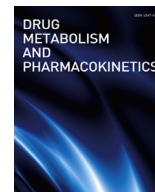




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## Review article

## Difference in the lipid nanoparticle technology employed in three approved siRNA (Patisiran) and mRNA (COVID-19 vaccine) drugs

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## ARTICLE INFO

## Article history:

Received 18 July 2021

Received in revised form

4 October 2021

Accepted 8 October 2021

Available online 10 October 2021

## Keywords:

Lipid nanoparticles

Ionizable lipid

LNP

mRNA vaccine

COVID-19

Patisiran

Tozinameran

Elasomeran

COVID-19 vaccine moderna

SARS-CoV-2

## ABSTRACT

Nucleic acid therapeutics are developing into precise medicines that can manipulate specific genes. However, the development of safe and effective delivery system for the target cells has remained a challenge. Lipid nanoparticles (LNPs) have provided a revolutionary delivery system that can ensure multiple clinical translation of RNA-based candidates. In 2018, Patisiran (Onpattro) was first approved as an LNP-based siRNA drug. In 2020, during the coronavirus disease 2019 (COVID-19) outbreak, LNPs have enabled the development of two SARS-CoV-2 mRNA vaccines, Tozinameran (Comirnaty or Pfizer-BioNTech COVID-19 vaccine) and Elasmomeran (Spikevax or COVID-19 vaccine Moderna) for conditional approval. Here, we reviewed the state-of-the-art LNP technology employed in three approved drugs (one siRNA-based and two mRNA-based drugs) and discussed the differences in their mode of action, formulation design, and biodistribution.

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## 1. Introduction

In the 21st century, nucleic acids are attracting attention as next generation modality beyond a small molecule and antibody. Since the approval of the world's first nucleic acid drug "fomivirsen" in 1998, a total of 16 nucleic acid medicines (9 antisense oligonucleotides (ASO), 4 short interfering RNAs (siRNA), 1 aptamer, and 2 messenger RNAs (mRNA)) have been approved in some areas of US, EU, and Japan, as of May 2021 [1]. More than 80% of these drugs (13 out of 16) have been approved since 2016. This rapid expansion of nucleic acid therapeutics has been triggered by scientific breakthrough in drug-delivery systems. Tremendous efforts toward the development of delivery carriers [2–9] have identified lipid nanoparticles (LNPs) as a clinically validated platform technology that can deliver both long (i.e. mRNA [10,11]) and short nucleic acids (i.e. siRNA [12]) into target cells. Lipid nanoparticle technology has

played a central role in the development of the world's first drugs associated with two completely independent phenomena, namely RNA interference by siRNA and vaccination by mRNA. In this review, we have focused on three approved drugs using lipid nanoparticle technology, namely Patisiran as a siRNA medicine, and Tozinameran and Elasmomeran mRNA vaccines. Their development history [13,14], recent clinical trials and perspectives [15], gene regulation [16], structure of ionizable lipid [17], and mRNA vaccine [18–20] have already been reported in previous studies. This review has partially referred to the information from an interview form archived in Pharmaceuticals and Medical Devices Agency (PMDA) [21,22,23].

Table 1 provides an overview of the three approved drugs. Patisiran (trade name: Onpattro, developed by Alnylam) was first approved in 2018 for the treatment of hereditary transthyretin (hATTR) amyloidosis. siRNA is designed to target a sequence within the untranslated region (3'UTR) of transthyretin (TTR) mRNA. LNP-formulated siRNA is administered intravenously to patients at a dose of 0.3 mg/kg every three weeks [24]. The risk of infusion reactions and adverse events often triggered by nanomedicine [25]

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**Table 1**  
Overview of the three approved drugs.

