Relationships of p16 Immunohistochemistry and Other Biomarkers With Diagnoses of Cervical Abnormalities:

Implications for LAST Terminology

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Abstract

Context.——Lower Anogenital Squamous Terminology (LAST) standardization recommended p16^{INK4a} immunohistochemistry (p16 IHC) for biopsies diagnosed morphologically as cervical intraepithelial neoplasia (CIN) grade 2 (CIN2) to classify them as low-grade or high-grade squamous intraepithelial lesions (HSILs).

Objective.——To describe the relationships of p16 IHC and other biomarkers associated with cervical cancer risk with biopsy diagnoses.

Design.——A statewide, stratified sample of cervical biopsies diagnosed by community pathologists (CPs), including 1512 CIN2, underwent a consensus, expert pathologist panel (EP) review (without p16 IHC results), p16 IHC interpretation by a third pathology group, and human papillomavirus (HPV) genotyping, results of which were grouped hierarchically according to cancer risk. Antecedent cytologic interpretations were also available.

Results.——Biopsies were more likely to test p16 IHC positive with increasing severity of CP diagnoses, overall (P_{trend} .001) and within each HPV risk group (P_{trend} .001 except for low-risk HPV [$P_{\text{trend}} < .010$]). All abnormal grades of CP-diagnosed biopsies were more likely to test p16 IHC positive with a higher HPV risk group ($P_{\text{trend}} < .001$), and testing p16 IHC positive was associated with higher HPV risk group than testing p16 IHC negative for each grade of CP-diagnosed biopsies ($P < .001$). p16 IHC–positive, CP-diagnosed CIN2 biopsies were less likely than CP-diagnosed CIN3 biopsies to test HPV16 positive, have an antecedent HSIL$^+$ cytology, or to be diagnosed as CIN3$^+$ by the EP ($P < .001$ for all). p16 IHC–positive, CP-diagnosed CIN1 biopsies had lower HPV risk groups than p16 IHC–negative, CP-diagnosed CIN2 biopsies ($P < .001$).

Conclusions.——p16 IHC–positive, CP-diagnosed CIN2 appears to be lower cancer risk than CP-diagnosed CIN3. LAST classification of "HSIL" diagnosis, which includes p16 IHC–positive CIN2, should annotate the morphologic diagnosis (CIN2 or CIN3) to inform all management decisions, which is especially important for young (<30 years) women diagnosed with CIN2 for whom surveillance rather than treatment is recommended.

Persistent cervical infections by 12 to 15 high-risk human papillomavirus (HPV) genotypes cause nearly all cervical cancers[1] and most of the immediate precursor cervical abnormalities, including cervical intraepithelial neoplasia (CIN) grade 2 (CIN2), grade 3

(CIN3), and adenocarcinoma in situ. HPV16 and HPV18 are the most carcinogenic HPV genotypes, with HPV16 causing approximately 50% to 60% of cervical cancers and HPV18 causing 10% to 15% of cervical cancers.[2] The other 10 to 13 HPV types cause the remaining 25% to 40% of cervical cancers.[2] With increasing severity of the cervical abnormality, attributable fractions due to HPV16 and HPV18 increase while those due to other types concomitantly decrease.[3]

CIN2 has been the threshold for cervical treatment, by either excision or ablative treatment.[4] However, recently there has been an increasing recognition that the hematoxylin-eosin (H&E) diagnosis of CIN2 is an equivocal diagnosis with significant interobserver variability and likely represents an admixture of (misclassified) HPV infection/CIN1 and precancer (CIN3)[5] rather than a biological intermediate step in the progression from CIN1 to CIN3 as was originally thought.[6] The uncertainty of the meaning of this diagnosis is perhaps reflected in its poor diagnostic reproducibility between pathologists.[7–11] Because CIN2 likely has overall low immediate potential to become invasive cancer, frequently regresses especially in young women (aged <30 years),[12] and excisional treatment is possibly associated with an increased risk of preterm delivery,[13,14] current management guidelines in the United States recommend "wait and watch" rather than treatment for CIN2 diagnosed in young women (aged <30 years) of reproductive potential when the squamocolumnar junction can be visualized in its entirety.[4]

There has been great interest in using adjunctive biomarkers to improve the classification and reliability of histopathologic diagnoses, based on H&E staining, of cervical abnormalities, especially to reduce the overdiagnosis of CIN2 on H&E and clarify the clinical significance of CIN2 (ie, distinguish between benign CIN2 diagnoses potentially destined to regress or not progress from CIN2 diagnoses that reflect the presence of high-grade cervical abnormalities that for safety should be treated to reduce the risk of cancer development). Some of the biomarkers that have been investigated for clarifying the meaning of an H&E diagnosis of CIN2 on biopsy include (but are not limited to) HPV16,[15] HPV L1,[16,17] Ki-67,[7,16] E4,[18,19] and p16INK4a (p16)[7,11,16] detection.

Immunohistochemistry (IHC) for in situ detection of p16 (p16 IHC) has emerged as an adjunctive biomarker to aid in the diagnosis of cervical abnormalities. p16 IHC has been shown to be sensitive for CIN2 and CIN3[7,14] and its interpretation is much more reliable/reproducible than morphology based on H&E staining alone.[11,20,21] Recommendations from the Lower Anogenital Squamous Terminology (LAST) Standardization Project include the use of p16 IHC in the following specific circumstance[22]: "If the pathologist is entertaining an H&E morphologic interpretation of –IN 2 (under the old terminology, which is a biologically equivocal lesion falling between the morphologic changes of HPV infection [low-grade lesion] and high-grade cervical abnormalities), p16 IHC is recommended to help clarify the situation. Strong and diffuse block-positive p16 IHC results support a categorization of precancer. Negative or non–block-positive staining strongly favors an interpretation of low-grade disease or a non–HPV-associated pathology." LAST recommended a switch from the 3-tier categorization—CIN1, CIN2, and CIN3—to a 2-tier system of categorization of low-grade squamous intraepithelial lesion, which includes CIN1

and p16 IHC–negative CIN2, and high-grade squamous intraepithelial lesion (HSIL), which includes CIN3 and p16 IHC–positive CIN2.

