Circulating and Intratumoral Adrenal Androgens Correlate with Response to Abiraterone in Men with Castration-Resistant Prostate Cancer



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ABSTRACT

Purpose: In metastatic castration-resistant prostate cancer (mCRPC) low serum androgens prior to starting abiraterone acetate (AA) is associated with more rapid progression. We evaluated the effect of AA on androgens in castration-resistant prostate cancer (CRPC) metastases and associations of intratumoral androgens with response.

Experimental Design: We performed a phase II study of AA plus prednisone in mCRPC. The primary outcome was tissue testosterone at 4 weeks. Exploratory outcomes were association of steroid levels and genomic alterations with response, and escalating AA to 2,000 mg at progression.

Results: Twenty-nine of 30 men were evaluable. Testosterone in metastatic biopsies became undetectable at 4 weeks (P < 0.001). Serum and tissue dehydroepiandrosterone sulfate (DHEAS) remained detectable in many patients and was not increased at

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progression. Serum and tissue DHEAS in the lowest quartile (pretreatment), serum DHEAS in the lowest quartile (4 weeks), and undetectable tissue DHEAS (on-therapy) associated with rapid progression (20 vs. 48 weeks, P = 0.0018; 20 vs. 52 weeks, P = 0.0003; 14 vs. 40 weeks, P = 0.0001; 20 vs. 56 weeks, P = 0.02, respectively). One of 16 men escalating to 2,000 mg had a 30% PSA decline; 13 developed radiographic progression by 12 weeks. Among patients with high serum DHEAS at baseline, wild-type (WT) PTEN status associated with longer response (61 vs. 33 weeks, P = 0.02).

Conclusions: Low-circulating adrenal androgen levels are strongly associated with an androgen-poor tumor microenvironment and with poor response to AA. Patients with CRPC with higher serum DHEAS levels may benefit from dual androgen receptor (AR)-pathway inhibition, while those in the lowest quartile may require combinations with non–AR-directed therapy.

Introduction

While initially effective, treatment of prostate cancer with androgen deprivation therapy (ADT) is uniformly characterized by progression to castration-resistant prostate cancer (CRPC). Despite castrate-serum testosterone (T) levels, levels of T and DHT in CRPC metastases and patient-derived xenografts (PDX) are sufficient for androgen receptor (AR) activation (1, 2). Residual androgens in CRPC tumors may reflect *de novo* androgen synthesis or intratumoral uptake and conversion of adrenally derived steroid precursors to T and DHT (3, 4). In particular, serum levels of dehydroepiandrosterone sulfate (DHEAS), the primary circulating form of the adrenal androgen DHEA, are extremely high and are not suppressed by ADT (5, 6).

CYP17A, expressed in the adrenal gland, testes, and ovary, catalyzes sequential reactions converting pregnenolone and progesterone to the adrenal androgens DHEA and androstenedione (AED). The role of adrenal androgens in promoting CRPC is supported by the efficacy of the CYP17A inhibitor abiraterone acetate (AA) in decreasing circulating adrenal androgens and improving overall survival (OS) in CRPC (7-10). The proposed mechanism is suppression of androgen levels in tumor tissue as a result of suppressing testicular, adrenal, and tumoral CYP17A activity. This regimen markedly decreases prostate androgens below that achieved by standard ADT in the neoadjuvant setting (5) and studies using CRPC xenografts similarly demonstrate suppression of tissue androgens below castration alone (11). However, the efficacy of AA and prednisone in reducing intratumoral androgens in CRPC metastases has not been reported. Moreover, lower serum androgen levels prior to starting this regimen have been associated with worse outcomes in patients with CRPC (12, 13). Whether a similar association exists with intratumoral androgen levels prior to

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Translational Relevance

In men with metastatic castration-resistant prostate cancer (CRPC), low-circulating adrenal androgens levels are strongly associated with an androgen-poor tumor microenvironment, with lower androgen levels while on-therapy, and with poor response to abiraterone acetate (AA) plus prednisone. While response to AA is clearly linked with its ability to suppress circulating and intratumor androgen levels, the paradoxical association of lower adrenal androgen levels while on-therapy with more rapid radiographic progression suggests an intrinsic resistance to androgen receptor (AR)-pathway therapy related to tumor outgrowth in a low androgen microenvironment. These data suggest that clinical benefit from dual-AR pathway inhibition is likely to be observed in patients with higher serum adrenal androgen levels, but will be limited in those with low/undetectable levels in whom treatment combinations with non-AR-directed therapy are likely to be required. Assessment of baseline and on-treatment androgen levels can inform the stratification and interpretation of trials evaluating the efficacy of AR pathway-directed therapy.

starting therapy, or with on-treatment androgen levels in serum or tissue is unknown.

We carried out a phase II study to evaluate the efficacy of AA plus prednisone in reducing androgen levels in CRPC metastases. The primary outcome was tissue T levels at 4 weeks. Exploratory outcomes were the association of steroid levels and genomic alterations with response, and the impact of escalating AA to 2,000 mg at progression.

Materials and Methods

Study design and patient population

This was an investigator-initiated open-label single-center threearm study of AA (1,000 mg daily) and prednisone (5 mg twice daily) followed by escalation of AA to 2,000 mg daily at disease progression in men with metastatic castration-resistant prostate cancer (mCRPC). Eligible patients had metastatic prostate cancer resistant to ADT comprised of medical or surgical castration \pm standard antiandrogen (bicalutamide, flutamide, or nilutamide) and were not previously treated with AA or enzalutamide. Prior ketoconazole and docetaxel were allowed.

