

Article

Decreased Global EEG Synchronization in Amyloid Positive Mild Cognitive Impairment and Alzheimer's Disease Patients—Relationship to *APOE* $\epsilon 4$

Una Smailovic^{1,2,*}, Charlotte Johansson^{3,4}, Thomas Koenig⁵, Ingemar Kåreholt^{6,7}, Caroline Graff^{3,8} and Vesna Jelic^{1,4}

- ¹ Division of Clinical Geriatrics, Center for Alzheimer Research, Department of Neurobiology, Care Sciences and Society, Karolinska Institutet, 14152 Huddinge, Sweden; Vesna.Jelic@ki.se
- ² Department of Clinical Neurophysiology, Karolinska University Hospital, 14186 Huddinge, Sweden
- ³ Division of Neurogeriatrics, Center for Alzheimer Research, Department of Neurobiology, Care Sciences and Society, Karolinska Institutet, 14152 Huddinge, Sweden; charlotte.johansson@ki.se (C.J.); caroline.graff@ki.se (C.G.)
- ⁴ Clinic for Cognitive Disorders, Karolinska University Hospital, 14186 Huddinge, Sweden
- ⁵ Translational Research Center, University Hospital of Psychiatry, University of Bern, 3012 Bern, Switzerland; thomas.koenig@upd.unibe.ch
- ⁶ Aging Research Centre, Karolinska Institutet and Stockholm University, 17165 Solna, Sweden; ingemar.kareholt@ki.se
- ⁷ School of Health and Welfare, Aging Research Network—Jönköping (ARN-J), Institute for Gerontology, Jönköping University, 55111 Jönköping, Sweden
- ⁸ Unit for Hereditary Dementia, Karolinska University Hospital-Solna, 17176 Solna, Sweden
- * Correspondence: una.smailovic@ki.se



Citation: Smailovic, U.; Johansson, C.; Koenig, T.; Kåreholt, I.; Graff, C.; Jelic, V. Decreased Global EEG Synchronization in Amyloid Positive Mild Cognitive Impairment and Alzheimer's Disease Patients—Relationship to *APOE* $\epsilon 4$. *Brain Sci.* **2021**, *11*, 1359. <https://doi.org/10.3390/brainsci11101359>

Academic Editors: Jesús Poza and Carlos Gómez

Received: 3 September 2021

Accepted: 8 October 2021

Published: 16 October 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Abstract: The apolipoprotein E (*APOE*) $\epsilon 4$ allele is a risk factor for Alzheimer's disease (AD) that has been linked to changes in brain structure and function as well as to different biological subtypes of the disease. The present study aimed to investigate the association of *APOE* $\epsilon 4$ genotypes with brain functional impairment, as assessed by quantitative EEG (qEEG) in patients on the AD continuum. The study population included 101 amyloid positive patients diagnosed with mild cognitive impairment (MCI) ($n = 50$) and AD ($n = 51$) that underwent resting-state EEG recording and CSF A β 42 analysis. In total, 31 patients were *APOE* $\epsilon 4$ non-carriers, 42 were carriers of one, and 28 were carriers of two *APOE* $\epsilon 4$ alleles. Quantitative EEG analysis included computation of the global field power (GFP) and global field synchronization (GFS) in conventional frequency bands. Amyloid positive patients who were carriers of *APOE* $\epsilon 4$ allele(s) had significantly higher GFP beta and significantly lower GFS in theta and beta bands compared to *APOE* $\epsilon 4$ non-carriers. Increased global EEG power in beta band in *APOE* $\epsilon 4$ carriers may represent a brain functional compensatory mechanism that offsets global EEG slowing in AD patients. Our findings suggest that decreased EEG measures of global synchronization in theta and beta bands reflect brain functional deficits related to the *APOE* $\epsilon 4$ genotype in patients that are on a biomarker-verified AD continuum.

Keywords: quantitative electroencephalography; apolipoprotein E; Alzheimer's disease; mild cognitive impairment; amyloid pathology

1. Introduction

Alzheimer's disease (AD) is the most common form of dementia with distinct neuropathological changes and a heterogeneous clinical presentation. The disease is neuropathologically characterized by the gradual accumulation of amyloid- β 42 (A β 42) and hyperphosphorylated tau proteins in the brain tissue in the form of senile plaques and neurofibrillary tangles, respectively [1,2]. However, AD can present with different clinical symptomatology in its typical and atypical forms, depending on whether memory or other specific cognitive domains are affected [3]. These individual differences in the vulnerability

of the human brain to the AD-associated neuropathology depend on several elements, including genetic predisposition, environmental effect, and lifestyle factors [4–6].

Even though an increasing number of genetic variants have been associated with AD [6], the apolipoprotein $\epsilon 4$ (*APOE*) genotype remains the main known genetic risk factor for the more common sporadic form of AD [7,8]. The *APOE* gene encodes three alleles— $\epsilon 2$, $\epsilon 3$, and $\epsilon 4$ —with an allele-frequency ranging from 4–8%, 72–87%, and 9–19%, respectively, among the healthy population worldwide [7]. An extensive meta-analysis demonstrated that heterozygous and homozygous *APOE* $\epsilon 4$ carriers have a 1.1–5.6- and 2.2–33.1-fold increased risk, respectively, for developing AD compared to non-carriers [7]. The mechanism by which *APOE* $\epsilon 4$ exerts these effects remains to be fully elucidated; however, its physiological role in generating and maintaining synapses suggests impaired synaptic connections as the contributing factor, especially in the event of brain injury [8,9]. *APOE* $\epsilon 4$ has been additionally related to increased $A\beta$ accumulation [10], altered $A\beta$ clearance [11], interaction with tau protein [12], and neuroinflammation in brain tissue [13].

Previous studies have shown that *APOE* $\epsilon 4$ affects brain functional activity and connectivity, even before the presence of $A\beta$ pathology in the brain tissue [14,15]. These findings highlight the relationship between genetic factors and brain functional changes in subjects that are at increased risk for dementia. The heritability and genetic origin of brain functioning have been particularly emphasized by studies employing electroencephalography (EEG) [16–18].

EEG is a non-invasive diagnostic procedure that registers brain activity in real time [19] and that can be further used to quantify and localize changes in the temporal and spatial organization of brain neuronal networks [20]. Quantitative EEG (qEEG), therefore, provides objective and comprehensive assessment of brain functional activity commonly analyzed across its slow (delta and theta) and fast (alpha and beta) frequency bands that are thought to reflect distinct physiological processes [19]. Individual differences in these EEG signals were shown to have strong genetic origin [18], including a high concordance of EEG signals between identical twins [16,21,22].

A number of studies have assessed qEEG changes in patients with cognitive impairment and AD, coming from a standpoint that AD is a disease of synaptic failure [23,24] and that EEG is the only available method that can directly mirror synaptic activity with a millisecond time resolution [25]. The most consistent qEEG findings in patients on the clinical AD continuum include generalized EEG-slowing and reduced EEG synchronization in fast frequency bands [26–31]. These qEEG changes have been additionally related to the severity of cognitive impairment [32–35], future cognitive decline and progression to dementia [27,36,37], as well as with a cerebrospinal fluid (CSF) profile of AD biomarker changes [38,39].

