

Review



Arachidonic Acid 15-Lipoxygenase: Effects of Its Expression, Metabolites, and Genetic and Epigenetic Variations on Airway Inflammation

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ABSTRACT

Arachidonic acid 15-lipoxygenase (ALOX15) is an enzyme that can oxidize polyunsaturated fatty acids. ALOX15 is strongly expressed in airway epithelial cells, where it catalyzes the conversion of arachidonic acid to 15-hydroxyeicosatetraenoic acid (15-HETE) involved in various airway inflammatory diseases. Interleukin (IL)-4 and IL-13 induce ALOX15 expression by activating Jak2 and Tyk2 kinases as well as signal transducers and activators of transcription (STATs) 1/3/5/6. ALOX15 up-regulation and subsequent association with phosphatidylethanolamine-binding protein 1 (PEBP1) activate the mitogen-activated extracellular signal-regulated kinase (MEK)-extracellular signal-regulated kinase (ERK) pathway, thus inducing eosinophil-mediated airway inflammation. In addition, ALOX15 plays a significant role in promoting the migration of immune cells, such as immature dendritic cells, activated T cells, and mast cells, and airway remodeling, including goblet cell differentiation. Genome-wide association studies have revealed multiple ALOX15 variants and their significant correlation with the risk of developing airway diseases. The epigenetic modifications of the *ALOX15* gene, such as DNA methylation and histone modifications, have been shown to closely relate with airway inflammation. This review summarizes the role of ALOX15 in different phenotypes of asthma, chronic obstructive pulmonary disease, chronic rhinosinusitis, aspirin-exacerbated respiratory disease, and nasal polyps, suggesting new treatment strategies for these airway inflammatory diseases with complex etiology and poor treatment response.

Keywords: Arachidonic acid; respiratory tract; inflammation; genetics; epigenomics

INTRODUCTION

There is a high incidence of worldwide airway inflammatory diseases seriously affecting the quality of life of patients. Recent epidemiological surveys of the Chinese population have shown that the prevalence rates of asthma, chronic rhinosinusitis (CRS), and chronic obstructive pulmonary disease (COPD) are 4.2%, 2.1%, and 8.87%, respectively.¹⁻³ These airway diseases often co-exist in patients and share common molecular mechanisms, which should be focused on in order to develop effective treatment approaches. In this review, we systematically analyzed the role of arachidonic acid 15-lipoxygenase (ALOX15), including its

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genetic variations and epigenetic modifications, in eosinophilic inflammation, immune cell migration, and airway remodeling, which are the key processes underlying the development of different phenotypes of asthma, COPD, CRS, and aspirin-exacerbated respiratory disease (AERD). The clinical significance of ALOX15 as a potential drug target was also evaluated.

ALOX15 OVERVIEW

Enzymatic activity

Polyunsaturated fatty acids (PUFAs) and their metabolites play a vital role in the growth and development of normal cells. Lipoxygenases (LOXs) are heme-free dioxygenases that can catalyze the peroxidation of PUFAs to corresponding hydroperoxy derivatives.⁴ The human genome contains 6 functional LOX genes (*ALOX15*, *ALOX15B*, *ALOX12*, *ALOX12B*, *ALOX5*, and *ALOXE3*), each encoding a different LOX enzyme.⁵ ALOX15 (also called 15-LOX) uses physiological substrates such as arachidonic acid (AA), docosahexaenoic acid (DHA), eicosapentaenoic acid (EPA), α -linolenic acid (ALA), γ -linolenic acid (GLA), and linoleic acid (LA), which exist either in a free form or are incorporated into phospholipids, glycerides, or cholesterol esters.⁶ AA, an ω -6 PUFA representing a major component of the cell membrane phospholipids and the metabolic precursor of eicosanoids,⁷ is converted by ALOX15 into 15-[S]-hydroperoxyeicosatetraenoic acid (15-[S]-HPETE) and 15-hydroxyeicosatetraenoic acid (15-HETE), further metabolized into various bioactive molecules.⁶ The reactions of ALOX15 with DHA, EPA, and other substrates also produce metabolites with diverse physiological activities involved in the development of various diseases.^{8,10}

Physiological functions

Normally, ALOX15 is expressed in the cells of the hematopoietic, endocrine, immune, and respiratory systems, where it has different physiological functions and plays an important role in maintaining body homeostasis.^{11,14} In the respiratory system, ALOX15 has a higher level of expression in airway epithelial cells, where it releases chemokines acting on immune cells such as eosinophils, and regulates mucus secretion. In human airway epithelial cells, ALOX15 participates in the production of eoxins (EXs) that increase the permeability of endothelial cells *in vitro*: 15-[S]-HPETE produced by ALOX15 from AA is converted into EXA4, which is used by glutathione transferase P1-1 to generate EXC4 being further converted into EXD4 and EXE4.¹⁵ During mucociliary differentiation of normal human tracheobronchial epithelial cells, ALOX15 is not expressed at the early stage, but is strongly up-regulated at the late stage in fully differentiated cells, where it metabolizes AA and LA into 15-HETE and 13-hydroxyoctadecadienoic acid (13-HODE), respectively.¹⁶ Higher levels of ALOX15 in the epithelium correlate with the activation of extracellular signal-regulated kinase (ERK). ALOX15-generated 15-HETE is conjugated to phosphatidylethanolamine (PE) and then form 15-HETE-PE; both ALOX15 and 15-HETE-PE act as signaling molecules interacting with phosphatidylethanolamine-binding protein 1 (PEBP1) to activate ERK, thus resulting in the induction of interleukin (IL)-4 receptor α (IL-4R α) downstream pathways.¹⁷ Under homeostatic conditions, PEBP1, also known as Raf-1 kinase inhibitor, binds to Raf-1 and prevents its phosphorylation, thus inhibiting Raf-1-mediated mitogen-activated protein kinase (MAPK) signaling.¹⁸ Furthermore, ALOX15 expression is induced by IL-4 and IL-13 through activation of Jak2 and Tyk2 kinases as well as signal transducers and activators of transcription (STATs)-1/3/5/6.¹⁹ Thus, after induction with IL-4/IL-13, both ALOX15 and its product 15-HETE-PE cause competitive dissociation of Raf-1 from PEBP1 to activate ERK, which enhances the expression of IL-4R α signaling-dependent genes.¹⁷ Cumulatively, these data indicate that in

human airway epithelial cells, ALOX15 interacts with PEBP1 to sustain MAPK/ERK activation and enhance pro-inflammatory signaling pathways related to airway diseases.

