



# Homeobox Genes in Cancers: From Carcinogenesis to Recent Therapeutic Intervention

Yangyang Feng<sup>1,2</sup>, Tongyue Zhang<sup>1,2</sup>, Yijun Wang<sup>1,2</sup>, Meng Xie<sup>1,2</sup>, Xiaoyu Ji<sup>1,2</sup>, Xiangyuan Luo<sup>1,2</sup>, Wenjie Huang<sup>2,3\*</sup> and Limin Xia<sup>1,2\*</sup>

<sup>1</sup> Department of Gastroenterology, Institute of Liver and Gastrointestinal Diseases, Tongji Hospital of Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China, <sup>2</sup> Hubei Key Laboratory of Hepato-Pancreato-Biliary Diseases, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China, <sup>3</sup> Hepatic Surgery Center, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China

## OPEN ACCESS

### Edited by:

Alfredo Procino,  
Rudjer Boskovic Institute, Croatia

### Reviewed by:

Manfra Michele,  
University of Basilicata, Italy  
Mohammad Hojjat-Farsangi,  
Karolinska Institutet (KI), Sweden

### \*Correspondence:

Limin Xia  
xialimin@tjh.tjmu.edu.cn  
Wenjie Huang  
huangwenjie@tjh.tjmu.edu.cn

### Specialty section:

This article was submitted to  
Cancer Molecular Targets  
and Therapeutics,  
a section of the journal  
Frontiers in Oncology

**Received:** 03 September 2021

**Accepted:** 28 September 2021

**Published:** 14 October 2021

### Citation:

Feng Y, Zhang T, Wang Y,  
Xie M, Ji X, Luo X, Huang W  
and Xia L (2021) Homeobox Genes in  
Cancers: From Carcinogenesis to  
Recent Therapeutic Intervention.  
Front. Oncol. 11:770428.  
doi: 10.3389/fonc.2021.770428

The homeobox (HOX) genes encoding an evolutionarily highly conserved family of homeodomain-containing transcriptional factors are essential for embryogenesis and tumorigenesis. HOX genes are involved in cell identity determination during early embryonic development and postnatal processes. The deregulation of HOX genes is closely associated with numerous human malignancies, highlighting the indispensable involvement in mortal cancer development. Since most HOX genes behave as oncogenes or tumor suppressors in human cancer, a better comprehension of their upstream regulators and downstream targets contributes to elucidating the function of HOX genes in cancer development. In addition, targeting HOX genes may imply therapeutic potential. Recently, novel therapies such as monoclonal antibodies targeting tyrosine receptor kinases, small molecular chemical inhibitors, and small interfering RNA strategies, are difficult to implement for targeting transcriptional factors on account of the dual function and pleiotropic nature of HOX genes-related molecular networks. This paper summarizes the current state of knowledge on the roles of HOX genes in human cancer and emphasizes the emerging importance of HOX genes as potential therapeutic targets to overcome the limitations of present cancer therapy.

**Keywords:** homeobox genes, transcription factors, therapy, biomarker, cancer progression

**Abbreviation:** NSCLC, non-small-cell lung carcinoma; CRC, colorectal cancer; PTC, papillary thyroid cancer; LSCC, laryngeal squamous cell cancer; GC, gastric cancer; HCC, hepatocellular carcinoma; FASN, fatty acid synthase; LUAD, lung adenocarcinoma; HNSCC, head and neck squamous cell carcinoma; TNBC, triple negative breast cancer; CMM, cutaneous malignant melanoma; ESCC, esophageal squamous cell carcinoma; NPC, nasopharyngeal carcinoma; CHOL, cholangiocarcinoma.

## 1 INTRODUCTION

### 1.1 General Overview

The homeobox (HOX) genes were initially discovered in 1992 with the roles in embryogenesis in *Drosophila melanogaster*, where mutations in antennapedia and bithorax clusters were observed to lead to abnormal body development (1). These mutations lead one body segment to emerge similar to another segment and is originally called the term “homeotic” mutations since the 1990s (2).

The HOX genes encode a family of transcription factors that regulate embryogenesis and morphogenesis during the embryonic period and adulthood. These genes are highly conserved and contain homologous domains in almost all eukaryotic cells (3). The HOX genes comprise a highly conserved sequence of 180–183 base pairs encoding a homeodomain of 60 or 61 amino acids helix-turn-helix motif (2). The specific structure was initially characterized by magnetic resonance spectroscopy imaging.

In the human genome, according to the sequence similarity and position correlation in the chromosome, 39 HOX family genes can be divided into 4 clusters, namely HOXA, HOXB, HOXC, HOXD.

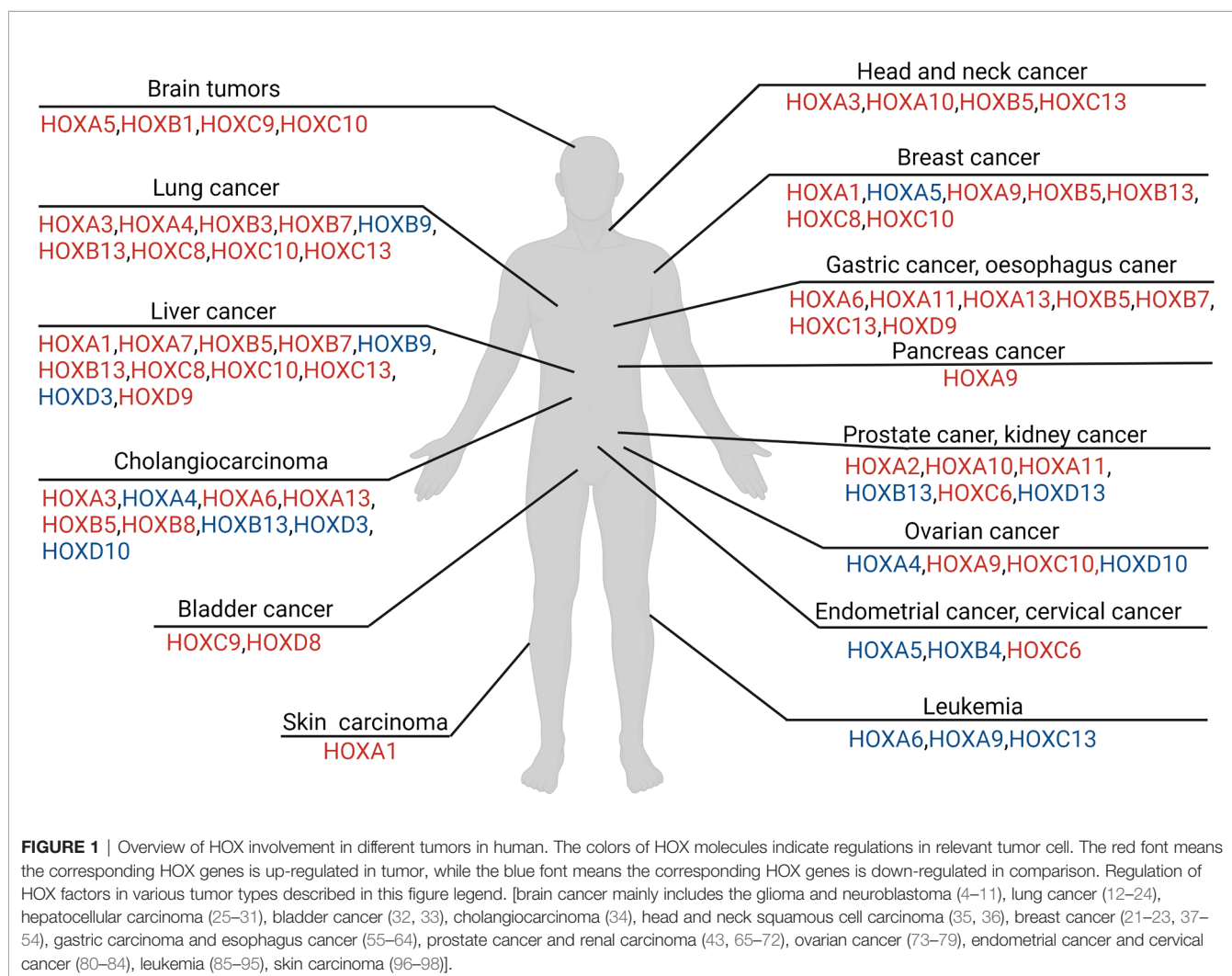
The number after the subgroup HOXA, HOXB, HOXC, HOXD increases orderly in a 3' to 5' orientation, which is shown in **Figure 1**. Each cluster has between 9 and 11 genes in a row on a homologous strand of DNA, which contains duplication and divergence of ancestral HOX genes.

### 1.2 Physiological Function of HOX Genes

HOX genes encode a family of master transcriptional regulators throughout growth and development in human tissues and organs. This period monitoring elicits distinct and temporospatial limb and organ developmental programs along the anterior-posterior axis (99). Researchers found the chromosomal arrangement of HOX genes was closely related to their localization order of genetic expression (100).

Plenty of scientific evidence suggests that the specific functions of individual HOX genes largely reflect their regional and restricted expression patterns. The disruption of the chromosome region related expression pattern may lead to developmental defects and diseases, especially human cancer (101).

HOX proteins encoded by HOX genes are key components of substantial metabolic processes such as lipidic metabolism, and their



roles in organogenesis and tumorigenesis have been studied in detail over the past decade (12, 37, 73, 102–105). Pluripotent embryonic carcinoma cells have been associated with the differentiation of a broad spectrum of tissues early in development in mice embryos (106). This process appears to be related to retinoic acid reactions, the chromosomal region determining the aforementioned relation was regulated by HOX gene (96, 107).

In conclusion, HOX genes act as primary regulators to regulate downstream target molecules (103). HOX genes play an essential role in embryonic development, including morphogenesis, organogenesis, and differentiation of a variety of tissues and organs (108).

## 2 DEREGULATION OF HOX GENES IN HUMAN CANCER

Over the past several decades, we have come to understand that a great many genes and proteins controlling embryogenesis usually partake indispensable effects in carcinogenesis likewise. Many genes identified as pivotal genes in carcinogenesis, for example, oncogenes or tumor suppressor genes, have been discovered to play primary roles in embryogenesis correspondingly. It indicates both processes are tightly linked (109). The familiar components of signal transduction pathways, such as the Sonic Hedgehog pathway, Wntless-Type MMTV Integration Site Family, Notch pathway, Paxillin pathway, and Sry-box transcription factors family members, closely act in accordance with the above rules.

This close relationship between embryogenesis and carcinogenesis supports the lineage-dependency theory. The theory proposes that the cellular mechanisms participating in lineage heredity and adaptability during development, potentially underlies tumorigenic mechanisms in humans (110). The HOX genes might be applicable to the lineage-dependency theory. Apart from the indispensable involvement of HOX genes in embryonic development in physiological status, their abnormal expression, has long been linked to tumors. Initially, these genes were thought to be related to human cancers in the hematologic system and embryo. Later, many other kinds of tumors were also noticed to be associated with deregulated HOX genes (85, 101, 111). In the context of cancer development, the abnormal expression of HOX gene may affect cell proliferation, differentiation, apoptosis, motility, angiogenesis, autophagy, and cell receptor signaling (112). HOX genes in the hallmarks of cancer are shown in **Figure 2**.

The protein products of HOX genes are initially thought to be transcription factors that accelerate cancerization since HOX proteins are deregulated in carcinogenesis. In the above process, HOX proteins govern the intricate balance of multiple signaling pathways on and off, and affect downstream targets of these pathways (113), so as to determine different cancer outcomes.

Subsequent studies have shown that HOX genes act not only as transcriptional activators but also as transcriptional repressors in cancer. This abnormal regulation of HOX genes in cancer suggests that HOX genes expression is an integral part of the regulatory network (103, 114, 115).

