



Effect of adenoid hypertrophy on the upper airway and craniomaxillofacial region

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Background: In recent years, annual incidences of adenoid hypertrophy (AH), a highly common tissue lesion in children, have increased. Currently, research on AH has focused on its obstruction of nasal cavity function, and little has been written on its influence on the upper airway's bone structure. For this reason, our present study seeks to determine the influence of AH on both the morphological development characteristics of the upper airway and the craniofacial features in children, with the goal being to offer more choices for diagnosing and treating the condition in the future.

Methods: From June 2019 to December 2020 in Department of Orthodontics, Beijing Stomatological Hospital, Capital Medical University, 38 children with AH admitted to the Department of Otolaryngology [research group (RG)] and 35 children [control group (CG)] who underwent orthodontic treatment over the same time span were selected as the research objects. X-ray examination of the lateral position of the head, observation of the maxillofacial structure, and detection of the children's height, growth factors, and sleep status, and analysis of the differences between the two groups.

Results: The height of RG, insulin-like growth factor-1 (IGF-1) as well as insulin-like growth factor binding protein-3 (IGFBP-3) were all lower than CG ($P < 0.05$), the upper airway became narrower, and the malocclusion was aggravated ($P < 0.05$). Cephalometric measurement showed that the angle between the subspinale and sella at nasion (SNA angle) and the angle between the subspinale and supraemental at nasion (ANB angle) of RG children decreased, and the angle between the supraemental and sella at nasion (SNB angle) increased ($P < 0.05$). In addition, the sleep quality of RG was significantly lower than that of CG ($P < 0.05$).

Conclusions: AH can change a child's breathing mode and function by giving rise to upper airway stenosis, and by inducing deformities of their craniomaxillofacial region and oral cavity, thus disrupting their normal growth and development.

Keywords: Adenoid hypertrophy (AH); upper airway; craniomaxillofacial region; growth and development

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Introduction

Adenoid hypertrophy (AH) is a highly common tissue lesion in children and is mainly related to nasopharyngitis (1). According to statistics, worldwide there are over 600,000 new cases of AH each year, and the trend is increasing annually (2). AH mainly causes a series of uncomfortable and inflammatory changes in the ear, nose, and throat. However, it can also lead to deformities in the development of the craniomaxillofacial region as it forces patients to engage in long-term open mouth breathing (3). In addition, AH also greatly impacts eating, so for younger patients it may also compromise their intake of nutrients and thus disrupt their growth and development (4). At this stage, the examination and diagnosis of AH is still complicated, and nasal endoscope, nasopharynx palpation, and X-ray are all required to confirm AH (5). In clinical practice, resection is recommended for the treatment of AH as it is the most effective treatment (6). However, for children whose body function is not fully developed, an operation will bring about serious post-operative traumatic stress and require a long rehabilitation cycle (7). Therefore, a deeper understanding of the pathogenic mechanism and influence of AH is key to finding a new and effective diagnosis and treatment plan for the condition.

Thus far, research on AH has focused on its obstruction of nasal cavity function (8,9), while little has been written on its influence on the upper airway's bone structure. As we all know, adenoid facies is the most common complication of AH, and is caused by AH hindering the function of the nasal respiratory tract (10). Moreover, an earlier study has found that tracheitis and respiratory tract infection are also potentially bound up with the structural changes of the upper airway in children (11). Understanding how AH alters the upper airway may be the key to finding a new diagnosis and treatment plan for AH. For this reason, our study sought to determine the influence of AH on both the morphological development characteristics of the upper airway and the craniofacial features in children, with the goal being to offer more choices for diagnosing and treating the condition in the future. We present the following article in accordance with the STROBE reporting checklist (available at <https://dx.doi.org/10.21037/tp-21-437>).