ALOX15 type B (ALOX15B), first found by Brash *et al.*²⁰ in the human skin, has about 40% homology with ALOX15 and shows similar enzymatic activity towards AA; it differs from ALOX15 in the catalytic profile as well as in tissue distribution. Thus, the expression of ALOX15 in human macrophages is inducible, whereas that of ALOX15B is constitutive, suggesting that ALOX15B is involved in the maintenance of lipid homeostasis in macrophages; however, its role in the respiratory system is not as significant as that of ALOX15.^{21,22}

ROLE OF ALOX15 IN AIRWAY INFLAMMATORY DISEASES

ALOX15 and its metabolites are important factors in the pathophysiology of diseases of both the lower respiratory tract (asthma and COPD) and the upper respiratory tract (CRS and AERD) because of their role in the regulation of eosinophil-mediated inflammation, migration of immune cells (immature dendritic cells, activated T cells, and mast cells), and airway remodeling including goblet cell differentiation. Here, we summarize the pathological effects of ALOX15 and its metabolites in various airway diseases (**Table**) as well as the molecular mechanisms underlying airway inflammation (**Fig. 1**).

ALOX15 promotes eosinophilic inflammation

Although ALOX15 is not expressed in unstimulated human peripheral macrophages, it is significantly up-regulated by type 2 inflammatory factors. In patients with asthma, airway macrophages are the main source of 15-HETE, whose level correlates with tissue eosinophil numbers, sub-basement membrane thickness, and expression of tissue inhibitor of metalloproteinase-1 in bronchoalveolar lavage fluid (BALF).^{23,24} Another ALOX15 product in macrophages, 12-[S]-HETE, has been shown to cause bronchial epithelial damage in asthma.²⁵

In addition to macrophages, ALOX15 is strongly expressed in airway epithelial cells under physiological and pathological conditions. Ordovas-Montanes *et al.*²⁶ carried out comprehensive expression profiling in CRS using single-cell RNA sequencing and revealed the difference in ALOX15 expression between non-polyp and polyp tissues. In non-polyp tissues, ALOX15 was found to be mainly expressed in ciliated cells, whereas in polyps, its expression was high in ciliated, apical, basal, and glandular cells.

Several studies have proved that ALOX15 participates in eosinophil-mediated inflammation, which plays a key role in the occurrence and progression of asthma and COPD. A high

Table. The involvement of ALOX15 and its metabolites in the pathogenesis of airway inflammatory diseases

Disease	ALOX15 expression	Main metabolites	Pathogenesis	Reference
Asthma	Upregulated in bronchial epithelium and macrophages	15-HETE	Tissue eosinophilic inflammation	23,24
		15-HETE, 5,15-diHETE, 13-HODE	Airway hyperresponsiveness	31
		12-[S]-HETE	Bronchial epithelial damage	25
		15-HETE-PE	Goblet cell metaplasia and mucus hypersecretion	44,45
CRS	Upregulated in nasal epithelium	15-HETE	Tissue eosinophilic inflammation	35
		ND	Epithelial-mesenchymal transition	49
COPD	Upregulated in whole blood	ND	Tissue eosinophilic inflammation	27
AERD	Upregulated in nasal epithelium	15-HETE	Abnormal metabolism of AA	37-39

AA, arachidonic acid; AERD, aspirin-exacerbated respiratory disease; ALOX15, arachidonic acid 15-lipoxygenase; COPD, chronic obstructive pulmonary disease; CRS, chronic rhinosinusitis; 5,15-diHETE, 5S,15S-dihydroxyeicosatetraenoic acid; ERK, extracellular signal-regulated kinase; 15-HETE, 15-hydroxyeicosatetraenoic acid; PE, phosphatidylethanolamine; 13-HODE, 13-hydroxyoctadecadienoic acid; 12-[S]-HETE, 12-[S]-hydroxyeicosatetraenoic acid; ND, not determined.

