#### **REVIEW ARTICLE**



# Neutrophils: fast and furious—the nucleotide pathway

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#### **Abstract**

Nucleotide signaling is a key element of the neutrophil activation pathway. Neutrophil recruitment and migration to injured tissues is guided by purinergic receptor sensitization, mostly induced by extracellular adenosine triphosphate (ATP) and its hydrolysis product, adenosine (ADO), which is primarily produced by the CD39-CD73 axis located at the neutrophil cell surface. In inflammation unrelated to cancer, neutrophil activation via purinergic signaling aims to eliminate antigens and promote an immune response with minimal damage to healthy tissues; however, an antagonistic response may be expected in tumors. Indeed, alterations in purinergic signaling favor the accumulation of extracellular ATP and ADO in the microenvironment of solid tumors, which promote tumor progression by inducing cell proliferation, angiogenesis, and escape from immune surveillance. Since neutrophils and their N1/N2 polarization spectrum are being considered new components of cancer-related inflammation, the participation of purinergic signaling in pro-tumor activities of neutrophils should also be considered. However, there is a lack of studies investigating purinergic signaling in human neutrophil polarization and in tumor-associated neutrophils. In this review, we discussed the human neutrophil response elicited by nucleotides in inflammation and extrapolated its behavior in the context of cancer. Understanding these mechanisms in cancerous conditions may help to identify new biological targets and therapeutic strategies, particularly regarding tumors that are refractory to traditional chemo- and immunotherapy.

**Keywords** Human neutrophils · Purinergic activation · Neutrophil migration · Purinergic signaling · Neutrophil modulation · Activation spectrum · N1/N2 profile

### Highlights

- 1. Activation and upregulation of the purinergic system could favor protumor neutrophil activity.
- 2. Purinergic receptors P2Y<sub>2</sub>, A2a, and A3 guide neutrophil migration through an ATP concentration gradient, TLR4 stimulation, or IL-8 secretion
- 3. Neutrophil migration to injured sites is impaired by the decrease in extracellular adenosine levels mediated by CD73 inhibition.
- 4. Extracellular adenosine plays a key role in NET production via A1 and A3 receptor sensitization.
- $5. P2Y_6$  signaling upregulates the Bcl-xl-mediated anti-apoptotic pathway and inhibits neutrophil apoptosis.
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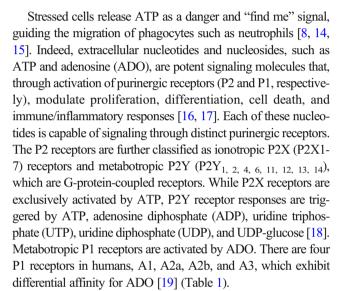


## Introduction

Normal tissues are composed of different types of cellular, molecular, and microenvironmental signals that work together to ensure homeostasis and proper tissue functioning [1]. In nonphysiological conditions such as infection, tissue damage, or inflammatory processes, the initiation, triggering, or recruitment of innate immune cells and plasma proteins occurs at the sensitized site [2]. Although tissues are resistant to many disorders, the tumorigenesis process is capable of disrupting homeostasis to the point of no possible restoration [1].

The origins of solid cancer are not completely understood; however, its functional relationship with inflammation has been widely discussed [3, 4]. Chronic inflammation contributes to tumor stabilization based on the release of cytokines into the microenvironment. Tumor growth is sustained by the presence of immune cells, growth and angiogenesis factors, and DNA damage-promoting agents [4-6]. Cell proliferation caused by tissue regeneration after injury increases until tissue repair [7]. In contrast, cells continue to grow and develop in a chronic inflammatory microenvironment, establishing irreparable lesions. The power of inflammatory cells in tumor progression is undeniable as they promote neoplastic processes and provide an attractive tumor microenvironment (TME) [4–7]. The immune system is a regulated and integrated cellular network that preserves and restores homeostasis, and purinergic signaling helps to adjust the functions of immune cells [8].

Neutrophils, which are part of the polymorphonuclear (PMN) leukocyte family, have a major role during the early stages of the inflammatory response. They are the first leukocytes recruited to the injured site within a few hours of the damage. In addition, pathogens are eliminated through a variety of mechanisms such as degranulation, necrosis, and phagocytosis [9–11]. Specific chemokines and exogenous ligands are common mechanisms of neutrophil recruitment to injured sites [11, 12]. However, promoters of early migration of PMN cells to distant sites of metastasis in the absence of detectable inflammation are not yet defined [13]. In this regard, neutrophils can be associated with a quick response to any disturbance.



Due to its proinflammatory actions, extracellular ATP is considered to be a damage-associated molecular pattern (DAMP) [14, 15, 33]. Conversely, extracellular ADO, which is mainly generated by the hydrolysis of ATP by ectonucleotidases, triggers immunosuppressive and immunomodulatory responses [19, 34]. There are two major ectonucleotidases responsible for the control of ATP and ADO levels in the bloodstream and at the surface of leukocytes, the ecto-nucleoside triphosphate diphosphohydrolase-1 (NTPDase1/CD39) and the ecto-5'-nucleotidase (CD73). The CD39-CD73 axis is also present on the surface of tumor cells. Together, these two enzymes convert extracellular ATP to ADO in a sequential manner [35] (Fig. 1).

The neutrophil activation spectrum, classified as antitumor (N1) and pro-tumor (N2) neutrophil phenotypes, is similar to that proposed for macrophage polarization. However, these studies are preliminary, and there are no distinctive markers of maturation, activation, or polarization states of neutrophils; the effector mechanisms that modulate the leukocyte functional behavior and its role in disease perpetuation are not completely understood as well [6, 7, 12].

In line with the complexity of neutrophil polarization in solid tumors, some studies have shown that tumor-



 Table 1
 Purinergic receptors involved in neutrophil physiology

A1 High ADO affinity (EC <sub>50</sub> 0.2-0.5 μM)	Inhibition of cAMP formation by Gi/o-coupled protein	↑ Adhesion	[19, 20]
		↑ Chemotaxis	
		↓ Neutrophils extravasation	
		↑ A <sub>2</sub> upregulation	
A2a Low ADO affinity (EC $_{50}$ 0.6-0.9 $\mu M$ )	Promotion of cAMP formation by Gs-coupled protein	↓ Chemotaxis	[20, 21]
		↓ Adhesion	
		↓ ROS production	
		↓ Degranulation	
A2b	Promotion of cAMP production by Gs-coupled protein		[22]
		↓ Transendothelial migration	
		↓ Oxidative burst	
		↓ NET formation	
A3 High ADO affinity (EC $_{50}$ 0.2-0.5 $\mu M$ )	Inhibition of cAMP production and stimulation of IP3 production by Gi/o and Gq-coupled proteins	↓ Migration	[23, 24]
		↑ Chemotaxis	
		Regulation of directional movement	
P2Y <sub>2</sub> ATP	Gq protein increases cytosolic Ca <sup>2+</sup> through interaction with the actin cytoskeleton	↑ Chemotaxis	[23–26]
		↑ Orientation in chemoattractant gradients	
		,	
P2Y <sub>6</sub> UDP	Ga protein		
	stimulation causes PLCβ activation, Ca <sup>2+</sup> mobilization, and IP3 formation		
		* 6 3	
P2Y <sub>11</sub> ATP and NAD <sup>+</sup>	Promotion of cAMP production by Gs protein	E	[29, 30]
		<b>↓</b>	
P2X1 ATP		Chemotaxis in response to LPS-induced	uced [8, 31]
	Na <sup>+</sup> , K <sup>+</sup> , and Ca <sup>2+</sup>		
P2X7 ATP	Ion channels permeable for Na <sup>+</sup> , K <sup>+</sup> , and Ca <sup>2+</sup>		[32]
	Low ADO affinity (EC <sub>50</sub> 0.6-0.9 μM)  Very low ADO affinity (EC <sub>50</sub> 16-64 μM)  High ADO affinity (EC <sub>50</sub> 0.2-0.5 μM)  ATP	Low ADO affinity $(EC_{50} \ 0.6\text{-}0.9 \ \mu\text{M})$ Promotion of cAMP formation by Gs-coupled protein  Promotion of cAMP production by Gs-coupled protein  Promotion of cAMP production by Gs-coupled protein  Inhibition of cAMP production and stimulation of IP3 production by Gi/o and Gq-coupled proteins  ATP  Gq protein increases cytosolic $Ca^{2+}$ through interaction with the actin cytoskeleton  UDP  Gq protein stimulation causes PLC $\beta$ activation, $Ca^{2+}$ mobilization, and IP3 formation  ATP and NAD+  Promotion of cAMP production by Gs protein  Ion channels permeable for Na+, K+, and Ca^2+  Ion channels permeable for	