Active ingredient	Patisiran	Tozinameran	Elasomeran
Trade name	Onpattro	Comirnaty	Spikevax
Company	Alnylam	Pfizer/BioNTech	Moderna
Development code	ALN-TTR02	BNT162b2	mRNA-1273
First approval year	Aug-2018	Dec-2020 (conditional)	Dec-2020 (conditional)
Indication	hATTR amyloidosis	Prevention of COVID-19	Prevention of COVID-19
Target protein	TTR	Spike of SARS-CoV-2 <sup>a</sup>	Spike of SARS-CoV-2 <sup>a</sup>
LNP-formulated RNA	siRNA	mRNA	mRNA
Administration route	Intravenous	Intramuscular	Intramuscular
Administration schedule	every 3 weeks	2 doses, 3 weeks apart	2 doses, 4 weeks apart
Dosage strength	0.3 mg/kg siRNA <sup>b</sup>	30 µg mRNA	100 µg mRNA
Dosage volume	Total 200 mL <sup>c</sup>	0.3 mL <sup>d</sup>	0.5 mL
Administration time	over 70 min	immediately	immediately
Premedication	required	Not required	Not required
Storage condition	2 °C to 8 °C (do not freeze)	-90 °C to -60 °C	-50 °C to -15 °C

The information is derived from the interview form archived in PMDA as of May 2021 [21–23]. hATTR, hereditary transthyretin; SARS-CoV-2, severe acute respiratory syndrome coronavirus 2; COVID-19, coronavirus disease 2019; LNP, lipid nanoparticles.

<sup>a</sup> Spike proteins are modified by 2 proline substitutions to produce prefusion-stabilized SARS-CoV-2 spike proteins, S-2P.

<sup>b</sup> Body weight is less than 103 kg; 31.2 mg siRNA, when 104 kg or more.

<sup>c</sup> Diluted by saline from 2 mg/mL siRNA in supplied vial.

<sup>d</sup> Diluted by saline from 0.5 mg/mL mRNA in supplied vial to 0.1 mg/mL for injection.

are mitigated by two approaches, namely premedication and slow infusion [26]. Tozinameran (trade name: Comirnaty, developed by Pfizer/BioNTech) was granted Emergency Use Authorization (EUA) by the United States Food and Drug Administration (FDA) in December 2020 for the prevention of coronavirus disease 2019, COVID-19 [27]. mRNA is designed to encode the stabilized prefusion SARS-CoV-2 spike protein, a key molecule for eliciting neutralizing antibodies. LNP-formulated mRNA is administered intramuscularly to human at a dose of 30 µg in a series of two doses (0.3 mL each) three weeks apart. Elasomeran (trade name: Spikevax, developed by Moderna) was also granted EUA at approximately the same time in 2020 [28]. The two mRNA vaccines share multiple similarities, with slight differences in dosages and administration schedules; none required premedication. Differences in the storage conditions of all 3 drugs should, however, be noted. While the siRNA-LNP can be stored in a standard medical refrigerator (2 °C to 8 °C), the mRNA-LNP products need to be stored in a freezer (approximately -20 °C) or in ultra-cold storage (-80 °C to -60 °C).

## 2. Difference in mode of action

Since Tozinameran and Elasomeran have very similar modes of action, the 3 drugs have been classified into two categories in this section, namely siRNA-LNP and mRNA-LNP (Table 2).

### 2.1. Chemistry of siRNA and mRNA

The siRNA in Patisiran is a short, double-stranded RNA with 21 bases in each strand while mRNA in the two vaccines is a long, single-stranded RNA with over 4000 bases (Fig. 1) [14]. Human body recognizes RNA as a toxic virus, not as medicines, via cellular sensors meant for biological defense against invading pathogens. Chemical modification can allow RNA to avoid this response triggered by pattern recognition receptors (PRRs), such as toll-like receptors (TLRs) and retinoic acid-inducible gene I receptors (RIG-I) [29,30]. In siRNA, uridine and cytosine are partially replaced by 2'-O-methyl uridine and cytosine. In mRNA, uridine is fully replaced by N1-methyl pseudouridine [31] (Fig. 1). Since innate immune