Several studies have further addressed a potential relationship between qEEG changes and the *APOE* status in patients with AD; however, the results have been conflicting thus far, reporting accentuated EEG slowing in *APOE* $\epsilon 4$ carriers [40,41], more severe EEG slowing in *APOE* $\epsilon 4$ non-carriers [42], and no differences in relation to the *APOE* status [43]. Notably, these studies involved patients whose diagnoses were based on clinical criteria and without the biomarker evidence of AD pathology, which, considering the pathophysiological heterogeneity of cognitive disorders, may have affected the interpretation of results.

The aim of the present study was to investigate relationship between *APOE* genotype and brain functional impairment in memory clinic patients on a biomarker-verified AD continuum. We employed two qEEG measures of global power and synchronization that have been utilized previously in studies on AD and cognitive disorders [28,29,34,38]. Our hypothesis was that the *APOE* genotype has an intrinsic effect on brain oscillatory activity in MCI and dementia patients that have positive biomarkers of AD pathology. Furthermore, we investigated whether comprehensive qEEG analyses, including measures of both power and synchronization of EEG oscillations, may provide complementary information on these brain functional changes.

2. Materials and Methods

2.1. Study Population

The study population included in total 101 patients clinically diagnosed with MCI ($n = 50$) according to the Winblad et al., 2004 criteria [44] or with AD ($n = 51$) according to the ICD-10 criteria [45]. All patients were recruited at the Clinic for Cognitive Disorders, Karolinska University Hospital Huddinge, Stockholm, Sweden and underwent comprehensive clinical assessment, computed tomography (CT) and/or magnetic resonance brain imaging (MRI), resting-state EEG recording, CSF sampling, analysis of AD biomarkers (A β 42, phospho tau (p -tau), and total tau (t-tau)), and *APOE* genotyping of peripheral blood-DNA.

All diagnostic tests were part of the baseline cognitive assessment and patients were, therefore, drug naïve with respect to AD medication. The severity of cognitive impairment was, among other tests, assessed using the Mini-Mental State Examination (MMSE) [46]. The exclusion criteria included the presence of any significant psychiatric or neurological comorbidity, history of brain trauma, use of antiepileptic or neuroleptic medications, and any other dementia diagnosis.

Patients were stratified into three groups based on their *APOE* status and numbers of $\epsilon 4$ alleles including *APOE* $\epsilon 4$ non-carriers ($n = 31$), *APOE* $\epsilon 4$ heterozygous carriers—one allele ($n = 42$) and *APOE* $\epsilon 4$ homozygous carriers—two alleles ($n = 28$). All MCI and AD patients included in this study were amyloid positive according to their CSF A β 42 levels and, therefore, meet the research and biomarker criteria for Alzheimer's disease continuum [3,47]. MCI and AD patients were pooled in the following analyses due to a limited number of patients that prevented separate analyses of diagnostic groups. The number of MCI and AD patients within each *APOE* genotype group are presented in Table 1.

Table 1. Demographic and clinical characteristics in *APOE* $\epsilon 4$ non-carriers and carriers.

	<i>APOE</i> $\epsilon 4$ Non-Carriers	<i>APOE</i> $\epsilon 4$ Heterozygous Carriers (One Allele)	<i>APOE</i> $\epsilon 4$ Homozygous Carriers (Two Alleles)	<i>p</i> -Value
N (total)	31	42	28	
MCI	13	22	15	
AD	18	20	13	
Age (years)	65.03 \pm 9.17	65.79 \pm 8.54	64.04 \pm 5.31	0.766
Sex (M/F)	15/16	17/25	9/19	0.447
Education (years)	11.97 \pm 3.80	12.39 \pm 3.80	12.68 \pm 3.42	0.631
MMSE ^a	24.73 \pm 4.32	26.71 \pm 2.76	25.11 \pm 4.14	0.092

Data presented as the means \pm standard deviation. ^a Missing values for MMSE variables: one patient per each *APOE* $\epsilon 4$ group. Independent-Samples Kruskal–Wallis Test and Chi-Square test over the three *APOE* genotype groups as appropriate. AD = Alzheimer's disease; *APOE* = Apolipoprotein E; CSF = cerebrospinal fluid; MCI = mild cognitive impairment; and MMSE = Mini Mental State Examination.

2.2. CSF Analysis

CSF samples were obtained by a standard lumbar puncture procedure between the L3/L4 or L4/L5 intervertebral space. All CSF samples were sampled using a 25-gauge needle, collected in 12 mL polypropylene tube, centrifuged at 1000 rpm (10 min), and frozen at -70 °C. Conventional CSF markers of AD pathology, including A β 42, p -tau, and t-tau protein concentrations, were analyzed at the clinical chemistry laboratory Karolinska University Hospital, Huddinge, using xMAP technology and the INNO-BIA AlzBio3 kit (Innogenetics) [48]. Patients on the AD continuum present with decreased CSF A β 42 levels, which is thought to reflect increased deposition and reduced clearance of A β 42 into the CSF [49]. The clinical cut-off for pathological CSF levels, defined by Karolinska University Hospital laboratory, was CSFA β 42 levels <550 ng/L. MCI and AD patients with pathological CSF A β 42 levels were defined as amyloid positive in the present study.

2.3. Resting-State EEG Recordings and Analyses

All patients underwent resting-state EEG recordings at the Department of Clinical Neurophysiology within 6 months of the baseline clinical assessment. EEGs were recorded on the Nervus system (NicoletOne EEG Reader v5.93.0.424, Natus NicoletOne, Pleasanton, CA, USA) using 21 electrodes and standard electrode placement according to the 10/20 system. The electrode impedances were kept below 5 k Ω . The EEGs were recorded with a sampling rate of 256 Hz and band-pass filter between 0.5 and 70 Hz.

All digital EEG files were exported for research purposes. The EEG recordings were first assessed for any physiological and non-physiological artifacts as well as periods of drowsiness by visual inspection. Artifacts and periods of drowsiness were then removed by manual selection and the rejection of referred EEG segments. Eye movements and electrocardiographic artifacts were additionally removed using independent component analysis algorithm. The EEGs were first segmented in the 2-s artifact-free epochs. Next, Fast Fourier Transform (FFT) was performed on all available 2-s EEG epochs in order to translate EEG data into the frequency domain.

