

Determination of H⁺-ATPase Activity in *Arabidopsis* Guard Cell Protoplasts through H⁺-pumping Measurement and H⁺-ATPase Quantification

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[Abstract] The opening of stomata in plants in response to blue light is driven by the plasma membrane H⁺-ATPase in guard cells. To evaluate the activation of the H⁺-ATPase *in vivo*, we can use H⁺-pumping by guard cells in response to blue light and fusicoccin. To do this, it is required to prepare a large amount of guard cell protoplasts and measure H⁺-pumping in the protoplasts. It is also necessary to determine the protein amount of H⁺-ATPase. In this protocol, we describe the procedures required for these preparations and measurements.

Keywords: *Arabidopsis thaliana*, Blue light, Fusicoccin, H⁺-pumping, Plasma membrane H⁺-ATPase

[Background] The opening of stomata in response to blue light is driven by membrane hyperpolarization mediated through H⁺-pumping across the plasma membrane in guard cells (Assmann *et al.*, 1985; Shimazaki *et al.*, 1986), and is brought about by the plasma membrane H⁺-ATPase (Kinoshita and Shimazaki, 1999). The H⁺-ATPase generates an electrochemical gradient across the membrane, and provides the energy required for numerous secondary transports in plant cells. However, it is not easy to measure the activity of H⁺-ATPase *in vivo*. Taking advantage of the blue light-sensitive properties of guard cells, our method makes it possible to measure H⁺-pumping as an *in vivo* H⁺-ATPase activity using *Arabidopsis* guard cell protoplasts (Ueno *et al.*, 2005). Together with H⁺-ATPase quantification by Western blot (Yamauchi *et al.*, 2016), this method allows comparing H⁺-ATPase activity under different conditions or mutant backgrounds.

Materials and Reagents

1. Planter (640 x 230 x 185 mm) (Iris Oyama E-type, PR650EMK)
2. Vermiculite (particle size 3-6 mm)
3. Soil (potting soil containing peat moss)
4. Cellophanes (Asahi KASEI)
5. Nylon mesh (10 μm, 25 μm, 58 μm and 94 μm) (Kyoshin Riko)
6. Micro slide glass (Matsunami Glass, catalog number: S011260)
7. Glass test tube (10 ml) (IWAKI, catalog number: TEST15-105NP)
8. Gel loading tip (Thermo Fisher Scientific, Thermo Scientific, catalog number: 010-Q)

9. Sample tube (1.5 ml) (INA•OPTIKA, catalog number: ST-0105F)
10. Micro cuvette (100 μ l) (Beckman Coulter, catalog number: 523270)
Note: This product has been discontinued. Any micro cuvette (100 μ l) that can be fixed in the spectrophotometer can be substituted.
11. Blender cup (100 ml) (Waring Lab, catalog number: MC2)
12. Glass pipette (10 ml) (Sansyo, catalog number: 73-0045)
13. Glass centrifuge tube (50 ml) (Sansyo, catalog number: 84-0182)
14. Orange-colored film (Kodak, Cinemoid 5A)
15. Filter paper (3MM CHR CHROMATOGRAPHY PAPER, GE Healthcare, catalog number: 3030-928)
16. *Arabidopsis thaliana* ecotype Col-0
17. *Arabidopsis thaliana* mutant *aha1-10*
18. Bradford protein assay dye reagent concentrate (Bio-Rad Laboratories, catalog number: 5000006JA)
19. Aprotinin (Merck, Calbiochem, catalog number: 616399)
20. Protease inhibitor cocktail set III (Merck, Calbiochem, catalog number: 539134)
21. MG132 (Sigma-Aldrich, catalog number: M8699-1MG)
22. Trichloroacetic acid (TCA) (NACALAI TESQUE, catalog number: 06275-24)
23. Anti-rabbit IgG HRP (Bio-Rad Laboratories, catalog number: 1706515)
24. Anti H⁺-ATPase antibodies (Kinoshita and Shimazaki, 1999) or Agrisera anti H⁺-ATPase antibodies (Agrisera, catalog number: AS07 260)
25. Clarity Western ECL substrate (Bio-Rad Laboratories, catalog number: 1705061)
26. Cellulase R-10 (Yakult, Pharmaceutical Industry, Tokyo, Japan)
27. Macerozyme R-10 (Yakult, Pharmaceutical Industry, Tokyo, Japan)
28. Polyvinylpyrrolidone K 30 (PVP-K30) (NACALAI TESQUE, catalog number: 28314-95)
29. Bovine serum albumin (BSA) (Thermo Fisher Scientific, Gibco™, catalog number: 30036727)
30. Mannitol (Wako Pure Chemical Industries, catalog number: 130-00855)
31. Calcium chloride dihydrate (CaCl₂·2H₂O) (Wako Pure Chemical Industries, catalog number: 031-00435)
32. 2-(N-morpholino) ethanesulfonic acid (MES) (NACALAI TESQUE, catalog number: 02442-44)
33. Cellulase RS (Yakult, Pharmaceutical Industry, Tokyo, Japan)
34. Hydrochloric acid (HCl) (Wako Pure Chemical Industries, catalog number: 081-03475)
35. Potassium chloride (KCl) (NACALAI TESQUE, catalog number: 28513-85)
36. Fusicoccin (Fc) (Sigma-Aldrich, catalog number: F0537-1MG)
37. DMSO (Wako Pure Chemical Industries, catalog number: 046-21981)
38. Tris (hydroxymethyl) aminomethane (Wako Pure Chemical Industries, catalog number: 514-98121)
39. Sodium chloride (NaCl) (Wako Pure Chemical Industries, catalog number: 191-01665)
40. Sodium dodecyl sulfate (SDS) (NACALAI TESQUE, catalog number: 31606-75)

