GENOME SEQUENCES



Complete Genome Sequence of a Clinical Isolate of Acinetobacter baumannii Harboring 11 Plasmids

Arata Sakiyama,^a Yuki Matsumoto,^a [®]Taishi Tsubouchi,^{a,b,c} [®]Masato Suzuki,^d Makoto Niki,^{a,e} [®]Mamiko Niki,^{a,b} [®]Ken-Ich Oinuma,^{a,b} [®]Hiroshi Kakeya,^e [®]Yukihiro Kaneko^{a,b}

Microbiology[®]

Resource Announcements

^aDepartment of Bacteriology, Graduate School of Medicine, Osaka City University, Abeno, Osaka, Japan ^bResearch Center for Infectious Disease Sciences, Graduate School of Medicine, Osaka City University, Abeno, Osaka, Japan ^cToneyama Institute for Tuberculosis Research, Graduate School of Medicine, Osaka City University, Toneyama, Osaka, Japan ^dAntimicrobial Resistance Research Center, National Institute of Infectious Diseases, Aoba, Higashi-Murayama, Tokyo, Japan ^eDepartment of Infection Control and Prevention, Osaka City University Hospital, Abeno, Osaka, Japan

ABSTRACT Here, we report the complete genome sequence of an *Acinetobacter baumannii* isolate harboring 11 plasmids, obtained at a hospital in Japan in 2016. The complete 4.07-Mbp genome sequence (1 chromosome and 11 plasmids) was analyzed by a combination of long-read (Flongle) and short-read (NovaSeq 6000) sequencing.

A cinetobacter spp. are naturally competent at DNA uptake and transformation using type IV pili (1), contributing to the horizontal gene transfer (HGT) of antimicrobial resistance genes and virulence genes. We isolated an *Acinetobacter* sp. strain, obtained from the venous blood of a patient in Japan in 2016 (IRB approval number 3568; 30 September 2016). When we extracted and electrophoresed the genomic DNA (gDNA), we found that the strain had multiple plasmids. We thought that sequencing the genome of this strain would help elucidate the plasmid acquisition mechanism of *Acinetobacter* species.

The blood culture became positive at 21.9 h, and the bacteria were then isolated on sheep blood agar and MacConkey agar solid medium at 35°C for 24 h. The isolates were analyzed using a MicroScan WalkAway 40 system equipped with a Neg NF combo panel (Beckman Coulter, Brea, CA, USA) and determined to be an *Acinetobacter* sp.

OCU_Ac18 was streaked onto a Mueller-Hinton II agar plate and incubated at 37°C for 16 h. gDNA was extracted using the DNeasy blood and tissue kit (Qiagen). Wholegenome sequencing was performed using the MinION system with FLO-FLG001 (Oxford Nanopore Technologies [ONT]) and the NovaSeq 6000 system (Illumina). The gDNA was sheared to 8 kbp using a g-TUBE device (M&S Instruments) and prepared using a ligation sequencing kit (SQK-LSK108) for the ONT library. The Illumina library (paired-end format; insert size, 500 to 900 bp) was prepared using the Nextera XT DNA library prep kit. The ONT reads were base called using Guppy v3.2.1 in high-accuracy mode. The read N_{50} value and the number of reads were 6.92 kbp and 87,219, respectively. The ONT reads were filtered using Filtlong v0.2.0 (https://github.com/rrwick/ Filtlong; length, \geq 1,000) to keep only 90% of reads, with a target output of 500 Mb $(N_{50}, 7.08 \text{ kbp})$. The Illumina reads (read length, 151 bp; number of reads, 31,907,116) were filtered using Fastp v0.20.1 to a quality score of \geq 30, trimming 5 bases off the 3' end (2). Both sets of filtered reads were assembled de novo using Unicycler v0.4.8 (3). The assembly yielded 12 circular contigs, and the *dnaA* gene and *rep*-like gene on OCUAc18-4 and pOCUAc18-6 were rotated to start at the first gene. The rep-like genes of other contigs were annotated using the RAST v2.0 server (4) and were rotated manually to put the rep-like gene first. Sequencing errors such as indels were corrected four times using Pilon v1.24 (5). The complete genome sequence of OCU_Ac18 was

Citation Sakiyama A, Matsumoto Y, Tsubouchi T, Suzuki M, Niki M, Niki M, Oinuma K-I, Kakeya H, Kaneko Y. 2021. Complete genome sequence of a clinical isolate of *Acinetobacter baumannii* harboring 11 plasmids. Microbiol Resour Announc 10:e00695-21. https://doi.org/10.1128/MRA.00695-21.

