

Review

Towards Next Generation Biomarkers in Natural Killer/T-Cell Lymphoma

Jason Yongsheng Chan ^{1,2,*}, Jing Quan Lim ^{3,4}  and Choon Kiat Ong ^{3,4,5,*} ¹ Division of Medical Oncology, National Cancer Centre Singapore, Singapore 169610, Singapore² SingHealth Duke-NUS Blood Cancer Centre, Singapore 169857, Singapore³ Lymphoma Genomic Translational Research Laboratory, Division of Cellular and Molecular Research, National Cancer Centre Singapore, Singapore 169610, Singapore; lim.jing.quan@nccs.com.sg⁴ Cancer and Stem Cell Biology, Duke-NUS Medical School, Singapore 169857, Singapore⁵ Genome Institute of Singapore, A*STAR (Agency for Science, Technology and Research), Singapore 138672, Singapore

* Correspondence: jason.chan.y.s@singhealth.com.sg (J.Y.C.); cmrock@nccs.com.sg (C.K.O.)

Abstract: Natural killer/T-cell lymphoma (NKTCL) is an Epstein–Barr virus-associated non-Hodgkin lymphoma linked to an aggressive clinical course and poor prognosis. Despite an improvement in survival outcomes with the incorporation of novel agents including immune checkpoint inhibitors in the treatment of NKTCL, a significant proportion of patients still relapse or remain refractory to treatment. Several clinical prognostic models have been developed for NKTCL patients treated in the modern era, though the optimal approach to risk stratification remains to be determined. Novel molecular biomarkers derived from multi-omic profiling have recently been developed, with the potential to improve diagnosis, prognostication and treatment of this disease. Notably, a number of potential biomarkers have emerged from a better understanding of the tumor immune microenvironment and inflammatory responses. This includes a recently described 3'UTR structural variant in the *PD-L1* gene, which confers susceptibility to checkpoint immunotherapy. In this review, we summarize the biomarker landscape of NKTCL and highlight emerging biomarkers with the potential for clinical implementation.

Keywords: precision oncology; PD-L1; immunotherapy; prognosis; EBV



Citation: Chan, J.Y.; Lim, J.Q.; Ong, C.K. Towards Next Generation Biomarkers in Natural Killer/T-Cell Lymphoma. *Life* **2021**, *11*, 838. <https://doi.org/10.3390/life11080838>

Academic Editor: Andrew Edet Ekpenyong

Received: 25 July 2021

Accepted: 14 August 2021

Published: 16 August 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Natural killer/T-cell lymphoma (NKTCL) is an aggressive subtype of non-Hodgkin lymphoma with a propensity for extranodal involvement. Generally considered to be a rare cancer in the West, NKTCL demonstrates a geographic distribution that is predominant in East Asia and South America [1]. Clinically, NKTCL is most frequently characterized by involvement of the nasal cavity, and the neoplastic lymphoid cells are invariably infected with Epstein–Barr virus (EBV). In the past decade, the standard treatment for NKTCL has made steady progress with the adoption of L-asparaginase-based chemotherapy regimens, including SMILE (dexamethasone, methotrexate, ifosfamide, L-asparaginase, and etoposide) [2,3], P-GEMOX (pegaspargase, gemcitabine, and oxaliplatin) [4], DDGP (dexamethasone, cisplatin, gemcitabine, and pegaspargase) [5], and AspaMetDex (pegaspargase, methotrexate, and dexamethasone) [6]. More recently, newer therapies, such as immune checkpoint inhibitors and histone deacetylase (HDAC) inhibitors, have emerged in the armamentarium against NKTCL [7]. Despite improvements in survival outcomes with the incorporation of these agents in the management of NKTCL, a significant proportion of patients still relapse or remain refractory to treatment. Furthermore, these multi-agent/multimodality regimens often carry significant toxicities including severe myelosuppression and hypersensitivity reactions. An optimized approach to prognostication and risk stratification is thus warranted so as to provide the best outcomes while minimizing adverse effects. In this review,

we summarize the various model systems developed for this purpose and highlight novel biomarkers with the potential for clinical implementation (Table 1).

Table 1. Summary of recent studies on molecular biomarkers in NKTCL.

Reference	Biomarker Type	Study Design	Main Findings
[8]	Diagnostic	GWAS	Germline SNPs at the loci of the <i>HLA-DPB1</i> locus associate with increased risk of NKTCL
[9]	Diagnostic	GWAS	Germline SNPs at two novel loci of <i>HLA-DRB1</i> and <i>IL18RAP</i> identified to confer risk of NKTCL
[10]	Diagnostic	Pedigree analysis and WES	Recessive germline SNPs in <i>FAM160A1</i> reported in male siblings who were both diagnosed with NKTCL
[11]	Prognostic	WES and targeted sequencing	Identified recurrent mutations in <i>DDX3X</i> and <i>TP53</i> , both conferring worse OS and PFS.
[12]	Prognostic	Targeted sequencing	Mutations in <i>KMT2D</i> and <i>TP53</i> associated with worse survival outcomes, and may enhance the prognostic value of the IPI model
[13]	Prognostic	Multi-omic profiling	Identified three molecular subtypes—Tumor-suppressor/immune-modulator (TSIM), MYC-related (MB) and Histone epigenetic altered (HEA) groups. HEA and MB subtypes were associated with the best and worst OS and PFS, respectively.
[14]	Prognostic	SNP genotype microarray	A 7-SNP-based classifier predicts the survival of patients with NKTCL, and improves existing risk stratification systems based on clinicopathological variables
[15]	Prognostic	RNA expression (NanoString) and IHC	Identified four immune subgroups—immune-tolerant (high T-reg cells and FOXP3 expression, early stage, best prognosis), immune evasion-A and immune evasion-B (high cytotoxic T-cells, high PD-L1, low T-reg), and immune-silenced (depleted immune response). The immune-silenced group represented patients with advanced disease and poor prognosis.
[16]	Predictive	WGS	Identified cryptic rearrangements of the <i>PD-L1</i> gene disrupting the 3'-UTR to be a potential predictive biomarker of response to pembrolizumab in relapsed or refractory NKTCL

Abbreviations: GWAS, Genome-wide association study; WES, whole exome sequencing; SNP, single-nucleotide polymorphism; OS, overall survival; PFS, progression-free survival; IHC, immunohistochemistry; WGS, whole genome sequencing.

2. Clinical Risk Stratification and Prognostication Models for NK/T-Cell Lymphoma

Contemporary treatment of localized NKTCL with radiotherapy and chemotherapy results in a five-year overall survival rate of over 70% [17], while patients with advanced disease fare significantly worse with survival rates under 50% [7]. Several clinical features have been recognized to confer adverse prognosis, including extranasal compared to nasal subtypes [18], as for primary sites involving the gastrointestinal tract [19] and skin [20]. Recently, several prognostic scoring systems have been developed for NKTCL patients treated in the modern era, moving on from more traditional models including the International Prognostic Index (IPI) and Korean Prognostic Index (KPI) [21–23]. One of the first of such systems to be proposed was the Prognostic Index of Natural Killer lymphoma (PINK) model, which was derived from a retrospective study of patients treated with non-anthracycline-based regimens in the International NK/T-Cell Lymphoma Project. A total of four risk factors: age more than 60 years, stage III/IV disease, distant lymph node involvement and non-nasal subtype were strongly correlated with worse survival. The addition of detectable pre-treatment EBV DNA levels to the PINK model (PINK-E) further separated patients into distinctive prognostic groups [24]. More recently, a new central nervous system-prognostic index of natural killer (CNS-PINK) model was developed for the prediction of CNS relapse in extranodal NKTCL, by incorporating two or more extranodal sites involvement into the PINK model [25]. In a multicenter retrospective study, a high

peripheral blood neutrophil–lymphocyte ratio (NLR) was shown to confer poor survival, in addition to age more than 60 years, stage III/IV disease, as well as the presence of B symptoms (NABS model) [26]. Similarly, another prognostic model, the Chinese Prognostic Index for Natural Killer Cell Lymphoma (C-PINK), which consisted of three risk factors (disease stage, hemoglobin concentration below 100 g/L, and local invasiveness) was also described [27].

On top of these clinical factors, a recent meta-analysis demonstrated that several indices derived from fluorodeoxyglucose-positron emission tomography/computerized tomography (FDG-PET/CT) imaging could predict survival outcomes in patients with extranodal NKTCL, suggesting that FDG-PET/CT is not only useful for the assessment of treatment response but also potentially valuable for the prognostication in clinical practice [28]. The inclusion of imaging-based indicators such as SUVmax values and Deauville scores into clinical prognostic models have been suggested to complement clinical prognostic models [29]. Interestingly, PET/CT imaging patterns may also predict tumor immune microenvironment subtype in NKTCL, with the immune-silenced PET/CT pattern conferring worse overall survival compared to others [30].

While these prognostic models have improved over the years and may provide risk stratification for patients with NKTCL to a certain degree, there is probably still some way to go before they can achieve the level of consistency to enable risk-adapted approaches for treatment. Several other disease or host-related factors, including inflammatory responses, hematological indices [31–34], as well as nutritional indices [35–40], have been shown to affect outcomes in patients with NKTCL as well, and could possibly be integrated into the prognostic models. Nonetheless, these clinical prognostic systems, while inexpensive and easily accessible, ultimately suffer from modest discriminatory ability [23]. With advances in the understanding of the molecular pathobiology of NKTCL, incorporating novel molecular biomarkers may represent a potential strategy to further refine these prognostic models in order for their adoption in clinical decision making (Figure 1).