41. Ammonium peroxodisulfate (Wako Pure Chemical Industries, catalog number: 012-20503)
42. 30% (w/v) Acrylamide/Bis mixed solution (37.5:1) (Wako Pure Chemical Industries, catalog number: 018-25625)
43. *N,N,N',N'*-tetramethylethylenediamine (NACALAI TESQUE, catalog number: 33401-72)
44. Dithiothreitol (DTT) (NACALAI TESQUE, catalog number: 14112-94)
45. *N*^α-Tosyl-Lys chloromethyl ketone (Merck, Calbiochem, catalog number: 616382)
46. Ponceau S (NACALAI TESQUE, Catalog number: 28322-72)
47. Acetic acid (Wako Pure Chemical Industries, catalog number: 017-00251)
48. Tween 20 (MP Biomedicals, catalog number: 0210316890)
49. Skim milk (Snow Brand Milk Products, Ltd.)
50. Glycine (Wako Pure Chemical Industries, catalog number: 077-00735)
51. Methanol (Wako Pure Chemical Industries, catalog number: 137-01823)
52. Ethylenediaminetetraacetate acid (EDTA) (Wako Pure Chemical Industries, catalog number: 343-01861)
53. Sucrose (Wako Pure Chemical Industries, catalog number: 196-00015)
54. Coomassie Brilliant Blue G-250 (NACALAI TESQUE, catalog number: 09409-42)
55. 2-Mercaptoethanol (NACALAI TESQUE, catalog number: 21417-52)
56. 1st digestion solution (see Recipes)
57. 2nd digestion solution (see Recipes)
58. 2x H⁺-pumping buffer (see Recipes)
59. Reaction mixture for H⁺ pumping (see Recipes)
60. Fc stock solution (see Recipes)
61. Reaction mixture for inhibitor treatment (see Recipes)
62. HCl stock solution (see Recipes)
63. 10x TBS (see Recipes)
64. 4x separation buffer (see Recipes)
65. 4x stacking buffer (see Recipes)
66. Ammonium peroxodisulfate stock solution (see Recipes)
67. 10% acrylamide gel (see Recipes)
68. DTT stock solution (see Recipes)
69. TLCK stock solution (see Recipes)
70. Ponceau S (see Recipes)
71. Tween 20 stock solution (see Recipes)
72. Blocking buffer (see Recipes)
73. T-TBS (see Recipes)
74. Transfer buffer (see Recipes)
75. 3x SDS buffer (see Recipes)
76. 1x SDS sample buffer (see Recipes)

Equipment

1. Metal shelf made by stainless steel (120 x 61 x 230cm) (Iris Oyama, MR-230P)
2. Blender (Waring Lab, model: 51BL31, catalog number: 7011BU)*
3. Funnel (TGK, catalog number: 416-09-24-04)*
4. Erlenmeyer flask (500 ml) (IWAKI, catalog number: 82-0088)
5. Erlenmeyer flask (300 ml) (IWAKI, catalog number: 82-0087)
6. Refrigerated centrifuge (TOMY, model: AX-310)*
7. Refrigerated centrifuge (TOMY, model: MX-100)*
8. Incubator with shaker (TAITEC, model: Personal Lt10)
9. Unit thermostat bath (TAITEC, THERMO MINDER model: JR 80)*
10. Spectrophotometer (Beckman Coulter, model: DU800)*
11. pH glass electrode (Beckman Coulter, catalog number: 39532)*
12. pH meter (Beckman Coulter, model: Φ71 pH meter)*
13. Chart recorder (YOKOGAWA ELECTRIC WORKS, model: 3066)*
14. Magnetic stirrer (RANK BROTHERS, model: Model 300)
15. Chiller bath circulator (Thermo Fisher Scientific, Thermo Scientific™, model: RTE7)
16. LED panel (CCS, model: ISL 150X150 H4RRHB, Made by order)

Note: LEDs for both red and blue are fixed on the same panel, and each LED can be turned on and off by the power supply.

17. LED power supply (CCS, model: ISC-201-2)
18. Microscope (Nikon Instruments, model: Eclipse TS100)
19. Pipetman (P200)
20. Power supply (Bio-Rad Laboratories, model: PowerPac™ HC High-Current Power Supply)
21. Transfer Cell (Bio-Rad Laboratories, model: Trans-Blot® SD Semi-Dry Transfer Cell)
22. Electrophoresis vessel (ATTO, model: AE6200, catalog number: 2392385)*
23. Chemical Luminescence analyzer (Bio-Rad Laboratories, model: ChemiDoc™ Touch Imaging System)
24. Gel cassette (ATTO, model: AE-6210)

**Note: This product has been discontinued.*

Software

1. ImageJ

Procedure

- A. Preparation of guard cell protoplasts from *Arabidopsis* leaves
1. Sow 200 *Arabidopsis* seeds per planter on soil and vermiculite mixture (1:1).
 2. Grow *Arabidopsis* plants for 4- to 6-weeks under white light ($60 \mu\text{mol m}^{-2} \text{sec}^{-1}$) with 14/10 h light/dark cycle at 24 °C (see Figure 1).