Study procedures and treatment

Patients underwent pretreatment biopsy of a bone, node, or softtissue metastasis and were then alternately enrolled in cohorts undergoing repeat biopsy at 4 weeks (cohort 1) or 12 weeks (cohort 2) until 10 patients were enrolled in each cohort (Study Schema provided in Supplementary Fig. S1). Patients with radiographic or clinical progression prior to planned biopsy were reassigned to cohort 3 (biopsy at progression) and another patient placed in cohort 1 or 2. After cohorts 1 and 2 were filled, patients were sequentially assigned to cohort 3 with biopsy at progression, until a total of 10 patients were enrolled in cohort 3 (of these, 2 had been reassigned from cohort 1 or 2 for clinical progression at 4 and 8 weeks). Patients remained on ADT and received AA 1,000 mg orally daily with prednisone 5 mg orally twice daily until development of radiographic or clinical progression. CT of the chest, abdomen, and pelvis, and bone scan were obtained before enrollment and every 12 weeks while on study treatment. Radiographic progression was determined based on Prostate Cancer Working Group 2 (PCWG2) criteria. Serum for PSA was drawn monthly. At radiographic progression asymptomatic patients were offered dose escalation of AA to 2,000 mg daily. Therapy with standard or dose-escalated AA was continued until radiographic or clinical progression or for a maximum of 2 years (104 weeks).

Steroid and abiraterone measurements

Methods for determination of steroids and abiraterone in serum and prostate tissue by mass spectrometry were as previously reported (14). Similar methods were used for detection of the abiraterone metabolites D4-abiraterone, and 3-keto-5 α -abiraterone. Additional information on assay methodology and limits of detection and quantitation is provided in Supplementary Fig. S2. For purposes of calculation, analyte values below the lower limit of detection were set at the lower limit of quantitation specific for that analyte.

Genomic analysis

Genomic DNA was prepared from clinical samples (buffy coat, mCRPC tumor tissue) using DNeasy Blood and Tissue Kit (QIAGEN). Next-generation sequencing (NGS) was performed on CRPC tissue biopsies using the clinically validated UW-OncoPlex platform (15). We determined *HSD3B1* genotype in DNA extracted from buffy coat using a melting assay with an unlabeled, locked, nucleic acid oligonucleotide probe in an asymmetric PCR as previously described (16).

Statistical analysis

A sample size of 6 patients per arm provided 94% power to detect the anticipated 0.660 pg/mg difference in tissue T levels relative to baseline, based on a 2-sided paired *t* test with alpha 5%. Ten patients per cohort were enrolled to account for potentially unproductive biopsies. Demographic and clinical characteristics were summarized using descriptive statistics. Comparisons of continuous variables among groups were assessed using the nonparametric Wilcoxon rank–sum test (Mann–Whitney test). Comparison of continuous variables between baseline and subsequent timepoints were assessed using the Wilcoxon matched-pairs test. Progression-free survival (PFS) was estimated using Kaplan–Meier methods and compared using the Gehan–Wilcoxan test. The statistical significance was set at P < 0.05. Due to the small sample size, all findings were considered hypothesis generating and no multiple-testing adjustments were performed. All statistical analyses were done using GraphPad Prism (version 8.3.0).

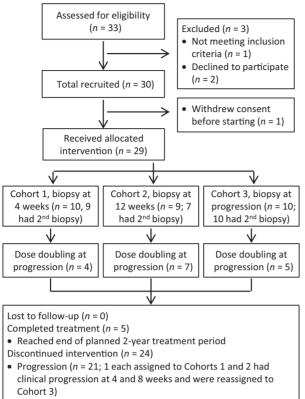
Study approval

All procedures were carried out in accordance with the U.S. Common Rule and approved by the institutional review board of the University of Washington. All subjects signed written informed consent. The trial was registered with the clinicaltrials.gov identifier NCT01503229.

Results

Patient characteristics

Thirty patients enrolled and 29 were evaluable for analysis (**Fig. 1**). Baseline characteristics are shown in **Table 1**. Median age was 71 years (range 44–83 years). The median PSA was 78 ng/mL (range 2–908). In the 18 patients with soft-tissue \pm bone disease, the biopsy site was equally divided between bone and lymph node based solely on tumor location most accessible to biopsy. Eleven patients had received previous definitive therapy for localized disease while 19 were metastatic at diagnosis. Eleven patients had received one or more treatments beyond bicalutamide for CRPC.



 Withdrew (n = 3; 1 to pursue alternative therapy, 1 due to religious reasons, 1 due to diagnosis of a different cancer)

Analyzed (n = 29)

- Efficacy evaluable population (n = 29)
- Safety evaluable population (n = 25)
 Safety evaluable population (n = 29)
- Excluded from analysis (n = 0)

Figure 1.

Flow diagram of patient recruitment, enrollment, and participation. Diagram depicts participant flow through the study process from patient screening to data analysis.