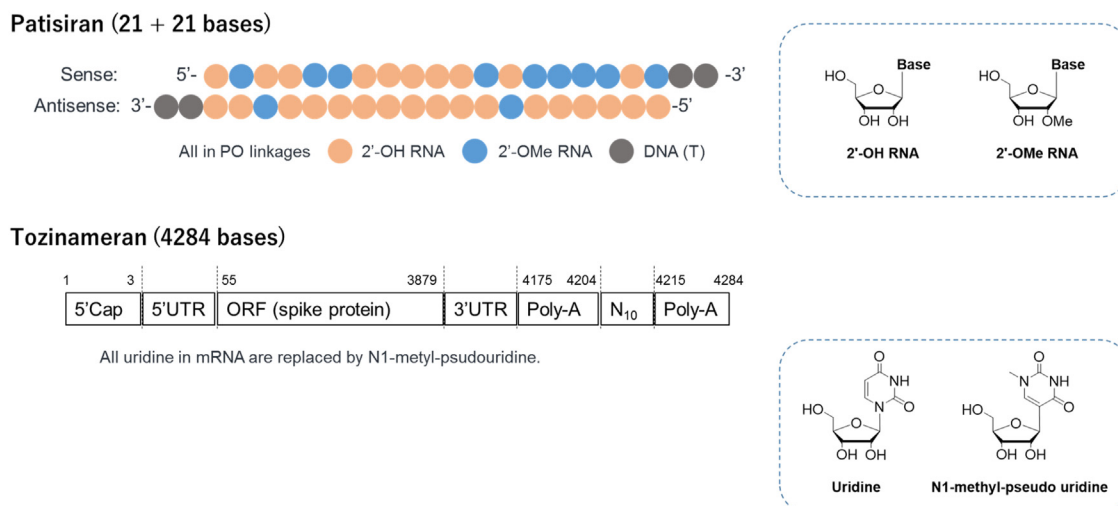
**Table 2**  
Difference in mode of action.

Active ingredient (Company)	Patisiran (Alnylam)	Tozinameran (Pfizer/BioNTech), Elasomeran (Moderna)
Drug substance	siRNA	mRNA
Strand	double strand	single strand
RNA length	42 (21 + 21)	4284 (Tozinameran) <sup>a</sup>
Base modification	2'-OMe-uridine, 2'-OMe-cytosine	N1-methyl-pseudouridine
Purpose of base modification	suppress the innate immune response to mitigate off-target effect	suppress the innate immune response to increase protein production
Mode of action	stop production of target protein by degrading mRNA	initiate production of target protein by supplying mRNA, which triggers the adaptive immunity
Target protein	Transthyretin (TTR)	spike of SARS-CoV-2 <sup>b</sup>
Endogenous machinery	RNA interference	translation
Location of machinery	cytoplasm	cytoplasm
Primary target cell of LNP	hepatocytes	antigen presenting cells (APCs)

LNP, lipid nanoparticles; SARS-CoV-2, severe acute respiratory syndrome coronavirus 2.

<sup>a</sup> Structure of Elasomeran is not described in detail.

<sup>b</sup> Target spike proteins are modified by 2 proline substitutions to produce prefusion-stabilized SARS-CoV-2 spike protein.



**Fig. 1.** Structure of the drugs. For both mRNA designs, S–2P spike proteins were modified by 2 proline substitutions to produce prefusion-stabilized SARS-CoV-2 spike proteins. UTR, untranslated region; ORF, open reading frame.

responses lead to downregulation of mRNA translation through phosphorylation of eIF2a by dsRNA-dependent protein kinase [32], N1-methyl pseudouridine modification enhances protein production, resulting in increased immunogenicity [31,33].

In principle, siRNA can be chemically manufactured by phosphoramidite method using an automated solid-phase DNA/RNA synthesizer [34]. mRNA is enzymatically manufactured by in-vitro transcription (IVT) from a plasmid DNA template encoding mRNA sequences [35,36].

## 2.2. Mechanism of action of siRNA-LNP after administration

Intravenously administered LNP-formulated siRNA accumulates in the liver, the primary site of transthyretin (TTR) production. After the uptake of LNPs by hepatocytes in the liver, siRNA harnesses the endogenous RNA interference pathway, degrades TTR mRNA, and reduces the production of TTR protein.

## 2.3. Mechanism of action of mRNA-LNP after administration

The EMA assessment report has detailed the process associated with mRNA vaccine at the injection site as follows: “Intramuscular administration of LNP-formulated RNA vaccines results in transient local inflammation that drives recruitment of neutrophils and antigen presenting cells (APCs) to the site of delivery. Recruited APCs are capable of LNP uptake and protein expression (i.e. spike of SARS-CoV-2) by translation and can subsequently migrate to the local draining lymph nodes where T cell priming occurs [37].”

