

PAPER

Inhibition of testosterone synthesis induced by oral TiO₂ NPs is associated with ROS-MAPK(ERK1/2)-StAR signaling pathway in SD rat

Shanji Liu,¹ Yizhou Tang,¹ Bolu Chen,¹ Yu Zhao,¹ Zoraida P. Aguilar,² Xueying Tao^{1,*} and Hengyi Xu^{1,*}

¹State Key Laboratory of Food Science and Technology, Nanchang University, Nanchang 330047, China and

²Zystein, LLC., Fayetteville, AR 72704, USA

*Correspondence address. State Key Laboratory of Food Science and Technology, Nanchang University, 235 Nanjing East Road, Nanchang 330047, China. Tel: +0086-791-8830-4447-ext-9520; Fax: +0086-791-8830-4400; E-mail: kidyxu@163.com, HengyiXu@ncu.edu.cn (Hengyi Xu); Tel: +0086-791-8833-4578; Fax: +0086-791-8833-3708; E-mail: 1027txy@163.com, 407867172@qq.com (Xueying Tao)

Abstract

Titanium dioxide nanoparticles (TiO₂ NPs) have been widely used in food, medical, and other fields; their reproductive toxicity has been reported in numerous studies. However, the relevant toxicity mechanism still requires further exploration. In this paper, the effect of oral exposure to 500 mg/kg TiO₂ NPs (anatase and rutile) in adult male SD rats was studied over 3 and 7 days. Results showed that the total sperm count and testosterone level of 7 days of exposure in serum decreased in the experimental group. Testicular tissue lesions, such as disappearance of Leydig cells, disorder of arrangement of spermatogenic cells in the lumen of convoluted seminiferous tubules, and disorder of arrangement of germ cells, were observed. Meanwhile, the expression of steroidogenic acute regulatory (StAR; the key factors of testosterone synthesis), MAPK (ERK1/2), and phosphorylated ERK1/2 in testes of SD rats after exposure to TiO₂ NPs for 7 days decreased, while the malondialdehyde content increased and superoxide dismutase activity decreased in serum. The present study showed that TiO₂ NPs could cause reproductive toxicity. Notably, anatase is more toxic than rutile. In addition, exposure to 500 mg/kg TiO₂ NPs for 7 days inhibited testosterone synthesis in male rat, which may be related to the reactive oxygen species (ROS)-MAPK (ERK1/2)-StAR signal pathway. Warning that the use of TiO₂ NPs should be regulated.

Highlights

- Titanium dioxide nanoparticles (TiO₂ NPs) induced reproductive toxicity in the SD rats.
- Inhibition of MAPK (ERK1/2) expression of ROS induced by TiO₂ NPs.
- Inhibition of testosterone synthesis induced by oral TiO₂ NPs is associated with ROS-MAPK(ERK1/2)-StAR signaling pathway in SD rats.

Key words: titanium dioxide nanoparticle, smale reproductive toxicity, reactive oxygen species, ERK1/2, testosterone synthesis

Received: 12 April 2021; Revised: 6 July 2021; Accepted: 19 July 2021

© The Author(s) 2021. Published by Oxford University Press. All rights reserved. For Permissions, please email: journals.permissions@oup.com

Introduction

In recent years, nanomaterials have been widely used in various fields, especially in food additives and food contact materials [1]. Nano-silver, titanium dioxide, and silicon dioxide are the most commonly mentioned nanomaterials in literature [2]. Titanium dioxide nanoparticles (TiO₂ NPs) is one of the most widely used nanomaterials in varying fields, including medicine [3], agriculture [4], bacteriostasis [5], wastewater treatment [6], personal care products [7], cosmetics, sunscreen, toothpaste, paint, and food [8]. TiO₂ NPs are inevitably released into the environment due to widespread use, which will harm individual organisms and ecosystems. TiO₂ NPs mainly exist in three different natural crystal forms, namely, anatase, rutile, and brookite [9]. Anatase and rutile are more commonly used than brookite; moreover, studies have reported that the surface activities of anatase and rutile differ [10]. Thus, these two kinds of crystal materials were selected for this study. Given the wide range of uses and special physical properties (small size), TiO₂ NPs are transferred to the human body through inhalation, environmental intake, and medical applications (skin exposure) [11, 12] and then enter different organs through blood circulation, such as the heart, liver, lung, brain, and testis [13, 14]. According to previous studies, after continuous exposure for 14 days, the deposition of TiO₂ NPs in various organs of mice is in the following order: liver > kidney > spleen > lung > brain > heart [15]. TiO₂ NPs enter the testis through the blood–testis barrier (BTB) due to their small size and then migrate to the testicular microenvironment composed of Sertoli cells, sperm cells, and Leydig cells (LCs). They then destroy the normal structure of the testis [16, 17] and inhibit testosterone production; lead to a decline in sperm quality; and cause a decrease in LCs and sperm count, sperm motility, and other male fertility problems [18, 19]. However, studies on the mechanism of TiO₂ NPs inhibiting testosterone production are few.

The process of testosterone synthesis has been reported [20]. First, cholesterol is carried from the outer membrane of the mitochondria to the intima through steroidogenic acute regulatory (StAR), and its side chain is cleaved and converted into pregnenolone by cytochrome P450 family 11 (Cyp11a1) in LCs [21]. Pregnenolone is transported to the smooth endoplasmic reticulum in the cytoplasm and forms testosterone through the reaction catalyzed by 3 β -hydroxysteroid dehydrogenase (3 β HSD) and 17 β HSD [22, 23]. StAR is an acute regulatory protein [24], which regulates the transfer of cholesterol to the mitochondria. Otherwise, it is considered a rate-limiting step in testosterone synthesis and is regulated by extracellular regulated protein kinase 1/2 (ERK1/2) [25, 26]. More specifically, the upstream of StAR gene expression is regulated by phosphorylated ERK1/2 (Only phosphorylated ERK1/2 is active) [27]. ERK1/2, a member of the mitogen-activated protein kinase (MAPK) family [28], has been shown to promote the transcription of the StAR gene and may be involved in testosterone synthesis [29, 30]. In addition, previous studies have shown that MAPK is activated by reactive oxygen species (ROS) [31, 32], which may affect the normal expression of MAPK (ERK1/2) in the testis and affect testosterone synthesis. Notably, a large number of studies proved that the entry of TiO₂ NPs into the body may break the oxidation balance and generate ROS, resulting in a series of adverse effects, such as hepatotoxicity, reproductive toxicity, and genotoxicity [33, 34].

Here, we hypothesized that oral exposure to TiO₂ NPs in male rats may lead to the production of ROS and further activation of MAPK (ERK1/2) by ROS. ERK and StAR regulate each other [35]. Interference with the expression of ERK1/2 will lead to the

inability of StAR to transport cholesterol to the mitochondria normally, resulting in the inhibition of testosterone synthesis in the testis.

In this study, the adverse effects of anatase and rutile TiO₂ NPs on male reproductive function were evaluated by analyzing testicular histopathology, sperm count, and testosterone levels. The potential mechanism underlying the decreased testosterone synthesis pathway induced by TiO₂ NPs was investigated through 3 and 7 days of oral exposure in male rats.