Abbreviations: A2: α-2 adrenergic G-protein-coupled receptor; ADO: adenosine; ADP: Adenosine diphosphate; ATP: Adenosine triphosphate; cAMP: cyclic adenosine monophosphate; HNP1: human neutrophil peptide; IL-1β: *interleukin 1* beta; IL-8: *interleukin;* IP3: inositol triphosphate; LPS: lipopolysaccharide; MSU: monosodium urate crystals; NAD<sup>+</sup>: nicotinamide adenine dinucleotide oxidized; NET: neutrophil extracellular traps; NLRP3: NOD-, LRR- and pyrin domain-containing protein 3; PLCβ: phospholipase C; ROS: reactive oxygen species; UDP: uridine diphosphate

associated neutrophils (TANs) exhibit mixed characteristics of N1/N2 polarized cells [36]. Additionally, recent evidence points to arginase-1 and LOX-1 to be hallmarks of PMN myeloid-derived suppressor cells (PMN-MDSCs) [37]. MDSCs are a heterogeneous group of immature myeloid cells related to either the neutrophil (PMN-MDSCs) or monocyte (M-MDSC) differentiation pathways, which promote tumor growth by suppressing immune surveillance [38].

PMN-MDSCs, also recently proposed by some authors as neutrophils with proven immunosuppressive activity or, alternatively, as pathologically activated neutrophils [38, 39], inhibit T cell function, myeloid, and natural killer (NK) cells; enhance angiogenesis through the production of metalloproteinase-9 (MMP-9), prokineticin 2, and vascular endothelial growth factor (VEGF); and promote tumor metastasis [39]. The effects of extracellular purines on immunosuppressor cells have raised interest. For example,

MDSCs overexpress P2X7 receptor that upon ATP binding induces arginase-1, reactive oxygen species (ROS), and transforming growth factor  $\beta$  (TGF- $\beta$ ) release, resulting in unexpected ATP immunosuppressive activity [40]. Additionally, ADO promotes the expansion of the MDSC population by engaging A2b receptors that are expressed on myeloid precursor cells [41]. It is important to note that the discovery of LOX-1 and arginase-1 as hallmarks of PMN-MDSCs may facilitate the understanding of immunosuppressive mechanisms of neutrophils in TME as well as its tumor-promoting role [37].

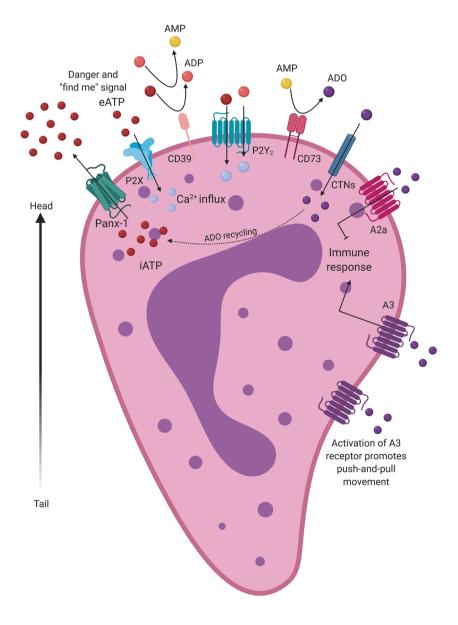
Neutrophil polarization to a pro-tumor phenotype may also be dependent on purinergic signaling. The increase in neutrophil activation and oxidative burst also depends on the autocrine mechanism previously described [23, 24, 42]. In addition, the extracellular ATP levels of non-self-source, such as the microenvironment or the injured site, could result in P2X



and P2Y stimulation in a paracrine manner [8, 43]. Interestingly, the P2X7 expression and function also directly impacts on ATP content in the TME, which further determine the behavior of tumor-infiltrated immune cells [44]. Under desensitization conditions, such as prolonged stimulation with high ATP levels present in the TME, P2Y<sub>2</sub> receptors expressed by neutrophils are useful for sustaining the signaling, due to the lower response, retaining cellular viability, which has already been demonstrated in macrophages [45]. Purinergic signaling is widely studied because it is found on the surface of most cells and is a fundamental component of the immune/inflammatory response. Recent studies explored the role of purinergic signaling in cancer-infiltrating immune cells, including macrophages, CD4+/CD8+ lymphocytes, NK, and MDSC cells. These investigations point that the

modulation of purinergic signaling in tumor-associated immune cells supports proliferation, chemotaxis, and cytokine release [44, 46–49]. Although the well-known participation of purinergic signaling in neutrophil function regulation, little was investigated about how this pathway affects the neutrophil behavior in the TME. Here, we have reviewed which purinergic receptors contribute to neutrophil functions, in light of their diversity and plasticity. The discussion is focused on studies performed on human neutrophils, in view of the high heterogeneity in neutrophils, including immature, mature, aged neutrophils, PMN-MDSCs, and the lack of specific markers to define these subsets. Purinergic signaling in neutrophils under both acute and chronic inflammatory diseases has been explored and further extrapolated in the context of cancer-related inflammation.

Fig. 1 Neutrophil migration through purinergic pathway activation. Pannexin-1 (PNX-1) releases ATP (red balls), a danger, and "find me" signal. The increase in extracellular ATP potentiates neutrophil migration. ATP is hydrolyzed to ADP (pink balls) and ADP to AMP (yellow balls) by CD39. CD73 hydrolyzes AMP to adenosine (ADO) (purple balls). ATP recognizes P2X receptors and ATP/ADP/UTP/UDP binds P2Y2 receptors in neutrophils, inducing cell activation via intracellular Ca<sup>2+</sup> release. Besides, extracellular adenosine binds to two main receptors: A2a and A3. The responses elicited by ATP and adenosine generate a movement of "push and pull" that regulates neutrophil phenotype and orients it migration





# Driving license: purinergic regulation of neutrophil migration and chemotaxis

Neutrophil migration depends on a frontal excitatory and a back inhibitory signal on the cell surface. ATP release via Pannexin-1 (PNX-1) induces the chemotaxis of neutrophils at the front edge by autocrine stimulation of the P2Y<sub>2</sub> receptor. Subsequently, ADO is recognized by P1 type receptors, localized on the neutrophil tail, such as A2a, which blocks chemoattractant signaling and alternatively binds to A3 receptors, which stimulate immune migration. This purinergic feedback loop promotes neutrophil movement toward the chemoattractant source [8, 23]. The rapid conversion of extracellular ATP into ADO by neutrophils allows the activation of A2a receptors, providing an important counterpoint to the stimulation of P2Y<sub>2</sub> and A3 receptors. Suppressive actions of A2a receptors provide a limiting mechanism for the main functions of neutrophils [50]. In summary, P2Y<sub>2</sub> and A2a receptors mainly provide excitatory and inhibitory responses, respectively, producing a push-pull movement, thereby allowing neutrophil migration [23, 43]. As neutrophils are recruited in response to different stimuli, including bacterial products, complement proteins (C5a), immune complexes, chemokines, and cytokines [10], a phenotypic adaptation to these different microenvironments is inevitable [12].

P2Y receptors have been shown to be major influencers of neutrophil activation. Indeed, the release of IL-8 a major chemokine for neutrophils is regulated by P2 receptors sensitization. In this regard, P2Y<sub>6</sub> induces IL-8 secretion from human monocytes, which in turn controls in vitro neutrophil migration [51]. Moreover, P2 receptor activation, particularly P2Y<sub>2</sub>, is required for IL-8-induced neutrophil chemotaxis [52]. Finally, Kukulski and colleagues demonstrated with a transwell apparatus that P2Y<sub>2</sub> receptor activation is necessary for TLR4-induced in vitro transendothelial neutrophil migration, which was potentiated by UTP, a P2Y<sub>2</sub> agonist. However, in opposite to which was expected, this phenomenon was mostly regulated by Rho kinase pathway, rather than IL-8 release [53]. Therefore, extracellular nucleotides participate of crosstalk among immune and endothelial cells, orchestrating the neutrophil responses.