Clinical response to standard and dose-escalated AA and prednisone

The median time-to-radiographic/clinical progression in the entire cohort was 36 weeks (range 4-104; Fig. 2A). Twenty of 29 (69%) achieved a PSA decline of \geq 30% (PSA30) after 12 weeks of standarddose AA plus prednisone (Fig. 2B), with 22 (75%) eventually doing so. The median time-to-progression was longer in those achieving PSA30 decline at 12 weeks (40 vs. 34 weeks; P = 0.05; with a similar but nonsignificant trend in those achieving a PSA50 decline; Supplementary Fig. S3A and S3B; ref. 17). Notably, 4 of the 9 patients without PSA30 at 12 weeks had responses lasting 36, 40, 41, and 104 months, underscoring the importance of radiographic progression versus PSA decline as a primary endpoint. Three patients withdrew while responding to therapy (1 each to pursue alternative therapy, for religious reasons, and due to diagnosis of an unrelated malignancy) and were censored at the time of study withdrawal (at 28, 24, and 20 months). Three patients did not progress on standard-dose therapy by the end of the 2-year treatment period and were censored at 104 months.

Patients. <i>n</i>	29
Median age (range), years	71 (44-83)
Median PSA (range), ng/mL	78 (2-908)
Extent of disease	
Bone only	10 (33%)
Bone and LAD	15 (50%)
Bone and LAD/visceral disease	3 (10%)
LAD only	2 (7%)
Median tumor fraction in prestudy biopsy	55% (4-88%)
Previous therapies, <i>n</i>	
Radical prostatectomy	5
Definitive radiotherapy	6
Neither (metastatic at diagnosis)	19
Systemic therapy	
Combined androgen blockade	8
Bicalutamide	9
Nilutamide/flutamide	3
High-dose ketoconazole	5
Diethylstilbestrol	2
Sipuleucel-T	4
Docetaxel	4 (1 CSPC)
Lines of therapy for CRPC	
0	7
1	14
2	3
3–4	5

 Table 1. Baseline clinical characteristics at time of study enrollment.

Abbreviations: CSPC, castration-sensitive prostate cancer; LAD, lymphadenopathy.

At radiographic progression, 16 of 29 patients underwent dose escalation of AA to 2,000 mg per day (with continuation of prednisone). There was no decrease in the median PSA (Supplementary Fig. S3C), with only 1 patient achieving a PSA30 decline (**Fig. 2C**). Thirteen patients had clinical or radiographic progression and discontinued therapy by week 12 after dose escalation. Two patients remained on dose-escalated therapy for 56 weeks and 36 weeks, respectively, prior to second radiographic progression, while 1 patient remained on dose-escalated therapy for 20 weeks at which time he reached the end of the 2-year treatment period and was censored at 104 months. No consistent findings with regards to PSA, steroid, or abiraterone levels, or tumor genomic alterations were observed in these 3 patients.

Steroid and abiraterone levels during standard and doseescalated AA therapy

Steroid levels in serum and tissue prior to and during AA and prednisone therapy are summarized in Supplementary Tables S1 and S2. Serum levels showed a significant increase in steroids upstream of CYP17A (pregnenolone) and decreases in downstream steroids (DHEAS, DHEA, AED, T, and DHT) at all timepoints (**Fig. 2D**; see steroid metabolism schema, Supplementary Fig. S4), consistent with prior observations in serum and urine (7). While median levels of DHEAS and DHEA decreased by over two orders of magnitude in response to therapy in all patients, there appeared to be two modes of response. In one, levels became undetectable by week 4 and remained undetectable, while in the other subset, levels were still detectable at week 4 and remained detectable at all time points of treatment. AED, T, and DHT in serum were largely undetectable at all time points after starting therapy. Serum steroids were not increased at progression compared with earlier time points.

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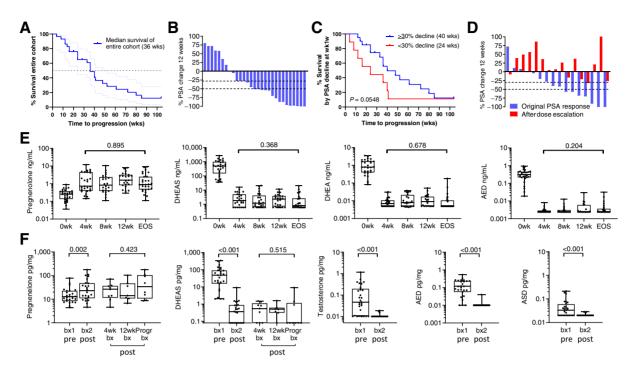


Figure 2.

Clinical response and steroid levels in serum and metastatic tissue on AA plus prednisone. **A**, Kaplan-Meier plot of time to radiographic progression on standard dose AA. **B**, Waterfall plot showing percent change in PSA at 12 weeks. **C**, Kaplan-Meier plot of time to radiographic progression comparing patients with or without a 30% PSA decline at 12 weeks. **D**, Waterfall plot comparing the original change in PSA (blue bars) to the percent change in PSA after dose escalation of AA to 2,000 mg per day (red bars). **E**, Change in serum steroid levels after standard dose AA plus prednisone at baseline (0 weeks); at 4, 8 and 12 weeks (4wk, 8wk, 12wk); and at end of study (EOS) at the time of radiographic progression. **F**, Change in steroid levels in metastatic tissue (tx) biopsies prior to therapy (biopsy 1, bx1) and while on therapy (biopsy 2, bx2). Biopsy 2 was taken at either 4 weeks (4wk bx), 12 weeks (12wk bx), or at progression (Progr bx). *P* values for the indicated comparison calculated via the Wilcoxon matched-pairs test. Data are shown as box and whisker plots, where horizontal lines indicate median values, white boxes denote the 75th (upper margin) and 25th percentiles (lower margin), and upper and lower bars indicate the minimum and maximum values, respectively. AED, androstenedione; DHEA, dehydroepiandrosterone; DHEAS, dehydroepiandrosterone sulfate.

