

A Catalog of 5' Fusion Partners in *ROS1*-Positive NSCLC Circa 2020



Sai-Hong Ignatius Ou, MD PhD,^{a,*} Misako Nagasaka, MD^{b,c}

^a*Chao Family Comprehensive Cancer Center, Department of Medicine, Division of Hematology-Oncology, University of California Irvine School of Medicine, Orange, California*

^b*Department of Oncology, Karmanos Cancer Institute, Wayne State University, Detroit, Michigan*

^c*Department of Neurology, St. Marianna University Graduate School of Medicine, Kawasaki, Japan*

Received 18 March 2020; revised 13 April 2020; accepted 22 April 2020

Available online - 28 April 2020

ABSTRACT

ROS1 fusion-positive (*ROS1*+) NSCLC was discovered in 2007, the same year as the discovery of *ALK*-positive (*ALK*+) NSCLC but has trailed *ALK*+ NSCLC in terms of development. There seems to be a differential response to *ROS1* inhibitors, which depend on fusion partners (CD74, SLC34A2, or SDC4); thus, knowledge of the fusion partners in *ROS1*+ NSCLC is important. To date (end of February 2020), we have identified 24 unique 5' fusion partners of *ROS1* in *ROS1*+ NSCLC from published literature and congress proceedings. Thus, we published this catalog for easy reference.

© 2020 The Authors. Published by Elsevier Inc. on behalf of the International Association for the Study of Lung Cancer. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Keywords: *ROS1* fusion partner; Next-generation sequencing; *ROS1*-positive NSCLC

Introduction

ROS1 fusion-positive (*ROS1*+) NSCLC was discovered in 2007,—the same year as *ALK* fusion-positive (*ALK*+) NSCLC.¹ It constitutes about 2.9% of all adenocarcinomas of the lung.² The development of *ROS1* TKIs has followed the development of *ALK* TKIs; but to date, there are only two U.S. Food and Drug Administration-approved *ROS1* TKIs (crizotinib and entrectinib).^{3,4} Neel et al.⁵ reported that different *ROS1* fusion partners determine the subcellular localization of the *ROS1* fusion variant and the subsequent oncogenic potency of that *ROS1* fusion variant. In addition, Li et al.⁶ suggested that *ROS1* fusion partners (*CD74-ROS1* versus non-*CD74-ROS1*) have a differential response to crizotinib, and,

more importantly, have a predilection for central nervous system metastasis. Thus, it is important to have a catalog of fusion partners of *ROS1* in *ROS1*+ NSCLC.

Methods and Results

We extensively searched publications in PubMed, conference abstracts and presentations, and the cBio-Portal for Cancer Genomics website to identify novel *ROS1* fusion partners (including noncoding RNAs). We included only 5' fusion partners that retained the 3'-*ROS1* kinase domain. Overall, a total of 24 distinct *ROS1* fusion partners were identified in the literature by the end of February 2020 (Table 1). We did not include one case report, in which the *ROS1* fusion variant arose as a resistance mechanism to EGFR TKI, but the fusion partner to *ROS1* was a 3' fusion partner (*ROS1-ADGRG6*). In that *ROS1* fusion variant, the *ROS1-ADGRG6* fusion

*Corresponding author.

Disclosure: Dr. Ou has stock ownership and was on the scientific advisory board of Turning Point Therapeutics, Inc. (until Feb 28, 2019); received speaker honorarium from Merck, Roche/Genentech, Astra Zeneca, Takeda/ARIAD and Pfizer; and received advisory fees from Roche/Genentech, Astra Zeneca, Takeda/ARIAD, Pfizer, Foundation Medicine Inc., Daiichi-Sankyo, and Spectrum Pharmaceuticals. Dr. Nagasaka has received honoraria from Astra Zeneca, Caris Life Sciences, Daiichi-Sankyo, Takeda, and Tempus.