The present study employed two global frequency domain qEEG measures that have been validated previously in the context of cognitive impairment and AD [28,36,38] named GFP and GFS. GFP is a single and generalized measure of the strength of scalp potential fields [25,50]. It can be further employed in the frequency domain where its formula corresponds to the root mean of squared spectral amplitudes across all EEG channels and, therefore, summarizes global EEG power across pre-defined frequency bands [25,28,38]. GFS is a global measure of brain functional connectivity that reflects the phase synchrony of EEG oscillations across all electrode sites [51]. In more detail, FFT analysis of EEG epochs yields, for a given frequency, sine and cosine coefficients of all electrodes that can be entered into a sine–cosine diagram. Multichannel-EEG recording, at a given time point or time epoch, can, therefore, be presented as a cloud spread of electrode points in the sine–cosine diagram. For the computation of GFS at a given frequency, these points were submitted to a principal component analysis (PCA): GFS is then defined as the ratio of the two resulting PCA eigenvalues. The more of the variance is explained by the first principal component, the more of the electrode entry points approximate the straight line and the closer GFS is to 1, which would imply that all EEG sources observable at the given frequency oscillated either in phase or in antiphase. GFS is, therefore, a measure of a common phase of EEG oscillations at all electrode sites at a certain frequency point and can obtain a value between 0 and 1. Both GFP and GFS measures were averaged over all EEG epochs and conventional frequency bands were defined as delta (1–3.5 Hz), theta (4–7.5 Hz), alpha (8–11.5 Hz), and beta (12–19.5 Hz). EEG preprocessing and quantitative analysis were performed in Brain Vision Analyzer, version 2.0, software (Gilching, Germany).

2.4. Statistical Analysis

Statistical analyses were performed in SPSS (version 26, IBM, New York, NY, USA) and STATA (version 16.1, StataCorp LLC, College Station, TX, USA). Demographics (age, gender, and education) and MMSE levels were compared between *APOE* $\epsilon 4$ non-carriers, heterozygous, and homozygous carriers using the Independent-samples Kruskal–Wallis Test for continuous variables and Chi-Square test for categorical variables.

qEEG measure GFP in all frequency bands was transformed with zero-skewness natural logarithmic transformation in order to obtain non-skewed data distributions. In the figures, the differences in GFP and GFS across *APOE* groups were presented using the original untransformed data. P-values are based on one-way analysis of variance (ANOVA) on the transformed variables. Since the assumption of homogeneity of variance was not met for all GFP/GFS variables (Levene's test < 0.05), the *p*-values from Welch's ANOVA were reported. Welch's ANOVA was run separately for GFP/GFS measures in each frequency band. The level of statistical significance was $p < 0.05$.

3. Results

3.1. Demographics and Clinical Characteristics

Demographics and clinical characteristics of our study population and across the three *APOE* $\epsilon 4$ genotype groups (non-carriers, carriers with one allele, and carriers with two alleles) are presented in Table 1. There were no statistically significant differences in age ($p = 0.766$), education ($p = 0.631$), or distribution of females versus males ($p = 0.447$) between the three groups. The *APOE* $\epsilon 4$ non-carriers, heterozygous, and homozygous carriers did not differ significantly in the global severity of cognitive impairment as assessed by MMSE ($p = 0.092$).

3.2. Relationship between Global EEG Power and *APOE* Genotype in Amyloid Positive MCI and AD Patients

GFP medians and interquartile ranges in four conventional frequency bands across *APOE* $\epsilon 4$ genotype groups are presented in Figure 1. One outlying GFP alpha data point from an *APOE* $\epsilon 4$ heterozygous carrier was excluded from the analysis (GFP alpha > 2 μV). There was a statistically significant gradient-like increase in GFP beta ($p = 0.001$) in amyloid positive *APOE* $\epsilon 4$ heterozygous and homozygous carriers compared to non-carriers (Figure 1). Interestingly, there were no significant differences in GFP in delta ($p = 0.065$), theta ($p = 0.491$), or alpha bands ($p = 0.084$) between amyloid positive *APOE* genotype groups (Figure 1).

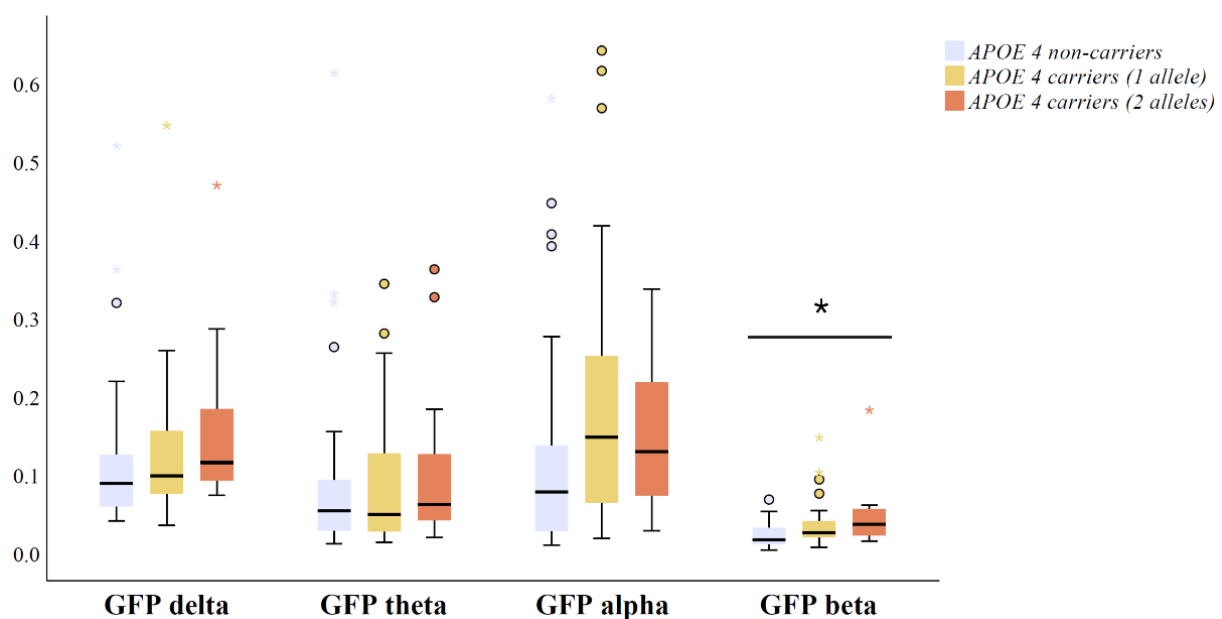


Figure 1. Differences in qEEG measures of global field power (GFP) between *APOE* $\epsilon 4$ carriers with two alleles ($n = 28$), carriers with one allele ($n = 42$) and non-carriers ($n = 31$) including CSF A β 42 positive (<550 ng/L) MCI and AD patients. The original data on GFP are presented as the median (solid line), interquartile range (box), and minimum and maximum values (whiskers) across four conventional frequency bands. p -Values are based on ANOVA over the three genotype groups using GFP measures transformed with zero skewness natural log-transformation; * $p < 0.05$. Outlier and extreme values are denoted as circles and stars, respectively. Abbreviations: ANOVA = analysis of variance; and *APOE* = apolipoprotein E.