Materials and Methods

Characterization of TiO₂ NPs

TiO₂ NPs of anatase and rutile types (40 nm \pm 5 nm) were obtained from Aladdin industrial Corporation (Shanghai, China). The size of TiO₂ NPs was characterized by emission scanning electron microscopy (SEM) (JSM 6701F, JEOL Ltd, USA) after being dispersed by anhydrous ethanol and more than 100 nanoparticles sizes were measured by Image J software.

Animals treatment

Approximately 8-week-old adult male Sprague Dawley (SD) rats were purchased from the experimental animal center of Nanchang University (Nanchang, China). And then, the animals were kept in non-toxic and harmless cages in a room with a temperature at 22 \pm 2°C and humidity of 45 \pm 5% and 12 h cycle of dark and light. Animals have free access to clean drinking water and commercial food. Before the end of the experiment, clean the cage every 3 days in order to maintain a comfortable environment. The animal trials program was carried out with reference to the guidelines for experimental animal welfare and approved by the animal care review committee (approval No. 0064257), Nanchang University.

Adapting for a week, the animals were randomly divided into control, anatase, and rutile groups ($n = 5$). The study of Ali showed that exposure to 500 mg/kg TiO₂ NPs had oxidative damage in mice [36]. The rats in the experimental group were oral administered with 500 mg/kg TiO₂ NPs for 3 and 7 days in this study. Meanwhile, the control group was fed with the same amount of solvent. The clinical symptoms of rats were observed every day during the experiment. Twenty-four hours after the last administration, serum and testicular tissue samples of rats were obtained and stored in the refrigerator at -80°C .

Sperm counts

The assessment of testicular sperm count was adopted from the studies reported by Kisin *et al.* [37], and we made minor modifications. The isolated left epididymis was completely cut up and then put into 1 ml of saline and incubated at 37°C for 15 min in order to fully dissociate the sperm. 200 μl sperm sample was killed with hot water at 90°C, and then 10 μl sample was put into the blood cell counter to observe and determine the sperm count under light microscope (Nikon eclipse Ti). All studies were in triplicate.

Assay for testosterone in serum

The effects of TiO₂ NPs exposure over 3 and 7 days on the testosterone levels in serum were evaluated. According to the

instructions provided by the manufacturer, we used ELISA kit (Item No. 582701) to determine the content of testosterone in serum samples. Absorbance was recorded between 405 and 420 nm with the microplate reader (Thermo Scientific, USA). The plate should be read when the absorbance of the B₀ wells is in the range of 0.3–1.0 A.U (blank subtracted). The accurate testosterone content was calculated according to the standard curve.

Histopathology examination

Histopathological damage of testis exposed to TiO₂ NPs was evaluated. The testes of rat were extracted and stored in Bonn fixed solution for 24 h, then 75% alcohol was replaced for storage. The paraffin blocks were obtained by embedding the sample in paraffin after dehydration of ethanol and transparency of xylene, which were cut at 5 μm thickness and stained with hematoxylin and eosin. The stained sections were fixed on glass slides and magnified 200 and 400 times, respectively, under optical microscope to observe the seminiferous tubules, lumen, interstitial area, and germ cells.

Real-time quantitative polymerase chain reaction

According to Axyprep™ Multisource Total RNA Miniprep kit instructions (Takara Bio Inc.), total RNA was extracted from testis and reverse transcribed into cDNA. The primers were designed with NCBI Primer and Oligo Primer Analysis Software version 7.0 (Molecular Biology Insights, Inc.; DBA Oligo, Inc.). And then, the primers were synthesized by Qingke Biology Co, Ltd (Shanghai, China). In the AriaMx Real-time quantitative polymerase chain reaction (RT-qPCR) system (MY 19435252), we used the three-step method, that is 1 min at 95°C followed by 40 cycles of 95°C for 5 s, 59°C for 60s, 72°C for 30 s. The relative quantification of mRNA was calculated by 2^{-ΔΔCt} method and GAPDH was used as the internal reference gene.

Immunohistochemistry (IHC) analysis

Paraffin sections of testicular tissue were dewaxed with xylene and ethanol and then placed in citric acid antigen repair buffer (pH 6.0) to repair antigens. The sections were incubated in 3% hydrogen peroxide and kept away from light at room temperature for 25 min in order to block the endogenous peroxidase. Next, add 3% BSA covering tissue and seal at room temperature for 30 min. After dripping primary antibody against StAR and pERK1/2 (1:500) and incubating overnight at 4°C, and adding corresponding secondary antibody (HRP marked, Servicebio, GB23303) for 50 min at room temperature. Finally, the sections were developed with diaminobenzidine (DAB). IHC was quantified with reference to the previous method and quantified with Image J software. The results obtained were converted into scores using $[\sum Pi(i + 1)]$ formula (i: staining intensity score; Pi: percentage of stained contribution, negative: 0, low positive: 1, positive: 2, high positive: 3).

Detection of superoxide dismutase, catalase and malondialdehyde

The activities of superoxide dismutase (SOD) and catalase (CAT) as well as the content of malondialdehyde (MDA) in

serum were measured according to the instructions of the kit manufacturer, which purchased from Jiancheng Bio-tech Co. Ltd (Nanjing, China).

Statistical analysis

All the reported data were displayed with mean ± standard deviation (SD), which was analyzed using the SPSS (version 22) software. Differences between groups were analyzed by one-way analysis of variance. Significant differences were expressed by * (*P < 0.05, **P < 0.01, ***P < 0.001). Specially, at least three independent parallel experiments were conducted in each group to ensure the authenticity of the reported data.

Results

Characterization of TiO₂ NPs

To explore the dispersion of nanoparticles in solution and the true particle size and shape, this study was observed by SEM. The results showed that the two types of TiO₂ NPs were spherical with an average diameter of anatase of 37 nm and rutile 46 nm (Fig. 1).

Sperm counts

After exposure to different crystalline TiO₂ NPs for 3 and 7 days, the spermatozoa of the left epididymis were collected and counted with the blood cell count plate. Compared with the control group, the total sperm count of the anatase and rutile groups after oral administration for 3 (Fig. 2A) and 7 days (Fig. 2B) decreased significantly (P < 0.001). In addition, the total sperm count of the anatase group was significantly lower than that of the rutile group (P < 0.05). The results showed that acute exposure to different crystal forms of TiO₂ NPs had adverse effects on sperm survival in the testes of SD male rats, and the effect of anatase was greater than that of rutile.

Histopathological evaluation

The histopathological evaluations of the testicular sections and histomorphology of the control group revealed normal findings. The seminiferous tubules were closely connected and arranged neatly; the interstitial region was intact; germ cells were abundant in the control group; and spermatogonia, spermatocytes, and sperm cells were clearly observed (Fig. 3A-a and B-a). In the anatase group exposed for 3 (Fig. 3A-b) and 7 days (Fig. 3B-b), the number of LCs disappeared; spermatogenic disorder appeared in the lumen of seminiferous tubules; interestingly, germ cells were randomly arranged, the number decreased, and spermatocytes were exfoliated and vacuolated. The pathological condition in the rutile group was similar to that in the anatase group, but Fig. 3A-c and B-c shows that the pathological condition was not poor. Therefore, acute exposure to TiO₂ NPs could cause damage to the testis of animals and interfere with the production of spermatozoa.