Address for correspondence: Sai-Hong Ignatius Ou, MD, PhD, Chao Family Comprehensive Cancer Center, Department of Medicine, Division of Hematology-Oncology, University of California Irvine School of Medicine, 200 South Manchester Avenue, Suite 400, Room 407, Orange, CA 92868-3298. E-mail: siou@hs.uci.edu

Cite this article as: Ou S-H and Nagasaka M. A Catalog of 5' Fusion Partners in *ROS1*-Positive NSCLC Circa 2020. *JTO Clin Res Rep* 1:100048

© 2020 The Authors. Published by Elsevier Inc. on behalf of the International Association for the Study of Lung Cancer. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

ISSN: 2666-3643

<https://doi.org/10.1016/j.jtocr.2020.100048>

Table 1. Catalog of Fusion Partners in ROS1-Positive NSCLC

No.	Fusion Partner	Year Published in Print/Presented	Chromosomal Location	Fusion Breakpoint	Response to ROS1 TKI at the Time of Publication	Tumor Source	Method of Detection	Variant Frequency in Tumor (%)	FISH/IHC	References
1	CD74	2007	5q33.1	(C6, R34)	Not treated with ROS1 TKI	FFPE	5' RACE RT-PCR	NR	+/NR	Rikova et al. ¹
2	SLC34A2	2007	4p15.2	(S4, R34)	Not treated with ROS1 TKI	HCC78 cell line	5' RACE RT-PCR	NR	+/NR	Rikova et al. ¹
3	EZR	2012	6q25.3	(E10, R34)	Not treated with ROS1 TKI	FFPE	5' RACE RT-PCR	NR	+/NR	Takeuchi et al. ¹⁶
4	LRIG3	2012	12q14.1	(L16, R35)	Not treated with ROS1 TKI	FFPE	5' RACE RT-PCR	NR	+/NR	Takeuchi et al. ¹⁶
5	SDC4	2012	20q13.12	(S2, R32) (S4, R32) (S4, R34)	Not treated with ROS1 TKI	FFPE	5' RACE RT-PCR	NR	+/NR	Takeuchi et al. ¹⁶
6	TPM3	2012	1q21.3	(T8, R35)	Not treated with ROS1 TKI	FFPE	5' RACE RT-PCR	NR	+/NR	Takeuchi et al. ¹⁶
7	GOPC (FIG)	2012	6q22.1	NR	Not treated with ROS1 TKI	FFPE	RT-PCR	NR	+/+	Rimkunas et al. ¹⁷
		2012	6q22.1	(G7, R35)	Not treated with ROS1 TKI	FFPE	RT-PCR,	NR	NR/NR	Suehara et al. ¹⁸
8	KDREL2	2012	7p22.1	NR	Not treated with ROS1 TKI	FFPE	DNA NGS	NR	NR/NR	Govindan et al. ¹⁹
9	CCDC6	2012	10q21.2	(C6, R34)	Not treated with ROS1 TKI	FFPE	DNA NGS	NR	NR/NR	Seo et al. ²⁰
10	LIMA1 ^a	2012	12q13.12	NR	Response to crizotinib	FFPE	NR	NR	+/NR	Shaw et al. ²¹
11	MSN ^a	2012	Xq12	(M9, R34)	NR	FFPE	Targeted RNA sequencing	NR	+/NR	Zheng et al. ²²
		2012	Xq12	NR	Response to crizotinib	FFPE	Targeted RNA sequencing	NR	+/NR	Shaw et al. ²¹
12	CLTC	2014	17q23.1	(C31, R35)	Not treated with ROS1 TKI	FFPE	RNA sequencing	NR	NR/NR	TCGA ²³
13	TMEM106B	2015	7p21.3	(T3, R35)	Not treated with ROS1 TKI	FFPE	DNA NGS	NR	NR/NR	Ou et al. ²⁴
14	TPD52L1	2016	6q22.31	(T3, R33)	Not treated with ROS1 TKI	FFPE	DNA NGS	NR	NR/NR	Zhu et al. ²⁵
15	SLC6A17	2017	1p13.3	NR	NR	FFPE	NGS	NR	NR/NR	Zehir ²⁶ www.cbiportal.org ⁹
16	CEP72	2018	5p15.33	(C11, R23)	Not treated with ROS1 TKI	FFPE	DNA NGS	NR	NR/NR	Zhu et al. ²⁷
17	ZCCHC8	2018	12q24.31	NR	Not treated with ROS1 TKI	FFPE	NGS	NR	NR/NR	Park et al. ²⁸
		2018	12q24.31	(Z2, R36)	Response to crizotinib	FFPE	NGS	NR	+/NR	Hicks et al. ²⁹
		2018	12q24.31	(Z2, R36)	Response to crizotinib ^b	FFPE	NGS	NR	NR/NR	Zhu et al. ³⁰
18	SLMAP	2018	3p14.3	(S?, R35)	Not treated with ROS1 TKI	FFPE	NGS	NR	NR/NR	Park et al. ²⁸
19	MYO5C	2018	15q21.2	(M?, R35)	Not treated with ROS1 TKI	FFPE	NGS	NR	NR/NR	Park et al. ²⁸
20	TFG	2018	3q12.2	NR	Not treated with ROS1 TKI	FFPE	NGS	NR	NR/NR	Park et al. ²⁸
21	WNK1	2019	12p13.33	(W25, R34)	PR to crizotinib	FFPE	NGS	19.3	NR/NR	Liu et al. ³¹

