

Review Article

Silica/Protein and Silica/Polysaccharide Interactions and Their Contributions to the Functional Properties of Derived Hybrid Wound Dressing Hydrogels

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Multifunctional and biocompatible hydrogels are on the focus of wound healing treatments. Protein and polysaccharides silica hybrids are interesting wound dressing alternatives. The objective of this review is to answer questions such as why silica for wound dressings reinforcement? What are the roles and contributions of silane precursors and silica on the functional properties of hydrogel wound dressings? The effects of tailoring the porous, morphological, and chemical characteristics of synthetic silicas on the bioactivity of hybrid wound dressings hydrogels are explored in the first part of the review. This is followed by a commented review of the mechanisms of silica/protein and silica/polysaccharide interactions and their impact on the barrier, scaffold, and delivery matrix functions of the derived hydrogels. Such information has important consequences for wound healing and paves the way to multidisciplinary researches on the production, processing, and biomedical application of this kind of hybrid materials.

1. Introduction: Key Definitions and Analysis Techniques Related to Silica and Hydrogels

Silica is a silicon oxide inorganic network, bearing a low population of silanol groups on the surface. This material has a zero-point charge \sim pH 2.2–3.0. The geminal and single silanol groups have pKa around 5.6 and 10.5, respectively [1]. Silica can be synthesized by bottom-up approximations from common precursors such as alkoxysilanes (tetraethylortosilane, TEOS; tetramethylortosilane, TMOS), organosilanes also known as organoalkoxysilanes or alkylalkoxysilanes (3-glycidoxypropyltrimethoxysilane, GPTMS; 3-amino-propyltriethoxysilane, APTES), and/or sodium silicate [2]. The morphology, porosity, specific surface area, and chemical composition can be modulated by selecting the synthesis conditions and postsynthesis modifications [3]. Some usual characterization techniques are summarized in Figure 1.

Silica is usually present in the form of films, colloidal suspensions, and powders of particles and fibers. The sodium silicate and Ludox[®] are commercial examples of colloidal suspensions, which have between 30 and 50 wt% SiO₂, nonporous nanoparticles (above 25 nm), and Na⁺ or NH₄⁺ counterions in a basic aqueous medium. The charge of the deprotonated silanol groups in this medium is responsible for the electrostatic colloidal stability [4]. The temperature increasing and/or pH lowering favor the interparticle silanol condensation, leading to particle aggregation/growing. These groups behave as primary alcohols (-Si-OH), and their condensation results in -Si-O-Si- and -Si-O-R siloxane networks (analogous to organic esters, where *M* can be a metal or carbon). This reaction is responsible for the silica roles as binder and promoter of adhesion [5]. Moreover, the same reactions are also involved in the silica postfunctionalization with organic groups by silanization or silylation reactions with organoalkoxysilanes and chlorosilanes.

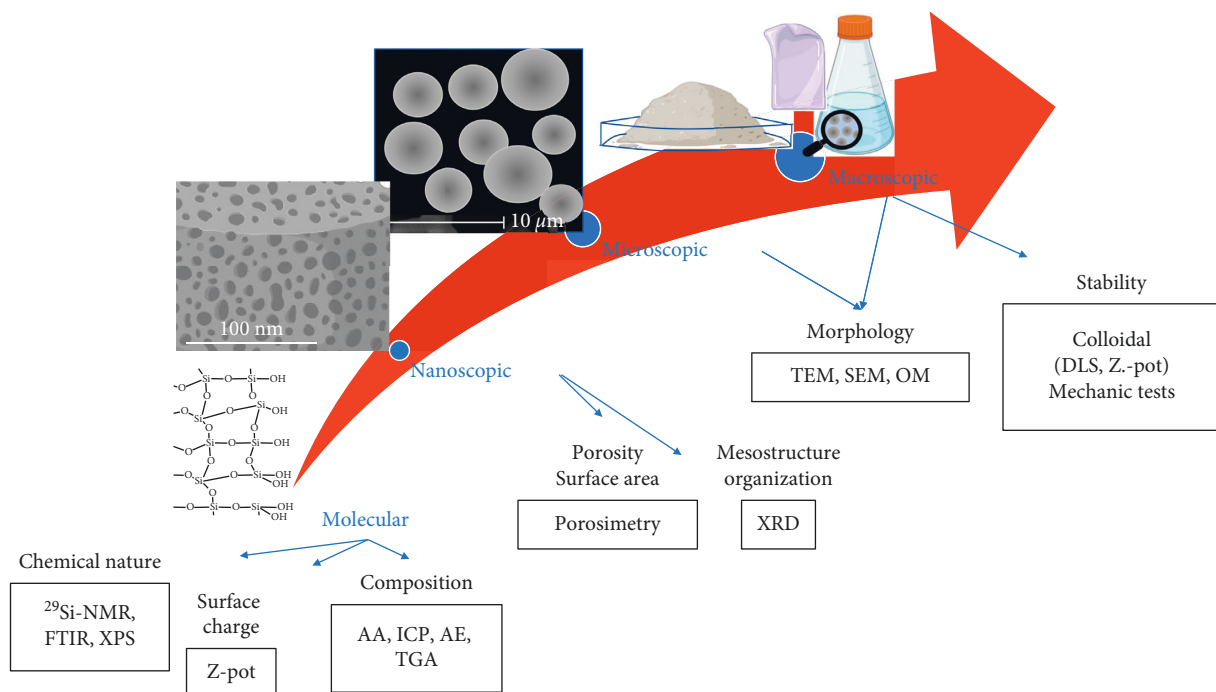


FIGURE 1: Silica properties and common characterization techniques at different levels.

Frequently, the silica particles are added as reinforcers to polymers for favoring the hardness, microbial, and attrition resistance of the resultant composites or hybrid materials. This has motivated their use in hydrogels for production of wound dressings, as reported for similar natural clays and other related metal oxides [6–10]. The impact of these properties will be exemplified on the subsequent sections of this review. Moreover, the silica has osteogenic and angiogenic effects that are interesting for tissue engineering [11, 12].

Especially emphasis has been put on the production and properties characterization of hydrogel wound dressings derived from proteins (fibrin, collagen, elastin, gelatin, silk fibroin, and keratin) [13, 14] and polysaccharides (cellulose, chitosan, chitin, pullulan, starch, and β -glucan) [7, 15]. These kinds of hydrogels could mimic tissues and could provide an adequate environment for tissue regeneration, due to their physicochemical and biological characteristics. These hydrogels can adsorb high quantities of water and can interact with extracellular matrix moieties to drive cells–hydrogels interactions through the presence of hydrophilic groups (e.g., carboxyl, hydroxyl, ether, and amino groups).

The microstructure of both types of hydrogels relies on the physical and/or chemical crosslinking [16], which in turn affect their physicochemical properties [17]. The wound dressing functions are also directly affected by the crosslinking degree. Physicochemical properties related to the crosslinking such as the mechanical stability of these hydrogels can determine their use so that they can be properly manipulated [18]. Their morphology can make these hydrogels efficient as cell culture supports and favor growth, proliferation, and even differentiation of cells that

allow tissue regeneration [19, 20]. Other structural properties such as swelling ratio, glass transition, and protein denaturation could determine the efficient nutrient transport and cellular interactions (adhesion) inside these hydrogels, which make them good candidates for wound dressings. A quick guide for crosslinking related properties characterization is given in Table 1.

Then, the question that addresses this review is how silica properties and its physicochemical interactions with proteins or polysaccharides affect the mechanical, chemical, and biological properties of the derived wound dressing hybrid hydrogels. The objective is to explain the contributions of tailoring the porous, morphological, and chemical characteristics of synthetic silicas on the bioactivity of hybrid wound dressings. This is followed by a critical review of the mechanisms of silica/protein and silica/polysaccharide interactions and their impact on the barrier, scaffold, and delivery matrix functions of the derived hydrogels, which are important for wound healing.

The novelty, originality, and relevance of the review rely on the following aspects.

- (i) This review explains how to tailor silica properties to enhance bioactivity in wound healing processes and encourage the use of siliceous materials in wound dressings.
- (ii) This is the first review that focuses on the mechanisms of silica/protein and silica/polysaccharide interactions involved in the physical entrapped or covalent bonding of preformed silicas or their silane precursors onto hydrogels and explains their functional properties as wound dressings

TABLE 1: Protein and polysaccharide derivates hydrogels crosslinking related properties characterization.