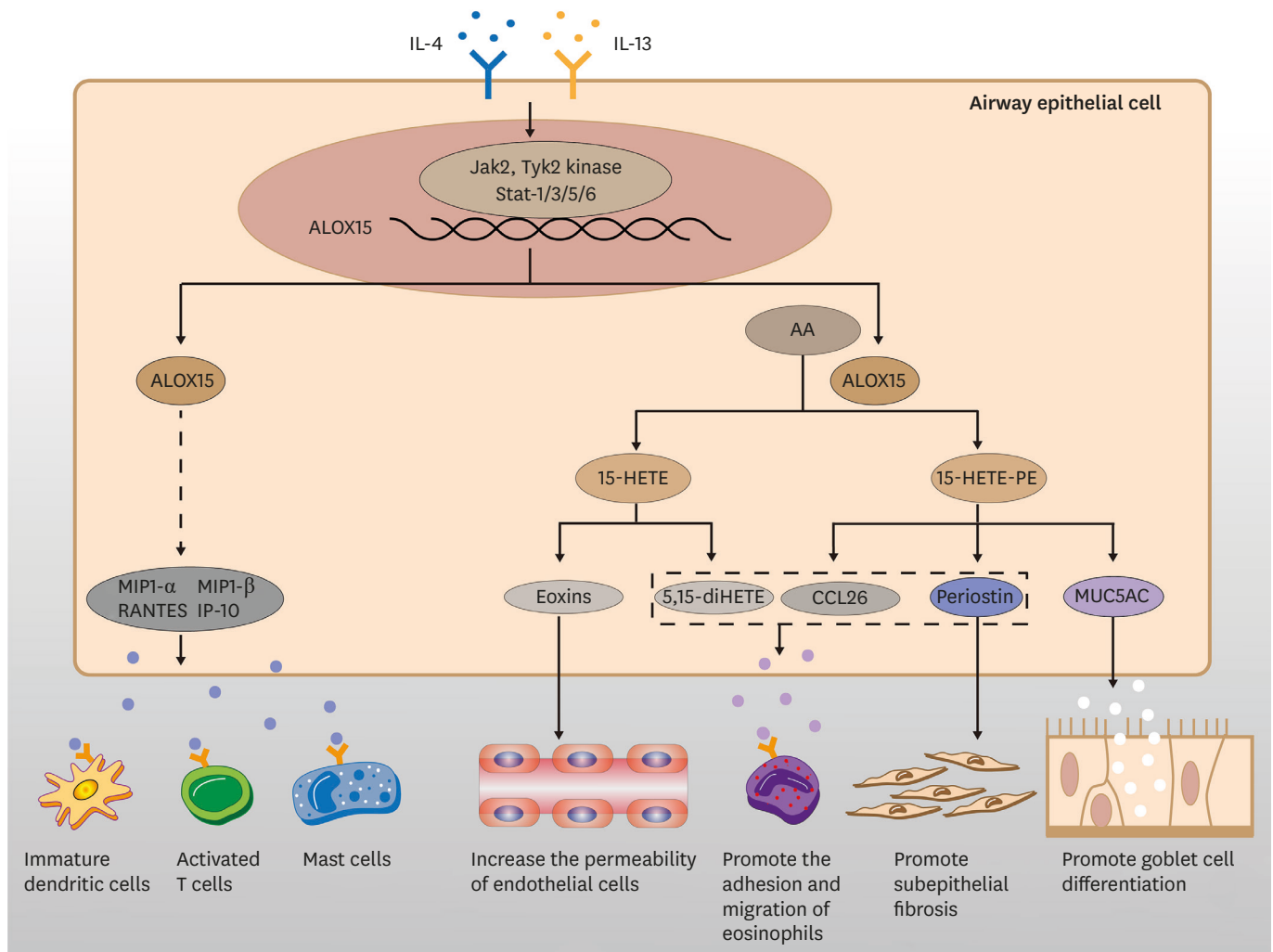


Fig. 1. Specific molecular metabolic pathways of ALOX15 and its reaction products in airway inflammation. ALOX15 induces the expression of chemokines such as MIP-1 α , MIP-1 β , RANTES, and IP-10, thus promoting the migration of immature dendritic cells, activated T cells, and mast cells. ALOX15 expressed in human airway epithelial cells catalyzes the conversion of AA into 15-HETE, which is then conjugated with PE to form 15-HETE-PE or further metabolized into biologically active molecules such as eoxins and 5,15-diHETE. 15-HETE-PE induces the expression of CCL26, MUC5AC, and periostin to promote the migration of eosinophils, the differentiation of goblet cells, the adhesion and migration of eosinophils, and subepithelial fibrosis, whereas eoxins increase the permeability of endothelial cells, and 5,15-diHETE promotes eosinophil infiltration.

ALOX15, arachidonic acid 15-lipoxygenase; MIP, macrophage inflammatory protein; RANTES, Regulated upon Activation, Normal T Cell Expressed and Presumably Secreted; IP-10, interferon- γ -inducible protein 10; AA, arachidonic acid; 15-HETE, 15-hydroxyeicosatetraenoic acid; PE, phosphatidylethanolamine; 5,15-diHETE, 5S,15S-dihydroxyeicosatetraenoic acid; CCL26, chemokine ligand 26; MUC5AC, mucin 5AC.

proportion of patients with COPD (20%–40%) have eosinophilic inflammation and demonstrate the up-regulation of ALOX15 expression.^{27,28} Similarly, most patients with asthma have a pattern of allergic inflammation in the airways characterized by eosinophilia.²⁹ In the BALF of patients with asthma, the level of 15-HETE, which is the main metabolite of the ALOX15 pathway, is significantly higher than in that of normal individuals, and highest in patients having severe asthma with airway eosinophilia.²⁴ Transcriptome analysis of blood samples from 298 patients with moderate to severe asthma revealed that ALOX15 was one of the 50 genes showing the strongest correlation with the absolute eosinophil count in peripheral blood, and that its expression could predict eosinophilic airway inflammation, blood eosinophil numbers, and anti-IL-13 therapeutic effect.³⁰ In patients with asthma treated with benralizumab, a humanized non-glycosyl monoclonal antibody targeting the

IL-5 receptor, ALOX15 expression was significantly decreased, and this effect was stronger in patients with high eosinophil counts than with low eosinophil counts.²⁷ In a mouse model of asthma induced by house dust mites, the levels of various ALOX15 products, such as 15-HETE, 5S,15S-dihydroxyeicosatetraenoic acid (5,15-diHETE), and 13-HODE were significantly increased, resulting in airway hyperresponsiveness and pulmonary eosinophilia.³¹ Overall, these findings indicate that ALOX15 and its metabolites stimulate eosinophilia in airway diseases, thus playing a key role in subsequent airway inflammation.

In addition to lower respiratory tract diseases, ALOX15 has a similar pro-inflammatory function in upper respiratory tract diseases including CRS and AERD. CRS is a complex heterogeneous condition usually characterized by type 2 inflammation driven by IL-5 and IL-13, which activate inflammatory cells such as mast cells and eosinophils.^{32,33} It has been shown that ALOX15 expression is significantly increased in all patients with CRS compared to healthy controls.³⁴ A recent study has indicated that the expression of ALOX15 is significantly up-regulated in nasal polyp (NP) epithelial cells and middle turbinate epithelial cells of patients with chronic rhinosinusitis with nasal polyps (CRSwNP) and co-localized with that of eosinophil chemokine ligand 26 (CCL26) which mediates the recruitment and activation of eosinophils at inflammation sites.³⁵ In addition, increasing evidence suggests that ALOX15-produced 15-HETE plays an important role in aspirin intolerance. Furthermore, ALOX15 expression and 15-HETE production have been shown to be increased in patients with CRS and aspirin intolerance.³⁶⁻³⁹

Altogether these data indicate that ALOX15 participates in the airway diseases of the upper and lower respiratory tracts by regulating eosinophilic inflammation. In airway epithelial cells of patients with asthma or CRSwNP, IL-4 and/or IL-13 not only induce ALOX15 expression and 15-HETE-PE production, but also up-regulate CCL26 through activation of the MAPK/ERK pathway.^{17,40} In turn, CCL26 activates C-C chemokine receptor 3 (CCR3), thus stimulating the infiltration of eosinophils from the circulation into tissues, which is closely related to the pathogenesis of airway inflammatory diseases.⁴¹

ALOX15 promotes the migration of immune cells

The increased infiltration of immature dendritic cells, activated T cells, and mast cells into airway tissues is an important feature of airway inflammatory diseases such as asthma and COPD. In an A549 cell model, Liu *et al.*⁴² found that the ectopic expression of ALOX15 resulted in the production of 15-HETE, 8S,15S-diHETE, and 15-keto-6Z,8Z,11Z,13E-eicosatetraenoic acid (15-KETE), as well as the secretion of chemokines macrophage inflammatory protein (MIP)-1 α (CCL3), MIP-1 β (CCL4), RANTES (CCL5), and IP-10 (CXCL10), which promoted the migration of immature dendritic cells, activated T cells, and mast cells in the chemotaxis experiment. Antibody neutralization experiments have shown that the enhanced immune cell recruitment mainly depends on the up-regulation of RANTES and MIP-1 release induced by ALOX15. The siRNA-mediated knockdown of ALOX15 leads to reduced levels of 15-HETE and the down-regulation of MIP-1 α , RANTES, and IP-10. These results indicate that ALOX15 overexpression in human airway epithelial cells can cause transcriptional activation of chemokines-encoding genes, which results in the enhanced migration of immune cells to inflammatory sites. Although the underlying mechanism is still unclear, it could play an important role in the occurrence and progression of airway inflammatory diseases.