Interestingly, Gabl and colleagues argued that P2Y<sub>2</sub> down-regulation in neutrophils probably originates from inside by a novel cytoskeleton-dependent mechanism [25]. They demonstrated that receptors occupied by their ligands undergo an agonist-induced conformational change, which elevates intracellular Ca<sup>2+</sup> levels by coupled G-protein signaling. The authors also proposed that the P2Y<sub>2</sub> receptor blockade inhibits the NADPH oxidation signaling pathway [25].

Neutrophil chemoattraction involves chemokines, lipids, anaphylatoxins from the complement system (C5a-C3a), and platelet activation factor (PAF), but IL-8 promotes a more potent binding with CXCR1 and CXCR2. There are also

additional mediators that can work as recruiters of neutrophils [48, 52, 53]. Thus, the purinergic system also plays an important role in activating neutrophil migration. Besides, this pathway contains signaling molecules that modulate differentiation and proliferation and that are able to control inflammatory events, which might be responsible for neutrophil activation.

The release of microenvironment chemokines plays an essential role in tumor progression. Considering that in glioblastoma cells the purinergic signaling is active, a study from our group showed that the spontaneous and lipopolysaccharide (LPS)-mediated IL-8 release by tumor cells is dependent on P2Y<sub>6</sub> and P2X7, which in turn promotes glioma cell proliferation [54] and may induce in vivo neutrophil recruitment. In addition, the neutrophil expression of CD39 may facilitate the molding of the immune response [55]. Indeed, IL-8 production is controlled by the activity of CD39 expressed by human neutrophils. Interestingly, ATP is a potent stimulus for IL-8 release by neutrophils only upon CD39 inhibition, suggesting that at physiological condition neutrophils remain unresponsive to nucleotide stimulation due to its intrinsic CD39 activity [55]. Neutrophil participation in tumor progression has been investigated in preclinical studies of animals. However, few studies have correlated tumors, neutrophils, and purinergic signaling in humans.

The purinergic cascade produces a very important metabolite in neutrophil activation and migration control, the ADO. A study showed that ADO promotes chemotaxis while inhibiting the activation and the consequent release of ROS [56]. Hence, neutrophils migrate to the site of infection without damaging healthy tissues along their path.

CD39 hydrolyzes ATP to AMP in a sequential manner; a second enzyme, CD73, hydrolyzes AMP to ADO. Therefore, these enzymes profoundly influence immune response [35]. The abnormal activity of CD39 and CD73 produces high amounts of ADO and may favor an immunosuppressive environment, which reinforces cancer development by impairing immune surveillance [57]. Maintaining the harmony between inflammatory and anti-inflammatory responses prevents exacerbated immunosuppression or uncontrolled inflammation, as the CD39-CD73 axis can promote the self-tolerance mechanism. The outcome of the activity escalation of these enzymes generates elevated levels of extracellular immunosuppressive ADO [35, 57].

# Survival of the dead: neutrophil modulation through purinergic activation

Neutrophils are commonly believed to remain viable in circulation for approximately 4 days, followed by apoptosis [12, 58]. Understanding the mechanisms that affect the life span of neutrophils may help to identify new therapeutic targets. The following paragraphs highlight the importance of the



neutrophil life span, in view of the fact that nucleotides affect neutrophil apoptosis.

Neutrophils have few mitochondria in their cytosol and therefore produce energy mainly through glycolysis. Thus, mitochondria rarely participate in ATP formation [42, 50]. However, for a higher level of intracellular ATP production, the cell relies on the tricarboxylic acid cycle (TCA), probably by the activation of the mTOR pathway, an important metabolic pathway that regulates biological and physiological processes such as proliferation, growth, cell survival, and autophagy. This allows the flow of Ca<sup>2+</sup> into the neutrophil mitochondria, which is related to the activation of the P2Y<sub>2</sub> receptor, leading to the production of intracellular ATP. The ATP produced is externalized by PNX-1, which impacts the P2Y<sub>2</sub> receptors in an autocrine manner, potentiating neutrophil migration [42].

P2Y<sub>2</sub> overstimulation is unfavorable under different circumstances. An investigation showed that increased levels of systemic ATP in sepsis impair neutrophil functions by disrupting the endogenous purinergic signaling mechanisms that regulate cell activation and chemotaxis mediated by P2Y<sub>2</sub>. The authors proposed that targeting systemic ATP may improve neutrophil function and host defenses, as a new therapeutic strategy for sepsis treatment [43].

Upon activation, TCA significantly increases the production of ATP. In this scenario, the NADH produced is oxidized in the respiratory chain reaction, ATP is synthesized, and NAD+ returns to the cycle. A study observed that increased NAD+ levels are directly related to neutrophil aging, probably because of increased energy demand. Moreover, intracellular ATP levels are not consistent with expectations. It is argued that there may be an increase in ATP synthesis but also an increase in consumption, and therefore the final energy balance is lower [59]. Thus, ATP levels are decreased in aged neutrophils. Considering that intracellular ATP is produced by activating purinergic signaling and the few existing mitochondria, the decrease in ATP may be caused by increased energy demand or decreased production [59].

Another P2Y receptor, P2Y<sub>6</sub>, has been drawing attention for its relationship to neutrophil apoptosis inhibition. The study conducted by Nagaoka and colleagues evaluated the interaction between the P2Y<sub>6</sub> antagonist (MRS2578) and apoptotic behavior [27]. The authors observed that apoptosis was reactivated in the presence of the P2Y<sub>6</sub> antagonist. The P2Y<sub>6</sub> ligand, UDP, induced suppression of programmed cell death when bound to the receptor. MRS2578 also prevented the binding of P2Y<sub>6</sub> to its ligand, allowing neutrophil apoptosis, suggesting that the induction by HNP-1 downregulated pro-apoptotic and upregulated the anti-apoptotic activities by Bcl-xl, which in turn inhibited apoptosis. The mitochondrial membrane potential and caspase-3 activity resulted in decreased pro-apoptotic signals through the P2Y<sub>6</sub> signaling pathway [27].

In cancer, increased survival of TAN has been proposed to play an important role in the development and growth of tumor mass [60]. Therefore, further studies on the life span of TAN in cancer and the purinergic pathway in neutrophil activation and migration, as well as its connection with cellular death, may help in identifying new molecular targets for cancer therapeutics.

# Suicide squad: extracellular nucleotide levels in inflammation and cancer

Injured tissues, whether inflamed or infected, secrete neutrophil recruitment chemokines that signal the attack site to peripheral blood circulating neutrophils. Neutrophils are the first immune cells to reach the damaged tissue. It can be said that these cells are the infantry of our immune system. Upon arrival at the injured site, neutrophils begin the process of receptor-mediated respiratory burst and degranulation, leading to neutrophil apoptosis [61].

An investigation performed by Patel and collaborators [13] observed the chemotactic activity of PMN-MDSCs from cancer patients when compared to that of control neutrophils. The study found that there was less chemotactic activity in PMN-MDSCs, probably due to the lack of extracellular ADO, suggesting that ATP hydrolysis might be slowed down in this situation [13].

Cancer can be characterized as chronic inflammation. ATP levels in cancer are higher than those under physiological conditions, as a result of ATP release from necrotic, stromal, and cancer cells as well as from stress and hypoxia factors [14, 44, 62]. In addition, the mechanism by which ATP is secreted in the extracellular medium is crucial for P2-mediated responses [14]. Extracellular ATP has a dual role in cancer, which includes an antitumor immune response inducing tumor cell death and a pro-tumor response that increases the proliferation and metastasis of cancer cells [63]. Hypoxia is a tumor condition that increases CD39 and CD73 expression, and consequently ADO formation, which is associated with resistance to chemotherapy due to its immunosuppressor effect [34]. Thus, purinergic signaling can modulate cancer progression by activating P2 and P1 receptors expressed by tumors as well as immune-associated cells [40, 41, 54, 63].

