https://doi.org/10.1093/bib/bbab216 Review

Predicting MHC class I binder: existing approaches and a novel recurrent neural network solution

Limin Jiang, Hui Yu, Jiawei Li, Jijun Tang, Yan Guo and Fei Guo

Corresponding authors: Fei Guo, School of Computer Science and Engineering, Central South University, Changsha, China. E-mail: guofeieileen@163.com; Yan Guo, Comprehensive cancer center, Department of Internal Medicine, University of New Mexico, Albuquerque, NM, USA. E-mail: yanguo1978@gmail.com

Abstract

Major histocompatibility complex (MHC) possesses important research value in the treatment of complex human diseases. A plethora of computational tools has been developed to predict MHC class I binders. Here, we comprehensively reviewed 27 up-to-date MHC I binding prediction tools developed over the last decade, thoroughly evaluating feature representation methods, prediction algorithms and model training strategies on a benchmark dataset from Immune Epitope Database. A common limitation was identified during the review that all existing tools can only handle a fixed peptide sequence length. To overcome this limitation, we developed a bilateral and variable long short-term memory (BVLSTM)-based approach, named BVLSTM-MHC. It is the first variable-length MHC class I binding predictor. In comparison to the 10 mainstream prediction tools on an independent validation dataset, BVLSTM-MHC achieved the best performance in six out of eight evaluated metrics. A web server based on the BVLSTM-MHC model was developed to enable accurate and efficient MHC class I binder prediction in human, mouse, macaque and chimpanzee.

Key words: variable recurrent neural network; Position-specific scoring matrix; long short-term memory; MHC class I

Introduction

Major histocompatibility complex (MHC) is a set of genes coding for cell surface proteins essential for immune surveillance. Due to variation in function, molecular structure and distribution, MHC molecules are classified into MHC class I, MHC class II and MHC class III. MHC class I proteins are transported to the cell surface in almost all cells and they display an antigen to provide signals for cytotoxic T lymphocytes including cluster of differentiation (CD8+). Found on the special antigen-presenting immune cells including macrophages, dendritic cells and B cells, MHC class II proteins typically bind with CD4+ receptors on the helper T cells to clear exogenous antigens. MHC class III genes are interspaced with class I and II genes on the short arm of chromosome 6, but their proteins play different physiologic roles. Of the three MHC classes, class I received the most attention in biomedical research due to its universality. For example, reduction in MCH class I is associated with poor prognosis [1]. Recently, a study [2] showed that tumors can escape T-cell responses by losing MHC class I proteins. MHC genes are highly polymorphic, and a particular variant of an MHC gene/protein is usually termed an MHC allele. Each MHC class contains multiple alleles. For example, in MHC class I, groups of human leukocyte antigen (HLA) A, B and C are defined, and each group is composed of multiple alleles. To determine the binding between an antigenic peptide and an MHC protein, researchers usually use such gold standard methods as enzyme-linked immunosorbent spot, intracellular cytokine staining, competitive binding assays, the direct binding assay and the real-time kinetic binding assay. Beyond these experimental methods, computational algorithms can help with inferring the binding affinity between an MHC and a potential antigen. In particular, computational approaches can efficiently prioritize plausible candidates for an optimal study

Hui Yu is a research fellow at the Department of Internal Medicine, University of New Mexico.

Limin Jiang is a PhD candidate at Tianjin University. Her research interests include Bioinformatics and Machine Learning.

Jiawei Li is currently a master's degree candidate at Tianjin University. His research interests include Deep Learning and Machine Learning.

Jijun Tang is a professor at the University of South Carolina. His main research interests include Computational Biology and Algorithm.

Yan Guo is an associate professor in the Department of Internal Medicine, University of New Mexico. He is also the director of Bioinformatics Shared Resources of the University of New Mexico, Comprehensive Cancer Center.

Fei Guo is a professor at Central South University. Her research interests include Bioinformatics and Computational Biology. Submitted: 5 March 2021; Received (in revised form): 14 May 2021

[©] The Author(s) 2021. Published by Oxford University Press. All rights reserved. For Permissions, please email: journals.permissions@oup.com



Figure 1. Usage frequency of major MHC binding databases to miscellaneous prediction tools.

design, thus alleviating both time and financial burden of the major wet-lab validation experiments. Based on known MHCbinding sequences accumulated from decades of experimental validation efforts, around 30 MHC binding prediction tools have been developed to predict new binding sequences of MHC proteins. A majority of these tools [3–6] focused on MHC class I, because it has more specific properties than class II in most molecular processes, including cancer vaccines [7], cytosolic double-stranded DNA (dsDNA) in rat thyroid epithelial cells [8] and peptide fragments of the foreign protein [9].

Here, we review numerous MHC binding prediction tools and outline various aspects entailed in these computational approaches, including database sources, feature representation, machine learning strategies, etc. Our review is centered on MHC class I and it achieves a wide coverage by encompassing 13 MHC binding databases and 27 prediction tools. Moreover, we classified existing methods into two broad classes, machine learning and heuristic score. While reviewing the existing MHC class I binding tools, we identified a shared methodological limitation that all current approaches are limited to tackling a fixed peptide sequence length. To overcome this limitation, we developed a novel recurrent neural network solution based on bilateral and variable long short-term memory (BVLSTM), which achieves MHC class I prediction of variable peptide length. The implemented software product, with a specific application named BVLSTM-MHC. In the later portion of this review, we introduce the design of BVLSTM-MHC and demonstrate its superior performance in relation to existent MHC class I binding prediction tools.

Existing approaches

MHC binding databases

Experimentally verified MHC binding peptides were curated into databases. Over the last three decades, 13 MHC binding databases have been developed (Table 1). As of March 2021, nine databases are still functional, including two that had changed the hosting domain names. Two databases, Immune Epitope Database (IEDB) and Immuno Polymorphism Database of MHC (IPD-MHC), published updates within the prior 12 months. In terms of data quantity, the IEDB [10] hosts the largest collection of MHC-binding peptides with more than 900 000 peptides, followed by MHCBN [11] with 25 860 peptides. All existing databases focus on MHC class I and II binder, except for IMGT/MHC. And all databases mostly curate human data. These databases of experimentally verified MHC-binding sequences provide a foundation for the development of computational methods to predict an affinity score between a novel peptide and an MHC allele. Over the years, many MHC binding prediction tools have been developed based on data from these MHC binding databases. In following section, we focus on 27 mainstream prediction tools. The reliance on respective databases by various MHC class I binding prediction tools is illustrated in Figure 1. The largest database IEDB [10] has been used most frequently to MHC class I prediction tools, contributing to the development of 19 tools.

Strategies for existing MHC class I prediction methods

There are two major strategies for MHC binding prediction: machine learning and calculation score. Machine learning-based methods usually follow the following three steps: 1) feature representation; 2) training; and 3) evaluation. Calculation score based methods replace the training step with a scoring model. In this review, we discuss 27 MHC class I binding prediction methods. These methods' classifications are available in Table 2.