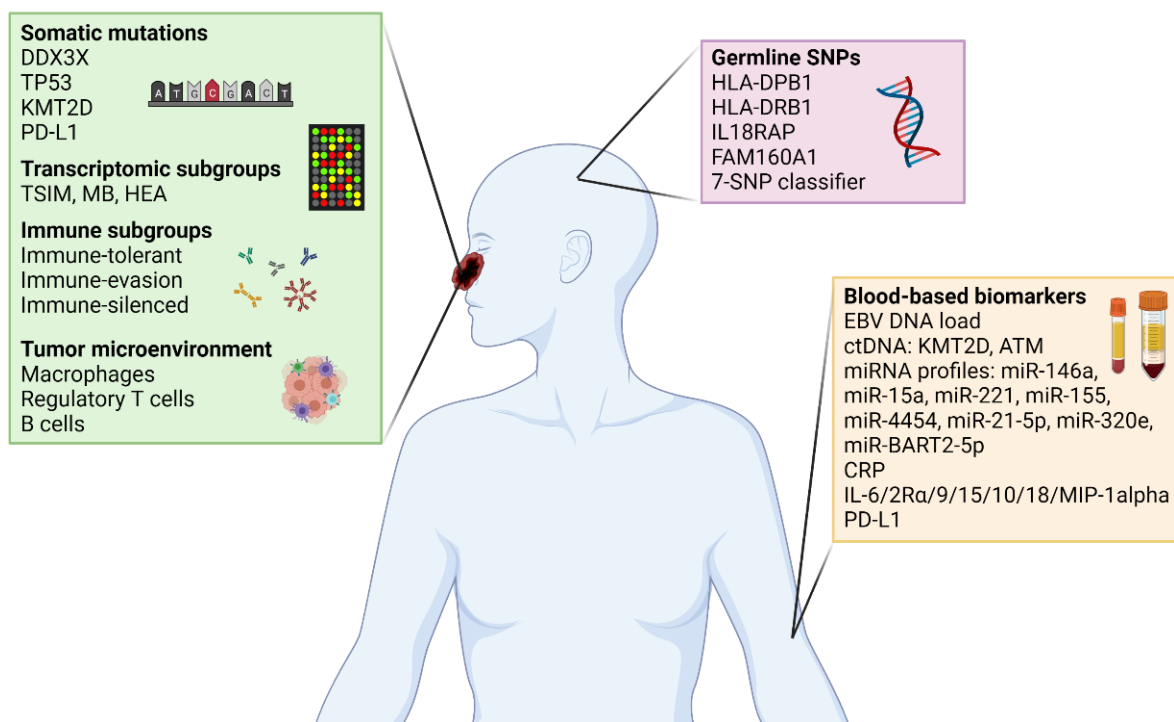


Figure 1. Emerging biomarker landscape for NKTCL.

3. Immunohistochemical Protein Expression as Potential Biomarkers

Apart from clinical features, several biomarkers have been proposed based on protein expression on tumor cells or features in the microenvironment using immunohistochemical assays. However, the search for reliable and consistent biomarkers has been challenging due to the limitations of study design, with most being retrospective studies consisting of small patient cohorts receiving heterogeneous treatments. Differences in assigning cut-off values defining positive expression, as well as inter-observer variation, further add to difficulties in interpreting the results. For example, tumor expression of CD30, a member of the tumor necrosis factor (TNF) receptor family, has been investigated in several studies. While some studies have correlated CD30 positivity with inferior survival outcomes [41,42], others have instead demonstrated favorable results [43–45] or no association [46,47]. CD38, a transmembrane glycoprotein strongly expressed in NKTCL, has been associated with poor prognosis in one study [48]. The prognostic utility of these biomarkers in NKTCL remains unclear and will need to be further validated.

Ki-67 is a nuclear and nucleolar protein expressed in proliferating cells but not resting cells, and high expression levels have been demonstrated to be associated with extra-aerodigestive tract primary site, B symptoms (any of the following: unexplained fever $>38^{\circ}\text{C}$, drenching night sweats, or loss of $>10\%$ body weight within 6 months), and tumor bulk [49], conferring worse survival outcomes [49,50]. In keeping with these findings, expression profiles of cell cycle related proteins in NKTCL, specifically p-ATM (phospho-ATM Serine/Threonine Kinase) and CHK2 (Checkpoint kinase 2), were recently shown to correlate with worse overall survival [51]. Chen et al. showed that human trophoblastic cell surface antigen 2 (Trop2), a tumor-related protein with oncogenic functions, was overexpressed in NKTCL and correlated with adverse survival outcomes [52]. More recently, overexpression of enhancer of zeste homolog 2 (EZH2), an H3K27-specific histone methyltransferase with an oncogenic role in NKTCL, was shown to be significantly associated with higher tumor cell proliferation, advanced stage, and higher risk of death [53]. In a study on NKTCL and PTCL, expression of phosphatidylinositol 3-kinase (PIK3) isoforms was evaluated and, in particular, high PIK3 α expression was significantly associated with poor survival [54].

The EBV-encoded proteins, latent membrane proteins (LMP), LMP1, and LMP2A have been examined as potential prognostic biomarkers in NKTCL. Results from several studies, however, have been conflicting. High expression of LMP1 has been associated with worse survival in two studies [55,56] but, conversely, correlated with better outcomes in two other studies [57,58]. LMP2A has been associated with poorer survival [56]. Activation of the NF- κ B pathway, which is downstream of LMP1 signaling, has been correlated with chemoresistance and worse survival in NKTCL [59]. High expression of Y box binding protein 1 (YB-1), regulated by LMP1 and NF- κ B, has similarly been shown to increase the potential for relapse, and conferred poor disease-free survival and reduced overall survival [60].

Taken together, results from studies evaluating histopathological protein expression in NKTCL and their prognostic significance, while promising, should be regarded as hypothesis generating and their clinical utility will require further validation.

4. Immune Microenvironment and Inflammatory Responses

The tumor immune microenvironment of NKTCL has been suggested to influence disease biology and clinical outcomes. A high number of tumor-associated macrophages have been shown to be associated with a higher Ki-67 proliferative index [61]. In keeping with this observation, a high degree of tumor-associated macrophages at diagnosis correlated with worse clinicopathological features, such as B symptoms and elevated serum LDH (lactate dehydrogenase) levels, as well as lower complete remission rates and dismal survival [62,63]. On the other hand, high levels of tumor-infiltrating FOXP3-positive regulatory T-cells showed prolonged overall and progression-free survival [64]. Patients with a high density of CD20+ B lymphocytes had early stage cancer and the tumors contained a

low Ki-67 index. High infiltration of B lymphocytes in the tumor tissues correlated with better overall survival [65].

Apart from tumor-infiltrating immune cells, various pro-inflammatory cytokines and interleukins (IL) expressed within the tumor microenvironment or circulation may also affect patient outcomes. C-reactive protein (CRP) is an acute-phase protein secreted by hepatocytes during the inflammatory response and is regulated by pro-inflammatory cytokines. NKTCL patients with elevated serum CRP levels have been associated with adverse clinical characteristics and worse survival outcomes [18,66]. Likewise, other inflammatory factors in the serum such as ferritin [67], ORM1 (Orosomucoid-1), S100A9 [68], 14-3-3 epsilon [69] have been shown to be associated with poor survival. In recent studies, high IL-6 expression levels, both in the tumor microenvironment [70] and serum [71] were correlated with poor prognosis. Other cytokines associated with worse prognosis include high serum IL-2R α [72], IL-9 [73], IL-15 [74], IL-10 [75], macrophage inflammatory protein 1 alpha (MIP-1alpha) [76], and IL-18 [77]. The mechanisms underlying the relationship between these pro-inflammatory cytokines and prognosis in NKTCL remain unclear. These inflammatory cytokines may potentially activate pro-oncogenic pathways, enhance tumor cell proliferation, and/or confer resistance to drug-evoked apoptosis [18,70]. IL-6 for example, has been suggested to activate the JAK/STAT pathway and also upregulate the expression of programmed cell death ligand-1 (PD-L1), thereby conferring immunosuppression in the microenvironment of NKTCL [70]. Future studies are needed to confirm the exact pathobiological mechanisms.

Targeting the Programmed cell death 1 (PD-1)/PD-1 ligand 1 (PD-L1) immune checkpoint pathway has emerged as a promising strategy for NKTCL treatment, and several studies have been conducted to evaluate its prognostic significance as well. In one study, PD-L1 expression on NKTCL tumor cells was shown to be an independent, favorable prognostic marker for overall survival in advanced NKTCL [78]. However, several others have shown either the opposite [79–82] or no effect [83]. These varying results may be due to a lack of standard guidelines for the interpretation of PD-L1 expression in NKTCL, as well as the use of different anti-PD-L1 antibodies and variable threshold cut-offs for positive staining [84]. Most importantly, the interpretation of its prognostic significance has to be considered in the context of the treatment type received by the patient cohort. In parallel, circulating PD-L1 analytes have been investigated in several studies [79,80,85]. In early stage NKTCL patients treated with asparaginase, patients with high pre-treatment serum soluble PD-L1 (sPD-L1) levels were associated with poor treatment response and high post-treatment levels and had shorter progression-free survival and overall survival [85]. In another study on early stage NKTCL patients treated with induction chemotherapy followed by consolidative radiotherapy, patients who had a high concentration of serum sPD-L1 showed lower complete response rates to treatment and poorer survival than those with a low sPD-L1 concentration [79]. Levels of PD-L1 mRNA in peripheral blood mononuclear cells and sPD-L1 were significantly associated with several adverse clinicopathological factors [86]. High pre-treatment plasma exosomal PD-L1 concentration was associated with higher SUVmax level and recurrence rate. Similarly, high sPD-L1 group was also associated with some adverse clinical parameters, including advanced stage, elevated LDH levels, B symptoms, high IPI score, and PINK score [87]. It has been suggested that soluble forms of ligands are often generated by the proteolytic cleavage of membrane-bound proteins, including PD-L1. sPD-L1 levels have been positively correlated with the total number of PD-L1-expressing tumor cells and may thus be reflective of tumor load. In addition, sPD-L1 may also be produced by immune cells, potentially signifying a suppressed T-cell mediated immune response in the tumor microenvironment [85–87].

