i> , and <i>HOTAIR</i> is an independent factor for predicting CRC prognosis	Liu et al. (2020)
<i>MSC-AS1</i>	Up	LC	Tissues	123 LC patients (111 tumor tissues, 12 adjacent normal samples)	—	—	—	—
<i>MSC-AS1</i>	RT-PCR	Diagnosis and prognosis	<i>MSC-AS1</i> may be used as a potential biomarker of LC.	Liu et al. (2021b)	—	—	—	—
<i>HELIS</i> <i>LINC01093</i> , <i>CYTOR</i>	<i>HELIS</i> and <i>LINC01093</i> (Down), <i>CYTOR</i> (Up)	HCC	Tissues	82 paired tissue samples from patients with HCC	RT-PCR	Prognosis; disease monitoring	Down-regulated <i>HELIS</i> and <i>LINC01093</i> , up-regulated <i>CYTOR</i> are perspectives for differential diagnostics of HCC	Burenina et al. (2021)
<i>SNHG18</i>	Up	HCC	Tissues, Plasma	71 paired HCC patients	RT-PCR	Diagnosis	—	—
<i>DLG2-AS1</i>	Down	LUAD	Tissues	70 LUAD patients	RT-PCR	Prognosis; disease monitoring	<i>DLG2-AS1</i> serves as a good diagnostic biomarker for LUAD patients	Arenas et al. (2020)
<i>MIAT</i> , <i>LINC00460</i> , and <i>LINC00443</i>	<i>MIAT</i> and <i>LINC00460</i> (Up) <i>LINC00443</i> (Down)	KIRC	Tissues	530 KIRC patients	RT-PCR	Prognosis; disease monitoring	The LPM based on three-lncRNAs could serve as independent prognostic factors with a tremendous predictive ability for KIRC patients	Zhang et al. (2020a)
<i>SAMMSON</i>	Up	OSCC, GBM	Tissues, Plasma	90 OSCC patients	—	—	—	—
56 patients with GBM (34 males and 22 females)	RT-PCR	Diagnosis and prognosis	<i>SAMMSON</i> might play a critical role in OSCC progression and serve as a novel prognostic and diagnostic biomarker in OSCC.	—	—	—	—	—
Plasma <i>SAMMSON</i> has diagnostic value for GBM	Xie et al. (2019); Zheng et al. (2020)	—	—	—	—	—	—	—
<i>LUCAT1</i>	Up	PTC	Tissues	61 PTC patients	RT-PCR	Diagnosis and prognosis	<i>LUCAT1</i> can act as a novel prognostic biomarker for patients with PTC	Luzón-Toro et al. (2019)
<i>PTENP1</i>	Down	BC	Plasma	50 patients with BC and 60 healthy controls	RT-PCR	Diagnosis	Exosomal <i>PTENP1</i> is a potential novel biomarker that can be used for the	Zheng et al. (2018)

(Continued on following page)

TABLE 2 | (Continued) Summary of data from relevant lncRNAs-based biomarkers in human multiple tumors.

Biomarkers	Dys-regulation	Tumors	Sample type	Sample size	Technological approach	Application	Comments	References
PANDAR, FOXD2-AS1, SMARCC2	Up	GC	Plasma	109 GC patients and 106 healthy controls	RT-PCR	Diagnosis	clinical detection of BC. Plasma PANDAR, FOXD2-AS1, and SMARCC2 may be appropriate diagnostic biomarkers for GC.	Yang et al. (2019b)



target occurrence in addition to RNA interference technology (Maruyama and Yokota, 2020). For example, *MALAT1* targeted ASO has been developed, and its inhibitory effect has been identified using animal models of malignancy (Amodio et al., 2018). Moreover, besides the method that targets lncRNA itself, controlling lncRNA function by inhibiting its interaction with the RNA-binding proteins has also been attempted (Kung et al., 2013; Bhat et al., 2016). However, note that RNA interference therapeutics have recently been progressed through preclinical development into clinical trials (Bobbitt and Rossi, 2016). Thus, applying these as ideal clinical therapeutics requires the development of safe and effective delivery systems.

Small molecules have been extensively used for the therapeutic targeting of various diseases. These compounds have greater cellular uptake and fewer administrative challenges than antisense oligonucleotides and viral vectors for RNAi delivery. Small molecule inhibitors target lncRNAs by preventing them from binding to their RNA-binding proteins (RBPs). After analysing the lncRNA expression profiles from lncRNA modulator atlas in pan-cancer (lncMAP) database by bioinformatics analysis, the lncRNA network consists of 1,206 nodes and 4,770 drug-lncRNA associations to examine the global relationship between small molecule drugs and their affected lncRNAs (He et al., 2019). In addition, small molecules were screened to modulate the lncRNA HOX transcript antisense RNA (*HOTAIR*)-enhancer of zeste homolog2 (*EZH2*) interaction using alphaScreen technology (Pedram Fatemi et al., 2015). The interaction was inhibited with *HOTAIR*-polycomb repressive complex 2 (PRC2) binding through small-molecule intervention resulting in reduced metastatic phenotypes in many cancers, including breast (Gupta et al., 2010), colorectal (Kogo et al., 2011), and hepatocellular carcinomas (El-Khazragy et al., 2020). However, it is necessary to investigate the lncRNA-protein interaction and pharmacological trends further to develop more effective small molecule drugs (Figure 3).

CONCLUSION AND FUTURE PERSPECTIVES

Recent studies have shown that lncRNAs are involved in various lipid-related diseases (Table 1), thereby opening up a new research field and providing insight for lncRNAs as important eukaryotic transcripts. Concerning the correlation between lncRNAs regulation and lipid-related diseases, atherosclerosis is the most frequently studied disease (Ye et al., 2021). The occurrence of lipid-related diseases is due to the inactivation of suppressor genes and the activation of pathogenic genes. Thus, screening and identifying candidate biomarkers for prognosis, monitoring, and evaluating patients' responses to therapies is required to develop novel strategies for lipid-related disease therapies. Also, ncRNAs (miRNAs and lncRNAs), DNA methylation, and histone modifications can epigenetically regulate gene expression. lncRNAs have recently served as important regulators of lipid-related diseases via various biological processes, including lipid metabolism, lipid

accumulation, lipid synthesis, and cholesterol efflux (Sallam et al., 2018; Chen X. et al., 2019; Wang Z. et al., 2020; Zuo et al., 2020). Thus, there is a considerable thrill in using lncRNAs as a critical therapeutic target in treating lipid-related diseases.

Recent studies have demonstrated that lncRNAs could be detected in the blood plasma, tumor tissue, and urine, making them serve as promising biomarkers for development as disease, including atherosclerosis, MI, and cancer diseases (Dastmalchi et al., 2020; Fattahi et al., 2020). Genome-wide sequencing techniques have emerged as an important technology and reported a large number of newly dysregulated lncRNAs, implying promising results about the broad application prospects of lncRNAs in the prognosis and diagnosis of lipid-related diseases. Deregulation of many lncRNAs, such as *H19* (Pan, 2017), *TUG1* (Li et al., 2018), *GAS5* (Chen et al., 2017), *RPIA* (Sun et al., 2020), *MIAT* (Ye et al., 2019), *CASC11* (Tao et al., 2019), *NEXN-AS1* (Hu Y.-W. et al., 2019), and *lnc00113* (Yao et al., 2018), has been detected in patients with atherosclerosis. LncRNAs including *H19*, *TUG1*, *MIAT*, and *CASC11* could be detected in serum samples as a potential diagnostic marker in patients with atherosclerosis. In addition to establishing the functional role of lncRNAs in diagnosis, some lncRNAs such as *AL117190.1*, *COL4A2-AS1*, *LINC00184*, *MEG3* and *MIR22HG* could function as crucial prognostic markers for patients (Yao et al., 2019). Besides, as diagnostic and prognostic markers, lncRNAs such as *H19* (Yörüker et al., 2018), *MEG3* (Wan and Zhao, 2020), *PVT1* (Pan et al., 2019), *FAM83H* antisense RNA 1 (*FAM83H-AS1*) (El-Ashrawy et al., 2020), *SNHG1* (Xiao et al., 2018), and *LUCAT1* (Xing et al., 2021) are involved in the process of various cancer progression. Thus, we speculate that dysregulated lncRNAs may be used as biomarkers to provide diagnosis and prognostic of lipid-related diseases but also are useful in therapeutic applications.

