

1 **Title:** A Third Dose of SARS-CoV-2 Vaccine Increases Neutralizing Antibodies Against
2 Variants of Concern in Solid Organ Transplant Recipients

3 **Authors:** Andrew H. Karaba¹, Xianming Zhu², Tao Liang¹, Kristy H. Wang¹, Alex G.
4 Rittenhouse¹, Olivia Akinde², Yolanda Eby², Jessica E. Ruff², Joel N. Blankson¹, Aura T.
5 Abedon³, Jennifer L. Alejo³, Andrea L. Cox^{1,4,5}, Justin R. Bailey¹, Elizabeth A.
6 Thompson^{1,5}, Sabra L. Klein^{1,4}, Daniel S. Warren³, Jacqueline M. Garonzik-Wang⁶,
7 Brian J. Boyarsky³, Ioannis Sitaras⁴, Andrew Pekosz^{1,4}, Dorry L. Segev³, Aaron A.R.
8 Tobian^{2*}, William A. Werbel^{1*}

9 **Affiliations:**

10 ¹Department of Medicine, Johns Hopkins University School of Medicine, Baltimore, MD 21287, USA.

11 ²Department of Pathology, Johns Hopkins University School of Medicine, Baltimore, MD 21287, USA

12 ³Department of Surgery, Johns Hopkins University School of Medicine, Baltimore, MD 21287, USA.

13 ⁴W. Harry Feinstone Department of Molecular Microbiology and Immunology, Johns Hopkins University
14 Bloomberg School of Public Health, Baltimore, MD 21287, USA

15 ⁵Bloomberg Kimmel Institute for Cancer Immunotherapy, Johns Hopkins University School of Medicine,
16 Baltimore, MD 21287, USA.

17 ⁶Department of Surgery, University of Wisconsin School of Medicine and Health, Madison, WI 53792,
18 USA

19 *These authors contributed equally

20

21

22 **Corresponding author email:** wwerbel1@jhmi.edu

23

24

25

26

27

28

29

30

31 **Abbreviations:**

- 32 Anti-N: Anti-nucleocapsid antibody
- 33 Anti-RBD: Anti-Receptor Binding Domain antibody
- 34 Anti-S: Anti-Spike antibody
- 35 AU: Arbitrary Unit
- 36 AUC: Area Under the Curve
- 37 CFR: Case Fatality Rate
- 38 CM: Complete Media
- 39 ELISA: enzyme-linked immunosorbent assay
- 40 HC: Healthy Control
- 41 IM: Infection Media
- 42 MSD: Meso Scale Diagnostics
- 43 nAb: Neutralizing antibody
- 44 NT50: 50% Neutralization Titer
- 45 OD: Optical Density
- 46 SOTR: Solid Organ Transplant Recipient
- 47 TCID50: 50% Tissue Culture Infectious Dose
- 48 VOC: Variant of Concern

49

50 **Word Count: 2,898**

51

52

53

54 **ABSTRACT:**

55 Vaccine-induced SARS-CoV-2 antibody responses are attenuated in solid organ
56 transplant recipients (SOTRs) and breakthrough infections are more common.
57 Additional SARS-CoV-2 vaccine doses increase anti-spike IgG in some SOTRs, but it is
58 uncertain whether neutralization of variants of concern (VOCs) is enhanced. We tested
59 47 SOTRs for clinical and research anti-spike IgG, pseudoneutralization (ACE2
60 blocking), and live-virus neutralization (nAb) against VOCs before and after a third
61 SARS-CoV-2 vaccine dose (70% mRNA, 30% Ad26.COV2.S) with comparison to 15
62 healthy controls after two mRNA vaccine doses. We used correlation analysis to
63 compare anti-spike IgG assays and focused on thresholds associated with neutralizing
64 activity. A third SARS-CoV-2 vaccine dose increased median anti-spike (1.6-fold) and
65 receptor-binding domain (1.5-fold) IgG, as well as pseudoneutralization against VOCs
66 (2.5-fold versus Delta). However, IgG and neutralization activity were significantly lower
67 than healthy controls ($p < 0.001$); 32% of SOTRs had zero detectable nAb against Delta
68 after third vaccination. Correlation with nAb was seen at anti-spike IgG > 4 AU on the
69 clinical assay and $> 10^4$ AU on the research assay. These findings highlight benefits of
70 a third vaccine dose for some SOTRs and the need for alternative strategies to improve
71 protection in a significant subset of this population.

72

73

74

75

76

77 **1. INTRODUCTION:**

78 Solid organ transplant recipients (SOTRs) are at increased risk for severe
79 COVID-19¹. For example, while the case fatality rate (CFR) for the general population in
80 the United States is 1-2%², CFRs range from 10-30% in SOTRs, due to a combination
81 of chronic disease and immunosuppressive medications³. Therefore, effective and
82 optimized vaccines that prevent COVID-19 disease in this group are critical.
83 Unfortunately, these patients were excluded from the phase III trials of the mRNA based
84 COVID-19 vaccines,^{4,5} and recent publications suggest that breakthrough disease is
85 more common among fully-vaccinated SOTRs than the general population^{6,7}.
86 Furthermore, it has been demonstrated that many SOTRs develop weak SARS-CoV-2
87 antibody responses after the recommended two doses of an mRNA-based vaccine⁸⁻¹¹.
88 This led to the hypothesis that a third dose of mRNA-based SARS-CoV-2 vaccine may
89 improve the immune response to SARS-CoV-2 and protection from COVID-19. Although
90 third doses have been authorized for immunocompromised persons in several
91 countries, including the United States (US), published data on neutralizing capacity of
92 SOTR plasma after additional vaccine doses are limited¹²⁻¹⁴. In particular, it is unknown
93 if augmented immune responses to a third vaccine dose would result in protection
94 against more transmissible variants of concern (VOCs) that exhibit immune escape,
95 including the Delta variant which currently comprises >99% of new cases in the US¹⁵. In
96 an effort to assess whether a third dose of SARS-CoV-2 vaccine in SOTRs would
97 improve the SARS-CoV-2 specific neutralizing response, we measured total SARS-
98 CoV-2 specific IgG and neutralizing activity against the vaccine strain and four VOCs
99 before and after a third dose of SARS-CoV-2 vaccine and compared this to IgG levels

100 and neutralizing capacity of healthy controls who received a standard two-dose mRNA-
101 based vaccine series.