In tissue as in serum, steroids upstream of CYP17A (pregnenolone) were increased in the on-treatment tissue biopsies, while downstream steroids [DHEAS, AED, T, androsterone (ASD)] were significantly decreased (**Fig. 2E**). Levels of AED, T, and ASD in tissue fell below the limit of detection by week 4 and remained essentially undetectable. DHEAS in tissue remained detectable in the majority of biopsies obtained at 4 and 12 weeks (6 of 9 and 6 of 7, respectively), but was undetectable in 6 of 9 samples at progression. DHT levels were below the limit of detection in 24 of 29 tissue samples at baseline and undetectable in all tissue after therapy. These observations suggest that an increase in tissue levels of androgens is not responsible for resistance to AA and prednisone and explains the lack of response to AA dose escalation.

AA is a prodrug that rapidly dissociates to abiraterone once ingested, therefore, all serum and tissue measurements are of abiraterone, not AA (summarized in Supplementary Table S3). Abiraterone is converted by 3β HSD1 to D4-abiraterone, and then by SRD5A to 3-keto- 5α -abiraterone, metabolites with AR antagonist and ARagonist activity, respectively (18). Levels of abiraterone metabolites in serum and tissue were similar at all time points and were not decreased at progression compared with earlier time points (Supplementary Fig. S5A and S5B). These observations suggest that resistance to AA is not associated with an increase in serum steroid levels or decrease in serum or tissue drug levels at the time of progression.

Dose escalation of AA from 1,000 mg to 2,000 mg resulted in the expected pharmacokinetic changes with a nonsignificant increase in

the median level of abiraterone (22.4 to 78.2 ng/mL), and a significant increase in D4-abiraterone (1.48 to 4.0; P = 0.012) and 3-keto-5 α -abiraterone (10.6 to 14.3 ng/mL; P = 0.005) at 4 weeks (Supplementary Fig. S5C). Accordingly, serum levels of DHEAS and DHEA decreased in 9 of 16 patients, and AED levels remained or became undetectable in 15 of 16 (Supplementary Fig. S5D), although the overall decrease in median levels was nonsignificant for DHEAS and DHEA, and of borderline significance for AED (P = 0.21, 0.36, and 0.06, respectively). These observations suggest that the lack of clinical response to AA dose escalation is not accounted for by a failure of pharmacokinetic or pharmacodynamic responses in serum. Moreover, in the 3 patients that experienced prolonged responses, serum abiraterone and androgen levels after dose escalation were variably altered (decreased in 2 and increased in 1), although changes in tissue levels before and after dose escalation were not assayed.

Correlation of steroid levels in serum and metastatic tissue before and after therapy with AA and prednisone

Serum levels of DHEAS, DHEA, AED, ASD, T, and DHT, were highly correlated with each other and with levels of DHEAS, AED, and ASD in pretreatment biopsies (**Fig. 3A**). Tissue levels of DHEAS and AED, DHEAS and ASD, and AED and ASD correlated with each other (r = 0.51, 0.56, and 0.46, respectively; P < 0.05 for all), while tissue T levels were not significantly correlated with steroid levels in either serum or tissue. After starting AA plus prednisone, levels of DHEAS,

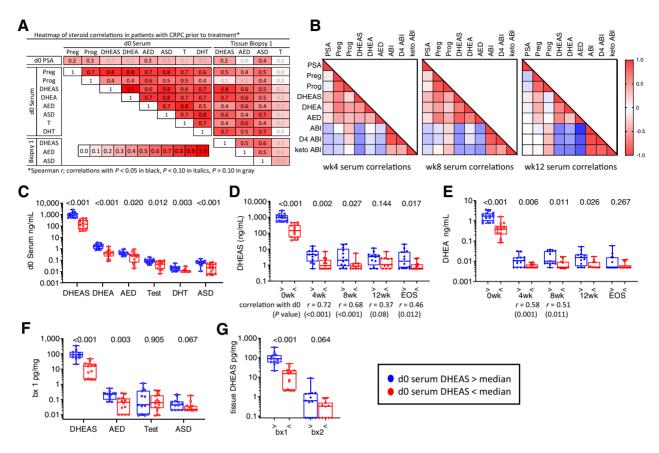


Figure 3.

Correlation of steroid levels in serum and metastatic tissue before and after treatment with AA plus prednisone. **A**, Heatmap of steroid correlations in pretreatment serum (day 0) and metastatic tissue biopsies (Biopsy 1). The Spearman *r* value for each correlation is shown in the box. Correlations with P < 0.05 in black, P < 0.10 in italics, P > 0.10 in gray. **B**, Heatmap of Spearman correlations for steroids in serum at week 4 (wk4), week 8 (wk8), and week 12 (wk12). **C**, Comparison of baseline serum steroid levels stratified by serum DHEAS levels above (blue) versus below (red) the median at baseline (d0). **D** and **E**, Comparison of DHEAS levels (**D**) and DHEA levels (**E**) in serum stratified by baseline serum DHEAS levels at baseline (0 weeks); at 4, 8, and 12 weeks; and at EOS. Significant Spearmen correlations between on-treatment and baseline values at each time point is indicated below the timepoint. **F**, Comparison of steroid levels in pretreatment tissue biopsies (bx1) stratified by serum DHEAS levels above (blue) vs. below (red) the median at baseline. **G**, Comparison of tissue (tx) DHEAS levels in pretreatment tissue biopsies (bx1 and bx2). *P* values for the indicated comparison calculated via nonparametric Mann-Whitney *t* tests. ABI, abiraterone; D4 ABI, D4-abiraterone; Keto ABI, 3-keto-5\alpha-abiraterone; Preg, pregnenolone; Preg, progesterone; Test, testosterone.