Methods

Research participants

In total, 73 children who were admitted to the Department

of Orthodontics, Beijing Stomatological Hospital, Capital Medical University between June 2019 and December 2020 were enrolled in the study. Thirty-eight of these children had AH and were included in our research group (RG), while 35 underwent orthodontic treatment and were included in the control group (CG). The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). This experiment was carried out with permission from the Ethics Committee of Beijing Stomatological Hospital, Capital Medical University, and informed consent forms were signed by the guardians of all enrolled children.

Inclusion and exclusion criteria

Children were included in the RG if their AH was confirmed by examination in Department of Orthodontics, Eye Hospital of Hebei, were between 4 and 12 years old, snored or engaged in open mouth breathing, and their detailed case data were available. Children were excluded if they had a history of chronic rhinitis, turbinate hypertrophy, temporomandibular joint, or craniofacial trauma; received ear, nose, or throat surgery, or craniomaxillofacial orthodontic treatment; and if they had congenital developmental disorders. The inclusion and exclusion criteria were the same for the CG, the only difference being children enrolled in the CG were not confirmed to have AH.

Study methods

Height measurement

During admission to hospital, we measured the height of each child in the two groups using an automatic height detector.

Growth factor detection

A sample of fasting venous blood (2 mL) was taken from each child and placed in a coagulation-promoting tube, followed by 10 min centrifugation (1,505 ×g, 4 °C) to obtain serum after 30 min still standing at indoor temperature, and IGF1 and IGFBP-3 in the serum were quantified using the electrochemical luminescence method.

X-ray examination

A lateral cranial image was taken from each child at a voltage of 86 kV, current of 16 mA, and exposure time of 16 s (*Figure 1*). In the image, the following items were then measured: the vertical length of the tangent line from the lowest end of the lower part of the adenoid to the cranial lateral side of

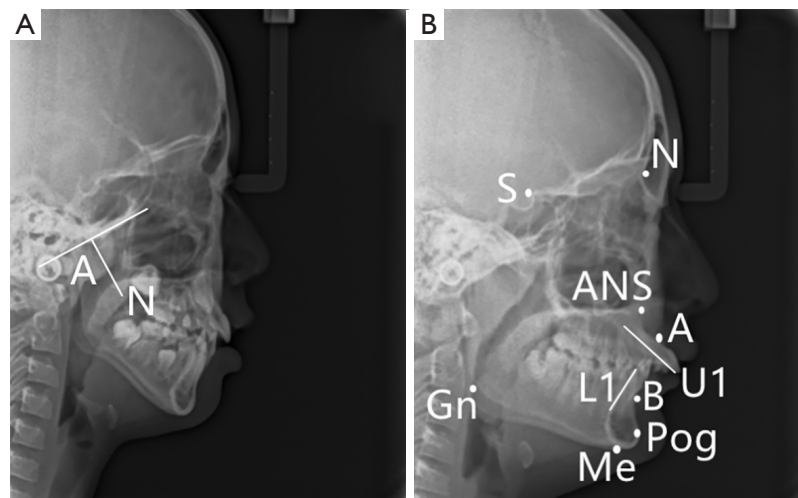


Figure 1 X-ray results. (A) Adenoid index. A: thickness of adenoid; N: nasopharynx cavity width. (B) Lateral cranial markings. N: the most concave point of the bone between the anterior nasal spine and the edge of the upper alveolar; S: the most concave point of the bone between the anterior nasal spine and the edge of the upper alveolar; A: the most concave point of the bone between the anterior nasal spine and the edge of the upper alveolar; B: the most concave point of the bone between the anterior nasal spine and the edge of the upper alveolar; Me: the lowest point of the chin; Gn: the midpoint between the pre-chin point and the sub-chin point; L1: the midpoint between the pre-mental point and the submental point; U1: the midpoint between the pre-mental point and the submental point; Pog: the midpoint between the pre-chin point and the sub-chin point; ANS: the tip of the anterior nasal spine is used as one of the two landmark points to determine the palatal plane.