Hydrogels source	Measurement method	Crosslinking related property	References
Polysaccharide: alginate	Compressive test	Stiffness (elastic modulus)	[20]
Polysaccharide: alginate Protein/polysaccharides collagen/tyramine hyaluronan	Gravimetric analysis	Degradation	[18, 19]
Polysaccharide: alginate	DNA content Metabolic assay test	Cell proliferation	[17–21]
Proteins and polysaccharides: alginate, collagens, fibrin, hyaluronic acid, keratin, and laminin	Atomic force microscopy AFM	Stiffness (elastic modulus) Microstructure	[18, 21, 22]
Protein/polysaccharides collagen/tyramine hyaluronan Fibrin/alginate	Rheological measurements	Storage G' and loss G'' moduli	[19, 22]
Polysaccharide: inulin Protein/polysaccharides collagen/tyramine hyaluronan	Differential scanning calorimetry DSC	Glass transition Protein denaturation	[19, 23]
Polysaccharides: alginate; inulin Protein: fibrin Protein/polysaccharides collagen/tyramine hyaluronan	Scanning electron microscopy (SEM)	Porous diameter; morphology	[18, 19, 23]
Polysaccharides: inulin Protein/polysaccharides collagen/tyramine hyaluronan	Gravimetric analysis	Swelling ratio	[19, 23]

- (iii) This review widens the basic knowledge for designing hybrid wound dressings based on silica, protein, and polysaccharides hydrogels, starting from the tailoring of silica chemistry and modulating the molecular interactions during the synthesis and gelation processes, respectively.
- (iv) This review rationalizes the impact of silane and silica on the functional properties of protein and polysaccharides derived hydrogel from a chemical point of view.

2. Relationships between Silica Properties and Wound Healing Bioactivity

Nowadays, research focuses on the production of wound dressings that promote cellular migration/adhesion/proliferation, hemostasis, and tissue regeneration in conjunction with the passive task (barrier) of the gauzes. Recent reviews on wound healing highlight the participation of natural clays, minerals, and related materials in different stages of this biological process [10] and their use for the preparation of multifunctional bioactive inorganic/organic nanocomposites by different methods [7, 9]. Specifically, silicas synthesized in the laboratory become versatile materials because their chemical, porous, and morphological properties can be tailored according to the application, as it has been reported for multiple biomedical areas [3].

The next examples show that silica *per se* contributes to the bioactivity of wound dressings, emphasizing how their physicochemical properties affect the biological responses on different *in vivo* and *in vitro* experiments (Figure 2).

2.1. Specific Surface Area, Porosity, and Morphology. The surfactant-templated sol-gel or miniemulsion methodologies can be adapted for producing colloidal silicas with different particle (30–75 nm) and pore (2.7–6.5 nm) sizes.

They were stronger polymer gel adhesive than nonporous silica (LUDOX TM-50). The larger nanoparticles and higher pore diameters increased the polymer contact due to their higher total outer surface area (513.61 m²/g). The silanol groups can be interacting with the polymers or cells mainly through H-bonds and electrostatic forces, increasing the contact area between these surfaces. Moreover, the silica condensation under dehydration conditions or pH changes promotes the formation a silica network that eventually could also help to join the two surfaces, as it has been shown in other materials [24]. Therefore, the silica nanoparticles act as glue for biological tissues [25]. This helps to explain their adhesive role for wound closure when present in chitosan-derived hydrogels [5]. The silica nanoparticles also favor the collagen deposition without the risk of accumulation. The later is due to slow degradation in a physiological medium [26]. The high porosity of the silica is also responsible for the enhancement of the adhesion and proliferation of fibroblast cells in the drug-loaded chitosan–silica/poly(ϵ -caprolactone) wound dressing [27]. The porosity contributed to the adsorption and sustained release of keratinocyte growth factor from a hybrid chitosan-silica dressing, accelerating wound healing [28]. An increase of the surface area, due to the silica nanospheres on chitosan materials, was responsible for improved bleeding homeostasis. Silica promoted blood adsorption and platelet aggregation [29, 30].

2.2. Surface Charge. Undersaturated suspensions of silica (below 2 mM Si), prepared from sodium silicate, with primary particles ~ 10 nm normally exhibit negative charge at basic pHs. The negative character can be increased by the presence of L-(+)-lysine in the reaction mixture or be changed for being positive by postfunctionalization with APTES. Their use in an *in vitro* study showed that the positively charged nanoparticles stimulated the human skin fibroblasts (CCD-25SK) proliferation/migration and wound

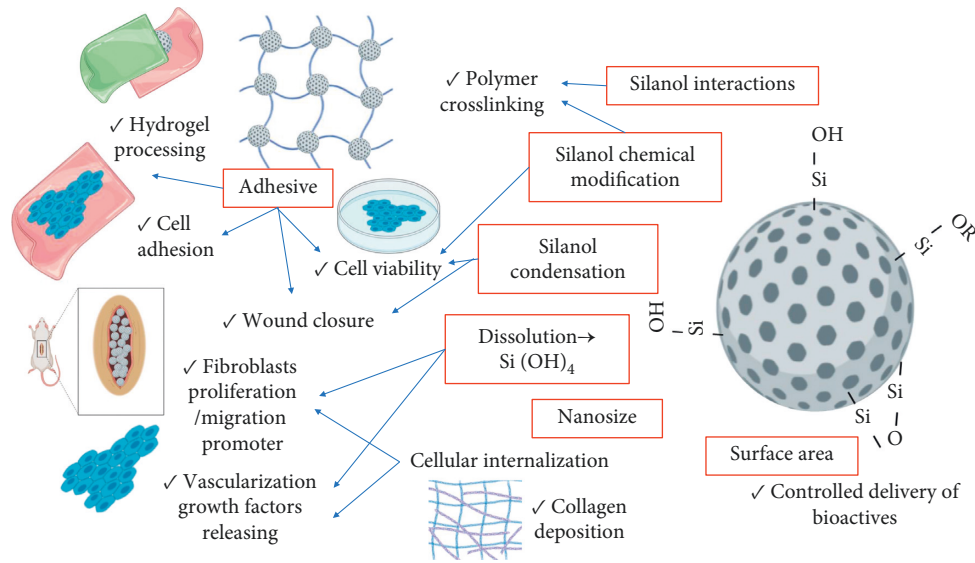


FIGURE 2: Silica properties (red squares) and biological responses on wound healing and wound dressings production.

closure. This is because of the easy cellular internalization and controlled intracellular dissolution, releasing the beneficial silicic acid and maintaining the cell viability [31]. The silicic acid ($1 \mu\text{g/mL Si}^{+4}$, extracted from a commercial bioglass) is not cytotoxic for human umbilical venous endothelial cells (CCK-8 assay) and promotes the release of vascularization growth factors (HIF-1 α , VEGF) [32].

2.3. Chemical Modifications. A high concentration of silanol groups would make the silica surface reactive towards the blood cells and can promote the reactive oxygen species formation. Therefore, the control of the chemical nature of the surface is important for avoiding hemolysis, especially if they are going to be used on wound dressings when the dermis is compromised. The silanol condensation by thermal treatments and the coating of nanoparticles with organic molecules are some strategies for avoiding this effect [33]. For example, the covalent attachment of chitosan through the isocyanatopropyltriethoxysilane linker and subsequent grafting of hydrocinnamic acid mediated by N-(3-dimethylaminopropyl)-N'-ethylcarbodiimide hydrochloride (EDC) reactant avoid the concentration-dependent cytotoxicity of silica nanoparticles. They improve also the clotting kinetics on whole blood and in vivo hemostasis because the chitosan promotes the amine-blood cells by electrostatic interactions and the catechol quinones can be bound covalently to carbohydrates and proteins (thiol, amine, and imidazole residues) of extracellular matrix, creating a physical barrier by accumulation [34]. The silica particles can be also marked with fluorophores for having smart wound dressings able to sensing bacterial infections [35] and follow the in vivo permeability and cellular uptake [30, 36, 37].

In addition to the bioactive effects of silicas, the inorganic nature, tailored surface area, and loading capacity favor their application as polymers' reinforcing agent and drug/cell carrier [3, 38, 39]. These aspects have also positive

impacts on the mechanical properties (for the processing, handling, etc.) and biological behavior of the hydrogels, as will be explained in the following section.

3. Mechanisms of Silica/Protein and Silica/Polysaccharide Interactions and Their Impacts on the Functional Properties of the Derived Hydrogels

The silica and its sources can be incorporated by physical entrapment or covalent bonding onto hydrogels (Figures 3–5). This is independent if the gels are prepared by three-dimensional polymerization of monomers, crosslinking of water-soluble polymers [16, 40], or self-assembly of proteins mediated by different kinds of molecular interactions and external stimulus [41, 42]. The next discussion will be focused on the interaction mechanisms and multiple contributions of silica to the bioactivity and passive action (barrier and selective permeability) of polysaccharides- and protein-derived hydrogels, including injectable and molded wound dressings and scaffolds.