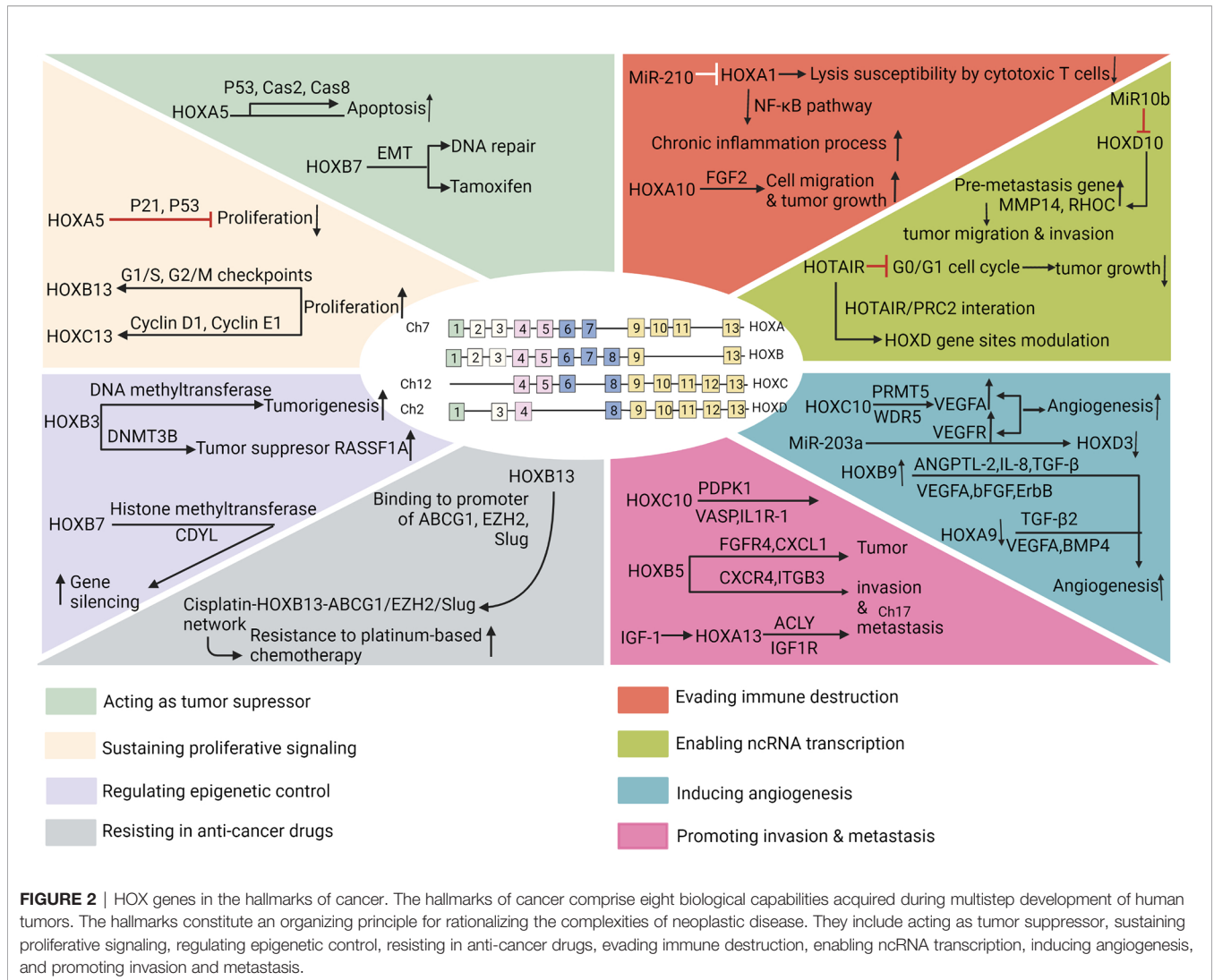
HOX genes in tumor show temporospatial deregulation pattern, different from that in normal tissues and organs. Besides, the gene dominance in expression level, that is, the aberrantly increased expression level of HOX genes in specific tissue types, the mechanism is also proposed to explain HOX genes relevant to cancer. The targets identified for HOX factors in human cancer are shown in **Table 1** (4–7, 13–20, 25–30, 32–35, 38–49, 55–62, 65–70, 74–76, 80–83, 86–91, 116–121, 127–130, 133–149).

### 2.1 Direct Role: As Oncogenes

The HOX genes have been reported to act as oncogenes and contribute to tumor progression in several types of cancer. For example, simultaneous overexpression of HOXA9 and MEIS1A induces acute myeloid leukemia in rats (92, 131). In breast cancer, HOXB7 has been reported as an oncogene because its upregulation appears to promote the expression of bFGF and induce the epithelial-mesenchymal transitions (EMT) (15, 27, 34). In colorectal cancer, HOXB5 overexpression mediated by CXCL12 chemokine ligand 12 facilitates metastasis through transactivating downstream protein CXCR4 and ITGB3 (134). HOXB13 seems to be an oncogene for ovarian and prostate cancer. Its knockout in ovarian cancer cells results in reduced tumor invasion (45, 65, 71, 138, 141). On the contrary, its ectopic expression promotes cell proliferation and non-anchoring. Mutations and growing resistance to tamoxifen-mediated apoptosis in the cell tumor antigens *P53*, *Myc*, and *Ras* are included in mouse ovarian cancer cells (43–46). In addition, HOXB13 overexpression appears to promote invasion of these types of cancers. In prostate cancer, HOXB13 regulates the prostate-derived ETS family members and also facilitates cell invasion (140). As HOXB3 in glioblastoma, its knockout leads to cancer cell suppression (150). In squamous carcinoma of the cervix, HOXC10, which is overexpressed and thought to be an oncogene, is knocked out to reduce invasiveness (6, 19, 29, 61). In colorectal cancer, HOXA13 overexpression mediated by insulin-like growth factor 1 promotes metastasis through upregulating downstream targets ATP-citrate lyase and insulin-like growth factor 1 receptor (133). Ectopic overexpression of ATP-citrate lyase and insulin-like growth factor 1 receptor rescues the decreased colorectal cancer metastasis induced by HOXA13 knockdown (133). HOXC10 overexpression mediated by Interleukin 1 $\beta$  facilitates hepatocellular carcinoma (HCC) metastasis (29). The above studies implicate HOX genes as oncogenes and prognostic biomarkers. Hence, targeting HOX genes' relevant pathways is likely to be the promising therapeutic option for clinical cancer prevention (29).

### 2.2 Direct Role: As Tumor-Suppressor Genes

Basing on our review of the literature, we found the expression of specific HOX genes in cancers tends to vary with tissue type and tumor sites. Besides, the HOX genes have been found to behave as tumor-suppressor genes and dedicate to tumor suppression in several cancers.



HOXC8 in nasopharyngeal cancer is silent and its ectopic expression causes the inhibition of tumor growth (143). *In vitro* and *in vivo*, in addition, HOXA5 is often down-regulated in breast cancer and appears to mediate apoptosis through p53 or caspases 2 and 8 in normal cells, thus having a tumor-like effect inhibition (50, 121, 122). Downregulation of HOXC9 in infant neuroblastoma appears to lead to increased cancer cell survival and tumor growth (5). HOXB1 is also significantly down-regulated in glioma cells, its knockdown promotes proliferation and invasion, inhibits apoptosis of cancer cells *in vitro*, and leads to poor survival (5, 8, 151).

The expression changes of HOXA4, HOXA9, and HOXD10 cause abnormal proliferation and differentiation of colorectal carcinoma cells and contribute to tumor development (14, 74, 87, 127). In addition, HOXB3 overexpression promotes the proliferation and invasion of glioblastoma cells, acute myeloid leukemia, pancreas, prostate, ovarian, and lung cancer (74, 93, 150, 152, 153). Excessive HOXB7 inducing MET in breast cancer cells also interferes with DNA repair and tamoxifen (136). HOXA5 down-regulation is discovered in multiple tumors,

including liposarcoma, cervical cancer, breast cancer, which suggests that HOXA5 may be an important tumor suppressor (21, 51, 84, 123). The promoter methylation and downregulation of HOXA5 during the epigenetic deregulation decrease RARβ-driven apoptosis mediated by caspase 2 and caspase 8. Therefore, HOXA5 consumption in breast cells causes a lack of epithelial cell characteristics as well as an increase of stem cell property and cellular plasticity, eventually leads to a more aggressive phenotype (119).

Therefore, the abnormal expression of the HOX genes in various cancers seems to indicate that random alterations are not necessary for the health maintenance and survival of cancer cells. Instead, abnormal expression of specific HOX genes in cancer seems to play a large part in the development of cancer.

However, HOX genes may show a predisposition. It is up-regulated or down-regulated in certain tumors, promoting cancer progression or inhibition in certain cases. Thus, they can be regarded as oncogenes or tumor suppressor genes, usually depending on the corresponding tumor microenvironment.



**TABLE 1 |** Targets identified for HOX factors in human cancer.

HOX protein	Role	Interference	Cancer type	Year	Reference
HOXA1	Oncogene	Sequester G9a/EZH2/Dnmts, sponge miR-193a-5p, via cyclin D1, via miR-100	Breast cancer, glioma, GC, lung cancer	2014,2016,2018	(38, 63, 97, 116)
HOXA3	Oncogene	Upregulate methylation level, promote differentiation to angiogenesis, confer cisplatin resistance	NSCLC, PTC, blood	2011,2019	(13, 117, 118)
HOXA4	Tumor suppressor	Downregulate $\beta$ -catenin, Cyclin D1, c-Myc and survivin, indicate inhibition of Wnt signaling, upregulate GSK3 $\beta$	Lung cancer, ovarian carcinoma	2009,2018	(14, 74, 79)
HOXA5	Tumor suppressor	Induce apoptosis mechanism mediated by cas2 and cas8; regulate E-cadherin and CD24; methylate promoter region & limit p53 expression	Breast cancer, mammary cancer, cervical cancer	2015-2021	(9, 21, 39–41, 50, 51, 80, 84, 119–126)
HOXA6	Oncogene	Coexpress with PBX2	GC, CRC, leukemia	2021	(55)
HOXA7	Oncogene	Combine to Snail promoter, cyclin E1/CDK2, activate Snail	Cervical cancer, HCC	2016,2020	(25, 81)
HOXA9	Oncogene	Pioneer factor at <i>de novo</i> enhancers and recruit CEBP $\alpha$ & MLL3/MLL4 complex, regulate BRCA1, activate JAK/STAT, induce 1GF1R expression	Pancreatic cancer, leukemia, NSCLC	2017-2020	(75, 77, 86–90, 92, 94, 127–132)
HOXA10	Oncogene	Suppress FASN transcription by forming a protein complex with AR and prevent AR recruitment to FASN gene promoter, hinder mir195	Prostate cancer, testicular cancer, HNSCC	2020	(35)
HOXA11	Oncogene	LncRNA HOXA11-AS recruit EZH2 along with the histone demethylase LSD1 or DNMT1	GC, LUAD, renal cancer	2016,2017,2018	(56, 57)
HOXA13	Oncogene	IGF-1	CRC, GC	2021	(133)
HOXB4	Tumor suppressor	Downregulate activity of Wnt/ $\beta$ -catenin signaling pathway, downregulate P-gp, MRP1 and BCRP expression	Cervical cancer, leukemia	2016,2021	(82, 91)
HOXB5	Oncogene	Transactivate CXCR4, ITGB3, FGFR4, CXCL1	HCC, CRC, breast cancer, HNSCC, GC	2015,2021	(26, 42, 134)
HOXB7	Oncogene	Reprogram to iPSC with comparable efficiency to LIN28B or c-MYC, activate TGF $\beta$ signaling pathway	Lung cancer, GC	2016,2018	(15, 27, 34, 58, 135, 136)
HOXB8	Oncogene	Instigate BACH1-mediated transcriptional cascade, viaZEB2 targets	GC, CRC	2017,2020	(59, 137)
HOXB13	Tumor suppressor	Suppress C-Myc expression to exert antitumor effects via $\beta$ -catenin/TCF4 signals, network with ABCG1/EZH2/Slug	Colon cancer, lung cancer, prostate cancer	2015-2019	(16, 43–46, 60, 65–68, 138–141)
HOXC6	Prognosis marker, oncogene	Enhance BCL2-mediated antiapoptotic effects, drive MET	Prostate cancer, cervical cancer,HCC	2019	(28, 69, 83)
HOXC8	Oncogene	Upregulate TGF $\beta$ 1, repress LMP1	NSCLC, TNBC	2018	(17, 47, 48, 142, 143)
HOXC9	Oncogene	Mediate autophagy, via mir-495/HOXC9 axis, promote multi-chemoresistance	Bladder cancer, neuroblastoma, NSCLC	2011,2015,2016	(4, 5, 32)
HOXC10	Oncogene	Via upregulating PDPK1, VASP, EMT, promote angiogenesis, induce immunosuppressive gene	HCC, lung cancer, ovarian cancer	2014-2020	(6, 7, 19, 29, 49, 61, 144, 145)
HOXC13	Oncogene	Modulate CCND1 & CCNE1	Lung adenocarcinoma	2017	(20)
HOXD3	Tumor suppressor	Inhibit HDAC1 via ITGA2 pathway & MAPK/AKT signaling	HCC, CRC	2019,2020	(146, 147)
HOXD8	Oncogene	Enhance LINC01116 contribution to progression of BCa via targeting ELK3 & HOXD8	Bladder cancer	2020	(33)
HOXD9	Oncogene	Transactivate RUFY3 & ZEB1, promote MET	GC, HCC	2015,2019	(30, 62)
HOXD10	Tumor suppressor	Inhibit RHOC/AKT/MAPK pathway, upregulate mir-10b	CRC	2019	(76, 148)
HOXD13	Tumor suppressor	Inhibit SMAD1, suppress BMP4	Prostate cancer	2021	(70)

## 2.3 Indirect Role: Epigenetic Control via HOX Genes

HOX proteins are also involved in chromatin posttranslational modifications, epigenetically affect the expression of crucial cancer progression genes. For example, HOXB3 regulates the expression of DNA methyltransferase, which can add methyl to the 5 positions of cytosine ring carbon in CpG dinucleotide (93, 152, 153). Therefore, HOX proteins can indirectly regulate the expression of other genes by controlling the expression of methyltransferase. Thus, the HOX genes may have an indirect effect on cancer by controlling the expression of DNA

methyltransferase, which seems to be of considerable relevance to understanding the mechanisms involved in tumorigenesis.