the basilar clivus (A), nasopharynx cavity width (N), distance between the nasopharynx apex (PNS) and R (point located at the intersection of the posterior pharyngeal wall and PNS-Hor line) (PNS-R), distance between the PNS and UPW (point located at the intersection of the posterior pharyngeal wall and PNS-Ba line) (PNS-UPW), distance between the SPP (point of intersection of the line from the soft palate center perpendicular to the posterior pharyngeal wall and posterior margin of the soft palate) and SPPW (point of intersection of the line from the soft palate center perpendicular to the posterior pharyngeal wall and posterior pharyngeal wall) (SPP-SPPW), distance between U (tip of soft palate) and MPW (foot point at the posterior pharyngeal wall of perpendicular line from point U) (U-MPW), the angle between the subspinale and sella at nasion (representing the position of the maxilla in relation to the cranium) (SNA angle), the angle between the supraemental and sella at nasion (representing the position of the mandible in relation to the cranium) (SNB angle), and the angle between the subspinale and supraemental at nasion (representing the relationship of maxilla and mandible in relation to the cranium) (ANB). In addition, Dolphin software was used to analyze the angles between the plane of the anterior skull base and the facial

plane (NPog-SN), the former and the mandibular plane (GoGn-SN), the upper central incisor long axis and the NA plane (U1-NA), the upper central incisor long axis and NB plane (L1-NB), and the upper central incisor long axis and mandibular plane (IMPA).

Electronic nasopharyngoscope examination

A Japanese Olympus ENF-VT2 electronic nasopharyngoscope was adopted for examination. The child was let to keep a sitting position, with the lower jaw slightly lift and the hands, feet and head positions all fixed, and then required to receive spray anesthesia for bilateral nasal cavity twice, at an interval of 3–5 min. The nasopharyngoscope was inserted into the nasal cavity through nostril to check nasal mucosa, and then the nasopharyngoscope was let to reach the nasopharyngeal cavity when nasal septum, chronic rhinitis, abnormal shape of turbinate can be ruled out. The child was instructed to breathe hard, followed by observation of the volume of adenoids in nostril. Synchronous images were collected, and transmitted to computer synchronously.

Sleep quality evaluation

The Pittsburgh Sleep Quality Index (PSQI) was used

Table 1 Baseline data

Baseline data	RG	CG	t or χ^2	P
Age	7.53±2.14	7.54±2.38	0.019	0.985
Gender			0.275	0.600
Boy/girl	24/14	20/15		
Childbirth			0.308	0.579
Natural childbirth/caesarean section	22/16	18/17		
Only child			0.145	0.704
Yes/no	29/9	28/7		
Family history of illness			0.015	0.902
Yes/no	4/34	4/34		
Sleep performance			0.085	0.959
Snoring/open mouth breathing/both	7/5/26	6/4/25		
Living environment			0.226	0.635
Town/rural	29/9	25/10		
Nationality			0.893	0.345
Han/minority	35/3	34/1		

RG, research group; CG, control group.

to evaluate the sleep quality, efficiency, and time of the children in the two groups (12). The scale comprised self-assessment items and other assessment items which include seven modules, namely, sleep quality, time to fall asleep, sleep time, sleep efficiency, sleep disorder, hypnotic drugs, and daytime dysfunction.

With a total score of 0–21 points, the scale indicates sleep disorder by a score >7 points, and a higher score indicates more serious sleep disorder.

Outcome measures

The outcomes of the two groups were determined using the height and growth factors [insulin-like growth factor-1 (IGF-1) and insulin-like growth factor binding protein-3 (IGFBP-3)], the measurement results of the sagittal diameter of the upper airway (A, N, PNS-P, PNS-UPW, SPP-SPPW, and U-MPW), the relative indexes of malocclusion (NPog-SN, GoGn-SN, U1-NA, L1-NB, and IMPA), the hard tissue cephalometric indexes (SNA angle, SNB angle, and ANB angle), and sleep quality.