102

103 **2. MATERIALS AND METHODS:**

104 *2.1 Cohorts:*

105 SOTR participants were enrolled in a national prospective, observational cohort:
106 COVID-19 Antibody Testing of Recipients of Solid Organ Transplants and Patients with
107 Chronic Diseases, Johns Hopkins IRB00248540, as previously described^{9,16}. Healthy
108 control participants were enrolled under Johns Hopkins IRB00027183¹⁷. SOTRs
109 submitted blood samples to the investigators 0-4 weeks before and 2 weeks after third
110 vaccine doses, which were independently obtained in the community. Blood was
111 collected in Acid Citrate Dextrose tubes and plasma was isolated by Ficoll centrifugation
112 and stored at -80°C.

113 *2.2 IgG Measurement:*

114 Plasma was tested using the clinically available EUROIMMUN anti-SARS-CoV-2
115 IgG enzyme-linked immunosorbent assay (ELISA) versus the S1 domain of spike
116 protein, performed per the manufacturers' protocols. Optical density (OD) of the sample
117 was divided by calibrator provided arbitrary unit (AU) ratio, for which ≥ 1.1 was
118 considered positive and ≥ 0.8 – 1.1 were considered indeterminate. Plasma was thawed
119 and anti-N, anti-RBD, and anti-S IgG was measured using the multiplex
120 chemiluminescent Meso Scale Diagnostics (MSD) V-PLEX COVID-19 Respiratory
121 Panel 3 Kit according to the manufactures' protocol at a dilution of 1:5000.

122

123 *2.3 Pseudoneutralization/ACE2 Inhibition Measurement:*

124 Plasma from study participants was thawed and ACE2 blocking was measured
125 using the ACE2 MSD V-PLEX SARS-CoV-2 Panel 6 and Panel 14 kits according to the
126 manufacturers' protocol at a dilution of 1:100.

127 *2.4 Viruses and cells.*

128 VeroE6-TMPRSS2 cells¹⁸ were cultured in complete media (CM) consisting of
129 DMEM containing 10% FBS (Gibco, Thermo Fisher Scientific), 1 mM glutamine
130 (Invitrogen, Thermo Fisher Scientific), 1 mM sodium pyruvate (Invitrogen, Thermo
131 Fisher Scientific), 100 U/mL penicillin (Invitrogen, Thermo Fisher Scientific), and 100
132 µg/mL streptomycin (Invitrogen, Thermo Fisher Scientific). Cells were incubated in a 5%
133 CO₂ humidified incubator at 37°C.

134 The SARS-CoV-2/USA-WA1/2020 virus was obtained from BEI Resources. The
135 delta variant of SARS-CoV-2 (hCoV19/USA/MD-HP05660/2021, EPI_ISL_2331507)
136 was isolated on Vero-E6-TMPRSS2 cells plated in 24-well dishes and grown to 75%
137 confluence. The CM was removed and replaced with 150 µL of infection medium (IM),
138 which is identical to CM but with the fetal bovine serum reduced to 2.5%, and 150 µL of
139 the viral transport media containing a swab from a patient confirmed to be SARS-CoV-2
140 positive was added to the culture. The cultures were incubated at 37°C for 2 hours, the
141 inoculum was aspirated and replaced with 0.5 mL of IM and the cells cultured at 37°C
142 for up to 5 days. When a cytopathic effect was visible in most of the cells, the IM was
143 harvested and stored at -70°C. The presence of SARS-CoV-2 was verified by
144 extracting RNA from the harvested supernatant using the Qiagen Viral RNA extraction
145 kit (Qiagen), and viral RNA detected using quantitative RT-PCR.¹⁹ The consensus

146 sequence of the virus isolate did not differ from the sequence derived from the clinical
147 specimen.

148 The infectious virus titer was determined on VeroE6-TMPRSS2 cells using a
149 50% tissue culture infectious dose (TCID₅₀) assay as previously described for SARS-
150 CoV.^{20,21} Serial 10-fold dilutions of the virus stock were made in IM, and then 100 µL of
151 each dilution was added to the cells in a 96-well plate in sextuplicate. The cells were
152 incubated at 37°C for 4 days, visualized by staining with naphthol blue-black, and
153 scored visually for cytopathic effect.