3.1. Physical Entrapment. The silica-hydrogels composites mediated by physical interactions can be prepared by mixing the protein or gel precursor with the (i) silica source or (ii) preformed silica nanoparticles (Figure 3).

3.1.1. Gelation in the Presence of the Silica Source. An example of route (i) is the foam membrane prepared by lyophilization of TMOS and chitosan colloidal dispersions, giving a composite foam [43]. The TMOS silanol groups ($\text{pK}_a > 5.6$) interact with the amine ($\text{pK}_a = 6.3\text{--}6.5$) and hydroxyl groups of this polysaccharide through electrostatic and Van der Waals/hydrogen bonds interactions during the gel formation (Figure 3(a)), as demonstrated for similar systems at neutral pH [44]. These interactions can reduce the

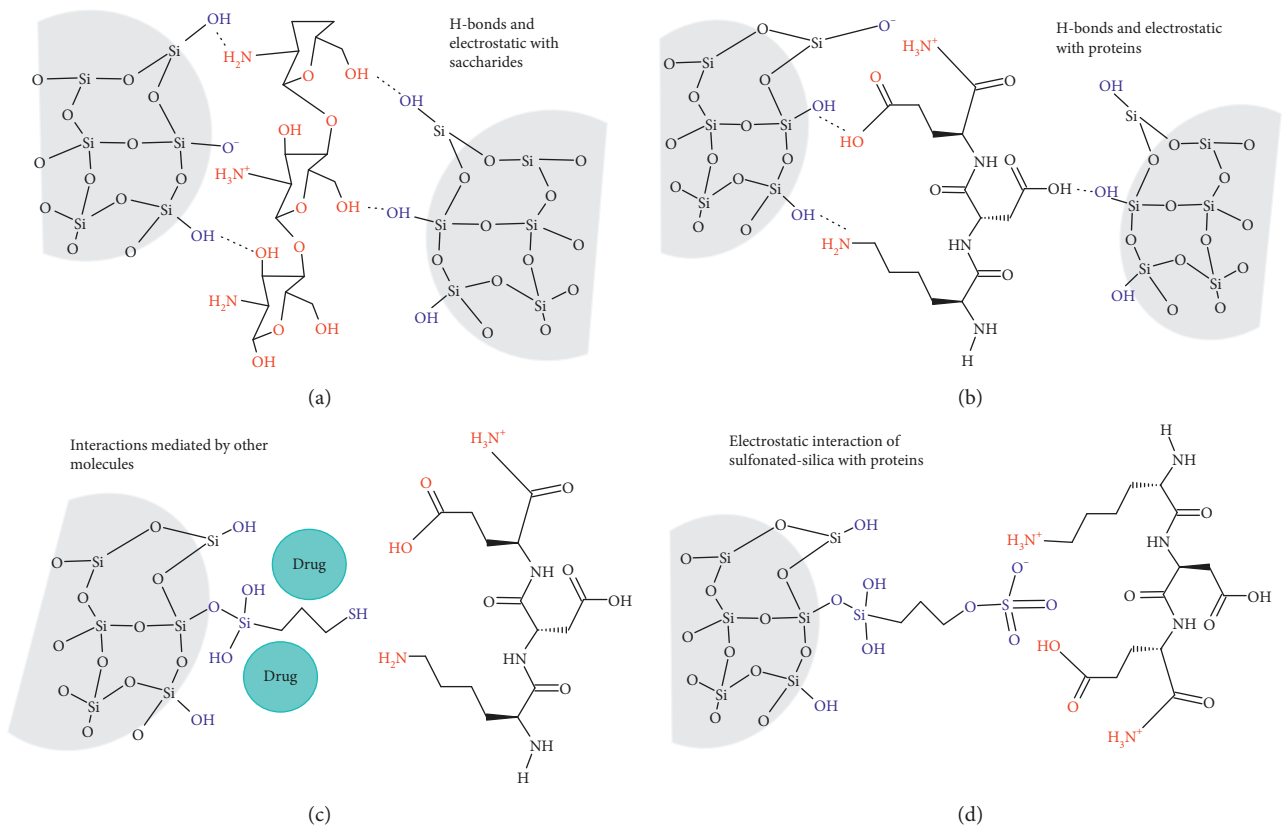


FIGURE 3: Physical interactions of pure (a,b) and silylated silicas (c,d) with polysaccharides and proteins by direct or mediated mechanisms.

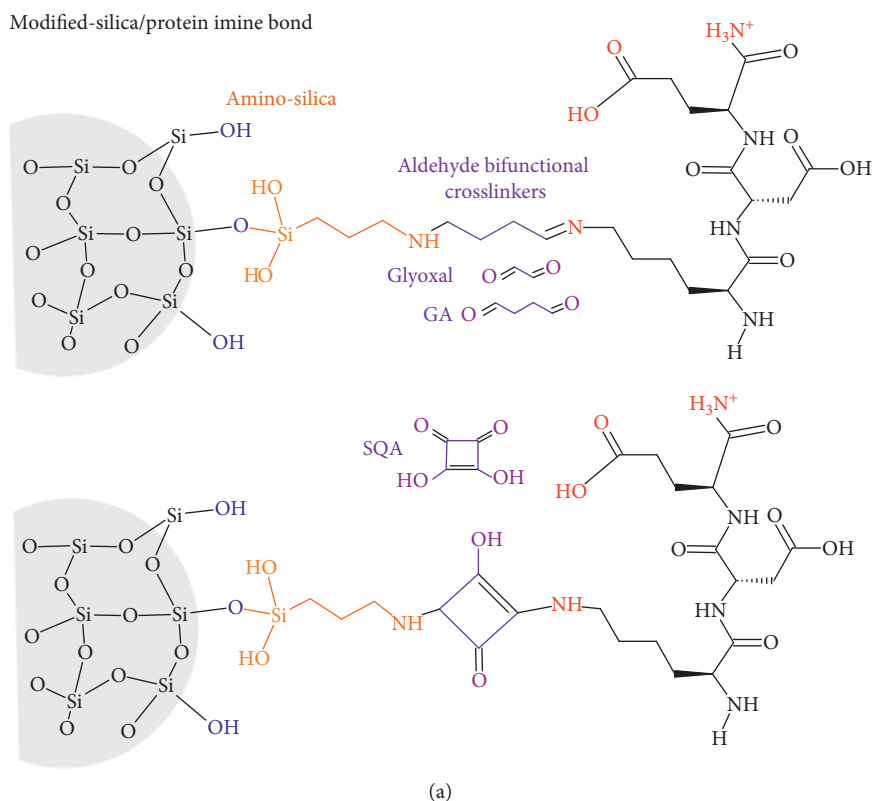
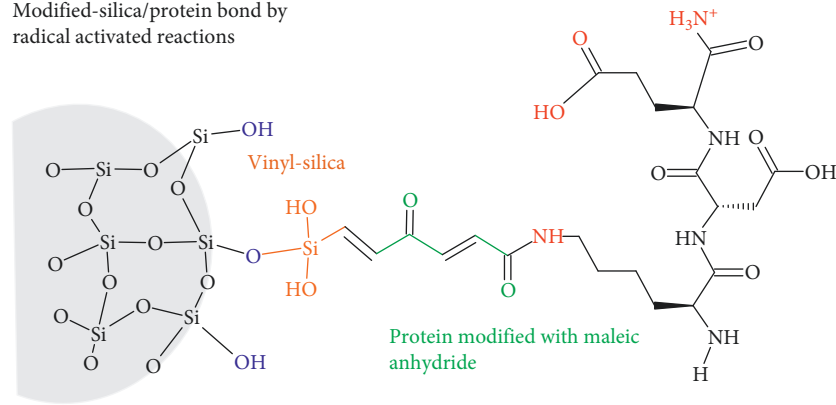


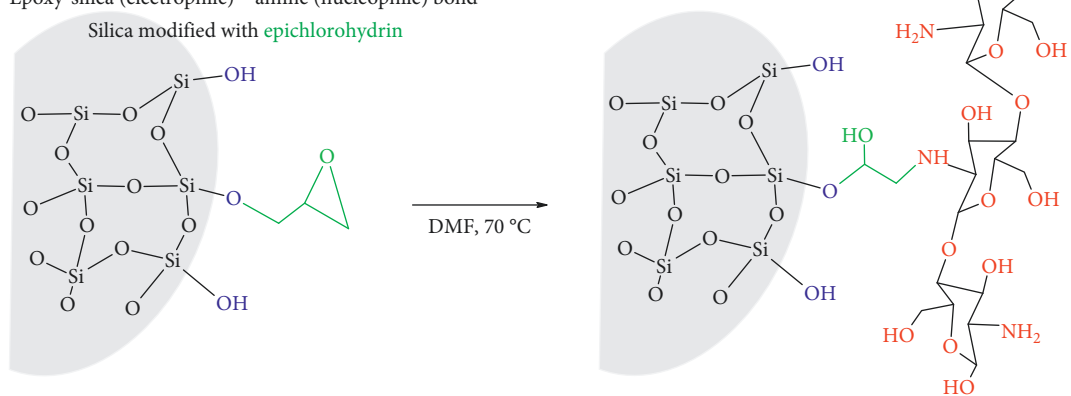
FIGURE 4: Continued.