## 2.4. Delivery of LNP from extracellular to intracellular environment

The biggest challenge for nucleic acid drugs is RNA delivery into the cytoplasm, where the endogenous machinery, for RNA interference and protein translation, is located. In principle, the LNP delivery mechanism is similar for siRNA [38–40] and mRNA [10,11,41]. The three main steps are as follows (Fig. 2). First, full encapsulation by LNP protects RNA from nuclease digestion. LNPs are neutral in physiological pH due to the ionizable lipid and polyethylene glycol (PEG)-lipid, thereby reducing non-specific interactions with serum proteins. Second, following dissociation of the PEG-lipid, cells take up LNPs via apolipoprotein E (ApoE)-

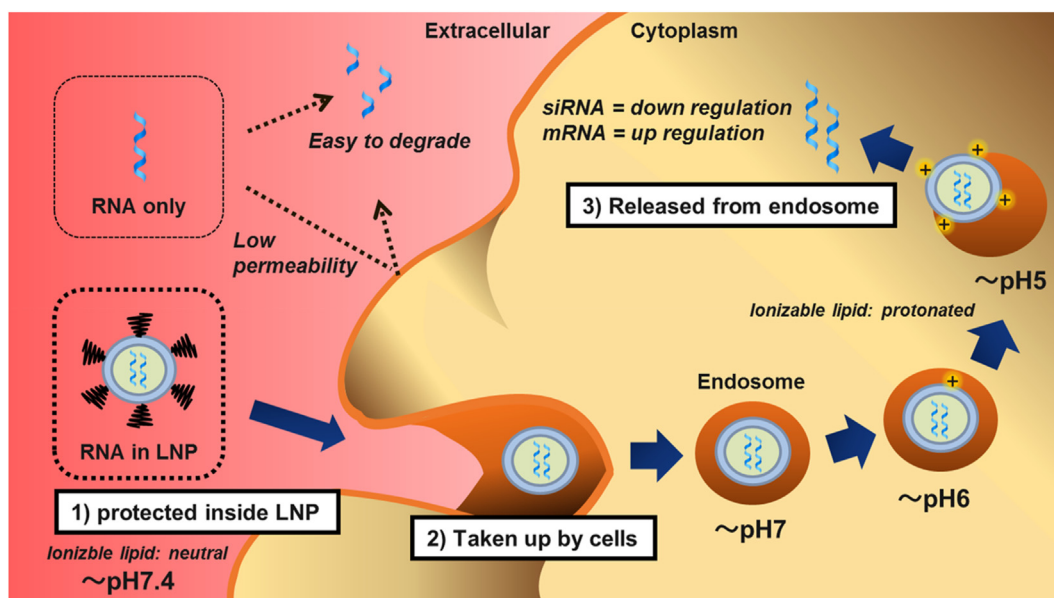
dependent and/or ApoE-independent pathways. Finally, protonated LNPs, upon acidification in the endosome, induce hexagonal phase structures, disrupt the membranes, and release RNA molecules into the cytoplasm. The released RNA molecules lead to either down regulation (siRNA) or up regulation (mRNA) of target proteins. Details regarding the uptake of LNPs into hepatocytes have already been reported, revealing that LNPs are internalized via LDL receptors following the ApoE-dependent pathway [38]. Details regarding the uptake into antigen-presenting cells have not yet been clarified; however, efficient localization of nucleic acids in the cytoplasm of antigen presenting cells has been reported [42].