5. Molecular Biomarkers for Diagnosis and Prognostication

Our understanding of the molecular landscape of NKTCL has greatly improved over the past years through advances in multi-omic profiling technologies as well as multinational collaboration, and efforts continue to be placed on their potential translational utility as biomarkers to facilitate the precise diagnosis, prognosis, and treatment of patients with NKTCL. Many of the early molecular studies on NKTCL were focused on the role of EBV in its pathogenesis and clinical behavior [88–91]. Presently, NKTCL tumor cells are thought to be invariably infected by EBV and contemporary diagnosis has to be supported by the presence of EBV, typically by in-situ hybridization [92]. As such, circulating EBV DNA in the blood or plasma from apoptotic tumor cells is an intuitive consequence of tumor load [93] and several studies have established EBV DNA in the blood as a significant independent predictor of poor prognosis [94–99].

Early studies in the Japanese population showed that HLA-A*02:01 was associated with a reduced risk of NKTCL [100]. More recently, genome-wide association studies revealed that germline single-nucleotide polymorphisms (SNPs) at the loci of the *HLA-DPB1*, *HLA-DRB1*, and *IL18RAP* were associated with increased risk of NKTCL [8,9]. Li et al. provided the first evidence of a common SNP at *HLA-DPB1* contributing to NKTCL risk, with each copy of the risk allele raising the disease risk by 1.84 fold compared with the baseline wild-type genotype. This risk modulation may be mediated by alteration of antigen recognition and tumor clearance, due to variation at the binding pocket of the HLA-DPB1 protein (amino acid positions 84–87) [8]. In a larger follow-up study on 1417 NKTCL cases and 20,402 controls, Lin et al. subsequently identified two novel loci significantly associated with NKTCL development, independent of the risk conferred by *HLA-DPB1*, implicating inflammation and immune regulation through the IL18–IL18RAP axis and antigen presentation involving HLA-DRB1 [9]. The relevance of the immune microenvironment in NKTCL pathogenesis is further supported by the identification of recessive germline SNPs in *FAM160A1* reported in a pair of male siblings from a non-consanguineous Chinese family who were both diagnosed with NKTCL. *FAM160A1* was expressed in CD68-positive histiocytes rather than the tumor cells, suggesting that variant *FAM160A1* contributes to NKTCL pathogenesis by immune dysregulation [10].

Next generation sequencing has led to a greater clarity of the genomic landscape of NKTCL, revealing recurrent somatic mutations such as *TP53*, *JAK3*, *STAT3*, and *DDX3X* in NKTCL [11,101–103]. Additionally, frequent somatic *PD-L1* structural rearrangements have also been recently reported [104]. However, only mutational status of the *DDX3X*, *TP53*, and *KMT2D* genes were found to be prognostic in patients treated with CHOP-based regimens [11,12]. Molecular subtyping based on transcriptomics was able to group NKTCLs into three subtypes with association to survival: Tumor-suppressor/immune-modulator (TSIM), MYC-related (MB), and Histone epigenetic altered (HEA) groups. The NKTCL tumors in the HEA and MB subgroups were associated with the best and worst overall survival and progression-free survival, respectively. The associations were also made more pronounced when compared within the advanced Ann Arbor stage patients with NKTCL [13]. Aside from somatic mutations, a composite 7-single nucleotide polymorphism (7-SNP) germline signature was recently developed for predicting the survival of patients with NKTCL. The composite 7-SNP signature was suggested to be an additional risk indicator in helping clinical decision making [14].

6. Current Treatment Strategies for NKTCL and Potential Predictive Biomarkers

The treatment of disseminated NKTCL has entered a paradigm shift from anthracycline/CHOP-based regimens, to asparaginase-based regimens and to the most recent immunotherapy with PD-L1/PD1 immune checkpoint blockade [105]. Contemporary first-line therapeutic options for advanced NKTCL utilizes L-asparaginase as the backbone, including regimens such as SMILE [2,3], P-GEMOX [4], DDGP [5], and AspaMetDex [6]. However, patient outcomes remain poor with relapse rates approaching 70%. Upon relapse, salvage treatment using platinum-based or gemcitabine-based regimens suffer from limited effi-

cacy [1]. As such, novel strategies incorporating immune checkpoint inhibitors and HDAC inhibitors have been explored in NKTCL. Blockade of the PD1/PDL1 axis in NKTCL has recently emerged as a promising treatment strategy. The combined objective response rate of three case series of patients with NKTCL being treated with pembrolizumab (PD-1 antibody) and avelumab (PD-L1 antibody) was as high as 54.3% (19/35) [106–108]. In an interim report, a phase II prospective study using sintilimab (PD-1 antibody) achieved an overall response rate of 68% in 28 relapsed or refractory NKTCL patients [109]. While these results are encouraging, predictive biomarkers are urgently required to help with patient stratification and treatment guidance.

In a recent study on the tumor immune microenvironment of NKTCL, Cho et al. analyzed the expression of 579 immune-related genes and characterized the immune cells using immunohistochemistry for FoxP3, PD-L1, and CD68. Four subgroups were identified, namely immune-tolerant (high T-reg cells and FOXP3 expression, early stage, best prognosis), immune evasion-A and immune evasion-B (high cytotoxic T-cells, high PD-L1, low T-reg), and immune-silenced (depleted immune response). The immune-silenced group represented patients with advanced disease and poor prognosis. Responses to pembrolizumab were observed for patients in the immune-tolerant group (1 out of 1) and immune-evasion group (3 out of 5) but none in the immune-silenced group (n = 5) [15]. Subsequently, Lee et al. identified *EGR1* upregulation in localized stage, low risk NKTCL. *EGR1* expression was highest in the immune-tolerant group and tended to decrease in the order of immune evasion-A, immune evasion-B, and immune-silenced subgroups, suggesting that *EGR1* contributes to early tumorigenesis by suppressing the immune response [110]. A retrospective multicenter clinicopathologic and genetic analysis of patients with relapsed or refractory NKTCL treated with pembrolizumab identified cryptic rearrangements of the PD-L1 gene disrupting the 3'-UTR to be a more reliable biomarker of the response to pembrolizumab than the immunohistochemical staining of membranous PD-L1 on NKTCL tumoral cells [16].

7. Developing Next Generation Biomarkers for NKTCL

Advances in next generation sequencing have led to the development of several novel molecular prognostic models and identification of potential predictive biomarkers of checkpoint immunotherapy in NKTCL. Liquid biopsy-based approaches may represent a non-invasive means of obtaining this information, bypassing the need for an invasive tissue biopsy [111]. In particular, plasma circulating tumor DNA (ctDNA) has recently come to the forefront as a dynamic marker reflecting disease burden, and represents a non-invasive means for disease assessment. In a recent study, Li et al. showed that the mutation spectrum of NKTCL could be adequately captured, and that serial monitoring could be useful for tracking residual disease. Furthermore, mutations in *KMT2D* and *ATM* were able to predict adverse outcomes [112].

MicroRNAs (miRNA) are small, noncoding RNAs that inhibit the translation of target genes and have been shown to play clinically important roles in NKTCL. For example, early studies showed that miR-21 and miR-155 were aberrantly overexpressed in NKTCL and activated oncogenic AKT signaling [113,114]. Low miR-146a expression was demonstrated to be an independent poor prognostic factor in NKTCL, while in NKTCL cell-lines, miR-146a overexpression inhibited NF κ B activity, suppressed cell proliferation, induced apoptosis, and enhanced chemosensitivity, suggesting it functions as a tumor suppressor [115]. Reduced miR-15a expression, possibly mediated by EBV LMP1, was associated with poor prognosis [116]. In line with these findings, circulating microRNA profiles have also been investigated for their potential clinical utility. Circulating plasma miR-221 was associated with poor overall survival outcomes [117]. Plasma miR-155 was higher in non-responders to treatment compared with responders, and was directly correlated with serum expression of IL-6, IL-13, and TNF- α [118]. Increased serum levels of exosomal miR-4454, miR-21-5p, and miR-320e were also associated with poor overall survival [119]. Circulating EBV-encoded miRNAs miR-BART2-5p, miR-BART7-3p, miR-BART13-3p, and

miR-BART1-5p discriminated NKTCL patients from healthy controls and significantly decreased after treatment. In addition, a high miR-BART2-5p level was associated with disease progression and poor prognosis [120].

8. Conclusions and Prospects

Moving forwards, high definition profiling technologies, including single cell sequencing and spatial transcriptomic assays, are expected to further unravel the complexities between tumor cells and the microenvironment of NKTCL, which may be especially relevant as immunotherapy evolves to the forefront of NKTCL treatment. The prospect of being able to dissect the molecular landscape of NKTCL at high resolution and the discovery of novel biomarkers are truly exciting as we enter a new era of precision medicine and immune oncology.