During noncancerous inflammation, ATP is released at high concentrations by injured cells as a "danger signal" or DAMP to restore tissue integrity [14, 15, 33, 40]. In this scenario, P2X receptors are upregulated in immune cells including neutrophils, macrophages, and lymphocytes [18]. The P2X7 receptor is particularly involved in inflammation by releasing proinflammatory cytokines such as IL-1 $\beta$  and tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) [32, 64]. In addition, P2X7 promotes PI3K/Akt activation, HIF1 $\alpha$  expression, and VEGF secretion, regulating MYCN oncogene which further implicate in cell proliferation and poor overall survival of patients with neuroblastoma [65, 66]. Regarding P1 receptor, A3



receptor plays an important role in the migration of neutrophils to inflammation sites [8].

The need for extracellular ATP for direct migration as well as its modulation in the release of proteolytic enzymes has been discussed in previous studies [13, 23]. On one hand, neutrophil recruitment is necessary for maintaining homeostasis, and on the other hand, neutrophil enzymes lack specificity for necrotic cells, and this causes damage to the adjacent tissues [3]. This nonspecific behavior may be one of the factors influencing cancer progression.

A feature of ADO that deserves attention is its ability to inhibit proinflammatory mediator production by monocytes and dendritic cells (DCs), such as ROS and TNF- $\alpha$ , in addition to the A2a-mediated immunosuppressive function in T-regulatory cells [47]. Moreover, P2X7 is overexpressed in several malignancies as well as in immune cells, where it participates in growth-promoting activity and contributes to TME composition via regulation of cytokine release, including TGF- $\beta$  and IL-1 $\beta$  [65, 67, 68]. P2X7 antagonism is also related to downregulation of CD39-CD73 axis in CD4<sup>+</sup> T-effector cells and DCs, further decreasing the ADO levels in TME [44]. Taken together, these characteristics may be involved in the maintenance of the TME, considering that extracellular ADO has immunosuppressive action while recruiting more leukocytes.

Neutrophil and monocyte modulation may also be related to purinergic signaling. In the case of neutrophil activation by gram-positive pathogens, in vitro a study showed that the inhibition of CD73 decreased the ability of PMN cells to kill bacteria, suggesting that the ablation of enzymes that generates extracellular ADO impairs both the recruitment and bactericidal activity of PMNs [69]. Therefore, ADO affects neutrophil-killing cellular functions.

Changes in the number of neutrophil extracellular traps (NETs) are related to autoimmunity promotion, the presence of vascular diseases, and thrombosis and contribute to tumor progression and metastasis. A relationship between elevated NET production and poor prognosis in human tumors has been shown in previous studies. Indeed, these NETs are capable of catching circulating tumor cells and favor metastatic implant formation [3, 70, 71].

# Tumor hustle: neutrophils wrap cancer cells

In solid tumors, the presence of DAMPs and cytokines in the TME may induce differential neutrophil responses [3, 7, 38]. Moreover, the lack of specificity of neutrophil enzymes may contribute to cancer progression, which is especially associated with purinergic pathway activation. ADO is a key molecule with an established role as an immunosuppressive agent, regulating immune cell recruitment, modulating neutrophil-killing features, and promoting cancer progression.

N1- and N2-like neutrophils represent extremes of different molecular phenotypes, which depend on the microenvironment [7]. Considering that purinergic signaling has great influence on neutrophil activation and migration, our hypothesis is that the neutrophil activation spectrum is related to the activation and signaling of purinergic receptors.

Few studies have shown the relationship between these phenotypes and the purinergic signaling. It is known that the immune system plays a fundamental role in tumor progression, although all the mechanisms are not yet well elucidated. In vivo and in vitro studies have found that these PMN leukocytes modulate the TME [5, 6]. The antitumor phenotype is characterized by enhanced expression of TNF- $\alpha$ , CCL3, and ICAM-1, and reduced arginase-1 production, inhibition of angiogenesis, and promotion of antitumor response of T lymphocytes [72, 73]. In contrast, it is discussed that the immunosuppressive phenotype is acquired by the presence of TGF-β, favoring the infiltration of neutrophils with high expression of CXCR4, VEGF-A, and MMP-9 [73]. Neutrophils are the major producers of VEGF-A and delivery high levels of MMP-9, which releases the active form of VEGF-A from the extracellular matrix. However, pro-tumor neutrophils are able to discharge MMP-9 even in the absence of a protease inhibitor, and increased levels contribute to angiogenesis and tissue invasion [74]. P2X7 is described as an important angiogenesis and immunossupressive mediator as its sensitization results in VEGF and TGF-β release in TME [44, 66]. Although the expression of P2X7 on human neutrophils is controversial [32, 75], the soluble factors present in the TME as a consequence of P2X7 activity certainly impact the function and phenotype of TANs.

In cancer, the formation of metastasis is linked to NET release, in which circulating tumor cells are trapped by neutrophils, facilitating their deposition at distant sites of metastasis [76–78]. The quantification of NETs in patients diagnosed with cancer remains challenging; however, the presence of NETs in the tumor niche is associated with a worse prognosis [79] and indirectly links with patient survival [80–82].

The participation of purinergic signaling in NETs formation has been investigated in inflammatory conditions, including deficiency of ADA2 and gout [28, 83, 84]. Indeed, ADO contributes to NETs release via A1 and A3 receptor activation expressed on neutrophils [83], while A2a receptor induces the opposite effect [84]. Regarding P2 receptors, a study performed by Sil and colleagues demonstrated that P2Y<sub>6</sub> receptor is essential for regulating neutrophil functions in gout disease. The investigation elucidated that P2Y<sub>6</sub>/store-operated Ca<sup>2+</sup> influx/IL-8 axis participates in MSU crystal-induced NET formation, suggesting P2Y<sub>6</sub> as an interesting target to modulate neutrophil function and activation [28]. Therefore, the antagonism of purinergic receptors may be an alternative to



debilitate the pro-tumor immune response of neutrophils in TME as well as its over-activation in inflammatory diseases.

# **Concluding remarks**

To summarize, as several studies have shown, the purinergic pathway profoundly influences neutrophil features. The driving license of neutrophils has a P2Y<sub>2</sub> stamp, seeing that excitatory and inhibitory responses from this receptor and A2a produce a push-pull movement, allowing neutrophil migration. In fact, IL-8 favors ATP-mediated P2Y<sub>2</sub> sensitization and regulates neutrophil migration. In addition, P2Y<sub>2</sub> activation increases intracellular Ca<sup>2+</sup> levels and blocks NADPH oxidation, inducing a conformational change in the cytoskeleton. Nevertheless, high ATP levels and P2Y<sub>2</sub> overstimulation can disrupt physiological responses [25, 43, 53].

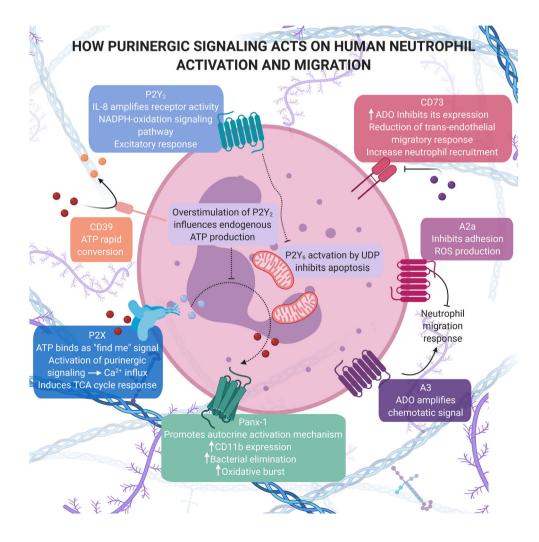
Although neutrophils are not "zombies," their aging process has subtle characteristics. Higher NAD<sup>+</sup> levels and smaller ATP levels are present, due to the increased energy demand in aging cells. Meanwhile, neutrophils can

act like zombies, as P2Y<sub>6</sub> pathway stimulation blocks neutrophil apoptosis via mitochondrial membrane potential, caspase-3 activity, and Bcl-xl upregulation, increasing neutrophil life span [27, 59].