ALOX15 promotes airway remodeling

Airway remodeling is an important process in the pathogenesis of airway diseases.⁴³ Studies in models of human asthma have shown that ALOX15 and its main product, 15-HETE-PE,

are essential for pulmonary tissue remodeling, especially for the differentiation of goblet cells. Thus, it has been shown that in airway epithelial cells, IL-13 not only induces ALOX15 expression and production of 15-HETE-PE which increases the synthesis of mucin 5AC (MUC5AC), but also up-regulates the expression of periostin,^{44,45} a protein that promotes the migration and adhesion of eosinophils, and promotes subepithelial fibrosis.^{45,46} At the same time, IL-13-induced ALOX15 expression correlates with the up-regulation of FOXA3 and downregulation of FOXA2.⁴⁷ FOXA3, a transcription factor binding to proximal promoters of genes associated with goblet cell metaplasia, was reported to induce goblet cell metaplasia and enhance MUC5AC expression, whereas FOXA2, another transcription factor functioning in airway epithelial cells, reduced goblet cell metaplasia and stimulated ciliated cell differentiation.^{47,48} The knockout of ALOX15 by siRNA significantly down-regulated the expression of FOXA3, MUC5AC, and periostin induced by IL-13, whereas exogenous 15-HETE-PE could up-regulate these factors even in the absence of IL-13 stimulation.⁴⁷ These data further prove that 15-HETE-PE generated by ALOX15 participates in airway tissue remodeling in asthma by stimulating goblet cell differentiation and periostin expression. Although the mechanisms by which ALOX15 controls the expression of FOXA transcription factors need further investigation, it is possible that ERK/MAPK activation and/or the interaction of membrane phospholipids with IL-4R α or its downstream signaling pathways may be involved.

In addition, it has been shown that in CRSwNP infiltrated by eosinophils, the basement membrane thickness was larger, and that the matrix metalloproteinase 1 expression positively correlates with ALOX15 transcription.⁴⁹

Thus, ALOX15 plays an important pathophysiological role in airway tissue remodeling associated with airway inflammation, suggesting that it is a critical factor in the natural course of airway diseases.

GENETIC VARIATIONS AND EPIGENETIC MODIFICATIONS IN ALOX15

The presence of multiple variants of the *ALOX15* gene and its epigenetic modifications, including DNA methylation and histone modification, are related to a variety of airway diseases (**Fig. 2**).

Genetic variations

Numerous studies indicate that there are several mutations at various positions of the *ALOX15* gene, which have different effects on airway diseases, increasing or decreasing the risk of disease development. Therefore, the assessment of the allele frequency of these *ALOX15* variants in the population and their pathophysiological role is essential for the development of effective treatment strategies.

Wittwer *et al.*⁵⁰ detected the c.-292C>T mutation in the *ALOX15* promoter region, which doubled *ALOX15* transcription in macrophages, suggesting that the presence of this *ALOX15* variant may increase the production of AA metabolites participating in the pathogenesis of asthma. Some *ALOX15* variants have been shown to exacerbate inflammation by regulating the synthesis and release of pro-inflammatory IL-6. Thus, Fairfax *et al.*⁵¹ reported that specific *ALOX15* polymorphisms (rs11078527 and rs11568131) were significantly associated with IL-6 production in lipopolysaccharide-stimulated primary human peripheral blood mononuclear

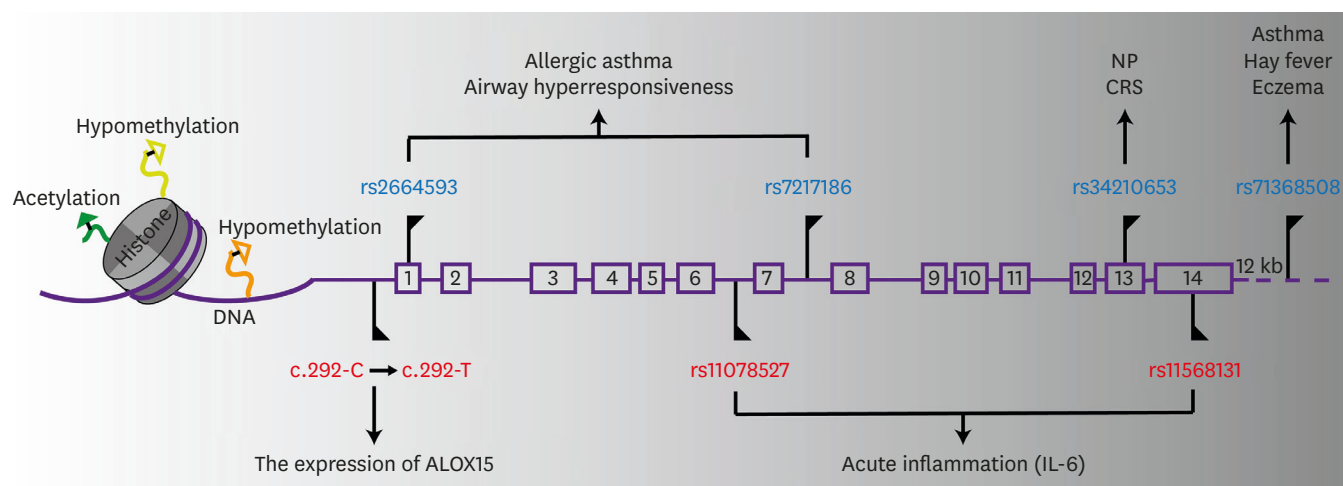


Fig. 2. The role of ALOX15 genetic variations and epigenetic modifications in airway inflammatory diseases. Multiple genetic variants of ALOX15 carrying mutations at different sites have distinct effects on airway diseases, with some increasing the risk of developing airway inflammation (c.292C>T, rs11078527, and rs11568131) and others decreasing it (rs2664593, rs7217186, rs34210653, and rs71368508). Epigenetic modifications regulating the transcription of the ALOX15 gene in the airway epithelium mainly include DNA methylation and histone modifications. DNA and histone (H3K27me3 and H3K9me2) hypomethylation and histone acetylation can increase ALOX15 expression. The numbers in the boxes represent the order of exons in the ALOX15 gene. ALOX15, arachidonic acid 15-lipoxygenase; NP, nasal polyp; CRS, chronic rhinosinusitis; IL, interleukin.

cells and could influence the expression of tumor necrosis factor (TNF) and IL-1 β through trans-acting effects, thereby promoting acute inflammation.