3.3. Relationship between Global EEG Synchronization and *APOE* Genotype in Amyloid Positive MCI and AD Patients

GFS medians and interquartile ranges in four conventional frequency bands across *APOE* genotype groups are presented in Figure 2. Amyloid positive *APOE* $\epsilon 4$ non-carriers, heterozygous and homozygous carriers differed in GFS measure in theta ($p = 0.041$) and beta bands ($p = 0.036$). That is, *APOE* $\epsilon 4$ carriers exhibited lower EEG synchronization in theta and beta bands compared to non-carriers (Figure 2). Even though a trend was

observed, the differences in GFS in delta ($p = 0.090$) and alpha ($p = 0.079$) band did not reach statistical significance (Figure 2).

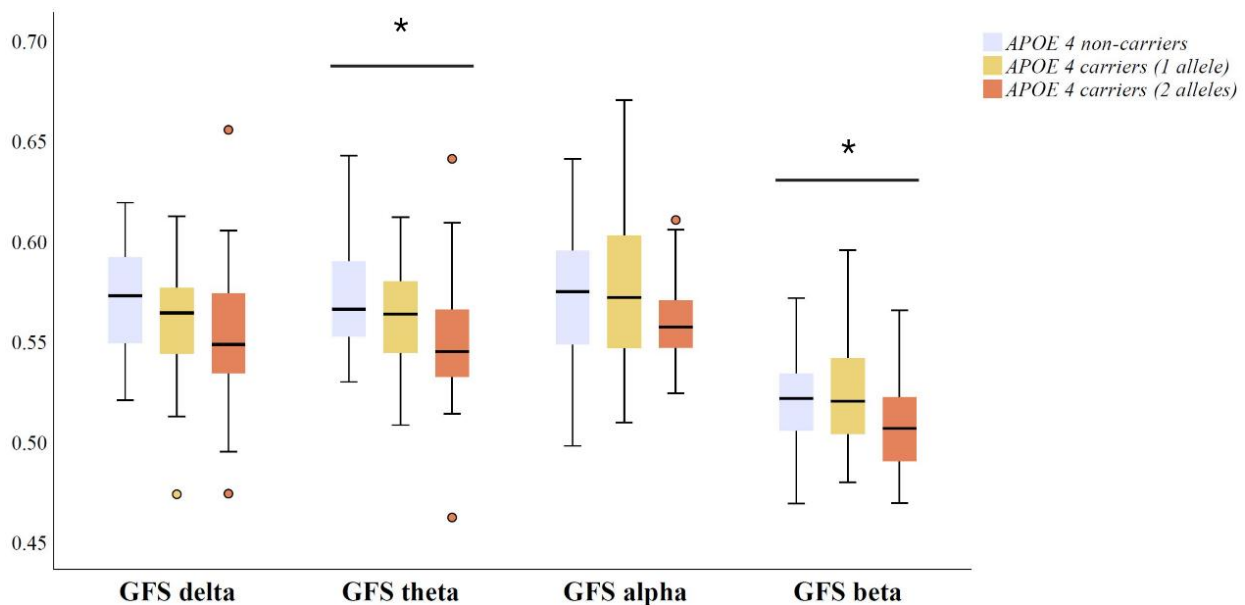


Figure 2. Differences in qEEG measures of global field synchronization (GFS) between *APOE* $\epsilon 4$ carriers with two alleles ($n = 28$), carriers with one allele ($n = 42$) and non-carriers ($n = 31$), including CSF A β 42 positive (<550 ng/L) MCI and AD patients. The original data on GFS are presented as the median (solid line), interquartile range (box), and minimum and maximum values (whiskers) across four conventional frequency bands. p -Values are based on ANOVA over the three genotype groups; * $p < 0.05$. Outlier and extreme values are denoted as circles and stars, respectively. Abbreviations: ANOVA = analysis of variance; and *APOE* = apolipoprotein E.

4. Discussion

The main finding of the present study was that a decrease in qEEG measures of global brain synchronization in theta and beta bands is associated with the presence of an *APOE* $\epsilon 4$ genotype in MCI and AD patients with amyloid biomarker changes indicative of AD pathology. Additionally, amyloid positive *APOE* $\epsilon 4$ carriers exhibited an increase in global EEG power in beta band compared to non-carriers. Our study, therefore, demonstrated an association between *APOE* $\epsilon 4$ genotypes and intrinsic brain activity and connectivity, as assessed by qEEG analyses, in patients with cognitive dysfunction that are on a biomarker-verified AD continuum.

Decreased global EEG synchronization in theta and beta bands may reflect more severe brain functional impairment in patients that are on the AD continuum and carry the *APOE* $\epsilon 4$ allele compared to non-carriers. Reduced EEG synchronization in fast frequencies, including alpha and beta bands, has been supported by numerous studies involving MCI and AD patients and is consistent across various qEEG measures of brain functional connectivity [29,43,52–54]. The level of decrease in alpha and beta synchronization has been additionally associated with the disease severity [29,34,35], performance on neuropsychological tests [31], and AD biomarker changes in CSF, including a decrease in A β 42 and increase in p -tau and t-tau levels [38]. Importantly, beta activity has been related to a number of cognitive processes including working and episodic memory [55–57], language processing [58], visual perception [59], decision making [60], and attentional processes [61,62]. Therefore, alterations in the idle, “resting-state” beta activity and synchronization may be associated with inadequate engagement of beta rhythms during various cognitive tasks and, consequently, with impairment of multiple cognitive domains that is characteristic of AD dementia.

We further report that $A\beta$ positive *APOE* $\epsilon 4$ carriers exhibit lower GFS in the theta band compared to *APOE* $\epsilon 4$ non-carriers. There is an overlap between the brain areas that are typically affected in amnesic syndromes and brain regions that are thought to generate theta rhythm including hippocampus and entorhinal cortex [19]. In this context, decreased global theta synchronization in amyloid positive *APOE* $\epsilon 4$ carriers may reflect a limbic predominant pathology associated with clinical presentation of typical AD. A recent meta-analysis that addressed heterogeneity of biologically subtypes of AD reported several characteristics of limbic-predominant AD, including amnesic syndrome, late-onset sporadic presentation, and the presence of the *APOE* $\epsilon 4$ genotype [63].

Several previous studies have assessed the relationship between *APOE* status and changes in EEG power across conventional frequency bands. Most of them reported a pattern of qEEG changes that are characteristic for “EEG slowing”, including increase in delta and theta and decrease in alpha and beta power and/or amplitude in MCI and AD patients that were *APOE* $\epsilon 4$ carriers compared to non-carriers [40,41,64,65]. One of the studies reported no changes in EEG power with respect to the *APOE* $\epsilon 4$ status; however, it included a single measure of EEG power ratio and a modest number of study participants [43].