Modified-silica/protein bond by radical activated reactions



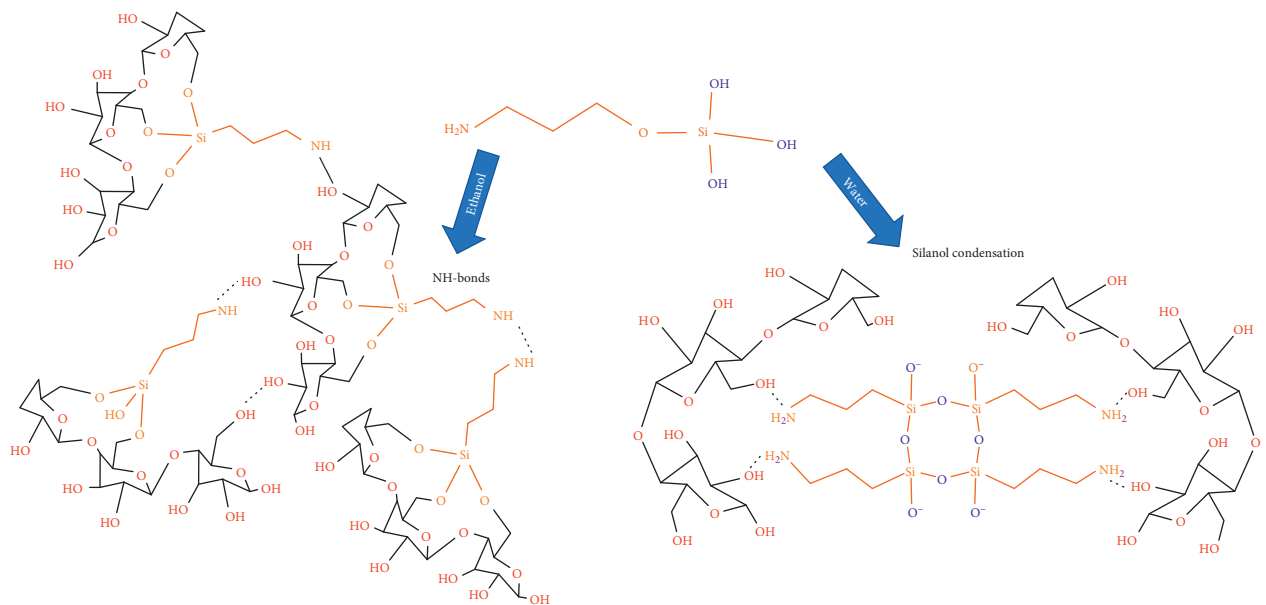
(b)

Epoxy-silica (electrophile) – amine (nucleophile) bond
Silica modified with epichlorohydrin



(c)

FIGURE 4: Chemical modification of silica and their contributions to their covalent bonding to proteins through (a) aminosilica-protein imine bonds mediated by bifunctional aldehyde crosslinkers (glyoxal; GA: glutaraldehyde; SQA: squarid acid); (b) radical activated reactions between vinyl-silica and maleic anhydride protein; (c) polysaccharide coupling via the epoxy opening in silica modified with epichlorohydrin at 70°C in dimethylformamide (DMF).



(a)

FIGURE 5: Continued.

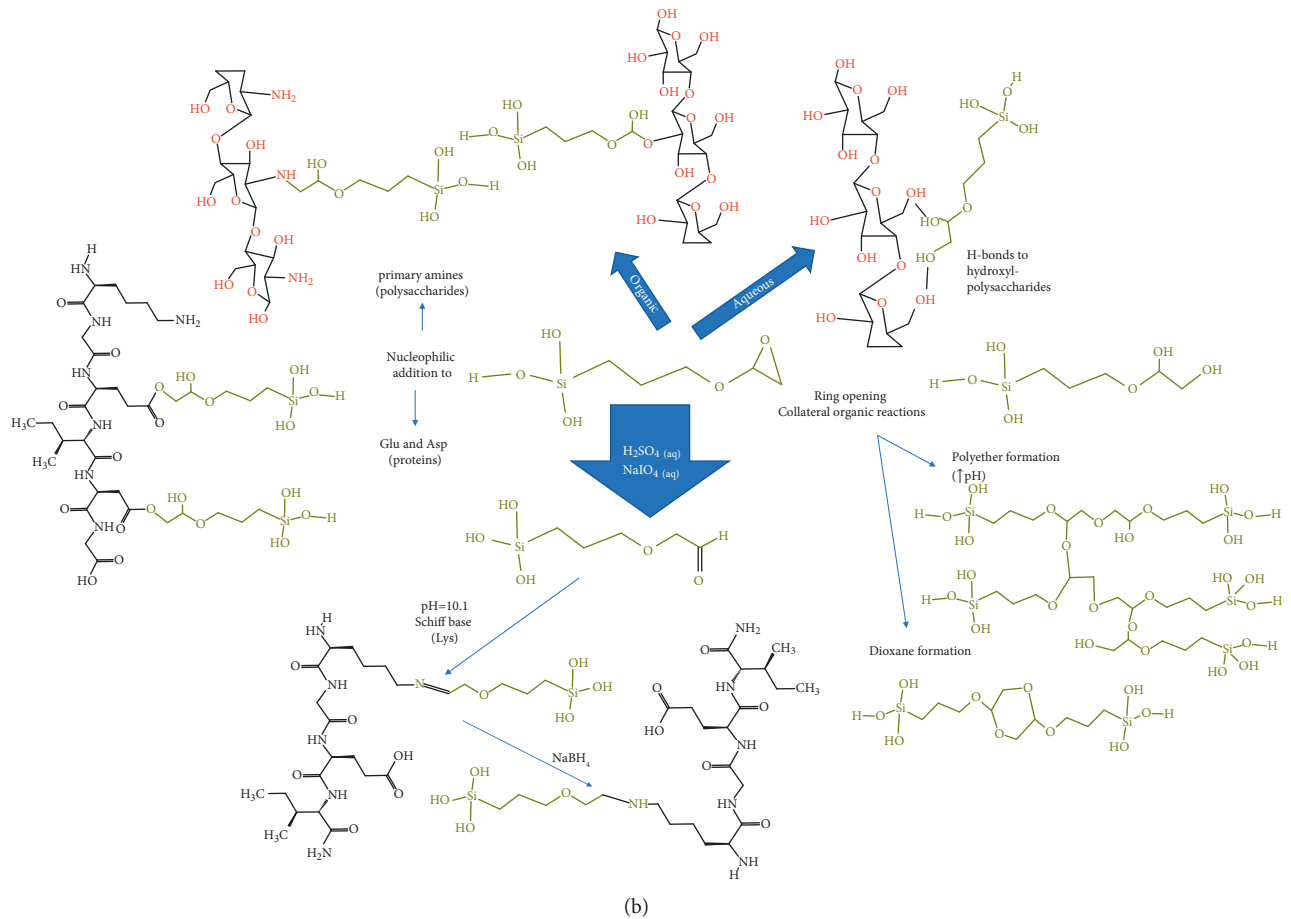


FIGURE 5: Multiple reactions and interaction mechanism of (a) APTES and (b) GPTMS silanes in the presence of proteins and polysaccharide hydrogel precursors.

polymer mobility, which explains why silica reinforces the gel, helping to maintain the foam structure after adsorption of larger wound exudate volumes compared with the pure chitosan gel [43, 45]. This water adsorption capability and faster wettability are induced by the silica surface hydrophilicity, which is also important for cell viability. The in vivo tests show that the silica presence enhances the fibroblast proliferation, secretion of TGF- β growth factor, and collagen deposition [43].