While some ALOX15 variants are pro-inflammatory and may increase the susceptibility to airway inflammatory diseases, others have been found to reduce the risk of airway inflammation. Tremblay *et al.*⁵² reported the protective effects of ALOX15 variants rs2664593 and rs7217186 in allergic asthma and airway hyperresponsiveness. Ferreira *et al.*⁵³ conducted a genome-wide association study (GWAS) on a wide range of allergic disease phenotypes in 180,129 patients with asthma, hay fever, and/or eczema in Europe and detected 136 independent risk-associated variants; among them, ALOX15 rs71368508 could reduce the risk of allergic diseases. Another ALOX15 variant has been suggested to have a significant protective effect against developing NPs and CRS. Kristjansson *et al.*⁵⁴ in a GWAS study with 4,366 NP and 5,608 CRS samples from Iceland and the UK revealed gene mutations related to NPs and CRS; among them, the highest association with the diseases was shown by low-frequency ALOX15 missense variant rs34210653 (p.T560M), which could reduce the risk of NPs and CRS by 68% and 36%, respectively. Schurmann *et al.*⁵⁵ showed that the p.T560M ALOX15 mutant lacked an OH group and could not form a hydrogen bridge, which significantly reduced its catalytic activity. This finding suggests that the p.T560M substitution in ALOX1 may decrease the production of pro-inflammatory mediators in eosinophils and nasal mucosal epithelial cells as well as reduce the number of circulating eosinophils, thus protecting against NPs and CRS. It is worth noting that the frequency of rs34210653 (p.T560M) varies among racial groups: it is highest in Hispanics (8.0%), followed by white Europeans (1.2%), lowest in Black/African Americans (0.5%), and not detected in East Asians (0%),⁵⁶ suggesting that the association of rs34210653 with CRS and NPs may be race-specific.

Epigenetic modifications

Several studies have indicated that epigenetic mechanisms regulate the transcription of the ALOX15 gene in the airway epithelium and that methylation and acetylation modifications are increased in inflammation. Thus, a recent study of children with allergic asthma found

significant changes in the methylation status of 47 allergy-associated genes involved in asthma, which inversely correlated with gene expression: in particular, the hypomethylation of *ALOX15* increased its expression, which was associated with the pathogenesis of asthma.⁵⁷ Besides DNA methylation, histone modifications at the *ALOX15* promoter site also play a key role in transcriptional regulation. There is evidence that in IL-4-treated epithelial cells, the hypomethylation in histone H3 trimethyllysine 27 (H3K27me3) at the *ALOX15* promoter upregulates ALOX15 expression.⁵⁸ It was found that in colon cancer cells, the acetylation of H3 and H4, and hypomethylation at the H3K9 site could activate ALOX15.⁵⁹ However, further studies are needed to confirm the role of histone modifications in airway diseases. These results characterize ALOX15 as an epigenetic marker for airway diseases because DNA hypomethylation and histone modifications up-regulating ALOX15 expression should result in the elevated production of pro-inflammatory metabolites involved in all stages of airway inflammation-related diseases.

EFFECTS OF ALOX15 INHIBITORS

Several natural compounds, such as flavonoids, coumarins, catechins, and emodin, have been shown to exert significant inhibitory effects on ALOX15.⁶⁰ These inhibitors act on distinct regulatory pathways and may potentially have therapeutic effects on airway inflammation. Flavonoids are powerful antioxidants with anti-allergic activity, which can suppress the release of pro-inflammatory mediators and Th2 cytokines (IL-4 and IL-13), and down-regulate CD40 ligand expression in high-affinity immunoglobulin E (IgE) receptor-positive cells (mast cells and basophils), thus making flavonoid intake beneficial for patients with asthma.⁶¹ Coumarin derivatives can reduce allergic inflammation by reducing the release of histamine and the secretion of TNF- α , IL-1 β , and IL-4.⁶² Epigallo-catechin-3-gallate is a catechin that can inhibit the epithelial-mesenchymal transition induced by transforming growth factor β 1 (TGF- β 1) and prevent the migration of human bronchial epithelial cells, thereby reducing airway remodeling in asthma.⁶³ In a mouse model of asthma, emodin was found to suppress the infiltration of inflammatory cells through inhibition of the Notch pathway and decrease the levels of inflammatory cytokines such as IL-5, IL-17, and interferon- γ in BALF and serum.⁶⁴ These data indicate that plant-derived ALOX15 inhibitors have therapeutic potential in the treatment of airway inflammatory diseases.

CONCLUSIONS

The role of ALOX15 in airway diseases is underlain by its regulation of eosinophil-mediated inflammation through the ALOX15-15-HETE-PE axis, which activates ERK and the downstream IL-4R α signaling pathway controlling eosinophil trafficking to the sites of allergic inflammation.⁶⁵ However, it should be noted that the targets of the metabolites produced by ALOX15 as well as the mechanisms regulating the expression of pro-inflammatory chemokines and airway remodeling-related transcription factors are still unclear. Therefore, future studies are needed to determine whether, in addition to the ERK pathway, other signaling mechanisms are regulated by ALOX15. Although GWAS studies on ALOX15 have revealed exciting data, they have mainly been conducted on the European population, and it is unclear whether the effects of the identified ALOX15 variants on airway diseases are race-specific; therefore, further research is needed to determine the clinical significance of ALOX15 mutations in different racial groups. Limited pharmacological

agents targeting the ALOX15 pathway have been approved for use so far. However, previously discovered protective loss-of-function variants such as proprotein convertase subtilisin/kexin type 9 (PCSK9) have been successfully used in clinical practice and have guided further drug development studies.⁶⁶ Leukotriene inhibitors are a class of drugs that act by inhibiting the action of 5-lipoxygenase (5-LOX), a member of the lipoxygenase family highly similar to ALOX15, which was approved for use in asthma and allergic rhinitis.⁶⁷ These data show that the compounds targeting ALOX15 present a new strategy for the treatment of allergic diseases, providing a foundation for the development of novel anti-allergy drugs, taking into consideration the genomic data. Furthermore, the findings of Forno *et al.*⁶⁸ support the feasibility of using the nasal methylome for future clinical applications, such as predicting the development of asthma among wheezing infants. The epigenetic modifications of ALOX15, which may serve as markers in the diagnosis and risk prediction of airway inflammatory diseases, should be further investigated. The development and testing of ALOX15-targeting drugs for therapeutic intervention in airway inflammatory diseases is the focus of our future work. Extensive studies on the genetic and epigenetic mechanisms would yield valuable information about the clinical potential and enhance our understanding of the etiology of airway inflammatory diseases.