However, the question remains about whether p16 IHC distinguishes between benign HPV infection and clinically significant CIN2, that is, those that have or will develop invasive potential, thereby representing a high-grade cervical abnormality. Obviously, it is not logistically or ethically possible to follow up a cohort of women diagnosed with CIN2 to see who develops cervical cancer to answer this question, as was done tragically with CIN3/carcinoma in situ.[23] The subsequent diagnosis of CIN3 in follow-up of CIN2 cases may not be true progression but rather a correction of a previously misclassified CIN2 diagnosis and sampling errors including missed CIN3 at colposcopy.

To better understand the cervical cancer risk stratification achieved by p16 IHC for routine diagnoses (community pathology [CP]) of CIN2 as well as other diagnoses, we conducted a large US population–based study of p16 IHC and its relationship to other biomarkers of cervical cancer risk, including an expert panel (EP) consensus review that has been shown to improve the certainty of high-grade cervical abnormalities and therefore the association with HPV,[24] tissue HPV genotyping, and antecedent cytology result. Increasing severity of histologic diagnosis rendered by an EP review of a CP diagnosis of CIN2 is associated with a CIN3 diagnosis on tissue from an excision procedure.[25] The percentage positive for the highest risk HPV genotypes, especially HPV16, increases with the severity of cervical diagnosis,[3,25] and the HPV genotype(s) detected in the diagnostic tissue generally is(are) considered the cause of the cervical abnormality. Cytologic interpretations of HSIL or more severe (HSIL$^+$) are more strongly associated with histologically confirmed CIN3 and cancer than less severe cytologic interpretations, and antecedent HSIL often precedes rare cases of invasive cervical cancer in the follow-up of women diagnosed with CIN2 and under surveillance (versus immediate treatment).[26,27] HSIL cytology is of sufficient clinical concern that treatment is considered acceptable even without histologic confirmation of CIN2 or more severe diagnoses (CIN2$^+$) on biopsy.[4]

Our main goal for this analysis was to assess whether biopsy diagnosed as CIN2 by morphology and tested as positive for p16 by IHC was similar enough to biopsy-diagnosed CIN3 in the distribution of these other biomarkers of cervical cancer risk such that making a distinction between the 2 would be unnecessary, that is, calling both HSIL without annotating the morphologic diagnosis of CIN2 or CIN3.

METHODS

Cervical biopsies used in the current study were part of a population-based study conducted in the state of New Mexico.[10] The biopsy with the most severe diagnosis for individual women who received diagnoses in the period of 2006–2009 was used. Of the 21 297 women receiving diagnoses in laboratories serving New Mexico's residents during the study period, a stratified sample of 6272 women was chosen to overrepresent CIN2 and CIN3 for additional characterization. This sample included 90.1% of all CIN2$^+$ diagnosed, which represented all adequate CIN2$^+$ biopsies that could be found, and random samples of 17.7% of all CIN1 and 6.3% of all negative histology biopsies diagnosed during that period.

Laboratory Testing

A "sandwich" technique was used to enable histopathologic review of tissue sections flanking the sections subjected to HPV genotyping and p16 IHC as follows: one 4-μM section was obtained for H&E staining, two 4-μM sections for HPV genotyping were collected into O-ringed microfuge tubes, a second 4-μM section was obtained for H&E staining, and then 4-μM section(s) adjacent to this second H&E was(were) obtained for biomarker staining including p16 IHC with sections collected onto Fisherbrand Superfrost Plus (ThermoFisher Scientific, Waltham, Massachusettes) glass slides.

Selection of the cases for the current study was limited to those in which 5 or more unstained slides were available to allow for potential unsatisfactory slides and the opportunity for further tests on the same subset. This resulted in a group of 4359 cases from which 4100 biopsy specimens from different women were selected randomly to create 41 sets of 100 slides, each of which was reviewed by 1 of 41 different pathologists participating in this study as volunteers (p16 IHC Study Group). Patient age and, when available, referral cytology (3563, 86.9% of cases) also were provided to the pathologists. p16 IHC was performed, as described below, on an unstained slide adjacent to the H&E-stained slide used for diagnosis from each case. Each set contained similar proportions of each diagnostic outcome.

p16 IHC.——Formalin-fixed, paraffin-embedded (FFPE) cervical tissue sections were stained by using 1 of 2 methods: manually by using the CINtec Histology Kit (Roche mtm laboratories, Heidelberg, Germany) or by using the automated BenchMark instrument platform (Ventana/Roche, Tucson, Arizona). Briefly, deparaffinization was performed by baking the slides at 65°C for 45 minutes followed by rehydration of the tissues in xylene and graded alcohol baths (95%, 70%, and 50%). Optimized epitope retrieval for archival tissues was performed at 95°C for 45 minutes in epitope retrieval solution. Epitope retrieval slides were either transferred to the BenchMark XT or manually stained. p16 IHC staining and visualization procedures followed the BenchMark p16 IHC protocol or the CINtec Histology Kit protocol specified by the manufacturer.

HPV Genotype Detection in FFPE Tissues.——Methods for HPV genotyping of the FFPE tissue sections were previously reported and are summarized here.[28] FFPE tissue sections were digested in a protein K digestion buffer at 65°C for 4 hours followed by overnight at 37°C. Before the polymerase chain reaction–based HPV genotyping, digested FFPE tissue was heated at 95°C for 15 minutes to inactivate the protein K and centrifuged briefly at $13\,000g$ to remove undigested material, and the supernatant (aqueous digest) was decanted and stored at −80°C until tested.

Two microliters and 5 μL (for 2 separate genotyping determinations) of the aqueous digest from each tissue specimen were used for genotyping with the LINEAR ARRAY HPV Genotyping Test (HPV LA, Roche Diagnostics, Indianapolis, Indiana), a qualitative HPV genotyping test for 37 HPV genotypes.[29–31] Using the Roche LA HPV detection kit, hybridizations were automated by using Tecan ProfiBlot-48 robots (Tecan, Grödig, Austria) as previously described.[32] The Roche LA HPV Genotyping Test detects 13 high- and 24

low-risk HPV types. HPV52 is not determined directly by a type-specific probe but by inference as previously described.[28,31] Two independent readers interpreted the presence of HPV genotypes by using a reference template, and any differences between the 2 readers were adjudicated by a third independent reader. The adjudicated result was taken as the final interpretation.