The chemical nature, the concentration of the silica source, and temperature affect the protein autoassembly, which has consequences on the hydrogel microstructure. For example, the APTES polycondensation during gelation of fibrinogen from bovine serum produced reinforced fibrin hydrogels with more homogenous fiber diameters than TEOS at lower concentrations, without affecting the myoblast growing in both cases. The differences were explained by the ammonium functionality APTES, which contribute to the electrostatic interactions with proteins and modulate the ionic strength of the medium, affecting the fiber thickness and stiffness [46]. On the other hand, the increase of the temperature above the denaturation protein (70°C) can be responsible of changes on the fibrinogen association and interaction strength with anionic silicate species (from sodium silicate) by the Arg and Lys cationic

rich region at pH 3. They affect the stiffness of the fibrin hydrogel and the silica condensation extent and microstructure [47].

3.1.2. Gelation in the Presence of the Preformed Silica Materials. The route (ii) involves the use of preformed pure or functionalized silica, having also physical and biological contributions. The hydrogen bond interactions between silica nanoparticles and glycol chitosan restricted the movement of polymer chains, which affected the sol-gel transition, elastic (G'), and loss (G'') moduli and improved the adhesive properties of this injectable hydrogel [5]. These particles can be nucleation agents for hydrogels polymerization and/or crosslinking. For example, the presence of pure silica on the blood plasma gelation induced by CaCl₂ and crosslinking with glutaraldehyde increase the fiber thickness and hydrogel porosity [48]. This is due to their interaction with fibrinogen (isoelectric point=5.5) especially through the electrostatic interactions (NH³⁺ vs. SiO⁻) in the α -domain, as studied by sodium silicate polycondensation in the presence of fibrinogen [47]. Other Van der Waals interactions and H-bonds can be involved (-NH and -OH vs. Si-OH and SiO⁻) (Figure 3(b)), usually in protein-derived hydrogels depending on the pH of the medium and

pKa of the amino acids [49]. The size and concentration of silica nanoparticle nuclei must be controlled for maintaining the fibrillar organization and avoid silica aggregation. This is because nanoparticles are reinforcing charges if they are homogeneously distributed through the gel [50]. On the contrary, the aggregates can be breakdown points under mechanical stress, as exemplified with elastic collagen hydrogels [51]. Silica also increases the elastic behavior of the injectable cellulose and chitosan hydrogels [5, 52] and the tensile strength of silica-cellulose membranes [53]. Another physical contribution of silica nanoparticles is related to the swelling. It is demonstrated how the highly organized porous structure of SBA-15 mesoporous silica added during the alkaline casting and acidic recovering of cellulose films improves the swelling rate, conferring a 1012 to 1186 $\text{g}\cdot\text{m}^{-2}\cdot\text{d}^{-1}$ moisture permeability. This is into the ideal values (904–1447 $\text{g}\cdot\text{m}^{-2}\cdot\text{d}^{-1}$) for wound dressings [53].

The nanoparticles can preserve the biological contributions discussed in the precedent section after their incorporation on the hydrogels. This is because their dissolution under physiological conditions produces silicic acid [26, 32], which can be progressively delivered from the hydrogel. For example, from fibrin scaffolds without affecting the cell behavior [48] and showing wound healing synergic effects on succinyl chitosan/dialdehyde-based oxidized alginate crosslinked injectable hydrogel, favoring collagen/myofibrils accumulation, new blood vessels formation, and L929 fibroblast viability [52]. On the other hand, silica can also be a reservoir of bioactive compounds, such as keratin growth factors and interleukins, improving and expanding the biological functions of the wound dressings [28, 54].

A special interest has been put in the controlled release of antimicrobial agents. They can be inorganic oxides such as ZnO incorporated during the synthesis of bioactive glass nanoparticles [52], Cu nanoparticles covered by silica in a starch medium [55], or organic antibiotics adsorbed on pure [51, 56, 57] or chemically-modified [39, 58–60] nanosilicas, before being added to the gel. Silica microspheres loaded with ferulic acid during TEOS polymerization conferred antimicrobial and antioxidant properties to fibrin/chitosan/keratin wound scaffolds [61]. The delivery profile and cytotoxicity are dependent of particle size/concentration, as exemplified with collagen hydrogels [51]. Moreover, the silica chemical modification can modulate the hydrophobicity and charge of the surface, which in turn affect the antibiotic diffusion from the wound dressing. This was the objective of silica functionalization with (3-mercaptopropyl)trimethoxysilane for having hydrophobic propyl-thiol groups (Figure 3(c)) and treatment with sulfuric acid for having negative charge (sulfonate; Figure 3(d)) groups [58]. These chemical modifications could affect also silica interactions with protein precursor; nevertheless, they continue being physical (Figure 3(c)) because the abundance of nucleophilic cysteines (pKa 8) is so low and there are not oxidant agents promoting a disulfide formation [62]. On the other hand, the delivery can be also driven by the silica dissolution and hydrogel degradation. This was shown with preloaded FICT-lysozyme silica supraparticles assembly,

coated with fibrin polymerized from fibrinogen in the presence of thrombin [63].

Recently, γ -chloropropyl-triethoxysilane (CPTES) was used as the cross-linking agent between N-halamine chemically modified silica particles and chitosan. The silanol groups of the poly(CPTES) chains, formed at 80°C, interact with the amine and hydroxyls groups of the chitosan. This physical entanglement leads to a stable, low density, 3D interconnected porous structure with additional swelling, hemostatic, and bactericidal properties [64].

3.2. Covalent Bonding. The silica can contribute to the covalent bonding of hydrogels in form of (i) silylated silicas, whose surface chemical group will act as coupling and crosslinker functionalities, and (ii) silsesquioxanes and oligomers formed after coupling an alkoxy silane to the protein or polysaccharide hydrogel precursor (Figure 4).

3.2.1. Functionalization of Silicas for Their Use in Hydrogels.

Silica particles or fibers can be chemically modified by condensation their silanol groups with organo-alkoxysilanes, chlorosilanes, or organochlorines. The APTES is a common organosilane [65] for coupling the silica to bifunctional crosslinkers such as glutaraldehyde, glyoxal [66, 67], and squaric acid [68], typical in the production of protein-derived hydrogels (Figure 4(a)). The silanization process allows controlling the amine density, nucleophilicity, and ionization degree for being physical and covalent reactive attachment multipoints in the silica surface [69]. The covalent coupling with glutaraldehyde occurs at a pH higher than 7.6, which is the pKa of the amino groups in this kind of amino-silica surface [70]. The modification degree can be measured indirectly by the free-amino groups by the OPA method [67]. After that, the aldehyde groups on the chemically modified silica particles condensate mainly with the primary amine groups of lysines (pKa 10.3), forming an imine bond (Figure 4(a)). For example, they lead to a highly dense chemical gelatin crosslinked microstructure with a more elastic behavior and lower swelling volume and water desorption at a lower temperature than the gel prepared in the presence of nonmodified silicas [49]. The silica functionalized with APTES/glutaraldehyde in the form of films can also support the formation of smooth and homogenous protein monolayer. That was the case of a keratin scaffold, promoting the adhesion and proliferation of human dermal fibroblasts and epidermal keratinocytes [71]. Another crosslinker is the genipin (Figure 4(a)), whose proposed crosslinking mechanism involves the nucleophilic attack of primary amines of the protein and dimerization [72], being a second-order reaction on fibrin hydrogels [73]. The chemical crosslinking mediated genipin-amino-silicas avoided the inorganic/organic phase separation due to nanoparticles aggregation in collagen/chitosan/hyaluronic hydrogels, increasing the hydrogel stiffness [74].

Even less common, the silica nanoparticles modified vinyltrimethoxysilane can be also reactive crosslinkers (Figure 4(b)). Their vinyl groups participate in radical

activated reactions with a maleic anhydride modified-casein and glycidyl methacrylate modified-chondroitin, started by sodium persulfate in N,N,N',N'-tetramethyl ethylenediamine and ended by combination or disproportion reactions. This gave a chemical crosslinked gel, where the silica nanospheres were well-dispersed in protein-derived hydrogels, and controlled the sustained release of the selected drug, as they interfere with the interactions between the drug and the protein matrix [75].

Organochlorine such as epichlorohydrin can be also used for the chemical modification of silica. It reacts with the free silanol, forming a carbon-oxygen bond and releasing HCl in organic solvents at 70°C. The epoxy functionality on these functionalized particles is an electrophile, which can react with amine groups in saccharides such as chitosan (Figure 4(c)), via the epoxy opening pathway, contributing to the polymer crosslinking. The hybrid nature has synergic effects on the wound healing, exudates adsorption, and gas exchange of the wound dressings [76].