Statistical analysis

All data were processed using SPSS (version 25.0) (IBM

Corp., Armonk, NY, USA). Counting data (n/%) were processed using a chi-square test. Measurement data ($\bar{x}\pm s$) in normal distribution were analyzed by the independent-samples *t*-test, and those not in normal distribution by the Whitney U test. $P<0.05$ denotes a remarkable difference.

Results

Clinical baseline data

The clinical baseline data (e.g., age and gender) was no notably different between the two groups ($P>0.05$). Under the electronic nasopharyngoscope, with the adenoid size occupying 50% of the posterior nostril as the threshold, there were 23 patients (60.5%) with the adenoid occupying $\geq 50\%$ of the posterior nostril and 15 patients (39.5% with the adenoid occupying $<50\%$ of the posterior nostril in the RG (Table 1).

Comparison of height and growth factors

According to the results, the height of the RG was lower than that of the CG ($P<0.05$), and the levels of IGF-1 (186.88 ± 22.86 ng/mL) and IGFBP-3 (5.42 ± 0.89 μ g/mL) in

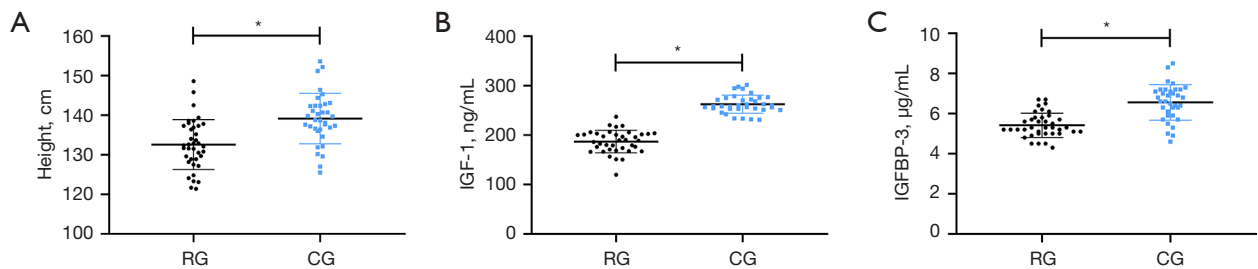


Figure 2 Comparison of height and growth factors. (A) Comparison of height. (B) Comparison of IGF-1. (C) Comparison of IGFBP-3. *, $P < 0.05$. IGF-1, insulin-like growth factor-1; IGFBP-3, insulin-like growth factor binding protein-3; RG, research group; CG, control group.