Above all, neutrophils are the immune suicide squad, since these cells are the first line of body defense and their attack mechanism results in neutrophil death, contributing to their short life span. In this case, a small change can impair the immune response and allow tumor progression. As shown in this review, the purinergic system plays a crucial role in neutrophil action. For instance, neutrophil activation can lead to extracellular ATP hydrolysis and consequently chemotaxis regulated by P1 receptors. ADO receptors also modulate migration, adhesion, and ROS release, allowing neutrophils to migrate without shattering other tissues [8, 14, 56, 61] (Fig. 2).

Although preclinical models are a great option for disease research, studies show that there are differences between human and murine immune cells that should be considered [85, 86]. In this regard, it is suggested that at least two different approaches should be used in the in vivo model so that the conclusions reflect the true contribution of neutrophils [87–89]. The worldwide

Fig. 2 Overview of purinergic activation in neutrophils





**CD73** 

that is responsible for converting AMP to

ADO in the purinergic system. It also acts as

a cell-cell and cell-matrix protein, important

for cell communication and migration, by

potentiating EGFR/Akt and VEGF/Akt pathways. In addition, it promotes invasion.

migration, and adhesion of tumor cells.

Hypoxia-inducible factor 1-alpha. It is a

transcriptional regulator of cellular and

the  $\alpha$ -defensin family of antimicrobial

Interleukin-8, a chemokine released by

and other immune cells to the tumor or

infection site. It is also involved in

macrophages and other cells of the innate immune response that attracts neutrophils

Human neutrophil peptide 1 belonging to

developmental response to hypoxia.

peptides.

movement around proposals that reduce the number of animals in research reinforces the need to promote actions that use alternatives; thus, the use of human neutrophils, when suitable, comes from a sample that generates little or no stress to the individual.

The literature regarding human neutrophils in a cancer context is interfered by the unclear identification of different subsets and with functions that might be misinterpreted. Understanding how the activation of purinergic receptors generates intracellular responses might be helpful in discovering novel drug targets. Tumor behavior in the presence of immune cells has attracted attention due to the close connection between them. In summary, more studies regarding neutrophil purinergic activation in human tumor sites are needed to provide new therapeutic strategies based in purine targets.

angiogenesis, cell proliferation, and tissue remodeling. LPS Lipopolysaccharide is a large molecule made of a lipid and a polysaccharide that Glossary occurs in the membrane of Gram-negative A2 α-2 adrenergic G-protein-coupled recepbacteria, acting as a trigger for the innate immune system; it is classified as a PAMP A1/A2a/A2b/A3 P1 purinergic receptors sensitized by (pathogen-associated molecular pattern). adenosine. N1 Neutrophils with antitumor activities. ADO Adenosine is a purine nucleoside that N2 Neutrophils with pro-tumor activities. participates in the purinergic system as a NAD+ The oxidized form of nicotinamide adenine form of extracellular signaling, modulating dinucleotide, a cofactor involved in redox proliferation, differentiation, cell death, reactions, transporting electrons from one and control of inflammatory response substrate to another. events, acting mainly as an The reduced form of NAD+, a cofactor NADH immunosuppressive/immunomodulatory involved in redox reactions. molecule via P1 receptor sensitization. NADP<sup>+</sup> A coenzyme called nicotinamide adenine ADP Adenosine diphosphate is a nucleotide that dinucleotide phosphate, acting as a also participates in the purinergic system as cofactor in anabolic metabolism. a form of extracellular signaling, inducing NETs Neutrophil extracellular traps are a defense platelet aggregation and microglial mechanism, where neutrophils release migration via P2Y<sub>12</sub> sensitization. chromatin to form an extracellular fibril AMP Adenosine monophosphate is a nucleotide matrix, which traps pathogens. formed in the extracellular environment P2 receptors (P2Y<sub>1</sub>, P2Y<sub>2</sub>, Receptors that are activated by purines (e.g. mainly via ATP hydrolysis mediated by P2Y<sub>6</sub>, P2X1, P2X7) ATP, ADP) or pyrimidines (e.g. UTP, UDP). NTPDase1/CD39 enzyme activity. Until PNX-1 Pannexin-1 is a large transmembrane now, no purine-receptor has been dechannel in the plasmatic membrane, scribed to be activated by this nucleotide. allowing the passage of ions and small ATP Adenosine triphosphate is a purine molecules, such as ATP. nucleotide involved in complex TLR4 Toll-like receptor 4 is a cell surface signaling pathways, including driving receptor activated by LPS derived from energy to the cells and being a Gram-negative bacteria or by endogenous precursor to DNA and RNA. In this ligands such as HMGB1, which elicit pocase, it participates in the purinergic tent innate immune responses in several system, a form of extracellular signaling cells such as macrophages, dendritic cells, via the P2 receptor agonist. and neutrophils. NTPDase1/CD39 Ecto-nucleoside triphosphate UDP-glucose Uridine diphosphate-glucose is a nucleodiphosphohydrolase-1 is an enzyme located tide sugar involved in glycosyl-transferase at the cell surface of immune cells and some reactions that activates one of the P2 cancer cells that hydrolyze the P2 receptor purinergic receptors. ligands ATP, ADP, UTP, and UDP to the VCAM-1 Vascular cell adhesion molecule-1 is a cell respective monophosphate-nucleosides by adhesion molecule expressed by the vasremoving one phosphate at a time. cular endothelium.

Ecto-5'-nucleotidase is an enzyme present on the cell surface of a large number of tissues HIF1α

HNP-1

IL-8



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#### **Declarations**

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### References

- Basanta D, Anderson ARA (2017) Homeostasis Back and Forth: An Ecoevolutionary Perspective of Cancer. Cold Spring Harb Perspect Med 7:a028332. https://doi.org/10.1101/cshperspect.a028332
- Medzhitov R (2008) Origin and physiological roles of inflammation. Nature. 454:428–435. https://doi.org/10.1038/nature07201
- Lecot P, Sarabi M, Pereira Abrantes M, Mussard J, Koenderman L, Caux C, Bendriss-Vermare N, Michallet M-C (2019) Neutrophil Heterogeneity in Cancer: From Biology to Therapies. Front Immunol 10. https://doi.org/10.3389/fimmu.2019.02155
- Coussens LM, Werb Z (2002) Inflammation and cancer. Nature. 420:860–867. https://doi.org/10.1038/nature01322
- Donskov F (2013) Immunomonitoring and prognostic relevance of neutrophils in clinical trials. Semin Cancer Biol 23:200–207. https://doi.org/10.1016/j.semcancer.2013.02.001
- Dumitru CA, Lang S, Brandau S (2013) Modulation of neutrophil granulocytes in the tumor microenvironment: Mechanisms and consequences for tumor progression. Semin Cancer Biol 23:141– 148. https://doi.org/10.1016/j.semcancer.2013.02.005
- Grecian R, Whyte MKB, Walmsley SR (2018) The role of neutrophils in cancer. Br Med Bull 128:5–14. https://doi.org/10.1093/ bmb/ldy029
- Wang X, Chen D (2018) Purinergic Regulation of Neutrophil Function. Front Immunol 9. https://doi.org/10.3389/fimmu.2018. 00399
- Sadik CD, Kim ND, Luster AD (2011) Neutrophils cascading their way to inflammation. Trends Immunol 32:452–460. https://doi.org/ 10.1016/j.it.2011.06.008
- Amulic B, Cazalet C, Hayes GL, Metzler KD, Zychlinsky A (2012) Neutrophil Function: From Mechanisms to Disease. Annu Rev Immunol 30:459–489. https://doi.org/10.1146/annurev-immunol-020711-074942
- Mantovani A, Cassatella MA, Costantini C, Jaillon S (2011) Neutrophils in the activation and regulation of innate and adaptive immunity. Nat Rev Immunol 11:519–531. https://doi.org/10.1038/ nri3024