Author Contributions: Conceptualization J.Y.C., J.Q.L. and C.K.O.; original draft preparation, J.Y.C. and J.Q.L.; writing, review and editing, J.Y.C., J.Q.L. and C.K.O. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by the Tanoto Foundation (NRDUKST18101), New Century Foundation (NCCRF-YR2014-SEP-SD2), Ling Foundation (NRDUKSN18101), Singapore Ministry of Health's National Medical Research Council Research (NMRC/Fellowship/0054/2017, NMRC-OFIRG16NOV090, TCR/010-NCC/2012, NMRC-OFLCG-18May0028), SHF-Foundation Research Grant (SHF/FG653P/2017), as well as the SingHealth Duke-NUS Academic Medical Centre and Oncology ACP Nurturing Clinician Scientist Scheme (08-FY2017/P1/14-A28).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Chan, J.Y.; Lim, S.T. Novel findings from the Asian Lymphoma Study Group: Focus on T and NK-cell lymphomas. *Int. J. Hematol.* **2018**, *107*, 413–419. [\[CrossRef\]](#)
- Yamaguchi, M.; Kwong, Y.-L.; Kim, W.S.; Maeda, Y.; Hashimoto, C.; Suh, C.; Izutsu, K.; Ishida, F.; Isobe, Y.; Sueoka, E.; et al. Phase II Study of SMILE Chemotherapy for Newly Diagnosed Stage IV, Relapsed, or Refractory Extranodal Natural Killer (NK)/T-Cell Lymphoma, Nasal Type: The NK-Cell Tumor Study Group Study. *J. Clin. Oncol.* **2011**, *29*, 4410–4416. [\[CrossRef\]](#)
- Kwong, Y.-L.; Kim, W.S.; Lim, S.T.; Kim, S.J.; Tang, T.; Tse, E.; Leung, A.Y.H.; Chim, C. SMILE for natural killer/T-cell lymphoma: Analysis of safety and efficacy from the Asia Lymphoma Study Group. *Blood* **2012**, *120*, 2973–2980. [\[CrossRef\]](#)
- Wang, J.-H.; Wang, L.; Liu, C.-C.; Xia, Z.-J.; Huang, H.-Q.; Lin, T.-Y.; Jiang, W.-Q.; Lu, Y. Efficacy of combined gemcitabine, oxaliplatin and pegaspargase (P-gemox regimen) in patients with newly diagnosed advanced-stage or relapsed/refractory extranodal NK/T-cell lymphoma. *Oncotarget* **2016**, *7*, 29092–29101. [\[CrossRef\]](#) [\[PubMed\]](#)
- Li, X.; Cui, Y.; Sun, Z.; Zhang, L.; Li, L.; Wang, X.; Wu, J.; Fu, X.; Ma, W.; Zhang, X.; et al. DDGP versus SMILE in Newly Diagnosed Advanced Natural Killer/T-Cell Lymphoma: A Randomized Controlled, Multicenter, Open-label Study in China. *Clin. Cancer Res.* **2016**, *22*, 5223–5228. [\[CrossRef\]](#) [\[PubMed\]](#)
- Jaccard, A.; Gachard, N.; Marin, B.; Rogez, S.; Audrain, M.; Suarez, F.; Tilly, H.; Morschhauser, F.; Thieblemont, C.; Ysebaert, L.; et al. Efficacy of L-asparaginase with methotrexate and dexamethasone (AspaMetDex regimen) in patients with refractory or relapsing extranodal NK/T-cell lymphoma, a phase 2 study. *Blood* **2011**, *117*, 1834–1839. [\[CrossRef\]](#) [\[PubMed\]](#)
- Wang, H.; Fu, B.B.; Gale, R.P.; Liang, Y. NK-/T-cell lymphomas. *Leukemia* **2021**. [\[CrossRef\]](#) [\[PubMed\]](#)
- Li, Z.; Xia, Y.; Feng, L.N.; Chen, J.R.; Li, H.M.; Cui, J.; Cai, Q.Q.; Sim, K.S.; Nairismägi, M.L.; Laurensia, Y.; et al. Genetic risk of extranodal natural killer T-cell lymphoma: A genome-wide association study. *Lancet Oncol.* **2016**, *17*, 1240–1247. [\[CrossRef\]](#)
- Lin, G.W.; Xu, C.; Chen, K.; Huang, H.Q.; Chen, J.; Song, B.; Chan, J.K.C.; Li, W.; Liu, W.; Shih, L.Y.; et al. Genetic risk of extranodal natural killer T-cell lymphoma: A genome-wide association study in multiple populations. *Lancet Oncol.* **2019**, *21*, 306–316. [\[CrossRef\]](#)
- Chan, J.Y.; Ng, A.Y.J.; Cheng, C.L.; Nairismägi, M.L.; Venkatesh, B.; Cheah, D.M.Z.; Li, S.T.; Chan, S.H.; Ngeow, J.; Laurensia, Y.; et al. Whole exome sequencing identifies recessive germline mutations in FAM160A1 in familial NK/T cell lymphoma. *Blood Cancer J.* **2018**, *8*, 111. [\[CrossRef\]](#)
- Jiang, L.; Gu, Z.H.; Yan, Z.X.; Zhao, X.; Xie, Y.Y.; Zhang, Z.G.; Pan, C.M.; Hu, Y.; Cai, C.P.; Dong, Y.; et al. Exome sequencing identifies somatic mutations of DDX3X in natural killer/T-cell lymphoma. *Nat. Genet.* **2015**, *47*, 1061–1066. [\[CrossRef\]](#)
- Gao, Y.; Li, Y.; Ma, G.; Zhao, G.; Liu, H. KMT2D and TP53 mutation status improve the prognostic value of the International Prognostic Index (IPI) stratification in ENKTL patients. *Neoplasma* **2020**, *67*, 636–643. [\[CrossRef\]](#)

13. Xiong, J.; Cui, B.W.; Wang, N.; Dai, Y.T.; Zhang, H.; Wang, C.F.; Zhong, H.J.; Cheng, S.; Ou-Yang, B.S.; Hu, Y.; et al. Genomic and Transcriptomic Characterization of Natural Killer T Cell Lymphoma. *Cancer Cell* **2020**, *37*, 403–419.e6. [[CrossRef](#)] [[PubMed](#)]
14. Tian, X.P.; Ma, S.Y.; Young, K.H.; Ong, C.K.; Liu, Y.; Li, Z.; Zhai, Q.; Huang, H.Q.; Lin, T.; Li, Z.; et al. A composite single-nucleotide polymorphism prediction signature for extranodal natural killer/T-cell lymphoma. *Blood* **2021**, *138*, 452–463. [[CrossRef](#)]
15. Cho, J.; Kim, S.J.; Park, W.-Y.; Kim, J.; Woo, J.; Kim, G.; Yoon, S.E.; Ko, Y.H.; Kim, W.S. Immune subtyping of extranodal NK/T-cell lymphoma: A new biomarker and an immune shift during disease progression. *Mod. Pathol.* **2019**, *33*, 603–615. [[CrossRef](#)] [[PubMed](#)]
16. Lim, J.Q.; Huang, D.; Tang, T.; Tan, D.; Laurensia, Y.; Peng, R.J.; Wong, E.K.Y.; Cheah, D.M.Z.; Chia, B.K.H.; Iqbal, J.; et al. Whole-genome sequencing identifies responders to Pembrolizumab in relapse/refractory natural-killer/T cell lymphoma. *Leukemia* **2020**, *34*, 3413–3419. [[CrossRef](#)] [[PubMed](#)]
17. Kwong, Y.; Kim, S.; Tse, E.; Oh, S.; Kwak, J.; Eom, H.; Do, Y.; Mun, Y.; Lee, S.; Shin, H.; et al. Sequential chemotherapy/radiotherapy was comparable with concurrent chemoradiotherapy for stage I/II NK/T-cell lymphoma. *Ann. Oncol.* **2017**, *29*, 256–263. [[CrossRef](#)]
18. Au, W.-Y.; Weisenburger, D.D.; Intragumtornchai, T.; Nakamura, S.; Kim, W.-S.; Sng, I.; Vose, J.; Armitage, J.O.; Liang, R. International Peripheral T-Cell Lymphoma Project Clinical differences between nasal and extranasal natural killer/T-cell lymphoma: A study of 136 cases from the International Peripheral T-Cell Lymphoma Project. *Blood* **2009**, *113*, 3931–3937. [[CrossRef](#)]
19. Kim, S.J.; Jung, H.A.; Chuang, S.-S.; Hong, H.; Guo, C.-C.; Cao, J.; Hong, X.-N.; Suzuki, R.; Kang, H.J.; Won, J.H.; et al. Extranodal natural killer/T-cell lymphoma involving the gastrointestinal tract: Analysis of clinical features and outcomes from the Asia Lymphoma study group. *J. Hematol. Oncol.* **2013**, *6*, 86. [[CrossRef](#)]
20. Takata, K.; Hong, M.-E.; Sitthinamsuwan, P.; Loong, F.; Tan, S.-Y.; Liao, J.-Y.; Hsieh, P.-P.; Ng, S.-B.; Yang, S.-F.; Pongpruttipan, T.; et al. Primary Cutaneous NK/T-cell Lymphoma, Nasal Type and CD56-positive Peripheral T-cell Lymphoma. *Am. J. Surg. Pathol.* **2015**, *39*, 1–12. [[CrossRef](#)]
21. Lee, J.; Suh, C.; Park, Y.H.; Ko, Y.H.; Bang, S.M.; Lee, J.H.; Lee, D.H.; Huh, J.; Oh, S.Y.; Kwon, H.-C.; et al. Extranodal Natural Killer T-Cell Lymphoma, Nasal-Type: A Prognostic Model From a Retrospective Multicenter Study. *J. Clin. Oncol.* **2006**, *24*, 612–618. [[CrossRef](#)]
22. Yang, Y.; Zhang, Y.-J.; Zhu, Y.; Cao, J.-Z.; Yuan, Z.-Y.; Xu, L.-M.; Wu, J.-X.; Wang, W.; Wu, T.; Lu, B.; et al. Prognostic nomogram for overall survival in previously untreated patients with extranodal NK/T-cell lymphoma, nasal-type: A multicenter study. *Leukemia* **2015**, *29*, 1571–1577. [[CrossRef](#)]
23. Chen, S.-Y.; Yang, Y.; Qi, S.-N.; Wang, Y.; Hu, C.; He, X.; Zhang, L.-L.; Wu, G.; Qu, B.-L.; Qian, L.-T.; et al. Validation of nomogram-revised risk index and comparison with other models for extranodal nasal-type NK/T-cell lymphoma in the modern chemotherapy era: Indication for prognostication and clinical decision-making. *Leukemia* **2021**, *35*, 130–142. [[CrossRef](#)]
24. Kim, S.J.; Yoon, D.H.; Jaccard, A.; Chng, W.J.; Lim, S.T.; Hong, H.; Park, Y.; Chang, K.M.; Maeda, Y.; Ishida, F.; et al. A prognostic index for natural killer cell lymphoma after non-anthracycline-based treatment: A multicentre, retrospective analysis. *Lancet Oncol.* **2016**, *17*, 389–400. [[CrossRef](#)]
25. Kim, H.; Jeong, H.; Yamaguchi, M.; Sohn, I.; Yoon, S.E.; Byeon, S.; Hur, J.Y.; Koh, Y.; Yoon, S.-S.; Kim, S.J.; et al. Prediction and prevention of central nervous system relapse in patients with extranodal natural killer/T-cell lymphoma. *Blood* **2020**, *136*, 2548–2556. [[CrossRef](#)]
26. Tan, K.M.; Chia, B.; Lim, J.Q.; Khoo, L.P.; Cheng, C.L.; Tan, L.; Poon, E.; Somasundaram, N.; Farid, M.; Tang, T.P.L.; et al. A clinicohaematological prognostic model for nasal-type natural killer/T-cell lymphoma: A multicenter study. *Sci. Rep.* **2019**, *9*, 14961. [[CrossRef](#)]
27. Hong, H.; Huang, H.; Fang, X.; Wang, Z.; Ye, S.; Zhang, H.; Huang, Y.; Guo, H.; Chen, X.; Liang, C.; et al. A prognostic index for nasal-type early-stage extranodal natural killer/T-cell lymphoma: A multicenter study. *Am. J. Hematol.* **2019**, *94*, E122–E124. [[CrossRef](#)] [[PubMed](#)]
28. Wang, H.; Shen, G.; Jiang, C.; Li, L.; Cui, F.; Tian, R. Prognostic value of baseline, interim and end-of-treatment 18F-FDG PET/CT parameters in extranodal natural killer/T-cell lymphoma: A meta-analysis. *PLoS ONE* **2018**, *13*, e0194435. [[CrossRef](#)] [[PubMed](#)]
29. Huang, H.L.; Ngam, P.I.; Tan, K.M.; Ng, D.C.E.; Lim, S.T.; Chan, J.Y. The exact Deauville score, NABS score and high SUVmax predicts outcome in extranodal natural killer/T-cell lymphoma. *Ann. Nucl. Med.* **2021**, *35*, 557–568. [[CrossRef](#)] [[PubMed](#)]
30. Lim, C.H.; Yoon, S.E.; Kim, S.J.; Cho, J.; Ko, Y.H.; Lee, K.-H.; Kim, W.S. Metabolic activity of extranodal NK/T cell lymphoma on 18F-FDG PET/CT according to immune subtyping. *Sci. Rep.* **2021**, *11*, 5879. [[CrossRef](#)] [[PubMed](#)]
31. Wang, L.; Xia, Z.-J.; Lu, Y.; Wang, Q.-X.; Niu, S.-Q.; Huang, H.-Q.; Zhang, Y.-J. A modified international prognostic index including pretreatment hemoglobin level for early stage extranodal natural killer/T cell lymphoma. *Leuk. Lymphoma* **2015**, *56*, 3038–3044. [[CrossRef](#)]
32. Cao, J.; Lan, S.; Shen, L.; Si, H.; Xiao, H.; Yuan, Q.; Li, X.; Li, H.; Guo, R. Hemoglobin level, a prognostic factor for nasal extranodal natural killer/T-cell lymphoma patients from stage I to IV: A validated prognostic nomogram. *Sci. Rep.* **2017**, *7*, 10982. [[CrossRef](#)]
33. Luo, H.; Quan, X.; Song, X.-Y.; Zhang, L.; Yin, Y.; He, Q.; Cai, S.; Li, S.; Zeng, J.; Zhang, Q.; et al. Red blood cell distribution width as a predictor of survival in nasal-type, extranodal natural killer/T-cell lymphoma. *Oncotarget* **2017**, *8*, 92522–92535. [[CrossRef](#)]
34. Zhang, Y.P.; Zhang, R.; Zhu, H.Y.; Wang, L.; Wu, Y.J.; Liang, J.H.; Shi, W.Y.; Liu, H.; Xu, W.; Li, J.Y. Circulating Low Absolute CD4+ T Cell Counts May Predict Poor Prognosis in Extranodal NK/T-Cell Lymphoma Patients Treating with Pegaspargase-Based Chemotherapy. *Cancer Res. Treat.* **2019**, *51*, 368–377. [[CrossRef](#)]