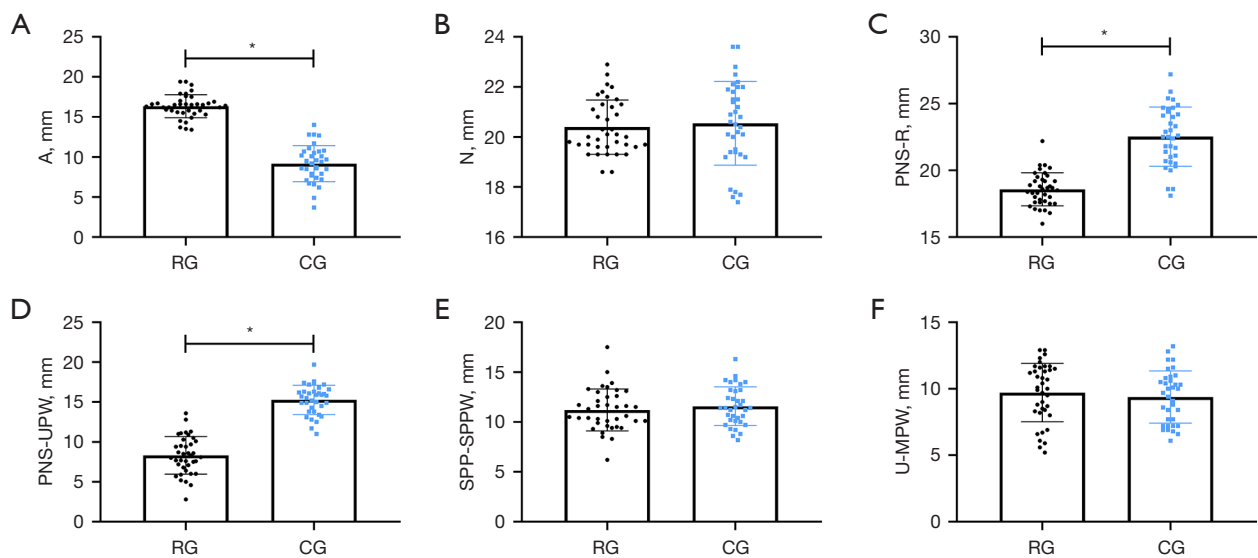


Figure 3 Measurement results of the sagittal diameter of the upper airway. (A) Comparison of A. (B) Comparison of N. (C) Comparison of PNS-R. (D) Comparison of PNS-UPW. (E) Comparison of SPP-SPPW. (F) Comparison of U-MPW. *, $P < 0.05$. A: the vertical length of the tangent line from the lowest end of the lower part of the adenoid to the cranial lateral side of the basilar clivus; N: nasopharynx cavity width; R: point located at the intersection of the posterior pharyngeal wall and PNS-Hor line; PNS: nasopharynx apex; PNS-R: distance between the PNS and R; UPW: point located at the intersection of the posterior pharyngeal wall and PNS-Ba line; PNS-UPW: distance between the PNS and UPW. RG, research group; CG, control group.

the RG were also lower than those in the CG (both $P < 0.05$) (Figure 2).

Comparison of the sagittal diameter of the upper airway measurement results

According to the measurement results, the two groups did not greatly differ in N, SPP-SPPW, and U-MPW (all $P > 0.05$), but the RG showed higher A and lower PNS-R and PNS-UPW than the CG (both $P < 0.05$) (Figure 3).

Comparison of relative indexes of malocclusion

We found no notable difference between the two groups in NPog-SN, GoGn-SN, and IMPA (all $P > 0.05$), but U1-NA and L1-NB in the RG were both higher than those in the CG ($P < 0.05$) (Figure 4).