Although it is well established that high concentrations of serum cholesterol levels facilitate the development of atherosclerosis (Johnston et al., 2017), the association of LDL-C or other lipids with atherosclerosis remains controversial. To date, a large number of lncRNAs associated with lipid metabolism and lipid-related diseases have been identified through RNA-seq and bioinformatics analyses. The functions of these lncRNAs may have important clinical implications in lipid metabolism and lipid-related diseases since they provide a myriad of possibilities for the diagnostics and treatment of these diseases. Furthermore, lncRNAs have been described as high tissue- and cell type-specific expression patterns (Kopp and Mendell, 2018; Antonov et al., 2019), which could be classified as different subclasses of lipid-related diseases or even predict responses to treatments. However, our current knowledge of the effect of lncRNAs on lipid-related diseases is

possibly only the tip of the iceberg. Thus, more comprehensive investigations should be conducted to better understand how lncRNAs affect lipid-related diseases and develop new therapies.

The study of lncRNAs involved in controlling the cholesterol levels, specifically lncRNAs that directly interact with target genes or epigenetic proteins at the transcriptional level, may contribute to developing novel drugs to treat lipid-related diseases. Importantly, the latest next-generation sequencing-based big data research has identified numerous lncRNAs associated with various lipid-related diseases (Ye et al., 2021). However, further molecular biological research is needed to deepen the understanding of the association between various lncRNAs discovered and actual genetic mechanisms.

This review summarized various lipid-related lncRNAs and their target genes that play essential roles in lipid metabolism and lipid-related diseases. The involvement of lncRNAs was abnormally expressed in certain disease conditions, including atherosclerosis (Gao and Guo, 2021), myocardial infarction (Li J. et al., 2021), non-alcoholic fatty liver disease (Li J.-z. et al., 2021), and hypercholesterolemia (Tontonoz et al., 2017). Furthermore, a large number of lncRNAs identified from various studies were found to be associated with a diverse range of diseases. As lncRNAs are structurally and functionally conserved, further research is required to develop more effective diagnostics and therapeutics in this field or reveal the mechanism of certain diseases (see **Figure 4** and **Table 1**). Altogether, advancing the knowledge of these lncRNAs and their functions is crucial for developing novel detection and modification methods.

AUTHOR CONTRIBUTIONS

W-CY and X-FP conceived and outlined the article. W-CY, S-FH, CH, and LJ surveyed the literature and wrote the article. W-CY and S-FH researched the literature and provided suggestions. W-CY and X-FP conceived ideas and the initial design. All the authors have approved the manuscript for submission.

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REFERENCES

- Amodio, N., Stamato, M. A., Juli, G., Morelli, E., Fulciniti, M., Manzoni, M., et al. (2018). Drugging the lncRNA MALAT1 via LNA gapmer ASO Inhibits Gene Expression of Proteasome Subunits and Triggers Anti-multiple Myeloma Activity. *Leukemia* 32, 1948–1957. doi:10.1038/s41375-018-0067-3
- Antonov, I. V., Mazurov, E., Borodovsky, M., and Medvedeva, Y. A. (2019). Prediction of lncRNAs and Their Interactions with Nucleic Acids: Benchmarking Bioinformatics Tools. *Brief. Bioinformatics* 20, 551–564. doi:10.1093/bib/bby032