In contrast, a more recent large-scale study by de Waal and colleagues included 320 AD patients and 246 healthy controls and demonstrated higher EEG delta and theta and lower alpha power in AD patients that were *APOE* $\epsilon 4$ non-carriers compared to carriers, indicating more pronounced EEG slowing in *APOE* $\epsilon 4$ non-carriers [42]. Our results may provide some insights on these contradictory reports. We reported an increase in beta power (and a non-significant trend towards an increase in alpha and delta power) as the only significant change in global EEG power measures in amyloid positive *APOE* $\epsilon 4$ heterozygous and homozygous carriers compared to non-carriers, partly supporting recent reports from de Waal and colleagues that demonstrated less severe EEG slowing in *APOE* $\epsilon 4$ carriers. Increases in EEG beta power may appear counterintuitive in the context of qEEG changes in AD; however, these results stem from MCI and AD patient groups that have not been contrasted to healthy controls, i.e., the comparison was made to *APOE* $\epsilon 4$ negative and amyloid positive MCI and AD patients. In that regard, increased EEG beta power may reflect a mechanism of brain functional compensation in amyloid positive *APOE* $\epsilon 4$ carriers that are at increased risk for future cognitive deterioration [8,66]. Overall, our results suggest that the main drawback of previous studies is lack of biological and pathological characterization of the study population.

The increases in delta and theta power in previous studies may reflect a non-specific qEEG alterations in cognitively impaired *APOE* $\epsilon 4$ carriers, driven by a number of subjects without underlying AD pathology. As it is known from the literature and clinical practice, a number of MCI subjects will progress to other types of dementias, including dementia with Lewy bodies (DLB), Parkinson disease dementia (PDD), and vascular dementia or remain cognitively stable over time [67,68], while a substantial proportion of clinically diagnosed AD patients may have underlying vascular, mixed, or tau-related pathologies [69–71]. Interestingly, *APOE* $\epsilon 4$ has also been associated with increased risk for Lewy Body disease, including DLB and PDD [72], as well as with vascular dementia [73].

It would be of interest to further investigate these associations in cognitively healthy *APOE* $\epsilon 4$ carriers with and without evidence of AD pathology. These studies would elucidate whether *APOE* $\epsilon 4$ carriers exhibit disturbances in brain functional connectivity even before the clinical appearance of symptoms and amyloid pathology as reported by some of the functional MRI studies [14,15]. A limitation of the present study is that a rather conservative and binary cut-off for CSF amyloid positivity was used for the stratification of both MCI and AD patients. These biomarker cut-offs were initially derived from AD patient cohorts in order to clinically support a diagnosis of AD dementia, and a less stringent cut-off may be required for mild cognitive disorders [47,74].

Modification of brain activity and functional connectivity by the *APOE* genotype could further aid selection of qEEG parameters that may contribute to the prediction of AD

or even different clinico-biological AD subtypes. In the quest for such sensitive biomarkers, qEEG analysis could be extended to sleep EEG and different provocation methods during standard EEG recordings, such as hyperventilation and photostimulation. Interestingly, Ponomareva and colleagues reported that cognitively healthy relatives of AD patients that were *APOE* $\epsilon 4$ carriers had higher occurrence of synchronous delta and theta as well as sharp-waves during hyperventilation condition compared to relatives that were *APOE* $\epsilon 4$ non-carriers [65]. This study indicated that an EEG activation paradigm may be required to accentuate early brain functional impairment in cognitively healthy patients who are at a genetically increased risk for dementia.

In conclusion, our study demonstrated that decreased EEG global synchronization in theta and beta bands reflect brain functional deficits related to the *APOE* $\epsilon 4$ genotype in patients with a cognitive dysfunction and biomarker-verified AD pathology. Future studies on novel qEEG approaches and both established and new AD risk genes are required to demonstrate whether qEEG parameters could serve as potential endophenotypes for cognitive disorders due to AD.

Author Contributions: Conceptualization, U.S. and V.J.; methodology, U.S., I.K., T.K. and V.J.; software, U.S., T.K. and I.K.; formal analysis, U.S. and V.J.; investigation, U.S., V.J., C.J. and C.G.; resources, V.J. and U.S.; writing—original draft preparation, U.S.; writing—review and editing, V.J. and I.K., T.K., C.J. and C.G.; visualization, U.S.; supervision, V.J. and C.G.; funding acquisition, U.S. and V.J. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by Gun and Bertil Stohne’s Research Scholarship (US), Gun and Bertil Stohne’s Research Grant (US), Gamla Tjänarinnor grant (US, VJ), and Swedish State Support for Clinical Research (#ALF-591660) (VJ).

Institutional Review Board Statement: The study was conducted according to the guidelines of the Declaration of Helsinki and approved by the Regional Ethical Review Board in Stockholm, Sweden (Dnr: 2020-00678, 2011/1978-31/4).

Informed Consent Statement: Informed consent was obtained from all subjects involved in the study.

Data Availability Statement: The datasets used and/or analyzed during this study are available from the study’s senior and corresponding authors on reasonable request.

Acknowledgments: We thank Lars Hyllienmark and Atif Sepic from Department of Clinical Neurophysiology at Karolinska University Hospital for their kind help and support during data collection and Bengt Winblad for all the scientific discussions and kind support of our research studies.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Selkoe, D.J.; Hardy, J. The amyloid hypothesis of Alzheimer’s disease at 25 years. *EMBO Mol. Med.* **2016**, *8*, 595–608. [[CrossRef](#)] [[PubMed](#)]
2. Braak, H.; Braak, E. Neuropathological stageing of Alzheimer-related changes. *Acta Neuropathol.* **1991**, *82*, 239–259. [[CrossRef](#)]
3. Dubois, B.; Feldman, H.H.; Jacova, C.; Hampel, H.; Molinuevo, J.L.; Blennow, K.; DeKosky, S.T.; Gauthier, S.; Selkoe, D.; Bateman, R.; et al. Advancing research diagnostic criteria for Alzheimer’s disease: The IWG-2 criteria. *Lancet Neurol.* **2014**, *13*, 614–629. [[CrossRef](#)]
4. Lourida, I.; Hannon, E.; Littlejohns, T.J.; Langa, K.M.; Hyppönen, E.; Kuzma, E.; Llewellyn, D.J. Association of Lifestyle and Genetic Risk With Incidence of Dementia. *JAMA* **2019**, *322*, 430–437. [[CrossRef](#)]
5. Grant, W.B.; Campbell, A.; Itzhaki, R.F.; Savory, J. The significance of environmental factors in the etiology of Alzheimer’s disease. *J. Alzheimers Dis.* **2002**, *4*, 179–189. [[CrossRef](#)]
6. Karch, C.M.; Goate, A.M. Alzheimer’s disease risk genes and mechanisms of disease pathogenesis. *Biol. Psychiatry* **2015**, *77*, 43–51. [[CrossRef](#)]
7. Farrer, L.A.; Cupples, L.A.; Haines, J.L.; Hyman, B.; Kukull, W.A.; Mayeux, R.; Myers, R.H.; Pericak-Vance, M.A.; Risch, N.; van Duijn, C.M. Effects of age, sex, and ethnicity on the association between apolipoprotein E genotype and Alzheimer disease. A meta-analysis. APOE and Alzheimer Disease Meta Analysis Consortium. *JAMA* **1997**, *278*, 1349–1356. [[CrossRef](#)]
8. Liu, C.C.; Liu, C.C.; Kanekiyo, T.; Xu, H.; Bu, G. Apolipoprotein E and Alzheimer disease: Risk, mechanisms and therapy. *Nat. Rev. Neurol.* **2013**, *9*, 106–118. [[CrossRef](#)]