154 *2.5 Neutralization assay.*

155 The neutralizing antibody (nAb) levels were determined as described for SARS-
156 CoV.²² Two-fold dilutions of plasma (starting at a 1:20 dilution) were made in IM.
157 Infectious virus was added to the plasma dilutions at a final concentration of 1×10^4
158 TCID₅₀/mL (100 TCID₅₀ per 100 µL). The samples were incubated for 1 hour at room
159 temperature, and then 100 µL of each dilution was added to 1 well of a 96-well plate of
160 VeroE6-TMPRSS2 cells in sextuplet for 6 hours at 37°C. The inocula were removed,
161 fresh IM was added, and the plates were incubated at 37°C for 2 days or until complete
162 cytopathic effect was visible in wells exposed to virus without plasma. The cells were
163 fixed by the addition of 100 µL of 4% formaldehyde per well, incubated for at least 4
164 hours at room temperature, and then stained with Naphthol Blue Black (MilliporeSigma).
165 The nAb titer was calculated as the highest serum dilution that eliminated the cytopathic
166 effect in 50% of the wells and area under the curve (AUC) was calculated using
167 GraphPad Prism.

168

169 *2.6 Statistical analysis:*

170 Only SOTRs with available demographic and immunological data on pre and
171 post third dose of SARS-CoV-2 vaccine were included in the analysis. Wilcoxon signed
172 rank test was used to compare the median of SARS-CoV-2 anti-Spike and anti-RBD
173 IgG level and percent ACE2 inhibition before and after third dose of vaccine among
174 SOTRs. The median of IgG level and ACE2 inhibition between SOTRs and HCs were
175 compared using Wilcoxon rank sum test. Pearson correlation was used to evaluate the
176 linear association between Spike IgG and percent ACE2 inhibition among SOTRs. A
177 spline knot was added at $4 \log_{10}(\text{AU})$ MSD IgG. Bonferroni correction was conducted to
178 control multiple comparison when analyzing variants ($p < 0.01$ was considered
179 statistically significant). The analysis was also stratified by type of third dose vaccine,
180 age, sex, and graft transplanted to evaluate effect measure modification. Missing values
181 were treated using available case strategy in subgroup analysis.

182

183 **3. RESULTS:**

184 *3.1* Pre- and post-third dose samples were available for 47 SOTRs followed in
185 our ongoing longitudinal observational cohort studying immunogenicity and safety of
186 SARS-CoV-2 vaccination. Most of these participants had previously undergone anti-
187 spike antibody testing using two clinically available assays¹⁶. The median age was 63
188 (interquartile range 49-70) years and 55% were female. Most SOTRs were kidney
189 transplant recipients (64%) and all initially received two doses of an mRNA-based
190 vaccine (23 Moderna mRNA-1273, 24 Pfizer BNT162b2). Most were taking a
191 calcineurin inhibitor-based maintenance immunosuppression regimen (77%) and 30%

192 were on “triple immunosuppression” with a calcineurin inhibitor, an antimetabolite, and
193 corticosteroids. 70% of SOTRs received a third mRNA vaccine dose and 30% received
194 the Janssen Ad26.COVID.S vaccine. None reported a known history of COVID-19.
195 Among mRNA-vaccinated healthy controls (HC, N=15), none had known medical
196 conditions, and all received two doses of BNT162b2. See **Table 1** for full demographic
197 and clinical data.

198 Anti-S1-Receptor binding domain (RBD), anti-Spike (S), and anti-Nucleocapsid
199 (N) total IgG were measured in plasma using a research assay (MSD) with FDA-verified
200 sero-positivity cutoffs before and after a third dose of SARS-CoV-2 vaccine in SOTRs
201 and after two doses of an mRNA-based vaccine in HCs. No participants had a positive
202 anti-N response at before or after a third dose of vaccine (**Supplemental Figure 1**).
203 Prior to a third dose of vaccine, 17 (36%) and 11 (23%) SOTRs were sero-positive for
204 anti-RBD and anti-S, respectively (**Figure 1A**). After the third dose, these numbers
205 increased to 36 (77%) and 34 (72%), respectively, and there was a significant increase
206 in the median total anti-S (1.6 fold change) and anti-RBD (1.5 fold change) IgG levels
207 compared to matched pre-third dose samples (**Figure 1A**). The median anti-RBD and
208 anti-S IgG values of SOTRs receiving a third dose remained significantly lower than the
209 median responses in fully vaccinated HCs after the two-dose series (**Figure 1B**). In
210 comparison to all other transplant recipients, kidney transplant recipients had
211 significantly lower anti-S IgG (**Figure 1C**). Eight (57%) of those receiving Ad26.COVID.S
212 as a third dose and 26 (79%) who received an mRNA-based vaccine as a third dose
213 became sero-positive. When stratifying by type of third dose received (mRNA versus
214 Ad.COVID.S), however, we did not observe a significant difference in median anti-S IgG

215 value (**Supplemental Figure 2C**). In exploratory analysis, median IgG levels did not
216 differ by other key clinical or demographic parameters such as age or sex, though
217 subgroup sizes were small (**Supplemental Figures 2A and 2B**). Notably, seven female
218 kidney recipients had the lowest post third dose IgG levels of all SOTRs in the study. All
219 were taking anti-metabolite maintenance immunosuppression, but they did not
220 otherwise share clinical or demographic factors.

221 3.2 Next, we investigated the neutralizing potential of SOTR plasma versus major
222 SARS-CoV-2 VOCs after three vaccine doses with comparison to that of healthy
223 individuals after two vaccine doses. This was assessed initially via an assay measuring
224 capacity of plasma to inhibit S protein binding to the ACE2 receptor, termed
225 pseudoneutralization. There was a significant increase in the median
226 pseudoneutralization of all variants after a third vaccine dose among SOTRs: fold
227 changes 2.5, 2.2, 1.6, 1.5, and 2.5 for vaccine, Alpha, Beta, Gamma, Delta variants,
228 respectively (**Figure 2A**). However, pseudoneutralization of all variants was significantly
229 lower than that of healthy controls after two doses of an mRNA-based vaccine (**Figure**
230 **2B**). For example, only two (6%) SOTRs had pseudoneutralization values for the Delta
231 variant above the first quartile of the healthy control pseudoneutralization values; the
232 majority were below 20% inhibition for all variants. When stratified by type of organ
233 received, kidney transplant recipients had significantly lower ACE2 inhibition versus the
234 vaccine strain and Alpha variant compared to all other organs (**Figure 2C**). Stratification
235 by age, sex, or vaccine platform, did not identify any significant differences in
236 pseudoneutralization (**Supplemental Figure 3**).