Hard tissue cephalometric indexes

The angles of SNA and ANB in the RG were both lower

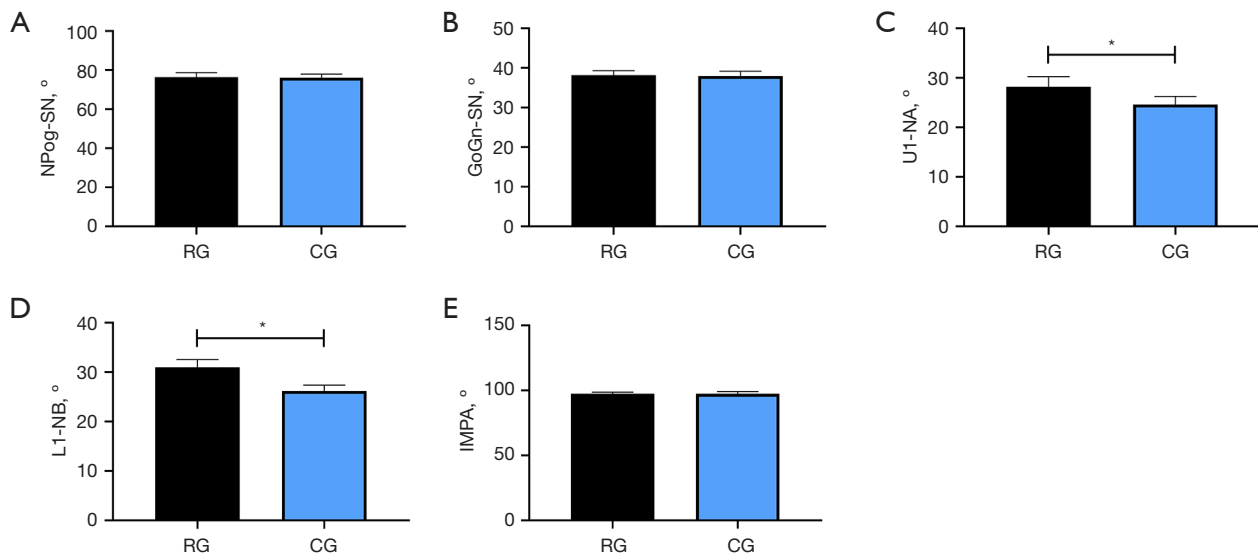


Figure 4 Comparison of the relative indexes of malocclusion. (A) Comparison of NPog-SN. (B) Comparison of GoGn-SN. (C) Comparison of U1-NA. (D) Comparison of L1-NB. (E) Comparison of IMPA. *, $P < 0.05$. NPog-SN: the angles between the plane of the anterior skull base and the facial plane; GoGn-SN: the former and the mandibular plane; U1-NA: the upper central incisor long axis and the NA plane; L1-NB: the upper central incisor long axis and NB plane; IMPA: the upper central incisor long axis and mandibular plane. RG, research group; CG, control group.

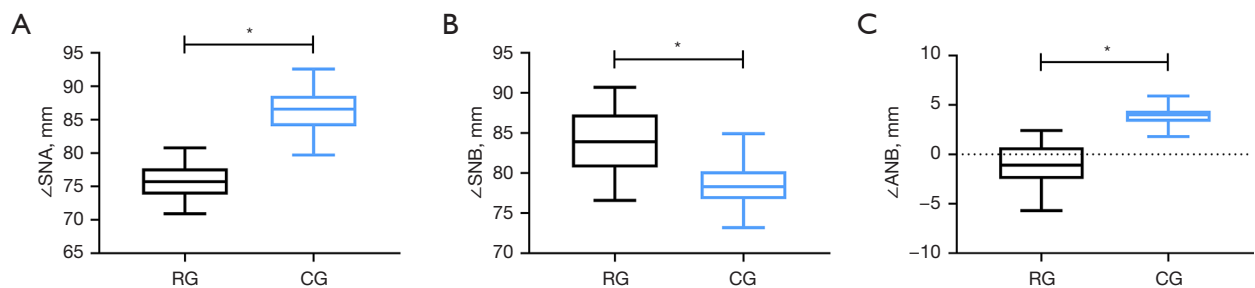


Figure 5 Comparison of hard tissue cephalometric indexes. (A) Comparison of SNA. (B) Comparison of SNB. (C) Comparison of ANB. * $P < 0.05$. SNA: the angle between the subspinale and sella at nasion (indicating the site of the maxilla in relation to the cranium); ANB: the angle between the subspinale and supraaemental at nasion (indicating the association of maxilla and mandible in relation to the cranium); SNB: the angle between the supraaemental and sella at nasion (indicating the site of the mandible in relation to the cranium). RG, research group; CG, control group.

than those in the CG ($P < 0.05$), while the angle of SNB in the former was higher than that in the latter ($P < 0.05$) (Figure 5).

Comparison of sleep quality

Following our sleep quality evaluation, we found that the time to fall asleep (23.16 ± 4.05 min) and the PSQI score (14.00 ± 4.25 points) of the RG were higher than those of the

CG (all $P < 0.05$) (Figure 6).