Figure 1. Four weeks old *Arabidopsis* grown under white light. *Arabidopsis* plants were grown for 4-weeks under white light ($60 \mu\text{mol m}^{-2} \text{sec}^{-1}$) with 14/10 h light/dark cycle at 24 °C. The bar represents 1 cm.

3. Harvest the fully expanded rosette leaves from the 200 plants and remove the main petioles (~2 cm-length, see Figure 2) before bolting.

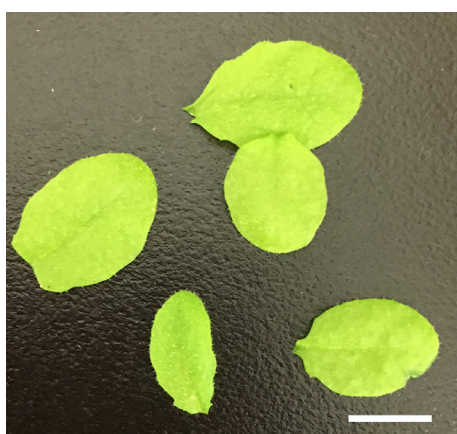


Figure 2. Fully expanded leaves from 4 weeks old *Arabidopsis*. Fully expanded leaves were harvested from *Arabidopsis* and main petioles were removed before bolting. The bar represents 1 cm.

- Place the leaves (25 g) in ice-cold water (see Figure 3).



Figure 3. Leaves in ice-cold water. Harvested leaves were put into a glass beaker filled with ice-cold water.

- Move the ice-cold leaves into the cup of Waring blender (see Figure 4A). Homogenize the leaves in the blender covered with two layers of cellophane (see Figure 4B) at full speed for 45 sec in 70 ml ice-cold distilled water. Pour the resulting homogenate onto the nylon mesh placed on a funnel (see Figure 5). Discard the filtrate (flow-through) throughout the protocol.

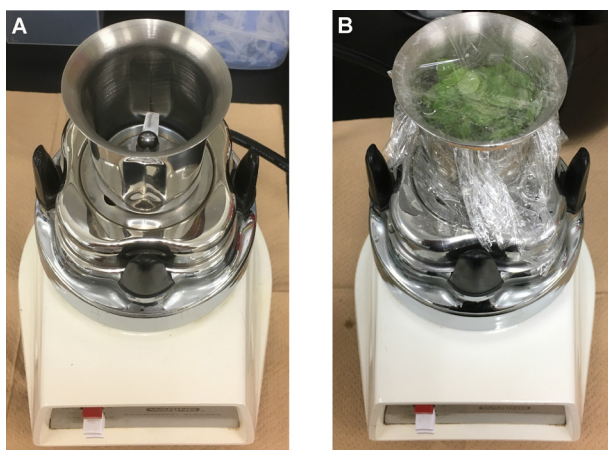


Figure 4. Waring blender with blender cup. A. Picture of Waring blender with blender cup; B. Fully expanded rosette leaves were put into blender cup filled with ice-cold water. Blender cup was sealed with cellophane. Leaves were homogenized at maximum speed for 45 sec.

- Transfer the epidermal tissues retained on the 58 μ m mesh (see Figure 5) into the cup of Waring blender by pouring ice-cold distilled water (see Video 1) and subject them to homogenization again for 1 min in 70 ml water at full speed, and collect them by filtering through the 58 μ m nylon mesh as above. The nylon mesh can be reused after cleaning with distilled water.

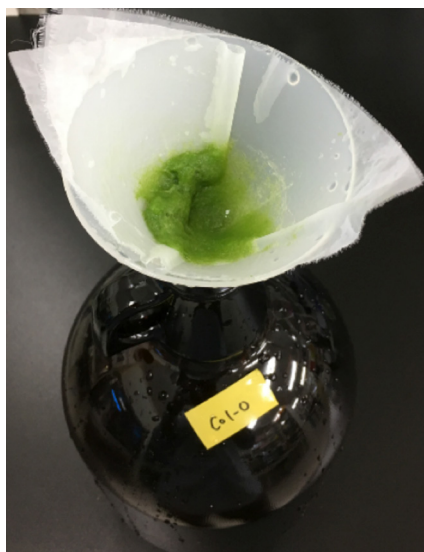


Figure 5. Epidermal tissues collected with layers of 58 μ m nylon mesh. Homogenized epidermal tissues were collected with layers of 58 μ m nylon mesh after 45 sec homogenization. Collected tissues were placed into the blender cup again and homogenized for 1 min (see Video 1). The epidermal tissues were collected again with the 58 μ m nylon mesh and resuspended in the 1st digestion solution.



Video 1. Transfer the epidermal fragments by pouring the solution onto the fragments on the mesh using a wash bottle (or 10 ml Komagome pipette, not shown here)

7. Transfer 12-13 g of the wet epidermal fragments on the mesh (see Figure 7) into a 500 ml Erlenmeyer flask by pouring the 1st digestion solution (see Recipes) using a wash bottle or a 10 ml Komagome pipette (see Figure 6 and Video 1). Adjust the total volume of the digestion solution to 100 ml by adding the same solution.