In fact, all cancer progression has common abnormalities, such as the massive epigenetic silencing of tumor suppressor genes. The mechanisms behind this phenomenon are remained to be fully understood. The DNA methyltransferase protein family consists of DNA methyltransferase (DNMT)-1, DNMT3A, DNMT3B, and DNMT3L (154). The last is associated with *de novo* methylation of DNMT3A/B, which is required during embryonic development, imprinting, and X chromosome inactivation. DNMT1 maintains the DNA methylation profile of the genome during each cell

division. DNMT3A and DNMT3B can actively add *de novo* methyl to DNA sequences to control gene expression and play a key role in cancer progression.

Palakurthy has shown that DNMT3B is the target of HOXB3 protein and is involved in the epigenetic regulation of tumor suppressor gene RASSF1A in lung adenocarcinoma and other cancers (153). HOXB3 also directly interacts with DNMT3B to promote the occurrence of leukemia in acute myeloid leukemia.

In addition, HOX genes may also play a role in post-translational modification of chromatin and affect gene expression. For instance, Heinonen has shown that HOXB7 can directly bind to chromodomain protein Y-like and enhance the histone methyltransferase activity of Polycomb Complex 2 and induce gene silencing as typical for trimethylate lysine 27 of histone H3 (155).

## 2.4 Tumor Proliferation, Invasion, and Metastasis

Proliferation phenotype has been found in various HOX-related researches, especially in leukemia, where HOX upregulated expression is often resulted from translocation mutations or altered regulation in the trithorax homologue myeloid-lymphoid leukemia (112). Invasion and metastasis phenotypes caused by abnormal HOX gene expression have been mostly studied in solid tumors, where HOX gene deregulation is usually due to loss-of-function mutations or gain-of-function mutations, or undefined mutations in upstream regulators.

In recent work, HOXC13 promotes cell proliferation by modulating the expression of cyclin D1 and cyclin E1 in lung adenocarcinoma. HOXA5 inhibits cell proliferation by regulating p53 expression in liposarcomas and p21 in non-small lung cancers (20, 63, 84, 120). Although HOXB13 overexpression promotes prostate cancer metastasis by downregulating intracellular zinc and upregulating NF-kappaB signaling pathways, HOXB13 consumption promotes proliferation of PC-3 and LNCaP cells by controlling G1/S and G2/M checkpoints (16, 60, 67, 138, 141).

HOX proteins affect cell cycle process by regulating cell cycle-related proteins (156), thus also affect proliferation and apoptosis in cancer progression. Growing evidence is noticeable that many HOX transcription factors are abnormally expressed in cancer, and their dysregulation significantly promotes tumor invasion and metastasis.

HOXD9 interacts with the promoter region of zinc-finger E-box binding homeobox (ZEB)-1, inhibition of ZEB1 induced by HOXD9 suppresses HCC cell migration, invasion as well as EMT (30). In addition, according to microarray analysis, ZEB2 may be a downstream cofactor of HOXB8 (59). Patients enrolled in studies have shown that high HOXB13 expression promotes the progression of lung adenocarcinoma and predicts poor prognosis (16).

In epithelial ovarian cancer cells, HOXA9 not only promotes the growth of epithelial ovarian cancer cells *in vivo* by activating transcriptional activity of the gene encoding transforming growth factor  $\beta$  (TGF $\beta$ )-2, but also binds to the promoter of the cadherin3 gene that encodes P-cadherin to induce

intraperitoneal dissemination (77, 94). Many reports suggest the temporospatial deregulation of HOXA9 is associated with primary tumors and specific histological subtypes. HOXA9 is of therapeutic potency, while the potency is limited by the low membrane permeability.

## 2.5 Angiogenesis

Angiogenesis is an important link during tumor progression. HOX proteins affect angiogenesis mainly by regulating VEGF expression (7). For example, HOXC10 level is statistically correlated with VEGFA expression in gliomas. HOXC10 upregulates VEGFA expression transcriptionally by binding to its promoter, and the post-translational modification of histones mediated by protein arginine methyltransferase 5 and WD repeat domain 5 is required in angiogenesis (7). MiR-203a negatively targets HOXD3 directly by targeting the VEGFR promoter region and increases VEGFR expression.

HOX protein expression also plays an important role in endothelial cells (EC) (146). For instance, HOXA5 is expressed in static EC but not in activated angiogenic EC. HOXA5 continuously increases the TSP-2 expression and decreases the VEGF expression, thereby inhibiting histopathological angiogenesis (124, 125). In addition, HOXA5 is absent in EC of proliferative hemangioma (9). HOXA5 increases the mRNA and protein expression of Akt1, further enhances Akt activity by coordinating the down-regulation of PTEN, thereby increasing the stability of the capsular patellar junction (126).

## 2.6 Resistance in Anti-Cancer Drugs

HOX proteins are involved in resistance to the anti-cancer drugs in cancers, especially in myeloid leukemia, HCC, breast cancer, and lung cancer. HOX proteins regulate various non-coding RNAs (ncRNAs) which influence cancer cells chemotherapy resistance. In particular, we take the example of the role of HOXB13 in mediating chemotherapy resistance in lung adenocarcinoma. HOXB13 upregulates a series of drug-transfer and drug-resistance-related genes by directly binding to their promoters and forming the cisplatin-HOXB13-ABCG1/EZH2/Slug network, including *ATP Binding Cassette Subfamily G Member 1*, *Enhancer Of Zeste 2 Polycomb Repressive Complex 2*, and *Slug* (16, 139). Levels of HOXB13 and its target genes may help predict sensitivity of platinum-based chemotherapy in lung adenocarcinoma patients (16).

## 2.7 HOX-Mediated Molecular Crosstalk During Tumorigenesis

### 2.7.1 Post-Translational Modifications

A variety of signaling pathways that regulate proliferation, apoptosis, differentiation, movement, and angiogenesis, interact with HOX transcription factors family. Post-translational modifications of HOX protein also play a key role in tumorigenesis. HOX post-translational modifications are an under-valued project. In most eukaryotic proteins, the turn-over, intracellular localization, molecular interactions and activity are modulated by post-translational modifications. The post-translational modifications of HOX proteins in cancer mainly

contributes to the modulation of protein stability, DNA binding, transcriptional activator-like effectors interaction, transcriptional activation capacity, unidentified cellular impact (157).

### 2.7.2 MicroRNA and Non-Coding RNA

Of the 39 HOX genes, 30 HOX genes contain more than one conserved nucleotide sequences that are expected to be targets of miRNAs in vertebrate (2). We found that the HOX targeting genes are mainly located on the 3' side of each HOX miRNA site. This study shows that HOX miRNAs help inhibiting abundant pre-gene expression, hence enhancing the prevalence of post-genes. In patients with gastrointestinal stromal tumors, the level of miRNA196a is positively related to higher tumor histologic grade, higher recurrence rate, and lower survival rate (64).

In addition, HOX transcribed antisense RNA (HOTAIR) also contributes to HOX expression regulation. HOTAIR is a 2.2-kilobase long non-coding RNA that is transcribed from the antisense strand of HOXC clusters, and its function is to inhibit the transcription of 40 kilobases of HOXD clusters (158, 159). The mechanism of action of these long non-coding RNAs has not been fully determined. However, HOTAIR has been shown to regulate chromatin state and kinetics by binding to specific chromatin modification complexes. The trimethylation of histone H3 lysine 27 at the HOXD site requires HOTAIR/PRC2 interactions. Knocking out HOTAIR activates HOXD gene transcriptional activity on human chromosome 2 (158, 160, 161). Thus, it is possible to modulate the HOXD gene by modulating HOTAIR levels, which has implications in cancer therapy. For example, the expression of HOTAIR is closely related to the neoplasm staging and poor prognosis of glioma. Reducing HOTAIR expression induces inhibition of colony formation, G0/G1 cell cycle arrest, and inhibition of tumor growth *in situ* (10). Thus, HOTAIR seems to serve as a prognostic factor for survival, as well as a biomarker for identifying molecular subtypes in cancer (162).

Recent evidence underscored the function of long noncoding RNAs (lncRNAs)-driven hepatocarcinogenesis. The expression level of lncRNA HOXA transcript at the distal tip (HOTTIP) and HOXA13 is associated with metastasis and survival in HCC patients. It indicates the prospective potency of HOTTIP and HOXA13 to be the predictive biomarker in HCC (163).

Summing up the above, we highlight the specific roles of miRNAs and lncRNAs in human cancer. They perform not only as the intermediary between DNA and protein, including chromosome remodeling, transcription, and post-transcriptional processing, but as leading characters in body balance adjustment during congenic malformation, oncogenesis, metabolic processes, and deregulation of cell cycle (164).

## 3 HOX TRANSCRIPTION FACTORS AS THERAPEUTIC TARGETS

### 3.1 HOX Genes Act as a Key Intermediate Point in the Anti-Cancer Progression

HOX proteins interact with specific cofactors to select their downstream binding sites in the genome. In vertebrates, those

including pre-B cell leukemia transcription factor (PBX) and mouse heterotopic integration, belong to the tricarboxylic acid cycle family of homologous domain protein. In addition, these HOX cofactors can increase the nuclear translocation of HOX protein from the cytoplasm to the nucleus. Nuclear translocation of HOX protein is inhibited by suppressing the formation of HOX/PBX dimer, which impairs the function of HOX transcript factors (165). Therefore, considering the use of HOX protein as cancer therapeutic targets, its interaction with cofactors needs to be determined. Therapeutic values of HOX factors in human cancer are seen in **Table 2** (4, 7, 13, 14, 17, 18, 22, 23, 25–27, 29, 31, 32, 35, 41, 45, 56, 57, 68, 71, 72, 81, 82, 84, 87, 88, 90, 97, 116, 117, 127, 128, 132–135, 137, 141, 147, 148, 153, 166, 167).

Morgan et al. have found HXR9 peptides specifically target the interaction between HOX and PBX (78, 168). HXR9 inhibits HOX function by preventing its binding to PBX, which leads to apoptosis in multiple mouse breast cancer derived cell lines (168). In addition, the interaction between HOX and HXR9 has been shown to cause apoptosis in numerous cancers, including melanoma, mesothelioma, myeloma, renal cancer, prostate cancer, lung cancer, ovarian cancer, pancreatic cancer, squamous cell cancer of the head and neck, and oral cancer (58, 78, 95, 98, 168). Kaspar and Reichert have already used HOX/HXR9-based treatments in cancer treatments, and Morgan and others are exploring a variant of HXR9 for intratumoral injection in clinical trials (168). The HOX gene targeting therapy is also being explored through RNA interference approaches (158). For example, MicroRNAs transcribed in HOX clusters, namely miR10A32 and miR196B33, regulate HOX expression through RNA interference (158). These miRNAs cleave or inhibit the translation of HOX mRNA. The use of these miRNAs in cancer cells *in vitro* seems to modulate HOX gene expression and its effect on cancer progression.

In fact, HOTAIR has the potential to be an effective therapeutic target for many types of cancer, where abnormal HOXD expression has been found to be associated with cancer progressions, such as cancers in breast, stomach, colon, cervix, lung, and liver (11, 12, 24, 36, 52–54, 158, 159, 169, 170). For example, HOTAIR is highly expressed in cervical cancer compared to normal cells (158, 159, 169). However, its knockout in cervical carcinoma cells induces apoptosis and inhibits tumor proliferation, migration, and invasion.

### 3.2 HOX Genes as Therapeutic Targets

Previous studies have shown that HOX protein exerts carcinogenic activity not only through its reversed transcription ability but through its protein interaction network.

When designing domain-specific HOX inhibitors, gene targeting must be taken into account. The problem in bringing monoclonal antibodies into clinical therapies is their nuclear localization. To address the above issue, the focus of next-generation immunotherapies is to develop smaller monoclonal antibody fragments or totally new entities to improve tissue permeability and subcellular localization. These crucial immunotherapeutic strategies are also focused on addressing the treatment of hematologic tumors and solid tumors.