Discussion

The adenoid is an organ of lymphoid tissue located at the top of the nasopharynx and posterior pharyngeal wall (13). Adenoids are in exuberant growth in children between 2 and 6 years old, and they gradually start to shrink in children after the age of 10 years (14). Therefore, AH usually occurs in

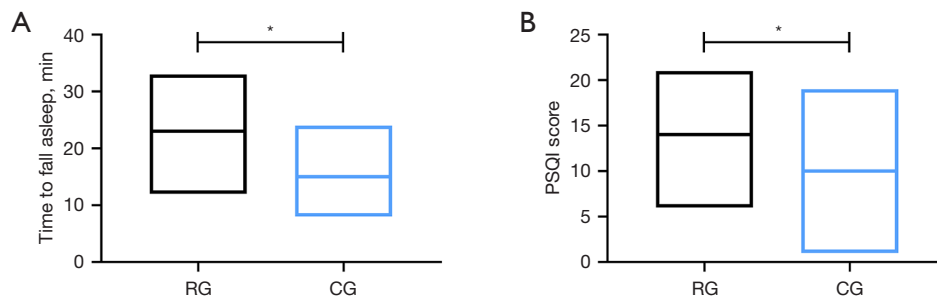


Figure 6 Comparison of sleep quality. (A) Comparison of time to fall asleep. (B) Comparison of PSQI score. *, $P < 0.05$. PSQI, Pittsburgh Sleep Quality Index; RG, research group; CG, control group.

children aged 2 to 6 years (15). The condition can give rise to nasal congestion and open mouth breathing, especially during sleep, and can even result in sleep apnea. For this reason, it is considered a particularly serious condition for children as they are in the most important stage of growth and development (16). Current clinical research on AH focuses on symptoms of oppression and obstruction. For instance, one study by Lou *et al.* (17) found a close relationship between AH and allergic rhinitis in children, while another study by Gomaa *et al.* revealed that AH can give rise to thickening of the laryngeal mucosa (18). There are also studies on AH-induced changes of the upper airway (19), but they are highly limited, and have not evaluated and analyzed the upper airway structure of children in detail. In response to increasingly high incidences of AH, understanding its pathological mechanism and potential influence as soon as possible is the key to preventing and treating AH in the future. This study determined to evaluate the impact of AH on the upper airway and craniomaxillofacial region, and thus provide a reliable reference for future clinical and follow-up research.

In this study, we first measured the height of the two groups and determined their IGF-1 and IGFBP-3. As a result, we found lower IGF-1 and IGFBP-3 in the RG than those in the CG, indicating that children with AH have a notably slower growth and development. As we know, a child's growth and development is regulated and influenced by many factors, with hormones released by the hypothalamus playing one of the most crucial parts (20). IGF-1 is one of the active factors secreted by hypothalamus, which has the function of transmitting growth hormones and promoting growth (21). IGFBP-3 is the specific binding protein of IGF-1, which can promote the bone activation of IGF-1 and accelerate bone development (22). Moreover, one study has revealed that a child's ability to secrete growth

hormones during nighttime sleep is much stronger than that during daytime (23). This also speaks to the inhibition of growth and development in the RG, who demonstrated that children with AH slept poorly at night, and that as a result their body could not enter a deep-sleep state to stimulate the hypothalamus and accelerate the secretion of the growth hormone. This result has also been verified in our PSQI-based investigation results about the sleep quality of the two groups, and the results of previous research are consistent with our study (24,25). To understand the influence of AH on the upper airway, we evaluated the upper airway and craniomaxillofacial region of the two groups by X-ray. The results revealed an increase in A and decreases in PNS-P and PNS-UPW in the RG, which suggested that the upper respiratory tract of children with AH obviously narrowed as a result of AH, and that this was also the main reason for children to breathe with their mouths open during sleep. We speculated that the narrowing of the upper airway had changed the respiratory function and mode of the children, resulting in a certain degree of changes in the oral muscles, as well as damage in the balance between the oral muscles and the jaws, which eventually gave rise to abnormal growth of the jaws and teeth. In addition, because of the stenosis of the upper airway, the air pressure during breathing may cause the soft tissue of the upper airway of children to collapse, causing excessive swelling of the soft palate, tongue root, and pharynx, before finally forming an obstruction. As everyone knows, the wrong way of breathing exerts a heavily negative impact on the growth and physical health of maxillofacial region. According to one study (26), the growth of maxillofacial region is a response to functional needs, and local and regional factors heavily impact maxillofacial growth and development, while the growth of cartilage and bone is a compensatory response to functional matrix (26).