Figure 6. Komagome pipette (10 ml) with silicone rubber bulb

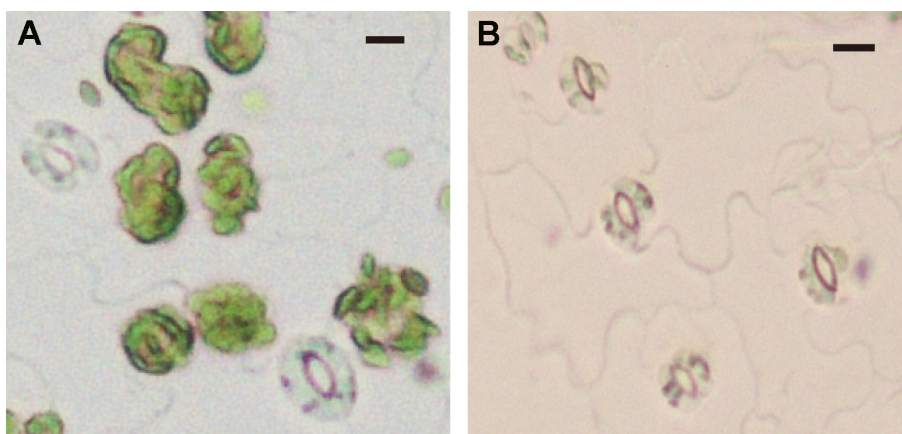


Figure 7. Mesophyll cell removal by 1st digestion solution. A. Epidermal fragment before 1st digestion solution treatment. Mesophyll cells adhered to epidermal tissue. B. Epidermal fragment after 1st digestion solution treatment. All mesophyll cells are detached from the epidermal cells. The bars represent 10 µm.

8. Pass the epidermal fragments gently into 100 ml of 1st digestion solution in the Erlenmeyer flask through the same Komagome pipette several times up and down to thoroughly mix the suspension and to ensure homogeneous digestion (see Video 2).



Video 2. Pipetting the partially digested epidermal fragments to remove the mesophyll cells using Komagome pipette

9. Incubate the suspension in the Erlenmeyer flask at 24 °C for 30 min, shaking at 70 strokes min^{-1} .
10. Pipette the partially digested epidermal fragments in the 1st digestion solution 30 times up and down through Komagome pipette to remove the attached mesophyll cells (see Video 2). Inspect the mesophyll cells attached to the epidermal fragments on a micro slide glass using a microscope (150x).
11. Repeat Step A9 until all mesophyll cells attached to the epidermis are removed (see Figure 7).
12. Pass the suspension through a 58 μm nylon mesh, and keep the epidermal fragments retained on the mesh.
13. Transfer the epidermal fragments into 0.3 M mannitol and 1 mM CaCl_2 solution by pouring the solution onto the fragments on the mesh (see Video 1). Keep the fragments in the solution for 30 min to adapt them to the high osmotic pressure.
14. Pass the osmotically adapted epidermis through the 58 μm nylon mesh. Transfer the epidermis retained on the mesh into a 300 ml Erlenmeyer flask by pouring the 2nd digestion solution (see Recipes) on the epidermis using Komagome pipette (see Video 1). Adjust the total volume to 50 ml by adding the same 2nd digestion solution.

Note: Since the epidermal peels were partially digested and fragile, do not touch the epidermis in this step.

15. Incubate the epidermis in the 2nd digestion solution with 50 strokes min^{-1} for 50-60 min at 27 °C.
16. Using several epidermal fragments, inspect whether most of the guard cells have become spherical under a microscope (see Figure 8).

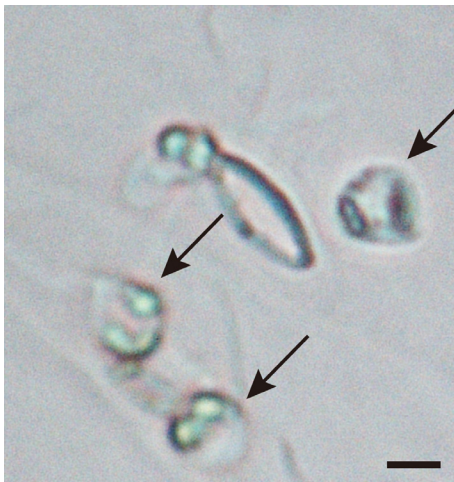


Figure 8. Guard cell spheroidization by 2nd digestion solution. Epidermal fragments were incubated with 2nd digestion solution. The spherical nature of the guard cells was examined under the microscope (Black arrows). The bar represents 10 μm .

17. Place the Erlenmeyer flask containing the digested epidermis on ice, and pipette the suspension gently up and down 60 times through a Komagome pipette to detach guard cell

- protoplasts from the epidermis (see Video 2).
18. Pass the suspension through two layers of piled nylon meshes of 94 and 25 μm (the 94 μm mesh is on top of the 25 μm one) to separate guard cell protoplasts from the epidermal remnants.
 19. Pass the separated guard cell protoplasts through two layers of 10 μm nylon mesh to remove contamination by other small tissues.
 20. Centrifuge the suspension of guard cell protoplasts at 420 x g for 18 min at 4 °C to collect guard cell protoplasts as a pellet. Discard the supernatant.
 21. Resuspend the pellet in a 50 ml solution of 0.4 M mannitol and 1 mM CaCl_2 .
 22. Repeat Steps A19 and A20 twice (total three times) to remove the residual enzymes.
 23. Resuspend the pellet of guard cell protoplasts (see Figure 9) in 200 to 500 μl of 0.4 M mannitol and 1 mM CaCl_2 in 10 ml test tube and keep them on ice.