DHEA, and AED remained significantly correlated with each other and showed the expected inverse correlations with abiraterone and its metabolites (**Fig. 3B**), while progesterone showed the expected positive correlation with abiraterone and its metabolites. These treatmentrelated correlations were present by 4 weeks and became notably more pronounced at 8 and 12 weeks. Abiraterone and metabolite levels were strongly correlated with each other in serum and tissue at all time points (Supplementary Fig. S6A).

These baseline and on-treatment steroid correlations are further illustrated by the observation that subjects with baseline serum DHEAS levels above versus below the median also had higher baseline levels of DHEA, AED, T, DHT, and ASD (**Fig. 3C**) and maintained significantly higher serum levels of DHEAS (**Fig. 3D**) and DHEA (**Fig. 3E**) at all time points [including, for DHEAS, at the end-of-study (EOS) measurement taken at progression]. Moreover, strong correlations between baseline and on-treatment serum levels were observed at all time points for DHEAS (**Fig. 3D**), and at 4 and 8 weeks for DHEA (**Fig. 3E**). Likewise, subjects with baseline serum DHEAS above versus below the median had higher tissue levels of DHEAS, AED, and ASD at baseline (Fig. 3F), as well as higher DHEAS in on-treatment tissue biopsies (Fig. 3G). These observations demonstrate that serum DHEAS levels prior to AA therapy can identify patients with CRPC with concomitantly higher levels of all androgens in serum and tissue, and that this subset of patients with high baseline androgen levels maintains persistently higher serum and tissue DHEAS levels after treatment with AA and prednisone.

Association of steroid and abiraterone levels with PSA decline and PFS

Compared with patients who achieved a PSA30 decline at 12 weeks, those with less than a PSA30 decline had numerically lower pretreatment levels of serum and tissue androgens (**Fig. 4A**). These differences were strongest for T (P = 0.055) and DHT (P = 0.069) in serum, and for AED (P = 0.063), T (P = 0.034), and ASD (P = 0.088) in tissue. Accordingly, the rate of achieving a PSA30 decline at 12 weeks was 37% (3/8) in patients in the lowest quartile of serum DHEAS, but 81% (17/21) in patients in the top 3 quartiles (**Fig. 4C**; PSA levels at baseline were not associated

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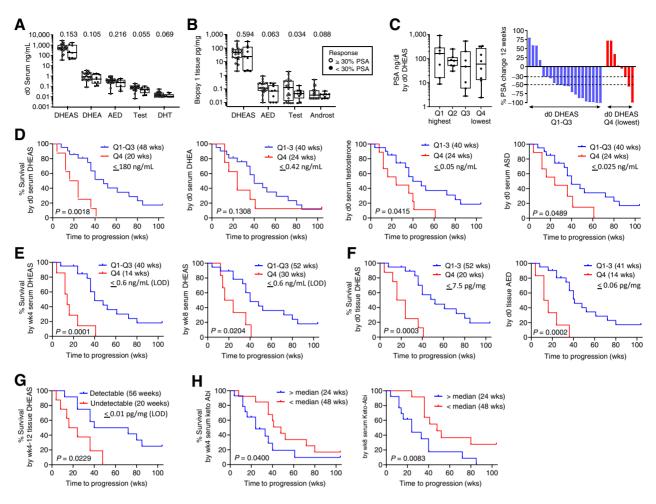


Figure 4.

Association of steroid and abiraterone levels with PSA decline and radiographic progression on AA plus prednisone. **A** and **B**, Comparison of steroid levels in serum (**A**) and metastatic tissue biopsies (**B**), based on achieving a 30% PSA decline at 12 weeks. **C**, Distribution of pretreatment PSA levels and waterfall plot showing percent change in PSA at 12 weeks by quartile (Q1-Q4) of pretreatment serum DHEAS levels. *P* values for the indicated comparison calculated via nonparametric Wilcoxon rank-sum test (Mann-Whitney test). **D**, Radiographic PFS (rPFS) as a function of baseline serum androgen levels comparing subjects in the lowest vs. highest 3 quartiles (Q4 vs. Q1-3). **E**, rPFS as a function of on-treatment serum DHEAS levels at week 4 (wk4) and week 8 (wk8) comparing subjects in the lowest vs. highest 3 quartiles (Q4 vs. Q1-3). **F**, rPFS as a function of pretreatment tissue DHEAS and AED levels comparing subjects in the lowest vs. highest 3 quartiles (Q4 vs. Q1-3). **F**, rPFS as a function of pretreatment tissue DHEAS and AED levels. The quartiles were separately assessed in the pre- and on treatment populations. **G**, rPFS comparing subjects with detectable vs. undetectable levels of DHEAS in tissue biopsies taken at 4 and 12 weeks of therapy. **H**, rPFS as a function of serum 3-keto-5α-abiraterone levels (keto-Abi) at wk4 and wk8 comparing subjects above vs. below the median. PFS was estimated using Kaplan-Meier methods and compared using the Gehan-Wilcoxan test. Androst, androsterone; Q, quartile.

with serum DHEAS levels). These observations suggest that being in the lowest quartile of serum androgens prior to starting AA therapy is associated with a decreased incidence of PSA30 decline at 12 weeks.