Testosterone levels

The change in the testosterone level determined by an ELISA kit is shown in Fig. 4. The results showed that the testosterone levels in serum exposed for 7 days were lower in the treatment group than in the control group, and the anatase levels were lower

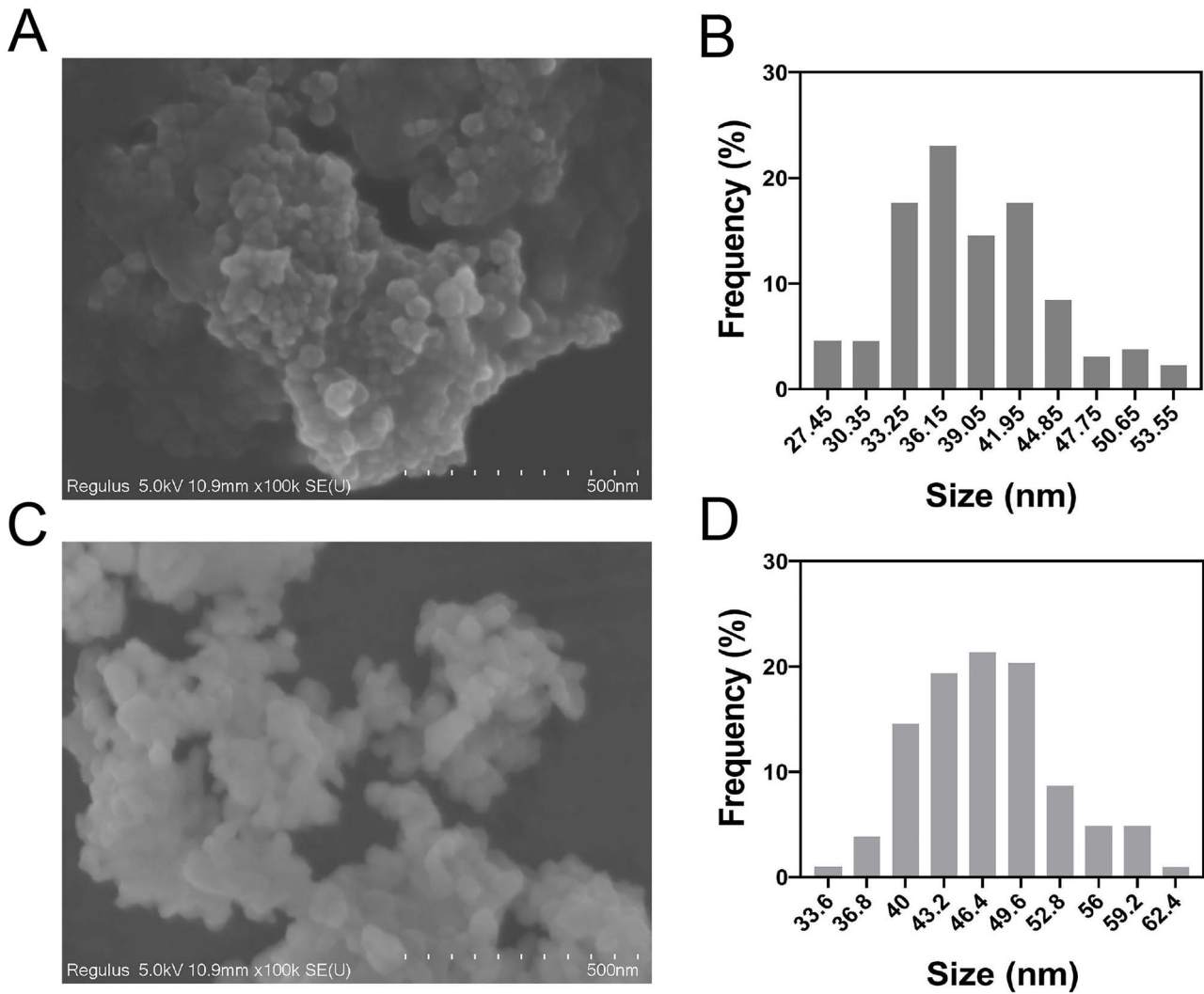


Figure 1: Characterization of TiO₂ NPs. (A, C) SEM image of anatase and rutile TiO₂ NPs. (B, D) The frequency of the size distribution of anatase and rutile TiO₂ NPs.

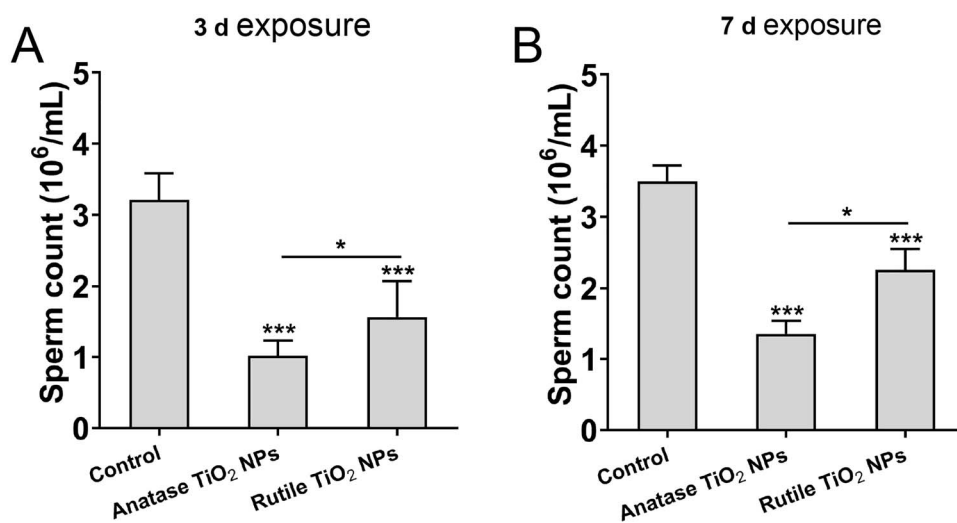


Figure 2: The number of sperms in the left epididymis of male rat after oral administration of 40 nm TiO₂ NPs. The data are presented in the form of sperm per milliliter of saline. (A) 3-day exposure, (B) 7-day exposure. Data are expressed as mean ± SD. *p < 0.05, ***p < 0.001 compared with control.