Since HOX protein is closely linked to brain malignancies where the blood brain barrier is a major challenge for drug

**TABLE 2** | Therapeutic values of HOX factors in human cancer.

Gene	Down regulation effects	Downstream regulated molecules or pathways	Upstream regulatory molecules	Tumor x cell lines x normal tissues	Related targeting molecules	Intervention therapy	Clinical trial number & Reference
HOXA1	Proliferation, migration, invasion, metastasis	Wiping out epigenetic silencing of HOXA1	HOTAIRM, G9a, EZH2	GBM	Histone modification of H3K9, H3K27	/	(116)
HOXA1	Metastasis, invasion	MITF	TGF- $\beta$	Melanoma	TGF- $\beta$ signaling	/	(97)
HOXA1	Chemoresistance(up)	/	MiR-100	SCLC	HOXA1	/	(22)
HOXA3	Occurrence, development	/	HOXA-AS2, miR-15a-5p	PTCC	HOXA-AS2/miR-15a-5p/HOXA3 axis	/	(118)
HOXA3	Chemoresistance	EMT& Cisplatin resistance	HOXA-A3	NSCLC	HOXA-A3	/	(13)
HOXA4	Growth, migration, invasion	$\beta$ -catenin, cyclin D1, c-Myc, survivin	GSK3 $\beta$	Lung cancer	GSK3 $\beta$	LiCl	(14)
HOXA5	Angiogenesis	/	MiR-130-3p	HCC	SP1/MiR-130-3p/HOXA5	SP1 inhibitor	(22)
HOXA5	Growth, migration, invasion	CAF	Exosomal miR-181d-5p	Breast cancer	HOXA5/CDX2	/	(41)
HOXA5	Arresting cell cycle	TP53, P21	Binding to TAAT motif within promoter of TP53	Cervical cancer	Wnt/ $\beta$ -catenin	/	(84)
HOXA7	Migration, invasion	Snail	/	HCC	HOXA7/Snail	/	(25)
HOXA7	Proliferation, invasion	/	CircSLC26A4	Cervical cancer	QKI/circSLC26A4/miR-1287-5p/HOXA7 axis	/	(81)
HOXA9	Leukemogenesis	STAT5, AP1	JAK3/STAT5	Leukemia	Mutant JAK/STAT/HOXA9	PIM1 kinase	(88)
HOXA9	Leukemogenesis	CEBP $\alpha$ , MLL3, MLL4	HOXA9/Meis	Leukemia	Histone H3K4 methylation	/	(128)
HOXA9	Leukemogenesis	Meis, Syk	MiR-146a	Leukemia	HOXA9/Meis/Syk feedback loop	/	(87)
HOXA9	Leukemogenesis	Erg	Trib1	Leukemia	HOXA9/Trib1/Erg	JQ1	(90)
HOXA9	Regulating glucose metabolism reengineering	HIF1 $\alpha$ , HK2, GLUT1, PDK1	Onco-miR-365	CSCC	MiR-365-HOXA9-HIF1 $\alpha$ axis	/	(127)
HOXA9	Tumor aggression	HOXA9	BRCA1	Breast cancer	/	/	(132)
HOXA10	Tumor aggression	HOXA10, CCSCs	LINC00355/miR-195	HNSCC	LINC00355	/	(35)
HOXA11	Proliferation, cell-cell adhesion pathway	LncRNA HOXA11-AS, LSD1	EZH2	GC	EZH2/HOXA11-AS/LSD, HOXA11-AS/miR1297/EZH2	/	(56)
HOXA11	Metastasis, invasion	$\beta$ -catenin, P21, KLF2	WDR5/EZH2/STAU1	GC	HOXA11-AS	/	(57)
HOXA13	Metastasis	ACLY, IGF1R	IGF1	CRC	IGF1R/ACLY	Linsitinib/ETC-1002	(133, NCT01154335, NCT01016860)
HOXB3	Tumor aggression	RASSFIA, DNMT3B	POL2	Lung adenocarcinoma	RASSFIA	RASSFIA epigenetic silencing	(153)
HOXB3	Tumor aggression	CDCA3	/	Primary prostate cancer/PC-3/LNCaP	HOXB3	/	(72)
HOXB4	Arresting cell cycle	$\beta$ -catenin, TCF, c-Myc	/	Cervical cancer	Wnt/ $\beta$ -catenin	/	(82)
HOXB5	Metastasis	ITGB3, CXCR4	CXCL12	CRC	CXCR4	AMD3100	(134, NCT02179970)
HOXB5	Metastasis	CXCL1, FGFR4	FGF19	HCC	FGFR4	BLU-554	(26, NCT02508467, CT04194801)
HOXB7	Metastasis	TGF $\beta$ 2, SMADS	/	Breast cancer	MET	/	(135)
HOXB7	Metastasis	c-Myc, Slug	/	HCC	MET	/	(27)
HOXB8	Invasiveness	BACH1	/	CRC	/	/	(137)
HOXB9	Angiogenesis	IL-6, VEGF	/	Colorectal cancer	VEGF	Bevacizumab	(166)
HOXB9	Migration, tumor growth	JMJD6	Ack27	Lung adenocarcinoma	Ack27-HOXB9	/	(23)
HOXB9	Migration, tumor growth	EZH2	Ack27-HOXB9	Clone cancer	Ratio of Ack27-HOXB9/HOXB9	/	(167)
HOXB13	Tumor aggression	/	/	Prostate cancer	HOXB13, G84E variant	/	(71)
HOXB13	Metastasis	RFX6	Rs339331 at 6q22	Prostate cancer	RFX6	/	(68)

(Continued)



TABLE 2 | Continued

Gene	Down regulation effects	Downstream regulated molecules or pathways	Upstream regulatory molecules	Tumor x cell lines x normal tissues	Related targeting molecules	Intervention therapy	Clinical trial number & Reference
HOXB13	Tamoxifen resistance of breast cancer	IL-6	HBXIP	Breast cancer	HBXIP	Aspirin	(45)
HOXB13	Tumor aggression	$\beta$ -catenin, TCF4, c-Myc	DNMT3B	RCC	DNMT3B-HOXB13-c-Myc axis	/	(141)
HOXC8	Cisplatin chemoresistance, anti-apoptosis	TGF $\beta$ 1	/	NSCLC	HOXC8	/	(17)
HOXC9	Proliferation, migration	DAPK1-beclin1	/	Glioblastoma	HOXC9/autophagy	/	(4)
HOXC9	Chemoresistance	SRSF2, PLAU, HIC2	MIR-193a-3p	Bladder cancer	MIR-193a-3p/HOXC9/ DNA damage	/	(32)
HOXC10	Aberrant expression	HOXC10	PRC2	NSCLC	Kras-mutant/HOXC10	BET/MEK inhibitor	(18)
HOXC10	Angiogenesis	VEGFA	/	Gliomas	HOXC10/VEGFA	Bevacizumab	(7)
HOXC10	Invasiveness	VASP, PDPK1	IL1 $\beta$	HCC	IL1R1	Anakinra	(29)
HOXD3	Tumor aggression	Integrin $\beta$ 3	HOXD-AS1	CRC	HOXD-AS1-HOXD3-integrin $\beta$ 3	/	(147)
HOXD10	Invasion	Snail, Slug, MMP2, MMP9, MMP14, E-cadherin	MIR-23a	Gliblastoma	MIR-23a/HOXD10	/	(148)

infiltration, this provides instructive thinking of treatment. Another strategy for targeting oncogenes in cancer cells is to use small interfering RNAs (siRNAs) (171). Many studies have shown inhibition of HOX expression by siRNAs can distinctly retard tumor growth and aggressiveness. Similarly, the use of siRNA strategies may be an effective therapeutic approach for targeting HOX genes *in vivo*.

Although the strategy is not as progressive as small molecular chemical compound inhibitors or monoclonal antibodies, efforts have been made over the past decade to implement it in cancer treatment. At present, poor uptake of cells, side effects of packaging-related methods are major obstacles to HOX targeting clinical application. As a result, current efforts are made to address a range of novel strategies for delivering siRNAs *in vivo*.

The small-molecule chemical inhibitors are of anticancer potential. We highlight the recent therapies targeting HOX genes-downstream proteins especially in HCC and colorectal cancer. Overexpression of HOXB5 transactivates downstream protein expression of FGFR4 and CXCL1, hence promoting HCC metastasis. The small chemical compounds application of FGFR4 inhibitor BLU-554 and CXCR2 inhibitor SB265610 sharply inhibits HCC metastasis mediated by HOXB5 (26). Integrins are also involved in HOX-induced cancer metastasis. In CAOV-3 cells, HOXA4 overexpression inhibited migration and increased the protein level of  $\beta$ 1 integrin, suggesting that the  $\beta$ 1 integrin may be involved in HOXA4's inhibitory effect on cell motility (79). In HOXC10-VASP/IL-1R1 mediated HCC metastasis, daily administration of IL-1R1 antagonist anakinra dramatically prolongs survival time (29). In colorectal cancer, the molecule network IGF1-HOXA13-ACLY/IGF1R and CXCL12-HOXB5-CXCR4/ITGB3, targeted blocking the downstream protein with small molecular compounds serves as a promising anticancer therapy (133, 134).

Furthermore, natural and synthetic drugs regulating HOX genes network are associated with anticancer and antibacterial activities. For instance, researchers synthesized the new molecule 5H-pyrido [3,2-a] phenoxazine-5-one in the laboratory, evaluated its ability to regulate the activity of lncRNA HOTAIR and HOXC locus genes from HOXC9 to HOXC13 in MCF-7 human breast cancer cell lines, and confirmed that 5H-pyrido[3,2-a] phenoxazine-5-one was able to inhibit the relevant HOX gene expression and counteract the pathogenesis of breast cancer (172).

The discovery of targeting downstream molecules with chemical compounds has enhanced the potential to specifically target HOX genes intermediated network and eliminate the cancer development, which may improve clinical outcomes in cancers.

## 4 CONCLUSION

Although it is difficult to intercept various transcription factors, alternative strategies based on the exploitation of the associated molecular networks are emerging. The HOX genes play an important role in cancer progression, showing great plasticity and interfering with many molecular mechanisms. Abnormal HOX expression by altering its homologous box methylation profile, is often associated with cancer. In addition, different HOX gene regulatory features describe different cancer types and are increasingly being used as cancer biomarkers. Their role in cancer progression is based on the ability to control gene expression either directly as transcriptional regulators or indirectly through epigenetic control. In fact, HOX proteins have been shown to be involved in different epigenetic mechanisms involved in DNA and histone methylation, which may be related to the epigenetic regulation of multiple cancer related genes. Therefore, it is very important to seek pharmacological agents that are synthetically lethal in conjunction

with HOX overexpression. Therapies targeting HOX molecules should intake the functional redundancy among the different HOX family members, so that appropriate therapeutic combination can be created. There is an urgent need for a more comprehensive understanding of their biology, and identifying their gene targets and molecular networks involved in tumor progression.

## AUTHOR CONTRIBUTIONS

Conceptualization, YF, and LX. Writing—original draft preparation, YF. Writing—review and editing, YF, TZ, YW,

MX, XJ, WH, and LX. Supervision, TZ, MX, XJ, XL, and LX. Project administration, WH and LX. Funding acquisition, WH and LX. All authors contributed to the article and approved the submitted version.

## FUNDING

The research was supported by grants from the National Natural Science Foundation of China No. 81871911 (WH), No. 81972237 (LX), and No.81772623 (LX), and the National Key Research and Development Program of China 2018YFC1312103 (LX).