237 We also examined the correlation between anti-S IgG and pseudoneutralization
238 for all the variants. We found a strong correlation between anti-S IgG and
239 pseudoneutralization, but the relationship only became linear around 4 log₁₀(arbitrary
240 unit, AU) IgG, suggesting that values below this may not correlate with neutralizing
241 response (**Figure 2D**).

242 3.3 Finally, we used live-virus neutralization (nAb) to assess 50% neutralization
243 titer (NT50) and area under the curve (AUC) against the vaccine strain and the Delta
244 variant before and after a third vaccine dose in SOTRs and in two-dose vaccinated
245 HCs. For SOTRs, median (IQR) NT50s were 40 (10-120) versus vaccine strain and 20
246 (10-40) versus Delta (**Figure 3A**), with median (IQR) AUC of 50 (2-145) and 9 (3-50),
247 respectively (**Figure 3B**) after a third vaccine dose. This corresponded to a fold change
248 in NT50 of 1.6 and 1.3 and a fold change in AUC 50.2 and 8.4 versus the vaccine strain
249 and Delta variant, respectively. Compared to HCs, NT50s and AUC versus vaccine and
250 Delta variant strains were significantly lower among SOTRs (**Figure 3D**). Fully 32% of
251 SOTRs had nAb NT50s at or below the limit of detection versus the Delta variant after a
252 third vaccine dose (as compared to 0% of HCs). There were two female liver transplant
253 recipients with very high neutralizing titers, even beyond those of the HCs.

254 3.4 We assessed inter-assay correlation for both the vaccine strain (**Figure 4A**)
255 and Delta variants (**Figure 4B**) among clinical (EUROIMMUN) and research (MSD) anti-
256 spike IgG assays, as well as pseudoneutralization and nAb AUC for SOTRs and HCs.
257 For the vaccine strain, EUROIMMUN and MSD IgG showed excellent positive
258 correlation, particularly above the clinical manufacturer threshold for seropositivity (1.1
259 AU). Correlation of pseudoneutralization with both IgG assays was strong above a

260 threshold of 20% ACE2 blocking. Below this, there was marked variation in
261 corresponding IgG levels among SOTRs (e.g., EUROIMMUN IgG ranged 0-7.5 AU)
262 (**Supplemental Figure 4**). Correlation of both IgG assays and nAb AUC was moderate,
263 though markedly improved when restricting to higher IgG cutoffs (4 AU on the
264 EUROIMMUN assay and 4 log₁₀(AU) on the MSD assay). Overall correlation of
265 pseudoneutralization and nAb was stronger, particularly when restricting each to one
266 patient group (SOTR or HC). These cross-correlation patterns were similar when
267 considering the Delta variant, although pseudoneutralization and nAb AUC correlation
268 was overall stronger as compared to the vaccine strain, reflecting reduction in ACE2
269 blocking for HCs.

270

271 **4. DISCUSSION**

272 Here, we provide evidence that a third dose of COVID-19 vaccine increases
273 plasma neutralization against VOCs for some SOTRs, including versus the highly
274 transmissible and now dominant Delta variant. This was robustly characterized using a
275 combination of clinical and research IgG assays, pseudoneutralization, and gold-
276 standard live-virus neutralization. Although median plasma neutralizing capacity did
277 increase for SOTRs, levels were generally far below that of HCs after the two-dose
278 mRNA series and 32% showed no nAb against the Delta variant using the live-virus
279 assay.

280 Other key findings include lower neutralization among kidney transplant
281 recipients versus other transplant recipients, potentially reflecting heavier maintenance
282 immunosuppression. Other factors previously associated with improved sero-response

283 such as younger age or third dose vaccine platform (i.e., mRNA) were not clearly
284 associated with response. Importantly, although there was significant variability in IgG
285 responses in SOTRs, we found evidence through correlation analysis that certain IgG
286 cutoffs were associated with clear increases in ACE2 blocking, as well as in nAb. This is
287 an early step toward establishing thresholds for high-throughput assays that may
288 indicate protection from COVID-19, including the Delta variant, though this will need to
289 be tested by assessing risk of infection in real-world cohort and clinical trial settings.

290 The observed variable humoral response to additional vaccine doses in this high-
291 risk group indicates that alternative strategies, such as immunosuppressive modulation
292 or using emerging vaccine platforms, may be necessary to induce a protective response
293 to vaccination. While some SOTRs clearly produce an antibody response on par with
294 HCs, they were a small minority in this study. Though we identified kidney
295 transplantation as a risk factor for decreased responsiveness, the mechanism
296 underlying this association is unknown. This is evident in the seven female kidney
297 recipients who all had very low IgG and neutralizing responses, yet had no clear pattern
298 in demographic features, age, or vaccine platform received as a third dose. Additional
299 investigations and deeper immunological analyses are warranted to understand in a
300 personalized fashion why some SOTRs respond to additional antigen exposure, while
301 others do not.