- Arenas, A., Cuadros, M., Andrades, A., García, D., Coira, I., Rodríguez, M., et al. (2020). LncRNA DLG2-AS1 as a Novel Biomarker in Lung Adenocarcinoma. *Cancers* 12. doi:10.3390/cancers12082080
- Atanasovska, B., Rensen, S. S., van der Sijde, M. R., Marsman, G., Kumar, V., Jonkers, I., et al. (2017). A Liver-specific Long Noncoding RNA with a Role in Cell Viability Is Elevated in Human Nonalcoholic Steatohepatitis. *Hepatology* 66, 794–808. doi:10.1002/hep.29034
- Bai, Y., Zhang, Q., Su, Y., Pu, Z., and Li, K. (2019). Modulation of the Proliferation/Apoptosis Balance of Vascular Smooth Muscle Cells in Atherosclerosis by lncRNA-MEG3 via Regulation of miR-26a/Smad1 Axis. *Int. Heart J.* 60, 444–450. doi:10.1536/ihj.18-195
- Bao, J., Zhang, S., Meng, Q., and Qin, T. (2020). SNHG16 Silencing Inhibits Neuroblastoma Progression by Downregulating HOXA7 via Sponging miR-128-3p. *Neurochem. Res.* 45, 825–836. doi:10.1007/s11064-020-02955-x
- Bhat, S. A., Ahmad, S. M., Mumtaz, P. T., Malik, A. A., Dar, M. A., Urwat, U., et al. (2016). Long Non-coding RNAs: Mechanism of Action and Functional Utility. *Non-coding RNA Res.* 1, 43–50. doi:10.1016/j.ncrna.2016.11.002
- Bin, X., Hongjian, Y., Xiping, Z., Bo, C., Shifeng, Y., and Binbin, T. (2018). Research Progresses in Roles of LncRNA and its Relationships with Breast Cancer. *Cancer Cel. Int.* 18, 179. doi:10.1186/s12935-018-0674-0
- Bobbin, M. L., and Rossi, J. J. (2016). RNA Interference (RNAi)-Based Therapeutics: Delivering on the Promise? *Annu. Rev. Pharmacol. Toxicol.* 56, 103–122. doi:10.1146/annurev-pharmtox-010715-103633
- Brown, M. S., and Goldstein, J. L. (1999). A Proteolytic Pathway that Controls the Cholesterol Content of Membranes, Cells, and Blood. *Proc. Natl. Acad. Sci.* 96, 11041–11048. doi:10.1073/pnas.96.20.11041
- Burenina, O. Y., Lazarevich, N. L., Kustova, I. F., Shavochkina, D. A., Moroz, E. A., Kudashkin, N. E., et al. (2021). Panel of Potential lncRNA Biomarkers Can Distinguish Various Types of Liver Malignant and Benign Tumors. *J. Cancer Res. Clin. Oncol.* 147, 49–59. doi:10.1007/s00432-020-03378-5
- Butt, A. A., Yan, P., Chew, K. W., Currier, J., Corey, K., Chung, R. T., et al. (2017). Risk of Acute Myocardial Infarction Among Hepatitis C Virus (HCV)-Positive and HCV-Negative Men at Various Lipid Levels: Results from ERCHIVES. *Clin. Infect. Dis.*, 65, 557–565. doi:10.1093/cid/cix359
- Chen, L., Zheng, S. Y., Yang, C. Q., Ma, B. M., and Jiang, D. (2019a). MiR-155-5p Inhibits the Proliferation and Migration of VSMCs and HUVECs in Atherosclerosis by Targeting AKT1. *Eur. Rev. Med. Pharmacol. Sci.* 23, 2223–2233. doi:10.26355/eurrev_201903_17270
- Chen, L., Yang, W., Guo, Y., Chen, W., Zheng, P., Zeng, J., et al. (2017). Exosomal lncRNA GAS5 Regulates the Apoptosis of Macrophages and Vascular Endothelial Cells in Atherosclerosis. *PLoS one* 12, e0185406. doi:10.1371/journal.pone.0185406
- Chen, R., Shi, P., Zhang, Y., Wu, H., Li, X., Yang, W., et al. (2020). Long Non-coding RNAE330013P06 Promotes Progression of Breast Cancer with Type 2 Diabetes. *J. Clin. Lab. Anal.* 34, e23172. doi:10.1002/jcla.23172
- Chen, X., Tan, X.-R., Li, S.-J., and Zhang, X.-X. (2019b). LncRNA NEAT1 Promotes Hepatic Lipid Accumulation via Regulating miR-146a-5p/ROCK1 in Nonalcoholic Fatty Liver Disease. *Life Sci.* 235, 116829. doi:10.1016/j.lfs.2019.116829
- Chi, X., Gatti, P., and Papoian, T. (2017). Safety of Antisense Oligonucleotide and siRNA-Based Therapeutics. *Drug Discov. Today* 22, 823–833. doi:10.1016/j.drudis.2017.01.013
- Cho, H., Li, Y., Archacki, S., Wang, F., Yu, G., Chakrabarti, S., et al. (2020). Splice Variants of lncRNA RNA ANRIL Exert Opposing Effects on Endothelial Cell Activities Associated with Coronary Artery Disease. *RNA Biol.* 17, 1391–1401. doi:10.1080/15476286.2020.1771519
- Choi, S.-H., Agatista-Boyle, C., Gonen, A., Kim, A., Kim, J., Alekseeva, E., et al. (2021). Intracellular AIBP (Apolipoprotein A-I Binding Protein) Regulates Oxidized LDL (Low-Density Lipoprotein)-Induced Mitophagy in Macrophages. *Atvb* 41, e82–e96. doi:10.1161/atvbaha.120.315485
- Costet, P., Luo, Y., Wang, N., and Tall, A. R. (2000). Sterol-dependent Transactivation of the ABC1 Promoter by the Liver X Receptor/Retinoid X Receptor. *J. Biol. Chem.* 275, 28240–28245. doi:10.1074/jbc.m003337200
- Cui, M., Xiao, Z., Wang, Y., Zheng, M., Song, T., Cai, X., et al. (2015). Long Noncoding RNA HULC Modulates Abnormal Lipid Metabolism in Hepatoma Cells through an miR-9-Mediated RXRA Signaling Pathway. *Cancer Res.* 75, 846–857. doi:10.1158/0008-5472.can-14-1192
- Dastmalchi, N., Safaralizadeh, R., and Nargesi, M. M. (2020). LncRNAs: Potential Novel Prognostic and Diagnostic Biomarkers in Colorectal Cancer. *Cmc* 27, 5067–5077. doi:10.2174/0929867326666190227230024
- de Carvalho, C. C. R., and Caramujo, M. J. (2018). *The Various Roles of Fatty Acids*, 23. Basel, Switzerland: Molecules. doi:10.3390/molecules23102583
- Di Mauro, S., Scamporrino, A., Petta, S., Urbano, F., Filippello, A., Ragusa, M., et al. (2019). Serum Coding and Non-coding RNAs as Biomarkers of NAFLD and Fibrosis Severity. *Liver Int.* 39, 1742–1754. doi:10.1111/liv.14167
- Dong, Z., Li, C., Yin, C., Xu, M., Liu, S., and Gao, M. (2019). LncRNA PU.1 AS Regulates Arsenic-Induced Lipid Metabolism through EZH2/Sirt6/SREBP-1c Pathway. *J. Environ. Sci.* 85, 138–146. doi:10.1016/j.jes.2019.05.019
- Duan, W., Nian, L., Qiao, J., and Liu, N. N. (2019). LncRNA TUG1 Aggravates the Progression of Cervical Cancer by Binding PUM2. *Eur. Rev. Med. Pharmacol. Sci.* 23, 8211–8218. doi:10.26355/eurrev_201910_19128
- Dłubek, J., Rysz, J., Jabłonowski, Z., Gluba-Brzózka, A., and Franczyk, B. (2021). The Correlation between Lipid Metabolism Disorders and Prostate Cancer. *Curr. Med. Chem.* 28, 2048–2061. doi:10.2174/0929867327666200806103744
- El-Ashmawy, N. E., Hussien, F. Z., El-Feky, O. A., Hamouda, S. M., and Al-Ashmawy, G. M. (2020). Serum LncRNA-ATB and FAM83H-AS1 as Diagnostic/prognostic Non-invasive Biomarkers for Breast Cancer. *Life Sci.* 259, 118193. doi:10.1016/j.lfs.2020.118193
- El-Khazragy, N., Elshimy, A. A., Hassan, S. S., Shaaban, M. H., Bayoumi, A. H., El Magdoub, H. M., et al. (2020). Lnc-HOTAIR Predicts Hepatocellular Carcinoma in Chronic Hepatitis C Genotype 4 Following Direct-Acting Antivirals Therapy. *Mol. Carcinog* 59, 1382–1391. doi:10.1002/mc.23263
- Fasolo, F., Patrucco, L., Volpe, M., Bon, C., Peano, C., Mignone, F., et al. (2019). The RNA-Binding Protein ILF3 Binds to Transposable Element Sequences in SINEUP lncRNAs. *FASEB J.* 33, 13572–13578. doi:10.1096/fj.201901618rr
- Fattahi, S., Kosari-Monfared, M., Golpour, M., Emami, Z., Ghasemiyani, M., Nouri, M., et al. (2020). LncRNAs as Potential Diagnostic and Prognostic Biomarkers in Gastric Cancer: A Novel Approach to Personalized Medicine. *J. Cel Physiol* 235, 3189–3206. doi:10.1002/jcp.29260
- Fernandes, J. C. R., Acuña, S. M., Aoki, J. I., Floeter-Winter, L. M., and Muxel, S. M. (2019). Long Non-coding RNAs in the Regulation of Gene Expression: Physiology and Disease. *Noncoding RNA* 5 (1), 17. doi:10.3390/ncrna5010017
- Fhu, C. W., and Ali, A. (2020). Fatty Acid Synthase: An Emerging Target in Cancer, 25. *Molecules*. 25, 3935. doi:10.3390/molecules25173935
- Freyre, C., Rauher, P. C., Ejsing, C. S., and Klemm, R. W. J. M. C. (2019). MIGA2 Links Mitochondria, the ER, and Lipid Droplets and Promotes De Novo Lipogenesis in Adipocytes. *Mol. Cel.* 76, 811–825. doi:10.1016/j.molcel.2019.09.011
- Fu, X., Cong, H., Zhao, S., Li, Y., Liu, T., Sun, Y., et al. (2020). Construction of Glycometabolism- and Hormone-Related lncRNA-Mediated Feedforward Loop Networks Reveals Global Patterns of lncRNAs and Drug Repurposing in Gestational Diabetes. *Front. Endocrinol.* 11, 93. doi:10.3389/fendo.2020.00093
- Gao, H., and Guo, Z. (2021). LncRNA XIST Regulates Atherosclerosis Progression in Ox-LDL-Induced HUVECs. *Open Med. (Warsaw, Poland)* 16, 117–127. doi:10.1515/med-2021-0200
- Garde, S., Akhter, R., Nguyen, M., Chow, C., and Eberhard, J. (2019). Periodontal Therapy for Improving Lipid Profiles in Patients with Type 2 Diabetes Mellitus: A Systematic Review and Meta-Analysis. *Int. J. Mol. Sci.* 20. doi:10.3390/ijms20153826
- Gluchowski, N. L., Becuwe, M., Walther, T. C., and Farese, R. V. (2017). Lipid Droplets and Liver Disease: from Basic Biology to Clinical Implications. *Nat. Rev. Gastroenterol. Hepatol.* 14, 343–355. doi:10.1038/nrgastro.2017.32
- Go, G. W., and Mani, A. (2012). Low-density Lipoprotein Receptor (LDLR) Family Orchestrates Cholesterol Homeostasis. *Yale J. Biol. Med.* 85, 19–28.
- Gong, X., Zhu, Y., Chang, H., Li, Y., and Ma, F. (2019). Long Noncoding RNA MALAT1 Promotes Cardiomyocyte Apoptosis after Myocardial Infarction via Targeting miR-144-3p. *Biosci. Rep.* 39. doi:10.1042/bsr20191103
- Gotto, A. M. (2011). Jeremiah Metzger Lecture: Cholesterol, Inflammation and Atherosclerotic Cardiovascular Disease: Is it All LDL? *Trans. Am. Clin. Climatol Assoc.* 122, 256–289.
- Guo, B. H., Zhao, Q., and Li, H. Y. (2019). TUG1 Promotes the Development of Prostate Cancer by Regulating RLIM. *Eur. Rev. Med. Pharmacol. Sci.* 23, 1926–1933. doi:10.26355/eurrev_201903_17230