- Giese MA, Hind LE, Huttenlocher A (2019) Neutrophil plasticity in the tumor microenvironment. Blood. 133:2159–2167. https://doi. org/10.1182/blood-2018-11-844548
- Patel S, Fu S, Mastio J, Dominguez GA, Purohit A, Kossenkov A, Lin C, Alicea-Torres K, Sehgal M, Nefedova Y, Zhou J, Languino LR, Clendenin C, Vonderheide RH, Mulligan C, Nam B, Hockstein N, Masters G, Guarino M, Schug ZT, Altieri DC, Gabrilovich DI (2018) Unique pattern of neutrophil migration and function during tumor progression. Nat Immunol 19:1236–1247. https://doi.org/10. 1038/s41590-018-0229-5
- Dosch M, Gerber J, Jebbawi F, Beldi G (2018) Mechanisms of ATP Release by Inflammatory Cells. Int J Mol Sci 19:1222. https://doi. org/10.3390/ijms19041222
- Elliott MR, Chekeni FB, Trampont PC, Lazarowski ER, Kadl A, Walk SF, Park D, Woodson RI, Ostankovich M, Sharma P, Lysiak JJ, Harden TK, Leitinger N, Ravichandran KS (2009) Nucleotides released by apoptotic cells act as a find-me signal to promote phagocytic clearance. Nature. 461:282–286. https://doi.org/10.1038/ nature08296
- Moesta AK, Li X-Y, Smyth MJ (2020) Targeting CD39 in cancer. Nat Rev Immunol 20:739–755. https://doi.org/10.1038/s41577-020-0376-4
- Boison D, Yegutkin GG (2019) Adenosine Metabolism: Emerging Concepts for Cancer Therapy. Cancer Cell 36:582–596. https://doi. org/10.1016/j.ccell.2019.10.007
- Burnstock G (2041) Introduction to Purinergic Signaling. Methods Mol Biol 2020:1–15. https://doi.org/10.1007/978-1-4939-9717-6
- Peleli M, Fredholm BB, Sobrevia L, Carlström M (2017) Pharmacological targeting of adenosine receptor signaling. Mol Asp Med 55:4–8. https://doi.org/10.1016/j.mam.2016.12.002
- Dianzani C, Brunelleschi S, Viano I, Fantozzi R (1994) Adenosine modulation of primed human neutrophils. Eur J Pharmacol 263: 223–226. https://doi.org/10.1016/0014-2999(94)90547-9
- Fredholm BB, Zhang Y, van der Ploeg I (1996) Adenosine A2A receptors mediate the inhibitory effect of adenosine on formyl-Met-Leu-Phe-stimulated respiratory burst in neutrophil leucocytes. Naunyn Schmiedeberg's Arch Pharmacol 354:262–267. https://doi.org/10.1007/BF00171056
- Bazzichi L, Trincavelli L, Rossi A, De Feo F, Lucacchini A, Bombardieri S, Martini C (2005) A2B adenosine receptor activity is reduced in neutrophils from patients with systemic sclerosis. Arthritis Res Ther 7:R189–R195. https://doi.org/10.1186/ar1468
- Chen Y, Corriden R, Inoue Y, Yip L, Hashiguchi N, Zinkernagel A, Nizet V, Insel PA, Junger WG (2006) ATP Release Guides Neutrophil Chemotaxis via P2Y2 and A3 Receptors. Science (80-) 314:1792–1795. https://doi.org/10.1126/science.1132559
- Corriden R, Chen Y, Inoue Y, Beldi G, Robson SC, Insel PA, Junger WG (2008) Ecto-nucleoside Triphosphate Diphosphohydrolase 1 (E-NTPDase1/CD39) Regulates Neutrophil Chemotaxis by Hydrolyzing Released ATP to Adenosine. J Biol Chem 283:28480-28486. https://doi.org/10. 1074/jbc.M800039200
- Gabl M, Winther M, Welin A, Karlsson A, Oprea T, Bylund J, Dahlgren C, Forsman H (2015) P2Y2 receptor signaling in neutrophils is regulated from inside by a novel cytoskeleton-dependent mechanism. Exp Cell Res 336:242–252. https://doi.org/10.1016/j. yexcr.2015.07.014
- Önnheim K, Christenson K, Gabl M, Burbiel JC, Müller CE, Oprea TI, Bylund J, Dahlgren C, Forsman H (2014) A novel receptor cross-talk between the ATP receptor P2Y2 and formyl peptide receptors reactivates desensitized neutrophils to produce superoxide. Exp Cell Res 323:209–217. https://doi.org/10.1016/j.yexcr.2014.01.023
- Nagaoka (2010) Evaluation of the effect of α-defensin human neutrophil peptides on neutrophil apoptosis. Int J Mol Med 26. https://doi.org/10.3892/ijmm 00000544



- Sil P, Hayes CP, Reaves BJ, Breen P, Quinn S, Sokolove J, Rada B (2017) P2Y6 Receptor Antagonist MRS2578 Inhibits Neutrophil Activation and Aggregated Neutrophil Extracellular Trap Formation Induced by Gout-Associated Monosodium Urate Crystals. J Immunol 198:428–442. https://doi.org/10.4049/jimmunol.1600766
- Pliyev BK, Ivanova AV, Savchenko VG (2014) Extracellular NAD+ inhibits human neutrophil apoptosis. Apoptosis. 19:581– 593. https://doi.org/10.1007/s10495-013-0948-x
- Vaughan KR, Stokes L, Prince LR, Marriott HM, Meis S, Kassack MU, Bingle CD, Sabroe I, Surprenant A, Whyte MKB (2007) Inhibition of Neutrophil Apoptosis by ATP Is Mediated by the P2Y 11 Receptor. J Immunol 179:8544

  –8553. https://doi.org/10. 4049/jimmunol.179.12.8544
- Wang X, Qin W, Xu X, Xiong Y, Zhang Y, Zhang H, Sun B (2017) Endotoxin-induced autocrine ATP signaling inhibits neutrophil chemotaxis through enhancing myosin light chain phosphorylation. Proc Natl Acad Sci 114:4483–4488. https://doi.org/10.1073/pnas. 1616752114
- Karmakar M, Katsnelson MA, Dubyak GR, Pearlman E (2016) Neutrophil P2X7 receptors mediate NLRP3 inflammasomedependent IL-1β secretion in response to ATP. Nat Commun 7: 10555. https://doi.org/10.1038/ncomms10555
- Pandolfi F, Altamura S, Frosali S, Conti P (2016) Key Role of DAMP in Inflammation, Cancer, and Tissue Repair. Clin Ther 38:1017–1028. https://doi.org/10.1016/j.clinthera.2016.02.028
- Allard D, Chrobak P, Allard B, Messaoudi N, Stagg J (2019) Targeting the CD73-adenosine axis in immuno-oncology. Immunol Lett 205:31– 39. https://doi.org/10.1016/j.imlet.2018.05.001
- Giuliani AL, Sarti AC, Di Virgilio F (2020) Ectonucleotidases in Acute and Chronic Inflammation. Front Pharmacol 11:619458. https://doi.org/10.3389/fphar.2020.619458
- 36. Ponzetta A, Carriero R, Carnevale S, Barbagallo M, Molgora M, Perucchini C, Magrini E, Gianni F, Kunderfranco P, Polentarutti N, Pasqualini F, Di Marco S, Supino D, Peano C, Cananzi F, Colombo P, Pilotti S, Alomar SY, Bonavita E, Galdiero MR, Garlanda C, Mantovani A, Jaillon S (2019) Neutrophils Driving Unconventional T Cells Mediate Resistance against Murine Sarcomas and Selected Human Tumors. Cell 178:346–360.e24. https://doi.org/10.1016/j.cell.2019.05.047
- Si Y, Merz SF, Jansen P, Wang B, Bruderek K, Altenhoff P, Mattheis S, Lang S, Gunzer M, Klode J, Squire A, Brandau S (2019) Multidimensional imaging provides evidence for downregulation of T cell effector function by MDSC in human cancer tissue. Sci Immunol 4:eaaw9159. https://doi.org/10.1126/ sciimmunol.aaw9159
- Jaillon S, Ponzetta A, Di Mitri D, Santoni A, Bonecchi R, Mantovani A (2020) Neutrophil diversity and plasticity in tumour progression and therapy. Nat Rev Cancer 20:485–503. https://doi. org/10.1038/s41568-020-0281-y
- Zhou J, Nefedova Y, Lei A, Gabrilovich D (2018) Neutrophils and PMN-MDSC: Their biological role and interaction with stromal cells. Semin Immunol 35:19–28. https://doi.org/10.1016/j.smim.2017.12. 004
- Di Virgilio F, Vuerich M (2015) Purinergic signaling in the immune system. Auton Neurosci 191:117–123. https://doi.org/10.1016/j. autneu.2015.04.011
- Ryzhov S, Novitskiy SV, Goldstein AE, Biktasova A, Blackburn MR, Biaggioni I, Dikov MM, Feoktistov I (2011) Adenosinergic Regulation of the Expansion and Immunosuppressive Activity of CD11b + Gr1 + Cells. J Immunol 187:6120–6129. https://doi.org/ 10.4049/jimmunol.1101225
- Bao Y, Ledderose C, Seier T, Graf AF, Brix B, Chong E, Junger WG (2014) Mitochondria Regulate Neutrophil Activation by Generating ATP for Autocrine Purinergic Signaling. J Biol Chem 289:26794–26803. https://doi.org/10.1074/jbc.M114.572495