## REFERENCES

- Mallo M, Alonso CR. The Regulation of Hox Gene Expression During Animal Development. *Dev (Cambridge England)* (2013) 140:3951–63. doi: 10.1242/dev.068346
- Holland PW. Evolution of Homeobox Genes. *Wiley Interdiscip Rev Dev Biol* (2013) 2:31–45. doi: 10.1002/wdev.78
- Montavon T, Soshnikova N. Hox Gene Regulation and Timing in Embryogenesis. *Semin Cell Dev Biol* (2014) 34:76–84. doi: 10.1016/j.semcdb.2014.06.005
- Xuan F, Huang M, Liu W, Ding H, Yang L, Cui H. Homeobox C9 Suppresses Beclin1-Mediated Autophagy in Glioblastoma by Directly Inhibiting the Transcription of Death-Associated Protein Kinase 1. *Neuro-Oncology* (2016) 18:819–29. doi: 10.1093/neuonc/nov281
- Mao L, Ding J, Zha Y, Yang L, McCarthy BA, King W, et al. HOXC9 Links Cell-Cycle Exit and Neuronal Differentiation and Is a Prognostic Marker in Neuroblastoma. *Cancer Res* (2011) 71:4314–24. doi: 10.1158/0008-5472.Can-11-0051
- Li S, Zhang W, Wu C, Gao H, Yu J, Wang X, et al. HOXC10 Promotes Proliferation and Invasion and Induces Immunosuppressive Gene Expression in Glioma. *FEBS J* (2018) 285:2278–91. doi: 10.1111/febs.14476
- Tan Z, Chen K, Wu W, Zhou Y, Zhu J, Wu G, et al. Overexpression of HOXC10 Promotes Angiogenesis in Human Glioma via Interaction With PRMT5 and Upregulation of VEGFA Expression. *Theranostics* (2018) 8:5143–58. doi: 10.7150/thno.27310
- Han L, Liu D, Li Z, Tian N, Han Z, Wang G, et al. HOXB1 Is a Tumor Suppressor Gene Regulated by miR-3175 in Glioma. *PLoS One* (2015) 10:e0142387. doi: 10.1371/journal.pone.0142387
- Zhu Y, Cuevas IC, Gabriel RA, Su H, Nishimura S, Gao P, et al. Restoring Transcription Factor HoxA5 Expression Inhibits the Growth of Experimental Hemangiomas in the Brain. *J Neuropathol Exp Neurol* (2009) 68:626–32. doi: 10.1097/NEN.0b013e3181a491ce
- Zhang L, He A, Chen B, Bi J, Chen J, Guo D, et al. A HOTAIR Regulatory Element Modulates Glioma Cell Sensitivity to Temozolomide Through Long-Range Regulation of Multiple Target Genes. *Genome Res* (2020) 30:155–63. doi: 10.1101/gr.251058.119
- Angelopoulou E, Paudel YN, Piperi C. Critical Role of HOX Transcript Antisense Intergenic RNA (HOTAIR) in Gliomas. *J Mol Med (Berlin Germany)* (2020) 98:1525–46. doi: 10.1007/s00109-020-01984-x
- Li L, Wang Y, Song G, Zhang X, Gao S, Liu H. HOX Cluster-Embedded Antisense Long Non-Coding RNAs in Lung Cancer. *Cancer Lett* (2019) 450:14–21. doi: 10.1016/j.canlet.2019.02.036
- Lin S, Zhang R, An X, Li Z, Fang C, Pan B, et al. LncRNA HOXA-AS3 Confers Cisplatin Resistance by Interacting With HOXA3 in Non-Small-Cell Lung Carcinoma Cells. *Oncogenesis* (2019) 8:60. doi: 10.1038/s41389-019-0170-y
- Cheng S, Qian F, Huang Q, Wei L, Fu Y, Du Y. HOXA4, Down-Regulated in Lung Cancer, Inhibits the Growth, Motility and Invasion of Lung Cancer Cells. *Cell Death Dis* (2018) 9:465. doi: 10.1038/s41419-018-0497-x
- MX, XJ, WH, and LX. Supervision, TZ, MX, XJ, XL, and LX. Project administration, WH and LX. Funding acquisition, WH and LX. All authors contributed to the article and approved the submitted version.
- Monterisi S, Lo Riso P, Russo K, Bertalot G, Vecchi M, Testa G, et al. HOXB7 Overexpression in Lung Cancer Is a Hallmark of Acquired Stem-Like Phenotype. *Oncogene* (2018) 37:3575–88. doi: 10.1038/s41388-018-0229-9
- Zhan J, Wang P, Li S, Song J, He H, Wang Y, et al. HOXB13 Networking With ABCG1/EZH2/Slug Mediates Metastasis and Confers Resistance to Cisplatin in Lung Adenocarcinoma Patients. *Theranostics* (2019) 9:2084–99. doi: 10.7150/thno.29463
- Liu H, Zhang M, Xu S, Zhang J, Zou J, Yang C, et al. HOXC8 Promotes Proliferation and Migration Through Transcriptional Up-Regulation of Tgfb1 in Non-Small Cell Lung Cancer. *Oncogenesis* (2018) 7:1. doi: 10.1038/s41389-017-0016-4
- Guerra SL, Maertens O, Kuzmickas R, De Raedt T, Adeyemi RO, Guild CJ, et al. A Deregulated HOX Gene Axis Confers an Epigenetic Vulnerability in KRAS-Mutant Lung Cancers. *Cancer Cell* (2020) 37:705–719.e6. doi: 10.1016/j.ccell.2020.03.004
- Tang XL, Ding BX, Hua Y, Chen H, Wu T, Chen ZQ, et al. HOXC10 Promotes the Metastasis of Human Lung Adenocarcinoma and Indicates Poor Survival Outcome. *Front Physiol* (2017) 8:557. doi: 10.3389/fphys.2017.00557
- Yao Y, Luo J, Sun Q, Xu T, Sun S, Chen M, et al. HOXC13 Promotes Proliferation of Lung Adenocarcinoma via Modulation of CCND1 and CCNE1. *Am J Cancer Res* (2017) 7:1820–34.
- Durisetti S, Winnard PTJr, Mironchik Y, Vesuna F, Raman A, Raman V. HOXA5 Regulates Hmlh1 Expression in Breast Cancer Cells. *Neoplasia (New York N.Y.)* (2006) 8:250–8. doi: 10.1593/neo.05766
- Xiao F, Bai Y, Chen Z, Li Y, Luo L, Huang J, et al. Downregulation of HOXA1 Gene Affects Small Cell Lung Cancer Cell Survival and Chemoresistance Under the Regulation of miR-100. *Eur J Cancer (Oxford England: 1990)* (2014) 50:1541–54. doi: 10.1016/j.ejca.2014.01.024
- Wan J, Xu W, Zhan J, Ma J, Li X, Xie Y, et al. PCAF-Mediated Acetylation of Transcriptional Factor HOXB9 Suppresses Lung Adenocarcinoma Progression by Targeting Oncogenic Protein JMJD6. *Nucleic Acids Res* (2016) 44:10662–75. doi: 10.1093/nar/gkw808
- Loewen G, Jayawickramarajah J, Zhuo Y, Shan B. Functions of lncRNA HOTAIR in Lung Cancer. *J Hematol Oncol* (2014) 7:90. doi: 10.1186/s13045-014-0090-4
- Tang B, Qi G, Sun X, Tang F, Yuan S, Wang Z, et al. HOXA7 Plays a Critical Role in Metastasis of Liver Cancer Associated With Activation of Snail. *Mol Cancer* (2016) 15:57. doi: 10.1186/s12943-016-0540-4
- He Q, Huang W, Liu D, Zhang T, Wang Y, Ji X, et al. Homeobox B5 Promotes Metastasis and Poor Prognosis in Hepatocellular Carcinoma, via FGFR4 and CXCL1 Upregulation. *Theranostics* (2021) 11:5759–77. doi: 10.7150/thno.57659
- Huan HB, Yang DP, Wen XD, Chen XJ, Zhang L, Wu LL, et al. HOXB7 Accelerates the Malignant Progression of Hepatocellular Carcinoma by Promoting Stemness and Epithelial-Mesenchymal Transition. *J Exp Clin Cancer Res: CR* (2017) 36:86. doi: 10.1186/s13046-017-0559-4
- Li PD, Chen P, Peng X, Ma C, Zhang WJ, Dai XF. HOXC6 Predicts Invasion and Poor Survival in Hepatocellular Carcinoma by Driving Epithelial-Mesenchymal Transition. *Aging* (2018) 10:115–30. doi: 10.18632/aging.101363