302 This study was limited by its observational nature and small number of
303 participants with demographic and immunosuppressive heterogeneity. Additionally, HC
304 comparators were younger than the SOTR group, which may contribute to observed
305 differences in humoral response. Although patient survey and anti-N IgG were used to

306 rule out prior COVID-19, it is possible that subclinical infections occurred in some
307 patients before or after vaccination. Furthermore, mucosal immune responses and
308 cellular immune responses were not characterized in this study.

309 In summary, a third dose of a SARS-CoV-2 vaccine increases anti-spike and
310 anti-RBD IgG levels and plasma neutralizing capability, including against the Delta
311 variant, in some SOTRs. Yet, a significant portion of SOTRs have limited or no
312 neutralizing activity against the dominant VOC indicating that a third dose of vaccine
313 may not be a fully effective strategy for a large portion of immunocompromised patients.
314 These data also inform how research and clinical anti-spike IgG measurements might
315 be used to estimate neutralizing ability and potential sero-protection thresholds. This is
316 novel and timely information regarding the potential improvement of immune protection
317 against SARS-CoV-2 variants in a highly vulnerable population amidst ongoing
318 community surges.

319

320

321 **ACKNOWLEDGEMENTS:**

322 This work was supported by the Ben-Dov family, the Johns Hopkins COVID-19
323 Vaccine-related Research Fund, the National Cancer Institute (U54CA260491), grants
324 T32DK007713 (JLA), F32DK124941 (BJB), and K23DK115908 (JMGW) from the
325 National Institute of Diabetes and Digestive and Kidney Diseases, and grants
326 K24AI144954 (DLS), K08AI156021 (AHK), K23AI157893 (WAW),
327 HHSN272201400007C (AP) and R01AI120938S1 (AART) from the National Institute of
328 Allergy and Infectious Disease.

329

330 **AUTHOR CONTRIBUTIONS:**

331 AHK and WAW conceived of the study and design. OA, JER, and YE processed the
332 samples and prepared them for the assays. AHK, KHW, and AGR performed the MSD
333 assays and collected the antibody data. AP and IS performed the live virus
334 neutralization. ATA, JLA, JNB, DW, and BJB assisted with participant enrollment and
335 collection of clinical data. XZ and TL performed the analysis. EAT assisted with sample
336 curation and data interpretation. AHK and WAW wrote the original manuscript. JMG,
337 ALC, JNB, SLK, AP, JRB, DLS, AART, and WAW supervised the studies, provided
338 material support, and contributed to the interpretation of results. All authors aided in
339 editing the manuscript.

340

341 **DISCLOSURES:**

342 DLS has the following financial disclosures: consulting and speaking honoraria from
343 Sanofi, Novartis, CSL Behring, Jazz Pharmaceuticals, Veloxis, Mallinckrodt, Thermo
344 Fisher Scientific. None of the other authors have any relevant competing interests.

345

346 **DATA AVAILABILITY:**

347 Reasonable requests for deidentified data to the corresponding author will be granted.

348

349

350

351

352 **FIGURE LEGENDS**

353 **Figure 1. Changes in SARS-CoV-2 Specific IgG After a Third Dose of SARS-CoV-2**

354 **Vaccine**

355

356 **A.** Total SARS-CoV-2 S1 RBD (left) and Spike (right) specific IgG in SOTRs before
357 and after a third dose of vaccine. The dashed line represents the assay
358 manufacturer's cut-off for seropositivity based on convalescent samples.

359 **B.** Total SARS-CoV-2 S1 RBD (left) and Spike (right) specific IgG in fully mRNA
360 vaccinated healthy controls (HCs) (n = 15) and SOTRs after a third dose of
361 vaccine (n = 47).

362 **C.** Total SARS-CoV-2 Spike specific IgG in SOTRs who received kidney (n = 30)
363 and non-kidney (n = 17) transplant.

364
365 The boxplots represent the IQR. The median is represented by a horizontal line in
366 the box. The lower and upper whiskers represent 1.5x the IQR beyond the quartiles.
367 Each dot represents an individual sample. Statistical differences between groups
368 were determined by Wilcoxon signed rank test for panel A, and Wilcoxon rank sum
369 test for panel B and C. P-values of <0.05 were considered significant.

370

371 **Figure 2. SARS-CoV-2 Pseudoneutralization After a Third Dose of COVID-19**

372 **Vaccine in SOTRs**

373

374 **A.** Pseudoneutralization of full-length SARS-CoV-2 Spike variants (indicated in top
375 header of each panel) before and after a third dose of vaccine among SOTRs.

376 **B.** Pseudoneutralization of full-length SARS-CoV-2 Spike variants (indicated in top
377 header of each panel) in SOTRs (n = 47) after a third dose of vaccine compared
378 to fully vaccinated healthy controls (n = 15).

379 **C.** Comparison of pseudoneutralization of full-length SARS-CoV-2 Spike variants
380 (indicated in top header of each panel) in SOTRs who received kidney (n = 30)
381 and non-kidney (n = 17) transplant.

382 **D.** Correlation between total SARS-CoV-2 Spike IgG and pseudoneutralization of
383 full-length SARS-CoV-2 Spike variants among SOTRs receiving a third dose of
384 vaccine.

385 In panel A-C, the boxplots represent the IQR. The median is represented by a
386 horizontal line in the box. The lower and upper whiskers represent 1.5x the IQR
387 beyond the quartiles. Each dot represents an individual sample. Statistical
388 differences between groups were determined by Wilcoxon signed rank test for panel
389 A, and Wilcoxon rank sum test for panel B and C. Pearson correlation coefficient
390 were generated for panel D. P-values of <0.01 were considered significant after
391 Bonferroni correction.