35. Watanabe, T.; Kinoshita, T.; Itoh, K.; Yoshimura, K.; Ogura, M.; Kagami, Y.; Yamaguchi, M.; Kurosawa, M.; Tsukasaki, K.; Kasai, M.; et al. Pretreatment total serum protein is a significant prognostic factor for the outcome of patients with peripheral T/natural killer-cell lymphomas. *Leuk. Lymphoma* **2010**, *51*, 813–821. [[CrossRef](#)] [[PubMed](#)]
36. Li, Y.J.; Jiang, W.Q.; Huang, J.J.; Xia, Z.J.; Huang, H.Q.; Li, Z.M. The Glasgow Prognostic Score (GPS) as a novel and significant predictor of extranodal natural killer/T-cell lymphoma, nasal type. *Am. J. Hematol.* **2013**, *88*, 394–399. [[CrossRef](#)] [[PubMed](#)]
37. Cai, Q.; Luo, X.; Liang, Y.; Rao, H.; Fang, X.; Jiang, W.; Lin, T.; Huang, H. Fasting blood glucose is a novel prognostic indicator for extranodal natural killer/T-cell lymphoma, nasal type. *Br. J. Cancer* **2013**, *108*, 380–386. [[CrossRef](#)] [[PubMed](#)]
38. Cai, Q.; Luo, X.; Zhang, G.; Huang, H.; Huang, H.; Lin, T.; Jiang, W.; Xia, Z.; Young, K.H. New prognostic model for extranodal natural killer/T cell lymphoma, nasal type. *Ann. Hematol.* **2014**, *93*, 1541–1549. [[CrossRef](#)]
39. Chen, K.L.; Liu, Y.H.; Li, W.Y.; Chen, J.; Gu, Y.K.; Geng, Q.R.; Jiang, W.Q.; Huang, H.Q.; Lin, T.Y.; Xia, Z.J.; et al. The prognostic nutritional index predicts survival for patients with extranodal natural killer/T cell lymphoma, nasal type. *Ann. Hematol.* **2015**, *94*, 1389–1400. [[CrossRef](#)]
40. Mao, J.; Yin, H.; Wang, L.; Wu, J.Z.; Xia, Y.; Zhu, H.Y.; Fan, L.; Li, J.Y.; Liang, J.H.; Xu, W. Prognostic value of 25-hydroxy vitamin D in extranodal NK/T cell lymphoma. *Ann. Hematol.* **2020**, *100*, 445–453. [[CrossRef](#)]
41. Hong, J.; Park, S.; Baek, H.L.; Jung, J.H.; Kang, I.G.; Sym, S.J.; Park, J.; Ahn, J.Y.; Cho, E.K.; Kim, S.T.; et al. Tumor cell nuclear diameter and CD30 expression as potential prognostic parameter in patients with extranodal NK/T-cell lymphoma, nasal type. *Int. J. Clin. Exp. Pathol.* **2012**, *5*, 939–947.
42. Li, P.; Jiang, L.; Zhang, X.; Liu, J.; Wang, H. CD30 expression is a novel prognostic indicator in extranodal natural killer/T-cell lymphoma, nasal type. *BMC Cancer* **2014**, *14*, 890. [[CrossRef](#)]
43. Mraz-Gernhard, S.; Natkunam, Y.; Hoppe, R.T.; LeBoit, P.; Kohler, S.; Kim, Y.H. Natural Killer/Natural Killer-Like T-Cell Lymphoma, CD56+, Presenting in the Skin: An Increasingly Recognized Entity with an Aggressive Course. *J. Clin. Oncol.* **2001**, *19*, 2179–2188. [[CrossRef](#)]
44. Kim, W.Y.; Nam, S.J.; Kim, S.; Kim, T.M.; Heo, D.S.; Kim, C.-W.; Jeon, Y.K. Prognostic implications of CD30 expression in extranodal natural killer/T-cell lymphoma according to treatment modalities. *Leuk. Lymphoma* **2014**, *56*, 1778–1786. [[CrossRef](#)]
45. Lee, W.J.; Moon, I.J.; Shin, H.J.; Won, C.H.; Chang, S.E.; Choi, J.H.; Lee, M.W. CD30-positive cutaneous extranodal natural killer/T-cell lymphoma: Clinicopathological features and survival outcomes. *Int. J. Dermatol.* **2018**, *58*, 688–696. [[CrossRef](#)]
46. Gaal, K.; Sun, N.C.J.; Hernandez, A.M.; Arber, D.A. Sinonasal NK/T-cell Lymphomas in the United States. *Am. J. Surg. Pathol.* **2000**, *24*, 1511–1517. [[CrossRef](#)] [[PubMed](#)]
47. Kuo, T.T.; Shih, L.Y.; Tsang, N.M. Nasal NK/T cell lymphoma in Taiwan: A clinicopathologic study of 22 cases, with analysis of histologic subtypes, Epstein-Barr virus LMP-1 gene association, and treatment modalities. *Int. J. Surg. Pathol.* **2004**, *12*, 375–387. [[CrossRef](#)] [[PubMed](#)]
48. Wang, L.; Wang, H.; Li, P.-F.; Lu, Y.; Xia, Z.-J.; Huang, H.-Q.; Zhang, Y.-J. CD38 expression predicts poor prognosis and might be a potential therapy target in extranodal NK/T cell lymphoma, nasal type. *Ann. Hematol.* **2015**, *94*, 1381–1388. [[CrossRef](#)] [[PubMed](#)]
49. Jiang, L.; Li, P.; Wang, H.; Liu, J.; Zhang, X.; Qiu, H.; Zhang, B. Prognostic significance of Ki-67 antigen expression in extranodal natural killer/T-cell lymphoma, nasal type. *Med. Oncol.* **2014**, *31*, 218. [[CrossRef](#)]
50. Kim, S.J.; Kim, B.S.; Choi, C.W.; Choi, J.; Kim, I.; Lee, Y.H.; Kim, J.S. Ki-67 expression is predictive of prognosis in patients with stage I/II extranodal NK/T-cell lymphoma, nasal type. *Ann. Oncol.* **2007**, *18*, 1382–1387. [[CrossRef](#)]
51. Ye, Q.; Chen, H.; Wen, Z.; Guo, W.; Huang, Y.; Mo, X. Abnormal expression of p-ATM/CHK2 in nasal extranodal NK/T cell lymphoma, nasal type, is correlated with poor prognosis. *J. Clin. Pathol.* **2020**, *74*, 223–227. [[CrossRef](#)]
52. Chen, R.; Lu, M.; Wang, J.; Zhang, D.; Lin, H.; Zhu, H.; Zhang, W.; Xiong, L.; Ma, J.; Mao, Y.; et al. Increased expression of Trop2 correlates with poor survival in extranodal NK/T cell lymphoma, nasal type. *Virchows Arch.* **2013**, *463*, 713–719. [[CrossRef](#)]
53. Liu, J.; Liang, L.; Huang, S.; Nong, L.; Li, D.; Zhang, B.; Li, T. Aberrant differential expression of EZH2 and H3K27me3 in extranodal NK/T-cell lymphoma, nasal type, is associated with disease progression and prognosis. *Hum. Pathol.* **2018**, *83*, 166–176. [[CrossRef](#)]
54. Huang, D.; Song, T.L.; Nairismägi, M.L.; Laurensia, Y.; Pang, W.L.; Zhe, D.C.M.; Wong, E.K.Y.; Wijaya, G.G.; Tan, J.; Tan, S.H.; et al. Evaluation of the PIK3 pathway in peripheral T-cell lymphoma and NK/T-cell lymphoma. *Br. J. Haematol.* **2020**, *189*, 731–744. [[CrossRef](#)]
55. Cao, W.; Liu, Y.; Zhang, H.; Wang, S.; Zhang, L.; Zhang, L.; Sun, B. Expression of LMP-1 and Cyclin D1 protein is correlated with an unfavorable prognosis in nasal type NK/T cell lymphoma. *Mol. Med. Rep.* **2008**, *1*, 363–368. [[CrossRef](#)] [[PubMed](#)]
56. Mao, Y.; Zhang, D.W.; Zhu, H.; Lin, H.; Xiong, L.; Cao, Q.; Liu, Y.; Li, Q.D.; Xu, J.R.; Xu, L.F.; et al. LMP1 and LMP2A are potential prognostic markers of extranodal NK/T-cell lymphoma, nasal type (ENKTL). *Diagn. Pathol.* **2012**, *7*, 178. [[CrossRef](#)] [[PubMed](#)]
57. Kanemitsu, N.; Isobe, Y.; Masuda, A.; Momose, S.; Higashi, M.; Tamaru, J.; Sugimoto, K.; Komatsu, N. Expression of Epstein-Barr virus-encoded proteins in extranodal NK/T-cell Lymphoma, nasal type (ENKL): Differences in biologic and clinical behaviors of LMP1-positive and -negative ENKL. *Clin. Cancer Res.* **2012**, *18*, 2164–2172. [[CrossRef](#)]
58. Yamaguchi, M.; Takata, K.; Yoshino, T.; Ishizuka, N.; Oguchi, M.; Kobayashi, Y.; Isobe, Y.; Ishizawa, K.; Kubota, N.; Itoh, K.; et al. Prognostic biomarkers in patients with localized natural killer/T-cell lymphoma treated with concurrent chemoradiotherapy. *Cancer Sci.* **2014**, *105*, 1435–1441. [[CrossRef](#)] [[PubMed](#)]
59. Liu, X.; Wang, B.; Ma, X.; Guo, Y. NF-kappaB activation through the alternative pathway correlates with chemoresistance and poor survival in extranodal NK/T-cell lymphoma, nasal type. *Jpn. J. Clin. Oncol.* **2009**, *39*, 418–424. [[CrossRef](#)] [[PubMed](#)]