## 3. Difference in formulation

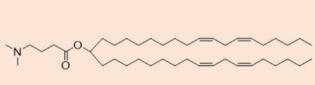
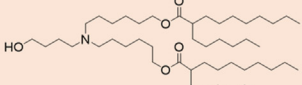
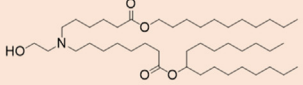
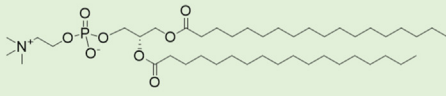
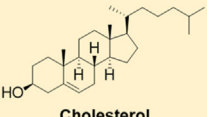
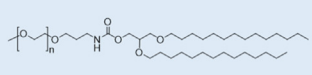
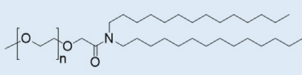
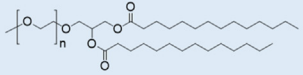
The three LNP-based drugs share multiple similarities in their formulation, and hence, behave similarly as nanoparticles in vivo. Importantly, all LNPs are composed of four types of lipids; ionizable lipid, phospholipid, cholesterol, and PEG-lipid (Fig. 3). All 3 ionizable lipids have tertiary amine group with pKa 6.0–6.7. These lipids switch its charge from neutral to cationic based on the neutral pH in the blood and the acidic pH in endosomes. The 3 PEG-lipids have dialkyl chains 14-carbon long, which are important for the rapid dissociation from the surface of LNPs once inside the body [43]. The biodegradable design of ALC-0315 [44] and SM-102 [11] is described later.

Lipid composition of the three drug products is summarized in Table 3. All three LNPs were composed of four lipids in similar molar ratios. The major differences are in the storage condition of drug products. A recent review estimated that mRNAs, not LNPs, are the main factors for low storage stability of drug products, and thus mRNA vaccines need to be supplied at freezing (about –20 °C) or ultra-freezing (–80 to –60 °C) conditions to slow down mRNA hydrolysis [45]. Sucrose is used as a cryoprotectant to maintain the physical properties of LNPs during freeze-thaw cycles [46]. In contrast to the two mRNA vaccines, Patisiran needs to avoid freezing temperatures, probably because lipid nanoparticles dissolved in PBS, not in Sucrose, cannot tolerate freezing.

In principal, assembly of LNPs is achieved by rapid mixing of four lipids in ethanol with RNA molecules in an aqueous buffer (near pH 4) in a microfluidic or T-junction mixer [47–49].



**Fig. 2.** Delivery mechanism of lipid nanoparticles. First, full encapsulation by lipid nanoparticles (LNP) protects RNA from nuclease digestion. LNPs are neutral in physiological pH due to the ionizable lipid and PEG-lipid, thereby reducing non-specific interactions with serum proteins. Second, following dissociation of the PEG-lipid, cells take up LNPs via apolipoprotein E (ApoE)-dependent and/or ApoE-independent pathways. Finally, protonated LNPs, upon acidification in the endosome, induce hexagonal phase structures, disrupt the membranes, and release RNA molecules into the cytoplasm.

Active ingredient (Company)	Patisiran (Alynam)	Tozinameran (Pfizer/BioNTech)	Elasomeran (Moderna)
Ionizable lipid	 DLin-MC3-DMA	 ALC-0315	 SM-102
Phospholipid	 DSPC		
Sterol	 Cholesterol		
PEG-lipid	 PEG <sub>2000</sub> -C-DMG	 ALC-0159	 PEG <sub>2000</sub> -DMG

**Fig. 3.** Chemical structure of lipids in lipid nanoparticles. ALC-0159 has PEG<sub>2000</sub>. All 3 ionizable lipids have tertiary amine groups, namely DLin-MC3-DMA (MC3), pKa 6.44 [12] or pKa 6.35 [11]; ALC-0315, pKa 6.09 [44]; and SM-102, pKa 6.68 [11]. The related patents are as follows: DLin-MC3-DMA, WO/2010/144740; ALC-0315, WO/2017/075531 (Lipid No. 3); and SM-102, WO/2017/049245 (Compound 25).

#### 4. Difference in biodistribution

The major difference in biodistribution of the three LNP-based drugs arises from the administration route (Table 4). Regarding Patisiran, a study on rats using <sup>14</sup>C-labeled ionizable lipid (<sup>14</sup>C-MC3) had revealed that with a single intravenous dose of siRNA-LNP, approximately 90% of the administered radioactivity was detected in the liver 4 h after administration [21]. Regarding Tozinameran, a

study on rats using <sup>3</sup>H-labeled cholesteryl hexadecyl ether (<sup>3</sup>H-CHE) had revealed that with a single intramuscular dose of mRNA-LNP (luciferase mRNA as surrogate), radioactivity concentration was the highest at the injection site between 15 min and 48 h (i.e. the entire measurement period) after administration [22]. Besides that in the administration site, the total radioactivity recovery rate for the dose was highest in the liver (up to 18%) 8–48 h after administration [22]. Subcutaneously administered liposomes