(continued)

Table 1. Continued

No.	Fusion Partner	Year Published in Print/Presented	Chromosomal Location	Fusion Breakpoint	TKI at the Time of Publication	Tumor Source	Method of Detection	Variant Frequency in Tumor (%)	FISH/IHC	References
22	MLL3 (KMT2C)	2019	7q36.1	NR	NR	Plasma	NGS	NR	NR/NR	Dagogo-Jack et al. ³²
23	CTD-2021J15.1 (LINC00973)	2019	3	NR	NR	Plasma	NGS	NR	NR/NR	Dagogo-Jack et al. ³²
24	RBPM5	2020	8p12	(R1, R32)	Response to crizotinib	FFPE	NGS	23.7	NR/NR	Zhang et al. ³³

^aBoth fusions were detected and treated in the crizotinib phase 2 trial. The *MSN-ROS1* fusion identified in the 2 reports was likely the same identical fusion variant. One report described the technique of its identification while the other report reported its response to crizotinib in the expand crizotinib phase 1 trial.

^bWith concurrent de novo *MET* amplification.

^{5'} RACE RT-PCR; 5' rapid amplification of CDA ends reverse transcription polymerase chain reaction; CCDC6, coiled-coil domain containing 6; CD74, cluster of differentiation 74; CEP72, centrosomal protein 72; CLTC, clathrin heavy chain; DCBLD1, discoidin, CUB and LCCL domain containing 1; EZR, ezrin; FFPE, formalin-fixed paraffin embedded; FISH, fluorescence in situ hybridization; GOPC (FIG), golgi associated PDZ and coiled-coil motif containing; IHC, immunohistochemistry; KM12C (MLL3), lysine methyltransferase 2C; LIMA1, LIM domain and actin binding 1; LINC00973 (CTD-2021J15.1), long intergenic nonprotein coding RNA 973; LRIG3, leucine rich repeats and immunoglobulin-like domains 3; MSN, moesin; MYO5C, myosin VC; NGS, next-generation sequencing; NR, not reported; TFG, trafficking from ER to golgi regulator, TMEM108B, transmembrane protein 108B; TPM3, tropomyosin 3; PR, partial response; TKI, tyrosine kinase inhibitor; WNK1, WNK lysine deficient protein kinase 1; ZCHC8; zinc finger CGHC-type containing 8.

variant was generated by the fusion of exons 1 to 33 of *ROS1*, which did not contain the *ROS1* kinase domain to exons 2 to 26 of *ADGRG6*. However, as the patient responded to crizotinib treatment, there was likely a potential presence of a 3' *ROS1* fusion variant.⁷ Another case reported FAM135B as a fusion partner in *ROS1*+ NSCLC.⁸ However, on verification of the data in the cBioPortal for Cancer Genomics,⁹ it was noted that the patient sample (P-0006921-T01-IM5) contained an *SLC34A2-ROS1* and a *ROS1-FAM135B* variant. In addition, the fusion breakpoint of *ROS1-FAM135B* was not recorded in the cBioPortal for Cancer Genomics. Given the nomenclature listed on the said portal, we interpreted, with the limited information available, that FAM135B would be a 3' fusion partner generated from a nonreciprocal translocation rather than a true 5' *ROS1* fusion partner. Only one intergenic rearrangement has been reported in *ROS1*+ NSCLC (Table 2).