Note: It takes 6-8 h to prepare guard cell protoplasts from 25 g leaves. The protoplasts can be stored in the refrigerator overnight under darkness until measurement. Keep the protoplasts in the dark for at least 1 h on ice for immediate measurement. The time required for one measurement is 2-3 h because it takes 1-2 h until the pH of the protoplast suspension shows a constant value under red light (RL). The measurement time for Arabidopsis is longer than for Vicia faba (Shimazaki et al., 1986).

24. Determine protein concentration by Bradford method (Bradford, 1976) using 10 μl of the protoplast suspension.

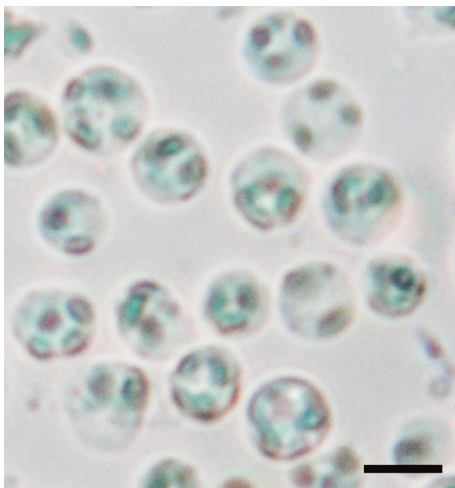


Figure 9. Guard cell protoplasts in 0.4 M mannitol and 1 mM CaCl_2 . Guard cell protoplasts isolated from *Arabidopsis*. Guard cell protoplasts were resuspended in 200 to 500 μl 0.4 M mannitol and 1 mM CaCl_2 . The typical yield of guard cell protoplasts was 4.3×10^7 cells per 5,000 leaves with a purity of 98%. The bar represents 10 μm .

B. Measurement of blue light- and fusicoccin-dependent H⁺-pumping

1. Blue light-dependent H⁺-pumping

- a. Keep guard cell protoplasts in the dark at 4 °C for 2 h or overnight.
- b. Circulate water at 24 °C in a Plexiglas water jacket to maintain the temperature. Mix X ml of guard cell protoplasts suspension (50 µg proteins) with 0.5-X ml of 0.4 M mannitol and 1 mM CaCl₂ in an open glass vessel (see Figure 10A) enclosed in the water jacket. Add 0.5 ml of 2x H⁺-pumping buffer (see Recipes) to the suspension to give a final volume of 1 ml reaction mixture in the vessel. Stir the protoplasts in the reaction mixture by a small magnetic stirrer inside the vessel.

Note: Add the protoplasts in the absence of blue light. Remove blue portion of light by covering the fluorescent lamp with orange-colored film, which prevents blue light penetration.

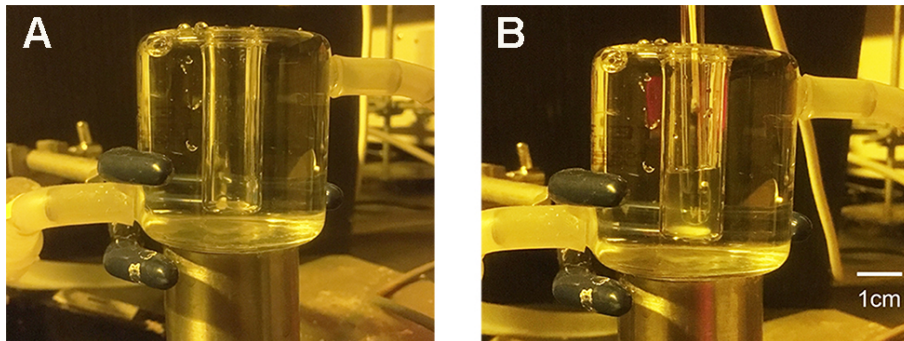


Figure 10. Open glass vessel surrounded by water jacket. Water was circulated around the vessel in the water jacket through a nylon (or silicone) tube. A. Empty glass vessel; B. pH glass electrode and guard cell protoplasts in the vessel.

- c. Place a pH glass electrode in the reaction mixture, and record the signal from the electrode on a recorder via a pH meter (see Figure 10B).
- d. Use the LED illuminator (see Figure 11) to apply red light (600 µmol m⁻² sec⁻¹) to protoplasts for 2 h by turning on the switch for the red photodiode until the pH shows an almost constant value (see Figure 13). The distance from the illuminator to the sample is about 10 cm. Regulate light intensity at the sample by the voltage controller.

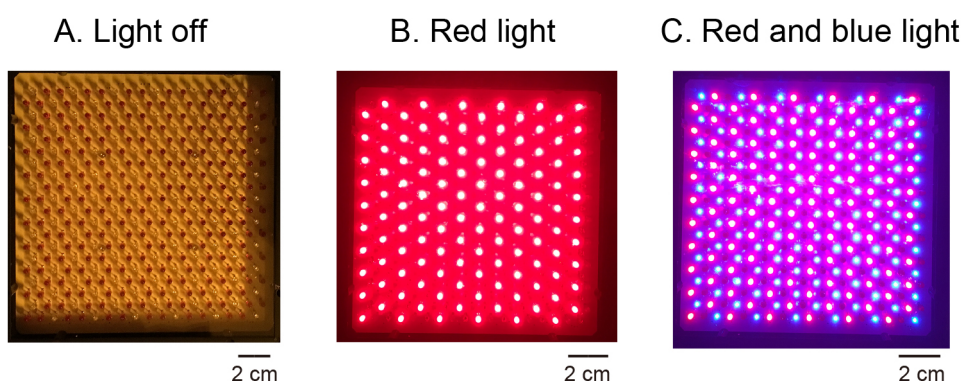


Figure 11. LED illuminator. Both red and blue LEDs were fixed on a plate. When the LEDs were turned off, all LED bulbs were seen as 242-243 dark-brown spots (A. Light off). When red LED was turned on, red-colored light could be seen (B. Red light). When both red and blue LEDs were turned on, the pink-colored light could be seen (C. Red and blue light).