**Table 3**  
Composition of lipid nanoparticles.

Active ingredient (Company)	Patisiran (Alnylam)	Tozinameran (Pfizer/BioNTech)	Elasomeran (Moderna)
Single dosage	0.3 mg/kg siRNA	0.030 mg mRNA	0.10 mg mRNA
Unit	5 mL	0.45 mL	0.5 mL
Drug substance	10 mg <sup>a</sup>	0.225 mg	0.10 mg
Ionizable lipid	65.0 mg	3.23 mg	1.075 mg
Phospholipid	16.5 mg	0.7 mg	0.275 mg
Cholesterol	31.0 mg	1.4 mg	0.47 mg
PEG-lipid	8.0 mg	0.4 mg	0.115 mg
Total lipid	120.5 mg	5.7 mg	1.94 mg
Total lipid/RNA (wt/wt)	12.1	25.5	19.4
Drug concentration	2.0 mg/mL	0.5 mg/mL	0.2 mg/mL
Sucrose in formulation	NA	46 mg	43.5 mg
Sucrose concentration	NA	102 g/L	87 g/L
pH	6.4–7.5	6.9–7.9	7.0–8.0
State of drug product	liquid	frozen suspension	frozen suspension
Shelf life of drug product (unopened vials) <sup>b</sup>	27 months (2–8 °C, avoid freeze)	6 months (–90 to –60 °C)	6 months (–25 to –15 °C)

The information is derived from the interview form archived in PMDA as of May 2021 (21–23).

<sup>a</sup> As Patisiran, without sodium salt.

<sup>b</sup> Shelf life of each drug is updated by ongoing storage stability tests.

**Table 4**  
Topics related to the pharmacokinetics of lipid nanoparticles.

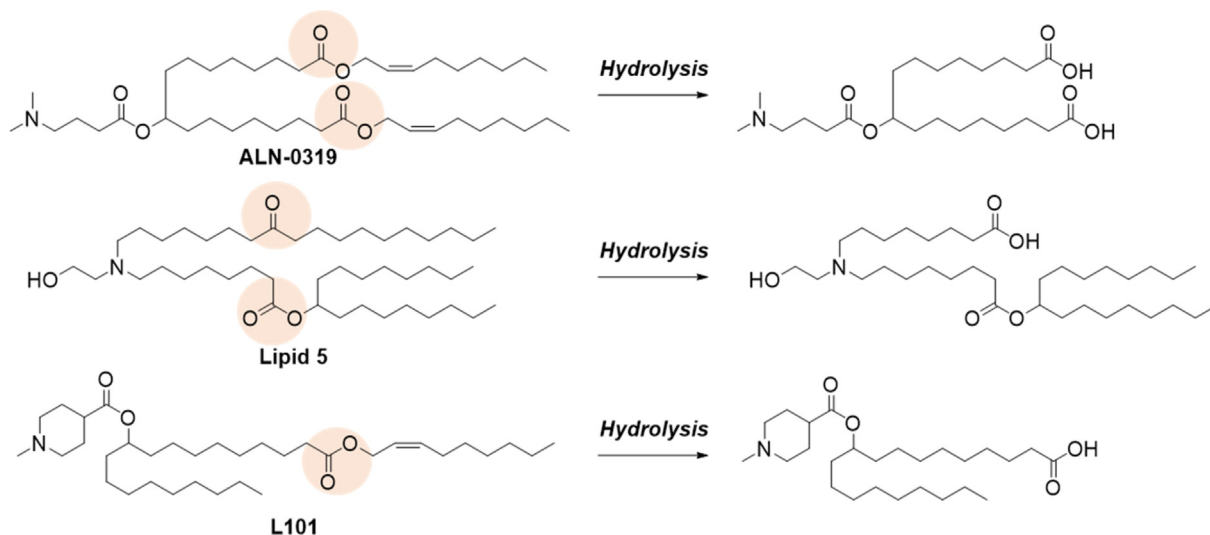
Active ingredient (Company)	Patisiran (Alnylam)	Tozinameran (Pfizer/BioNTech)
Administration route	intravenous	intramuscular
Primary biodistribution	liver	injection site
Biodegradable property in ionizable lipid	no	yes
Anti-drug antibody (ADA) assessments	anti-PEG antibody	not reported