Pathology Reviews

EP Reviews.——Expert panel pathologists rendered an adjudicated consensus diagnosis of these biopsies.[10] The EP diagnosis review was based only on an H&E staining of a new section and masked to any other data including p16 IHC results when available.

Volunteer Pathologist Reviews.——Forty-one pathologists (p16 IHC Study Panel Group; here referred to as VPs [volunteer pathologists]) practicing throughout the United States and Canada agreed to review the H&E and p16 IHC for 100 cervical biopsies each. Recruitment of VPs was either via direct invitation for College of American Pathologists committee members or through general advertisement (eg, flyers) at professional society meetings.

VPs were given a 2-page instructions sheet developed by Ventana to guide their interpretation of p16 IHC. Diffuse p16 IHC staining was considered positive ("a continuous staining of cells of the basal and parabasal cell layers of the cervical squamous epithelium, with or without staining of the intermediate or intermediate to superficial cell layers"). p16 IHC resulting in a focal ("either a staining of isolated cells or small cell clusters; that is, a noncontinuous staining, particularly not of the basal and parabasal cells") or negative ("the p16 stained slide shows no staining reaction") staining pattern was considered negative. Only the p16 IHC interpretations of VPs were used in these analyses.

The p16 IHC interpretations of VPs were used, whereas morphologic diagnoses of the VPs, though performed for studies of p16 utilization, were not included in these analyses. Use of p16 interpretations rendered by the independent VP was preferred to reduce inherent interpretation biases when comparing community and EP diagnoses.

Analysis

The primary aim of the analysis was to examine whether the CP-diagnosed CIN2 that subsequently tested p16 IHC positive was the risk equivalent to the CP-diagnosed CIN3, which is routinely treated in clinical practice. Thus, CP diagnoses, with and without stratification by p16 IHC as read by the VP, were compared to the EP diagnoses (masked to p16 IHC results), tissue HPV genotyping, and antecedent cytologic interpretations.

Of the statewide sample of 21 297 cervical biopsies, there were 21 187 cervical biopsy tissues after excluding adenocarcinoma in situ, adenocarcinoma, and cancers other than SCC histology. A consort diagram of the biopsies included in this analysis is shown in the Figure. Sampling fractions of the statewide population for each grade of biopsy diagnosis by the CP used in these analyses are shown in Supplemental Table 1 (see supplemental digital content containing 4 table files at www.archivesofpathology.org in the June 2020 table of contents) for reference. For simplicity, these analyses did not correct for sampling fractions (of the CP

diagnoses) (except where otherwise noted), as all CIN2^{+} biopsies that could be located were collected and were assumed to be representative, that is, cases were missing at random. A random sample of CP-diagnosed negative and CIN1 histology was included from the entire state of New Mexico.

Of the 4100 samples tested by p16 IHC, 65 were excluded (from above) from this analysis because they were diagnosed by CP as glandular disease, adenocarcinoma in situ, or adenocarcinoma, which are not included in the LAST recommendations. An additional 25 were classified as "Technically unsatisfactory" by any (CP, EP, and/or VP) pathology review, thus excluded. As noted, the distribution of HPV genotypes causing SCC was included for reference under the assumption that the more closely the profile of biomarkers for a precursor diagnosis resembled SCC, the better proxy it was for cancer risk, that is, high-grade cervical abnormalities with invasive potential. After the aforementioned exclusions, the resulting sample size was 4010.

HPV genotype results were categorized hierarchically according to their a priori cancer risk[2,3,33] for simplicity of presentation and to account for detection of multiple HPV genotypes: (1) HPV16 positive; (2) HPV16 negative and HPV18 and/or HPV45 (HPV18/45) positive; (3) HPV16, 18, and 45 negative and HPV31, 33, 35, 39, 51, 52, 56, 58, 59, and/or 68 positive (other high-risk HPV) positive; (4) negative for all high-risk HPV types and HPV26, 53, 66, 67, 70, 73, and/or 82 (intermediate-risk HPV) positive; (5) high- or intermediate-risk HPV negative and HPV6, 11, 40, 42, 54, 55, 61, 62, 64, 69, 71, 72, 81, 82v, 83, 84, and/or 89 (low-risk HPV) positive; or (6) HPV negative for all measured types. A shift in a distribution to higher or lower HPV risk groups was considered to be related to a greater or lesser risk of cervical cancer, respectively.

The proportion and binomial exact 95% CI of p16 IHC–positive cases by grade of biopsy diagnosis by CP was calculated. A nonparametric test of trend[34] or trend test using weighted logistic regression model was used to test for trends in p16 IHC or HPV risk groups within or between diagnostic categories by the CP or EP. Trends in the percentage that was p16 IHC positive, HPV16 positive, and with an antecedent HSIL[+] cytology—according to The Bethesda System for cytologic classification[35]—for paired CP and EP diagnoses, were calculated. The percentage that was p16 IHC positive, HPV16 positive, with an antecedent HSIL[+] cytology, and with CIN3 or more severe (CIN3^{+}) or CIN2^{+} diagnosed by the EP, was compared between p16 IHC–negative and p16 IHC–positive CIN2 and between p16 IHC–positive CIN2 and CIN3 diagnosed by the CP, using a Fisher exact test. Finally, trends in HPV risk groups were compared for CIN2 diagnosed by the CP for all 4 combinations of cytology results (<HSIL versus HSIL[+]) and p16 IHC results (negative and positive).

P values of <.05 were considered significant. STATA (versions 13.1 and 15.1, StataCorp LLC, College Station, Texas) was used for analyses.