3.2.2. Functionalization of Protein or Polysaccharide Hydrogel Precursor with Silanes. The coupling of organoalkoxysilanes to the protein or saccharide hydrogel precursor can occur by physical and covalent interactions (Figure 5). These organoalkoxysilanes behave as bifunctional reagents, whose chemical grafting and the extent of organic and inorganic polymerization depend on the reaction conditions. For example, the chemical grafting of APTES through the silanol groups to cellulose nanofibrils is favored in ethanol and drying at 105°C; by the contrary, the reaction in water at room temperature promotes the H-bonds interactions between the primary amine of the alkylalkoxysilane with the hydroxyl of the polysaccharide [77]. Then, the selection of the solvent and temperature is the key for having a free amine or silanol groups, contributing to the chemical crosslinking of the hydrogel [77] through the H-bonds or siloxane formation (Figure 5(a)). The amine behaves as a basic catalyst, promoting particle aggregation [78]. Therefore, special care must be put in controlling its concentration for avoiding the formation of breakdown clusters that affect the hydrogel mechanical properties.

The GPTMS is another alkoxysilane, in which the two electrophilic carbons on the epoxy ring are prone to the nucleophilic attack of primary amine groups on polysaccharides and proteins, forming secondary amines (Figure 5(b)). However, the chemical grafting depends on the polymer type, reactants concentration, pH, temperature, and addition of silica [79]. In the case of silica-chitosan scaffolds, the ¹³C and ²⁹Si NMR show the nucleophilic addition is through the primary amine of chitosan pH 2–4 and 40°C, accompanied by the epoxy ring-opening and diol formation. The condensation degree of the free silanol groups depends on the pH and the addition of hydrolyzed TEOS, which in turn affect the dissolution, porosity, and mechanical properties of the scaffolds form by freezing casting [80]. The increasing of GPTMS concentration improves the chemical grafting and crosslinking degree on the chitosan composites (followed by FTIR and ²⁹Si-NMR),

decreasing the porosity and retarding the scaffold dissolution [81].

The probability that the glyoxyl group could be covalently attached to the hydroxyl and silanol groups is lower compared with the amines, taking into account that the reaction rates towards epoxy groups follow the tendency $RSH > RNH_2, R_2NH > RCOOH > SiOH \gg ROH > H_2O$ [82]. Therefore, the H-bonds interactions occur with the diols formed under acid conditions [82], contributing also to the polysaccharides-derived hydrogel crosslinking (Figure 5(b)). On the other hand, reactions in organic solvents [83] allow the glyoxyl chemical modification of hydroxyl-rich polysaccharides. For example, the further free silanol groups' condensation with silica nanofibers in water improves the mechanical properties and water diffusion of silanized hydroxypropylmethylcellulose hydrogel [84, 85].

The GPTMS for protein chemical modification occurs mainly through the glutamic and aspartic amino acids instead of lysines and arginines (Figure 5(b)). This is because acid residues are the most abundant and better nucleophiles at pH between 5 and 7, where this protein modification is commonly made. That was the case of gelatin-derived hydrogels, where the epoxy reactions and the condensation of the silane also occur simultaneously in the presence of the Bronsted catalyst at pH 5, producing a gel when the reaction time is long and temperature is higher than 40°C [86]. This kind of scaffolds is biocompatible, and the porosity and mechanical strength can be controlled by the inorganic-organic coupling for being scaffolds on tissue engineering [87, 88]. Moreover, the chemical crosslinking increases the hydrophobicity and thermal stability of these scaffolds [89]. The GPTMS hydrolysis at highly acid pH (H₂SO₄ aqueous solution) and oxidation with NaIO₄ transform the epoxy into an aldehyde functionality, which forms a Schiff base with lysine residues at pH higher 10.1 (pK of the Lys). Then, it can be stabilized by reduction with sodium borohydride (Figure 5(b)). At basic pH, the kinetically controlled inorganic condensation of 3-glycidoxypropyltrimethoxysilane predominates, affecting the opening and organic reactions of the epoxy ring. The sol is stable for months [90]. The ring-opening and organic reactions are more efficient when the NaOH is increased [91] and can occur also in the polyether formation (Figure 5(b)). Then, self-reactions and interactions with proteins and polysaccharides are competing. Therefore, the selection of the medium conditions must be carefully chosen for promoting the hydrogel crosslinking.

4. Concluding Remarks and Perspectives

The silanes and silicas contribute to the barrier, scaffold, and delivery matrix functionalities of wound dressings hydrogels. They modify the physicochemical and biological characteristics of the hydrogel. The adhesive properties of these inorganic compounds by silanol condensation can be useful for the hydrogel formation and cells/hydrogels interactions. Physical and covalent mechanisms of silane and silica interactions with protein and polysaccharides could modulate the crosslinking degree and bioactivity of the

derived hydrogels. The former has consequences on the elasticity and stiffness mechanical properties, wetting, permeability (nutrient transport), and degradation of the hybrid hydrogel. Moreover, silica has positive effects on the bioactivity of the wound dressing, directly through the dissolution up to silicic acid or by carrying bioactive molecules (growth factors, antioxidants, and antibiotics).

The reinforcement of polysaccharide and protein derived-hydrogels for having multifunctional wound dressings is an open interdisciplinary field of research. The chemical modification of silica with crosslinkers, markers, and bioactive molecules gives opportunities of having first, second, and third-generation hydrogels (analogous to the hierarchical nanocarriers classification), which is beneficial to produce smart materials. The high relevance of silica on the mechanical properties of these natural hydrogels encourages the research of new processing and characterization methods for increasing the yield efficiency and sensible testing. The medical staff and users have more alternatives to conventional wound healing treatments.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