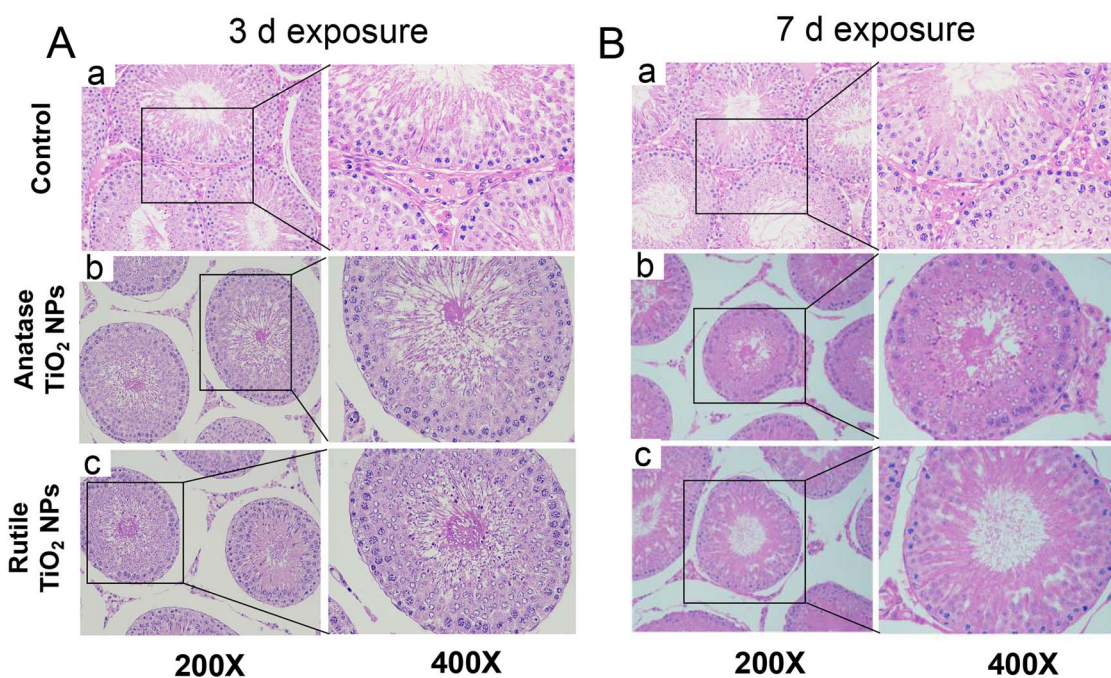


Figure 3: Light microscopy of cross sections of hematoxylin and eosin-stained testes from male rat. (A) 3-day exposure, (B) 7-day exposure.

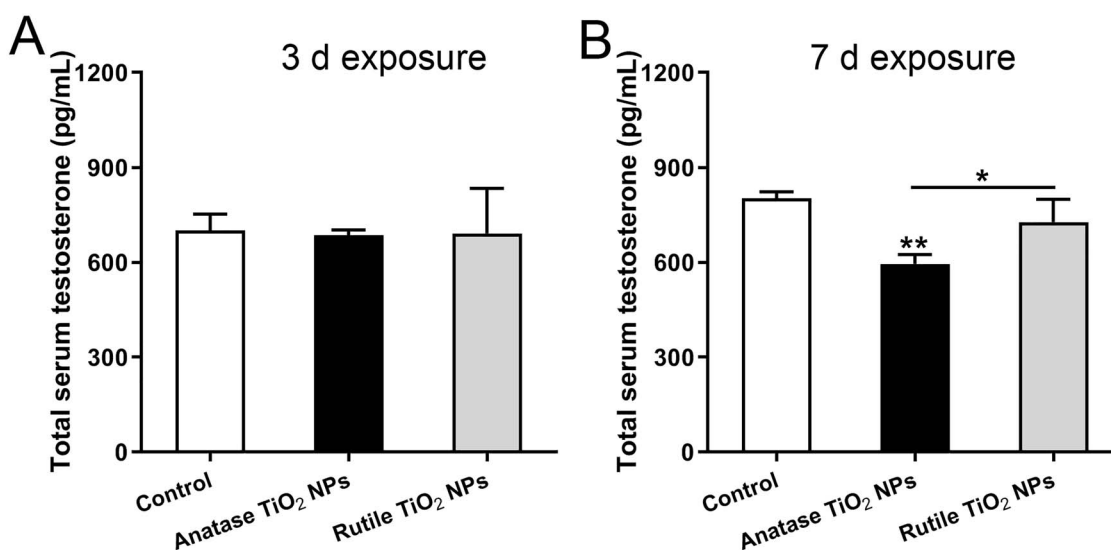


Figure 4: Testosterone concentration in male rat serum detected by ELISA. (A) 3-day exposure (B) 7-day exposure. Data are expressed as mean \pm SD. * $P < 0.05$, ** $P < 0.01$ compared with control.

in the treatment group than in the rutile group. These findings indicated that acute exposure to different crystal forms of TiO_2 NPs could affect the synthesis of testosterone, and the effect of anatase was more serious than that of rutile.

Change in relative gene quantification in testis

In this study, the changes in gene quantification related to ROS, MAPK (ERK1/2), and testosterone production were assessed by RT-qPCR. The primer information of related genes is shown in Table 1. In the 3-day exposure group, the expression levels of ROS-associated gene of nuclear factor erythroid 2-related factor 2 (*Nrf2*), NAD(P)H quinone dehydrogenase 1 (*NQO1*), glutathione

(GSH), CAT, superoxide dismutase 2 (*SOD2*), and MAPK (ERK1/2) were significantly downregulated in the experimental groups compared with the control groups. The testosterone production-associated gene of *StAR* was markedly upregulated in the experimental groups. By contrast, *Cyp11a1* was markedly downregulated in the anatase group and upregulated in the rutile group, whereas *17 β HSD* was significantly upregulated in the anatase group and downregulated in the rutile group. Similarly, in the 7-day exposure group, the ROS-associated gene of GSH was significantly upregulated, whereas *HO-1* was downregulated in the experimental groups. CAT and *NQO1* were downregulated in the rutile group, while *Nrf2* showed no significant alteration. MAPK (ERK1/2) was significantly downregulated in the anatase groups. The testosterone