RESULTS

Correlations of HPV categories, p16 IHC results, and histologic diagnosis by the CP are shown in Table 1. The percentage that was p16 IHC positive increased with an increasing

severity of histologic diagnoses by the CP: 7.4% (95% CI, 4.7%–11.1%) for negatives, 26.6% (95% CI, 23.7%–29.5%) for CIN1, 71.9% (95% CI, 69.6%–74.2%) for CIN2, and 90.7% (95% CI, 88.9%–92.2%) for CIN3 ($P_{trend} < .001$); for reference, 94.5% (95% CI, 86.6%–98.5%) of the SCCs tested p16 IHC positive. The percentage that was p16 IHC positive increased with an increasing severity of biopsy diagnosis by the CP for each HPV risk group (ie, HPV16 > HPV18/45 > other high-risk HPV > intermediate-risk HPV > low-risk HPV > HPV negative) (P_{trend} .001 except for low-risk HPV where P_{trend} was < .010). The percentage that was p16 IHC positive increased with an increasing severity of biopsy diagnosis by the CP even among HPV negatives ($P_{trend} < .001$), suggestive of some false-negative HPV genotyping. The percentage that was p16 IHC positive increased with higher-risk HPV groups for each diagnosis ($P_{trend} < .001$). Testing p16 IHC positive was associated with higher HPV risk group than testing p16 IHC negative for each grade of CP-diagnosed biopsies ($P < .001$). That is, both HPV genotype and histologic diagnosis were independent determinants of testing p16 IHC positive.

We found no meaningful differences in the distribution of HPV genotypes detected or p16 IHC results, based on the study subsample, compared to those same results when extrapolated to the whole sample (Supplemental Table 2). HPV genotype–specific results stratified by the CP diagnosis and p16 IHC results are presented in Supplemental Table 3.

For all abnormal histologic findings (CIN1 or more severe), p16 IHC–positive cases had higher-risk HPV than p16 IHC–negative diagnoses. Notably, p16 IHC–positive CIN2 had lower-risk HPV than CIN3 ($P_{trend} < .001$); 38.2% (415 of 1087) of p16 IHC–positive CIN2 tested positive for HPV16, whereas 54.5% (658 of 1208) of all CIN3, regardless of the p16 IHC, tested positive for HPV16. p16 IHC–negative CIN2 had lower-risk HPV than p16 IHC–positive CIN2 ($P_{trend} < .001$) but higher-risk HPV than all CIN1 ($P_{trend} < .001$), p16 IHC–positive CIN1 ($P < .001$), or negative histology ($P_{trend} < .001$) (ie, regardless of the p16 IHC).

p16 IHC–positive CIN3 had higher-risk HPV than p16 IHC–negative CIN3 ($P_{trend} < .001$). p16 IHC–positive CIN3 was less likely to test positive for HPV18/45 ($P < .001$) than SCC, again demonstrating that HPV18/45 tends to be underrepresented in CIN3, compared to its attributable fraction in SCC.[3,36,37] p16 IHC–negative CIN3 had higher-risk HPV than p16 IHC–positive CIN2 ($P_{trend} = .02$). Similar patterns of p16 IHC and HPV risk groups were observed for the EP (Supplemental Table 4).

Table 2 shows the pairwise diagnoses by the CP and EP and the correlation with testing p16 IHC positive or HPV16 positive, or having an antecedent HSIL+ cytology. Increasing severity of the EP diagnosis for a given CP diagnosis, and vice versa, was associated with increased likelihood of the biopsy testing p16 IHC positive or HPV16 positive ($P_{trend} < .001$ for both), with exception for when SCC was diagnosed by either the CP or the EP, or negative by the CP ($P_{trend} = .090$ for p16 IHC and $P_{trend} = .40$ for HPV16). Increasing severity of the CP diagnosis for a given EP diagnosis was also increasingly likely to have an antecedent HSIL+ cytology ($P_{trend} < .001$ for negative, CIN1, CIN2, and CIN3; and $P = .05$ for SCC). Increasing severity of the EP diagnosis for a CP diagnosis of CIN3 was more likely to have an antecedent HSIL+ cytology ($P_{trend} < .001$). Surprisingly, increasing severity

of the EP diagnosis for a CP diagnosis of negative, CIN2, and cancer was not associated with having an antecedent HSIL$^+$ cytology ($P_{trend} > .05$ for all).

Table 3 compares the percentage that was HPV16 positive, had antecedent HSIL$^+$ cytology, and CIN3$^+$ and CIN2$^+$ diagnosis by the EP between CP-diagnosed CIN2, stratified on p16 IHC status, and CIN3. p16 IHC–positive CIN2 was less likely than CIN3 to test HPV16 positive (38.18% versus 54.47%, respectively; $P < .001$), have an antecedent HSIL$^+$ cytology (21.02% versus 42.53%, respectively; $P < .001$), or be diagnosed on review by the EP as CIN3$^+$ (22.91% versus 65.31%, respectively; $P < .001$) or CIN2$^+$ (64.40% versus 88.08%, respectively; $P < .001$). p16 IHC–positive CIN2 was less likely than CIN3 to be positive for at least 1 of these biomarkers (HPV16, antecedent HSIL$^+$ cytology, and/or CIN3$^+$ diagnosed by EP) (58.23% versus 85.93%, respectively; $P < .001$). p16 IHC–positive CIN2 was less likely than CIN3 to be positive for all 3 biomarkers (2.48% versus 15.89%, respectively; $P < .001$). p16 IHC–negative CIN2 was less likely than p16 IHC–positive CIN2 to be positive for any individual biomarker ($P < .001$), with exception of having antecedent HSIL$^+$ cytology ($P > .99$). p16 IHC–negative CIN2 was less likely than p16 IHC–positive CIN2 to be positive for at least 1 biomarker ($P < .001$) or all 3 biomarkers ($P < .001$).

Table 4 compares the HPV risk group distribution for CP-diagnosed CIN2 that tested p16 IHC negative or positive with antecedent less than HSIL cytology (<HSIL) or HSIL$^+$ cytology. Notably, p16 IHC–positive CIN2 with an antecedent HSIL$^+$ cytology has lower-risk HPV than all CP-diagnosed CIN3 ($P = .01$). p16 IHC–negative CIN2 with an antecedent less than HSIL cytology has higher-risk HPV than all CP-diagnosed CIN1 ($P < .001$).