392

393 **Figure 3. Neutralizing antibody (nAb) versus SARS-CoV-2 vaccine strain and**
394 **Delta variant**

395
396 A. nAb NT50 versus SARS-CoV-2 vaccine strain and Delta variant before and after
397 a third dose SARS-CoV-2 vaccine among SOTRs.

398 B. nAb area under curve (AUC) versus SARS-CoV-2 vaccine strain and Delta
399 variant before and after a third dose SARS-CoV-2 vaccine among SOTRs.

400 C. Comparison of nAb reciprocal NT50 versus SARS-CoV-2 vaccine strain and
401 Delta variant between SOTRs after a third dose of SARS-CoV-2 vaccine and
402 HCs after two mRNA vaccine doses

403 D. Comparison of nAb AUC of SARS-CoV-2 vaccine strain and Delta variant
404 between SOTRs after a third dose of SARS-CoV-2 vaccine and HCs after two
405 mRNA vaccine doses.

406
407 In panel A-D, the boxplots represent the IQR. The median is represented by a
408 horizontal line in the box. The lower and upper whiskers represent 1.5x the IQR
409 beyond the quartiles. Each dot represents an individual sample. Statistical
410 differences between groups were determined by Wilcoxon signed rank test for panel
411 A and B, and Wilcoxon rank sum test for panel C and D.

412
413
414
415

416 **Figure 4. Correlations between neutralizing antibody (nAb), percent ACE2**
417 **inhibition, MSD anti-spike IgG and EUROIMMUN anti-spike IgG of SARS-CoV-2**
418 **among SOTRs and HCs**

419
420 A. Correlations between neutralizing and IgG assays versus the SARS-CoV-2
421 vaccine strain among SOTRs after a third dose of vaccine and HCs after two
422 doses.

423 B. Correlations between neutralizing and IgG assays versus the Delta variant
424 among SOTRs after a third dose of vaccine and HCs after two doses.

425 Each point on the scatter plots represents an individual sample. Pearson correlation
426 coefficients between assays are presented in the upper panels. “Corr” represents
427 the correlation across all samples. “HCs” (in red) represents the correlation among
428 only HCs. “SOTR” (in blue) represents the correlation among only SOTRs. * $p <$
429 0.05 ; ** $p < 0.01$; *** $p < 0.001$. Density plots of SOTRs and HCs are shown in
430 diagonal panels. Unit of analysis: nAb AUC, \log_{10} AUC; ACE2: percent ACE2
431 inhibition; MSD IgG, \log_{10} IgG AU/mL; EUROIMMUN IgG, AU/mL.

432

433 **Table 1. Clinical and Demographic Characteristics of SOTRs and Healthy**

434 **Controls.**

	Overall n = 62	SOTR n = 47	Healthy controls, n = 15
Age, years			
20-39	10 (16)	3 (6)	7 (47)
40-59	26 (42)	18 (38)	8 (53)
60-79	26 (42)	26 (55)	0 (0)
Sex			
Female	31 (50)	26 (55)	5 (33)
Male	31 (50)	21 (45)	10 (67)
Race			
White	57 (92)	46 (98)	11 (73)
Asian	4 (6)	1 (2)	3 (20)
African American	1 (2)	0 (0)	1 (7)
Graft transplanted			
Kidney*	-	30 (64)	-
Liver	-	10 (21)	-
Heart	-	4 (9)	-
Lung	-	2 (4)	-
Pancreas	-	1 (2)	-
Anti-rejection medication†			
Prednisone	-	22 (47)	-
Calcineurin Inhibitors	-	36 (77)	-
mTOR inhibitors	-	7 (15)	-
anti-metabolites	-	30 (64)	-
Type of the third dose vaccine			
mRNA	-	33 (70)‡	-
Ad26.COV2.S	-	14 (30)	-
Days between second dose and third dose vaccine	-	102 (70 - 124)	-
Days between transplant and third dose vaccine	-	1778 (930 - 4419)	-
Days post second dose vaccine	-	-	8 (7 - 10)

435 Note: all study participants received mRNA vaccine for the first two doses. Categorical variables were presented in
 436 n (%), and continuous variables were presented in median (interquartile range). *1 person had both kidney and
 437 pancreas transplanted and has been grouped into kidney category. †Anti-rejection medication use was not mutually
 438 exclusive. ‡10 (30%) of the 33 participants received a third mRNA vaccine that differed from their initial two-dose
 439 series.

440

441

442

443

444

445

446

447 **Supplemental Figure 1.**

448 Total SARS-CoV-2 Nucleocapsid specific IgG in SOTRs before and after a third dose of
449 vaccine. The dashed line represents the assay manufacturer's cut-off for positivity
450 based on convalescent samples. P-values were calculated using Wilcoxon rank sum
451 test.

452

453 **Supplemental Figure 2.**

454
455 Total SARS-CoV-2 Spike specific IgG in SOTRs who received a third dose of COVID-19
456 vaccine stratified by age (<60 n = 21 and ≥60 years n = 26), sex (female n = 26 and
457 male n = 21), and type of third dose vaccine received (mRNA n = 33 and Ad26.COV2.S
458 n = 14). P-values were calculated using Wilcoxon rank sum test and should be
459 considered exploratory given the small subgroups.

460

461 **Supplemental Figure 3.**

462 Pseudoneutralization of full-length SARS-CoV-2 Spike variants in SOTRs who received
463 a third dose of COVID-19 vaccine stratified by age (<60 n = 21 and ≥60 years n = 26),
464 sex (Female n = 26 and male n = 21), and type of third dose vaccine received (mRNA n
465 = 33 and Ad26.COV2.S n = 14). P-values were calculated using Wilcoxon rank sum test
466 and should be considered exploratory given the small subgroups.