- e. Use the LED illuminator (see Figure 11) to apply blue light pulse ($100 \mu\text{mol m}^{-2} \text{sec}^{-1}$) to protoplasts for 30 sec by turning on the switch for the blue photodiode superimposed on red light. Incubate the protoplasts for about 20 min under red light to follow the pH change in the mixture (Figure 13).
 - f. Add 3 or 5 μl of 1 mM HCl to the mixture to calculate the magnitude of H^+ -pumping in response to the blue light.
2. Fusicoccin (Fc)-dependent H^+ -pumping
 - a. Steps from B1a-B1f follow the same procedure as above. Use the same samples used above.
 - b. Add 2.5 μl of 4 mM Fc in DMSO to the suspension (at 10 μM) through a gel loading tip instead of the usual tip and induce H^+ -pumping. Calculate the rate after the pumping becomes constant (5 min after the addition of Fc).
- C. Determination of total H^+ -ATPase amount
1. Suspend 10 μg proteins of guard cell protoplasts in the reaction mixture (150 μl , see Recipes) in a 1.5 ml sampling tube. The concentration of guard cell proteins had been determined as per the measurement Steps A22-A23.
 2. Mix the protoplasts suspension gently by pipetting up and down.
 3. Add the inhibitors of protease or proteasome to the suspension and mix gently by pipetting up and down every 15 min.
- Inhibitors as indicated below were separately administered to the protoplasts:
- a. 10 μM aprotinin (inhibitor for serine protease) and 2 mM DTT.
 - b. 1 mM N^α -Tosyl-Lys Chloromethyl Ketone (TLCK; inhibitor for serine protease).
 - c. 1% (v/v) protease inhibitor cocktail set III (contains various protease inhibitors).
 - d. 50 μM MG132 (inhibitor of proteasome).

4. Incubate the protoplast suspension containing the inhibitors or without inhibitor as controls for 30 min at 24 °C with pipetting up and down every 15 min.
5. Add 10% trichloroacetic acid (TCA) to the suspension to cease the reaction and pipette the suspension up and down gently through PIPETMAN (P200) 10 times to denature the proteins.
6. Place the suspension on ice for 10 min to avoid excessive denaturalization.
7. Centrifuge the suspension at 17,800 x g for 10 min at 4 °C to collect the proteins.
8. Remove the supernatant and suspend the pellet with 200 µl of 50 mM Tris-HCl (pH 8.0). Repeat Steps C7-C8 twice to remove TCA.
9. Add 30 µl 1x SDS sample buffer (see Recipes) to the pellet and solubilize it.
10. Load 5 µg of the solubilized protein of guard cell protoplasts on SDS-PAGE. Separate the proteins by SDS-PAGE using a 10% acrylamide gel (see Recipes).
11. Transfer the proteins onto a nitrocellulose membrane.
 - a. Immerse the nitrocellulose membrane, the acrylamide gel that contains separated proteins, and 6 filter papers of the same size as that of the gel in 200 ml transfer buffer for 15 min.
 - b. Stack them by placing 3 layers of filter papers on the bottom, the nitrocellulose membrane, the gel, and 3 layers of filter papers on the top in the Transfer Cell.
 - c. Transfer the proteins from the gel onto the nitrocellulose membrane electrically using the Trans blot transfer cell.
12. Stain the membrane by 0.5 % Ponceau S in 1 % acetic acid for 5 sEC (see Recipes) and use as the loading control of proteins by taking a picture (see Figure 12, lower panel).

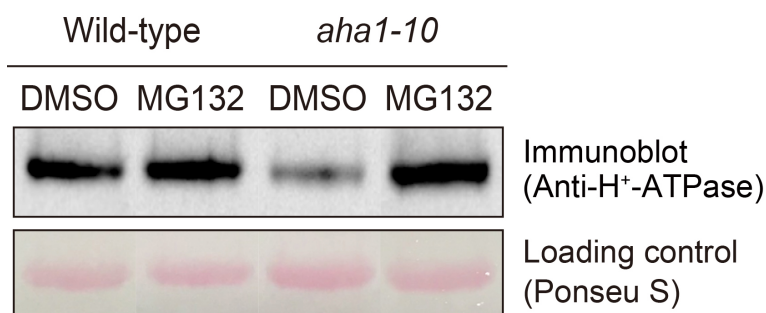


Figure 12. Determination of H⁺-ATPase by the immunological method. Immunodetection of H⁺-ATPase of guard cell after treatment with MG132 (Yamauchi *et al.*, 2016). Guard cell protoplasts from wild-type and *aha1-10* mutant were incubated with 50 µM MG132 for 30 min at 24 °C. Upper panel shows immunoblot using H⁺-ATPase antibodies. Lower panel shows Ponceau S staining as loading control. Each lane contains 5 µg guard cell protein.

13. Wash the membrane by distilled water and incubate it in the blocking buffer (see Recipes) for 30 min.
14. Incubate the membrane in the presence of anti-H⁺-ATPase antibodies (dilute 3,000x by blocking buffer) overnight at 4 °C without shaking.
15. Wash the membrane in T-TBS (see Recipes) for 10 min by incubation.