DISCUSSION

In the largest case series to include HPV genotyping and p16 IHC immunostaining of biopsy specimens to date, we were able to show the detailed relationship of these biomarkers with community diagnoses of precursors to cervical cancer, with a focus on CIN2. Key observations from our analyses were as follows: (1) most CP-diagnosed CIN2 and CIN3, and a significant proportion of CIN1, tested p16 IHC positive; (2) p16 IHC–positive, CP-diagnosed CIN2 was less likely to test HPV16 positive, to have an antecedent HSIL$^+$ cytology, and to be called CIN3$^+$ or CIN2$^+$ by the EP than CP-diagnosed CIN3; (3) p16 IHC–negative CIN2 had lower-risk HPV than p16 IHC–positive CIN2 but higher-risk HPV than CIN1; and (4) p16 IHC–negative CIN3 had higher-risk HPV than CIN2 or even p16 IHC–positive CIN2.

These data also confirm that p16 IHC corrects some of the errors in diagnosing high-grade cervical abnormalities but does so imperfectly. Approximately 28% of the CP-diagnosed CIN2 tested p16 IHC negative in this study; other studies have reported the percentage of p16 IHC–negative CIN2 ranging from approximately 20%[16] to less than 10%.[21,38] Based on HPV risk group distribution, these cases were indeed lower risk and are less likely to progress to cancer. That is, p16 IHC–negative, CP-diagnosed CIN2 was more like CIN1 than CIN3. It is therefore justifiable to downgrade p16 IHC–negative, CP-diagnosed CIN2 to low-grade squamous intraepithelial lesion as recommended by LAST.[22] Conversely, p16 IHC–positive, CP-diagnosed CIN2 was more like CP-diagnosed CIN3 than CIN1.

However, these data also indicate that some fraction of the p16 IHC–positive, CP-diagnosed CIN2 is not a high-grade cervical abnormality. The implication of these data is that if LAST terminology[22] is to be used in routine practice (equivalent to CP), the use of HSIL categorization for CIN3 or p16 IHC–positive CIN2 must include annotation of the H&E (morphologic) diagnosis, for example, HSIL(CIN3) or HSIL(CIN2), respectively, which was suggested as optional by LAST. It is clear from these data that p16 IHC–positive CIN2 is NOT the clinical equivalent of CIN3. That is, a p16 IHC–positive CIN2 does not have the same clinical meaning (invasive potential) as CIN3, and therefore the 2 cannot be considered 1 clinical entity and should not be conflated with one another. When EP diagnosed the CP-diagnosed CIN2 biopsy as CIN3, the fraction that tested HPV16 and p16 IHC positive was close to that of the CP-diagnosed CIN3 biopsy, suggesting that these were high-grade cervical abnormalities. However, a consensus review by a panel of expert pathologists is not typically available in routine clinical practice.

Recent reports confirm the dramatic difference in risk of subsequent invasive cancer between CIN2[12,28] and CIN3[23] diagnoses. The disparity in HPV genotype distribution that we report provides a credible biologic explanation for this difference. Moreover, our findings emphasize that making this distinction between CIN2 and CIN3 for a HSIL diagnosis is necessary for clinical decision-making on whether to treat women with precursor lesions that might otherwise regress on their own. This is especially true in young women diagnosed with CIN2 for whom conservative management (wait and watch) is preferred[4] owing possibly to the potential of added risk of negative reproductive outcomes (eg, preterm delivery) associated with excision treatments.[13,14] In most cases, p16 IHC–positive CIN2 is likely to regress on its own, given that approximately 70% to 80% of CIN2 cases test p16 IHC positive,[16,39] as observed here, but approximately 50% of all CIN2, and approximately 60% of CIN2 diagnosed in women younger than 30 years, will regress.[12] Arithmetically, even if all p16 IHC–negative CIN2 cases were regressive, a significant proportion of p16 IHC–positive CIN2 must also be regressive. A prospective study of women diagnosed with CIN2 reported that 57% of p16 IHC–positive CIN2 regressed in 12 months.[39] Retrospective study of women diagnosed with pathology review–confirmed CIN2 followed up for 2 years found that 50% of regressive CIN2 cases were initially p16 IHC positive and 18% of p16 IHC–positive lesions regressed.[40]

In addition to causing the unnecessary treatment of some women with CIN2, losing the distinction between (p16-positive) CIN2 and CIN3 would also have long-term negative implications on the opportunity for future improvements to the diagnosis of high-grade cervical abnormalities. As new biomarkers are being developed that might better distinguish between HPV infection and high-grade cervical abnormalities and therefore might be applied to CIN2 or even p16 IHC–positive CIN2, it will be important to be able to easily identify such cases by qualifying whether they were diagnosed as CIN2 or CIN3.

These data also underscore the importance of the LAST recommendation *not* to perform p16 IHC testing systematically on all CIN3 or CIN1.[22] For CP-diagnosed CIN3, less than 10% of community diagnoses of CIN3 tested p16 IHC negative. CP-diagnosed CIN3 that tested p16 IHC negative had a less risky HPV group distribution (P_{trend} < .001) but were similarly likely to have antecedent high-grade cytology (25.5% versus 24.8%, P = .89) as

CP-diagnosed CIN2 that was diagnosed as CIN3 by the EP. Thus, p16 IHC–negative CIN3 is unlikely to be at sufficiently low risk to change its management, that is, there is no clinical utility, only added cost.

Nor is there evidence that p16 IHC testing of CIN1 provides clinically meaningful risk stratification or predicts progression to CIN2+ as previously shown.[41–43] Here, based on HPV risk group distribution, p16 IHC–positive CIN1 was higher risk than p16 IHC–negative CIN1 but not even as high risk as p16 IHC–negative CIN2. Moreover, p16 IHC–positive CIN1 was similarly unlikely to have an antecedent high-grade cytology as p16 IHC–negative CIN1 (3.2% versus 5.1%, respectively; P = .27) (data not shown).