60. Zhao, Z.; Liao, Y.; Li, J.; Wu, J.; Zhang, Y.; Feng, G.; Tan, B.; Reng, S.; Zhang, Z.; Feng, X.; et al. Association between higher expression of YB-1 and poor prognosis in early-stage extranodal nasal-type natural killer/T-cell lymphoma. *Biomarkers Med.* **2014**, *8*, 581–588. [[CrossRef](#)]
61. Liu, Y.; Fan, L.; Wang, Y.; Li, P.; Zhu, J.; Wang, L.; Zhang, W.; Zhang, Y.; Huang, G. Tumor-associated macrophages promote tumor cell proliferation in nasopharyngeal NK/T-cell lymphoma. *Int. J. Clin. Exp. Pathol.* **2014**, *7*, 5429–5435. [[PubMed](#)]
62. Lin, Z.X.; Bai, B.; Cai, Q.C.; Cai, Q.Q.; Wang, X.X.; Wu, X.Y.; Huang, H.Q. High numbers of tumor-associated macrophages correlate with poor prognosis in patients with mature T- and natural killer cell lymphomas. *Med. Oncol.* **2012**, *29*, 3522–3528. [[CrossRef](#)]
63. Wang, H.; Li, P.; Wang, L.; Xia, Z.; Huang, H.; Lu, Y.; Li, Z. High numbers of CD68+ tumor-associated macrophages correlate with poor prognosis in extranodal NK/T-cell lymphoma, nasal type. *Ann. Hematol.* **2015**, *94*, 1535–1544. [[CrossRef](#)]
64. Kim, W.Y.; Jeon, Y.K.; Kim, T.M.; Kim, J.E.; Kim, Y.A.; Lee, S.H.; Kim, D.W.; Heo, D.S.; Kim, C.W. Increased quantity of tumor-infiltrating FOXP3-positive regulatory T cells is an independent predictor for improved clinical outcome in extranodal NK/T-cell lymphoma. *Ann. Oncol.* **2009**, *20*, 1688–1696. [[CrossRef](#)] [[PubMed](#)]
65. Chen, M.M.; Zeng, G.P.; Li, J.; Fu, J.H.; Long, Y.Y.; Pan, J.Y.; Wei, L.Z.; Guan, J.Y.; Lin, Y.X.; You, H.; et al. High infiltration of CD20(+) B lymphocytes in extranodal natural killer/T-cell lymphoma is associated with better prognosis. *Br. J. Haematol.* **2020**, *191*, e116–e120. [[CrossRef](#)] [[PubMed](#)]
66. Li, Y.J.; Li, Z.M.; Xia, Y.; Huang, J.J.; Huang, H.Q.; Xia, Z.J.; Lin, T.Y.; Li, S.; Cai, X.Y.; Wu-Xiao, Z.J.; et al. Serum C-reactive protein (CRP) as a simple and independent prognostic factor in extranodal natural killer/T-cell lymphoma, nasal type. *PLoS ONE* **2013**, *8*, e64158. [[CrossRef](#)] [[PubMed](#)]
67. Yamazaki, E.; Tomita, N.; Koyama, S.; Ogusa, E.; Ishii, Y.; Takahashi, H.; Miyashita, K.; Matsuura, S.; Tachibana, T.; Takasaki, H.; et al. Serum ferritin level is prognostic of patient outcome in extranodal NK/T cell lymphoma, nasal type. *Med. Oncol.* **2014**, *31*, 149. [[CrossRef](#)]
68. Zhou, Z.; Li, Z.; Sun, Z.; Zhang, X.; Lu, L.; Wang, Y.; Zhang, M. S100A9 and ORM1 serve as predictors of therapeutic response and prognostic factors in advanced extranodal NK/T cell lymphoma patients treated with pegaspargase/gemcitabine. *Sci. Rep.* **2016**, *6*, 23695. [[CrossRef](#)] [[PubMed](#)]
69. Qiu, Y.; Zhou, Z.; Li, Z.; Lu, L.; Li, L.; Li, X.; Wang, X.; Zhang, M. Pretreatment 14-3-3 epsilon level is predictive for advanced extranodal NK/T cell lymphoma therapeutic response to asparaginase-based chemotherapy. *Proteom. Clin. Appl.* **2017**, *11*, 1600111. [[CrossRef](#)] [[PubMed](#)]
70. Rong, Q.; Gao, Y.; Cai, Q.; Wang, X.; Bai, B.; Ping, L.; He, H.; Rao, H.; Zhang, Y.; Li, Z.; et al. High IL-6 expression in the tumor microenvironment is associated with poor prognosis of patients with extranodal natural/killer T-cell lymphoma (ENKTL). *Expert Rev. Anticancer. Ther.* **2021**, *21*, 121–127. [[CrossRef](#)]
71. Bao, C.; Zhou, D.; Zhu, L.; Qian, W.; Ye, X. Increased serum level of interleukin-6 correlates with negative prognostic factors in extranodal NK/T-cell lymphoma. *Transl. Cancer Res.* **2020**, *9*, 2378–2389. [[CrossRef](#)]
72. Wang, L.; Liao, D.Z.; Zhang, J.; Xia, Z.J.; Peng, X.W.; Lu, Y. Clinical significance of serum soluble interleukin-2 receptor- α in extranodal natural killer/T-cell lymphoma (ENKTL): A predictive biomarker for treatment efficacy and valuable prognostic factor. *Med. Oncol.* **2013**, *30*, 723. [[CrossRef](#)] [[PubMed](#)]
73. Zhang, J.; Wang, W.-D.; Geng, Q.-R.; Wang, L.; Chen, X.-Q.; Liu, C.-C.; Lv, Y. Serum Levels of Interleukin-9 Correlate with Negative Prognostic Factors in Extranodal NK/T-Cell Lymphoma. *PLoS ONE* **2014**, *9*, e94637. [[CrossRef](#)]
74. Wang, H.; Zhu, J.Y.; Liu, C.C.; Zhu, M.Y.; Wang, J.H.; Geng, Q.R.; Lu, Y. Increased serum levels of interleukin-15 correlate with negative prognostic factors in extranodal NK/T cell lymphoma. *Med. Oncol.* **2014**, *32*, 370. [[CrossRef](#)]
75. Wang, H.; Wang, L.; Wuxiao, Z.; Huang, H.; Jiang, W.; Li, Z.; Lu, Y.; Xia, Z. Increased serum levels of interleukin-10 predict poor prognosis in extranodal natural killer/T-cell lymphoma patients receiving asparaginase-based chemotherapy. *Oncotargets Ther.* **2015**, *8*, 2589–2599. [[CrossRef](#)]
76. Kim, H.S.; Ryu, K.J.; Ko, Y.H.; Kim, H.J.; Kim, S.H.; Kim, W.S.; Kim, S.J. Macrophage inflammatory protein 1 alpha (MIP-1alpha) may be associated with poor outcome in patients with extranodal NK/T-cell lymphoma. *Hematol. Oncol.* **2016**, *35*, 310–316. [[CrossRef](#)] [[PubMed](#)]
77. Lim, S.W.; Ryu, K.J.; Lee, H.; Ko, Y.H.; Kim, W.S.; Kim, S.J. Serum IL18 is associated with hemophagocytosis and poor survival in extranodal natural killer/T-cell lymphoma. *Leuk. Lymphoma* **2018**, *60*, 317–325. [[CrossRef](#)]
78. Kim, W.Y.; Jung, H.Y.; Nam, S.J.; Kim, T.M.; Heo, D.S.; Kim, C.W.; Jeon, Y.K. Expression of programmed cell death ligand 1 (PD-L1) in advanced stage EBV-associated extranodal NK/T cell lymphoma is associated with better prognosis. *Virchows Arch.* **2016**, *469*, 581–590. [[CrossRef](#)]
79. Bi, X.W.; Wang, H.; Zhang, W.W.; Wang, J.H.; Liu, W.J.; Xia, Z.J.; Huang, H.Q.; Jiang, W.Q.; Zhang, Y.J.; Wang, L. PD-L1 is upregulated by EBV-driven LMP1 through NF- κ B pathway and correlates with poor prognosis in natural killer/T-cell lymphoma. *J. Hematol. Oncol.* **2016**, *9*, 109. [[CrossRef](#)]
80. Nagato, T.; Ohkuri, T.; Ohara, K.; Hirata, Y.; Kishibe, K.; Komabayashi, Y.; Ueda, S.; Takahara, M.; Kumai, T.; Ishibashi, K.; et al. Programmed death-ligand 1 and its soluble form are highly expressed in nasal natural killer/T-cell lymphoma: A potential rationale for immunotherapy. *Cancer Immunol. Immunother.* **2017**, *66*, 877–890. [[CrossRef](#)]
81. Zeng, L.; Huang, W.; Cao, Z.; Zheng, B.; Liu, X.; Guo, L.; Feng, X. The correlation of clinicopathological features and prognosis in extranodal natural killer/T cell lymphoma: A report of 42 cases in the early stage. *Ann. Hematol.* **2019**, *98*, 1467–1476. [[CrossRef](#)]