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REFERENCES

1. Huang K, Yang T, Xu J, Yang L, Zhao J, Zhang X, et al. Prevalence, risk factors, and management of asthma in China: a national cross-sectional study. *Lancet* 2019;394:407-18.
[PUBMED](#) | [CROSSREF](#)
2. Wang XD, Zheng M, Lou HF, Wang CS, Zhang Y, Bo MY, et al. An increased prevalence of self-reported allergic rhinitis in major Chinese cities from 2005 to 2011. *Allergy* 2016;71:1170-80.
[PUBMED](#) | [CROSSREF](#)
3. Zhu B, Wang Y, Ming J, Chen W, Zhang L. Disease burden of COPD in China: a systematic review. *Int J Chron Obstruct Pulmon Dis* 2018;13:1353-64.
[PUBMED](#) | [CROSSREF](#)
4. Kuhn H, Humeniuk L, Kozlov N, Roigas S, Adel S, Heydeck D. The evolutionary hypothesis of reaction specificity of mammalian ALOX15 orthologs. *Prog Lipid Res* 2018;72:55-74.
[PUBMED](#) | [CROSSREF](#)
5. Ivanov I, Kuhn H, Heydeck D. Structural and functional biology of arachidonic acid 15-lipoxygenase-1 (ALOX15). *Gene* 2015;573:1-32.
[PUBMED](#) | [CROSSREF](#)
6. Singh NK, Rao GN. Emerging role of 12/15-lipoxygenase (ALOX15) in human pathologies. *Prog Lipid Res* 2019;73:28-45.
[PUBMED](#) | [CROSSREF](#)

7. Markworth JF, Mitchell CJ, D'Souza RF, Aasen KMM, Durainayagam BR, Mitchell SM, et al. Arachidonic acid supplementation modulates blood and skeletal muscle lipid profile with no effect on basal inflammation in resistance exercise trained men. *Prostaglandins Leukot Essent Fatty Acids* 2018;128:74-86.
[PUBMED](#) | [CROSSREF](#)
8. Kutzner L, Goloshchapova K, Heydeck D, Stehling S, Kuhn H, Schebb NH. Mammalian ALOX15 orthologs exhibit pronounced dual positional specificity with docosahexaenoic acid. *Biochim Biophys Acta Mol Cell Biol Lipids* 2017;1862:666-75.
[PUBMED](#) | [CROSSREF](#)
9. Ramon S, Baker SF, Sahler JM, Kim N, Feldsott EA, Serhan CN, et al. The specialized proresolving mediator 17-HDHA enhances the antibody-mediated immune response against influenza virus: a new class of adjuvant? *J Immunol* 2014;193:6031-40.
[PUBMED](#) | [CROSSREF](#)
10. Kumar N, Gupta G, Anilkumar K, Fatima N, Karnati R, Reddy GV, et al. 15-Lipoxygenase metabolites of α -linolenic acid, [13-(S)-HPOTrE and 13-(S)-HOTrE], mediate anti-inflammatory effects by inactivating NLRP3 inflammasome. *Sci Rep* 2016;6:31649.
[PUBMED](#) | [CROSSREF](#)
11. Rapoport SM, Schewe T, Wiesner R, Halangk W, Ludwig P, Janicke-Höhne M, et al. The lipoxygenase of reticulocytes. Purification, characterization and biological dynamics of the lipoxygenase; its identity with the respiratory inhibitors of the reticulocyte. *Eur J Biochem* 1979;96:545-61.
[PUBMED](#) | [CROSSREF](#)
12. Chen M, Yang ZD, Smith KM, Carter JD, Nadler JL. Activation of 12-lipoxygenase in proinflammatory cytokine-mediated beta cell toxicity. *Diabetologia* 2005;48:486-95.
[PUBMED](#) | [CROSSREF](#)
13. Rothe T, Gruber F, Uderhardt S, Ipseiz N, Rössner S, Oskolkova O, et al. 12/15-Lipoxygenase-mediated enzymatic lipid oxidation regulates DC maturation and function. *J Clin Invest* 2015;125:1944-54.
[PUBMED](#) | [CROSSREF](#)
14. Quan MY, Song XJ, Liu HJ, Deng XH, Hou HQ, Chen LP, et al. Amlexanox attenuates experimental autoimmune encephalomyelitis by inhibiting dendritic cell maturation and reprogramming effector and regulatory T cell responses. *J Neuroinflammation* 2019;16:52.
[PUBMED](#) | [CROSSREF](#)
15. Brunnström Å, Tryselius Y, Feltenmark S, Andersson E, Leksell H, James A, et al. On the biosynthesis of 15-HETE and eoxin C4 by human airway epithelial cells. *Prostaglandins Other Lipid Mediat* 2015;121:83-90.
[PUBMED](#) | [CROSSREF](#)
16. Hill EM, Eling T, Nettesheim P. Changes in expression of 15-lipoxygenase and prostaglandin-H synthase during differentiation of human tracheobronchial epithelial cells. *Am J Respir Cell Mol Biol* 1998;18:662-9.
[PUBMED](#) | [CROSSREF](#)
17. Zhao J, O'Donnell VB, Balzar S, St Croix CM, Trudeau JB, Wenzel SE. 15-Lipoxygenase 1 interacts with phosphatidylethanolamine-binding protein to regulate MAPK signaling in human airway epithelial cells. *Proc Natl Acad Sci U S A* 2011;108:14246-51.
[PUBMED](#) | [CROSSREF](#)
18. Yeung K, Seitz T, Li S, Janosch P, McFerran B, Kaiser C, et al. Suppression of Raf-1 kinase activity and MAP kinase signalling by RKIP. *Nature* 1999;401:173-7.
[PUBMED](#) | [CROSSREF](#)
19. Xu B, Bhattacharjee A, Roy B, Xu HM, Anthony D, Frank DA, et al. Interleukin-13 induction of 15-lipoxygenase gene expression requires p38 mitogen-activated protein kinase-mediated serine 727 phosphorylation of Stat1 and Stat3. *Mol Cell Biol* 2003;23:3918-28.
[PUBMED](#) | [CROSSREF](#)
20. Brash AR, Boeglin WE, Chang MS. Discovery of a second 15S-lipoxygenase in humans. *Proc Natl Acad Sci U S A* 1997;94:6148-52.
[PUBMED](#) | [CROSSREF](#)
21. Snodgrass RG, Brüne B. Regulation and functions of 15-lipoxygenases in human macrophages. *Front Pharmacol* 2019;10:719.
[PUBMED](#) | [CROSSREF](#)
22. Snodgrass RG, Zezina E, Namgaladze D, Gupta S, Angioni C, Geisslinger G, et al. A novel function for 15-lipoxygenases in cholesterol homeostasis and CCL17 production in human macrophages. *Front Immunol* 2018;9:1906.
[PUBMED](#) | [CROSSREF](#)
23. Profita M, Sala A, Riccobono L, Paternò A, Mirabella A, Bonanno A, et al. 15-Lipoxygenase expression and 15(S)-hydroxyeicosatetraenoic acid release and reincorporation in induced sputum of asthmatic subjects. *J Allergy Clin Immunol* 2000;105:711-6.
[PUBMED](#) | [CROSSREF](#)