Discussion

The number of *ROS1* fusion partners identified in *ROS1*+ NSCLC as of February 2020 is approximately 24, which is lower than that reported for *ALK*+ and *RET*+ NSCLC.^{10,11} It is quite surprising, given the fact that *ROS1*+ NSCLC was discovered in 2007, whereas *RET*+ NSCLC was discovered only in 2012, although *RET* fusions have been identified in other solid tumors, especially in thyroid cancer. The *ROS1* gene is located on chromosome 6q22.1 and only two fusion partners are located near *ROS1* (GOPC, TPD52L1), and one fusion partner, ERZ, is located on 6q25.3. Unlike *ALK*+ and *RET*+ NSCLC, only one intergenic rearrangement has been reported in *ROS1*+ NSCLC (Table 2).

Another unique feature of *ROS1*+ NSCLC is the high incidence of venous thromboembolic events.¹²⁻¹⁴ Given the potential role of fusion partners in affecting different oncogenic potencies on the *ROS1* fusion variant,⁵ the potential differential response to crizotinib, and the predilection for central nervous system metastasis,⁶ identifying *ROS1* fusion partners is essential to further advance the science and management of *ROS1*+ NSCLC. Although five fusion partners (CD74, SLC34A2, SDC4, ERZ, TPM3) made up most of the *ROS1*+ patients with NSCLC who were enrolled in the entrectinib trials, 23% of the patients diagnosed with *ROS1*+ NSCLC had unknown fusion partners.⁴ Thus, it is important for future prospective studies of *ROS1* TKIs to identify the fusion partners as much as possible, so that future translational studies can be performed from hypotheses generated from the subgroup analysis of these trials.

Table 2. List of Chromosomal Location of Intergenic Translocations With Potential ROS1 Fusion Partners

No.	Year Published in Print/Presented	Chromosomal Location	Potential Fusion Partner Gene	RET Fusion	Exon	Response to ALK TKI At the Time of Publication	Tumor Source	Method of Detection	Variant Frequency in Tumor	FISH/IHC	References
1	2019	6q22.1	DCBLD1 ^c	R35	NR	FFPE	DNA NGS	NR	NR/NR	NR/NR	Xu et al. ³⁴

^cDCBLD1 intergenic rearrangement-ROS1 was identified as a potential resistance RTK fusion to osimertinib in an EGFR+ patient with NSCLC (Del 19, T790M) in addition to RP11-565P22.6-NTRK1 fusion.

FFPE, formalin-fixed paraffin embedded; FISH, fluorescence in situ hybridization; IHC, immunohistochemistry; NGS, next-generation sequencing; NR, not reported; TKI, tyrosine kinase inhibitor.

Conclusions

1. *ROS1+* NSCLC is a heterogeneous disease with at least 24 distinct fusion partners identified in the literature up until February 2020; but fewer fusion partners were identified compared with *ALK+* and *RET+* NSCLC.
2. It is likely that many more fusion partners and intergenic rearrangements will be identified with the ever-increasing adoption of targeted RNA sequencing and whole transcriptome sequencing owing to the increasing demands of identifying rare, actionable fusions, such as *NTRK* and *NRG1* fusions.
3. We recommend clinicians worldwide to continue to report these novel fusions/intergenic rearrangements, with information on exon breakpoints/fusions, response to ROS1 TKI and allele frequency, and, if possible, whether the tumor is *ROS1*-positive on fluorescence in situ hybridization (FISH) and immunohistochemistry (IHC).
4. In this *ROS1* fusion partner catalog, most of the *ROS1+* NSCLC did not undergo any FISH or IHC testing. Currently, the companion diagnostic test for *ROS1* rearrangement approved by the U.S. Food and Drug Administration is next-generation sequencing (Oncomine Dx Target test, PMA number P160045).¹⁵ But given that FISH and IHC are still routinely used to detect *ROS1* fusion, we continue to encourage clinicians when they report novel 5' *ROS1* fusion partners to describe the FISH or IHC results if they had been performed.