29. Dang Y, Chen J, Feng W, Qiao C, Han W, Nie Y, et al. Interleukin 1 $\beta$ -Mediated HOXC10 Overexpression Promotes Hepatocellular Carcinoma Metastasis by Upregulating PDKP1 and VASP. *Theranostics* (2020) 10:3833–48. doi: 10.7150/thno.41712
30. Lv X, Li L, Lv L, Qu X, Jin S, Li K, et al. HOXD9 Promotes Epithelial-Mesenchymal Transition and Cancer Metastasis by ZEB1 Regulation in Hepatocellular Carcinoma. *J Exp Clin Cancer Res: CR* (2015) 34:133. doi: 10.1186/s13046-015-0245-3
31. Liao Y, Wang C, Yang Z, Liu W, Yuan Y, Li K, et al. Dysregulated Sp1/miR-130b-3p/HOXA5 Axis Contributes to Tumor Angiogenesis and Progression of Hepatocellular Carcinoma. *Theranostics* (2020) 10:5209–24. doi: 10.7150/thno.43640
32. Lv L, Li Y, Deng H, Zhang C, Pu Y, Qian L, et al. MiR-193a-3p Promotes the Multi-Chemoresistance of Bladder Cancer by Targeting the HOXC9 Gene. *Cancer Lett* (2015) 357:105–13. doi: 10.1016/j.canlet.2014.11.002
33. Meng L, Xing Z, Guo Z, Liu Z. LINC01106 Post-Transcriptionally Regulates ELK3 and HOXD8 to Promote Bladder Cancer Progression. *Cell Death Dis* (2020) 11:1063. doi: 10.1038/s41419-020-03236-9
34. Dai L, Hu W, Yang Z, Chen D, He B, Chen Y, et al. Upregulated Expression of HOXB7 in Intrahepatic Cholangiocarcinoma Is Associated With Tumor Cell Metastasis and Poor Prognosis. *Lab Investigation; J Tech Methods Pathol* (2019) 99:736–48. doi: 10.1038/s41374-018-0150-4
35. Lu S, Sun Z, Tang L, Chen L. LINC00355 Promotes Tumor Progression in HNSCC by Hindering MicroRNA-195-Mediated Suppression of HOXA10 Expression. *Mol Ther Nucleic Acids* (2020) 19:61–71. doi: 10.1016/j.omtn.2019.11.002
36. Sun S, Wu Y, Guo W, Yu F, Kong L, Ren Y, et al. STAT3/HOTAIR Signaling Axis Regulates HNSCC Growth in an EZH2-Dependent Manner. *Clin Cancer Res: an Off J Am Assoc Cancer Res* (2018) 24:2665–77. doi: 10.1158/1078-0432.Ccr-16-2248
37. de Bessa Garcia SA, Araújo M, Pereira T, Mouta J, Freitas R. HOX Genes Function in Breast Cancer Development. *Biochim Biophys Acta Rev Cancer* (2020) 1873:188358. doi: 10.1016/j.bbcan.2020.188358
38. Li J, Zeng T, Li W, Wu H, Sun C, Yang F, et al. Long Non-Coding RNA SNHG1 Activates HOXA1 Expression via Sponging miR-193a-5p in Breast Cancer Progression. *Aging* (2020) 12:10223–34. doi: 10.18632/aging.103123
39. Teo WW, Merino VF, Cho S, Korangath P, Liang X, Wu RC, et al. HOXA5 Determines Cell Fate Transition and Impedes Tumor Initiation and Progression in Breast Cancer Through Regulation of E-Cadherin and CD24. *Oncogene* (2016) 35:5539–51. doi: 10.1038/onc.2016.95
40. Hussain I, Deb P, Chini A, Obaid M, Bhan A, Ansari KI, et al. HOXA5 Expression Is Elevated in Breast Cancer and Is Transcriptionally Regulated by Estradiol. *Front Genet* (2020) 11:592436. doi: 10.3389/fgene.2020.592436
41. Wang H, Wei H, Wang J, Li L, Chen A, Li Z. MicroRNA-181d-5p-Containing Exosomes Derived From CAFs Promote EMT by Regulating CDX2/HOXA5 in Breast Cancer. *Mol Ther Nucleic Acids* (2020) 19:654–67. doi: 10.1016/j.omtn.2019.11.024
42. Lee JY, Hur H, Yun HJ, Kim Y, Yang S, Kim SI, et al. HOXB5 Promotes the Proliferation and Invasion of Breast Cancer Cells. *Int J Biol Sci* (2015) 11:701–11. doi: 10.7150/ijbs.11431
43. Shah N, Jin K, Cruz LA, Park S, Sadik H, Cho S, et al. HOXB13 Mediates Tamoxifen Resistance and Invasiveness in Human Breast Cancer by Suppressing Er $\alpha$  and Inducing IL-6 Expression. *Cancer Res* (2013) 73:5449–58. doi: 10.1158/0008-5472.Can-13-1178
44. Jansen MP, Sieuwerts AM, Look MP, Ritstier K, Meijer-van Gelder ME, van Staveren IL, et al. HOXB13-To-IL17BR Expression Ratio Is Related With Tumor Aggressiveness and Response to Tamoxifen of Recurrent Breast Cancer: A Retrospective Study. *J Clin Oncology: Off J Am Soc Clin Oncol* (2007) 25:662–8. doi: 10.1200/jco.2006.07.3676
45. Liu B, Wang T, Wang H, Zhang L, Xu F, Fang R, et al. Oncoprotein HBXIP Enhances HOXB13 Acetylation and Co-Activates HOXB13 to Confer Tamoxifen Resistance in Breast Cancer. *J Hematol Oncol* (2018) 11:26. doi: 10.1186/s13045-018-0577-5
46. Jerevall PL, Jansson A, Fornander T, Skoog L, Nordenskjöld B, Stål O. Predictive Relevance of HOXB13 Protein Expression for Tamoxifen Benefit in Breast Cancer. *Breast Cancer Res: BCR* (2010) 12:R53. doi: 10.1186/bcr2612
47. Shah M, Cardenas R, Wang B, Persson J, Mongan NP, Grabowska A, et al. HOXC8 Regulates Self-Renewal, Differentiation and Transformation of Breast Cancer Stem Cells. *Mol Cancer* (2017) 16:38. doi: 10.1186/s12943-017-0605-z
48. Zhang Y, Yang C, Zhang M, Liu H, Gong C, Zhang J, et al. Interleukin Enhancer-Binding Factor 3 and HOXC8 Co-Activate Cadherin 11 Transcription to Promote Breast Cancer Cells Proliferation and Migration. *Oncotarget* (2017) 8:107477–91. doi: 10.18632/oncotarget.22491
49. Pathiraja TN, Nayak SR, Xi Y, Jiang S, Garee JP, Edwards DP, et al. Epigenetic Reprogramming of HOXC10 in Endocrine-Resistant Breast Cancer. *Sci Trans Med* (2014) 6:229ra41. doi: 10.1126/scitranslmed.3008326
50. Chen H, Chung S, Sukumar S. HOXA5-Induced Apoptosis in Breast Cancer Cells Is Mediated by Caspases 2 and 8. *Mol Cell Biol* (2004) 24:924–35. doi: 10.1128/mcb.24.2.924-935.2004
51. Stasinopoulos IA, Mironchik Y, Raman A, Wildes F, Winnard P Jr., Raman V. HOXA5-Twist Interaction Alters P53 Homeostasis in Breast Cancer Cells. *J Biol Chem* (2005) 280:2294–9. doi: 10.1074/jbc.M411018200
52. Cantile M, Di Bonito M, Cerrone M, Collina F, De Laurentis M, Botti G. Long Non-Coding RNA HOTAIR in Breast Cancer Therapy. *Cancers* (2020) 12. doi: 10.3390/cancers12051197
53. Xue X, Yang YA, Zhang A, Fong KW, Kim J, Song B, et al. LncRNA HOTAIR Enhances ER Signaling and Confers Tamoxifen Resistance in Breast Cancer. *Oncogene* (2016) 35:2746–55. doi: 10.1038/onc.2015.340
54. Zhao W, Geng D, Li S, Chen Z, Sun M. LncRNA HOTAIR Influences Cell Growth, Migration, Invasion, and Apoptosis via the miR-20a-5p/HMGA2 Axis in Breast Cancer. *Cancer Med* (2018) 7:842–55. doi: 10.1002/cam4.1353
55. Lin J, Zhu H, Hong L, Tang W, Wang J, Hu H, et al. Coexpression of HOXA6 and PBX2 Promotes Metastasis in Gastric Cancer. *Aging* (2021) 13:6606–24. doi: 10.18632/aging.202426
56. Sun M, Nie F, Wang Y, Zhang Z, Hou J, He D, et al. LncRNA HOXA11-AS Promotes Proliferation and Invasion of Gastric Cancer by Scaffolding the Chromatin Modification Factors PRC2, LSD1, and DNMT1. *Cancer Res* (2016) 76:6299–310. doi: 10.1158/0008-5472.Can-16-0356
57. Liu Z, Chen Z, Fan R, Jiang B, Chen X, Chen Q, et al. Over-Expressed Long Noncoding RNA HOXA11-AS Promotes Cell Cycle Progression and Metastasis in Gastric Cancer. *Mol Cancer* (2017) 16:82. doi: 10.1186/s12943-017-0651-6
58. Shen LY, Zhou T, Du YB, Shi Q, Chen KN. Targeting HOX/PBX Dimer Formation as a Potential Therapeutic Option in Esophageal Squamous Cell Carcinoma. *Cancer Sci* (2019) 110:1735–45. doi: 10.1111/cas.13993
59. Ding WJ, Zhou M, Chen MM, Qu CY. HOXB8 Promotes Tumor Metastasis and the Epithelial-Mesenchymal Transition via ZEB2 Targets in Gastric Cancer. *J Cancer Res Clin Oncol* (2017) 143:385–97. doi: 10.1007/s00432-016-2283-4
60. Zhang E, Han L, Yin D, He X, Hong L, Si X, et al. H3K27 Acetylation Activated-Long Non-Coding RNA CCAT1 Affects Cell Proliferation and Migration by Regulating SPRY4 and HOXB13 Expression in Esophageal Squamous Cell Carcinoma. *Nucleic Acids Res* (2017) 45:3086–101. doi: 10.1093/nar/gkw1247
61. Li J, Tong G, Huang C, Luo Y, Wang S, Zhang Y, et al. HOXC10 Promotes Cell Migration, Invasion, and Tumor Growth in Gastric Carcinoma Cells Through Upregulating Proinflammatory Cytokines. *J Cell Physiol* (2020) 235:3579–91. doi: 10.1002/jcp.29246
62. Zhu H, Dai W, Li J, Xiang L, Wu X, Tang W, et al. HOXD9 Promotes the Growth, Invasion and Metastasis of Gastric Cancer Cells by Transcriptional Activation of RUFY3. *J Exp Clin Cancer Res: CR* (2019) 38:412. doi: 10.1186/s13046-019-1399-1
63. Yuan C, Zhu X, Han Y, Song C, Liu C, Lu S, et al. Elevated HOXA1 Expression Correlates With Accelerated Tumor Cell Proliferation and Poor Prognosis in Gastric Cancer Partly via Cyclin D1. *J Exp Clin Cancer Res: CR* (2016) 35:15. doi: 10.1186/s13046-016-0294-2
64. Niinuma T, Suzuki H, Nojima M, Noshio K, Yamamoto H, Takamaru H, et al. Upregulation of miR-196a and HOTAIR Drive Malignant Character in Gastrointestinal Stromal Tumors. *Cancer Res* (2012) 72:1126–36. doi: 10.1158/0008-5472.Can-11-1803
65. Chen Z, Wu D, Thomas-Ahner JM, Lu C, Zhao P, Zhang Q, et al. Diverse AR-V7 Cistromes in Castration-Resistant Prostate Cancer Are Governed by



- Hoxb13. *Proc Natl Acad Sci USA* (2018) 115:6810–5. doi: 10.1073/pnas.1718811115
66. Kim YR, Kim IJ, Kang TW, Choi C, Kim KK, Kim MS, et al. HOXB13 Downregulates Intracellular Zinc and Increases NF- $\kappa$ B Signaling to Promote Prostate Cancer Metastasis. *Oncogene* (2014) 33:4558–67. doi: 10.1038/ncr.2013.404
  67. VanOpstall C, Perike S, Brechka H, Gillard M, Lamperis S, Zhu B, et al. MEIS-Mediated Suppression of Human Prostate Cancer Growth and Metastasis Through HOXB13-Dependent Regulation of Proteoglycans. *eLife* (2020) 9. doi: 10.7554/eLife.53600
  68. Huang Q, Whittington T, Gao P, Lindberg JF, Yang Y, Sun J, et al. A Prostate Cancer Susceptibility Allele at 6q22 Increases RFX6 Expression by Modulating HOXB13 Chromatin Binding. *Nat Genet* (2014) 46:126–35. doi: 10.1038/ng.2862
  69. Zhou J, Yang X, Song P, Wang H, Wang X. HOXC6 in the Prognosis of Prostate Cancer. *Artif Cells Nanomedicine Biotechnol* (2019) 47:2715–20. doi: 10.1080/21691401.2019.1635136
  70. Xu F, Shanguan X, Pan J, Yue Z, Shen K, Ji Y, et al. HOXD13 Suppresses Prostate Cancer Metastasis and BMP4-Induced Epithelial-Mesenchymal Transition by Inhibiting SMAD1. *Int J Cancer* (2021) 148:3060–70. doi: 10.1002/ijc.33494
  71. Ewing CM, Ray AM, Lange EM, Zuhlke KA, Robbins CM, Tembe WD, et al. Germline Mutations in HOXB13 and Prostate-Cancer Risk. *N Engl J Med* (2012) 366:141–9. doi: 10.1056/NEJMoa1110000
  72. Chen J, Zhu S, Jiang N, Shang Z, Quan C, Niu Y. HoxB3 Promotes Prostate Cancer Cell Progression by Transactivating CDCA3. *Cancer Lett* (2013) 330:217–24. doi: 10.1016/j.canlet.2012.11.051
  73. Idaikkadar P, Morgan R, Michael A. HOX Genes in High Grade Ovarian Cancer. *Cancers* (2019) 11:1107. doi: 10.3390/cancers11081107
  74. Miller KR, Patel JN, Zhang Q, Norris EJ, Symanowski J, Michener C, et al. HOXA4/HOXB3 Gene Expression Signature as a Biomarker of Recurrence in Patients With High-Grade Serous Ovarian Cancer Following Primary Cytoreductive Surgery and First-Line Adjuvant Chemotherapy. *Gynecologic Oncol* (2018) 149:155–62. doi: 10.1016/j.ygyno.2018.01.022
  75. Ko SY, Naora H. HOXA9 Promotes Homotypic and Heterotypic Cell Interactions That Facilitate Ovarian Cancer Dissemination via Its Induction of P-Cadherin. *Mol Cancer* (2014) 13:170. doi: 10.1186/1476-4598-13-170
  76. Nakayama I, Shibasaki M, Yashima-Abo A, Miura F, Sugiyama T, Masuda T, et al. Loss of HOXD10 Expression Induced by Upregulation of miR-10b Accelerates the Migration and Invasion Activities of Ovarian Cancer Cells. *Int J Oncol* (2013) 43:63–71. doi: 10.3892/ijo.2013.1935
  77. Ko SY, Barengo N, Ladanyi A, Lee JS, Marini F, Lengyel E, et al. HOXA9 Promotes Ovarian Cancer Growth by Stimulating Cancer-Associated Fibroblasts. *J Clin Invest* (2012) 122:3603–17. doi: 10.1172/jci62229
  78. Morgan R, Plowright L, Harrington KJ, Michael A, Pandha HS, Targeting HOX. And PBX Transcription Factors in Ovarian Cancer. *BMC Cancer* (2010) 10:89. doi: 10.1186/1471-2407-10-89
  79. Klausen C, Leung PC, Auersperg N. Cell Motility and Spreading are Suppressed by HOXA4 in Ovarian Cancer Cells: Possible Involvement of Beta1 Integrin. *Mol Cancer research: MCR* (2009) 7:1425–37. doi: 10.1158/1541-7786.Mcr-08-0466
  80. Wang Z, Yu C, Wang H. HOXA5 Inhibits the Proliferation and Induces the Apoptosis of Cervical Cancer Cells via Regulation of Protein Kinase B and P27. *Oncol Rep* (2019) 41:1122–30. doi: 10.3892/or.2018.6874
  81. Ji F, Du R, Chen T, Zhang M, Zhu Y, Luo X, et al. Circular RNA Circslc26a4 Accelerates Cervical Cancer Progression via miR-1287-5p/HOXA7 Axis. *Mol Ther Nucleic Acids* (2020) 19:413–20. doi: 10.1016/j.omtn.2019.11.032
  82. Lei D, Yang WT, Zheng PS. HOXB4 Inhibits the Proliferation and Tumorigenesis of Cervical Cancer Cells by Downregulating the Activity of Wnt/ $\beta$ -Catenin Signaling Pathway. *Cell Death Dis* (2021) 12:105. doi: 10.1038/s41419-021-03411-6
  83. Wang Y, Wang C, Liu N, Hou J, Xiao W, Zhao L, et al. HOXC6 Promotes Cervical Cancer Progression via Regulation of Bcl-2. *FASEB Journal: Off Publ Fed Am Societies Exp Biol* (2019) 33:3901–11. doi: 10.1096/fj.201801099RR
  84. Ma HM, Cui N, Zheng PS. HOXA5 Inhibits the Proliferation and Neoplasia of Cervical Cancer Cells via Downregulating the Activity of the Wnt/ $\beta$ -Catenin Pathway and Transactivating TP53. *Cell Death Dis* (2020) 11:420. doi: 10.1038/s41419-020-2629-3
  85. Falini B, Brunetti L, Sportoletti P, Martelli MP. NPM1-Mutated Acute Myeloid Leukemia: From Bench to Bedside. *Blood* (2020) 136:1707–21. doi: 10.1182/blood.2019004226
  86. Zhang H, Zhang Y, Zhou X, Wright S, Hyle J, Zhao L, et al. Functional Interrogation of HOXA9 Regulome in MLLr Leukemia via Reporter-Based CRISPR/Cas9 Screen. *eLife* (2020) 9. doi: 10.7554/eLife.57858
  87. Mohr S, Doebele C, Comoglio F, Berg T, Beck J, Bohnenberger H, et al. Hoxa9 and Meis1 Cooperatively Induce Addiction to Syk Signaling by Suppressing miR-146a in Acute Myeloid Leukemia. *Cancer Cell* (2017) 31:549–562.e11. doi: 10.1016/j.ccell.2017.03.001
  88. de Bock CE, Demeyer S, Degryse S, Verbeke D, Sweron B, Gielen O, et al. HOXA9 Cooperates With Activated JAK/STAT Signaling to Drive Leukemia Development. *Cancer Discov* (2018) 8:616–31. doi: 10.1158/2159-8290.Cd-17-0583
  89. Whelan JT, Ludwig DL, Bertrand FE. HoxA9 Induces Insulin-Like Growth Factor-1 Receptor Expression in B-Lineage Acute Lymphoblastic Leukemia. *Leukemia* (2008) 22:1161–9. doi: 10.1038/leu.2008.57
  90. Yoshino S, Yokoyama T, Sunami Y, Takahara T, Nakamura A, Yamazaki Y, et al. Trib1 Promotes Acute Myeloid Leukemia Progression by Modulating the Transcriptional Programs of Hoxa9. *Blood* (2021) 137:75–88. doi: 10.1182/blood.2019004586
  91. Wang H, Jia XH, Chen JR, Yi YJ, Wang JY, Li YJ, et al. HOXB4 Knockdown Reverses Multidrug Resistance of Human Myelogenous Leukemia K562/ADM Cells by Downregulating P-Gp, MRP1 and BCRP Expression via PI3K/Akt Signaling Pathway. *Int J Oncol* (2016) 49:2529–37. doi: 10.3892/ijo.2016.3738
  92. Calvo KR, Knoepfler PS, Sykes DB, Pasillas MP, Kamps MP. Meis1a Suppresses Differentiation by G-CSF and Promotes Proliferation by SCF: Potential Mechanisms of Cooperativity With Hoxa9 in Myeloid Leukemia. *Proc Natl Acad Sci USA* (2001) 98:13120–5. doi: 10.1073/pnas.231115398
  93. Bi L, Zhou B, Li H, He L, Wang C, Wang Z, et al. A Novel miR-375-HOXB3-CDCA3/DNMT3B Regulatory Circuitry Contributes to Leukemogenesis in Acute Myeloid Leukemia. *BMC Cancer* (2018) 18:182. doi: 10.1186/s12885-018-4097-z
  94. Quéré R, Karlsson G, Hertwig F, Rissler M, Lindqvist B, Fioretos T, et al. Smad4 Binds Hoxa9 in the Cytoplasm and Protects Primitive Hematopoietic Cells Against Nuclear Activation by Hoxa9 and Leukemia Transformation. *Blood* (2011) 117:5918–30. doi: 10.1182/blood-2010-08-301879
  95. Daniels TR, Neacato II, Rodríguez JA, Pandha HS, Morgan R, Penichet ML. Disruption of HOX Activity Leads to Cell Death That Can be Enhanced by the Interference of Iron Uptake in Malignant B Cells. *Leukemia* (2010) 24:1555–65. doi: 10.1038/leu.2010.142
  96. Sarkar D, Leung EY, Baguley BC, Finlay GJ, Askarian-Amiri ME. Epigenetic Regulation in Human Melanoma: Past and Future. *Epigenetics* (2015) 10:103–21. doi: 10.1080/15592294.2014.1003746
  97. Wardwell-Ozgo J, Dogruluk T, Gifford A, Zhang Y, Heffernan TP, van Doorn R, et al. HOXA1 Drives Melanoma Tumor Growth and Metastasis and Elicits an Invasion Gene Expression Signature That Prognosticates Clinical Outcome. *Oncogene* (2014) 33:1017–26. doi: 10.1038/ncr.2013.30
  98. Morgan R, Simpson G, Gray S, Gillett C, Tabi Z, Spicer J, et al. HOX Transcription Factors Are Potential Targets and Markers in Malignant Mesothelioma. *BMC Cancer* (2016) 16:85. doi: 10.1186/s12885-016-2106-7
  99. Gaunt SJ. Hox Cluster Genes and Collinearities Throughout the Tree of Animal Life. *Int J Dev Biol* (2018) 62:673–83. doi: 10.1387/ijdb.180162sg
  100. Lewis EB. A Gene Complex Controlling Segmentation in *Drosophila*. *Nature* (1978) 276:565–70. doi: 10.1038/276565a0
  101. Quinonez SC, Innis JW. Human HOX Gene Disorders. *Mol Genet Metab* (2014) 111:4–15. doi: 10.1016/j.ymgme.2013.10.012
  102. Paço A, Aparecida de Bessa Garcia S, Leitão Castro J, Costa-Pinto AR, Freitas R. Roles of the HOX Proteins in Cancer Invasion and Metastasis. *Cancers* (2020) 13:10. doi: 10.3390/cancers13010010
  103. Li B, Huang Q, Wei GH. The Role of HOX Transcription Factors in Cancer Predisposition and Progression. *Cancers* (2019) 11:528. doi: 10.3390/cancers11040528
  104. Kuo TL, Cheng KH, Chen LT, Hung WC. Deciphering The Potential Role of Hox Genes in Pancreatic Cancer. *Cancers* (2019) 11:734. doi: 10.3390/cancers11050734