Feature representation

Feature representation is a technique to digitally code properties of an amino acid sequence as an MHC binding target. Over the last three decades, many feature representation methods have

Database	Address	Online	Last update	Entries	Species	Class
IEDB [10]	http://www.iedb.org/	Yes	2019	>900,000	Human, Mouse and Chimpanzee et al.	I, II
IPD-MHC [12]	https://www.ebi.ac.uk/ipd/mhc/	Yes	2019	_	Canines, Felines et al.	I, II
IPD-IMGT/HLA [13]	https://www.ebi.ac.uk/ipd/imgt/hla/	Yes	2019	_	Human	I, II
SYFPEITHI [14]	http://www.syfpeithi.de/	Yes	2012	>7000	Human	I, II
EPIMHC [15]	http://imed.med.ucm.es/epimhc/	Yes	2009	4875	Human	I, II
MHCBN [11]	http://crdd.osdd.net/raghava/mhcbn/	Yes	2009	25,860	Human, Mouse, rat et al.	I, II
AntiJen [16]	http://www.ddg-pharmfac.net/antijen/A ntiJen/antijenhomepage.htm	Yes	2005	15,454	Human, Mouse, Rat et al	I, II
Bcipep [17]	https://webs.iiitd.edu.in/raghava/bcipep/	Yes	2005	1230	Human, Mice et al	Ι
MPID-T [18]	http://variome.bic.nus.edu.sg/mpidt/inde x.html	Yes	2005	187	Human,Rat, Murine	I, II
MPID [19]	http://variome.bic.nus.edu.sg/mpid/	No	2003	86	Uknown	I, II
JenPep [20]	http://www.jenner.ac.uk/JenPep	No	2002	12,336	Human	I, II
FIMM [21]	http://sdmc.krdl.org.sg:8080/fimm	No	2002	1591	Human	Ι
MHCpep [22]	http://wehih.wehi.edu.au/mhcpep/ftp.	No	1994	>4000	Human, Mouse, rat, et al.	I, II

Table 1. MHC binding databases

been developed for predicting MHC binders. Here we discuss nine feature representation methods. Foremost, BLOcks SUbstitution Matrix (BLOSUM) [28, 34, 36-39, 41, 43] and positionspecific scoring matrix (PSSM) [30, 34, 35, 44, 47-49] have been popular choices. With L denoting the length of the peptide in question, BLOSUM is a 20 (amino acid) × L matrix that represents substitution frequency between each pair of amino acids. PSSM is a similar 20 (amino acid) × L matrix that captures evolutionary conservation within amino acid pairs. The runtime of PSSM is substantially longer than BLOSUM because the PSSM needs to be extracted by comparing a database with large protein sequence. Stabilized matrix method (SMM) has been used in PickPocket [27] to construct the PSSM with shortened runtime. PickPocket incorporates information of the MHC molecule per se into the representation of the binders. Another widely used sequence representation method is one-hot encoding [50], which transforms a peptide sequence into a 20 (amino acid) × L binary matrix with one row denoting each amino acid. Another common method is quantitative matrix (QM), which considers the contribution of each amino acid in a peptide sequence [29]. QM can be obtained from BIMAS [51] server by inputting amino acid sequences.

In addition to the four aforementioned conventional methods, other approaches have also been developed. For example, several studies proposed to use protein structure to model MHC class I binders. For example, Altuvia et al. [52] developed the first computational method based on protein structure to uncover MHC class I binders. Subsequently, Altuvia et al. [53] further improved the structure-based method to distinguish candidate peptides that bind to hydrophobic binding pockets of MHC molecules. Schueler-Furman et al. [54] extended the structure-based computational method to a wider range of MHC class I alleles. Other noticeable methods followed these pioneer works. Han et al. [26] merged information of the HLA molecule and the length of the target peptide into a 34×9 matrix, where 9 is the length of peptide and 34 is HLA-I contact residues proposed in NetMHCpan [38]. Kim et al. [25] developed the peptide:MHC binding energy covariance (PMBEC) matrix based on physicochemical properties (aromatic, hydrophobic and acidic etc.) to represent peptide sequence. Vang et al. [40]

developed the HLA-Vec method to map amino acids to a 15dimensional vector space to represent a peptide sequence. Bui et al. [55] constructed a 20 (amino acid) \times L average relative binding coefficient matrix depending on geometric average binding affinity, or half maximal inhibitory concentration (IC50) of peptides.

Feature representation is a necessary component of MHC binding prediction tools. In one prediction tool, one or multiple feature presentation methods can be applied. In an evaluation study in 2003, Nielsen et al. [37] applied information from two feature presentation methods to predict MHC T-cell class I epitope and accordingly demonstrated that multiple sources of information can improve performance. Subsequent prediction tools generally exercised this concept. For example, DeepLigand [4] and NetMHCpan [38] used the combination of one-hot encoding and BLOSUM. ProPred1 [23] applied two types of QM from BIMAS. NetMHCpan 4.0 [43] combined BLOSUM and length of peptide flanking regions. Zhao et al. [46] combined BLOSUM and physicochemical properties. Nielsen et al. [37] integrated information from one-hot encoding, BLOSUM and Hidden Markov models (HMM). The incorporation of various feature representation methods into various binding prediction tools is illustrated in Figure 2.

Score based methods

Heuristic score-based methods [14, 23, 30, 34] came as classical approaches to predicting MHC binding. They calculate a quasiprobability affinity score for a peptide as a potential MHC binder, using three strategies: 1) sequence similarity [56, 57]; 2) motif incorporation [14, 58]; 3) matrix formulation [23, 34]. In the end, an affinity score or probability is used to decide binding potential with a threshold.

Sequence similarity based methods summarize the amino acid composition of known MHC binding peptides into a sequence profile, and then compute a MHC binding probability index for a new peptide [57]. Similar to sequence similaritybased methods, motif-based methods first construct a score motif based on known MHC binding peptides, then a new peptide is compared to the motif to obtain binding affinity score. The