- Gupta, R. A., Shah, N., Wang, K. C., Kim, J., Horlings, H. M., Wong, D. J., et al. (2010). Long Non-coding RNA HOTAIR Reprograms Chromatin State to Promote Cancer Metastasis. *Nature* 464, 1071–1076. doi:10.1038/nature08975
- Hahne, J. C., and Valeri, N. (2018). Non-Coding RNAs and Resistance to Anticancer Drugs in Gastrointestinal Tumors. *Front. Oncol.* 8, 226. doi:10.3389/fonc.2018.00226
- Han, M., Lee, J., Kim, G., Lee, E., Lee, Y., Jang, S., et al. (2020). Expression of the Long Noncoding RNA GAS5 Correlates with Liver Fibrosis in Patients with Nonalcoholic Fatty Liver Disease. *Genes* 11. doi:10.3390/genes11050545
- Han, X., Huang, S., Xue, P., Fu, J., Liu, L., Zhang, C., et al. (2019). PTPRE-AS1 LncRNA Modulates M2 Macrophage Activation and Inflammatory Diseases by Epigenetic Promotion of PTPRE. *Sci. Adv.* 5, eaax9230. doi:10.1126/sciadv.aax9230
- He, W., Liang, B., Wang, C., Li, S., Zhao, Y., Huang, Q., et al. (2019). MSC-regulated lncRNA MACC1-AS1 Promotes Stemness and Chemoresistance through Fatty Acid Oxidation in Gastric Cancer. *Oncogene* 38, 4637–4654. doi:10.1038/s41388-019-0747-0
- Hennessy, E. J., van Solingen, C., Scacalossi, K. R., Ouimet, M., Afonso, M. S., Prins, J., et al. (2019). The Long Noncoding RNA CHROME Regulates Cholesterol Homeostasis in Primates. *Nat. Metab.* 1, 98–110. doi:10.1038/s42255-018-0004-9
- Hu, W., Ding, Y., Wang, S., Xu, L., and Yu, H. (2020). The Construction and Analysis of the Aberrant lncRNA-miRNA-mRNA Network in Adipose Tissue from Type 2 Diabetes Individuals with Obesity. *J. Diabetes Res.* 2020, 3980742. doi:10.1155/2020/3980742
- Hu, Y.-W., Guo, F.-X., Xu, Y.-J., Li, P., Lu, Z.-F., McVey, D. G., et al. (2019a). Long Noncoding RNA NEXN-AS1 Mitigates Atherosclerosis by Regulating the Actin-Binding Protein NEXN. *J. Clin. Invest.* 129, 1115–1128. doi:10.1172/jci98230
- Hu, Y., Jin, Y., Wu, X., Yang, Y., Li, Y., Li, H., et al. (2019b). LncRNA-HGBC Stabilized by HuR Promotes Gallbladder Cancer Progression by Regulating miR-502-3p/SET/AKT axis. *Mol. Cancer* 18, 167. doi:10.1186/s12943-019-1097-9
- Huang, C., Hu, Y.-W., Zhao, J.-J., Ma, X., Zhang, Y., Guo, F.-X., et al. (2016). Long Noncoding RNA HOXC-AS1 Suppresses Ox-LDL-Induced Cholesterol Accumulation through Promoting HOXC6 Expression in THP-1 Macrophages. *DNA Cel. Biol.* 35, 722–729. doi:10.1089/dna.2016.3422
- Huang, J., Chen, S., Cai, D., Bian, D., and Wang, F. (2018). Long Noncoding RNA lncARSR Promotes Hepatic Cholesterol Biosynthesis via Modulating Akt/SREBP-2/HMGCR Pathway. *Life Sci.* 203, 48–53. doi:10.1016/j.lfs.2018.04.028
- Huang, S., Tao, W., Guo, Z., Cao, J., and Huang, X. (2019a). Suppression of Long Noncoding RNA TTTY15 Attenuates Hypoxia-Induced Cardiomyocytes Injury by Targeting miR-455-5p. *Gene* 701, 1–8. doi:10.1016/j.gene.2019.02.098
- Huang, Y. (2018). The Novel Regulatory Role of lncRNA-miRNA-mRNA axis in Cardiovascular Diseases. *J. Cel Mol Med* 22, 5768–5775. doi:10.1111/jcmm.13866
- Huang, Y., Wang, L., Mao, Y., and Nan, G. (2019b). Long Noncoding RNA-H19 Contributes to Atherosclerosis and Induces Ischemic Stroke via the Upregulation of Acid Phosphatase 5. *Front. Neurol.* 10, 32. doi:10.3389/fneur.2019.00032
- Jacquemyn, J., Cascalho, A., and Goodchild, R. E. J. E. R. (2017). The Ins and Outs of Endoplasmic Reticulum \ ontrolled Lipid Biosynthesis, *EMBO Rep.*, 18, e201643426. doi:10.15252/embr.201643426
- Jeon, S.-M., Chandel, N. S., and Hay, N. (2012). AMPK Regulates NADPH Homeostasis to Promote Tumour Cell Survival during Energy Stress. *Nature* 485, 661–665. doi:10.1038/nature11066
- Jiang, C., Zhao, Q., Wang, C., Peng, M., Hao, G., Liu, Z., et al. (2021). Downregulation of Long Noncoding RNA LINC00261 Attenuates Myocardial Infarction through the miR-522-3p/Trinucleotide Repeat-Containing Gene 6a (TNRC6A) Axis. *Cardiovasc. Ther.* 2021, 6628194. doi:10.1155/2021/6628194
- Jiang, L., Shao, C., Wu, Q.-J., Chen, G., Zhou, J., Yang, B., et al. (2017). NEAT1 Scaffolds RNA-Binding Proteins and the Microprocessor to Globally Enhance Pri-miRNA Processing. *Nat. Struct. Mol. Biol.* 24, 816–824. doi:10.1038/nsmb.3455
- Jin, M., Feng, H., Wang, Y., Yan, S., Shen, B., Li, Z., et al. (2020). Gentiopicroside Ameliorates Oxidative Stress and Lipid Accumulation through Nuclear Factor Erythroid 2-Related Factor 2 Activation. *Oxidative Med. Cell. longevity* 2020, 2940746. doi:10.1155/2020/2940746
- Johnston, T. P., Korolenko, T. A., Pirro, M., and Sahebkar, A. (2017). Preventing Cardiovascular Heart Disease: Promising Nutraceutical and Non-nutraceutical Treatments for Cholesterol Management. *Pharmacol. Res.* 120, 219–225. doi:10.1016/j.phrs.2017.04.008
- Khor, V. K., Shen, W.-J., Kraemer, F. B., and Care, M. (2013). Lipid Droplet Metabolism. *Curr. Opin. Clin. Nutr. Metab. Care* 16, 632–637. doi:10.1097/mco.0b013e32832651106
- Kim, V. N., Han, J., and Siomi, M. C. (2009). Biogenesis of Small RNAs in Animals. *Nat. Rev. Mol. Cel Biol* 10, 126–139. doi:10.1038/nrm2632
- Kogo, R., Shimamura, T., Mimori, K., Kawahara, K., Imoto, S., Sudo, T., et al. (2011). Long Noncoding RNA HOTAIR Regulates Polycomb-dependent Chromatin Modification and Is Associated with Poor Prognosis in Colorectal Cancers. *Cancer Res.* 71, 6320–6326. doi:10.1158/0008-5472.can-11-1021
- Kopp, F., and Mendell, J. T. (2018). Functional Classification and Experimental Dissection of Long Noncoding RNAs. *Cell* 172, 393–407. doi:10.1016/j.cell.2018.01.011
- Kung, J. T. Y., Colognori, D., and Lee, J. T. (2013). Long Noncoding RNAs: Past, Present, and Future. *Genetics* 193, 651–669. doi:10.1534/genetics.112.146704
- Lan, X., Wu, L., Wu, N., Chen, Q., Li, Y., Du, X., et al. (2019). Long Noncoding RNA Lnc-HC Regulates PPAR γ -Mediated Hepatic Lipid Metabolism through miR-130b-3p. *Mol. Ther. - Nucleic Acids* 18, 954–965. doi:10.1016/j.omtn.2019.10.018
- Lan, X., Yan, J., Ren, J., Zhong, B., Li, J., Li, Y., et al. (2016). A Novel Long Noncoding RNA Lnc-HC Binds hnRNPA2B1 to Regulate Expressions of Cyp7a1 and Abca1 in Hepatocytic Cholesterol Metabolism. *Hepatology* 64, 58–72. doi:10.1002/hep.28391
- Lee, K., Hwang, H., and Cho, J. (2020). Long Non-coding RNA Associated with Cholesterol Homeostasis and its Involvement in Metabolic Diseases. *Int. J. Mol. Sci.* 21. doi:10.3390/ijms21218337
- Leti, F., Legendre, C., Still, C. D., Chu, X., Petrick, A., Gerhard, G. S., et al. (2017). Altered Expression of MALAT1 lncRNA in Nonalcoholic Steatohepatitis Fibrosis Regulates CXCL5 in Hepatic Stellate Cells. *Translational Res.* 190, 25–39. doi:10.1016/j.trsl.2017.09.001
- Li, F. P., Lin, D. Q., and Gao, L. Y. (2018). LncRNA TUG1 Promotes Proliferation of Vascular Smooth Muscle Cell and Atherosclerosis through Regulating miRNA-21/PEN axis. *Eur. Rev. Med. Pharmacol. Sci.* 22, 7439–7447. doi:10.26355/eurrev_201811_16284
- Li, J.-z., Ye, L.-h., Wang, D.-h., Zhang, H.-c., Li, T.-y., Liu, Z.-q., et al. (2021b). The Identify Role and Molecular Mechanism of the MALAT1/hsa-Mir-20b-5p/TXNIP axis in Liver Inflammation Caused by CHB in Patients with Chronic HBV Infection Complicated with NAFLD. *Virus. Res.* 298, 198405. doi:10.1016/j.virusres.2021.198405
- Li, J., Tong, Y., Zhou, Y., Han, Z., Wang, X., Ding, T., et al. (2021a). LncRNA KCNQ1OT1 as a miR-26a-5p Sponge Regulates ATG12-Mediated Cardiomyocyte Autophagy and Aggravates Myocardial Infarction. *Int. J. Cardiol.*
- Li, P., Yan, X., Xu, G., Pang, Z., Weng, J., Yin, J., et al. (2020a). A Novel Plasma lncRNA ENST00000416361 Is Upregulated in Coronary Artery Disease and Is Related to Inflammation and Lipid Metabolism. *Mol. Med. Rep.* 21, 2375–2384. doi:10.3892/mmr.2020.11067
- Li, X., Li, L., Dong, X., Ding, J., Ma, H., and Han, W. (2021c). Circ_GRN Promotes the Proliferation, Migration, and Inflammation of Vascular Smooth Muscle Cells in Atherosclerosis through miR-214-3p/FOXO1 Axis. *J. Cardiovasc. Pharmacol.* 77, 470–479. doi:10.1097/fjc.0000000000000982
- Li, Y., Ma, Z., Jiang, S., Hu, W., Li, T., Di, S., et al. (2017). A Global Perspective on FOXO1 in Lipid Metabolism and Lipid-Related Diseases. *Prog. Lipid Res.* 66, 42–49. doi:10.1016/j.plipres.2017.04.002
- Li, Y., Zhang, D., Zhang, Y., Xu, X., Bi, L., Zhang, M., et al. (2020b). Association of lncRNA Polymorphisms with Triglyceride and Total Cholesterol Levels Among Myocardial Infarction Patients in Chinese Population. *Gene* 724, 143684. doi:10.1016/j.gene.2019.02.085
- Lin, L., Li, H., Zhu, Y., He, S., and Ge, H. (2018). Expression of Metastasis-Associated Lung Adenocarcinoma Transcript 1 Long Non-coding RNA *In Vitro* and in Patients with Non-small Cell Lung Cancer. *Oncol. Lett.* 15, 9443–9449. doi:10.3892/ol.2018.8531