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References

- [1] A. M. Darlington and J. M. Gibbs-Davis, "Bimodal or trimodal? The influence of starting pH on site identity and distribution at the low salt aqueous/silica interface," *Journal of Physical Chemistry C*, vol. 119, no. 29, pp. 16560–16567, 2015.
- [2] A. Issa and A. Luyt, "Kinetics of alkoxy-silanes and organo-alkoxy-silanes polymerization: a review," *Polymers*, vol. 11, no. 3, p. 537, 2019.
- [3] J. G. Croissant, Y. Fatiev, A. Almalik, and N. M. Khashab, "Mesoporous silica and organosilica nanoparticles: physical chemistry, biosafety, delivery strategies, and biomedical applications," *Advanced Healthcare Materials*, vol. 7, no. 4, 2018.
- [4] S. V. Patwardhan, F. S. Emami, R. J. Berry et al., "Chemistry of aqueous silica nanoparticle surfaces and the mechanism of selective peptide adsorption," *Journal of the American Chemical Society*, vol. 134, no. 14, pp. 6244–6256, 2012.
- [5] F. Zhu, C. Wang, S. Yang et al., "Injectable tissue adhesive composite hydrogel with fibroblasts for treating skin defects," *Journal of Materials Chemistry B*, vol. 5, no. 13, pp. 2416–2424, 2017.
- [6] C. Feng, J. Li, G. S. Wu et al., "Chitosan-coated diatom silica as hemostatic agent for hemorrhage control," *ACS Applied Materials & Interfaces*, vol. 8, no. 50, pp. 34234–34243, 2016.
- [7] M. Naseri-Nosar and Z. M. Ziora, "Wound dressings from naturally-occurring polymers: a review on homopolysaccharide-based composites," *Carbohydrate Polymers*, vol. 189, pp. 379–398, 2018.
- [8] J. Kurczewska, P. Pecyna, M. Ratajczak, M. Gajęcka, and G. Schroeder, "Halloysite nanotubes as carriers of vancomycin in alginate-based wound dressing," *Saudi Pharmaceutical Journal*, vol. 25, no. 6, pp. 911–920, 2017.
- [9] X. Wang, J. Chang, and C. Wu, "Bioactive inorganic/organic nanocomposites for wound healing," *Applied Materials Today*, vol. 11, pp. 308–319, 2018.
- [10] F. García-Villén, I. M. S. Souza, R. de Melo Barbosa et al., "Natural inorganic ingredients in wound healing," *Current Pharmaceutical Design*, vol. 26, no. 6, pp. 621–641, 2020.
- [11] S. R. K. Meka, V. Agarwal, and K. Chatterjee, "In situ preparation of multicomponent polymer composite nanofibrous scaffolds with enhanced osteogenic and angiogenic activities," *Materials Science and Engineering: C*, vol. 94, pp. 565–579, 2019.
- [12] S. R. K. Meka, S. Kumar Verma, V. Agarwal, and K. Chatterjee, "In situ silication of polymer nanofibers to engineer multi-biofunctional composites," *Chemistry*, vol. 3, no. 13, pp. 3762–3773, 2018.
- [13] G. Suarato, R. Bertorelli, and A. Athanassiou, "Borrowing from nature: biopolymers and biocomposites as smart wound care materials," *Frontiers in Bioengineering and Biotechnology*, vol. 6, p. 137, 2018.
- [14] M. C. Catoira, L. Fusaro, D. Di Francesco, M. Ramella, and F. Boccafroschi, "Overview of natural hydrogels for regenerative medicine applications," *Journal of Materials Science: Materials in Medicine*, vol. 30, no. 10, p. 115, 2019.
- [15] J. Y. Ng, S. Obuobi, M. L. Chua et al., "Biomimicry of microbial polysaccharide hydrogels for tissue engineering and regenerative medicine - a review," *Carbohydrate Polymers*, vol. 241, Article ID 116345, 2020.
- [16] W. Hu, Z. Wang, Y. Xiao, S. Zhang, and J. Wang, "Advances in crosslinking strategies of biomedical hydrogels," *Biomaterials Science*, vol. 7, no. 3, pp. 843–855, 2019.
- [17] R. A. Batista et al., "Hydrogel as an alternative structure for food packaging systems," *Carbohydrate Polymers*, vol. 205, pp. 106–116, 2019.
- [18] N. Y. Becerra, L. M. Restrepo, Y. Galeano, A. C. Tobón, L. F. Turizo, and M. Mesa, "Improving fibrin hydrogels' mechanical properties, through addition of silica or chitosan-silica materials, for potential application as wound dressings," *International Journal of Biomaterials*, vol. 2021, pp. 1–11, Article ID 9933331, 2021.
- [19] A. Frayssinet, D. Petta, C. Illoul et al., "Extracellular matrix-mimetic composite hydrogels of cross-linked hyaluronan and fibrillar collagen with tunable properties and ultrastructure," *Carbohydrate Polymers*, vol. 236, Article ID 116042, 2020.
- [20] J. Jang, Y.-J. Seol, H. J. Kim, J. Kundu, S. W. Kim, and D.-W. Cho, "Effects of alginate hydrogel cross-linking density on mechanical and biological behaviors for tissue engineering," *Journal of the Mechanical Behavior of Biomedical Materials*, vol. 37, pp. 69–77, 2014.
- [21] C. D. Markert, X. Guo, A. Skardal et al., "Characterizing the micro-scale elastic modulus of hydrogels for use in regenerative medicine," *Journal of the Mechanical Behavior of Biomedical Materials*, vol. 27, pp. 115–127, 2013.
- [22] C. E. Vorwald, T. Gonzalez-Fernandez, S. Joshee, P. Sikorski, and J. K. Leach, "Tunable fibrin-alginate interpenetrating network hydrogels to support cell spreading and network formation," *Acta Biomaterialia*, vol. 108, pp. 142–152, 2020.
- [23] F. Afinjuomo, T. G. Barclay, Y. Song, A. Parikh, N. Petrovsky, and S. Garg, "Synthesis and characterization of a novel inulin

- hydrogel crosslinked with pyromellitic dianhydride,” *Reactive and Functional Polymers*, vol. 134, pp. 104–111, 2019.
- [24] R. K. Iler, *The Chemistry of Silica*, Wiley-Interscience, Hoboken, NJ, USA, 1979, http://link.springer.com/10.2991/978-94-6239-246-5_20.
- [25] S. Rose, A. PrevotEAU, P. Elzière, D. Hourdet, A. Marcellan, and L. Leibler, “Nanoparticle solutions as adhesives for gels and biological tissues,” *Nature*, vol. 505, no. 7483, pp. 382–385, 2014.
- [26] J.-H. Kim, H. Kim, Y. Choi, D. S. Lee, J. Kim, and G.-R. Yi, “Colloidal mesoporous silica nanoparticles as strong adhesives for hydrogels and biological tissues,” *ACS Applied Materials & Interfaces*, vol. 9, no. 37, pp. 31469–31477, 2017.
- [27] S. Zirak Hassan Kiadeh, A. Ghaee, A. Mashak, and J. Mohammadnejad, “Preparation of chitosan-silica/PCL composite membrane as wound dressing with enhanced cell attachment,” *Polymers for Advanced Technologies*, vol. 28, no. 11, pp. 1396–1408, 2017.
- [28] J.-S. Oh and E.-J. Lee, “Engineered dressing of hybrid chitosan-silica for effective delivery of keratin growth factor and acceleration of wound healing,” *Materials Science and Engineering: C*, vol. 103, Article ID 109815, 2019.
- [29] X. Sun, Y. Fang, Z. Tang, Z. Wang, X. Liu, and H. Liu, “Mesoporous silica nanoparticles carried on chitosan microspheres for traumatic bleeding control,” *International Journal of Biological Macromolecules*, vol. 127, pp. 311–319, 2019.
- [30] G. Patil, A. Torris, P. R. Suresha, S. Jadhav, M. V. Badiger, and V. Ghormade, “Design and synthesis of a new topical agent for halting blood loss rapidly: a multimodal chitosan-gelatin xerogel composite loaded with silica nanoparticles and calcium,” *Colloids and Surfaces B: Biointerfaces*, vol. 198, Article ID 111454, 2021.
- [31] S. Quignard, T. Coradin, J. J. Powell, and R. Jugdaohsingh, “Silica nanoparticles as sources of silicic acid favoring wound healing in vitro,” *Colloids and Surfaces B: Biointerfaces*, vol. 155, pp. 530–537, 2017.
- [32] L. Kong, Z. Wu, H. Zhao et al., “Bioactive injectable hydrogels containing desferrioxamine and bioglass for diabetic wound healing,” *ACS Applied Materials & Interfaces*, vol. 10, no. 36, pp. 30103–30114, 2018.
- [33] M. Singh Bakshi, “Nanotoxicity in systemic circulation and wound healing,” *Chemical Research in Toxicology*, vol. 30, no. 6, 2017.
- [34] J. Chen, J. Ai, S. Chen et al., “Synergistic enhancement of hemostatic performance of mesoporous silica by hydrocaffeic acid and chitosan,” *International Journal of Biological Macromolecules*, vol. 139, pp. 1203–1211, 2019.
- [35] B. Qiao, Q. Pang, P. Yuan, Y. Luo, and L. Ma, “Smart wound dressing for infection monitoring and NIR-triggered antibacterial treatment,” *Biomaterials Science*, vol. 8, no. 6, pp. 1649–1657, 2020.
- [36] A. L. Mohamed, H. Elmotasem, and A. A. A. Salama, “Colchicine mesoporous silica nanoparticles/hydrogel composite loaded cotton patches as a new encapsulator system for transdermal osteoarthritis management,” *International Journal of Biological Macromolecules*, vol. 164, pp. 1149–1163, 2020.
- [37] L. Pomeranic and D. Benayahu, “Mesenchymal cell growth and differentiation on a new biocomposite material: a promising model for regeneration therapy,” *Biomolecules*, vol. 10, no. 3, p. 458, 2020.
- [38] L. F. Giraldo, B. L. López, L. Pérez, S. Urrego, L. Sierra, and M. Mesa, “Mesoporous silica applications,” *Macromolecular Symposia*, vol. 258, no. 1, pp. 129–141, 2007.
- [39] J. Kurczewska, P. Sawicka, M. Ratajczak, M. Gajęcka, and G. Schroeder, “Will the use of double barrier result in sustained release of vancomycin? Optimization of parameters for preparation of a new antibacterial alginate-based modern dressing,” *International Journal of Pharmaceutics*, vol. 496, no. 2, pp. 526–533, 2015.
- [40] E. Caló and V. V. Khutoryanskiy, “Biomedical applications of hydrogels: a review of patents and commercial products,” *European Polymer Journal*, vol. 65, pp. 252–267, 2015.
- [41] L. Wang, C. Gong, X. Yuan, and G. Wei, “Controlling the self-assembly of biomolecules into functional nanomaterials through internal interactions and external stimulations: a review,” *Nanomaterials*, vol. 9, no. 2, 2019.
- [42] L. J. del Valle, A. Díaz, and J. Puiggali, “Hydrogels for biomedical applications: cellulose, chitosan, and protein/peptide derivatives,” *Gels*, vol. 3, no. 3, p. 27, 2017.
- [43] J.-U. Park, H.-D. Jung, E.-H. Song et al., “The accelerating effect of chitosan-silica hybrid dressing materials on the early phase of wound healing,” *Journal of Biomedical Materials Research Part B: Applied Biomaterials*, vol. 105, no. 7, pp. 1828–1839, 2017.
- [44] J. Diosa, J. C. Poveda-Jaramillo, F. Ramirez-Rodríguez, and M. Mesa, “Modeling surface chemistry and adsorption behavior of biomimetic chitosan/silica hybrid materials,” *Journal of Materials Research and Technology*, vol. 9, no. 4, pp. 8092–8103, 2020.
- [45] J.-U. Park, S.-H. Jeong, E.-H. Song, J. Song, H.-E. Kim, and S. Kim, “Acceleration of the healing process of full-thickness wounds using hydrophilic chitosan-silica hybrid sponge in a porcine model,” *Journal of Biomaterials Applications*, vol. 32, no. 8, pp. 1011–1023, 2018.
- [46] K. Wang, K. Albert, G. Mosser et al., “Self-assembly/condensation interplay in nano-to-microfibrillar silicified fibrin hydrogels,” *International Journal of Biological Macromolecules*, vol. 164, pp. 1422–1431, 2020.
- [47] N. Hassan, A. Soltero, D. Pozzo, P. V. Messina, and J. M. Ruso, “Bioinspired templates for the synthesis of silica nanostructures,” *Soft Matter*, vol. 8, no. 37, pp. 9553–9562, 2012.
- [48] F. Sani, F. Mehdipour, T. Talaei-Khozani, M. Sani, and V. Razban, “Fabrication of platelet-rich plasma/silica scaffolds for bone tissue engineering,” *Bioinspired, Biomimetic and Nanobiomaterials*, vol. 7, no. 2, pp. 74–81, 2018.
- [49] D. Rocha-García, M. d. L. Betancourt-Mendiola, A. Wong-Arce et al., “Gelatin-based porous silicon hydrogel composites for the controlled release of tramadol,” *European Polymer Journal*, vol. 108, pp. 485–497, 2018.
- [50] J. Yang, J.-J. Zhao, C.-R. Han, and J.-F. Duan, “Keys to enhancing mechanical properties of silica nanoparticle composites hydrogels: the role of network structure and interfacial interactions,” *Composites Science and Technology*, vol. 95, pp. 1–7, 2014.
- [51] G. S. Alvarez, C. Hélyary, A. M. Mebert, X. Wang, T. Coradin, and M. F. Desimone, “Antibiotic-loaded silica nanoparticle-collagen composite hydrogels with prolonged antimicrobial activity for wound infection prevention,” *Journal of Materials Chemistry B*, vol. 2, no. 29, pp. 4660–4670, 2014.
- [52] J. Zhu, G. Jiang, G. Song et al., “Incorporation of ZnO/bioactive glass nanoparticles into alginate/chitosan composite hydrogels for wound closure,” *ACS Applied Bio Materials*, vol. 2, no. 11, pp. 5042–5052, 2019.