(approximately  $<0.1 \mu\text{m}$ ) did not enter the blood capillary owing to their size; they remained at the injection site and were drained through lymphatic capillaries to nearby lymph nodes [50]. The innate biodistribution of nanoparticles is favorable for mRNA vaccines, since 1) adaptive immune response occurs at lymph nodes, and 2) unwanted systemic exposure is reduced. A detailed study had been conducted with intramuscularly administered mRNA-LNPs in non-human primates [51].

Ideal drug carriers are rapidly eliminated from the body once their purpose is served. Various LNPs incorporated biodegradable designs in an ionizable lipid as a means to facilitate their

elimination (Fig. 4) [10,52,53]. In contrast to MC3, ALC-0315 and SM-102 have ester linkages in the lipid tail. Intramuscular injection of mRNA-LNPs composed of SM-102 or MC3 resulted in faster clearance of SM-102, along with improved tolerability, over MC3 at the injection site [11]. The two ester linkages in ALC-0315 were also hydrolyzed in vivo [22].

Biologics, modified with PEG, produce anti-drug antibodies (ADA), more specifically anti-PEG antibodies, in vivo [54]. siRNA-LNP [55] and mRNA-LNP [56] have been shown to generate anti-PEG antibodies in mice, leading to accelerated blood clearance (ABC) upon repeated administration. In the phase 3 APOLLO trial,



**Fig. 4.** Biodegradable design in ionizable lipid. Pink circle in each chemical structure shows the hydrolysis site. Hydrolysis of ester linkage facilitates rapid elimination and improved tolerability. (For interpretation of the references to color in this figure legend, the reader is referred to the Web version of this article.)

ADAs to Patisiran were evaluated by measuring antibodies specific to PEG<sub>2000</sub>-C-DMG that is exposed on the surface of LNPs [57]. The overall incidence of treatment-emergent ADAs was 3.4% (5 of 145 patients) in the Patisiran group and 1.3% (1 of 77 patients) in the placebo group. Treatment-emergent ADAs to PEG<sub>2000</sub>-C-DMG occurred at a low frequency, were transient and appeared to have no effect on PK, PD, safety or efficacy [57]. To the best of our knowledge, no study has been reported on anti-PEG antibody yet, which causes accelerated blood clearance due to intramuscularly administered LNPs.

In relation to anti-PEG antibody, a severe allergic reaction, named anaphylaxis, has been reported to occur (though rarely; a couple of cases per million doses) due to the administration of two mRNA vaccines [58]. The trigger of anaphylaxis is considered to be due to anti-PEG IgE against PEG-lipid [59,60].

## 5. Conclusion

This review focused on the differences in LNP technology across three approved drugs. With the initial success of siRNA therapeutics [14], the technology platform has become well-established and its scope has expanded from siRNA to mRNA [15]. In early 2020, two pioneering candidates challenged to fight against COVID-19 outbreak, namely mRNA-1273 by Moderna [61] and BNT162 by Pfizer/BioNTech [62]. Following their examples, multiple mRNA vaccines leveraging LNP technology are currently in clinical trials, including CVnCoV by CureVac/BAYER [63,64], ARCT-021 by Arcturus Therapeutics [65], ARCoV by Valvax Biotechnology/Suzhou Abogen Biosciences [66], LNP-nCoVsaRNA by Imperial College London [67], and DS-5670 by Daiichi Sankyo/The University of Tokyo [68]. Recent clinical study of CVnCoV using mRNA with unmodified nucleotides may reveal the importance of mRNA chemistry [64]. A detailed understanding of these LNPs and RNA chemistry would expand the platform technology and open up pathways to next-generation therapeutics.

## Author contributions

Y.S. wrote manuscript. H.I. reviewed manuscript.

## Declaration of competing interest

The authors declare no conflicts of interest associated with this manuscript.

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