Editor David Rasko, University of Maryland School of Medicine

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Address correspondence to Yukihiro Kaneko, ykaneko@med.osaka-cu.ac.jp.

Received 8 July 2021 Accepted 25 August 2021 Published 30 September 2021

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TABLE 1 Sequencing metrics for the OCU_Ac18 genome

Genome element	Sequence length (bp)	G+C content (%)	No. of reads	DDBJ accession no.
OCU_Ac18chr	3,920,683	39.2	24,992,837	AP024802
pOCUAc18-1	90,090	40.4	594,136	AP024803
pOCUAc18-2	12,817	35.9	604,436	AP024804
pOCUAc18-3	10,831	35.8	735,450	AP024805
pOCUAc18-4	8,948	34.8	471,670	AP024806
pOCUAc18-5	7,104	37.8	516,595	AP024807
pOCUAc18-6	5,049	37.2	626,625	AP024808
pOCUAc18-7	4,293	41.1	476,726	AP024809
pOCUAc18-8	3,948	41.0	412,868	AP024810
pOCUAc18-9	3,496	36.8	390,525	AP024811
pOCUAc18-10	2,444	36.4	407,282	AP024812
pOCUAc18-11	2,278	39.9	436,954	AP024813

constructed as one chromosome and 11 plasmids (Table 1). A total of 3,863 coding DNA sequences (CDSs) were annotated using the DDBJ Fast Annotation and Submission Tool (DFAST) server (6). The complete genome sequence of OCU_Ac18 was compared to the complete genome sequence of *A. baumannii* ATCC 19606^T (GenBank accession number AP022839) using average nucleotide identity (ANI) methods, calculated using pyani v0.2.10. The ANI between the genomes was 97.50%, so we finally identified OCU_Ac18 as *A. baumannii*.

Data availability. The complete genome sequence of OCU_Ac18 has been deposited in DDBJ/ENA/GenBank under the accession numbers described in Table 1, and the raw sequence data are available in the Sequence Read Archive under the accession numbers DRX291375 and DRX291376.

ACKNOWLEDGMENTS

This research was supported by grants (JP21fk0108133 to Y. Kaneko, JP21fk0108133 to T. Tsubouchi, JP21fk0108133 to H. Kakeya, and JP21fk0108093, JP21fk0108139, JP21fk0108133, JP21wm0325003, JP21wm0325022, JP21wm0225004, and JP21wm0225008 to M. Suzuki) from the Japan Agency for Medical Research and Development (AMED) and grants (21K07008 to Y. Kaneko, 21H03007 to T. Tsubouchi, and 20K07509 to M. Suzuki) from the Ministry of Education, Culture, Sports, Science and Technology (MEXT), Japan.

REFERENCES

- Wilharm G, Piesker J, Laue M, Skiebe E. 2013. DNA uptake by the nosocomial pathogen Acinetobacter baumannii occurs during movement along wet surfaces. J Bacteriol 195:4146–4153. https://doi.org/10.1128/JB.00754-13.
- Chen S, Zhou Y, Chen Y, Gu J. 2018. fastp: an ultra-fast all-in-one FASTQ preprocessor. Bioinformatics 34:i884–i890. https://doi.org/10.1093/bioinformatics/ bty560.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. https://doi.org/10.1371/journal.pcbi.1005595.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL,

Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75. https://doi.org/10.1186/1471-2164-9-75.

- Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. PLoS One 9:e112963. https://doi.org/10.1371/journal.pone.0112963.
- Tanizawa Y, Fujisawa T, Nakamura Y. 2018. DFAST: a flexible prokaryotic genome annotation pipeline for faster genome publication. Bioinformatics 34:1037–1039. https://doi.org/10.1093/bioinformatics/btx713.