105. Procino A, Cillo C. The HOX Genes Network in Metabolic Diseases. *Cell Biol Int* (2013) 37:1145–8. doi: 10.1002/cbin.10145
106. Jambhekar A, Dhall A, Shi Y. Roles and Regulation of Histone Methylation in Animal Development. *Nat Rev Mol Cell Biol* (2019) 20:625–41. doi: 10.1038/s41580-019-0151-1
107. Cunningham TJ, Duester G. Mechanisms of Retinoic Acid Signalling and Its Roles in Organ and Limb Development. *Nat Rev Mol Cell Biol* (2015) 16:110–23. doi: 10.1038/nrm3932
108. Gaunt SJ. Hox Cluster Genes and Collinearities Throughout the Tree of Animal Life. *Cell Mol Life Sci: CMLS* (2018) 62:673–83. doi: 10.1387/ijdb.180162sg
109. Peters JM, Gonzalez FJ. The Evolution of Carcinogenesis. *Toxicological Sci: an Off J Soc Toxicol* (2018) 165:272–6. doi: 10.1093/toxsci/kfy184
110. Cairns RA, Harris IS, Mak TW. Regulation of Cancer Cell Metabolism. *Nat Rev Cancer* (2011) 11:85–95. doi: 10.1038/nrc2981
111. Magli MC, Largman C, Lawrence HJ. Effects of HOX Homeobox Genes in Blood Cell Differentiation. *J Cell Physiol* (1997) 173:168–77. doi: 10.1002/(sici)1097-4652(199711)173:2<168::Aid-jcp16>3.0.Co;2-c
112. Shah N, Sukumar S. The Hox Genes and Their Roles in Oncogenesis. *Nat Rev Cancer* (2010) 10:361–71. doi: 10.1038/nrc2826
113. Grier DG, Thompson A, Kwasniewska A, McGonigle GJ, Halliday HL, Lappin TR. The Pathophysiology of HOX Genes and Their Role in Cancer. *J Pathol* (2005) 205:154–71. doi: 10.1002/path.1710
114. Kamkar F, Yaymardan M, Asli NS. Hox-Mediated Spatial and Temporal Coding of Stem Cells in Homeostasis and Neoplasia. *Stem Cells Dev* (2016) 25:1282–9. doi: 10.1089/scd.2015.0352
115. Luo F, Yu S, Jin LH. The Posterior Signaling Center Is an Important Microenvironment for Homeostasis of the Drosophila Lymph Gland. *Front Cell Dev Biol* (2020) 8:382. doi: 10.3389/fcell.2020.00382
116. Li Q, Dong C, Cui J, Wang Y, Hong X. Over-Expressed lncRNA HOTAIRM1 Promotes Tumor Growth and Invasion Through Up-Regulating HOXA1 and Sequestering G9a/EZH2/Dnmts Away From the HOXA1 Gene in Glioblastoma Multiforme. *J Exp Clin Cancer Res: CR* (2018) 37:265. doi: 10.1186/s13046-018-0941-x
117. Jiang L, Wu Z, Meng X, Chu X, Huang H, Xu C. LncRNA HOXA-AS2 Facilitates Tumorigenesis and Progression of Papillary Thyroid Cancer by Modulating the miR-15a-5p/HOXA3 Axis. *Hum Gene Ther* (2019) 30:618–31. doi: 10.1089/hum.2018.109
118. Mahdipour E, Charnock JC, Mace KA. Hoxa3 Promotes the Differentiation of Hematopoietic Progenitor Cells Into Proangiogenic Gr-1+CD11b+ Myeloid Cells. *Blood* (2011) 117:815–26. doi: 10.1182/blood-2009-12-259549
119. Ordóñez-Morán P, Dafflon C, Imajo M, Nishida E, Huelsken J. HOXA5 Counteracts Stem Cell Traits by Inhibiting Wnt Signaling in Colorectal Cancer. *Cancer Cell* (2015) 28:815–29. doi: 10.1016/j.ccell.2015.11.001
120. Liang Y, Zhou R, Fu X, Wang C, Wang D. HOXA5 Counteracts the Function of Pathological Scar-Derived Fibroblasts by Partially Activating P53 Signaling. *Cell Death Dis* (2021) 12:40. doi: 10.1038/s41419-020-03323-x
121. Chen YQ, Yang TQ, Zhou B, Yang MX, Feng HJ, Wang YL. HOXA5 Overexpression Promotes Osteosarcoma Cell Apoptosis Through the P53 and P38 $\alpha$  MAPK Pathway. *Gene* (2019) 689:18–23. doi: 10.1016/j.gene.2018.11.081
122. Lee DH, Forscher C, Di Vizio D, Koeffler HP. Induction of P53-Independent Apoptosis by Ectopic Expression of HOXA5 in Human Liposarcomas. *Sci Rep* (2015) 5:12580. doi: 10.1038/srep12580
123. Strathdee G, Sim A, Soutar R, Holyoake TL, Brown R. HOXA5 is Targeted by Cell-Type-Specific CpG Island Methylation in Normal Cells and During the Development of Acute Myeloid Leukaemia. *Carcinogenesis* (2007) 28:299–309. doi: 10.1093/carcin/bgl133
124. Arderiu G, Cuevas I, Chen A, Carrio M, East L, Boudreau NJ. HoxA5 Stabilizes Adherens Junctions via Increased Akt1. *Cell Adhesion Migration* (2007) 1:185–95. doi: 10.4161/cam.1.4.5448
125. Cuevas I, Layman H, Coussens L, Boudreau N. Sustained Endothelial Expression of HoxA5 *In Vivo* Impairs Pathological Angiogenesis and Tumor Progression. *PLoS One* (2015) 10:e0121720. doi: 10.1371/journal.pone.0121720
126. Feng F, Ren Q, Wu S, Saeed M, Sun C. Hoxa5 Increases Mitochondrial Apoptosis by Inhibiting Akt/mTORC1/S6K1 Pathway in Mice White Adipocytes. *Oncotarget* (2017) 8:95332–45. doi: 10.18632/oncotarget.20521
127. Zhou L, Wang Y, Zhou M, Zhang Y, Wang P, Li X, et al. HOXA9 Inhibits HIF-1 $\alpha$ -Mediated Glycolysis Through Interacting With CRIP2 to Repress Cutaneous Squamous Cell Carcinoma Development. *Nat Commun* (2018) 9:1480. doi: 10.1038/s41467-018-03914-5
128. Sun Y, Zhou B, Mao F, Xu J, Miao H, Zou Z, et al. HOXA9 Reprograms the Enhancer Landscape to Promote Leukemogenesis. *Cancer Cell* (2018) 34:643–58.e5. doi: 10.1016/j.ccell.2018.08.018
129. Gonçalves CS, Xavier-Magalhães A, Martins EP, Pinto AA, Pires MM, Pinheiro C, et al. A Novel Molecular Link Between HOXA9 and WNT6 in Glioblastoma Identifies a Subgroup of Patients With Particular Poor Prognosis. *Mol Oncol* (2020) 14:1224–41. doi: 10.1002/1878-0261.12633
130. Yu SL, Koo H, Lee HY, Yeom YI, Lee DC, Kang J. Recombinant Cell-Permeable HOXA9 Protein Inhibits NSCLC Cell Migration and Invasion. *Cell Oncol (Dordrecht)* (2019) 42:275–85. doi: 10.1007/s13402-019-00424-4
131. Kroon E, Kros J, Thorsteinsdottir U, Baban S, Buchberg AM, Sauvageau G. Hoxa9 Transforms Primary Bone Marrow Cells Through Specific Collaboration With Meis1a But Not Pbx1b. *EMBO J* (1998) 17:3714–25. doi: 10.1093/emboj/17.13.3714
132. Gilbert PM, Mouw JK, Unger MA, Lakins JN, Gbengono MK, Clemmer VB, et al. HOXA9 Regulates BRCA1 Expression to Modulate Human Breast Tumor Phenotype. *J Clin Invest* (2010) 120:1535–50. doi: 10.1172/jci39534
133. Qiao C, Huang W, Chen J, Feng W, Zhang T, Wang Y, et al. IGF1-Mediated HOXA13 Overexpression Promotes Colorectal Cancer Metastasis Through Upregulating ACLY and IGF1R. *Cell Death Dis* (2021) 12:564. doi: 10.1038/s41419-021-03833-2
134. Feng W, Huang W, Chen J, Qiao C, Liu D, Ji X, et al. CXCL12-Mediated HOXB5 Overexpression Facilitates Colorectal Cancer Metastasis Through Transactivating CXCR4 and ITGB3. *Theranostics* (2021) 11:2612–33. doi: 10.7150/thno.52199
135. Liu S, Jin K, Hui Y, Fu J, Jie C, Feng S, et al. HOXB7 Promotes Malignant Progression by Activating the Tgf $\beta$  Signaling Pathway. *Cancer Res* (2015) 75:709–19. doi: 10.1158/0008-5472.Can-14-3100
136. Errico MC, Jin K, Sukumar S, Carè A. The Widening Sphere of Influence of HOXB7 in Solid Tumors. *Cancer Res* (2016) 76:2857–62. doi: 10.1158/0008-5472.Can-15-3444
137. Ying Y, Wang Y, Huang X, Sun Y, Zhang J, Li M, et al. Oncogenic HOXB8 is Driven by MYC-Regulated Super-Enhancer and Potentiates Colorectal Cancer Invasiveness via BACH1. *Oncogene* (2020) 39:1004–17. doi: 10.1038/s41388-019-1013-1
138. Nguyen NUN, Canseco DC, Xiao F, Nakada Y, Li S, Lam NT, et al. A Calcineurin-Hoxb13 Axis Regulates Growth Mode of Mammalian Cardiomyocytes. *Nature* (2020) 582:271–6. doi: 10.1038/s41586-020-2228-6
139. Barresi V, Ieni A, Cardia R, Licata L, Vitarelli E, Reggiani Bonetti L, et al. HOXB13 as an Immunohistochemical Marker of Prostatic Origin in Metastatic Tumors. *APMIS: Acta Pathologica Microbiologica Immunologica Scandinavica* (2016) 124:188–93. doi: 10.1111/apm.12483
140. Kim JJ, Kang TW, Jeong T, Kim YR, Jung C. HOXB13 Regulates the Prostate-Derived Ets Factor: Implications for Prostate Cancer Cell Invasion. *Int J Oncol* (2014) 45:869–76. doi: 10.3892/ijo.2014.2485
141. Xie B, Bai B, Xu Y, Liu Y, Lv Y, Gao X, et al. Tumor-Suppressive Function and Mechanism of HOXB13 in Right-Sided Colon Cancer. *Signal Transduction Targeted Ther* (2019) 4:51. doi: 10.1038/s41392-019-0086-1
142. Adwan H, Zhivkova-Galunska M, Georges R, Eyol E, Kleeff J, Giese NA, et al. Expression of HOXC8 Is Inversely Related to the Progression and Metastasis of Pancreatic Ductal Adenocarcinoma. *Br J Cancer* (2011) 105:288–95. doi: 10.1038/bjc.2011.217
143. Jiang Y, Yan B, Lai W, Shi Y, Xiao D, Jia J, et al. Repression of Hox Genes by LMP1 in Nasopharyngeal Carcinoma and Modulation of Glycolytic Pathway Genes by Hoxc8. *Oncogene* (2015) 34:6079–91. doi: 10.1038/nc.2015.53
144. Suo D, Wang Z, Li L, Chen Q, Zeng T, Liu R, et al. HOXC10 Upregulation Confers Resistance to Chemoradiotherapy in ESCC Tumor Cells and Predicts Poor Prognosis. *Oncogene* (2020) 39:5441–54. doi: 10.1038/s41388-020-1375-4
145. Guan Y, He Y, Lv S, Hou X, Li L, Song J. Overexpression of HOXC10 Promotes Glioblastoma Cell Progression to a Poor Prognosis via the PI3K/AKT Signalling Pathway. *J Drug Targeting* (2019) 27:60–6. doi: 10.1080/1061186x.2018.1473408