82. Muhamad, H.; Suksawai, N.; Assanasen, T.; Polprasert, C.; Bunworasate, U.; Wudhikarn, K. Programmed Cell Death 1 and Programmed Cell Death Ligands in Extranodal Natural Killer/T Cell Lymphoma: Expression Pattern and Potential Prognostic Relevance. *Acta Haematol.* **2019**, *143*, 78–88. [[CrossRef](#)]
83. Jo, J.C.; Kim, M.; Choi, Y.; Kim, H.J.; Kim, J.E.; Chae, S.W.; Kim, H.; Cha, H.J. Expression of programmed cell death 1 and programmed cell death ligand 1 in extranodal NK/T-cell lymphoma, nasal type. *Ann. Hematol.* **2016**, *96*, 25–31. [[CrossRef](#)]
84. Chan, J.Y.; Lim, J.Q.; Ong, C.K. Checkpoint immunotherapy for NK/T cell lymphoma—Time for a showdown? *Precis. Clin. Med.* **2021**, *4*, 70–72. [[CrossRef](#)]
85. Wang, H.; Wang, L.; Liu, W.J.; Xia, Z.J.; Huang, H.Q.; Jiang, W.Q.; Li, Z.M.; Lu, Y. High post-treatment serum levels of soluble programmed cell death ligand 1 predict early relapse and poor prognosis in extranodal NK/T cell lymphoma patients. *Oncotarget* **2016**, *7*, 33035–33045. [[CrossRef](#)] [[PubMed](#)]
86. Feng, Y.; Jing, C.; Yu, X.; Cao, X.; Xu, C. Predicting treatment response of patients with extranodal natural killer/T-cell lymphoma based on levels of PD-L1 mRNA and soluble PD-L1. *Hematol. Oncol.* **2020**, *38*, 467–477. [[CrossRef](#)]
87. Li, J.W.; Wei, P.; Guo, Y.; Shi, D.; Yu, B.H.; Su, Y.F.; Li, X.Q.; Zhou, X.Y. Clinical significance of circulating exosomal PD-L1 and soluble PD-L1 in extranodal NK/T-cell lymphoma, nasal-type. *Am. J. Cancer Res.* **2020**, *10*, 4498–4512. [[PubMed](#)]
88. Lei, K.I.; Chan, L.Y.; Chan, W.Y.; Johnson, P.J.; Lo, Y.M. Diagnostic and prognostic implications of circulating cell-free Epstein-Barr virus DNA in natural killer/T-cell lymphoma. *Clin. Cancer Res.* **2002**, *8*, 29–34. [[PubMed](#)]
89. Au, W.Y.; Pang, A.; Choy, C.; Chim, C.S.; Kwong, Y.L. Quantification of circulating Epstein-Barr virus (EBV) DNA in the diagnosis and monitoring of natural killer cell and EBV-positive lymphomas in immunocompetent patients. *Blood* **2004**, *104*, 243–249. [[CrossRef](#)]
90. Ishii, H.; Ogino, T.; Berger, C.; Köchli-Schmitz, N.; Nagato, T.; Takahara, M.; Nadal, D.; Harabuchi, Y. Clinical usefulness of serum EBV DNA levels of BamHI W and LMP1 for Nasal NK/T-cell lymphoma. *J. Med. Virol.* **2007**, *79*, 562–572. [[CrossRef](#)]
91. Suzuki, R.; Yamaguchi, M.; Izutsu, K.; Yamamoto, G.; Takada, K.; Harabuchi, Y.; Isobe, Y.; Gomyo, H.; Koike, T.; Okamoto, M.; et al. Prospective measurement of Epstein-Barr virus–DNA in plasma and peripheral blood mononuclear cells of extranodal NK/T-cell lymphoma, nasal type. *Blood* **2011**, *118*, 6018–6022. [[CrossRef](#)] [[PubMed](#)]
92. Campo, E.; Swerdlow, S.H.; Harris, N.L.; Pileri, S.; Stein, H.; Jaffe, E.S. The 2008 WHO classification of lymphoid neoplasms and beyond: Evolving concepts and practical applications. *Blood* **2011**, *117*, 5019–5032. [[CrossRef](#)]
93. Kim, H.S.; Kim, K.H.; Kim, K.H.; Chang, M.H.; Ji, S.H.; Lim, D.H.; Kim, K.; Kim, S.J.; Ko, Y.; Ki, C.-S.; et al. Whole blood Epstein-Barr virus DNA load as a diagnostic and prognostic surrogate: Extranodal natural killer/T-cell lymphoma. *Leuk. Lymphoma* **2009**, *50*, 757–763. [[CrossRef](#)] [[PubMed](#)]
94. Wang, Z.Y.; Liu, Q.F.; Wang, H.; Jin, J.; Wang, W.H.; Wang, S.L.; Song, Y.W.; Liu, Y.P.; Fang, H.; Ren, H.; et al. Clinical implications of plasma Epstein-Barr virus DNA in early-stage extranodal nasal-type NK/T-cell lymphoma patients receiving primary radiotherapy. *Blood* **2012**, *120*, 2003–2010. [[CrossRef](#)] [[PubMed](#)]
95. Kwong, Y.L.; Pang, A.W.; Leung, A.Y.; Chim, C.S.; Tse, E. Quantification of circulating Epstein-Barr virus DNA in NK/T-cell lymphoma treated with the SMILE protocol: Diagnostic and prognostic significance. *Leukemia* **2013**, *28*, 865–870. [[CrossRef](#)]
96. Wang, L.; Wang, H.; Wang, J.H.; Xia, Z.J.; Lu, Y.; Huang, H.Q.; Jiang, W.Q.; Zhang, Y.J. Post-treatment plasma EBV-DNA positivity predicts early relapse and poor prognosis for patients with extranodal NK/T cell lymphoma in the era of asparaginase. *Oncotarget* **2015**, *6*, 30317–30326. [[CrossRef](#)] [[PubMed](#)]
97. Kim, S.J.; Choi, J.Y.; Hyun, S.H.; Ki, C.S.; Oh, D.; Ahn, Y.C.; Ko, Y.H.; Choi, S.; Jung, S.H.; Khong, P.L.; et al. Risk stratification on the basis of Deauville score on PET-CT and the presence of Epstein-Barr virus DNA after completion of primary treatment for extranodal natural killer/T-cell lymphoma, nasal type: A multicentre, retrospective analysis. *Lancet Haematol.* **2015**, *2*, e66–e74. [[CrossRef](#)]
98. Cho, J.; Kim, S.J.; Park, S.; Yoo, K.H.; Ki, C.S.; Ko, Y.; Kim, W.S. Significance of circulating Epstein-Barr virus DNA monitoring after remission in patients with extranodal natural killer T cell lymphoma. *Ann. Hematol.* **2018**, *97*, 1427–1436. [[CrossRef](#)]
99. Li, P.F.; Mao, Y.Z.; Bai, B.; Gao, Y.; Zhang, Y.J.; Li, Z.M.; Jiang, W.Q.; Huang, H.Q. Persistent peripheral blood EBV-DNA positive with high expression of PD-L1 and upregulation of CD4 + CD25 + T cell ratio in early stage NK/T cell lymphoma patients may predict worse outcome. *Ann. Hematol.* **2018**, *97*, 2381–2389. [[CrossRef](#)]
100. Kanno, H.; Kojya, S.; Li, T.; Ohsawa, M.; Nakatsuka, S.; Miyaguchi, M.; Harabuchi, Y.; Aozasa, K. Low frequency of HLA-A*0201 allele in patients with Epstein-Barr virus-positive nasal lymphomas with polymorphic reticulosis morphology. *Int. J. Cancer* **2000**, *87*, 195–199. [[CrossRef](#)]
101. Koo, G.C.; Tan, S.Y.; Tang, T.; Poon, S.L.; Allen, G.E.; Tan, L.; Chong, S.C.; Ong, W.S.; Tay, K.; Tao, M.; et al. Janus Kinase 3–Activating Mutations Identified in Natural Killer/T-cell Lymphoma. *Cancer Discov.* **2012**, *2*, 591–597. [[CrossRef](#)] [[PubMed](#)]
102. Lee, S.; Park, H.Y.; Kang, S.Y.; Kim, S.J.; Hwang, J.; Lee, S.; Kwak, S.H.; Park, K.S.; Yoo, H.Y.; Kim, W.S.; et al. Genetic alterations of JAK/STAT cascade and histone modification in extranodal NK/T-cell lymphoma nasal type. *Oncotarget* **2015**, *6*, 17764–17776. [[CrossRef](#)]
103. Song, T.L.; Nairismägi, M.L.; Laurensia, Y.; Lim, J.Q.; Tan, J.; Li, Z.M.; Pang, W.L.; Kizhakeyil, A.; Wijaya, G.C.; Huang, D.C.; et al. Oncogenic activation of the STAT3 pathway drives PD-L1 expression in natural killer/T-cell lymphoma. *Blood* **2018**, *132*, 1146–1158. [[CrossRef](#)] [[PubMed](#)]