Trainine						
strategy	Models	Dataset	Feature representation ^a	Prediction model ^b	Evaluation criteria ^c	Code or Webserver
One-to-One	SYFPEITHI [14] ProPred1 [23]	EMBL, RDBMS MHCBN	constructing motif pattern QM	Heuristic score Heuristic score	None Sensitivity, Specificity, Accuracy, MCC	http://www.syfpeithi.de/ http://www.imtech.res.in/raghava/propred1
	SVMHC [24]	MHCPEP, SYFPEITHI,	sparse encoding	SVM	MCC, Specificity, Sensitivity	http://www.sbc.su.se/svmhc/
	SMMPMBEC [25]	ENSEMBL Laboratories	PMBEC	SMM	AUC	http://www.mhc-pathway.net/smmpmbec.
	ConvMHC [26]	IEDB, Sette and Buus	physicochemical scores	CNN	F1, Precision, Sensitivity	http://jumong.kaist.ac.kr:8080/convmhc
	PickPocket [27]	laboratories IEDB	PSSM	Heuristic score	AUC. PCC	None
	Rasmussen et al. [28]	IEDB	BLOSUM matrix	ANN	AUC, PCC	http://www.cbs.dtu.dk/services/NetMHCstabpan
	Bhasin et al. [29]	MHCBN, SWISS-PROT	QM, BIMAS	ANN	Sensitivity, Specificity, Accuracy Precision	https://webs.iiitd.edu.in/raghava/nhlapred/neural.html
	RANKPEP [30]	MHCPEP, SYFPEITHI	PSSM	Heuristic score		www.mifoundation.org/Tools/rankpep.html
	ForestMHC [31]	PRIDE, SysteMHC, Other	BLOSUM matrix, one-hot,	RF	AUPR, Prec1%	None
			physicochemical			
	Javadi et al. [32]	IEDB	physicochemical,	RF	AUC, Accuracy, Sensitivity, DDV Specificity, NDV	None
	Wilson et al [5]	IEDR MassIVE	biochemical	RF	AIIC Accuracy Sensitivity	None
				2	Specificity	
	Mattsson et al. [33]	IEDB, SYSPETHI	BLOSUM matrix	ANN	PCC	None
One-to-Many	PSSMHCpan [34]	MHCBN, IEDB, SYFPEITHI,	BLOSUM matrix, PSSM	Heuristic score	ACC, AUC	https://github.com/BGI2016/PSSMHCpan
		Laboratories				
	NetMHC-3.0 [35]	IEDB, SYFPEITHI	PSSMs	ANN	PCC, AUC	http://www.cbs.dtu.dk/services/NetMHC-3.0
	NetMHC-4.0 [36]	IEDB, SYFPEITHI	BLOSUM matrix, sparse	ANN	PCC, AUC	http://www.cbs.dtu.dk/services/NetMHC-4.0
			encoding			
	NIELSEN et al. [37]	SYFPEITHI	sparse encoding, BLOSUM matrix	ANN	PCC, AUC, Sensitivity/PPV	None
	NetMHCpan [38]	IEDB	sparse encoding, BLOSUM	ANN	Specificity, Sensitivity, PCC	http://www.cbs.dtu.dk/services/NetMHCpan/
			matrix			
	NetMHCpan-3.0 [39]	IEDB, SYFPEITHI	BLOSUM matrix	ANN-based ML	PCC, AUC	www.cbs.dtu.dk/services/NetMHCpan-3.0
	HLA-CNN [40]	MHCBN, IEDB, Antijen, Syfpertrhi	HLA-Vec	CNN	SRCC, AUC	https://github.com/uci-cbcl/HLA-bind.
	NetH2nan [41]	TEDR	BLOSUM matrix	ANN	ALIC. Sensitivity	None
		TEDR IPD-IMGT datahase	BLOSITM matrix	CNN	PCC ALIC	httns://aith1h com/HYsxe/ACME
	NetWirCone [42]		Combination of MetMur	None		
		מעזוו	NetMHCpan, PickPocket			W W W.CUD. ULU. MILLEN INC. MILLEN IN W W.CUD. ULU.
	NetMHCpan-4.0 [43]	IEDB	BLOSUM matrix	ANN	AUC	http://www.cbs.dtu.dk/services/NetMHCpan-4.0/
	Lundegaard et al. [44]	IEDB	None	None	PCC, AUC	None
Many-to-Many	MHCflurry [45]	IEDB, Other	BLOSUM matrix	ANN	AUC, KRCC, PCC, PPV	https://github.com/openvax/mhcflurry
	CNN-NF [46]	IEDB, DB2013	Sequence, Hydropathy,	CNN	F1, AUC	https://github.com/zty2009/MHC-1/tree/master.
			Polarity, Length			

^bANN, Artificial Neural Network. CNN, Convolutional Neural Networks . ML, Machine Learning. HMM, Hidden Markov Models. SVM, Support Vector Machine. RF, Random Forest. ^cACC, Accuracy. AUC, Area Under the receiver-operating-characteristic Curve. AUPR, Area Under the Precision-Recall curve. MCC, Matthews Correlation Coefficient. PCC, Pearson Correlation Coefficient. SRCC, Spearman's Rank Correlation Coefficient. Prec1%, Precision in the top 1% of predictive Predictive Predictive Value. NPV, Negative Predictive Value.



Figure 2. Usage frequency of feature representation methods to MHC prediction tools.

score motif generally assumes integer values, with a bigger number for a higher occurrence frequency of the amino acid. To construct motifs from known sequences, existing methods all require the peptides to have an equal length. For example, SYFPEITHI [14] constructs a 20 (amino acid) \times 9 motif pattern, by combining the information from MHC ligands, where 9 is the fixed peptide length. Here the motif score ranges from 1 to 10. Similarly, Sette *et al.* [58] constructed a 20 (amino acid) \times 6 substitution motif, where 6 represents the length of the core region of peptides. Here the motif score ranges from 1 to 3, with a smaller number indicating substitutions with drastic effects on IA^d binding.

Lastly, matrix-based methods differ from sequence similaritybased and motif-based methods by constructing a 20 (amino acid) \times L matrix for a new peptide instead of known MHC binding peptides. For example, PSSMHCpan model [34] calculates a score to represent binding affinity based on PSSM. ProPred1 [23] uses QM to compute the score. Bui *et al.* [55] propose an average relative binding matrix to compute the score. All these methods proceed to convert the scores to quasi-IC50 values for making a binary decision with 500 as a typical threshold for IC50.

Traditional machine learning based methods

Traditional machine learning methods are frequently applied to develop MHC binding prediction tools. For example, SVMHC [24] encodes peptide sequence with one-hot encoding and uses support vector machine to predict MHC class I binding peptides. Liu et al. [59] applied physicochemical properties to a support vector machine regression model to construct the SVRMHC model for predicting binding affinity on mouse MHC I molecules. Kevin et al. [31], Javadi et al. [32] and Wilson et al. [5] used random forest (RF) model to predict immunogenic peptides, where a peptide sequence is encoded mainly through physicochemical properties. To improve the performance of MHC binding prediction, SMMPMBEC [25] applies a Bayesian framework to work with the peptide MHC binding energy covariance (PMBEC) similarity matrix.

Network-based machine learning has also been applied in MHC prediction. Luo *et al.* [60] constructed a network between HLA molecules and peptides, and thereby devised Nebula, a neighbor-edges-based and unbiased leverage algorithm, to discover new HLA–peptide binding. Later, Luo et al. [61] improved Nebula model to a derivative named sNebula.