- Li X, Kondo Y, Bao Y, Staudenmaier L, Lee A, Zhang J, Ledderose C, Junger WG (2017) Systemic Adenosine Triphosphate Impairs Neutrophil Chemotaxis and Host Defense in Sepsis. Crit Care Med 45:e97–e104. https://doi.org/10.1097/CCM.0000000000002052
- De Marchi E, Orioli E, Pegoraro A, Sangaletti S, Portararo P, Curti A, Colombo MP, Di Virgilio F, Adinolfi E (2019) The P2X7 receptor modulates immune cells infiltration, ectonucleotidases expression and extracellular ATP levels in the tumor microenvironment. Oncogene. 38:3636–3650. https://doi.org/10.1038/s41388-019-0684-y
- del Rey A, Renigunta V, Dalpke AH, Leipziger J, Matos JE, Robaye B, Zuzarte M, Kavelaars A, Hanley PJ (2006) Knock-out Mice Reveal the Contributions of P2Y and P2X Receptors to Nucleotide-induced Ca2+ Signaling in Macrophages. J Biol Chem 281:35147–35155. https://doi. org/10.1074/jbc.M607713200
- Adinolfi E, De Marchi E, Orioli E, Pegoraro A, Di Virgilio F (2019)
   Role of the P2X7 receptor in tumor-associated inflammation. Curr Opin Pharmacol 47:59

  –64. https://doi.org/10.1016/j.coph.2019.02.012
- Vijayan D, Young A, Teng MWL, Smyth MJ (2017) Targeting immunosuppressive adenosine in cancer. Nat Rev Cancer 17: 709–724. https://doi.org/10.1038/nrc.2017.86
- 48. Li X-Y, Moesta AK, Xiao C, Nakamura K, Casey M, Zhang H, Madore J, Lepletier A, Aguilera AR, Sundarrajan A, Jacoberger-Foissac C, Wong C, dela Cruz T, Welch M, Lerner AG, Spatola BN, Soros VB, Corbin J, Anderson AC, Effern M, Hölzel M, Robson SC, Johnston RL, Waddell N, Smith C, Bald T, Geetha N, Beers C, Teng MWL, Smyth MJ (2019) Targeting CD39 in Cancer Reveals an Extracellular ATP- and Inflammasome-Driven Tumor Immunity. Cancer Discov 9:1754–1773. https://doi.org/10.1158/2159-8290.CD-19-0541
- Yan J, Li X-Y, Roman Aguilera A, Xiao C, Jacoberger-Foissac C, Nowlan B, Robson SC, Beers C, Moesta AK, Geetha N, Teng MWL, Smyth MJ (2020) Control of Metastases via Myeloid CD39 and NK Cell Effector Function. Cancer Immunol Res 8: 356–367. https://doi.org/10.1158/2326-6066.CIR-19-0749
- Chen Y, Yao Y, Sumi Y, Li A, U.K. To, Elkhal A, Inoue Y, Woehrle T, Zhang Q, Hauser C, Junger WG (2010) Purinergic Signaling: A Fundamental Mechanism in Neutrophil Activation. Sci Signal 3:ra45. https://doi.org/10.1126/scisignal.2000549
- Kukulski F, Ben Yebdri F, Lefebvre J, Warny M, Tessier PA, Sévigny J (2007) Extracellular nucleotides mediate LPS-induced neutrophil migration in vitro and in vivo. J Leukoc Biol 81:1269– 1275. https://doi.org/10.1189/jlb.1206758
- Kukulski F, Ben Yebdri F, Lecka J, Kauffenstein G, Lévesque SA, Martín-Satué M, Sévigny J (2009) Extracellular ATP and P2 receptors are required for IL-8 to induce neutrophil migration. Cytokine. 46:166–170. https://doi.org/10.1016/j.cyto.2009.02.011
- Kukulski F, Ben Yebdri F, Bahrami F, Fausther M, Tremblay A, Sévigny J (2010) Endothelial P2Y2 receptor regulates LPS-induced neutrophil transendothelial migration in vitro. Mol Immunol 47: 991–999. https://doi.org/10.1016/j.molimm.2009.11.020
- 54. Braganhol E, Kukulski F, Lévesque SA, Fausther M, Lavoie EG, Zanotto-Filho A, Bergamin LS, Pelletier J, Bahrami F, Ben Yebdri F, Fonseca Moreira JC, Battastini AMO, Sévigny J (2015) Nucleotide receptors control IL-8/CXCL8 and MCP-1/CCL2 secretions as well as proliferation in human glioma cells. Biochim Biophys Acta Mol basis Dis 1852:120–130. https://doi.org/10.1016/j.bbadis.2014.10.014
- Kukulski F, Bahrami F, Ben Yebdri F, Lecka J, Martín-Satué M, Lévesque SA, Sévigny J (2011) NTPDase1 Controls IL-8 Production by Human Neutrophils. J Immunol 187:644–653. https://doi.org/10.4049/jimmunol.1002680
- Rose FR, Hirschhorn R, Weissmann G, Cronstein BN (1988)
   Adenosine promotes neutrophil chemotaxis. J Exp Med 167: 1186–1194. https://doi.org/10.1084/jem.167.3.1186
- Perrot I, Michaud H-A, Giraudon-Paoli M, Augier S, Docquier A, Gros L, Courtois R, Déjou C, Jecko D, Becquart O, Rispaud-Blanc