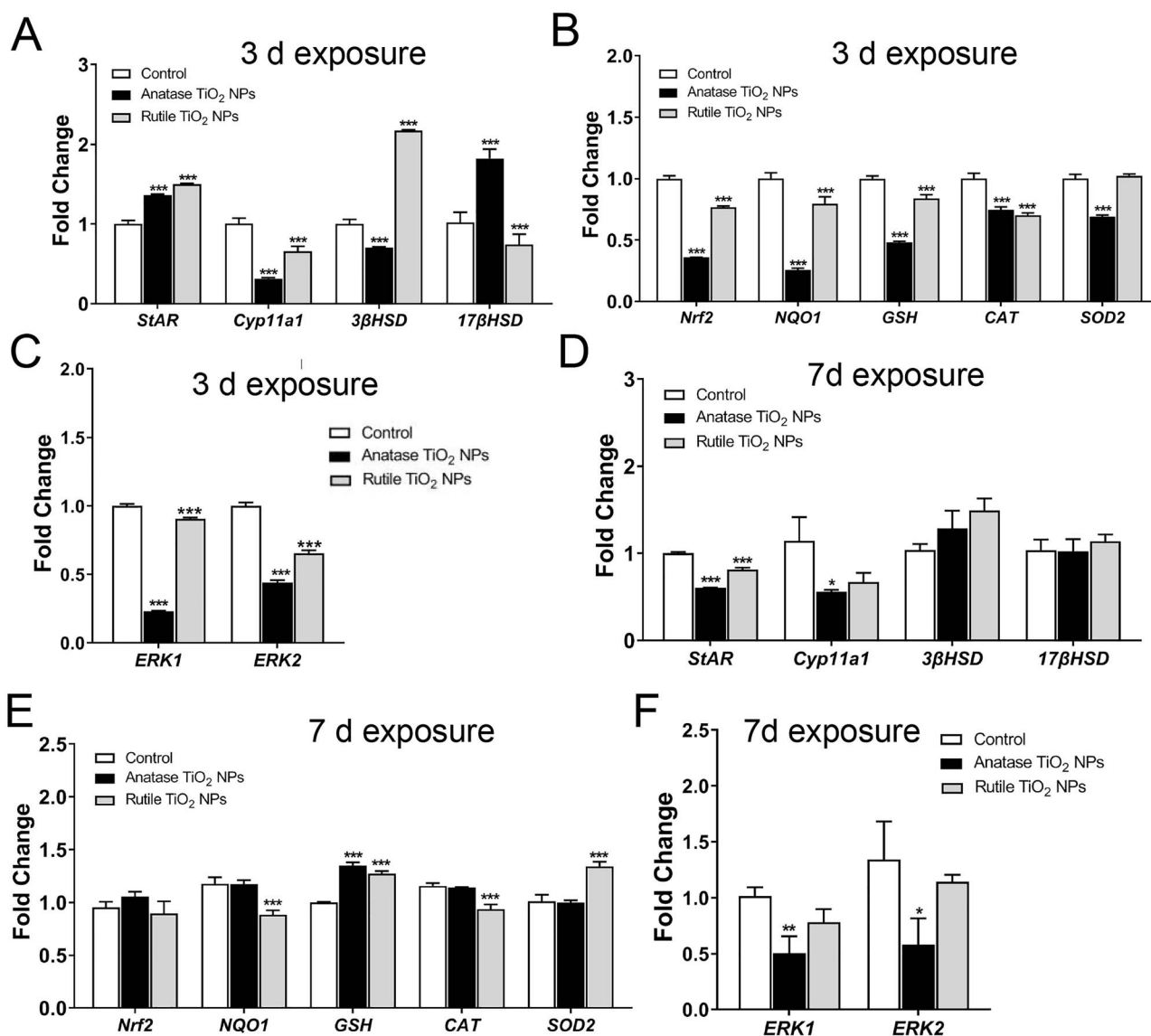


Figure 5: Gene expression of testosterone synthesis, ROS, and MAPK (ERK1/2) in testis. (A-C) 3-day exposure, (D-F) 7-day exposure. Data are expressed as mean \pm SD. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$ compared with control.

Table 1: Sequences of the primers used for quantitative real-time PCR

Target gene	Forward primer (5'-3')	Reverse primer (3'-5')
StAR	TCCTCGCTACGTTCAAGCTG	CGTCGAACTTGACCCATCCA
Cyp11a1	TAGCTTTGCCATGGGTGCGAG	AGTACCGGAAGTGGGTGGTA
3βHSD	ACACGGCTTCTGTCATGGATT	CCAATAGGTTCTGGGTACCTTTC
17βHSD	TGCTTGGGTTTGGCACATT	TCTCTCCAGGCACTGACGTA
ERK1	AATGGAAGGGCTATGACCG	AGCTTGAGAGGGAGAGGGTT
ERK2	ATGACCCAAGTGATGAGCCC	GAGCCCTTGCTGACCAATTT
Nrf2	AGACAAACATTC AAGCCGAT	CTCTCCTGCGTATATCTCGAA
NQO1	TTGCTTTCAGTTTTCGCCTT	CCCTAATCTGACCTCGTTC
GSH	ATCCCACTGCGCTCATGACC	AGCCAGCCATCACCAAGCC
CAT	ATAGCCAGAAGAGAAACCCACA	CCTCTCCATTCGCATTAACCAG
SOD2	ACTTGAAACGTGTAAC TAGGC	CTTTCATACAATACACAGTCCG
GAPDH	TCCTCAAGATTGTGACGAA	AGATCCACAACGGATACATT

production-associated gene of StAR was significantly downregulated in the experimental groups. Cyp11a1 was significantly

downregulated in the anatase groups, while 3βHSD and 17βHSD showed no significant alteration (Fig. 5).

IHC analysis and ROS index determination

The results of IHC analysis of StAR and pERK1/2 in testes (Fig. 6A and B) showed that after 7 days of exposure to TiO₂ NPs, the experimental group decreased significantly, and anatase was more obvious than rutile. In addition, the MDA content in serum of the experimental groups increased significantly, and the anatase group increased more significantly than the rutile group (Fig. 6C). MDA is one of the most important products of membrane lipid peroxide. SOD activity decreased significantly in the anatase group; a downward trend was observed in the rutile group compared with that in the control group, but the difference was insignificant (Fig. 6D). Moreover, there was no significant difference in CAT activity among the groups, but it was observed that there was a decreasing trend in the experimental group, especially in the anatase group (Fig. 6E). These results suggested