9. Kim, J.; Basak, J.M.; Holtzman, D.M. The role of apolipoprotein E in Alzheimer's disease. *Neuron* **2009**, *63*, 287–303. [[CrossRef](#)] [[PubMed](#)]
10. Reiman, E.M.; Chen, K.; Liu, X.; Bandy, D.; Yu, M.; Lee, W.; Ayutyanont, N.; Keppler, J.; Reeder, S.A.; Langbaum, J.B.; et al. Fibrillar amyloid-beta burden in cognitively normal people at 3 levels of genetic risk for Alzheimer's disease. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 6820–6825. [[CrossRef](#)] [[PubMed](#)]
11. Castellano, J.M.; Kim, J.; Stewart, F.R.; Jiang, H.; DeMattos, R.B.; Patterson, B.W.; Fagan, A.M.; Morris, J.C.; Mawuenyega, K.G.; Cruchaga, C.; et al. Human apoE isoforms differentially regulate brain amyloid-beta peptide clearance. *Sci. Transl. Med.* **2011**, *3*, 89ra57. [[CrossRef](#)]
12. Brecht, W.J.; Harris, F.M.; Chang, S.; Tesseur, I.; Yu, G.Q.; Xu, Q.; Dee Fish, J.; Wyss-Coray, T.; Buttini, M.; Mucke, L.; et al. Neuron-specific apolipoprotein e4 proteolysis is associated with increased tau phosphorylation in brains of transgenic mice. *J. Neurosci.* **2004**, *24*, 2527–2534. [[CrossRef](#)] [[PubMed](#)]
13. Lynch, J.R.; Morgan, D.; Mance, J.; Matthew, W.D.; Laskowitz, D.T. Apolipoprotein E modulates glial activation and the endogenous central nervous system inflammatory response. *J. Neuroimmunol.* **2001**, *114*, 107–113. [[CrossRef](#)]
14. Sheline, Y.I.; Morris, J.C.; Snyder, A.Z.; Price, J.L.; Yan, Z.; D'Angelo, G.; Liu, C.; Dixit, S.; Benzinger, T.; Fagan, A.; et al. APOE4 allele disrupts resting state fMRI connectivity in the absence of amyloid plaques or decreased CSF A β 42. *J. Neurosci.* **2010**, *30*, 17035–17040. [[CrossRef](#)] [[PubMed](#)]
15. Filippini, N.; MacIntosh, B.J.; Hough, M.G.; Goodwin, G.M.; Frisoni, G.B.; Smith, S.M.; Matthews, P.M.; Beckmann, C.F.; Mackay, C.E. Distinct patterns of brain activity in young carriers of the APOE- ϵ 4 allele. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 7209–7214. [[CrossRef](#)] [[PubMed](#)]
16. Stassen, H.H.; Lykken, D.T.; Propping, P.; Bomben, G. Genetic determination of the human EEG. *Hum. Genet.* **1988**, *80*, 165–176. [[CrossRef](#)]
17. Lykken, D.T.; Tellegen, A.; Thorkelson, K. Genetic determination of EEG frequency spectra. *Biol. Psychol.* **1974**, *1*, 245–259. [[CrossRef](#)]
18. Smit, D.J.; Posthuma, D.; Boomsma, D.I.; Geus, E.J. Heritability of background EEG across the power spectrum. *Psychophysiology* **2005**, *42*, 691–697. [[CrossRef](#)]
19. Schomer, D.L.; Lopes da Silva, F. *Niedermeyer's Electroencephalography: Basic Principles, Clinical Applications, and Related Fields*; Wolters Kluwer Health: Philadelphia, PA, USA, 2015.
20. Smailovic, U.; Jelic, V. Neurophysiological Markers of Alzheimer's Disease: Quantitative EEG Approach. *Neurol. Ther.* **2019**, *8*, 37–55. [[CrossRef](#)]
21. van Beijsterveldt, C.E.; Molenaar, P.C.; de Geus, E.J.; Boomsma, D.I. Heritability of human brain functioning as assessed by electroencephalography. *Am. J. Hum. Genet.* **1996**, *58*, 562–573.
22. van Beijsterveldt, C.E.; van Baal, G.C. Twin and family studies of the human electroencephalogram: A review and a meta-analysis. *Biol. Psychol.* **2002**, *61*, 111–138. [[CrossRef](#)]
23. Selkoe, D.J. Alzheimer's disease is a synaptic failure. *Science* **2002**, *298*, 789–791. [[CrossRef](#)] [[PubMed](#)]
24. Koffie, R.M.; Hyman, B.T.; Spiers-Jones, T.L. Alzheimer's disease: Synapses gone cold. *Mol. Neurodegener.* **2011**, *6*, 63. [[CrossRef](#)] [[PubMed](#)]
25. Michel, C.M. *Electrical Neuroimaging*; Cambridge University Press: Cambridge, UK, 2009; 238p.
26. Dierks, T.; Ihl, R.; Frolich, L.; Maurer, K. Dementia of the Alzheimer type: Effects on the spontaneous EEG described by dipole sources. *Psychiatry Res.* **1993**, *50*, 151–162. [[CrossRef](#)]
27. Jelic, V.; Johansson, S.E.; Almkvist, O.; Shigeta, M.; Julin, P.; Nordberg, A.; Winblad, B.; Wahlund, L.O. Quantitative electroencephalography in mild cognitive impairment: Longitudinal changes and possible prediction of Alzheimer's disease. *Neurobiol. Aging* **2000**, *21*, 533–540. [[CrossRef](#)]
28. Huang, C.; Wahlund, L.; Dierks, T.; Julin, P.; Winblad, B.; Jelic, V. Discrimination of Alzheimer's disease and mild cognitive impairment by equivalent EEG sources: A cross-sectional and longitudinal study. *Clin. Neurophysiol.* **2000**, *111*, 1961–1967. [[CrossRef](#)]
29. Koenig, T.; Prichep, L.; Dierks, T.; Hubl, D.; Wahlund, L.O.; John, E.R.; Jelic, V. Decreased EEG synchronization in Alzheimer's disease and mild cognitive impairment. *Neurobiol. Aging* **2005**, *26*, 165–171. [[CrossRef](#)]
30. Jelic, V.; Shigeta, M.; Julin, P.; Almkvist, O.; Winblad, B.; Wahlund, L.O. Quantitative electroencephalography power and coherence in Alzheimer's disease and mild cognitive impairment. *Dementia* **1996**, *7*, 314–323. [[CrossRef](#)]
31. Adler, G.; Brassens, S.; Jajcevic, A. EEG coherence in Alzheimer's dementia. *J. Neural Transm.* **2003**, *110*, 1051–1058. [[CrossRef](#)]
32. Pozzi, D.; Petracchi, M.; Sabe, L.; Golimstock, A.; Garcia, H.; Starkstein, S. Quantified electroencephalographic correlates of neuropsychological deficits in Alzheimer's disease. *J. Neuropsychiatry Clin. Neurosci.* **1995**, *7*, 61–67. [[CrossRef](#)]
33. Kim, J.S.; Lee, S.H.; Park, G.; Kim, S.; Bae, S.M.; Kim, D.W.; Im, C.H. Clinical implications of quantitative electroencephalography and current source density in patients with Alzheimer's disease. *Brain Topogr.* **2012**, *25*, 461–474. [[CrossRef](#)]
34. Park, Y.-M.; Che, H.-J.; Im, C.-H.; Jung, H.-T.; Bae, S.-M.; Lee, S.-H. Decreased EEG synchronization and its correlation with symptom severity in Alzheimer's disease. *Neurosci. Res.* **2008**, *62*, 112–117. [[CrossRef](#)]
35. Ma, C.C.; Liu, A.J.; Liu, A.H.; Zhou, X.Y.; Zhou, S.N. Electroencephalogram global field synchronization analysis: A new method for assessing the progress of cognitive decline in Alzheimer's disease. *Clin. EEG Neurosci.* **2014**, *45*, 98–103. [[CrossRef](#)]

