North Carolina Lineberger Comprehensive Cancer Center. Additional support includes grants from the Cystic Fibrosis Foundation (KATO20F0, MIKAMI19XX0, and OSTROW19G0).

References

- Suprynowicz FA, Upadhyay G, Krawczyk E, Kramer SC, Hebert JD, Liu X, et al. Conditionally reprogrammed cells represent a stem-like state of adult epithelial cells. Proc Natl Acad Sci USA 2012;109:20035–20040.
- Liu X, Ory V, Chapman S, Yuan H, Albanese C, Kallakury B, et al. ROCK inhibitor and feeder cells induce the conditional reprogramming of epithelial cells. Am J Pathol 2012;180:599–607.
- Koh KD, Siddiqui S, Cheng D, Bonser LR, Sun DI, Zlock LT, *et al.* Efficient RNP-directed human gene targeting reveals SPDEF is required for IL-13-induced mucostasis. *Am J Respir Cell Mol Biol* 2020;62:373–381.
- Rapiteanu R, Karagyozova T, Zimmermann N, Singh K, Wayne G, Martufi M, et al. Highly efficient genome editing in primary human bronchial epithelial cells differentiated at air-liquid interface. *Eur Respir J* 2020;55:1900950.
- Ranney ML, Griffeth V, Jha AK. Critical supply shortages the need for ventilators and personal protective equipment during the covid-19 pandemic. N Engl J Med 2020;382:e41.
- Bell CL, Quinton PM. Recycle those cell culture inserts. In Vitro Cell Dev Biol 1990;26:1123–1124.
- Fulcher ML, Randell SH. Human nasal and tracheo-bronchial respiratory epithelial cell culture. *Methods Mol Biol* 2013;945:109–121.
- Chen G, Sun L, Kato T, Okuda K, Martino MB, Abzhanova A, et al. IL-1β dominates the promucin secretory cytokine profile in cystic fibrosis. J Clin Invest 2019;129:4433–4450.
- Grubb BR, Rogers TD, Diggs PC, Boucher RC, Ostrowski LE. Culture of murine nasal epithelia: model for cystic fibrosis. Am J Physiol Lung Cell Mol Physiol 2006;290:L270–L277.
- Sisson JH, Stoner JA, Ammons BA, Wyatt TA. All-digital image capture and whole-field analysis of ciliary beat frequency. J Microsc 2003;211:103–111.
- Ostrowski LE, Yin W, Patel M, Sechelski J, Rogers T, Burns K, *et al.* Restoring ciliary function to differentiated primary ciliary dyskinesia cells with a lentiviral vector. *Gene Ther* 2014;21:253–261.
- Hsiau T, Conant D, Rossi N, Maures T, Waite K, Yang J, et al. Inference of CRISPR edits from Sanger trace data [preprint]. bioRxiv; 2019 [accessed 2020 Apr 4]. Available from: https://www.biorxiv.org/content/ 10.1101/251082v3.

Copyright © 2021 by the American Thoracic Society

Check for updates

COVID-19 and Coagulopathy

To the Editor:

We read with interest the recent review article by Rodriguez and colleagues regarding endothelial dysfunction and consequent thrombotic complication in patients with coronavirus disease (COVID-19) (1). Here, we would like to add a point. We will provide supplementary explanation on a molecular group (e.g., IL-1, IL-6, and TNF- α) that has been implicated in coagulation activation in the paper.

Supported by CREST, JST (CE19055-10).

We wish to explain the complex role of IL-6 in coagulopathy accompanied by severe inflammation. Using IL-6 knockout (IL- $6^{-/-}$) mice, we have previously demonstrated that IL-6 serves as a protector in coagulatory disturbance and thrombocytopenia during devastating systemic inflammation induced by intraperitoneal administration of bacterial endotoxins (LPS) (2), which was evidenced by prothrombin time and activated partial thromboplastin time. Furthermore, we have confirmed that in the presence of LPS, α_2 -plasmin inhibitor activity was significantly lower in $IL-6^{-/-}$ mice than in wild-type mice, indicating that IL-6 can partially inhibit LPS-provocated fibrinolysis by enhancing α_2 -plasmin inhibitor activity (3). Interestingly, fibrin degradation product was significantly lower in the $IL-6^{-/-}$ mice administered with LPS than in the wild-type mice, whereas D-dimer was comparable in both groups (unpublished observation; these complicated results require future research for further clarification). Regardless, IL-6 may not be introduced in the same way as other proinflammatory cytokines, such as IL-1 and TNF- α , in coagulatory disturbance with persistent inflammation.

Author disclosures are available with the text of this letter at www. atsjournals.org.

Ken-ichiro Inoue, M.D., Ph.D.* University of Shizuoka Shizuoka, Japan

Tomoya Sagawa, M.D. Hirohisa Takano, M.D., Ph.D. *Kyoto University Kyoto, Japan* ORCID IDs: 0000-0002-9723-5847 (K.-i.I.); 0000-0003-3048-0319 (T.S.). *Corresponding author (e-mail: inoue-k@u-shizuoka-ken.ac.jp).

References

- Rodriguez C, Luque N, Blanco I, Sebastian L, Barberà JA, Peinado VI, et al. Pulmonary endothelial dysfunction and thrombotic complications in patients with COVID-19. Am J Respir Cell Mol Biol 2021;64:407–415.
- Inoue K, Takano H, Yanagisawa R, Sakurai M, Shimada A, Morita T, et al. Protective role of interleukin-6 in coagulatory and hemostatic disturbance induced by lipopolysaccharide in mice. *Thromb Haemost* 2004;91:1194–1201.
- Inoue K, Takano H, Yanagisawa R, Sakurai M, Shimada A, Sato M, et al. Role of interleukin-6 in fibrinolytic changes induced by lipopolysaccharide in mice. Blood Coagul Fibrinolysis 2006;17:307–309.

Copyright © 2021 by the American Thoracic Society

Check for updates

Reply to: COVID-19 and Coagulopathy

From the Authors:

We read the letter from Dr. Inoue and colleagues with interest and recognize the importance of pointing out that IL-6

ิล

³This article is open access and distributed under the terms of the Creative Commons Attribution Non-Commercial No Derivatives License 4.0 (https://creativecommons.org/licenses/by-nc-nd/4.0/). For commercial usage and reprints, please contact Diane Gern (dgern@ thoracic.org).

Originally Published in Press as DOI: 10.1165/rcmb.2020-0588LE on March 26, 2021

This article is open access and distributed under the terms of the Creative Commons Attribution Non-Commercial No Derivatives License 4.0 (https://creativecommons.org/licenses/by-nc-nd/4.0/). For commercial usage and reprints, please contact Diane Gern (dgern@ thoracic.org).