Yet another category of machine learning methods frequently applied in MHC binding prediction is Artificial Neural Networks (ANN). In 2008, Lin et al. [62] demonstrated that ANN has better performance than score-based methods by comparatively evaluating 30 methods. Since then, multiple ANN-based tools have been developed. For example, NetMHC-3.0 [35] establishes a high-performance ANN web server for predicting peptide binders. NetMHCpan tools [38, 39, 43, 63] and MHCflurry [45] also used ANN to predict MHC binders, achieving excellent performance on multiple species. To prevent overfitting, NetMHCpan explored multiple ANN models with different number of neuron nodes and culminated on the optimal model with the highest area under the curve (AUC) or Pearson correlation coefficient (PCC) in 5-fold cross-validation. To improve the predicting performance of ANN, Nielsen et al. [37] and Zhang et al. [64] combined HMM and ANN to distinguish the MHC binding peptides. The dissection of MHC prediction tools by traditional machine learning models is illustrated in Figure 3.

Deep learning methods

Convolutional neural networks (CNN) [26, 40, 46] and Deep Residual Network [4] are two widely used deep learning models for predicting MHC binders. For example, HLA-CNN [40] and ConvMHC [26] use a CNN architecture and fully connected layers to predict MHC class I binders. DeepSeqPan [65] is a CNN based multi-task tool to calculate an IC50 affinity value and a binding probability simultaneously, which derive its feature information from the paired peptide and HLA molecule. Another CNN-based tool CNN-NF [46] is composed of two activation layers, two pool layers, one flatten layer and one full connection layer. DeepLigand [4] applies Deep Residual Network to construct an affinity prediction module and a peptide embedding module and then uses a fully connected layer to connect the two sub-modules for prediction.

Machine learning training strategy

Based on the sequence length of MHC binders, training strategies of machine learning methods can be classified into three categories of distinct training strategies: 1) One-to-One [14, 23, 24, 28,



Figure 3. Usage frequency of machine learning models to MHC prediction tools.

62, 66, 67], 2) One-to-Many [34, 38, 40–42, 49] and 3) Many-to-Many [45, 46] (Figure 4).

The One-to-One strategy limits the model to training peptides with the sample length. Thus, models of this type can only predict peptides of the same length as the model. Multiple models are necessary to predict peptides of various lengths. The most popular length choice for the One-to-One strategy is 9-mer [68], because in most cases binding affinity is measured for 9-mer peptides. The One-to-Many strategy was developed to overcome the challenge of length dependency. In 2008, Lundegaard *et al.* [44] proposed a strategy to encode 8-mer and10-mer peptides in addition to 9-mer peptides by considering insertion and deletion of amino acids. Subsequent tools [3, 34–36, 38, 39] extended the same idea to 11-mer peptides. Insertion and deletion of amino acids in One-to-Many and Many-to-Many strategies are illustrated in Figure 5.

Even though the One-to-Many strategy can be used to predict MHC binders with different amino acids, the models are trained on 9-mer peptides only. In addition, One-to-Many is limited to a maximum length of 11. The Many-to-Many strategy is a relatively novel approach. The MHCflurry [45] model proposed by O'Donnell et al. in 2018 encodes peptides of up to 15 amino acids as 15-mer sequences by inserting 'X' character, and uses 15-mer BLOSUM matrix to train the model. When inserting the 'X' characters, MHCflurry requires that no 'X' can be inserted into the first and last four amino acids of the peptide (Figure 5). Zhao et al. [46] applied the same principle as MHCflurry but trained the model with the CNN-NF algorithm. The Many-to-Many strategy has strength in that it allows the training of a single model for peptides of various lengths. Although these strategies to accommodate variable lengths have achieved good performance to predict MHC binders, the insertion or deletion of amino acids in peptide sequence can potentially alter the primary structure and thus lead to loss of information. It is worthwhile to train a length-independent model, which preserves peptide sequence structure. With this objective in mind, we developed BVLSTM-MHC, a bilateral and variable recurrent neural network-based method to overcome this limitation.

Evaluation parameters

In this review, we evaluated BVLSTM-MHC and existent MHC class I prediction tools using performance metrics including accuracy (ACC), Matthews correlation coefficient (MCC), precision, specificity, F1, sensitivity (also known as Recall), PCC area under the precision-recall curve (AUPR) and area under the receiver-operating-characteristic curve (AUC) [31]. The calculation of ACC, precision, specificity, sensitivity, F1 and MCC is described in Equations 1–6, where TP represents the number of true negative predictions, FP represents the number of true negative predictions, FP represents the number of false-negative predictions.

$$ACC = \frac{TP + TN}{TP + TN + FP + FN}$$
(1)

$$Precision = \frac{TP}{TP + FP}$$
(2)

$$Specificity = \frac{TP}{TN + FP}$$
(3)

$$Sensitivity = \frac{TP}{TP + FN}$$
(4)

$$F1 = \frac{2 \times (Precision \times Sensitivity)}{(Precision + Sensitivity)}$$
(5)

$$MCC = \frac{TP \times TN - FP \times FN}{\sqrt{(TP + FP) \times (TN + FN) \times (TP + FN) \times (TN + FP)}}$$
(6)

BVLSTM for MHC binder prediction

Datasets

Peptide–MHC binding relations from IEDB were extracted to train BVLSTM-MHC. Benchmark data, including 117 alleles of four species (Mouse, Human, Macaque and Chimpanzee), were also downloaded from IEDB. The sequence length of the IEDB entries ranges from 8 to 18 amino acids. An independent validation



Figure 4. Categorization of MHC prediction tools into three training strategies.



Figure 5. Changing length of peptides in One-to-Many and Many-to-Many strategies. For One-to-Many, 8-mer, 10-mer and 11-mer peptides are changed to 9-mer peptide by inserting 'X' character and deleting amino acid. For Many, 8–12-mer peptides are changed to 15-mer peptide by inserting 'X' character.

dataset, consisting of 42 MHC binders and 179 non-binders, was extracted from MHCBN [11]. Organization of positive examples (binders) and negative examples (binders) for MHC allele classes of four species is shown in Table 3 and Supplementary Figure 1, where species-omitted notations HLA-A, HLA-B and HLA-C all relate to human.

dataset	Species	MHC Allele class	Number of Alleles	9-mer binders	9-mer non-binder	non-9-mer binders	non-9-mer non-binder
IEDB	Mouse	H-2	6	1375	3165	1343	3899
dataset	Human	HLA-A	37	19,716	52,835	8438	16,640
		HLA-B	39	9367	36,256	2317	7510
		HLA-C	9	871	1163	8	162
	Macaque	Mamu-A Mamu-B	18	2716	3568	2256	5486
	Chimpanzee	Patr-A Patr-B	8	514	1440	544	1204
	-	Total	117	34,559	98,427	14,906	34,901
MHCBN	Human	HLA-A	10	13	106	10	30
dataset		HLA-B	5	8	22	11	21
		Total	15	21	128	21	51