Aside from the added cost for limited or no benefit to women diagnosed with CIN1, p16 IHC testing of biopsies diagnosed as CIN1 might result in incorrect, overinterpretation of a positive p16 IHC result as CIN2, which could then lead to unnecessary treatment and a concomitant increased risk of preterm delivery for those still considering childbearing.[13,14,42]

Limited p16 IHC testing, and/or possibly Ki-67 IHC testing, of some CIN1 might have some value for internal use as a laboratory quality control standard,[7,44] similar to the use of HPV to squamous intraepithelial lesion ratios for cytology,[45] to set the threshold of normal versus nonnormal histology. We observed that approximately one-quarter of the CP-diagnosed CIN1 tested p16 IHC positive in this study. Other studies have reported a percentage of p16 IHC–positive cases for CIN1 ranging from approximately 10% to almost 60%,[16,21,42,46–50] suggesting significant variability/unreliability in the morphologic interpretation of diagnosis criteria for CIN1 (versus negative or CIN2) compared to a more objective standard, that is, p16 IHC.

Likewise, some pathologists equivocate between CIN1 and CIN2 or diagnosis "CIN1/2." We did not separately analyze cases of CIN1/2, as there were small numbers in our dataset. However, the LAST recommendation for using p16 IHC was to clarify the clinical meaning of CIN2 by distinguishing those CIN2 that were higher risk (p16 IHC–positive CIN2) from those that were lower risk (p16 IHC–negative CIN2), not to clarify the meaning of CIN1. Here again, there would be potential for overutilization of p16 IHC. Pathologists uncertain whether a biopsy diagnosis is CIN1 or CIN2 might be tempted to call it CIN1/2 or even CIN2 for perceived greater safety, believing that using p16 IHC as an adjunctive test would correct any overcalls. However, because such a high percentage of CIN1 (and therefore also "CIN1/2") will also test p16 IHC positive, most of which is nothing more than low-grade, benign, regressive CIN1, these biopsies might then get misclassified as HSIL and women would receive unnecessary treatment.

A few other scenarios might exacerbate the inappropriate use of p16 IHC on CIN1 biopsies. First, pathologists, worried that future review of a CIN1 biopsy by a second pathologist might result in a CIN2 diagnosis, might be motivated to do p16 IHC on a CIN1 or a CIN1/2 that previously they would report as CIN1. In addition, p16 IHC on CIN1 or CIN1/2 may be used by pathologists as feedback to lower their criteria for a CIN2 diagnosis, resulting in more CIN1 being called CIN2.

We acknowledge an important limitation: this analysis was cross-sectional and therefore we could only make inferences related to true cervical cancer risk based on biomarker distributions. Nevertheless, these biomarkers included in this analysis are strong predictors of cervical cancer risk and if practical, could be incorporated into improved diagnostic classification of cervical abnormalities. Indeed, women whose Papanicolaou specimen was called high-grade cytology and tested HPV16 positive (the readout for which is provided by some HPV tests versus setting up laboratory testing of biopsy specimens) are at very high risk of CIN3, up to ~80%.[51,52]

Another potential limitation is that the EP diagnoses were based on H&E alone and were not informed by p16 IHC results. However, it is unknown whether p16 IHC–informed interpretations of H&E diagnoses would have resulted in improved classification of CIN2 versus H&E diagnosis rendered independently of p16 IHC results.

Future studies in cohorts with long-term follow-up will be needed to determine the risk stratification provided by p16 IHC testing of CIN2. These cohorts will need to be rather large because of the rarity of CIN2 (<1%) in the general population and losses to follow-up. Alternatively, retrospective analyses of conservatively managed CIN2 in younger women could be done in which the index CIN2 biopsies are tested by p16 IHC. Such studies would provide important information on its risk stratification, level of safety (versus invasive cancer) for women with p16 IHC–negative CIN2, and how much overtreatment is likely to occur if p16 IHC–positive CIN2 were to be treated immediately.

It is clear from these and other data that p16 IHC is a sensitive but nonspecific biomarker of CIN3, which is a more rigorous definition of "cervical precancer" than CIN2 and even p16 IHC–positive CIN2. Even so, many but not all CIN3 will develop into invasive cervical cancer if left untreated.[23] However, because of the lack of specificity of p16 IHC, presumably owing to its increased expression in response to productive HPV infections that may or may not progress,[53,54] many low-grade cervical abnormalities will still test p16 IHC positive even if they are not destined to progress to high-grade cervical abnormalities. LAST classification of "HSIL" diagnosis, which includes p16 IHC–positive CIN2, should annotate the morphologic diagnosis of CIN2 or CIN3 in routine clinical practice to inform all clinical management decisions. This is especially important for (but not limited to) those women younger than 30 years and/or considering childbearing who receive a diagnosis of CIN2 and for whom surveillance rather than treatment is recommended[4] and/or desirable, respectively. While neither "biomarker"—CIN diagnosis or p16 IHC—is perfect, together they further stratify the cervical cancer risk and if used correctly, better inform clinical-decision making than either can accomplish alone.

In summary, there is currently no reliable way to distinguish those CIN2 and even p16 IHC–positive CIN2 diagnoses that will progress or regress. The use of other biomarkers with or without p16 IHC may improve the diagnostic classification of cervical abnormalities in relation to their invasive cancer potential.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

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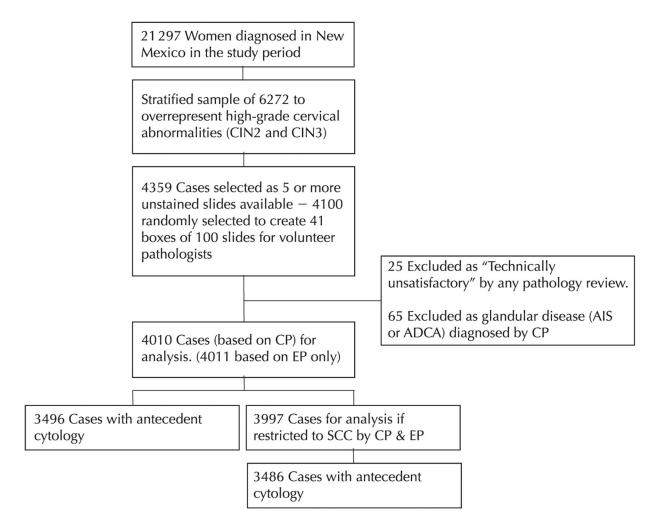
Figure.
Consort diagram of specimen inclusions and exclusions. Abbreviations: ADCA, adenocarcinoma; AIS, adenocarcinoma in situ; CIN, cervical intraepithelial neoplasia; CIN2, CIN grade 2; CIN3, CIN grade 3; CP, community pathologists; EP, expert pathologists; SCC, squamous cell carcinoma.