References

1. Rikova K, Guo A, Zeng Q, et al. Global survey of phosphotyrosine signaling identifies oncogenic kinases in lung cancer. *Cell.* 2007;131:1190-1203.
2. Zhu Q, Zhan P, Zhang X, Lv T, Song Y. Clinicopathologic characteristics of patients with *ROS1* fusion gene in non-small cell lung cancer: a meta-analysis. *Transl Lung Cancer Res.* 2015;4:300-309.
3. Kazandjian D, Blumenthal GM, Luo L, et al. Benefit-risk summary of crizotinib for the treatment of patients with *ROS1* alteration-positive, metastatic non-small cell lung cancer. *Oncologist.* 2016;21:974-980.
4. Drilon A, Siena S, Dziadziszko R, et al. Entrectinib in *ROS1* fusion-positive non-small-cell lung cancer: integrated analysis of three phase 1-2 trials. *Lancet Oncol.* 2020;21:261-270.
5. Neel DS, Allegaerten DV, Olivas V, et al. Differential subcellular localization regulates oncogenic signaling by *ROS1* kinase fusion proteins. *Cancer Res.* 2019;79:546-556.
6. Li Z, Shen L, Ding D, et al. Efficacy of crizotinib among different types of *ROS1* fusion partners in patients with *ROS1*-rearranged non-small cell lung cancer. *J Thorac Oncol.* 2018;13:987-995.
7. Xu S, Wang W, Xu C, et al. *ROS1-ADGRG6*: a case report of a novel *ROS1* oncogenic fusion variant in lung adenocarcinoma and the response to crizotinib. *BMC Cancer.* 2019;19:769.
8. Marks EI, Pamarthi S, Dizon D, et al. *ROS1-GOPC/FIG*: a novel gene fusion in hepatic angiosarcoma. *Oncotarget.* 2019;10:245-251.
9. cBioPortal for Cancer Genomics. www.cbioperl.org. Accessed March 3, 2020.
10. Ou S-HI, Zhu VW, Nagasaka M. Catalog of 5' fusion partners in *ALK*-positive NSCLC circa 2020. *JTO Clin Res Rep.* 2020;1:100015.
11. Ou S-HI, Zhu VW. Catalog of 5' fusion partners in *RET*-NSCLC circa 2020. *JTO Clin Res Rep.* 2020;1:100037.
12. Ng TL, Smith DE, Mushtaq R, et al. *ROS1* gene rearrangements are associated with an elevated risk of peridiagnosis thromboembolic events. *J Thorac Oncol.* 2019;14:596-605.
13. Chiari R, Ricciuti B, Landi L, et al. *ROS1*-rearranged non-small-cell lung cancer is associated with a high rate of venous thromboembolism: analysis from a phase II, prospective, multicenter, two-arms trial (METROS). *Clin Lung Cancer.* 2020;21:15-20.
14. Alexander M, Pavlakis N, John T, et al. A multicenter study of thromboembolic events among patients diagnosed with *ROS1*-rearranged non-small cell lung cancer. *Lung Cancer.* 2020;142:34-40.
15. United States Food and Drug Administration. List of Cleared or Approved Companion Diagnostic Devices (In Vitro and Imaging Tools). <https://www.fda.gov/medical-devices/vitro-diagnostics/list-cleared-or-approved-companion-diagnostic-devices-vitro-and-imaging-tools>. Accessed April 13, 2020.
16. Takeuchi K, Soda M, Togashi Y, et al. *RET*, *ROS1* and *ALK* fusions in lung cancer. *Nat Med.* 2012;18:378-381.