Table 3. Details of training data from IEDB and validation data from MHCBN

BVLSTM-MHC model

Here, we develop a recurrent neural network framework of strong scalability, BVLSTM-MHC, to predict MIC class I binders (Figure 6). MHC binders are first transformed into a 20 (amino acid) \times L matrix with one-hot encoding, where L is the number of amino acids of MHC binder. Then, one-hot encoding matrix and BLOSUM are combined to a 20 × L matrix to represent the peptide sequence. BLOSUM is used to initialize 20 convolution kernels. This process is shown in the section of the feature extraction layer in Figure 6. Then, the merged matrix is put into BVLSTM Encode Model, which is composed of two sets of LSTM. In the BVLSTM Encode Model, LSTM block dynamically changes with the sequence length. One set of LSTM processes the input matrix from left to right and the other set of LSTM processes the input matrix from right to left. This processing mechanism makes handling peptides of different lengths in one same model possible. Then, two fully-connected layers with 64 and 1 neurons, respectively, are used to handle the output vector of BVLSTM model. Finally, the Sigmoid function is used to normalize the output value to the predicted probability. All allele models shared these two fully-connected layers. In the training process, a dropout layer is applied to avoid over-fitting. The mean squared error is applied as the loss function. In the final models, the learning rate and the parameter of the dropout layer were set at 10^{-4} and 0.8, respectively, while a survey of a wide range of learning rates and dropout rates layer were shown in Supplementary Figure 2.

Evaluation of four feature representation methods

Four feature representation approaches, one-hot encoding, PSSM, 57 physicochemical properties [69], and the combination of the three were compared with the IEDB dataset. PSSM was extracted by NCBI blast-2.2.29 on UniProt [70] database. ANN and RF were performed to analyze the four approaches on 9mer peptides using five-fold cross-validation. Hidden neurons in ANN were set to 15 and the number of tree in RF was set to 500. PCC and AUC were used to assess the performance and results were shown in Figure 7. Data of 117 alleles were classified into six categories: HLA-A, HLA-B, HLA-C, Chimpanzee, Mouse and Macaque, where the first three categories belonged to Human. For ANN, the feature representation method PSSM achieved the highest medians for AUC and PCC in three categories. The combination of three individual methods achieved the best medians for AUC and PCC on the category of HLA-C, and One-hot encoding achieved the best medians of AUC and PCC for the categories of HLA-B and Chimpanzee. For RF, PSSM achieved the highest medians of AUC and PCC in five categories. The combination of three individual methods achieved the best performance of the median of AUC and PCC on the category of Chimpanzee and Mouse, respectively. Overall, PSSM had the best performance. Our analysis results also demonstrated that RF had higher AUCs and PCCs than ANN (Figure 7). This may be due to the fact that we used a fixed number of hidden neurons, as opposed to a variable number of hidden neurons.

Evaluation of four recurrent neural network models

We compared the performance of BVLSTM with three RNN models: bilateral variable gated recurrent units (BVGRU), LSTM and GRU. BVLSTM and BVGRU are bilateral variable-length models. LSTM and GRU are single-direction, fixed-length models. Four models were trained on 85 human alleles with 9-mer peptides. Bilateral Variable models performed better than LSTM and GRU models (Figure 8A). BVLSTM obtained the highest medians for AUC and PCC on HLA-A and HLA-B categories and the highest medians for AUPR on the HLA-A category. These results indicate that BVLSTM is a potentially optimal approach for predicting MHC binder peptides.

Cross-validation performance of BVLSTM-MHC

The IEDB dataset was split into 10 portions: seven were used for training, one was used as validation to select the optimal model, and the rest two were used for final testing. After five-fold cross-validation and validation, the final test results were as follows. Of the 117 alleles, 77 had AUC higher than 0.8 (62 human, 10 macaque, 3 mouse and 2 chimpanzee) (Figure 8B and Supplementary Table 1). For human HLA-A and HLA-B datasets, the medians of AUC were greater than 0.9 on both test datasets. The medians of AUC were above 0.8 for mouse, chimpanzee and HLA-C. AUCs of peptides with different lengths (8–18mers) were calculated to evaluate the variable-length performance of BVLSTM-MHC (Figure 8C and Supplementary Table 1). For human HLA-A and HLA-B, the median AUCs were greater than 0.9 regardless of peptide length. For HLA-C, mouse and chimpanzee, the median AUC of variable length peptides were closer to 0.8.

Performance of BVLSTM-MHC in comparison with existent MHC class I prediction methods

BVLSTM-MHC was compared to 10 popular MHC class I predictors: ANN [44], comblibsidney2008 [71], NetMHCcons [42],



Figure 6. Schema of BVLSTM-MHC. Two major processes are detailed, namely feature representation and BVLSTM Encode model.



Figure 7. Distribution of AUCs and PCCs returned by four feature representation methods. The four methods include PSSM, one-hot encoding, 57 physicochemical properties and combination of the former three (combination).



Figure 8. Prediction performance of the BVLSTM-MHC model in cross-validation. (A) Distribution of various performance metrics (AUC, AUPR and PCC) for various recurrent neural network models (BVGRU, BVLSTM, GRU and LSTM), stratified by MHC allele classes (HLA-A, HLA-B and HLA-C). (B) Distribution of AUC of the BVLSTM-MHC model on the test dataset. Results for chimpanzee and HLA-C were not plotted due to limited data points. (C) Distribution of AUC of the BVLSTM-MHC model on the test dataset, stratified by peptide length. Results for chimpanzee and HLA-C were not plotted due to limited data points. (D) Receiver-operating-characteristic curves of BVLSTM together with another 10 competitors. AUC values were annotated to each method.

Table 4. Performance of BVLSTM-MHC along with ten existent MHC cl	lass I predictors	on independent	MHCBN dataset
---	-------------------	----------------	---------------

Methods	ACC	AUC	F1	MCC	Specifici	ty Sensitivi	ty Precision	AUPR	# Postive examples	# Negative examples
ANN [44]	0.8827	0.9166	0.6866	0.6205	0.9507	0.6216	0.7667	0.7726	37	142
comblibsidney 2008 [71]	0.7419	0.1141	_	0.0000	1.0000	0.0000	0.0000	0.1591	16	46
NetMHCcons [42]	0.8883	0.9169	0.7222	0.6528	0.9366	0.7027	0.7429	0.8457	37	142
NetMHCpan [63]	0.8547	0.9024	0.5938	0.5173	0.9437	0.5135	0.7037	0.7471	37	142
NetMHCpan EL [43]	0.8156	0.8258	0.5075	0.3989	0.9085	0.4595	0.5667	0.5977	37	142
PickPocket [27]	0.8715	0.8429	0.6849	0.6043	0.9225	0.6757	0.6944	0.6378	37	142
SMM [72]	0.9268	0.9508	0.7692	0.7266	0.9635	0.7407	0.8000	0.8548	27	137
SMMPMBEC [25]	0.9146	0.9586	0.7308	0.6808	0.9562	0.7037	0.7600	0.8581	27	137
BVLSTM-MHC	0.9548	0.9512	0.8750	0.8490	0.9832	0.8333	0.9211	0.9112	42	179
CNN-NF [46]	0.8606	0.8513	0.6234	0.5449	0.8908	0.7059	0.5581	0.6383	34	174
MHCflurry [45]	0.7834	0.7526	0.4946	0.3633	0.8352	0.5610	0.4423	0.4569	41	176