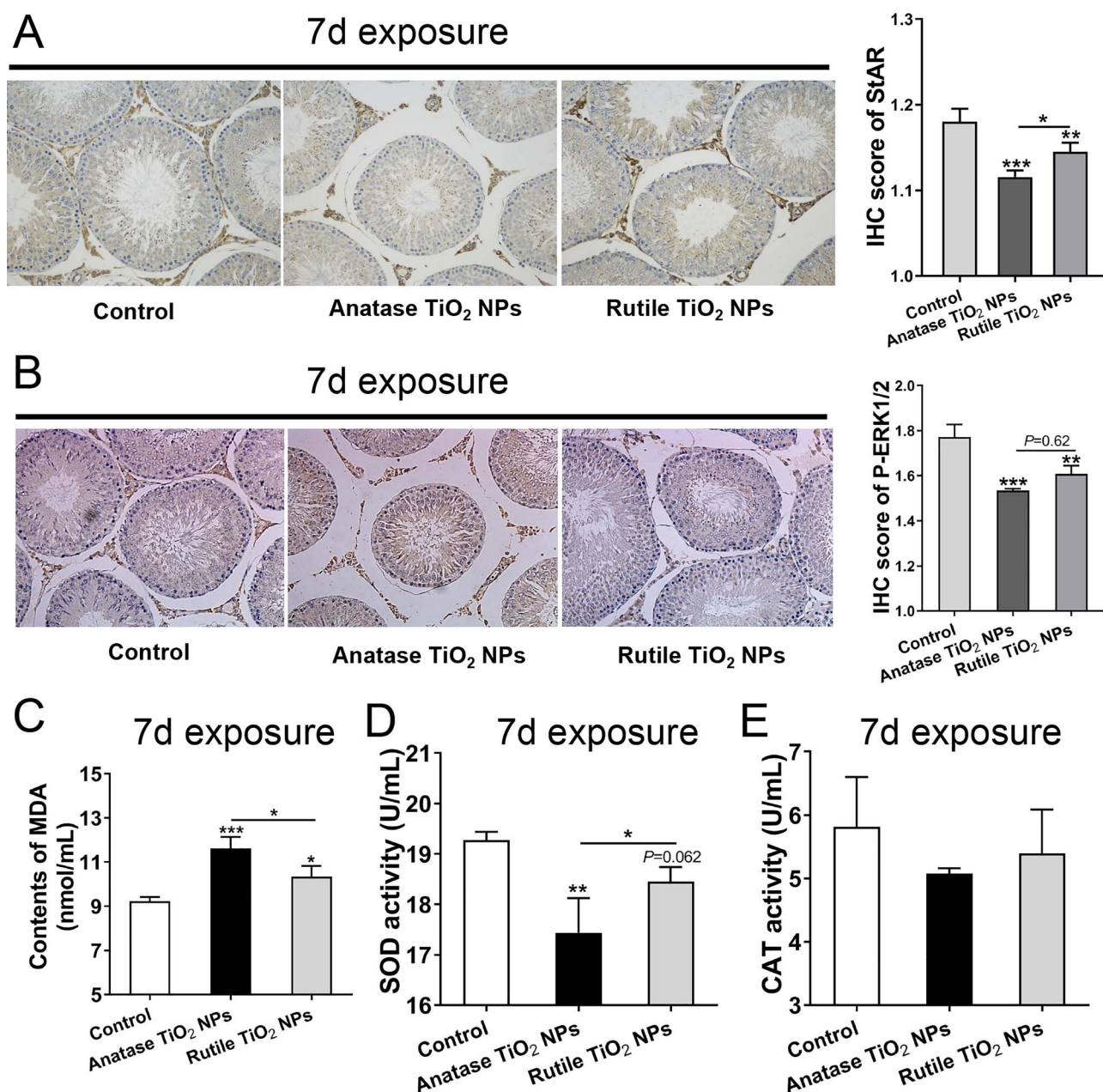


Figure 6: IHC analysis and ROS index determination. (A, B) IHC analysis of StAR and pERK1/2. (C-E) ROS index determination of MDA, SOD, and CAT in serum. Data are expressed as mean \pm SD. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$ compared with control.

that oral TiO₂ NPs for 7 days could induce ROS production and inhibit ERK1/2 phosphorylation and StAR expression.

Discussion

TiO₂ NPs inevitably enter the environment because they are widely used in different fields and enter the human body through different pathways, including oral administration, skin contact, and injections. Studies have shown that oral TiO₂ NPs are toxic to rodents and accumulate significantly over time due to slow tissue elimination and repeated exposure, notwithstanding low oral bioavailability [13]. Complementally, Hu et al. believes that it is easier to detect only higher doses of TiO₂ NPs with higher toxicity [38]. Male infertility is a global population health concern,

and global rates of male infertility range from 2.5 to 12% [39]. Thus, studies on the reproductive toxicity caused by TiO₂ NPs are crucial. Results of this study clearly proved that oral exposure to anatase and rutile TiO₂ NPs (500 mg/kg) for 7 days decreased the content of testosterone in serum compared with the control group (Fig. 3). This finding was consistent with the results of Jia et al. [19], who reported that TiO₂ NPs can inhibit the synthesis of testosterone. The main organ of testosterone synthesis is the testis. The results of our histopathological sections showed that the normal structure of the testis was destroyed, such as seminal cavity disorder and shedding and vacuolation of germ cells. Moreover, a large number of LCs disappeared (Fig. 3). Similar results were reported by Elnagar et al. [40]. LCs are the main sites of testosterone synthesis and secretion; they are mainly

distributed in the loose connective tissue of seminiferous tubules [41, 42] and are responsible for the production of androgens to maintain normal male development and reproductive function [43, 44]. In addition, testosterone is the main component of androgen, which is transported to the target organs of the body and makes an important contribution to reproductive function by binding to receptors [45, 46]. Previous studies reported that TiO₂ NPs directly trigger androgen imbalance, which results in estrogen imbalance, testicular dysfunction, and inhibition of spermatogenesis [17]. In this study, the sperm count also showed similar results; the total number of sperm in the experimental group decreased considerably compared with that in the control group (Fig. 2). In general, TiO₂ NPs can cause testicular damage and affect testosterone levels. A decrease in testosterone levels can affect the testes and exacerbate abnormal testicular spermatogenesis. In other words, the reproductive dysfunction and decrease in serum testosterone level in male SD rats are closely related to TiO₂ NPs.

The synthesis and secretion of testosterone are regulated by the expression of StAR, Cyp11a1, 3 β HSD, 17 β HSD, and other genes [47]. In short, cholesterol is transported from the outer membrane of the mitochondria to the inner membrane of the mitochondria through StAR and is bioconverted to pregnenolone by Cyp11a1, which is then catalyzed by 3 β HSD and 17 β HSD in the endoplasmic reticulum to produce testosterone [48]. StAR is considered a rate-limiting enzyme in the transport of cholesterol to the mitochondria, where its expression is regulated by ERK1/2, and testosterone synthesis is directly affected by it [21, 49]. In the present study, the results showed that anatase TiO₂ NPs significantly downregulated the expression of the ERK1/2, StAR, and Cyp11a1 genes in the 7-day exposure group (Fig. 4E and F). Similar results were obtained by Jia et al. [19] through oral administration of anatase TiO₂ NPs (10, 50, or 250 mg/kg) to mice. Interestingly, StAR was significantly upregulated (Fig. 4A) in the 3-day exposure group, which possibly contributed to the negative feedback regulation of the hormone synthesis pathway in short-term exposure.

ROS produced in the body has been proven to activate members of the MAPK family, which contains ERK1/2 [50]. RT-qPCR results showed that ROS-related genes in the testis changed significantly after exposure to TiO₂ NPs. For example, after 3 days of exposure, the expression of the GSH, CAT, and SOD2 antioxidant genes was significantly downregulated (Fig. 5B). Meena et al. [51] injected 50 mg/kg TiO₂ NPs (21 nm) intravenously in male Wistar rats; their results showed that antioxidant enzymes such as CAT, GSH, and SOD decreased significantly in the testis. In addition, the *Nrf2* and *NQO1* genes related to the antioxidant pathway were significantly downregulated. TiO₂ NPs induced ROS production and damaged the antioxidant capacity in the body. The MDA content in serum increased while SOD activity decreased after 7 days of exposure (Fig. 6C and D). TiO₂ NPs destroy the oxidative balance in the testes of SD rats, which may be the source of the inhibition of testosterone synthesis.