467

468

469

470 **Supplemental Figure 4. Correlation between percent ACE2 inhibition and**
471 **neutralizing antibody (nAb) and IgG of SARS-CoV-2 among SOTRs after a third**
472 **dose of COVID-19 vaccine, stratified by ACE2 inhibition level**

473 A. Pearson correlation between vaccine strain percent ACE2 inhibition and vaccine
474 strain neutralizing antibody (nAb), MSD IgG and EUROIMMUN IgG of SARS-
475 CoV-2 among SOTRs after a third dose of COVID-19 vaccine.

476 B. Pearson correlation between Delta variant percent ACE2 inhibition and Delta
477 variant neutralizing antibody (nAb), MSD IgG and EUROIMMUN IgG of SARS-
478 CoV-2 among SOTRs after a third dose of COVID-19 vaccine.

479 Units of analysis: nAb AUC, \log_{10} AUC; ACE2: percent ACE2 inhibition; MSD IgG,
480 \log_{10} IgG AU/mL; EUROIMMUN IgG, AU/mL.

481

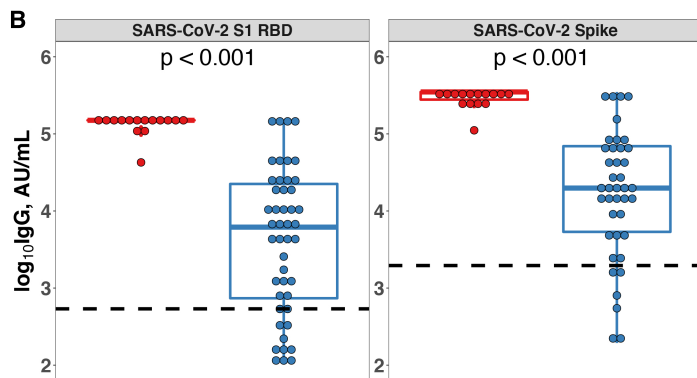
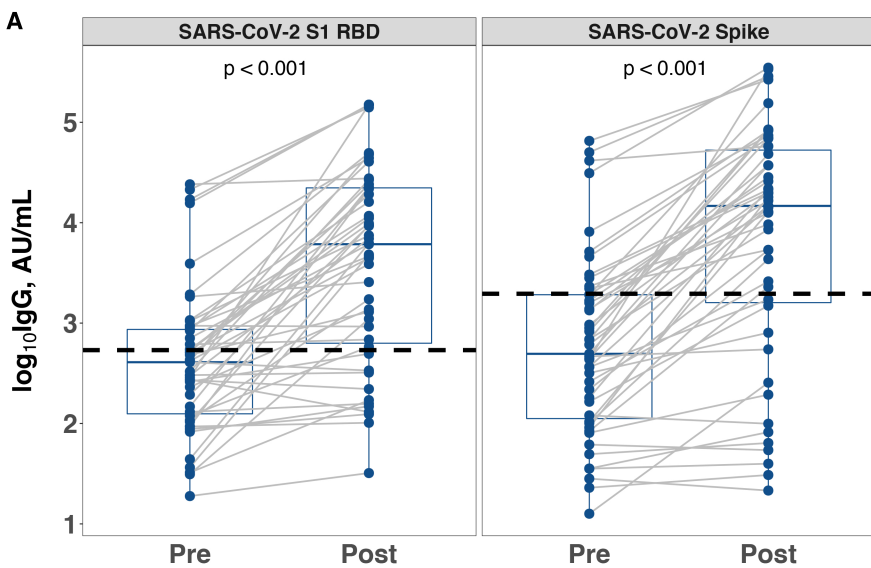
482