146. Wang L, Yao J, Yu T, Zhang D, Qiao X, Yao Z, et al. Homeobox D3, A Novel Link Between Bone Morphogenetic Protein 9 and Transforming Growth Factor Beta 1 Signaling. *J Mol Biol* (2020) 432:2030–41. doi: 10.1016/j.jmb.2020.01.043
147. Yang MH, Zhao L, Wang L, Ou-Yang W, Hu SS, Li WL, et al. Nuclear lncRNA HOXD-AS1 Suppresses Colorectal Carcinoma Growth and Metastasis via Inhibiting HOXD3-Induced Integrin  $\beta$ 3 Transcriptional Activating and MAPK/AKT Signalling. *Mol Cancer* (2019) 18:31. doi: 10.1186/s12943-019-0955-9
148. Yachi K, Tsuda M, Kohsaka S, Wang L, Oda Y, Tanikawa S, et al. miR-23a Promotes Invasion of Glioblastoma via HOXD10-Regulated Glial-Mesenchymal Transition. *Signal transduction targeted Ther* (2018) 3:333. doi: 10.1038/s41392-018-0033-6
149. Lin J, Teo S, Lam DH, Jayaseelan K, Wang S. MicroRNA-10b Pleiotropically Regulates Invasion, Angiogenicity and Apoptosis of Tumor Cells Resembling Mesenchymal Subtype of Glioblastoma Multiforme. *Cell Death Dis* (2012) 3:e398. doi: 10.1038/cddis.2012.134
150. Bi Y, Mao Y, Su Z, Du J, Ye L, Xu F. HOXB-AS1 Accelerates the Tumorigenesis of Glioblastoma via Modulation of HOBX2 and HOBX3 at Transcriptional and Posttranscriptional Levels. *J Cell Physiol* (2021) 236:93–106. doi: 10.1002/jcp.29499
151. Petrini M, Felicetti F, Bottero L, Errico MC, Morsilli O, Boe A, et al. HOXB1 Restored Expression Promotes Apoptosis and Differentiation in the HL60 Leukemic Cell Line. *Cancer Cell Int* (2013) 13:101. doi: 10.1186/1475-2867-13-101
152. Diehl F, Rössig L, Zeiher AM, Dimmeler S, Urbich C. The Histone Methyltransferase MLL Is an Upstream Regulator of Endothelial-Cell Sprout Formation. *Blood* (2007) 109:1472–8. doi: 10.1182/blood-2006-08-039651
153. Palakurthy RK, Wajapeyee N, Santra MK, Gazin C, Lin L, Gobeil S, et al. Epigenetic Silencing of the RASSF1A Tumor Suppressor Gene Through HOXB3-Mediated Induction of DNMT3B Expression. *Mol Cell* (2009) 36:219–30. doi: 10.1016/j.molcel.2009.10.009
154. Lorton BM, Shechter D. Cellular Consequences of Arginine Methylation. *Cell Mol Life Sci: CMLS* (2019) 76:2933–56. doi: 10.1007/s00018-019-03140-2
155. Boudadi E, Stower H, Halsall JA, Rutledge CE, Leeb M, Wutz A, et al. The Histone Deacetylase Inhibitor Sodium Valproate Causes Limited Transcriptional Change in Mouse Embryonic Stem Cells But Selectively Overrides Polycomb-Mediated Hoxb Silencing. *Epigenet Chromatin* (2013) 6:11. doi: 10.1186/1756-8935-6-11
156. Del Bene F, Wittbrodt J. Cell Cycle Control by Homeobox Genes in Development and Disease. *Semin Cell Dev Biol* (2005) 16:449–60. doi: 10.1016/j.semcdb.2005.02.001
157. Yu M, Zhan J, Zhang H. HOX Family Transcription Factors: Related Signaling Pathways and Post-Translational Modifications in Cancer. *Cell Signal* (2020) 66:109469. doi: 10.1016/j.cellsig.2019.109469
158. Gupta RA, Shah N, Wang KC, Kim J, Horlings HM, Wong DJ, et al. Long Non-Coding RNA HOTAIR Reprograms Chromatin State to Promote Cancer Metastasis. *Nature* (2010) 464:1071–6. doi: 10.1038/nature08975
159. Qu X, Alsager S, Zhuo Y, Shan B. HOX Transcript Antisense RNA (HOTAIR) in Cancer. *Cancer Lett* (2019) 454:90–7. doi: 10.1016/j.canlet.2019.04.016
160. Li Y, Ren Y, Wang Y, Tan Y, Wang Q, Cai J, et al. A Compound AC1Q3QWB Selectively Disrupts HOTAIR-Mediated Recruitment of PRC2 and Enhances Cancer Therapy of DZNep. *Theranostics* (2019) 9:4608–23. doi: 10.7150/thno.35188
161. Tsai MC, Manor O, Wan Y, Mosammamparast N, Wang JK, Lan F, et al. Long Noncoding RNA as Modular Scaffold of Histone Modification Complexes. *Sci (New York N.Y.)* (2010) 329:689–93. doi: 10.1126/science.1192002
162. Tan SK, Pastori C, Penas C, Komotar RJ, Ivan ME, Wahlestedt C, et al. Serum Long Noncoding RNA HOTAIR as a Novel Diagnostic and Prognostic Biomarker in Glioblastoma Multiforme. *Mol Cancer* (2018) 17:74. doi: 10.1186/s12943-018-0822-0
163. Quagliata L, Matter MS, Piscuoglio S, Arabi L, Ruiz C, Procino A, et al. Long Noncoding RNA HOTTIP/HOXA13 Expression Is Associated With Disease Progression and Predicts Outcome in Hepatocellular Carcinoma Patients. *Hepatol (Baltimore Md.)* (2014) 59:911–23. doi: 10.1002/hep.26740
164. Procino A. Class I Homeobox Genes, “The Rosetta Stone of the Cell Biology”, in the Regulation of Cardiovascular Development. *Curr medicinal Chem* (2016) 23:265–75. doi: 10.2174/0929867323666151207111302
165. Selleri L, Zappavigna V, Ferretti E. Building a Perfect Body: Control of Vertebrate Organogenesis by PBX-Dependent Regulatory Networks. *Genes Dev* (2019) 33:258–75. doi: 10.1101/gad.318774.118
166. Hoshino Y, Hayashida T, Hirata A, Takahashi H, Chiba N, Ohmura M, et al. Bevacizumab Terminates Homeobox B9-Induced Tumor Proliferation by Silencing Microenvironmental Communication. *Mol Cancer* (2014) 13:102. doi: 10.1186/1476-4598-13-102
167. Song J, Wang T, Xu W, Wang P, Wan J, Wang Y, et al. HOXB9 Acetylation at K27 Is Responsible for Its Suppression of Colon Cancer Progression. *Cancer Lett* (2018) 426:63–72. doi: 10.1016/j.canlet.2018.04.002
168. Morgan R, Boxall A, Harrington KJ, Simpson GR, Gillett C, Michael A, et al. Targeting the HOX/PBX Dimer in Breast Cancer. *Breast Cancer Res Treat* (2012) 136:389–98. doi: 10.1007/s10549-012-2259-2
169. Rajagopal T, Talluri S, Akshaya RL, Dunna NR. HOTAIR lncRNA: A Novel Oncogenic Propellant in Human Cancer. *Clinica Chimica Acta; Int J Clin Chem* (2020) 503:1–18. doi: 10.1016/j.cca.2019.12.028
170. Tatangelo F, Di Mauro A, Scognamiglio G, Aquino G, Lettierio A, Delrio P, et al. Posterior HOX Genes and HOTAIR Expression in the Proximal and Distal Colon Cancer Pathogenesis. *J Trans Med* (2018) 16:350. doi: 10.1186/s12967-018-1725-y
171. Singh A, Trivedi P, Jain NK. Advances in siRNA Delivery in Cancer Therapy. *Artif Cells Nanomedicine Biotechnol* (2018) 46:274–83. doi: 10.1080/21691401.2017.1307210
172. Manfra MFI, Bolognese A, Basilicata MG, Pepe G, Campiglia P, Procino A. Novel Anticancer Drug 5H-Pyro[3,2-a] Phenoxazin-5-One (PPH) Regulates lncRNA HOTAIR and HOXC Genes in Human MCF-7 Cells. *Bahrain Med Bull* (2021) 43:334–41.

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Publisher’s Note:** All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Feng, Zhang, Wang, Xie, Ji, Luo, Huang and Xia. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.