- H, Gauthier L, Rossi B, Chanteux S, Gourdin N, Amigues B, Roussel A, Bensussan A, Eliaou J-F, Bastid J, Romagné F, Morel Y, Narni-Mancinelli E, Vivier E, Paturel C, Bonnefoy N (2019) Blocking Antibodies Targeting the CD39/CD73 Immunosuppressive Pathway Unleash Immune Responses in Combination Cancer Therapies. Cell Rep 27:2411–2425.e9. https://doi.org/10.1016/j.celrep.2019.04.091
- Németh T, Sperandio M, Mócsai A (2020) Neutrophils as emerging therapeutic targets. Nat Rev Drug Discov 19:253–275. https://doi. org/10.1038/s41573-019-0054-z
- Richer BC, Salei N, Laskay T, Seeger K (2018) Changes in Neutrophil Metabolism upon Activation and Aging. Inflammation. 41:710–721. https://doi.org/10.1007/s10753-017-0725-z
- Shaul ME, Levy L, Sun J, Mishalian I, Singhal S, Kapoor V, Horng W, Fridlender G, Albelda SM, Fridlender ZG (2016) Tumorassociated neutrophils display a distinct N1 profile following TGFβ modulation: A transcriptomics analysis of pro- vs. antitumor TANs. Oncoimmunology 5:e1232221. https://doi.org/10.1080/ 2162402X.2016.1232221
- Grassi F (2010) Purinergic Control of Neutrophil Activation. J Mol Cell Biol 2:176–177. https://doi.org/10.1093/jmcb/mjq014
- Martins I, Wang Y, Michaud M, Ma Y, Sukkurwala AQ, Shen S, Kepp O, Métivier D, Galluzzi L, Perfettini J-L, Zitvogel L, Kroemer G (2014) Molecular mechanisms of ATP secretion during immunogenic cell death. Cell Death Differ 21:79–91. https://doi. org/10.1038/cdd.2013.75
- Campos-Contreras A d R, Díaz-Muñoz M, Vázquez-Cuevas FG (2020) Purinergic Signaling in the Hallmarks of Cancer. Cells 9: 1612. https://doi.org/10.3390/cells9071612
- 64. Mehta VB, Hart J, Wewers MD (2001) ATP-stimulated Release of Interleukin (IL)-1 β and IL-18 Requires Priming by Lipopolysaccharide and Is Independent of Caspase-1 Cleavage. J Biol Chem 276:3820–3826. https://doi.org/10.1074/jbc. M006814200
- Adinolfi E, Giuliani AL, De Marchi E, Pegoraro A, Orioli E, Di Virgilio F (2018) The P2X7 receptor: A main player in inflammation. Biochem Pharmacol 151:234–244. https://doi.org/10.1016/j. bcp.2017.12.021
- Amoroso F, Capece M, Rotondo A, Cangelosi D, Ferracin M, Franceschini A, Raffaghello L, Pistoia V, Varesio L, Adinolfi E (2015) The P2X7 receptor is a key modulator of the PI3K/GSK3β/VEGF signaling network: evidence in experimental neuroblastoma. Oncogene. 34:5240–5251. https://doi.org/10.1038/ onc.2014.444
- 67. De Marchi E, Orioli E, Dal Ben D, Adinolfi E (2016) P2X7 Receptor as a Therapeutic Target. pp. 39–79. https://doi.org/10.1016/bs.apcsb.2015.11.004
- Di Virgilio F, Sarti AC, Falzoni S, De Marchi E, Adinolfi E (2018) Extracellular ATP and P2 purinergic signalling in the tumour microenvironment. Nat Rev Cancer 18:601–618. https://doi.org/10.1038/s41568-018-0037-0
- Bou Ghanem EN, Clark S, Roggensack SE, McIver SR, Alcaide P, Haydon PG, Leong JM (2015) Extracellular Adenosine Protects against Streptococcus pneumoniae Lung Infection by Regulating Pulmonary Neutrophil Recruitment. PLoS Pathog 11:e1005126. https://doi.org/10.1371/journal.ppat.1005126
- Amulic B, Sollberger G (2018) Why Immune Cells Extrude Webs of DNA and Protein. NETS: two faced players in Immunity. Science 33:44–51
- Sorvillo N, Cherpokova D, Martinod K, Wagner DD (2019) Extracellular DNA NET-Works With Dire Consequences for Health. Circ Res 125:470–488. https://doi.org/10.1161/ CIRCRESAHA.119.314581
- Hurt B, Schulick R, Edil B, El Kasmi KC, Barnett C (2017) Cancerpromoting mechanisms of tumor-associated neutrophils. Am J Surg 214:938–944. https://doi.org/10.1016/j.amjsurg.2017.08.003

- Galdiero MR, Varricchi G, Loffredo S, Mantovani A, Marone G (2018) Roles of neutrophils in cancer growth and progression. J Leukoc Biol 103:457–464. https://doi.org/10.1002/JLB. 3MR0717-292R
- 74. Loffredo S, Borriello F, Iannone R, Ferrara AL, Galdiero MR, Gigantino V, Esposito P, Varricchi G, Lambeau G, Cassatella MA, Granata F, Marone G (2017) Group V Secreted Phospholipase A2 Induces the Release of Proangiogenic and Antiangiogenic Factors by Human Neutrophils. Front Immunol 8. https://doi.org/10.3389/fimmu.2017.00443
- Martel-Gallegos G, Rosales-Saavedra MT, Reyes JP, Casas-Pruneda G, Toro-Castillo C, Pérez-Comejo P, Arreola J (2010) Human neutrophils do not express purinergic P2X7 receptors. Purinergic Signal 6: 297–306. https://doi.org/10.1007/s11302-010-9178-7
- Park J, Wysocki RW, Amoozgar Z, Maiorino L, Fein MR, Jorns J, Schott AF, Kinugasa-Katayama Y, Lee Y, Won NH, Nakasone ES, Hearn SA, Kuttner V, Qiu J, Almeida AS, Perurena N, Kessenbrock K, Goldberg MS, Egeblad M (2016) Cancer cells induce metastasis-supporting neutrophil extracellular DNA traps. Sci Transl Med 8:361ra138. https://doi.org/10.1126/scitranslmed. aag1711
- Rayes RF, Mouhanna JG, Nicolau I, Bourdeau F, Giannias B, Rousseau S, Quail D, Walsh L, Sangwan V, Bertos N, Cools-Lartigue J, Ferri LE, Spicer JD (2019) Primary tumors induce neutrophil extracellular traps with targetable metastasis-promoting effects. JCI Insight 4. https://doi.org/10.1172/jci.insight.128008
- Arpinati L, Shaul ME, Kaisar-Iluz N, Mali S, Mahroum S, Fridlender ZG (2020) NETosis in cancer: a critical analysis of the impact of cancer on neutrophil extracellular trap (NET) release in lung cancer patients vs. mice. Cancer Immunol Immunother 69: 199–213. https://doi.org/10.1007/s00262-019-02474-x
- Berger-Achituv S, Brinkmann V, Abed UA, Kühn LI, Ben-Ezra J, Elhasid R, Zychlinsky A (2013) A proposed role for neutrophil extracellular traps in cancer immunoediting. Front Immunol 4. https://doi.org/10.3389/fimmu.2013.00048
- Perisanidis C, Kornek G, Pöschl PW, Holzinger D, Pirklbauer K, Schopper C, Ewers R (2013) High neutrophil-to-lymphocyte ratio is an independent marker of poor disease-specific survival in patients with oral cancer. Med Oncol 30:334. https://doi.org/10.1007/ s12032-012-0334-5
- Xiao W-K, Chen D, Li S-Q, Fu S-J, Peng B-G, Liang L-J (2014) Prognostic significance of neutrophil-lymphocyte ratio in hepatocellular carcinoma: a meta-analysis. BMC Cancer 14:117. https:// doi.org/10.1186/1471-2407-14-117
- Zhao J, Pan K, Wang W, Chen J, Wu Y, Lv L, Li J, Chen Y, Wang D, Pan Q, Li X, Xia J (2012) The Prognostic Value of Tumor-Infiltrating Neutrophils in Gastric Adenocarcinoma after Resection. PLoS One 7:e33655. https://doi.org/10.1371/journal.pone.0033655
- Carmona-Rivera C, Khaznadar SS, Shwin KW, Irizarry-Caro JA, O'Neil LJ, Liu Y, Jacobson KA, Ombrello AK, Stone DL, Tsai WL, Kastner DL, Aksentijevich I, Kaplan MJ, Grayson PC (2019) Deficiency of adenosine deaminase 2 triggers adenosinemediated NETosis and TNF production in patients with DADA2. Blood. 134:395–406. https://doi.org/10.1182/blood.2018892752
- 84. Xu K, Cooney KA, Shin EY, Wang L, Deppen JN, Ginn SC, Levit RD (2019) Adenosine from a biologic source regulates neutrophil extracellular traps (NETs). J Leukoc Biol 105:1225–1234. https://doi.org/10.1002/JLB.3VMA0918-374R
- Mestas J, Hughes CCW (2004) Of Mice and Not Men: Differences between Mouse and Human Immunology. J Immunol 172:2731– 2738. https://doi.org/10.4049/jimmunol.172.5.2731
- Wiesner O, Litwiller RD, Hummel AM, Viss MA, McDonald CJ, Jenne DE, Fass DN, Specks U (2005) Differences between human proteinase 3 and neutrophil elastase and their murine homologues



- are relevant for murine model experiments. FEBS Lett 579:5305–5312. https://doi.org/10.1016/j.febslet.2005.08.056
- Stackowicz J, Jönsson F, Reber LL (2020) Mouse Models and Tools for the in vivo Study of Neutrophils. Front Immunol 10. https://doi.org/10.3389/fimmu.2019.03130
- Sugawara T, Miyamoto M, Takayama S, Kato M (1995) Separation of neutrophils from blood in human and laboratory animals and comparison of the chemotaxis. J Pharmacol Toxicol Methods 33: 91–100. https://doi.org/10.1016/1056-8719(94)00062-9
- Soroush F, Tang Y, Mustafa O, Sun S, Yang Q, Kilpatrick LE, Kiani MF (2020) Neutrophil-endothelial interactions of murine cells is not a good predictor of their interactions in human cells. FASEB J 34:2691–2702. https://doi.org/10.1096/fj.201900048R

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