17. Rimkunas VM, Crosby KE, Li D, et al. Analysis of receptor tyrosine kinase ROS1-positive tumors in non-small cell lung cancer: identification of a FIG-ROS1 fusion. *Clin Cancer Res.* 2012;18:4449-4457.
18. Suehara Y, Arcila M, Wang L, et al. Identification of KIF5B-RET and GOPC-ROS1 fusions in lung adenocarcinomas through a comprehensive mRNA-based screen for tyrosine kinase fusions. *Clin Cancer Res.* 2012;18:6599-6608.
19. Govindan R, Ding L, Griffith M, et al. Genomic landscape of non-small cell lung cancer in smokers and never-smokers. *Cell.* 2012;150:1121-1134.
20. Seo JS, Ju YS, Lee WC, et al. The transcriptional landscape and mutational profile of lung adenocarcinoma. *Genome Res.* 2012;22:2109-2119.
21. Shaw AT, Ou SH, Bang YJ, et al. Crizotinib in ROS1-rearranged non-small-cell lung cancer. *N Engl J Med.* 2014;371:1963-1971.
22. Zheng Z, Liebers M, Zhelyazkova B, et al. Anchored multiplex PCR for targeted next-generation sequencing. *Nat Med.* 2014;20:1479-1484.
23. Cancer Genome Atlas Research Network. Comprehensive molecular profiling of lung adenocarcinoma. *Nature.* 2014;511:543-550.
24. Ou SH, Chalmers ZR, Azada MC, et al. Identification of a novel TMEM106B-ROS1 fusion variant in lung adenocarcinoma by comprehensive genomic profiling. *Lung Cancer.* 2015;88:352-354.
25. Zhu VW, Upadhyay D, Schrock AB, Gowen K, Ali SM, Ou SH. TPD52L1-ROS1, a new ROS1 fusion variant in lung adenosquamous cell carcinoma identified by comprehensive genomic profiling. *Lung Cancer.* 2016;97:48-50.
26. Zehir A, Benayed R, Shah RH, et al. Mutational landscape of metastatic cancer revealed from prospective clinical sequencing of 10,000 patients. *Nat Med.* 2017;23:703-713.
27. Zhu YC, Zhou YF, Wang WX, et al. CEP72-ROS1: A novel ROS1 oncogenic fusion variant in lung adenocarcinoma identified by next-generation sequencing. *Thorac Cancer.* 2018;9:652-655.
28. Park S, Ahn BC, Lim SW, et al. Characteristics and outcome of ROS1-positive non-small cell lung cancer patients in routine clinical practice. *J Thorac Oncol.* 2018;13:1373-1382.
29. Hicks JK, Boyle ALA, Albacker LA, Madison R, Frampton G, Creelan BC. Clinical activity of crizotinib in lung adenocarcinoma harboring a rare ZCCHC8-ROS1 fusion. *J Thorac Oncol.* 2018;13:e148-e150.
30. Zhu YC, Wang WX, Xu CW, et al. A novel co-existing ZCCHC8-ROS1 and de-novo MET amplification dual driver in advanced lung adenocarcinoma with a good response to crizotinib. *Cancer Biol Ther.* 2018;19:1097-1101.
31. Liu Y, Liu T, Li N, Wang T, Pu Y, Lin R. Identification of a novel WNK1-ROS1 fusion in a lung adenocarcinoma sensitive to crizotinib. *Lung Cancer.* 2019;129:92-94.
32. Dagogo-Jack I, Rooney M, Nagy RJ, et al. Molecular analysis of plasma from patients with ROS1 positive NSCLC. *J Thorac Oncol.* 2019;14:816-824.
33. Zhang Y, Yu M, Yuan M, Chen R, Huang MJ. Identification of a novel RBPM3-ROS1 fusion in an adolescent patient with microsatellite-unstable advanced lung adenocarcinoma sensitive to crizotinib: A case report. *Clin Lung Cancer.* 2020;21:e78-e83.
34. Xu H, Shen J, Xiang J, et al. Characterization of acquired receptor tyrosine-kinase fusions as mechanisms of resistance to EGFR tyrosine-kinase inhibitors. *Cancer Manag Res.* 2019;11:6343-6351.