104. Kataoka, K.; Miyoshi, H.; Sakata, S.; Dobashi, A.; Couronné, L.; Kogure, Y.; Sato, Y.; Nishida, K.; Gion, Y.; Shiraishi, Y.; et al. Frequent structural variations involving programmed death ligands in Epstein-Barr virus-associated lymphomas. *Leukemia* **2019**, *33*, 1687–1699. [[CrossRef](#)] [[PubMed](#)]
105. Hu, B.; Oki, Y. Novel Immunotherapy Options for Extranodal NK/T-Cell Lymphoma. *Front. Oncol.* **2018**, *8*, 139. [[CrossRef](#)]
106. Kwong, Y.L.; Chan, T.S.Y.; Tan, D.; Kim, S.J.; Poon, L.M.; Mow, B.; Khong, P.L.; Loong, F.; Au-Yeung, R.; Iqbal, J.; et al. PD1 blockade with pembrolizumab is highly effective in relapsed or refractory NK/T-cell lymphoma failing l-asparaginase. *Blood* **2017**, *129*, 2437–2442. [[CrossRef](#)]
107. Li, X.; Cheng, Y.; Zhang, M.; Yan, J.; Li, L.; Fu, X.; Zhang, X.; Chang, Y.; Sun, Z.; Yu, H.; et al. Activity of pembrolizumab in relapsed/refractory NK/T-cell lymphoma. *J. Hematol. Oncol.* **2018**, *11*, 15. [[CrossRef](#)]
108. Kim, S.J.; Lim, J.Q.; Laurensia, Y.; Cho, J.; Yoon, S.E.; Lee, J.Y.; Ryu, K.J.; Ko, Y.H.; Koh, Y.; Cho, D.; et al. Avelumab for the treatment of relapsed or refractory extranodal NK/T-cell lymphoma: An open-label phase 2 study. *Blood* **2020**, *136*, 2754–2763. [[CrossRef](#)]
109. Li, J.; Tao, R.; Fan, L.; Song, Y.; Hu, Y.; Zhang, W.; Wang, Y.; Xu, L.; Sun, X.; Zhou, H. Sintilimab for relapsed/refractory (r/r) extranodal NK/T cell lymphoma (ENKTL): Extended follow-up on the multicenter, single-arm phase II trial (ORIENT-4). *J. Clin. Oncol.* **2020**, *38*, 8050. [[CrossRef](#)]
110. Lee, J.Y.; Kim, J.H.; Bang, H.; Cho, J.; Ko, Y.H.; Kim, S.J.; Kim, W.S. EGR1 as a potential marker of prognosis in extranodal NK/T-cell lymphoma. *Sci. Rep.* **2021**, *11*, 10342. [[CrossRef](#)]
111. Rossi, D.; Spina, V.; Brusca, G.; Gaidano, G. Liquid biopsy in lymphoma. *Haematologica* **2019**, *104*, 648–652. [[CrossRef](#)] [[PubMed](#)]
112. Li, Q.; Zhang, W.; Li, J.; Xiong, J.; Liu, J.; Chen, T.; Wen, Q.; Zeng, Y.; Gao, L.; Zhang, C.; et al. Plasma circulating tumor DNA assessment reveals KMT2D as a potential poor prognostic factor in extranodal NK/T-cell lymphoma. *Biomark. Res.* **2020**, *8*, 27. [[CrossRef](#)]
113. Yamanaka, Y.; Tagawa, H.; Takahashi, N.; Watanabe, A.; Guo, Y.-M.; Iwamoto, K.; Yamashita, J.; Saitoh, H.; Kameoka, Y.; Shimizu, N.; et al. Aberrant overexpression of microRNAs activate AKT signaling via down-regulation of tumor suppressors in natural killer-cell lymphoma/leukemia. *Blood* **2009**, *114*, 3265–3275. [[CrossRef](#)] [[PubMed](#)]
114. Ng, S.B.; Yan, J.; Huang, G.; Selvarajan, V.; Tay, J.L.; Lin, B.; Bi, C.; Tan, J.; Kwong, Y.L.; Shimizu, N.; et al. Dysregulated microRNAs affect pathways and targets of biologic relevance in nasal-type natural killer/T-cell lymphoma. *Blood* **2011**, *118*, 4919–4929. [[CrossRef](#)] [[PubMed](#)]
115. Paik, J.H.; Jang, J.Y.; Jeon, Y.K.; Kim, W.Y.; Kim, T.M.; Heo, D.S.; Kim, C.W. MicroRNA-146a downregulates NFkappaB activity via targeting TRAF6 and functions as a tumor suppressor having strong prognostic implications in NK/T cell lymphoma. *Clin. Cancer Res.* **2011**, *17*, 4761–4771. [[CrossRef](#)] [[PubMed](#)]
116. Komabayashi, Y.; Kishibe, K.; Nagato, T.; Ueda, S.; Takahara, M.; Harabuchi, Y. Downregulation of miR-15a due to LMP1 promotes cell proliferation and predicts poor prognosis in nasal NK/T-cell lymphoma. *Am. J. Hematol.* **2013**, *89*, 25–33. [[CrossRef](#)]
117. Guo, H.Q.; Huang, G.L.; Guo, C.C.; Pu, X.X.; Lin, T.Y. Diagnostic and prognostic value of circulating miR-221 for extranodal natural killer/T-cell lymphoma. *Dis. Markers* **2010**, *29*, 251–258. [[CrossRef](#)]
118. Zhang, X.; Ji, W.; Huang, R.; Li, L.; Wang, X.; Li, L.; Fu, X.; Sun, Z.; Li, Z.; Chen, Q.; et al. MicroRNA-155 is a potential molecular marker of natural killer/T-cell lymphoma. *Oncotarget* **2016**, *7*, 53808–53819. [[CrossRef](#)] [[PubMed](#)]
119. Ryu, K.J.; Lee, J.Y.; Choi, M.E.; Yoon, S.E.; Cho, J.; Ko, Y.H.; Shim, J.H.; Kim, W.S.; Park, C.; Kim, S.J. Serum-Derived Exosomal MicroRNA Profiles Can Predict Poor Survival Outcomes in Patients with Extranodal Natural Killer/T-Cell Lymphoma. *Cancers* **2020**, *12*, 3548. [[CrossRef](#)]
120. Komabayashi, Y.; Kishibe, K.; Nagato, T.; Ueda, S.; Takahara, M.; Harabuchi, Y. Circulating Epstein-Barr virus-encoded microRNAs as potential biomarkers for nasal natural killer/T-cell lymphoma. *Hematol. Oncol.* **2016**, *35*, 655–663. [[CrossRef](#)]