Consistent with these observations, serum steroid levels in the lowest quartile prior to therapy were associated with more rapid progression. This was highly significant for DHEAS (20 vs. 48 weeks, P = 0.0018) with less significant trends noted for DHEA, T, and ASD (P = 0.13, 0.041, and 0.049 respectively; **Fig. 4D**). Notably, ontreatment levels of DHEAS in the lowest quartile remained associated with more rapid progression (14 vs. 40 weeks, P = 0.0001 at 4 weeks; 20 vs. 40 weeks, P = 0.02 at 8 weeks; **Fig. 4E**). Low adrenal androgens in pretreatment tissue biopsies were even more strongly associated with worse outcomes (20 vs. 52 weeks, P = 0.0003 for DHEAS; 14 vs. 41 weeks, P = 0.0002 for AED; **Fig. 4F**). Likewise, undetectable vs.

detectable DHEAS levels in biopsies taken at 4 and 12 weeks were associated with shorter time-to-progression (20 vs. 56 weeks, P = 0.023; **Fig. 4G**). These observations demonstrate that the association of lower serum adrenal androgen levels with more rapid progression on AA plus prednisone holds true for lower adrenal-androgen levels in pretreatment metastasis biopsies, and for lower on-treatment levels of DHEAS in serum and tissue.

Levels of abiraterone and D4-abiraterone in serum were not associated with differences in time-to-progression, nor were levels of abiraterone and its metabolites in tissue (Supplementary Fig. S6B and S6C). However, serum levels of 3-keto-5 α -abiraterone above the median at week 4 and 8 were associated with more rapid progression (24 vs. 48 weeks, P = 0.04, and 24 vs. 48 weeks, P = 0.008, respectively; **Fig. 4H**), consistent with the ability of this metabolite to act as an AR agonist.

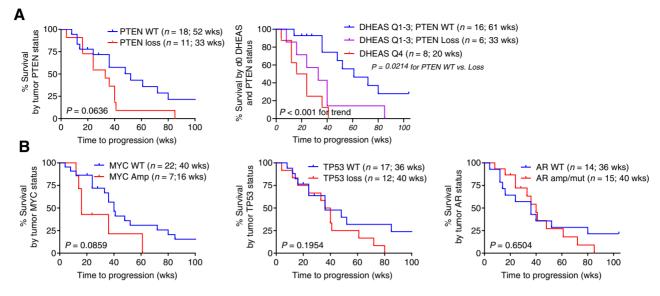


Figure 5.

Association of *PTEN* status with radiographic progression on AA plus prednisone. **A**, rPFS as a function of *PTEN* status alone or as a function of baseline serum DHEAS levels stratified by *PTEN* status, comparing subjects in the lowest vs highest three quartiles (Q4 vs Q1–3). **B**, rPFS as a function of *MYC*, *TP53*, or *AR* status.

Association of genomic alterations with radiographic PFS on AA and prednisone

All patients had pretreatment tumor biopsy tissue that was adequate for sequencing. The frequency and distribution of pathogenic alterations were consistent with mCRPC (Supplementary Table S4). Wild-type (WT) PTEN was weakly associated with longer time-to-progression (52 vs. 33 weeks, P = 0.06; Fig. 5A). Patients in the lowest quartile of serum DHEAS (3 PTEN WT, and 5 PTEN loss) were previously shown to have a poor response to AA (Fig. 4D) and this was not influenced by PTEN status. However, in the top 3 quartiles of serum DHEAS (with median survival 48 weeks; Fig. 4D), WT PTEN versus PTEN loss distinguished patients with a significantly more prolonged versus intermediate response (61 vs. 33 weeks; P = 0.02) versus those in the lowest quartile of DHEAS (20 weeks; P < 0.001 for trend; Fig. 5A). These findings demonstrate that patients in the lowest quartile of serum DHEAS levels have the most rapid progression on AA, regardless of PTEN status, while those with DHEAS levels in the top 3 quartiles can be meaningfully stratified by PTEN status. In this small study WT MYC was also weakly correlated with longer survival (40 vs. 16 weeks, P = 0.08) while TP53 status or the presence of AR mutation or amplification were not associated with progression (Fig. 5A).

As inhibition of AR signaling has been associated with reciprocal induction of PTEN signaling (19, 20), we determined if lower androgens were associated with PTEN loss. Median serum levels of T and DHT at baseline were numerically but not statistically lower in patients with *PTEN* loss versus WT *PTEN* (P = 0.09 and 0.084, respectively; Supplementary Fig. S7B), as were on-treatment levels of DHEAS in tissue (P = 0.09). Median serum levels of DHEAS and DHEA at baseline were also numerically lower in patients with *TP53* loss versus WT *TP53* (P = 0.053 and 0.047, respectively; Supplementary Fig. S7C). These observations suggest that while specific AR-independent tumor drivers such as *PTEN* and *TP53* may be selected to emerge in a low-androgen environment, a low-androgen environment is likely to promotes a more aggressive tumor biology by multiple mechanisms not limited to specific pathway alterations.