Table 1.

Human Papillomavirus (HPV) Genotyping Results, Categorized According to Cervical Cancer Risk, and p16 Immunohistochemistry (IHC) Results (p16 IHC Positive [p16+] or Negative [p16−]) for Biopsy Diagnosis by the Community Pathologist of Negative, Cervical Intraepithelial Neoplasia (CIN) 1, CIN2, and CIN3.

HPV Categories[a]	Negative				CIN1				CIN2				CIN3					SCC	
	p16 IHC Negative		p16 IHC Positive		p16 IHC Negative		p16 IHC Positive		p16 IHC Negative		p16 IHC Positive		p16 IHC Negative		p16 IHC Positive		P_{trend}[b]	All	
	N	% Col	N	% Col	N	% Col	N	% Col	N	% Col	N	% Col	N	% Col	N	% Col		N	% Col
HPV16	8	3.1	1	4.8	32	4.7	35	14.1	91	21.4	415	38.2	41	36.3	617	56.3	<.001	43	58.9
HPV18/45	5	1.9	2	9.5	24	3.5	25	10.1	29	6.8	82	7.5	8	7.1	57	5.2	<.001	10	13.7
Other high risk[c]	21	8.0	7	33.3	142	20.7	128	51.6	163	38.4	496	45.6	36	31.9	354	32.3	<.001	10	13.7
Intermediate risk[d]	9	3.4	2	9.5	54	7.9	28	11.3	29	6.8	56	5.2	3	2.7	31	2.8	<.001	2	2.7
Low risk[e]	4	1.5	0	0.0	35	5.1	4	1.6	18	4.2	7	0.6	4	3.5	3	0.3	.0097	1	1.4
HPV negative	215	82.1	9	42.9	399	58.2	28	11.3	95	22.4	31	2.9	21	18.6	33	3.0	<.001	7	9.6
Total	262	100	21	100	686	100	248	100	425	100	1087	100	113	100	1095	100	<.001	73	100
P_{trend}[f]	<.001				<.001				<.001				<.001						

Abbreviations: NA, not applicable; SCC, squamous cell carcinoma.

Results for all SCCs are shown for reference. "% Col" is the column percentage, that is, the number in cell divided by the total column number.

[a] Defined hierarchically according to cancer risk.

[b] Test of trend for testing p16 IHC positivity across diagnoses (excluding cancer) for each HPV risk group.

[c] HPV31, 33, 35, 39, 51, 52, 56, 58, 59, and 68.

[d] HPV26, 53, 66, 67, 70, 73, and 82.

[e] HPV6, 11, 40, 42, 54, 55, 61, 62, 64, 69, 71, 72, 81, 82v, 83, 84, and 89.

[f] Test of trend for HPV risk group by p16 IHC result for each diagnosis.

Table 2.

Pairwise Diagnoses by the Community Pathologist (CP) and Expert Panel (EP) and the Percentage p16 Immunohistochemistry (IHC)–Positive (%p16 IHC+), Human Papillomavirus Type 16 (HPV16)–Positive (HPV16+), and With an Antecedent High-Grade Squamous Intraepithelial Lesion (HSIL) or More Severe (HSIL+) Cytologic Interpretation

| | EP Diagnosis | | | | | | | | |
	Negative	CIN1	CIN2	CIN3	SCC	Total	P_{trend} (p16)	P_{trend} (HPV16)	P_{trend} (HSIL+ Cytology)
CP diagnosis									
Negative									
N	261	14	7	1	0	**283**	.009	.40	.15
%cell[a]	6.53	0.35	0.18	0.03	0.00	7.08			
%p16 IHC+	6.51	7.14	42.86	0.00	0.00	7.42			
%HPV16+	3.07	0.00	14.29	0.00	0.00	3.18			
%HSIL+ cytology[b]	7.58	33.33	14.29	0.00	0.00	8.84			
CIN1									
N	535	318	67	11	0	**931**	<.001	<.001	.029
%cell	13.39	7.96	1.68	0.28	0.00	23.29			
%p16 IHC+	10.47	42.77	68.66	81.82	0.00	26.53			
%HPV16+	3.74	11.32	16.42	0.00	0.00	7.20			
%HSIL+ cytology[c]	5.93	3.25	1.56	0.00	0.00	4.62			
CIN2									
N	260	431	547	271	0	**1509**	<.001	<.001	.43
%cell	6.50	10.78	13.69	6.78	0.00	37.75			
%p16 IHC+	34.23	68.68	82.45	91.88	0.00	71.90			
%HPV16+	18.46	24.59	40.04	47.97	0.00	33.33			
%HSIL+ cytology[d]	23.45	17.66	20.80	24.80	0.00	21.08			
CIN3									
N	90	48	275	787	2	**1202**	<.001	<.001	<.001
%cell	2.25	1.20	6.88	19.69	0.05	33.07			
%p16 IHC+	42.22	70.83	89.45	97.71	100.00	90.60			