36. Smailovic, U.; Kåreholt, I.; Koenig, T.; Ashton, N.J.; Winblad, B.; Höglund, K.; Nilsson, P.; Zetterberg, H.; Blennow, K.; Jelic, V. Synaptic molecular and neurophysiological markers are independent predictors of progression in Alzheimer's disease. *J. Alzheimers Dis.* **2021**, *83*, 355–366. [[CrossRef](#)] [[PubMed](#)]
37. Luckhaus, C.; Grass-Kapanke, B.; Blaeser, I.; Ihl, R.; Supprian, T.; Winterer, G.; Zielasek, J.; Brinkmeyer, J. Quantitative EEG in progressing vs stable mild cognitive impairment (MCI): Results of a 1-year follow-up study. *Int. J. Geriatr. Psychiatry* **2008**, *23*, 1148–1155. [[CrossRef](#)]
38. Smailovic, U.; Koenig, T.; Kåreholt, I.; Andersson, T.; Kramberger, M.G.; Winblad, B.; Jelic, V. Quantitative EEG power and synchronization correlate with Alzheimer's disease CSF biomarkers. *Neurobiol. Aging* **2018**, *63*, 88–95. [[CrossRef](#)]
39. Stomrud, E.; Hansson, O.; Minthon, L.; Blennow, K.; Rosen, I.; Londos, E. Slowing of EEG correlates with CSF biomarkers and reduced cognitive speed in elderly with normal cognition over 4 years. *Neurobiol. Aging* **2010**, *31*, 215–223. [[CrossRef](#)] [[PubMed](#)]
40. Lehtovirta, M.; Partanen, J.; Könönen, M.; Hiltunen, J.; Helisalminen, S.; Hartikainen, P.; Riekkinen Sr, P.; Soininen, H. A Longitudinal Quantitative EEG Study of Alzheimer's Disease: Relation to Apolipoprotein E Polymorphism. *Dement. Geriatr. Cogn. Disord.* **2000**, *11*, 29–35. [[CrossRef](#)] [[PubMed](#)]
41. Babiloni, C.; Benussi, L.; Binetti, G.; Cassetta, E.; Dal Forno, G.; Del Percio, C.; Ferreri, F.; Ferri, R.; Frisoni, G.; Ghidoni, R.; et al. Apolipoprotein E and alpha brain rhythms in mild cognitive impairment: A multicentric Electroencephalogram study. *Ann. Neurol.* **2006**, *59*, 323–334. [[CrossRef](#)]
42. de Waal, H.; Stam, C.J.; de Haan, W.; van Straaten, E.C.; Blankenstein, M.A.; Scheltens, P.; van der Flier, W.M. Alzheimer's disease patients not carrying the apolipoprotein E epsilon4 allele show more severe slowing of oscillatory brain activity. *Neurobiol. Aging* **2013**, *34*, 2158–2163. [[CrossRef](#)]
43. Jelic, V.; Julin, P.; Shigeta, M.; Nordberg, A.; Lannfelt, L.; Winblad, B.; Wahlund, L.O. Apolipoprotein E epsilon4 allele decreases functional connectivity in Alzheimer's disease as measured by EEG coherence. *J. Neurol. Neurosurg. Psychiatry* **1997**, *63*, 59–65. [[CrossRef](#)]
44. Winblad, B.; Palmer, K.; Kivipelto, M.; Jelic, V.; Fratiglioni, L.; Wahlund, L.O.; Nordberg, A.; Bäckman, L.; Albert, M.; Almkvist, O.; et al. Mild cognitive impairment—beyond controversies, towards a consensus: Report of the International Working Group on Mild Cognitive Impairment. *J. Intern. Med.* **2004**, *256*, 240–246. [[CrossRef](#)]
45. World Health Organization. *The ICD-10 Classification of Mental and Behavioural Disorders: Clinical Descriptions and Diagnostic Guidelines*; WHO: Geneva, Switzerland, 1992.
46. Folstein, M.F.; Folstein, S.E.; McHugh, P.R. "Mini-mental state". A practical method for grading the cognitive state of patients for the clinician. *J. Psychiatr. Res.* **1975**, *12*, 189–198. [[CrossRef](#)]
47. Jack, C.R., Jr.; Bennett, D.A.; Blennow, K.; Carrillo, M.C.; Dunn, B.; Haeberlein, S.B.; Holtzman, D.M.; Jagust, W.; Jessen, F.; Karlawish, J.; et al. NIA-AA Research Framework: Toward a biological definition of Alzheimer's disease. *Alzheimers Dement.* **2018**, *14*, 535–562. [[CrossRef](#)]
48. Olsson, A.; Vanderstichele, H.; Andreasen, N.; De Meyer, G.; Wallin, A.; Holmberg, B.; Rosengren, L.; Vanmechelen, E.; Blennow, K. Simultaneous measurement of beta-amyloid(1-42), total tau, and phosphorylated tau (Thr181) in cerebrospinal fluid by the xMAP technology. *Clin. Chem.* **2005**, *51*, 336–345. [[CrossRef](#)]
49. Motter, R.; Vigo-Pelfrey, C.; Kholodenko, D.; Barbour, R.; Johnson-Wood, K.; Galasko, D.; Chang, L.; Miller, B.; Clark, C.; Green, R.; et al. Reduction of beta-amyloid peptide42 in the cerebrospinal fluid of patients with Alzheimer's disease. *Ann. Neurol.* **1995**, *38*, 643–648. [[CrossRef](#)]
50. Lehmann, D.; Skrandies, W. Reference-free identification of components of checkerboard-evoked multichannel potential fields. *Electroencephalogr. Clin. Neurophysiol.* **1980**, *48*, 609–621. [[CrossRef](#)]
51. Koenig, T.; Lehmann, D.; Saito, N.; Kuginuki, T.; Kinoshita, T.; Koukkou, M. Decreased functional connectivity of EEG theta-frequency activity in first-episode, neuroleptic-naive patients with schizophrenia: Preliminary results. *Schizophr. Res.* **2001**, *50*, 55–60. [[CrossRef](#)]
52. Fonseca, L.C.; Tedrus, G.M.; Prandi, L.R.; Andrade, A.C. Quantitative electroencephalography power and coherence measurements in the diagnosis of mild and moderate Alzheimer's disease. *Arquivos de Neuro-Psiquiatria* **2011**, *69*, 297–303. [[CrossRef](#)] [[PubMed](#)]
53. Stam, C.J.; Van Der Made, Y.; Pijnenburg, Y.A.L.; Scheltens, P. EEG synchronization in mild cognitive impairment and Alzheimer's disease. *Acta Neurologica Scandinavica* **2003**, *108*, 90–96. [[CrossRef](#)] [[PubMed](#)]
54. Stam, C.J.; Nolte, G.; Daffertshofer, A. Phase lag index: Assessment of functional connectivity from multi channel EEG and MEG with diminished bias from common sources. *Hum. Brain Mapp.* **2007**, *28*, 1178–1193. [[CrossRef](#)]
55. Axmacher, N.; Schmitz, D.P.; Wagner, T.; Elger, C.E.; Fell, J. Interactions between medial temporal lobe, prefrontal cortex, and inferior temporal regions during visual working memory: A combined intracranial EEG and functional magnetic resonance imaging study. *J. Neurosci.* **2008**, *28*, 7304–7312. [[CrossRef](#)] [[PubMed](#)]
56. Hanslmayr, S.; Spitzer, B.; Bäuml, K.H. Brain oscillations dissociate between semantic and nonsemantic encoding of episodic memories. *Cereb. Cortex* **2009**, *19*, 1631–1640. [[CrossRef](#)]
57. Hanslmayr, S.; Staresina, B.P.; Bowman, H. Oscillations and Episodic Memory: Addressing the Synchronization/Desynchronization Conundrum. *Trends Neurosci.* **2016**, *39*, 16–25. [[CrossRef](#)]
58. Weiss, S.; Mueller, H.M. "Too Many betas do not Spoil the Broth": The Role of Beta Brain Oscillations in Language Processing. *Front. Psychol.* **2012**, *3*, 201. [[CrossRef](#)]