Note: Repeat this process three times.

16. Incubate the membranes in the presence of anti-rabbit IgG HRP (Bio-Rad; dilute 3,000x by blocking buffer) for 2 h at room temperature.
17. Wash the membrane by T-TBS for 10 min three times.
18. Incubate the membranes with Clarity Western ECL substrate (Bio-Rad) for 3min.
19. Detect the H⁺-ATPase by Chemical Luminescence analyzer (Bio-Rad)
20. Determine the band density on the membrane by ImageJ, and calculate the amount of the H⁺-ATPase.

Data analysis

Calculation of the magnitude and the maximum rate of H⁺-pumping (see Figure 13).

1. See Figure 13 that indicates H⁺-pumping (shown by decrease in pH over time) by guard cell protoplasts in response to a pulse of blue light (BL) under red light (RL).
2. Determine the amounts of acid equivalents by pH decrease caused by the addition of 3 nmol H⁺ (3 μ l of 1 mM HCl) to the suspension at the end of each experiment (see A). 3 nmol H⁺ corresponds to 3.2 cm on the chart recorder. Thus, 1 nmol H⁺ corresponds to 1.07 cm pH decrease on the chart.
3. Calculate the magnitude of H⁺-pumping in response to the pulse. The pH decrease is 4.20 cm per pulse (see B). The magnitude is calculated to be $4.20/1.07 = 3.93$ nmol H⁺ per pulse. Since the mixture contains 50 μ g protein, the magnitude of H⁺-pumping was calculated to be 0.079 nmol H⁺ per pulse μ g protein⁻¹.
4. Draw an approximate line at the steepest slope of H⁺-pumping, which occurs around 2.5 min after the pulse, to calculate the maximum rate of H⁺-pumping (see red line).
5. Determine the pH decrease on a chart on the basis of this line. It exhibited 4.0 cm of pH decrease on the chart in 3 min (see C).
6. Calculate the maximum rate of H⁺ pumping on the basis guard cell proteins used. Since the pH decrease per pulse is 1.33 cm min⁻¹ (4.0/3) and 1 nmol H⁺ corresponds to 1.07 cm of pH decrease, the maximum rate is calculated to be $1.33 \text{ cm}/1.07 \text{ cm} \times 60 \text{ nmol h}^{-1}/50 \mu\text{g protein}$ is 1.49 nmol H⁺ h⁻¹ μ g protein⁻¹.

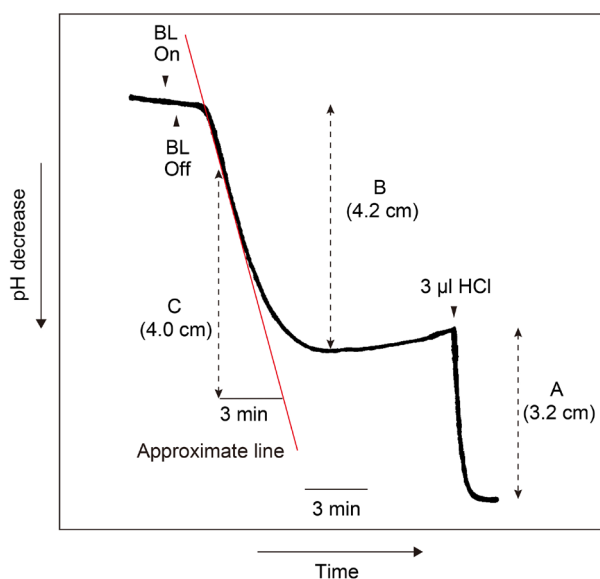


Figure 13. Time course of H⁺-pumping by guard cell protoplasts in response to blue light (BL). Guard cell protoplasts in the reaction mixture containing 50 µg proteins were illuminated with a 30 sec pulse of BL (100 µmol m⁻² sec⁻¹) superimposed on RL (600 µmol m⁻² sec⁻¹). The vertical and horizontal axes indicate pH and time, respectively. The pumping initiated a pH decrease 30 sec after the pulse and reached the maximum rate around 2-3 min after the pulse. The pumping was recorded on a chart recorder at the speed of 20 cm h⁻¹ (= 1 cm 3 min⁻¹). A: Magnitude of pH decrease by the addition of 3 nmol H⁺. B: Magnitude of H⁺-pumping in response to a pulse of BL. C: Magnitude of H⁺-pumping in 3 min in response to BL.

Notes

The H⁺-pumping activities vary with the viability of guard cell protoplasts. An important criterion to obtain viable guard cell protoplasts is enzyme quality (Cellulase RS and R-10). Activity and toxicity, which often cause reduction in protoplast recovery, vary with enzyme lot numbers. We usually check the enzyme activity on a small scale through preparation of the protoplasts and H⁺-pumping measurement. If the lot produces viable protoplasts, we purchase it in larger amounts.

Recipes

Note: Enzyme solutions for 1st and 2nd digestion should be prepared fresh.