	EP Diagnosis								
	Negative	CIN1	CIN2	CIN3	SCC	Total	P_{trend} (p16)	P_{trend} (HPV16)	P_{trend} (HSIL+ Cytology)
%HPV16+	33.33	25.00	49.45	60.36	50.00	54.41			
%HSIL+ cytology [e]	27.50	31.82	34.60	47.74	0.00	42.50			
SCC							.010	.36	.17
N	1	0	2	17	52	72			
%cell	0.03	0.00	0.05	0.43	1.30	1.80			
%p16 IHC+	0.00	0.00	100.00	100.00	96.15	95.83			
%HPV16+	0.00	0.00	50.00	58.82	59.62	58.33			
%HSIL+ cytology [f]	0.00	0.00	100.00	75.0	82.76	79.55			
Total							<.001	<.001	<.001
N	1147	811	898	1087	54	3997			
%cell	28.70	20.29	22.47	27.20	1.35	10.00			
%p16 IHC+	17.44	57.58	83.30	96.04	96.30	62.82			
%HPV16+	9.24	18.99	40.98	56.58	59.26	31.90			
%HSIL+ cytology [f]	12.08	13.15	23.46	41.61	80.00	23.61			
P_{trend} (p16)	<.001	<.001	<.001	<.001	.78	<.001			
P_{trend} (HPV16)	<.001	<.001	<.001	<.001	.79	<.001			
P_{trend} (HSIL+ cytology)	<.001	<.001	<.001	<.001	.046	<.001			

Abbreviations: CIN, cervical intraepithelial neoplasia; NA, not applicable; SCC, squamous cell carcinoma.

Thirteen cases excluded as diagnosed as adenocarcinoma in situ or adenocarcinoma by EP (3 CP CIN1, 3 CP CIN2, 6 CP CIN3, and 1 CP SCC).

[a] Cell percentage, that is, N_{cell}/N_{total}.

[b] Sixty-eight missing cytology (63 EP negative, 5 EP CIN1).

[c] One hundred eight missing cytology (63 EP negative, 41 EP CIN1, 3 EP CIN2, 1 EP CIN3).

[d] One hundred fifty-two missing cytology (34 EP negative, 46 EP CIN1, 47 EP CIN2, 25 EP CIN3).

[e] One hundred fifty-five missing cytology (10 EP negative, 4 EP CIN1, 38 EP CIN2, 102 EP CIN3, 1 EP SCC).

[f] Twenty-eight missing cytology (5 EP CIN3, 23 EP SCC).

Table 3.

The Relationships of Community Pathology–Diagnosed Cervical Intraepithelial Neoplasia (CIN) 3 and CIN2, Stratified on p16 Immunohistochemistry (IHC) Result, With Biomarkers of Cervical Cancer Risk: the Biopsy Testing Positive for Human Papillomavirus Type 16 (HPV16), an Antecedent High-Grade Intraepithelial Lesion (HSIL) or More Severe (HSIL[+]) Cytologic Interpretation, and an Expert Panel (EP) Review Histopathologic Diagnosis of CIN Grade 3 (CIN3) or More Severe (CIN3^{+}) or CIN Grade 2 (CIN2) or More Severe (CIN2^{+})

Biomarker Result	p16 IHC–Negative CIN2		p16 IHC–Positive CIN2		CIN3		P^a	P_{trend}^b
	N	%	N	%	N	%		
HPV16[+]	91	21.41	415	38.18	658	54.47	<.001	<.001
HSIL[+] cytology[c]	80	21.11	206	21.02	447	42.53	<.001	<.001
EP diagnosis of CIN3^{+}	22	5.18	249	22.91	789	65.31	<.001	<.001
EP diagnosis of CIN2^{+}	118	27.76	700	64.40	1064	88.08	<.001	<.001
HPV16[+], HSIL[+], and/or EP diagnosis of CIN3^{+}	163	38.35	633	58.23	1038	85.93	<.001	<.001
HPV16[+], HSIL[+], and EP diagnosis of CIN3^{+}	3	0.71	27	2.48	192	15.89	<.001	<.001

Below the individual biomarker results, the relationships of the diagnoses with combinations of any (or) or all (and) biomarkers are shown.

[a] p16 IHC–positive CIN2 versus CIN3.

[b] Trend for p16 IHC–negative CIN2 versus p16 IHC–positive CIN2 versus CIN3.

[c] Forty-six p16 IHC–negative CIN2, 107 p16 IHC–positive CIN2, and 157 CIN3 missing antecedent cytology.

Table 4.

The Relationships of Community Pathology (CP)–Diagnosed Cervical Intraepithelial Neoplasia Grade 2 (CIN2), Stratified by p16 Immunohistochemistry (IHC) Results and Antecedent Cytologic Interpretation Categorized as High-Grade Squamous Intraepithelial Lesion (HSIL) or More Severe (HSIL[+]) Versus Not (<HSIL), With Human Papillomavirus (HPV) Categories and Compared to CP-Diagnosed CIN3

	CP-Diagnosed CIN2							
	p16 IHC Negative				p16 IHC Positive			
	<HSIL Cytology		HSIL[+] Cytology		<HSIL Cytology		HSIL[+] Cytology	
HPV Risk Group [a]	N	% Col	N	% Col	N	% Col	N	% Col
HPV16	61	20.4	18	22.5	284	36.7	88	42.7
HPV18/45	16	5.4	9	11.3	54	7.0	17	8.3
Other high risk [b]	127	42.5	28	35.0	360	46.5	87	42.2
Intermediate risk [c]	21	7.0	5	6.3	46	5.9	8	3.9
Low risk [d]	15	5.0	1	1.3	6	0.8	0	0.0
HPV negative	59	19.7	19	23.8	24	3.1	6	2.9
P_{trend} (versus CIN3[e])		<.001		<.001		<.001		.012

One hundred fifty-three cases were missing antecedent cytology results. "% Col" is the column percentage, that is, the number in cell divided by the total column number.

[a] Defined hierarchically according to cancer risk.

[b] HPV31, 33, 35, 39, 51, 52, 56, 58, 59, and 68.

[c] HPV26, 53, 66, 67, 70, 73, and 82.

[d] HPV6, 11, 40, 42, 54, 55, 61, 62, 64, 69, 71, 72, 81, 82v, 83, 84, and 89.

[e] Compared to data combining p16 IHC–negative and p16 IHC–positive CIN3 from Table 1.