Breathing pattern determines the shape of maxillofacial, craniofacial and facial muscles to a large extent, and its change can bring about the change of jaw posture. When the original balance of perioral muscle strength is broken, after the adjustment of neuromuscular, teeth and jaw gradually adapt to the wrong balance state, resulting in other abnormal movements and growth (27). In addition, long-term mouth breathing promotes the head and neck of the child to gradually extend forward, resulting in an increase in the anteversion angle of the craniocervical angle (28). An anteversion angle greater than 5° is also one of the major reasons for deformities to develop in the craniomaxillofacial region, which is also a major reason for correcting the head posture and angle of patients in clinical orthodontic treatment (29). According to the comparison results of craniomaxillofacial growth between the two groups in our study, the RG showed larger angles of U1-NA, L1-NB, and SNB, and smaller angles of SNA and ANB, which verified our above speculation. From this, we can draw a preliminary conclusion that AH causes craniomaxillofacial deformity by causing upper airway stenosis. In addition, tonsil hypertrophy, as another common tissue lesion in children, is different from AH in that tonsil hypertrophy does not change the facial structure of children, and its clinical manifestations are mainly tinnitus, deafness, nasal congestion, runny nose, breathing with open mouth, coughing and spitting, and dysphagia (30). However, AH may give rise to inflammatory lesions of tonsil, which will trigger the development of tonsil hypertrophy. Therefore, more attention should be paid to the treatment of AH patients in clinic, which requires not only timely correction and intervention of maxillofacial deformities caused by AH, but also attention to the occurrence of tonsil lesions.

It is also important to mention the limitations of this study, including the fact only 73 patients in total were enrolled. This means the statistical calculation of some test results may not be representative. In addition, the baseline characteristics of the participants were relatively similar, so we were unable to see how ethnic and growth environment differences affect the test results. For instance, children living in high altitude areas such as Tibet and Yunnan province have adapted to the respiratory compensation under hypoxia since childhood, so their respiratory changes triggered by AH are probably not as significant as those in other areas. Moreover, the comparison is based on the examination results of the two groups at their admission, so it is impossible to evaluate the differences in various indicators of patients after treatment. In addition, this study

is based on the analysis results of X-ray. Cone-beam CT is an examination method extensively adopted in maxillofacial examination over the recent years, which can provide sub-millimeter resolution images by three-dimensional image reconstruction based on three-dimensional plan, and can deliver a higher resolution than X-ray. However, the cone beam CT has not been extensively adopted in Department of Orthodontics, Beijing Stomatological Hospital, Capital Medical University. We will supplement the correlation of AH with cone beam CT examination results as soon as possible. The above are the directions worth improving in subsequent research. In the future, we will conduct a longer follow-up investigation on the participants of this study, and a more comprehensive experimental analysis on the impact of AH on multiple respiratory organs such as the upper airway, throat, and nasal cavity to obtain more accurate results for clinical reference.

Conclusions

AH can change the breathing mode and function of children by giving rise to upper airway stenosis. From this, deformities of the craniomaxillofacial region and oral cavity can develop, disrupting the normal growth and development of a child.

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Footnote

Reporting Checklist: The authors have completed the STROBE reporting checklist. Available at <https://dx.doi.org/10.21037/tp-21-437>

Data Sharing Statement: Available at <https://dx.doi.org/10.21037/tp-21-437>

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at <https://dx.doi.org/10.21037/tp-21-437>). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The study was

conducted in accordance with the Declaration of Helsinki (as revised in 2013). This experiment was carried out with permission from the Ethics Committee of Beijing Stomatological Hospital, Capital Medical University, and informed consent forms were signed by the guardians of all enrolled children.

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