NetMHCpan [63], NetMHCpan EL [43], PickPocket [27], SMM [72], SMMPMBEC [25], CNN-NF [46] and MHCflurry [45] (Figure 8D). MHCBN dataset was used as an independent evaluation. Eight evaluation criteria (AUC, ACC, F1, MCC, specificity, sensitivity, precision and AUPR) were computed. The overall results can be viewed in Table 4 and the length specific results can be viewed in Supplementary Table 2. SMMPMBEC and obtained the best AUC (0.96), BVLSTM-MHC was closely behind with an AUC of 0.95. BVLSTM-MHC achieved the best performance for the other six parameters.



Figure 9. Binding prediction of BVLSTM-MHC is largely correlated with evolutionary conservation of amino acid positions. Using a 9-mer peptide of YMLDMTFPV sequence and the motif of allele HLA-A*02:19 as an example, we partially interpret the rationale of BVLSTM-MHC predicting MHC binding. For each position, we replace the amino acid with an 'X' character designating an unknown amino acid, thus generating 9 modulated sequences ('Sequence' column). BVLSTM-MHC reported a new binding probability score for each modulated sequence ('Predicted Value' column). The difference of MHC binding probability score between the original case and the modulated case was obtained and regarded as the impact value of each individual amino acid position ('Impact Value' column). Of note, we indicated decreased binding probability with red bars and increased binding probability with green bars. The two amino acid positions that displayed the highest impact values (the second and no positions) were highlighted in red for their predicted values. Referring to the sequence logo of the binding motif (the rightmost column), it is evident that BVLSTM-MHC model predicts binding propensity largely based on the evolutionary conservation of amino acid positions.

Association between BVLSTM-MHC and positional conservation

Despite high performance, deep learning models are often criticized for the lack of interpretability as a black box. Multiple tools such as LIME [73], SHAP [74] and DeepLIFT [75] have been designed to help understand the underlying mechanism of deep learning models. Unfortunately, our BVLSTM-MHC models were developed using a TensorFlow class that is not supported by existing interpretation tools yet. Nevertheless, inspired by these tools, we developed a strategy to interpret BVLSTM-MHC rationale. Using an exemplar 9-mer peptide and the BVLSTM-MHC model for allele HLA-A*02:19 as an example, we interpreted the rationale of BVLSTM-MHC prediction (Figure 9). Given the sequence logo for the motif of allele HLA-A*02:19 (extracted by ggseqlogo R package [76]), it was revealed that the impact of each position captured by BVLSTM-MHC matches the positionwise conservation level. This suggests that BVLSTM-MHC model predicts binding propensity largely based on the evolutionary conservation of amino acid positions.

BVLSTM-MHC webserver

Given all the above parameters and component optimization, we developed an online server BVLSTM-MHC to perform MHC

binder prediction. Because we reserved only MHC alleles showing an interim AUC of 0.8 or higher, the ultimate web server spans four species (Human, mouse, macaque and chimpanzee) and covers 77 MHC class I alleles in total. BVLSTM-MHC was developed with R, PHP and Python languages, and it is accessible at http://www.innovebioinfo.com/Proteomics/MHCBIB/MHCI. php.

Discussion

MHC binding prediction is a crucial step for identifying potential novel therapeutic strategies. Many tools have been developed for this purpose. We introduced BVLSTM-MHC, a variable-length BVLSTM RNN based method, to predict MHC class I binders. In comparison to the 10 existent MHC class I binding prediction tools, BVLSTM-MHC performed best in six of eight evaluation parameters on an independent dataset. Most MHC class I predictors limited the maximum length of the peptides to 11 or 12, whereas BVLSTM-MHC currently extends maximum peptide length to 30, and the built-in variable-length design spares users the trouble to switch models of different length parameter. BVLSTM-MHC performed the best overall in six of the eight performance parameters when compared to 10 existing MHC class I prediction tools. The reason for performance improvement is probably attributed to the enlarged sample size and the ability to handle variable length without modifying peptide sequences. For the One-to-One model, it is necessary to build a prediction model for each length, thus inevitably limiting the sample size of each length-specific model. For the One-to-Many model, 9mer peptides are used to train a prediction model and peptides with non-9-mer can be predicted by inserting 'X' character or deleting amino acids in the peptide sequence. For the Manyto-Many model, peptides of different lengths are all padded up to a uniform 15-mer length by inserting 'X' characters. One-to-Many and Many-to-Many models potentially alter the primary structure thus leading to loss of information, because primary structure determines tertiary structure to a large extent. Overall, this work yielded a web server based on the BVLSTM-MHC models, which can predict MHC class I binders of 77 alleles from four species.

Key points

- We comprehensively summarized 13 MHC databases and 27 prediction tools for MHC class I binding.
- We developed a variable-length MHC class I binding prediction tool based on remaining original peptide sequence, BVLSTM-MHC.
- BVLSTM-MHC performed best in six out of eight evaluated metrics when compared to the 10 mainstream MHC class I binding predictors.
- We developed a web server based on BVLSTM-MHC to predict human, mouse, macaque and chimpanzee MHC Class I alleles.

Supplementary Data

Supplementary data are available online at https://academic.oup.com/bib.

Funding

The National Natural Science Foundation of China (NSFC 61772362, 61972280); the National Key R&D Program of China (2020YFA0908400); the National Cancer Institute, USA (Grants P30CA118100 and R01ES030993-01A1 to Y.G.).

Data Availability

Data are available at http://www.innovebioinfo.com/Proteo mics/MHCBIB/MHCI.php.

References

- Roemer MGM, Advani RH, Redd RA, et al. Classical Hodgkin Lymphoma with Reduced beta M-2/MHC Class I Expression Is Associated with Inferior Outcome Independent of 9p24.1 Status. Cancer Immunol Res 2016;4:910–6.
- Garrido F, Aptsiauri N. Cancer immune escape: MHC expression in primary tumours versus metastases. *Immunology* 2019;158:255–66.
- 3. Hu Y, Wang Z, Hu H, et al. ACME: pan-specific peptide-MHC class I binding prediction through attention-based deep neural networks. Bioinformatics 2019;**35**:4946–54.
- Zeng H, Gifford DK. DeepLigand: accurate prediction of MHC class I ligands using peptide embedding. *Bioinformatics* 2019;35:i278–83.