24. Chu HW, Balzar S, Westcott JY, Trudeau JB, Sun Y, Conrad DJ, et al. Expression and activation of 15-lipoxygenase pathway in severe asthma: relationship to eosinophilic phenotype and collagen deposition. *Clin Exp Allergy* 2002;32:1558-65.
[PUBMED](#) | [CROSSREF](#)
25. Mabalirajan U, Rehman R, Ahmad T, Kumar S, Leishangthem GD, Singh S, et al. 12/15-Lipoxygenase expressed in non-epithelial cells causes airway epithelial injury in asthma. *Sci Rep* 2013;3:1540.
[PUBMED](#) | [CROSSREF](#)
26. Ordovas-Montanes J, Dwyer DF, Nyquist SK, Buchheit KM, Vukovic M, Deb C, et al. Allergic inflammatory memory in human respiratory epithelial progenitor cells. *Nature* 2018;560:649-54.
[PUBMED](#) | [CROSSREF](#)
27. Sridhar S, Liu H, Pham TH, Damera G, Newbold P. Modulation of blood inflammatory markers by benralizumab in patients with eosinophilic airway diseases. *Respir Res* 2019;20:14.
[PUBMED](#) | [CROSSREF](#)
28. Kolsum U, Ravi A, Hitchen P, Maddi S, Southworth T, Singh D. Clinical characteristics of eosinophilic COPD versus COPD patients with a history of asthma. *Respir Res* 2017;18:73.
[PUBMED](#) | [CROSSREF](#)
29. Wenzel SE. Emergence of biomolecular pathways to define novel asthma phenotypes. Type-2 immunity and beyond. *Am J Respir Cell Mol Biol* 2016;55:1-4.
[PUBMED](#) | [CROSSREF](#)
30. Choy DF, Jia G, Abbas AR, Morshead KB, Lewin-Koh N, Dua R, et al. Peripheral blood gene expression predicts clinical benefit from anti-IL-13 in asthma. *J Allergy Clin Immunol* 2016;138:1230-1233.e8.
[PUBMED](#) | [CROSSREF](#)
31. Kolmert J, Piñeiro-Hermida S, Hamberg M, Gregory JA, López IP, Fauland A, et al. Prominent release of lipoxygenase generated mediators in a murine house dust mite-induced asthma model. *Prostaglandins Other Lipid Mediat* 2018;137:20-9.
[PUBMED](#) | [CROSSREF](#)
32. Schleimer RP. Immunopathogenesis of chronic rhinosinusitis and nasal polyposis. *Annu Rev Pathol* 2017;12:331-57.
[PUBMED](#) | [CROSSREF](#)
33. Liao B, Liu JX, Li ZY, Zhen Z, Cao PP, Yao Y, et al. Multidimensional endotypes of chronic rhinosinusitis and their association with treatment outcomes. *Allergy* 2018;73:1459-69.
[PUBMED](#) | [CROSSREF](#)
34. Pérez-Novo CA, Watelet JB, Claeys C, Van Cauwenberge P, Bachert C. Prostaglandin, leukotriene, and lipoxin balance in chronic rhinosinusitis with and without nasal polyposis. *J Allergy Clin Immunol* 2005;115:1189-96.
[PUBMED](#) | [CROSSREF](#)
35. Li Z, Zeng M, Deng Y, Zhao J, Zhou X, Trudeau JB, et al. 15-Lipoxygenase 1 in nasal polyps promotes CCL26/eotaxin 3 expression through extracellular signal-regulated kinase activation. *J Allergy Clin Immunol* 2019;144:1228-1241.e9.
[PUBMED](#) | [CROSSREF](#)
36. Sanak M, Levy BD, Clish CB, Chiang N, Gronert K, Mastalerz L, et al. Aspirin-tolerant asthmatics generate more lipoxins than aspirin-intolerant asthmatics. *Eur Respir J* 2000;16:44-9.
[PUBMED](#) | [CROSSREF](#)
37. Kowalski ML, Ptasińska A, Bienkiewicz B, Pawliczak R, DuBuske L. Differential effects of aspirin and misoprostol on 15-hydroxyeicosatetraenoic acid generation by leukocytes from aspirin-sensitive asthmatic patients. *J Allergy Clin Immunol* 2003;112:505-12.
[PUBMED](#) | [CROSSREF](#)
38. Neighbour H. Mechanisms of aspirin-intolerant asthma: identifying inflammatory pathways in the pathogenesis of asthma. *Int Arch Allergy Immunol* 2014;163:1-2.
[PUBMED](#) | [CROSSREF](#)
39. Stevens WW, Staudacher AG, Hulse KE, Carter RG, Winter DR, Abdala-Valencia H, et al. Activation of the 15-lipoxygenase pathway in aspirin-exacerbated respiratory disease. *J Allergy Clin Immunol* 2021;147:600-12.
[PUBMED](#) | [CROSSREF](#)
40. Li Z, Zeng M, Deng Y, Zhao J, Zhou X, Trudeau JB, et al. 15-Lipoxygenase 1 in nasal polyps promotes CCL26/eotaxin 3 expression through extracellular signal-regulated kinase activation. *J Allergy Clin Immunol* 2019;144:1228-1241.e9.
[PUBMED](#) | [CROSSREF](#)
41. Provost V, Larose MC, Langlois A, Rola-Pleszczynski M, Flamand N, Laviolette M. CCL26/eotaxin-3 is more effective to induce the migration of eosinophils of asthmatics than CCL11/eotaxin-1 and CCL24/eotaxin-2. *J Leukoc Biol* 2013;94:213-22.
[PUBMED](#) | [CROSSREF](#)