- Liu, S., Cao, Q., An, G., Yan, B., and Lei, L. (2020). Identification of the 3-lncRNA Signature as a Prognostic Biomarker for Colorectal Cancer. *Int. J. Mol. Sci.* 21. doi:10.3390/ijms21249359
- Liu, S., Horlbeck, M., Cho, S., Birk, H., Malatesta, M., He, D., et al. (2017). *CRISPRi-Based Genome-Scale Identification of Functional Long Noncoding RNA Loci in Human Cells*. New York, N.Y.: Science, 355.
- Liu, S., Xu, D.-s., Li, M., Zhang, Y., Li, Q., Li, T.-t., et al. (2021a). Icarin Attenuates Endothelial-Mesenchymal Transition via H19/miR-148b-3p/ELF5 in Ox-LDL-Stimulated HUVECs. *Mol. Ther. - Nucleic Acids* 23, 464–475. doi:10.1016/j.omtn.2020.11.021
- Liu, Y., and Ding, Z. (2017). Obesity, a Serious Etiologic Factor for Male Subfertility in Modern Society. *Reproduction (Cambridge, England)* 154, R123–R131. doi:10.1530/rep-17-0161
- Liu, Y., Meng, W., Cao, H., and Wang, B. (2021b). Identification of MSC-AS1, a Novel lncRNA for the Diagnosis of Laryngeal Cancer. *Eur. Arch. Otorhinolaryngol.* 278, 1107–1118. doi:10.1007/s00405-020-06427-4
- Loewen, G., Jayawickramarajah, J., Zhuo, Y., and Shan, B. (2014). Functions of lncRNA HOTAIR in Lung Cancer. *J. Hematol. Oncol.* 7, 90. doi:10.1186/s13045-014-0090-4
- Lovren, F., Teoh, H., and Verma, S. (2015). Obesity and Atherosclerosis: Mechanistic Insights. *Can. J. Cardiol.* 31, 177–183. doi:10.1016/j.cjca.2014.11.031
- Luzón-Toro, B., Fernández, R., Martos-Martínez, J., Rubio-Manzanares-Dorado, M., Antiñolo, G., and Borrego, S. (2019). lncRNA LUCAT1 as a Novel Prognostic Biomarker for Patients with Papillary Thyroid Cancer. *Scientific Rep.* 9, 14374. doi:10.1038/s41598-019-50913-7
- Ma, Y., Zhang, J., Wen, L., and Lin, A. (2018). Membrane-lipid Associated lncRNA: A New Regulator in Cancer Signaling. *Cancer Lett.* 419, 27–29. doi:10.1016/j.canlet.2018.01.008
- Mannu, G. S., Zaman, M. J., Gupta, A., Rehman, H. U., and Myint, P. K. (2013). Evidence of Lifestyle Modification in the Management of Hypercholesterolemia. *Curr. Cardiol. Rev.* 9, 2–14. doi:10.2174/157340313805076313
- Maruyama, R., and Yokota, T. (2020). Knocking Down Long Noncoding RNAs Using Antisense Oligonucleotide Gapmers. *Methods Mol. Biol. (Clifton, N.J.)* 2176, 49–56. doi:10.1007/978-1-0716-0771-8_3
- Michos, E. D., McEvoy, J. W., and Blumenthal, R. S. (2019). Lipid Management for the Prevention of Atherosclerotic Cardiovascular Disease. *N. Engl. J. Med.* 381, 1557–1567. doi:10.1056/nejmra1806939
- Mumtaz, P. T., and Online, S. B. J. B. P. (2017). *lncRNAs and Immunity: Watchdogs for Host Pathogen Interactions*, 19. doi:10.1186/s12575-017-0052-7
- Musso, G., Gambino, R., and Cassader, M. (2013). Cholesterol Metabolism and the Pathogenesis of Non-alcoholic Steatohepatitis. *Prog. lipid Res.* 52, 175–191. doi:10.1016/j.plipres.2012.11.002
- Niu, Y., Ma, F., Huang, W., Fang, S., Li, M., Wei, T., et al. (2017). Long Non-coding RNA TUG1 Is Involved in Cell Growth and Chemoresistance of Small Cell Lung Cancer by Regulating LIMK2b via EZH2. *Mol. Cancer* 16, 5. doi:10.1186/s12943-016-0575-6
- Osielska, M. A., and Jagodziński, P. P. (2018). Long Non-coding RNA as Potential Biomarkers in Non-small-cell Lung Cancer: What Do We Know So Far? *Biomed. Pharmacother.* 101, 322–333. doi:10.1016/j.biopha.2018.02.099
- Ou, M., Li, X., Zhao, S., Cui, S., and Tu, J. (2020). Long Non-coding RNA CDKN2B-AS1 Contributes to Atherosclerotic Plaque Formation by Forming RNA-DNA Triplex in the CDKN2B Promoter. *EBioMedicine* 55, 102694. doi:10.1016/j.ebiom.2020.102694
- Pan, J. X. (2017). lncRNA H19 Promotes Atherosclerosis by Regulating MAPK and NF-κB Signaling Pathway. *Eur. Rev. Med. Pharmacol. Sci.* 21, 322–328.
- Pan, J. X., and sciences, p. (2017). lncRNA H19 Promotes Atherosclerosis by Regulating MAPK and NF-κB Signaling Pathway. *Eur. Rev. Med. Pharmacol. Sci.* 21, 322–328.
- Pan, X., Cheng, R., Zhu, X., Cai, F., Zheng, G., Li, J., et al. (2019). Prognostic Significance and Diagnostic Value of Overexpressed lncRNA PVT1 in Colorectal Cancer. *Clin. Lab.* 65. doi:10.7754/clin.lab.2019.190412
- Paredes, S., Fonseca, L., Ribeiro, L., Ramos, H., Oliveira, J., and Palma, I. (2019). Novel and Traditional Lipid Profiles in Metabolic Syndrome Reveal a High Atherogenicity. *Scientific Rep.* 9, 11792. doi:10.1038/s41598-019-48120-5
- Park, J., Kim, G., Jang, S., Lee, Y., Lee, E., Lee, H., et al. (2020). *LeXisPlasma Long Noncoding RNA Is a Potential Diagnostic Marker for Non-alcoholic Steatohepatitis*, 10. Basel, Switzerland: Life. doi:10.3390/life10100230
- Patil, D. P., Chen, C.-K., Pickering, B. F., Chow, A., Jackson, C., Guttman, M., et al. (2016). m6A RNA Methylation Promotes XIST-Mediated Transcriptional Repression. *Nature* 537, 369–373. doi:10.1038/nature19342
- Pedram Fatemi, R., Salah-Uddin, S., Modarresi, F., Khoury, N., Wahlestedt, C., and Faghihi, M. A. (2015). Screening for Small-Molecule Modulators of Long Noncoding RNA-Protein Interactions Using AlphaScreen. *J. Biomol. Screen.* 20, 1132–1141. doi:10.1177/1087057115594187
- Peng, L., Jiang, J., Tang, B., Nice, E. C., Zhang, Y.-Y., and Xie, N. (2020). Managing Therapeutic Resistance in Breast Cancer: from the lncRNAs Perspective. *Theranostics* 10, 10360–10377. doi:10.7150/thno.49922
- Piccinin, E., Cariello, M., and Moschetta, A. (2021). Lipid Metabolism in colon Cancer: Role of Liver X Receptor (LXR) and Stearoyl-CoA Desaturase 1 (SCD1). *Mol. aspects Med.* 78, 100933. doi:10.1016/j.mam.2020.100933
- Plączkowska, S., Pawlik-Sobecka, L., et al. Plączkowska, S., Pawlik-Sobecka, L., Kokot, I., Piwowar, A. (2014). [Incidence of Complex Metabolic Disorders Among Young People-Preliminary Report]. *Pol. Merkur Lekarski* 37, 269–273.
- Ratti, M., Lampis, A., Ghidini, M., Salati, M., Mirchev, M. B., Valeri, N., et al. (2020). MicroRNAs (miRNAs) and Long Non-coding RNAs (lncRNAs) as New Tools for Cancer Therapy: First Steps from Bench to Bedside. *Targ Oncol.* 15, 261–278. doi:10.1007/s11523-020-00717-x
- Rinn, J. L., and Chang, H. Y. (2012). Genome Regulation by Long Noncoding RNAs. *Annu. Rev. Biochem.* 81, 145–166. doi:10.1146/annurev-biochem-051410-092902
- Sallam, T., Jones, M. C., Gilliland, T., Zhang, L., Wu, X., Eskin, A., et al. (2016). Feedback Modulation of Cholesterol Metabolism by the Lipid-Responsive Non-coding RNA LeXis. *Nature* 534, 124–128. doi:10.1038/nature17674
- Sallam, T., Jones, M., Thomas, B. J., Wu, X., Gilliland, T., Qian, K., et al. (2018). Transcriptional Regulation of Macrophage Cholesterol Efflux and Atherogenesis by a Long Noncoding RNA. *Nat. Med.* 24, 304–312. doi:10.1038/nm.4479
- Santos, C. R., and Schulze, A. (2012). Lipid Metabolism in Cancer. *FEBS J.* 279, 2610–2623. doi:10.1111/j.1742-4658.2012.08644.x
- Shen, X.-h., Qi, P., and Du, X. (2015). Long Non-coding RNAs in Cancer Invasion and Metastasis. *Mod. Pathol.* 28, 4–13. doi:10.1038/modpathol.2014.75
- Shi, X., Wei, Y., Li, H., Jiang, T., Zheng, X., Yin, K., et al. (2020). Long Non-coding RNA H19 in Atherosclerosis: what Role? *Mol. Med. (Cambridge, Mass)* 26, 72. doi:10.1186/s10020-020-00196-w
- Shimano, H., and Sato, R. (2017). SREBP-regulated Lipid Metabolism: Convergent Physiology - Divergent Pathophysiology. *Nat. Rev. Endocrinol.* 13, 710–730. doi:10.1038/nrendo.2017.91
- Simion, V., Haemmig, S., and Feinberg, M. W. (2019). lncRNAs in Vascular Biology and Disease. *Vasc. Pharmacol.* 114, 145–156. doi:10.1016/j.vph.2018.01.003
- Simion, V., Zhou, H., Haemmig, S., Pierce, J. B., Mendes, S., Tesmenitsky, Y., et al. (2020). A Macrophage-specific lncRNA Regulates Apoptosis and Atherosclerosis by Tethering HuR in the Nucleus. *Nat. Commun.* 11, 6135. doi:10.1038/s41467-020-19664-2
- Spiroski, A.-M., Sanders, R., Meloni, M., McCracken, I. R., Thomson, A., Brittan, M., et al. (2021). The Influence of the LINC00961/SPAAR Locus Loss on Murine Development, Myocardial Dynamics, and Cardiac Response to Myocardial Infarction. *Int. J. Mol. Sci.* 22. doi:10.3390/ijms22020969
- Stackhouse, C., Gillespie, G., and Willey, C. (2020). Exploring the Roles of lncRNAs in GBM Pathophysiology and Their Therapeutic Potential. *Cells* 9, 236910.3390/cells9112369.
- Sukhorukov, V. N., Khotina, V. A., Chegodaev, Y. S., Ivanova, E., Sobenin, I. A., and Orekhov, A. N. (2020). Lipid Metabolism in Macrophages: Focus on Atherosclerosis. *Biomedicines* 8. doi:10.3390/biomedicines8080262
- Sun, C., Fu, Y., Gu, X., Xi, X., Peng, X., Wang, C., et al. (2020). Macrophage-Enriched lncRNA RAPIA. *Atvb* 40, 1464–1478. doi:10.1161/atvbaha.119.313749
- Sun, D., Yu, Z., Fang, X., Liu, M., Pu, Y., Shao, Q., et al. (2017). lnc RNA GAS 5 Inhibits Microglial M2 Polarization and Exacerbates Demyelination. *EMBO Rep.* 18, 1801–1816. doi:10.15252/embr.201643668
- Sun, R., and Zhang, L. (2019). Long Non-coding RNA MALAT1 Regulates Cardiomyocytes Apoptosis after Hypoxia/reperfusion Injury via Modulating miR-200a-3p/PDCD4 axis. *Biomed. Pharmacother.* 111, 1036–1045. doi:10.1016/j.biopha.2018.12.122