- Wilson EA, Krishna S, Anderson KS. A Random Forest based approach to MHC class I epitope prediction and analysis. The Journal of Immunology 2018;200:99.11–1.
- Jensen KK, Andreatta M. Improved methods for predicting peptide binding affinity to MHC class II molecules. 2018;154:394–406.
- Boraschi D, Italiani P, Palomba R, et al. Nanoparticles and innate immunity: new perspectives on host defence. Semin Immunol 2017;34:33–51.
- Yuqian Luo AY, Oda K, Ishido Y, et al. Naked DNA in cells: An inducer of major histocompatibility complex molecules to evoke autoimmune responses? World Journal of Translational Medicine 2016;5:46–52.
- Hudig D, Karimi R. Calreticulin in Cytotoxic Lymphocyte-Mediated Cytotoxicity. In: Eggleton P, Michalak M (eds). Calreticulin, Second edn. Boston, MA: Springer US, 2003, 142–50.
- Vita R, Mahajan S, Overton JA, et al. The Immune Epitope Database (IEDB): 2018 update. Nucleic Acids Res 2019;47: D339–d343.
- Bhasin M, Singh H, Raghava GP. MHCBN: a comprehensive database of MHC binding and non-binding peptides. Bioinformatics 2003;19:665–6.
- 12. Maccari G, Robinson J, Ballingall K, et al. IPD-MHC 2.0: an improved inter-species database for the study of the major histocompatibility complex. 2017;45:D860–d864.
- Robinson J, Halliwell JA, Hayhurst JD, et al. The IPD and IMGT/HLA database: allele variant databases. Nucleic Acids Res 2015;43:D423–31.
- 14. Rammensee H, Bachmann J, Emmerich NP, et al. SYFPEITHI: database for MHC ligands and peptide motifs. *Immunogenet*ics 1999;**50**:213–9.
- 15. Reche PA, Zhang H, Glutting JP, et al. EPIMHC: a curated database of MHC-binding peptides for customized computational vaccinology. *Bioinformatics* 2005;**21**:2140–1.
- 16. Toseland CP, Clayton DJ, McSparron H, et al. AntiJen: a quantitative immunology database integrating functional, thermodynamic, kinetic, biophysical, and cellular data. *Immunome Res* 2005;1:4.
- 17. Saha S, Bhasin M, Raghava GP. Bcipep: a database of B-cell epitopes. BMC *Genomics* 2005;**6**:79.
- Tong JC, Kong L, Tan TW, et al. MPID-T: database for sequence-structure-function information on T-cell receptor/ peptide/MHC interactions. Appl Bioinformatics 2006;5:111–4.
- Govindarajan KR, Kangueane P, Tan TW, et al. MPID: MHC-Peptide Interaction Database for sequence-structurefunction information on peptides binding to MHC molecules. Bioinformatics 2003;19:309–10.
- 20. Blythe MJ, Doytchinova IA, Flower DR. JenPep: a database of quantitative functional peptide data for immunology. Bioinformatics 2002;18:434–9.
- 21. Schönbach C, Koh JL, Sheng X, et al. FIMM, a database of functional molecular immunology. *Nucleic Acids Res* 2000;**28**: 222–4.
- Brusic V, Rudy G, Kyne AP, et al. MHCPEP-a database of MHCbinding peptides: update 1995. Nucleic Acids Res 1996;24: 242-4.
- 23. Singh H, Raghava GP. ProPred1: prediction of promiscuous MHC Class-I binding sites. Bioinformatics 2003;19:1009–14.
- 24. Dönnes P, Elofsson A. Prediction of MHC class I binding peptides. using SVMHC, BMC Bioinformatics 2002;**3**:25.
- 25. Kim Y, Sidney J, Pinilla C, et al. Derivation of an amino acid similarity matrix for peptide: MHC binding and its application as a Bayesian prior. BMC Bioinformatics 2009;10: 394.

- Han Y, Kim D. Deep convolutional neural networks for pan-specific peptide-MHC class I binding prediction. BMC Bioinformatics 2017;18:585.
- Zhang H, Lund O, Nielsen M. The PickPocket method for predicting binding specificities for receptors based on receptor pocket similarities: application to MHC-peptide binding. *Bioinformatics* 2009;25:1293–9.
- Rasmussen M, Fenoy E. Pan-Specific Prediction of Peptide-MHC Class I Complex Stability, a Correlate of T Cell Immunogenicity. 2016;197:1517–24.
- 29. Bhasin M, Raghava GP. A hybrid approach for predicting promiscuous MHC class I restricted T cell epitopes. *J Biosci* 2007;**32**:31–42.
- Reche PA, Glutting JP, Reinherz EL. Prediction of MHC class I binding peptides using profile motifs. Hum Immunol 2002;63:701–9.
- Boehm KM, Bhinder B, Raja VJ, et al. Predicting peptide presentation by major histocompatibility complex class I: an improved machine learning approach to the immunopeptidome. BMC Bioinformatics 2019;20:7.
- Javadi A, Khamesipour A, Monajemi F, et al. Computational Modeling and Analysis to Predict Intracellular Parasite Epitope Characteristics Using Random Forest Technique. Iran J Public Health 2020;49:125–33.
- Mattsson AH, Kringelum JV, Garde C, et al. Improved panspecific prediction of MHC class I peptide binding using a novel receptor clustering data partitioning strategy. Hla 2016;88:287–92.
- Liu G, Li D, Li Z, et al. PSSMHCpan: a novel PSSM-based software for predicting class I peptide-HLA binding affinity. *Gigascience* 2017;6:1–11.
- 35. Lundegaard C, Lamberth K, Harndahl M, et al. NetMHC-3.0: accurate web accessible predictions of human, mouse and monkey MHC class I affinities for peptides of length 8-11. Nucleic Acids Res 2008;36:W509–12.
- Andreatta M, Nielsen M. Gapped sequence alignment using artificial neural networks: application to the MHC class I system. Bioinformatics 2016;32:511–7.
- Nielsen M, Lundegaard C, Worning P, et al. Reliable prediction of T-cell epitopes using neural networks with novel sequence representations. Protein Sci 2003;12:1007–17.
- Nielsen M, Lundegaard C, Blicher T, et al. NetMHCpan, a method for quantitative predictions of peptide binding to any HLA-A and -B locus protein of known sequence. PLoS One 2007;2:e796.
- Nielsen M, Andreatta M. NetMHCpan-3.0; improved prediction of binding to MHC class I molecules integrating information from multiple receptor and peptide length datasets. *Genome Med* 2016;8:33.
- 40. Vang YS, Xie X. HLA class I binding prediction via convolutional neural networks. *Bioinformatics* 2017;**33**:2658–65.
- 41. DeVette CI, Andreatta M. NetH2pan: A Computational Tool to Guide MHC Peptide Prediction on Murine Tumors. 2018;6:636–44.
- 42. Karosiene E, Lundegaard C, Lund O, et al. NetMHCcons: a consensus method for the major histocompatibility complex class I predictions. *Immunogenetics* 2012;**64**:177–86.
- Jurtz V, Paul S. NetMHCpan-4.0: Improved Peptide-MHC Class I Interaction Predictions Integrating Eluted Ligand and Peptide Binding Affinity Data. 2017;199:3360–8.
- 44. Lundegaard C, Lund O, Nielsen M. Accurate approximation method for prediction of class I MHC affinities for peptides of length 8, 10 and 11 using prediction tools trained on 9mers. Bioinformatics 2008;24:1397–8.