42. Liu C, Xu D, Liu L, Schain F, Brunnström A, Björkholm M, et al. 15-Lipoxygenase-1 induces expression and release of chemokines in cultured human lung epithelial cells. *Am J Physiol Lung Cell Mol Physiol* 2009;297:L196-203.
[PUBMED](#) | [CROSSREF](#)
43. Kuhar HN, Tajudeen BA, Mahdavinia M, Gattuso P, Ghai R, Batra PS. Inflammatory infiltrate and mucosal remodeling in chronic rhinosinusitis with and without polyps: structured histopathologic analysis. *Int Forum Allergy Rhinol* 2017;7:679-89.
[PUBMED](#) | [CROSSREF](#)
44. Zhao J, Maskrey B, Balzar S, Chibana K, Mustovich A, Hu H, et al. Interleukin-13-induced MUC5AC is regulated by 15-lipoxygenase 1 pathway in human bronchial epithelial cells. *Am J Respir Crit Care Med* 2009;179:782-90.
[PUBMED](#) | [CROSSREF](#)
45. Li W, Gao P, Zhi Y, Xu W, Wu Y, Yin J, et al. Periostin: its role in asthma and its potential as a diagnostic or therapeutic target. *Respir Res* 2015;16:57.
[PUBMED](#) | [CROSSREF](#)
46. Johansson MW, Annis DS, Mosher DF. $\alpha_M\beta_2$ integrin-mediated adhesion and motility of IL-5-stimulated eosinophils on periostin. *Am J Respir Cell Mol Biol* 2013;48:503-10.
[PUBMED](#) | [CROSSREF](#)
47. Zhao J, Minami Y, Etling E, Coleman JM, Lauder SN, Tyrrell V, et al. Preferential generation of 15-HETE-PE induced by IL-13 regulates goblet cell differentiation in human airway epithelial cells. *Am J Respir Cell Mol Biol* 2017;57:692-701.
[PUBMED](#) | [CROSSREF](#)
48. Wan H, Kaestner KH, Ang SL, Ikegami M, Finkelman FD, Stahlman MT, et al. Foxa2 regulates alveolarization and goblet cell hyperplasia. *Development* 2004;131:953-64.
[PUBMED](#) | [CROSSREF](#)
49. Yan B, Wang Y, Li Y, Wang C, Zhang L. Inhibition of arachidonate 15-lipoxygenase reduces the epithelial-mesenchymal transition in eosinophilic chronic rhinosinusitis with nasal polyps. *Int Forum Allergy Rhinol* 2019;9:270-80.
[PUBMED](#) | [CROSSREF](#)
50. Wittwer J, Marti-Jaun J, Hersberger M. Functional polymorphism in ALOX15 results in increased allele-specific transcription in macrophages through binding of the transcription factor SPI1. *Hum Mutat* 2006;27:78-87.
[PUBMED](#) | [CROSSREF](#)
51. Fairfax BP, Vannberg FO, Radhakrishnan J, Hakonarson H, Keating BJ, Hill AV, et al. An integrated expression phenotype mapping approach defines common variants in LEP, ALOX15 and CAPNS1 associated with induction of IL-6. *Hum Mol Genet* 2010;19:720-30.
[PUBMED](#) | [CROSSREF](#)
52. Tremblay K, Daley D, Chamberland A, Lemire M, Montpetit A, Laviolette M, et al. Genetic variation in immune signaling genes differentially expressed in asthmatic lung tissues. *J Allergy Clin Immunol* 2008;122:529-36.e17.
[PUBMED](#) | [CROSSREF](#)
53. Ferreira MA, Vonk JM, Baurecht H, Marenholz I, Tian C, Hoffman JD, et al. Shared genetic origin of asthma, hay fever and eczema elucidates allergic disease biology. *Nat Genet* 2017;49:1752-7.
[PUBMED](#) | [CROSSREF](#)
54. Kristjansson RP, Benonisdottir S, Davidsson OB, Oddsson A, Tragante V, Sigurdsson JK, et al. A loss-of-function variant in ALOX15 protects against nasal polyps and chronic rhinosinusitis. *Nat Genet* 2019;51:267-76.
[PUBMED](#) | [CROSSREF](#)
55. Schurmann K, Anton M, Ivanov I, Richter C, Kuhn H, Walther M. Molecular basis for the reduced catalytic activity of the naturally occurring T560M mutant of human 12/15-lipoxygenase that has been implicated in coronary artery disease. *J Biol Chem* 2011;286:23920-7.
[PUBMED](#) | [CROSSREF](#)
56. Assimes TL, Knowles JW, Priest JR, Basu A, Borchert A, Volcik KA, et al. A near null variant of 12/15-LOX encoded by a novel SNP in ALOX15 and the risk of coronary artery disease. *Atherosclerosis* 2008;198:136-44.
[PUBMED](#) | [CROSSREF](#)
57. Yang IV, Pedersen BS, Liu AH, O'Connor GT, Pillai D, Kattan M, et al. The nasal methylome and childhood atopic asthma. *J Allergy Clin Immunol* 2017;139:1478-88.
[PUBMED](#) | [CROSSREF](#)
58. Han H, Xu D, Liu C, Claesson HE, Björkholm M, Sjöberg J. Interleukin-4-mediated 15-lipoxygenase-1 trans-activation requires UTX recruitment and H3K27me3 demethylation at the promoter in A549 cells. *PLoS One* 2014;9:e85085.
[PUBMED](#) | [CROSSREF](#)

59. Zuo X, Morris JS, Shureiqi I. Chromatin modification requirements for 15-lipoxygenase-1 transcriptional reactivation in colon cancer cells. *J Biol Chem* 2008;283:31341-7.
[PUBMED](#) | [CROSSREF](#)
60. Orafaie A, Matin MM, Sadeghian H. The importance of 15-lipoxygenase inhibitors in cancer treatment. *Cancer Metastasis Rev* 2018;37:397-408.
[PUBMED](#) | [CROSSREF](#)
61. Tanaka T, Takahashi R. Flavonoids and asthma. *Nutrients* 2013;5:2128-43.
[PUBMED](#) | [CROSSREF](#)
62. Li D, Wu L. Coumarins from the roots of *Angelica dahurica* cause anti-allergic inflammation. *Exp Ther Med* 2017;14:874-80.
[PUBMED](#) | [CROSSREF](#)
63. Yang N, Zhang H, Cai X, Shang Y. Epigallocatechin-3-gallate inhibits inflammation and epithelial-mesenchymal transition through the PI3K/AKT pathway via upregulation of PTEN in asthma. *Int J Mol Med* 2018;41:818-28.
[PUBMED](#) | [CROSSREF](#)
64. Hua S, Liu F, Wang M. Emodin alleviates the airway inflammation of cough variant asthma in mice by regulating the notch pathway. *Med Sci Monit* 2019;25:5621-9.
[PUBMED](#) | [CROSSREF](#)
65. Spencer LA, Melo RC, Perez SA, Bafford SP, Dvorak AM, Weller PF. Cytokine receptor-mediated trafficking of preformed IL-4 in eosinophils identifies an innate immune mechanism of cytokine secretion. *Proc Natl Acad Sci U S A* 2006;103:3333-8.
[PUBMED](#) | [CROSSREF](#)
66. Jaworski K, Jankowski P, Kosior DA. PCSK9 inhibitors - from discovery of a single mutation to a groundbreaking therapy of lipid disorders in one decade. *Arch Med Sci* 2017;13:914-29.
[PUBMED](#) | [CROSSREF](#)
67. Steinke JW, Culp JA. Leukotriene synthesis inhibitors versus antagonists: the pros and cons. *Curr Allergy Asthma Rep* 2007;7:126-33.
[PUBMED](#) | [CROSSREF](#)
68. Forno E, Wang T, Qi C, Yan Q, Xu CJ, Boutaoui N, et al. DNA methylation in nasal epithelium, atopy, and atopic asthma in children: a genome-wide study. *Lancet Respir Med* 2019;7:336-46.
[PUBMED](#) | [CROSSREF](#)