- Tao, K., Hu, Z., Zhang, Y., Jiang, D., and Cheng, H. (2019). LncRNA CASC11 Improves Atherosclerosis by Downregulating IL-9 and Regulating Vascular Smooth Muscle Cell Apoptosis and Proliferation. *Biosci. Biotechnol. Biochem.* 83, 1284–1288. doi:10.1080/09168451.2019.1597621
- Taufiq, Z., Pino, P., N'Dilimabaka, N., Arrouss, I., Assi, S., Soubrier, F., et al. (2011). Atorvastatin Prevents Plasmodium Falciparum Cytoadherence and Endothelial Damage. *Malar. J.* 10, 52. doi:10.1186/1475-2875-10-52
- Tontonoz, P., Wu, X., Jones, M., Zhang, Z., Salisbury, D., and Sallam, T. (2017). Long Noncoding RNA Facilitated Gene Therapy Reduces Atherosclerosis in a Murine Model of Familial Hypercholesterolemia. *Circulation* 136, 776–778. doi:10.1161/circulationaha.117.029002
- Tran, K.-V., Brown, E. L., DeSouza, T., Jespersen, N. Z., Nandrup-Bus, C., Yang, Q., et al. (2020). Human Thermogenic Adipocyte Regulation by the Long Noncoding RNA LINC00473. *Nat. Metab.* 2, 397–412. doi:10.1038/s42255-020-0205-x
- Vernon, G., Baranova, A., and Younossi, Z. M. (2011). Systematic Review: the Epidemiology and Natural History of Non-alcoholic Fatty Liver Disease and Non-alcoholic Steatohepatitis in Adults. *Aliment. Pharmacol. Ther.* 34, 274–285. doi:10.1111/j.1365-2036.2011.04724.x
- Wan, S., and Zhao, H. (2020). Analysis of Diagnostic and Prognostic Value of lncRNA MEG3 in Cervical Cancer. *Oncol. Lett.* 20, 183. doi:10.3892/ol.2020.12044
- Wang, C., Shi, H., Chen, L., Li, X., Cao, G., and Hu, X. (2019). Identification of Key lncRNAs Associated with Atherosclerosis Progression Based on Public Datasets. *Front. Genet.* 10, 123. doi:10.3389/fgene.2019.00123
- Wang, H., Gong, H., Liu, Y., and Feng, L. (2020a). Relationship between lncRNA-Ang362 and Prognosis of Patients with Coronary Heart Disease after Percutaneous Coronary Intervention. *Biosci. Rep.* 40. doi:10.1042/bsr20201524
- Wang, J., Yang, W., Chen, Z., Chen, J., Meng, Y., Feng, B., et al. (2018). Long Noncoding RNA lncSHGL Recruits hnRNPA1 to Suppress Hepatic Gluconeogenesis and Lipogenesis. *Diabetes* 67, 581–593. doi:10.2337/db17-0799
- Wang, S., Moustaid-Moussa, N., Chen, L., Mo, H., Shastri, A., Su, R., et al. (2014). Novel Insights of Dietary Polyphenols and Obesity. *J. Nutr. Biochem.* 25, 1–18. doi:10.1016/j.jnutbio.2013.09.001
- Wang, X., Wang, L., Ma, Z., Liang, W., Li, J., Li, Y., et al. (2020b). Early Expressed Circulating Long Noncoding RNA CHAT Is Associated with Cardiac Contractile Function in Patients with Acute Myocardial Infarction. *Int. J. Cardiol.* 302, 15–20. doi:10.1016/j.ijcard.2019.12.058
- Wang, Z., Yang, X., Kai, J., Wang, F., Wang, Z., Shao, J., et al. (2020c). HIF-1 α -upregulated lncRNA-H19 Regulates Lipid Droplet Metabolism through the AMPK α Pathway in Hepatic Stellate Cells. *Life Sci.* 255, 117818. doi:10.1016/j.lfs.2020.117818
- Wen, X., Ding, Y., Tan, Z., Wang, J., Zhang, D., and Wang, Y. (2020). Identification and Characterization of Cadmium Stress-Related lncRNAs from *Betula platyphylla*. *Plant Sci.* 299, 110601. doi:10.1016/j.plantsci.2020.110601
- Wu, D., Zhou, Y., Fan, Y., Zhang, Q., Gu, F., Mao, W., et al. (2019). LncRNA CAIF Was Downregulated in End-Stage Cardiomyopathy and Is a Promising Diagnostic and Prognostic Marker for This Disease. *Biomarkers* 24, 735–738. doi:10.1080/1354750x.2019.1677778
- Xiao, B., Huang, Z., Zhou, R., Zhang, J., and Yu, B. (2018). The Prognostic Value of Expression of the Long Noncoding RNA (lncRNA) Small Nucleolar RNA Host Gene 1 (SNHG1) in Patients with Solid Malignant Tumors: A Systematic Review and Meta-Analysis. *Med. Sci. Monit.* 24, 5462–5472. doi:10.12659/msm.911687
- Xiao, X., and Song, B.-L. (2013). SREBP: a Novel Therapeutic Target. *Acta Biochim. Biophys. Sinica* 45, 2–10. doi:10.1093/abbs/gms112
- Xie, H., Ma, H., and Zhou, D. (2013). Plasma HULC as a Promising Novel Biomarker for the Detection of Hepatocellular Carcinoma. *Biomed. Research International* 2013, 136106. doi:10.1155/2013/136106
- Xie, J., Wang, X., Liu, S., Chen, C., Jiang, F., Mao, K., et al. (2019). LncRNA SAMMSON Overexpression Distinguished Glioblastoma Patients from Patients with Diffuse Neurosarcoidosis. *Neuroreport* 30, 817–821. doi:10.1097/wnr.0000000000001278
- Xing, C., Sun, S.-G., Yue, Z.-Q., and Bai, F. (2021). Role of lncRNA LUCAT1 in Cancer. *Biomed. Pharmacother.* 134, 111158. doi:10.1016/j.biopha.2020.111158
- Xu, D., Wang, Z., Xia, Y., Shao, F., Xia, W., Wei, Y., et al. (2020). The Gluconeogenic Enzyme PCK1 Phosphorylates INSIG1/2 for Lipogenesis. *Nature* 580, 530–535. doi:10.1038/s41586-020-2183-2
- Xu, J., and Taubert, S. (2021). Beyond Proteostasis: Lipid Metabolism as a New Player in ER Homeostasis. *Metabolites* 11, 52. doi:10.3390/metabo11010052
- Yan, C., Chen, J., and Chen, N. (2016). Long Noncoding RNA MALAT1 Promotes Hepatic Steatosis and Insulin Resistance by Increasing Nuclear SREBP-1c Protein Stability. *Scientific Rep.* 6, 22640. doi:10.1038/srep22640
- Yan, C., Zhang, Y., Zhang, X., Aa, J., Wang, G., and Xie, Y. (2018). Curcumin Regulates Endogenous and Exogenous Metabolism via Nrf2-FXR-LXR Pathway in NAFLD Mice. *Biomed. Pharmacother.* 105, 274–281. doi:10.1016/j.biopha.2018.05.135
- Yan, L., Zhang, Y., Zhang, W., Deng, S.-Q., and Ge, Z.-R. (2020). lncRNA-NRF Is a Potential Biomarker of Heart Failure after Acute Myocardial Infarction. *J. Cardiovasc. Trans. Res.* 13, 1008–1015. doi:10.1007/s12265-020-10029-0
- Yang, Y., Li, Y., Ma, Z., Jiang, S., Fan, C., Hu, W., et al. (2016). A Brief Glimpse at CTRP3 and CTRP9 in Lipid Metabolism and Cardiovascular protection. *Prog. Lipid Res.* 64, 170–177. doi:10.1016/j.plipres.2016.10.001
- Yang, Y., Tang, F., Wei, F., Yang, L., Kuang, C., Zhang, H., et al. (2019a). Silencing of Long Non-coding RNA H19 Downregulates CTCF to Protect against Atherosclerosis by Upregulating PKD1 Expression in ApoE Knockout Mice. *Aging* 11, 10016–10030. doi:10.18632/aging.102388
- Yang, Z., Sun, Y., Liu, R., Shi, Y., and Ding, S. (2019b). Plasma Long Noncoding RNAs PANDAR, FOXD2-AS1, and SMARCC2 as Potential Novel Diagnostic Biomarkers for Gastric Cancer. *Cmar Vol.* 11, 6175–6184. doi:10.2147/cmar.s201935
- Yang, Z., Zhang, T., Han, S., Kusumanchi, P., Huda, N., Jiang, Y., et al. (2021). Long Noncoding RNA H19 - a New Player in the Pathogenesis of Liver Diseases. *Translational Res.* 230, 139–150. doi:10.1016/j.trsl.2020.11.010
- Yao, X., Yan, C., Zhang, L., Li, Y., and Wan, Q. (2018). LncRNA ENST00113 Promotes Proliferation, Survival, and Migration by Activating PI3K/Akt/mTOR Signaling Pathway in Atherosclerosis. *Medicine* 97, e0473. doi:10.1097/md.00000000000010473
- Yao, Y., Zhang, T., Qi, L., Zhou, C., Wei, J., Feng, F., et al. (2019). Integrated Analysis of Co-expression and ceRNA Network Identifies Five lncRNAs as Prognostic Markers for Breast Cancer. *J. Cel Mol Med* 23, 8410–8419. doi:10.1111/jcmm.14721
- Ye, J., Lin, Y., Yu, Y., and Sun, D. (2020). LncRNA NEAT1/microRNA-129-5p/SOCS2 axis Regulates Liver Fibrosis in Alcoholic Steatohepatitis. *J. translational Med.* 18, 445. doi:10.1186/s12967-020-02577-5
- Ye, W., Huang, S., Hou, L., Long, H., Yin, K., Hu, C., et al. (2021). Potential Therapeutic Targeting of lncRNAs in Cholesterol Homeostasis. *Front. Cardiovasc. Med.* 8, 688546. doi:10.3389/fcvm.2021.688546
- Ye, Z., Yang, S., Xia, Y., Hu, R., Chen, S., Li, B., et al. (2019). LncRNA MIAT Sponges miR-149-5p to Inhibit Efferocytosis in Advanced Atherosclerosis through CD47 Upregulation. *Cel Death Dis.* 10, 138. doi:10.1038/s41419-019-1409-4
- Yoon, J.-H., Abdelmohsen, K., Srikantan, S., Yang, X., Martindale, J. L., De, S., et al. (2012). LincRNA-p21 Suppresses Target mRNA Translation. *Mol. Cel.* 47, 648–655. doi:10.1016/j.molcel.2012.06.027
- Yörüker, E. E., Keskin, M., Kulle, C. B., Holdenrieder, S., and Gezer, U. (2018). Diagnostic and Prognostic Value of Circulating lncRNA H19 in Gastric Cancer. *Biomed. Rep.* 9, 181–186. doi:10.3892/br.2018.1116
- Yu, C., Li, L., Xie, F., Guo, S., Liu, F., Dong, N., et al. (2018). LncRNA TUG1 Sponges miR-204-5p to Promote Osteoblast Differentiation through Upregulating Runx2 in Aortic Valve Calcification. *Cardiovasc. Res.* 114, 168–179. doi:10.1093/cvr/cvx180
- Yu, F., Zheng, J., Mao, Y., Dong, P., Li, G., Lu, Z., et al. (2015). Long Non-coding RNA APTR Promotes the Activation of Hepatic Stellate Cells and the Progression of Liver Fibrosis. *Biochem. biophysical Res. Commun.* 463, 679–685. doi:10.1016/j.bbrc.2015.05.124
- Yu, H., Ma, S., Sun, L., Gao, J., and Zhao, C. (2019). TGF- β 1 U-pregulates the E-xpression of lncRNA-ATB to P-romote A-therosclerosis. *Mol. Med. Rep.* 19, 4222–4228. doi:10.3892/mmr.2019.10109
- Yu, X., Li, Z., Zheng, H., Chan, M., and Wu, W. (2017a). NEAT1: A Novel Cancer-Related Long Non-coding RNA. *Cel Prolif.* 50. doi:10.1111/cpr.12329
- Yu, Y., Nangia-Makker, P., Farhana, L., and Majumdar, A. (2017b). A Novel Mechanism of lncRNA and miRNA Interaction: CCAT2 Regulates miR-145 Expression by Suppressing its Maturation Process in colon Cancer Cells. *Mol. Cancer* 16, 155. doi:10.1186/s12943-017-0725-5