Association of *HSD3B1* genotype with steroid and abiraterone levels and radiographic PFS

The 1245C variant of HSD3B1 produces a more stable enzyme, increasing conversion of DHEA to AED, a precursor of T and DHT (Supplementary Fig. S4), as well as increasing conversion of abiraterone to D4-abiraterone (21). Germline DNA testing identified 14 men who were homozygous WT (48%), 10 men who were heterozygous (34%), and 6 who were homozygous for the allelic variant (21%). Consistent with the impact of the variant on enhancing conversion of abiraterone to D4-abiraterone, serum levels of this metabolite were highest in subjects with two variant alleles (median 2.7 ng/mL) versus one variant allele (2.1 ng/mL) versus two WT alleles (1.2 ng/ mL; ANOVA P = 0.02; Supplementary Fig. S8A). Serum levels of 3-keto-5\alpha-abiraterone did not differ by HSDB1 genotype, nor did levels of abiraterone metabolite in tissue (Supplementary Fig. S8A). There was no difference in serum or tissue-androgen levels based on HSD3B1 status (including the five tissue samples with detectable levels of DHT; Supplementary Fig. S8B), nor in time-to-progression (Supplementary Fig. S8C).

Discussion

In this phase II study we demonstrate that in mCRPC AA plus prednisone decreases T levels in metastatic tissue biopsies to undetectable by 4 and 12 weeks of therapy, consistent with the predicted mechanism of activity. Steroid and abiraterone levels in serum and tissue remained stable at progression compared with earlier time points, consistent with prior reports regarding serum adrenal androgens (8), but demonstrating for the first time that tissue-androgen levels also remain unchanged at progression. Overall, these findings suggest that the development of resistance to AA plus prednisone is not accounted for by a decrease in serum or tissue drug levels below the levels originally achieved in response to therapy. Nor is it accounted for by an increase in serum or intratumoral androgens above the levels originally achieved in response to AA plus prednisone. The observations that tumor steroid levels do not become reelevated at progression and that persistent serum DHEAS levels do not become undetectable after dose escalation of AA to 2,000 mg likely accounts for the lack of clinical response to high-dose AA at progression, now reported in two studies (22).

At baseline, serum levels of DHEAS and DHEA varied by over an order of magnitude (range 36–2,659 ng/mL) and demonstrated two patterns of response to treatment. While median levels of steroids downstream of CYP17A decreased by a similar order of magnitude in all patients, the subset of patients defined by higher serum DHEAS levels prior to therapy also had higher serum levels of other steroids at baseline; higher levels of DHEAS, AED, and ASD in pretreatment tissue biopsies; and maintained higher DHEAS levels in on-treatment serum samples and tissue biopsies. These observations demonstrate that serum DHEAS levels prior to therapy can distinguish patients with correspondingly lower or higher serum and tissue levels of all androgens and can identify those likely to maintain persistent serum and tissue DHEAS levels while on treatment with AA plus prednisone.

The correlation of low baseline with low on-treatment adrenalandrogen levels is of particular interest as lower adrenal androgens prior to therapy have been consistently associated with worse response to AA plus prednisone (13, 23–25). We show for the first time that this association with worse outcomes extends to low on-treatment level of DHEAS in serum and tissue, and with low adrenal androgens in pretreatment metastasis biopsies. At baseline, the association of worse response with lower levels was highly significant for DHEAS in serum (P = 0.0018) and for DHEAS and AED in pretreatment tissue biopsies (P = 0.0003 and P = 0.0002, respectively). On therapy, serum DHEAS in the lowest quartile at 4 and 8 weeks (P = 0.0001 and 0.024, respectively), as well as undetectable versus detectable DHEAS levels in metastatic biopsies taken at 4 and 12 weeks (P = 0.023 and P = 0.04, respectively) all associated with more rapid progression on AA therapy. These specific associations with DHEAS are consistent with the fact that DHEAS is the predominant steroid in serum and tissue prior to therapy, and remains the only steroid consistently present at measurable levels in metastatic tissue after AA therapy.

While response to AA is clearly linked with suppression of circulating and intratumor androgens, the paradoxical association of better outcomes with higher on-treatment androgen levels suggests that response to AA is not dictated solely by a pharmacodynamic effect on serum or tissue androgens, but may reflect an intrinsic resistance to AR-pathway therapy associated with tumor outgrowth in a low androgen microenvironment (26). These observations suggest that baseline serum androgen levels, by identifying patients with a more aggressive tumor biology that is less androgen dependent, may be prognostic as well as predictive of response to AR-pathway-directed therapy. As such, patients with higher androgens may have better survival independent of an effect of AA. This is consistent with the association of lower serum androgen levels with decreased survival in both the placebo and AA arms of the phase III study of AA plus prednisone in men with mCRPC (13), and with a recent meta-analysis demonstrating that PFS and OS were lower in patients with CRPC with lower versus higher T levels (27).

Importantly, the observation that lower androgen levels in CRPC associate with worse outcomes is not incompatible with data linking the achievement of lower T levels with improved time-to-progression in men with CSPC treated with ADT (28, 29). While time-to-development of CRPC may be delayed, CRPC tumors that emerge

in patients with lower T levels (due to low adrenal contribution and/or optimally suppressed T levels while on ADT) are likely to represent a more aggressive, less androgen-dependent phenotype compared with tumors that emerge with more rapid kinetics in context of higher androgen levels (due to more robust adrenal contribution and/or suboptimally suppressed T levels). While this may not influence response to subsequent non–AR-specific agents such as docetaxel, it is likely to adversely influence response to next-generation ARtargeted therapy. This is consistent with findings in the metaanalysis discussed above, in which lower T levels were a consistently poor prognostic factor for OS in patients with CRPC treated with AR-targeting agents, but not in those treated with chemotherapy (27).