1. 1st digestion solution
 - 0.5% (w/v) cellulase R-10
 - 0.05% (w/v) macerozyme R-10
 - 0.1% (w/v) polyvinyl pyrrolidone (PVP-K30)
 - 0.2% (w/v) BSA
 - 0.25 M mannitol

- 1 mM CaCl₂·2H₂O
- 10 mM MES-KOH (pH 5.4)
2. 2nd digestion solution
 - 1.5% (w/v) cellulase RS
 - 0.5% (w/v) macerozyme R-10
 - 0.2% (w/v) BSA
 - 0.4 M mannitol
 - 1 mM CaCl₂·2H₂O
 - 10 mM MES-KOH (pH 5.4)
 - Adjust pH to 5.4 with 2 N HCl
3. 2x H⁺-pumping buffer
 - 0.25 mM MES-KOH (pH 6.0)
 - 20 mM KCl
 - 0.4 M mannitol
 - 1 mM CaCl₂·2H₂O
 - Store at 4 °C
4. Reaction mixture for H⁺-pumping
 - 500 µl 2x H⁺-pumping buffer
 - X µl guard cell protoplasts (50-100 µg protein)
 - 500-X µl 0.4 M mannitol and 1 mM CaCl₂
5. Fc stock solution
 - 4 mM Fc in DMSO
 - Store at -20 °C
6. Reaction mixture for inhibitor treatment
 - 75 µl 2x H⁺-pumping buffer
 - X µl guard cell protoplasts (10 µg protein)
 - 75-X µl 0.4 M mannitol and 1 mM CaCl₂
7. HCl stock solution
 - 1 mM HCl
 - Store at room temperature
8. 10x TBS
 - 200 mM Tris-HCl (pH 7.4)
 - 1400 mM NaCl
 - Store at room temperature
9. 4x separation buffer
 - 1.5 M Tris-HCl (pH 8.8)
 - 4% (w/v) SDS
 - Store at room temperature

10. 4x stacking buffer
 - 1.0 M Tris-HCl (pH 6.8)
 - 4% (w/v) SDS
 - Store at room temperature
11. Ammonium peroxodisulfate stock solution
 - 10% (w/v) ammonium peroxodisulfate in distilled water
 - Store at 4 °C
12. 10% acrylamide gel
 - a. Separation gel
 - 6.7 ml Acrylamide/Bis mixed solution (30% [w/v], Wako Pure Chemical Industries)
 - 5.0 ml 4x separation buffer
 - 200 µl 1% (v/v) ammonium peroxodisulfate stock solution
 - 20 µl 0.1% (v/v) *N,N,N',N'*-tetramethylethylenediamine
 - Add distilled water to the solution until 20 ml
 - Prepare as required
 - b. Stacking gel
 - 1.67 ml acrylamide/Bis mixed solution (30% [w/v], Wako Pure Chemical Industries)
 - 2.5 ml 4x stack buffer
 - 100 µl ammonium peroxodisulfate stock solution
 - 10 µl *N,N,N',N'*-tetramethylethylenediamine
 - Add distilled water to the solution until 10 ml
 - Prepare as required
13. DTT stock solution
 - 1 M DTT in distilled water
 - Store at -20 °C
14. TLCK stock solution
 - 100 mM TLCK in DMSO
 - Store at -20 °C
15. Ponceau S
 - 0.5% (w/v) Ponceau S
 - 1% (v/v) acetic acid
 - Store at room temperature
16. Tween 20 stock solution
 - 25% (v/v) Tween 20 in 1x TBS buffer
 - Store at 4 °C
17. Blocking buffer
 - 5% (w/v) skim milk in T-TBS
 - Prepare as required

18. T-TBS
 0.5% (v/v) Tween 20 in 1x TBS
 Store at room temperature
19. Transfer buffer
 39 mM glycine
 48 mM Tris
 20% (v/v) methanol
 Store at room temperature
20. 3x SDS buffer
 30 mM Tris-HCl (pH 8.0)
 3 mM EDTA
 3% (w/v) SDS
 30% (w/v) sucrose
 Store at -20 °C
21. 1x SDS sample buffer
 10 mM Tris-HCl (pH 8.0)
 1 mM EDTA
 10% (w/v) sucrose
 1% (w/v) SDS
 0.04% (v/v) 2-mercaptoethanol
 0.003% (w/v) Coomassie Brilliant Blue G-250
 Prepare as required

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References

1. Assmann, S. M., Simoncini, L., and Schroeder, J. I. (1985). [Blue light activates electrogenic ion pumping in guard cell protoplasts of *Vicia faba*](#). *Nature* 318: 285-287.
2. Bradford, M. M. (1976). [A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding](#). *Anal Biochem* 72: 248-254.
3. Kinoshita, T. and Shimazaki, K. (1999). [Blue light activates the plasma membrane H⁺-ATPase by phosphorylation of the C-terminus in stomatal guard cells](#). *EMBO J* 18(20): 5548-5558.
4. Shimazaki, K., Iino, M. and Zeiger, E. (1986). [Blue light-dependent proton extrusion by guard-cell protoplasts of *Vicia faba*](#). *Nature* 319, 324-326.

5. Ueno, K., Kinoshita, T., Inoue, S., Emi, T. and Shimazaki, K. (2005). [Biochemical characterization of plasma membrane H⁺-ATPase activation in guard cell protoplasts of *Arabidopsis thaliana* in response to blue light.](#) *Plant Cell Physiol* 46(6): 955-963.
6. Yamauchi, S., Takemiya, A., Sakamoto, T., Kurata, T., Tsutsumi, T., Kinoshita, T. and Shimazaki, K. (2016). [The plasma membrane H⁺-ATPase AHA1 plays a major role in stomatal opening in response to blue light.](#) *Plant Physiol* 171(4): 2731-2743.