- Zeng, Y., Ren, K., Zhu, X., Zheng, Z., and Yi, G. (2018). Long Noncoding RNAs: Advances in Lipid Metabolism. *Adv. Clin. Chem.* 87, 1–36. doi:10.1016/bs.acc.2018.07.001
- Zhang, D., Zeng, S., and Hu, X. (2020a). Identification of a Three-Long Noncoding RNA Prognostic Model Involved Competitive Endogenous RNA in Kidney Renal clear Cell Carcinoma. *Cancer Cel. Int.* 20, 319. doi:10.1186/s12935-020-01423-4
- Zhang, L., Xu, W., Gao, X., Li, W., Qi, S., Guo, D., et al. (2020b). lncRNA Sensing of a Viral Suppressor of RNAi Activates Non-canonical Innate Immune Signaling in *Drosophila*. *Cell Host & Microbe* 27, 115–128. doi:10.1016/j.chom.2019.12.006
- Zhang, M., Chi, X., Qu, N., and Wang, C. (2018). Long Noncoding RNA lncARSR Promotes Hepatic Lipogenesis via Akt/SREBP-1c Pathway and Contributes to the Pathogenesis of Nonalcoholic Steatohepatitis. *Biochem. biophysical Res. Commun.* 499, 66–70. doi:10.1016/j.bbrc.2018.03.127
- Zhang, T., Liu, H., Mao, R., Yang, H., Zhang, Y., Zhang, Y., et al. (2020c). The lncRNA RP11-142A22.4 Promotes Adipogenesis by Sponging miR-587 to Modulate Wnt5 β Expression. *Cel Death Dis.* 11, 475. doi:10.1038/s41419-020-2550-9
- Zhao, J., Chen, F., Ma, W., and Zhang, P. (2020). Suppression of Long Noncoding RNA NEAT1 Attenuates Hypoxia-Induced Cardiomyocytes Injury by Targeting miR-378a-3p. *Gene* 731, 144324. doi:10.1016/j.gene.2019.144324
- Zhao, X., Xiong, X., Liu, T., Mi, L., Peng, X., Rui, C., et al. (2018). Long Noncoding RNA Licensing of Obesity-Linked Hepatic Lipogenesis and NAFLD Pathogenesis. *Nat. Commun.* 9, 2986. doi:10.1038/s41467-018-05383-2
- Zheng, R., Du, M., Wang, X., Xu, W., Liang, J., Wang, W., et al. (2018). Exosome-transmitted Long Non-coding RNA PTENP1 Suppresses Bladder Cancer Progression. *Mol. Cancer* 17, 143. doi:10.1186/s12943-018-0880-3
- Zheng, X., Tian, X., Zhang, Q., Shi, P., and Li, S. (2020). Long Non-coding RNA SAMMSON as a Novel Potential Diagnostic and Prognostic Biomarker for Oral Squamous Cell Carcinoma. *J. dental Sci.* 15, 329–335. doi:10.1016/j.jds.2019.11.008
- Zhou, L., Yu, M., Arshad, M., Wang, W., Lu, Y., Gong, J., et al. (2018). Coordination Among Lipid Droplets, Peroxisomes, and Mitochondria Regulates Energy Expenditure through the CIDE-ATGL-Ppar α Pathway in Adipocytes. *Diabetes* 67, 1935–1948. doi:10.2337/db17-1452
- Zhuo, L., Wen, Y., Wang, Y., Liang, Z., Wu, G., Nong, M., et al. (2019). lncRNA SNHG8 Is Identified as a Key Regulator of Acute Myocardial Infarction by RNA-Seq Analysis. *Lipids Health Dis.* 18, 201. doi:10.1186/s12944-019-1142-0
- Zuo, X., Chen, Z., Gao, W., Zhang, Y., Wang, J., Wang, J., et al. (2020). M6A-mediated Upregulation of LINC00958 Increases Lipogenesis and Acts as a Nanotherapeutic Target in Hepatocellular Carcinoma. *J. Hematol. Oncol.* 13, 5. doi:10.1186/s13045-019-0839-x