The significant variation in adrenal androgen levels is not well understood. Functional polymorphisms in genes encoding critical steroidogenic enzymes such as CYP17A, HSD3B1, SULT2A1, AKR1C3, SRD5A, and UGT2B17 influence enzyme activity and/or production of downstream steroids (21, 30–32). Population-based studies will be required to determine whether a composite haplotype of genes involved in adrenal androgen production exists that may account for the observed spectrum of serum androgen levels. Alternatively, the variability might reflect genetic variation in regulatory factors such as *CYB5A1*, *NR5A1* (also known as SF-1 or steroidogenic factor 1), or *MC2R* [the adrenocorticotropic hormone (ACTH) receptor] that are upstream of adrenal steroid synthesis (33–35).

We found no association of abiraterone levels with PSA decline or with time-to-radiographic progression, with nearly all patients having serum levels above the 8.4 ng/mL value previously proposed as a cutoff for PSA response (36). This contrasts with the observation of Friedlander and colleagues who found in a study of 41 men with mCRPC that patients who failed to show a PSA response to AA had lower abiraterone levels after 4 weeks and at progression (22). This difference may reflect variability in timing of the blood draws, which, while mandated before the daily dose, were not specifically recorded precluding correction for differences based on pharmacokinetics (18). However, our findings are consistent with the work of Smulewitz and colleagues, who also found no correlation between abiraterone levels and change in PSA in a study of 72 men with mCRPC randomized to standard or low-dose AA (37), as well as with an analysis performed by the FDA in which no association was found between trough abiraterone levels and OS (38).

The adrenal-permissive variant of HSD3B1 (promotes conversion of AED and T to downstream androgens, and of abiraterone to D4abiraterone and 3-keto-5\alpha-abiraterone) has been consistently associated with more rapid progression on ADT (39), and potentially with worse outcomes in response to AA and enzalutamide in some but not all studies (40-43). Although we did find an association of HSD3B1 status with D4-abiraterone (the immediate downstream product of HSD3B1), we did not observe an association of HSD3B1 genotype with response to abiraterone, nor the previously reported association with 3-keto-5 α -abiraterone, potentially reflecting that serum levels were not corrected for differences based on pharmacokinetics. (18) However, we did observe that serum levels of 3-keto- 5α -abiraterone above the median at weeks 4 and 8 were associated with a more rapid time-toprogression. These observations are consistent with the ability of this metabolite to act as an AR agonist, and suggest that dose escalation of AA may in fact be detrimental to patient outcomes. These findings provide support for the low-dose AA strategy proposed by Smulewitz (37), and provide impetus for clinical trials combining AA therapy with an SRD5A inhibitor to prevent conversion of D4-abiraterone to 5α -keto metabolite (44, 45).

NGS of CRPC tumors has identified aberrations in multiple genes including AR, TP53, PTEN, and SPOP which have been explored as predictive biomarkers of response to AR-pathway inhibition. In this small study we found no association of AR, TP53, or SPOP status with response to AA (46-48). In our cohort, WT PTEN was weakly associated with longer time-to-progression on AA plus prednisone, consistent with Ferraldeschi and colleagues who reported a retrospective study of 144 patients in which PTEN loss was associated with shorter OS (14 vs. 21 months; P = 0.004) and shorter duration of abiraterone treatment (24 vs. 28 weeks; P = 0.009; ref. 49). Moreover, whereas poor response to AA in men with the lowest serum DHEAS levels was not additionally influenced by PTEN status, in those with higher androgen levels WT PTEN versus PTEN loss conferred a significantly longer time-to-progression (61 vs. 33 weeks; P = 0.02). While our study is too small to meaningfully predict genomic alterations associated with response to AR-pathway inhibition, these data suggest that interpretation of tumor sequencing data may be more informative if analyzed in context of the risk conferred by low versus high serum androgen levels.

This study has several important limitations, including small sample size, that abiraterone and metabolite levels were not corrected for pharmacokinetic differences based on timing of trough blood draws, and that analyses were exploratory with no attempt to correct for multiple testing. As such, our observations must be considered hypothesis generating and require validation in larger data sets.

Our data show that higher circulating adrenal androgens in patients with CRPC are strongly associated with an androgenrich tumor microenvironment before and during therapy with AA and prednisone, and suggest that ambient androgen levels in the castrate tumor microenvironment are an important determinant of prostate-tumor biology and response to therapy (13). Baseline and on-treatment androgen levels are likely to be important in the stratification and interpretation of trials evaluating AR-pathwaydirected therapy. In particular, while addition of AA to enzalutamide did not prolong survival in metastatic CRPC (50), higher baseline DHEAS levels may distinguish a subset of patients who do benefit from dual-AR therapy. Conversely, patients with CRPC in the lowest quartile of serum DHEAS levels may warrant stratification to regimens that include non-AR-directed therapy. Whether adrenal androgens associate with response to AA in metastatic CSPC is unknown but requires exploration. Prospective studies evaluating baseline and on-treatment androgen levels as predictive biomarkers of response to AR-directed therapy are required to test these hypotheses.

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A.M. Matsumoto: Resources, data curation, supervision, writing-review and editing.
P.S. Nelson: Conceptualization, resources, funding acquisition, writing-review and editing.

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