59. Piantoni, G.; Kline, K.A.; Eagleman, D.M. Beta oscillations correlate with the probability of perceiving rivalrous visual stimuli. *J. Vis.* **2010**, *10*, 18. [[CrossRef](#)]
60. Wimmer, K.; Ramon, M.; Pasternak, T.; Compte, A. Transitions between Multiband Oscillatory Patterns Characterize Memory-Guided Perceptual Decisions in Prefrontal Circuits. *J. Neurosci.* **2016**, *36*, 489–505. [[CrossRef](#)] [[PubMed](#)]
61. Kamiński, J.; Brzezicka, A.; Gola, M.; Wróbel, A. β band oscillations engagement in human alertness process. *Int. J. Psychophysiol.* **2012**, *85*, 125–128. [[CrossRef](#)] [[PubMed](#)]
62. Gola, M.; Kamiński, J.; Brzezicka, A.; Wróbel, A. β band oscillations as a correlate of alertness—changes in aging. *Int. J. Psychophysiol.* **2012**, *85*, 62–67. [[CrossRef](#)] [[PubMed](#)]
63. Ferreira, D.; Nordberg, A.; Westman, E. Biological subtypes of Alzheimer disease: A systematic review and meta-analysis. *Neurology* **2020**, *94*, 436–448. [[CrossRef](#)]
64. Lehtovirta, M.; Partanen, J.; Könönen, M.; Soininen, H.; Helisalmi, S.; Mannermaa, A.; Ryyänen, M.; Hartikainen, P.; Riekkinen, P. Spectral analysis of EEG in Alzheimer's disease: Relation to apolipoprotein E polymorphism. *Neurobiol. Aging* **1996**, *17*, 523–526. [[CrossRef](#)]
65. Ponomareva, N.V.; Korovaitseva, G.I.; Rogae, E.I. EEG alterations in non-demented individuals related to apolipoprotein E genotype and to risk of Alzheimer disease. *Neurobiol. Aging* **2008**, *29*, 819–827. [[CrossRef](#)]
66. Olsson, B.; Lautner, R.; Andreasson, U.; Öhrfelt, A.; Portelius, E.; Bjerke, M.; Hölttä, M.; Rosén, C.; Olsson, C.; Strobel, G.; et al. CSF and blood biomarkers for the diagnosis of Alzheimer's disease: A systematic review and meta-analysis. *Lancet Neurol.* **2016**, *15*, 673–684. [[CrossRef](#)]
67. Mitchell, A.J.; Shiri-Feshki, M. Rate of progression of mild cognitive impairment to dementia—Meta-analysis of 41 robust inception cohort studies. *Acta Psychiatr. Scand.* **2009**, *119*, 252–265. [[CrossRef](#)] [[PubMed](#)]
68. Petersen, R.C. Mild cognitive impairment as a diagnostic entity. *J. Intern. Med.* **2004**, *256*, 183–194. [[CrossRef](#)]
69. DeTure, M.A.; Dickson, D.W. The neuropathological diagnosis of Alzheimer's disease. *Mol. Neurodegener.* **2019**, *14*, 32. [[CrossRef](#)]
70. Crary, J.F. Primary age-related tauopathy and the amyloid cascade hypothesis: The exception that proves the rule? *J. Neurol. Neuromedicine* **2016**, *1*, 53–57. [[CrossRef](#)]
71. Jack, C.R., Jr. PART and SNAP. *Acta Neuropathol.* **2014**, *128*, 773–776. [[CrossRef](#)]
72. Tsuang, D.; Leverenz, J.B.; Lopez, O.L.; Hamilton, R.L.; Bennett, D.A.; Schneider, J.A.; Buchman, A.S.; Larson, E.B.; Crane, P.K.; Kaye, J.A.; et al. APOE ϵ 4 increases risk for dementia in pure synucleinopathies. *JAMA Neurol.* **2013**, *70*, 223–228. [[CrossRef](#)]
73. Chuang, Y.F.; Hayden, K.M.; Norton, M.C.; Tschanz, J.; Breitner, J.C.; Welsh-Bohmer, K.A.; Zandi, P.P. Association between APOE epsilon4 allele and vascular dementia: The Cache County study. *Dement. Geriatr. Cogn. Disord.* **2010**, *29*, 248–253. [[CrossRef](#)]
74. Müller, E.G.; Edwin, T.H.; Stokke, C.; Navelsaker, S.S.; Babovic, A.; Bogdanovic, N.; Knapskog, A.B.; Revheim, M.E. Amyloid- β PET-Correlation with cerebrospinal fluid biomarkers and prediction of Alzheimer's disease diagnosis in a memory clinic. *PLoS ONE* **2019**, *14*, e0221365. [[CrossRef](#)] [[PubMed](#)]