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GLOSSARY

AMI Acute myocardial infarction	SQLE squalene epoxidase
ASH Alcoholic steatohepatitis	TNRC6A trinucleotide repeat-containing gene 6a
ATG12 autophagy-related 12 homologs	BC bladder cancer
CAIF Cardiac autophagy inhibitory factor	CYTOR cytoskeleton regulator RNA
CASIMO1 Cancer-associated small integral membrane open reading frame 1	FAM83H-AS1 FAM83H antisense RNA 1
CAVD calcific aortic valve disease	FOXD2-AS1 FOXD2 adjacent opposite strand RNA 1
CHAST cardiac hypertrophy-associated transcript	GBM glioblastoma
HF heart failure	GC gastric cancer
HMGCR HMG-CoA reductase	HCC hepatocellular carcinoma
HOXC6 homeobox C6	KIRC kidney renal clear cell carcinoma
HOXC-AS1 lncRNA HOXC cluster antisense RNA 1	LC laryngeal cancer
ITGB1 integrin β 1	LncRNA-ATB lncRNA activated by TGF β
JDP2 Jun dimerization protein 2	CRC colorectal cancer
LDLR low-density lipoprotein receptor	LPM lncRNA prognostic model
LRP2BP low-density lipoprotein related receptor 2 binding protein	LUAD lung adenocarcinoma
MALAT1 metastasis-associated lung adenocarcinoma transcript 1	MSC-AS1 MSC antisense RNA 1
MIAT myocardial infarction associated transcript	OSCC oral squamous cell carcinoma
MMP1 matrix metalloproteinase 1	PANDAR promoter of CDKN1A antisense DNA damage activated RNA
NEXN nexilin F-actin binding protein	PCa prostate cancer
NEXN-AS1 nexilin F-actin binding protein antisense RNA 1	PTC papillary thyroid cancer
PDCD4 programmed cell death 4	PTENP1 phosphatase and tensin homolog pseudogene 1
PPARA proliferator-activated receptor alpha	SAMMSON survival associated mitochondrial melanoma-specific oncogenic non-coding RNA
RAPIA associated with the progression and intervention of atherosclerosis	SMARCC2 SWI/SNF related, matrix associated, actin-dependent regulator of chromatin subfamily c member 2
SOCS2 cytokine signalling 2	SNHG18 small nucleolar RNA host gene 18