Thus, the small size of TiO₂ NPs could reach the testis through the BTB and induce ROS production. ROS may directly lead to apoptosis, inhibit the expression of the ERK1/2 gene, and reduce the phosphorylation level of ERK1/2, interfering with the normal transcription of the StAR gene. Finally, the synthesis of testosterone was inhibited (Fig. 7). Frankly, this study chose a high dose of TiO₂ NPs acute exposure to rats, which hardly reflect the exposure that animals will face in real scenarios. In that sense, it is meaningful to mention that these investigation provide experimental evidence for the potential risks of nanomaterials in daily use. However, it is necessary to study more realistic

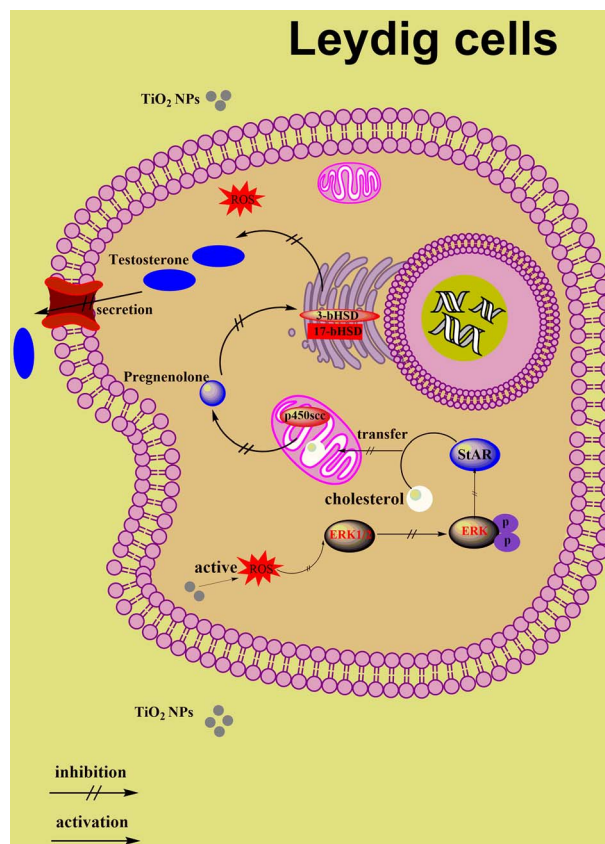


Figure 7: Schematic diagram for the suppression of testosterone production by TiO₂-NPs associated with ROS-MAPK(ERK1/2)-StAR signaling pathway. Acute oral exposure to high doses of TiO₂ NPs in male rats would lead to the production of ROS and further activation of MAPK (ERK1/2) by ROS. ERK and StAR regulate each other, and interference with the expression of ERK1/2 will lead to the inability of StAR to transport cholesterol to the mitochondria normally, resulting in the inhibition of testosterone synthesis in the testis. ("↑" activation, "+" inactivation, "P" phosphorylation)

situations in future research, namely subchronic and chronic exposures, as well as low doses similar to what we could be exposed to with commercial nanotechnology-based products.

Conclusion

The potential mechanism underlying oral exposure to different crystal forms of TiO₂ NPs for 3 and 7 days on testosterone synthesis in male SD rats was explored. The results showed that exposure to TiO₂ NPs was harmful to sperm production and inhibited the synthesis of testosterone. The destruction of testicular tissue was manifested by the disappearance of LCs and exfoliation and vacuolation of spermatogenic cells. In general, the results after exposure to anatase for 7 days were more significant than those after exposure to rutile. Thus, anatase led to greater toxicity than rutile, which may be attributed to the high surface activity of anatase [10]. RT-qPCR results showed that TiO₂ NPs inhibited the expression of antioxidant enzyme genes and induced ROS production in rats, which activated MAPK (ERK1/2). The expression of ERK1/2 and StAR was downregulated, resulting in the inhibition of testosterone synthesis. ROS-MAPK (ERK1/2)-StAR is a potential pathway in which testosterone synthesis is inhibited. Moreover, the synthesis and secretion of testosterone

are complex processes, and the compound dynamic route of TiO₂ NPs inhibiting testosterone synthesis needs to be further studied.

Funding

This work was supported by National Natural Science Foundation of China (81771658 and 81560537).

Conflict of interest statement

The author reports no conflicts of interest in this work.