483 REFERENCES

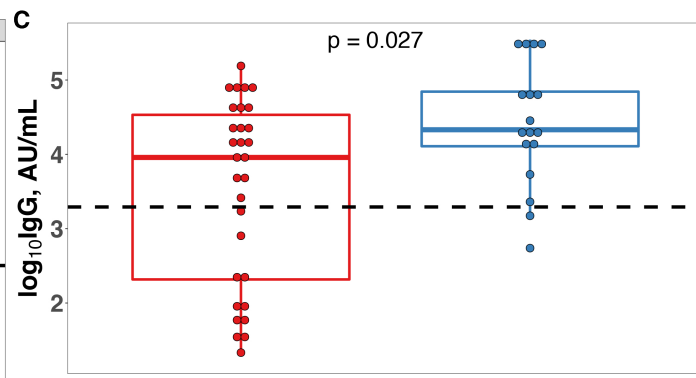
- 484 1. Fung M, Babik JM. COVID-19 in Immunocompromised Hosts: What We Know So
485 Far. *Clinical Infectious Diseases*. 2021;72(2):340-350. doi:10.1093/cid/ciaa863
- 486 2. COVID-19 Map. Johns Hopkins Coronavirus Resource Center. Accessed June 15,
487 2020. <https://coronavirus.jhu.edu/map.html>
- 488 3. Raja MA, Mendoza MA, Villavicencio A, et al. COVID-19 in solid organ transplant
489 recipients: A systematic review and meta-analysis of current literature.
490 *Transplantation Reviews*. 2021;35(1):100588. doi:10.1016/j.trre.2020.100588
- 491 4. Baden LR, El Sahly HM, Essink B, et al. Efficacy and Safety of the mRNA-1273
492 SARS-CoV-2 Vaccine. *New England Journal of Medicine*. 2021;384(5):403-416.
493 doi:10.1056/NEJMoa2035389
- 494 5. Polack FP, Thomas SJ, Kitchin N, et al. Safety and Efficacy of the BNT162b2
495 mRNA Covid-19 Vaccine. *New England Journal of Medicine*. 2020;383(27):2603-
496 2615. doi:10.1056/NEJMoa2034577
- 497 6. Aslam S, Adler E, Mekeel K, Little SJ. Clinical effectiveness of COVID-19
498 vaccination in solid organ transplant recipients. *Transplant Infectious Disease*.
499 2021;n/a(n/a):e13705. doi:10.1111/tid.13705
- 500 7. Qin CX, Moore LW, Anjan S, et al. Risk of Breakthrough SARS-CoV-2 Infections in
501 Adult Transplant Recipients. *Transplantation*. Published online July 23, 2021.
502 doi:10.1097/TP.0000000000003907
- 503 8. Boyarsky BJ, Werbel WA, Avery RK, et al. Immunogenicity of a Single Dose of
504 SARS-CoV-2 Messenger RNA Vaccine in Solid Organ Transplant Recipients.
505 *JAMA*. 2021;325(17):1784-1786. doi:10.1001/jama.2021.4385
- 506 9. Boyarsky BJ, Werbel WA, Avery RK, et al. Antibody Response to 2-Dose SARS-
507 CoV-2 mRNA Vaccine Series in Solid Organ Transplant Recipients. *JAMA*.
508 2021;325(21):2204-2206. doi:10.1001/jama.2021.7489
- 509 10. Hall VG, Ferreira VH, Ierullo M, et al. Humoral and cellular immune response and
510 safety of two-dose SARS-CoV-2 mRNA-1273 vaccine in solid organ transplant
511 recipients. *American Journal of Transplantation*. 2021;n/a(n/a).
512 doi:10.1111/ajt.16766
- 513 11. Sattler A, Schrezenmeier E, Weber UA, et al. Impaired humoral and cellular
514 immunity after SARS-CoV2 BNT162b2 (Tozinameran) prime-boost vaccination in
515 kidney transplant recipients. *J Clin Invest*. Published online June 8, 2021.
516 doi:10.1172/JCI150175

- 517 12. Hall VG, Ferreira VH, Ku T, et al. Randomized Trial of a Third Dose of mRNA-1273
518 Vaccine in Transplant Recipients. *N Engl J Med*. Published online August 11, 2021.
519 doi:10.1056/NEJMc2111462
- 520 13. Kamar N, Abravanel F, Marion O, Couat C, Izopet J, Del Bello A. Three Doses of
521 an mRNA Covid-19 Vaccine in Solid-Organ Transplant Recipients. *New England*
522 *Journal of Medicine*. 2021;0(0):null. doi:10.1056/NEJMc2108861
- 523 14. Benotmane I, Gautier G, Perrin P, et al. Antibody Response After a Third Dose of
524 the mRNA-1273 SARS-CoV-2 Vaccine in Kidney Transplant Recipients With
525 Minimal Serologic Response to 2 Doses. *JAMA*. 2021;326(11):1063.
526 doi:10.1001/jama.2021.12339
- 527 15. CDC. COVID Data Tracker. Centers for Disease Control and Prevention. Published
528 March 28, 2020. Accessed August 6, 2021. <https://covid.cdc.gov/covid-data-tracker>
- 529 16. Werbel WA, Boyarsky BJ, Ou MT, et al. Safety and Immunogenicity of a Third
530 Dose of SARS-CoV-2 Vaccine in Solid Organ Transplant Recipients: A Case
531 Series. *Ann Intern Med*. Published online June 15, 2021. doi:10.7326/L21-0282
- 532 17. Woldemeskel BA, Karaba AH, Garliss CC, et al. The BNT162b2 mRNA Vaccine
533 Elicits Robust Humoral and Cellular Immune Responses in People Living with HIV.
534 *Clinical Infectious Diseases*. 2021;(ciab648). doi:10.1093/cid/ciab648
- 535 18. Matsuyama S, Nao N, Shirato K, et al. Enhanced isolation of SARS-CoV-2 by
536 TMPRSS2-expressing cells. *Proc Natl Acad Sci USA*. 2020;117(13):7001.
537 doi:10.1073/pnas.2002589117
- 538 19. Waggoner JJ, Stittleburg V, Pond R, et al. Triplex Real-Time RT-PCR for Severe
539 Acute Respiratory Syndrome Coronavirus 2. *Emerg Infect Dis*. 2020;26(7):1633-
540 1635. doi:10.3201/eid2607.201285
- 541 20. Schaecher SR, Touchette E, Schriewer J, Buller RM, Pekosz A. Severe Acute
542 Respiratory Syndrome Coronavirus Gene 7 Products Contribute to Virus-Induced
543 Apoptosis. *Journal of Virology*. 2007;81(20):11054-11068. doi:10.1128/JVI.01266-
544 07
- 545 21. Schaecher SR, Mackenzie JM, Pekosz A. The ORF7b Protein of Severe Acute
546 Respiratory Syndrome Coronavirus (SARS-CoV) Is Expressed in Virus-Infected
547 Cells and Incorporated into SARS-CoV Particles. *Journal of Virology*.
548 2007;81(2):718-731. doi:10.1128/JVI.01691-06
- 549 22. Schaecher SR, Stabenow J, Oberle C, et al. An immunosuppressed Syrian golden
550 hamster model for SARS-CoV infection. *Virology*. 2008;380(2):312-321.
551 doi:10.1016/j.virol.2008.07.026

552



Cohort ● HCs ● SOTRs



Organ Transplanted ● Kidney ● Other