- O'Donnell TJ, Rubinsteyn A, Bonsack M, et al. MHCflurry: Open-Source Class I MHC Binding Affinity Prediction. Cell Syst 2018;7:129–132.e124.
- Zhao T, Cheng L, Zang T, et al. Peptide-Major Histocompatibility Complex Class I Binding Prediction Based on Deep Learning With Novel Feature. Front Genet 2019;10.
- Nielsen M, Lundegaard C, Lund O. Prediction of MHC class II binding affinity using SMM-align, a novel stabilization matrix alignment method. BMC Bioinformatics 2007;8:238.
- Nielsen M, Lund O. NN-align. An artificial neural networkbased alignment algorithm for MHC class II peptide binding prediction. BMC Bioinformatics 2009;10:296.
- Gfeller D, Guillaume P, Michaux J, et al. The Length Distribution and Multiple Specificity of Naturally Presented HLA-I Ligands. The Journal of Immunology 2018;201:3705–16.
- Davis MJ. Contrast Coding in Multiple Regression Analysis: Strengths, Weaknesses, and Utility of Popular Coding Structures. Journal of data science 2010;8:61–73.
- Parker KC, Bednarek MA, Coligan JE. Scheme for ranking potential HLA-A2 binding peptides based on independent binding of individual peptide side-chains. *The Journal of Immunology* 1994;152:163–75.
- 52. Altuvia Y, Schueler O, Margalit H. Ranking potential binding peptides to MHC molecules by a computational threading approach. J Mol Biol 1995;**249**:244–50.
- Altuvia Y, Sette A, Sidney J, et al. A structure-based algorithm to predict potential binding peptides to MHC molecules with hydrophobic binding pockets. *Hum Immunol* 1997;58:1–11.
- Schueler-Furman O, Altuvia Y, Sette A, et al. Structure-based prediction of binding peptides to MHC class I molecules: application to a broad range of MHC alleles. Protein Sci 2000;9:1838–46.
- Bui HH, Sidney J, Peters B, et al. Automated generation and evaluation of specific MHC binding predictive tools: ARB matrix applications. *Immunogenetics* 2005;57:304–14.
- 56. Celis E, Larson J, Otvos L, Jr. et al. Identification of a rabies virus T cell epitope on the basis of its similarity with a hepatitis B surface antigen peptide presented to T cells by the same MHC molecule (HLA-DPw4), J Immunol 1990;145:305–10.
- 57. Rothbard JB, Taylor WR. A sequence pattern common to T cell epitopes. EMBO J 1988;7:93–100.
- Sette A, Buus S, Appella E, et al. Prediction of major histocompatibility complex binding regions of protein antigens by sequence pattern analysis. Proc Natl Acad Sci U S A 1989;86:3296–300.
- Liu W, Meng X, Xu Q, et al. Quantitative prediction of mouse class I MHC peptide binding affinity using support vector machine regression (SVR) models. BMC Bioinformatics 2006;7:182.
- Luo H, Ye H, Ng H, et al. Understanding and predicting binding between human leukocyte antigens (HLAs) and peptides by network analysis. BMC Bioinformatics 2015;16(Suppl 13):S9.
- Luo H, Ye H, Ng HW, et al. sNebula, a network-based algorithm to predict binding between human leukocyte antigens and peptides. Sci Rep 2016;6:32115.
- 62. Lin HH, Ray S, Tongchusak S, et al. Evaluation of MHC class I peptide binding prediction servers: applications for vaccine research. BMC Immunol 2008;9:8.
- Hoof I, Peters B, Sidney J, et al. NetMHCpan, a method for MHC class I binding prediction beyond humans. *Immuno*genetics 2009;61:1–13.
- Zhang GL, Khan AM, Srinivasan KN, et al. MULTIPRED: a computational system for prediction of promiscuous HLA binding peptides. Nucleic Acids Res 2005;33:W172–9.

- 65. Liu Z, Cui Y, Xiong Z, et al. DeepSeqPan, a novel deep convolutional neural network model for pan-specific class I HLA-peptide binding affinity prediction. Sci Rep 2019;9:794.
- Adams HP, Koziol JA. Prediction of binding to MHC class I molecules. J Immunol Methods 1995;185:181–90.
- 67. Trolle T, Metushi IG, Greenbaum JA, et al. Automated benchmarking of peptide-MHC class I binding predictions. Bioinformatics 2015;**31**:2174–81.
- Lundegaard C, Lund O, Buus S, et al. Major histocompatibility complex class I binding predictions as a tool in epitope discovery. Immunology 2010;130:309–18.
- Song J, Tang J, Guo F. Identification of Inhibitors of MMPS Enzymes via a Novel Computational Approach. Int J Biol Sci 2018;14:863–71.
- Apweiler R, Bairoch A, Wu CH, et al. UniProt: the Universal Protein knowledgebase. Nucleic Acids Res 2004;32:D115–9.
- 71. Sidney J, Assarsson E, Moore C, et al. Quantitative peptide binding motifs for 19 human and mouse MHC class I molecules derived using positional scanning combinatorial peptide libraries. *Immunome Res* 2008;**4**:2.

- 72. Peters B, Sette A. Generating quantitative models describing the sequence specificity of biological processes with the stabilized matrix method. *BMC Bioinformatics* 2005;6: 132.
- 73. Ribeiro MT, Singh S, Guestrin C. "Why Should I Trust You?": Explaining the Predictions of Any Classifier. Proceedings of the 22nd ACM SIGKDD International Conference on Knowledge Discovery and Data Mining. San Francisco. California, USA: Association for Computing Machinery, 2016, 1135–44.
- 74. Lundberg SM, Lee S-I. A unified approach to interpreting model predictions. In: Proceedings of the 31st International Conference on Neural Information Processing Systems. Long Beach, California, USA: Curran Associates Inc., 2017, 4768–77.
- 75. Shrikumar A, Greenside P, Kundaje A. Learning Important Features Through Propagating Activation Differences. In: Doina P, Yee WT (eds). Proceedings of the 34th International Conference on Machine Learning. Proceedings of Machine Learning Research: PMLR, 2017, 3145–53.
- 76. Wagih O. ggseqlogo: a versatile R package for drawing sequence logos. Bioinformatics 2017;**33**:3645–7.