References

- Zhang J-R. Research on application of nanomaterials in food packaging design. *Adv Mater Sci Technol* 2019;1:1–6.
- Peters RJ, Bouwmeester H, Gottardo S et al. Nanomaterials for products and application in agriculture, feed and food. *Trends Food Sci Technol* 2016;54:155–64.
- Çeşmeli S, Biray Avci C. Application of titanium dioxide (TiO₂) nanoparticles in cancer therapies. *J Drug Target* 2019;27:762–6.
- Rodríguez-González V, Terashima C, Fujishima A. Applications of photocatalytic titanium dioxide-based nanomaterials in sustainable agriculture. *J Photochem Photobiol C* 2019;40:49–67.
- Zhu X, K Pathakoti, Hwang H. M. Green synthesis of titanium dioxide and zinc oxide nanoparticles and their usage for antimicrobial applications and environmental remediation. In: Shukla AK, Irvani S (eds.), *Green Synthesis, Characterization and Applications of Nanoparticles*. E-Publishing Inc, 2019, 223–263.
- Song T, Li R, Li N, Gao Y. Research progress on the application of nanometer TiO₂ photoelectrocatalysis technology in wastewater treatment. *Sci Adv Mater* 2019;11:158–65.
- Weir A, Westerhoff P, Fabricius L et al. Titanium dioxide nanoparticles in food and personal care products. *Environ Sci Technol* 2012;46:2242–50.
- Abdulla IT. Histological effects of titanium dioxide nanoparticles size 10 nm in mice testes. *Sci J Univ Zakho* 2017;5:158–61.
- Uboldi C, Urbán P, Gilliland D et al. Role of the crystalline form of titanium dioxide nanoparticles: rutile, and not anatase, induces toxic effects in Balb/3T3 mouse fibroblasts. *Toxicol in Vitro* 2016;31:137–45.
- de Matteis V, Cascione M, Brunetti V et al. Toxicity assessment of anatase and rutile titanium dioxide nanoparticles: the role of degradation in different pH conditions and light exposure. *Toxicol in Vitro* 2016;37:201–10.
- Zhang X, Li W, Yang Z. Toxicology of nanosized titanium dioxide: an update. *Arch Toxicol* 2015;89:2207–17.
- Shakeel M, Jabeen F, Shabbir S et al. Toxicity of nano-titanium dioxide (TiO₂-NP) through various routes of exposure: a review. *Biol Trace Elem Res* 2016;172:1–36.
- Heringa MB, Geraets L, van Eijkeren JCH et al. Risk assessment of titanium dioxide nanoparticles via oral exposure, including toxicokinetic considerations. *Nanotoxicology* 2016;10:1515–25.
- Hong F, Wang L. Nanosized titanium dioxide-induced premature ovarian failure is associated with abnormalities in serum parameters in female mice. *Int J Nanomedicine* 2018;13:2543.
- Liu H, Ma L, Zhao J et al. Biochemical toxicity of nano-anatase TiO₂ particles in mice. *Biol Trace Elem Res* 2009;129:170–80.
- Hong F, Wang Y, Zhou Y et al. Exposure to TiO₂ nanoparticles induces immunological dysfunction in mouse testis. *J Agric Food Chem* 2016;64:346–55.
- Sharafutdinova L, Fedorova AM, Bashkatov SA et al. Structural and functional analysis of the spermatogenic epithelium in rats exposed to titanium dioxide nanoparticles. *Bull Exp Biol Med* 2018;166:279–82.
- Gao G, Ze Y, Zhao X et al. Titanium dioxide nanoparticle-induced testicular damage, spermatogenesis suppression, and gene expression alterations in male mice. *J Hazard Mater* 2013;258-259:133–43.
- Jia F, Sun Z, Yan X et al. Effect of pubertal nano-TiO₂ exposure on testosterone synthesis and spermatogenesis in mice. *Arch Toxicol* 2014;88:781–8.
- Slaunwhite WR Jr, Burgett MJ. In vitro testosterone synthesis by rat testicular tissue. *Steroids* 1965;6:721–35.
- Stocco DM. StAR protein and the regulation of steroid hormone biosynthesis. *Annu Rev Physiol* 2001;63:193–213.
- Hoberman JM, Yesalis CE. The history of synthetic testosterone. *Sci Am* 1995;272:76–81.
- Payne AH, Youngblood GL, Sha L et al. Hormonal regulation of steroidogenic enzyme gene expression in Leydig cells. *J Steroid Biochem Mol Biol* 1992;43:895–906.
- Leers-Sucheta S, Stocco DM, Azhar S. Down-regulation of steroidogenic acute regulatory (StAR) protein in rat Leydig cells: implications for regulation of testosterone production during aging. *Mech Ageing Dev* 1999;107:197–203.
- Luo D-Y, Yang G, Liu JJ et al. Effects of varicocele on testosterone, apoptosis and expression of StAR mRNA in rat Leydig cells. *Asian J Androl* 2011;13:287–91.
- Martinat N, Crépieux P, Reiter E, Guillou F. Extracellular signal-regulated kinases (ERK) 1, 2 are required for luteinizing hormone (LH)-induced steroidogenesis in primary Leydig cells and control steroidogenic acute regulatory (StAR) expression. *Reprod Nutr Dev* 2005;45:101–8.
- Poderoso C, Converso DP, Maloberti P et al. A mitochondrial kinase complex is essential to mediate an ERK1/2-dependent phosphorylation of a key regulatory protein in steroid biosynthesis. *PLoS One* 2008;3:e1443.
- Seger R, Krebs EG. The MAPK signaling cascade. *FASEB J* 1995;9:726–35.
- Seger R, Hanoch T, Rosenberg R et al. The ERK signaling cascade inhibits gonadotropin-stimulated steroidogenesis. *J Biol Chem* 2001;276:13957–64.
- Matzkin ME, Yamashita S, Ascoli M. The ERK1/2 pathway regulates testosterone synthesis by coordinately regulating the expression of steroidogenic genes in Leydig cells. *Mol Cell Endocrinol* 2013;370:130–7.
- Son Y, Cheong YK, Kim NH et al. Mitogen-activated protein kinases and reactive oxygen species: how can ROS activate MAPK pathways? *J Signal Transduct* 2011;2011:1–6.
- Wang C, Li P, Xuan J et al. Cholesterol enhances colorectal cancer progression via ROS elevation and MAPK signaling pathway activation. *Cell Physiol Biochem* 2017;42:729–42.
- Proquin H, Rodríguez-Ibarra C, Moonen CGJ et al. Titanium dioxide food additive (E171) induces ROS formation and genotoxicity: contribution of micro and nano-sized fractions. *Mutagenesis* 2017;32:139–49.
- Shukla RK, Kumar A, Gurbani D et al. TiO₂ nanoparticles induce oxidative DNA damage and apoptosis in human liver cells. *Nanotoxicology* 2013;7:48–60.
- Castillo AF, Orlando U, Helfenberger KE et al. The role of mitochondrial fusion and StAR phosphorylation in the

- regulation of StAR activity and steroidogenesis. *Mol Cell Endocrinol* 2015;**408**:73–9.
36. Ali SA, Rizk MZ, Hamed MA et al. Assessment of titanium dioxide nanoparticles toxicity via oral exposure in mice: effect of dose and particle size. *Biomarkers* 2019;**24**: 492–8.
 37. Kisin ER, Yanamala N, Farcas MT et al. Abnormalities in the male reproductive system after exposure to diesel and biodiesel blend. *Environ Mol Mutagen* 2015;**56**:265–76.
 38. Hu H, Guo Q, Wang C et al. Titanium dioxide nanoparticles increase plasma glucose via reactive oxygen species-induced insulin resistance in mice. *J Appl Toxicol* 2015;**35**:1122–32.
 39. Agarwal A, Mulgund A, Hamada A, Chyatte MR. A unique view on male infertility around the globe. *Reprod Biol Endocrinol* 2015;**13**:37.
 40. Elnagar AMB, Ibrahim A, Soliman AM. Histopathological effects of titanium dioxide nanoparticles and the possible protective role of N-acetylcysteine on the testes of male albino rats. *Int J Fertil Steril* 2018;**12**:249.
 41. Kordić M, Tomić D, Soldo D et al. Reinke's crystals in perivascular and peritubular Leydig cells of men with non-obstructive and obstructive azoospermia: a retrospective case-control study. *Croat Med J* 2019;**60**:158–65.
 42. Ilacqua A, Francomano D, Aversa A. The physiology of the testis. In: Belfiore A, Leroith D (eds.), *Principles of Endocrinology and Hormone Action*. Italy: S-Publishing Inc, 2017, 1–38.
 43. Payne AH, Youngblood GL. Regulation of expression of steroidogenic enzymes in Leydig cells. *Biol Reprod* 1995;**52**: 217–25.
 44. Sharpe R, Maddocks S, Kerr J. Cell-cell interactions in the control of spermatogenesis as studied using Leydig cell destruction and testosterone replacement. *Am J Anat* 1990;**188**:3–20.
 45. Dohle G, Smit M, Weber R. Androgens and male fertility. *World J Urol* 2003;**21**:341–5.
 46. Elias M. Serum cortisol, testosterone, and testosterone-binding globulin responses to competitive fighting in human males. *Aggress Behav* 1981;**7**:215–24.
 47. Zhang L, Cui S. Effects of daidzein on testosterone synthesis and secretion in cultured mouse Leydig cells. *Asian Australas J Anim Sci* 2009;**22**:618–25.
 48. Hall PF, Osawa S, Mrotek J. The influence of calmodulin on steroid synthesis in Leydig cells from rat testis. *Endocrinology* 1981;**109**:1677–82.
 49. Wang H, Wang Q, Zhao XF et al. Cypermethrin exposure during puberty disrupts testosterone synthesis via downregulating StAR in mouse testes. *Arch Toxicol* 2010;**84**:53–61.
 50. Han A, Zou L, Gan X et al. ROS generation and MAPKs activation contribute to the Ni-induced testosterone synthesis disturbance in rat Leydig cells. *Toxicol Lett* 2018;**290**:36–45.
 51. Meena R, Kajal K, Paulraj R. Cytotoxic and genotoxic effects of titanium dioxide nanoparticles in testicular cells of male Wistar rat. *Appl Biochem Biotechnol* 2015;**175**:825–40.