- [53] Z. Shen et al., "Engineering sustainable antimicrobial release in silica-cellulose membrane with CaCO₃-aided processing for wound dressing application," *Polymers*, vol. 11, no. 5, pp. 1–18, 2019.
- [54] X. Wang, T. Coradin, and C. Héлары, "Modulating inflammation in a cutaneous chronic wound model by IL-10 released from collagen-silica nanocomposites via gene delivery," *Biomaterials Science*, vol. 6, no. 2, pp. 398–406, 2018.
- [55] M. E. Villanueva, A. M. d. R. Diez, J. A. González et al., "Antimicrobial activity of starch hydrogel incorporated with copper nanoparticles," *ACS Applied Materials & Interfaces*, vol. 8, no. 25, pp. 16280–16288, 2016.
- [56] R. K. Perumal, A. Gopinath, R. Thangam et al., "Collagen-silica bio-composite enriched with Cynodon dactylon extract for tissue repair and regeneration," *Materials Science and Engineering: C*, vol. 92, pp. 297–306, 2018.
- [57] H. Namazi, R. Rakhshaei, H. Hamishehkar, and H. S. Kafil, "Antibiotic loaded carboxymethylcellulose/MCM-41 nanocomposite hydrogel films as potential wound dressing," *International Journal of Biological Macromolecules*, vol. 85, pp. 327–334, 2016.
- [58] A. M. Mebert, G. S. Alvarez, R. Peroni et al., "Collagen-silica nanocomposites as dermal dressings preventing infection in vivo," *Materials Science and Engineering: C*, vol. 93, pp. 170–177, 2018.
- [59] R. Rakhshaei and H. Namazi, "A potential bioactive wound dressing based on carboxymethyl cellulose/ZnO impregnated MCM-41 nanocomposite hydrogel," *Materials Science and Engineering: C*, vol. 73, pp. 456–464, 2017.
- [60] S. Hashemikia, F. Farhangpazhouh, M. Parsa, M. Hasan, A. Hassanzadeh, and M. Hamidi, "Fabrication of ciprofloxacin-loaded chitosan/polyethylene oxide/silica nanofibers for wound dressing application: in vitro and in vivo evaluations," *International Journal of Pharmaceutics*, vol. 597, Article ID 120313, 2021.
- [61] S. Sivakumar, R. Murali, D. Arathanaikotti et al., "Ferulic acid loaded microspheres reinforced in 3D hybrid scaffold for antimicrobial wound dressing," *International Journal of Biological Macromolecules*, vol. 177, pp. 463–473, 2021.
- [62] E. Steen Redeker, D. T. Ta, D. Cortens, B. Billen, W. Guedens, and P. Adriaensens, "Protein engineering for directed immobilization," *Bioconjugate Chemistry*, vol. 24, no. 11, pp. 1761–1777, 2013.
- [63] Y. Ma, C. Cortez-Jugo, J. Li et al., "Engineering biocoatings to prolong drug release from supraparticles," *Biomacromolecules*, vol. 20, no. 9, pp. 3425–3434, 2019.
- [64] Y. Wang, M. Yin, X. Zheng, W. Li, and X. Ren, "Chitosan/mesoporous silica hybrid aerogel with bactericidal properties as hemostatic material," *European Polymer Journal*, vol. 142, Article ID 110132, 2021.
- [65] K. M. Koch, "Silane coupling agents," *Journal of Organometallic Chemistry*, vol. 246, no. 1, 1983.
- [66] M. N. Gupta, M. Perwez, and M. Sardar, "Protein cross-linking: uses in chemistry, biology and biotechnology," *Biocatalysis and Biotransformation*, vol. 38, no. 3, pp. 178–201, 2020.
- [67] C. M. Vaz, L. A. De Graaf, R. L. Reis, and A. M. Cunha, "Effect of crosslinking, thermal treatment and UV irradiation on the mechanical properties and in vitro degradation behavior of several natural proteins aimed to be used in the biomedical field," *Journal of Materials Science: Materials in Medicine*, vol. 14, no. 9, pp. 789–796, 2003.
- [68] J. Skopinska-Wisniewska, J. Kuderko, A. Bajek, M. Maj, A. Sionkowska, and M. Ziegler-Borowska, "Collagen/elastin hydrogels cross-linked by squaric acid," *Materials Science and Engineering: C*, vol. 60, pp. 100–108, 2016.
- [69] R. R. Bhat and J. Genzer, "Tuning the number density of nanoparticles by multivariant tailoring of attachment points on flat substrates," *Nanotechnology*, vol. 18, no. 2, 2007.
- [70] H. Zhang, H. X. He, J. Wang, T. Mu, and Z. F. Liu, "Force titration of amino group-terminated self-assembled monolayers using chemical force microscopy," *Applied Physics A: Materials Science & Processing*, vol. 66, no. SUPPL. 1, pp. S269–S271, 1998.
- [71] B. Y. Tan, L. T. H. Nguyen, H.-S. Kim, J.-H. Kim, and K. W. Ng, "Cultivation of human dermal fibroblasts and epidermal keratinocytes on keratin-coated silica bead substrates," *Journal of Biomedical Materials Research Part A*, vol. 105, no. 10, pp. 2789–2798, 2017.
- [72] V. G. Tacias-Pascacio et al., "Genipin as an emergent tool in the design of biocatalysts: mechanism of reaction and applications," *Catalysts*, vol. 9, no. 12, p. 1035, 2019.
- [73] C. Ninh, A. Iftikhar, M. Cramer, and C. J. Bettinger, "Diffusion-reaction models of genipin incorporation into fibrin networks," *Journal of Materials Chemistry B*, vol. 3, no. 22, pp. 4607–4615, 2015.
- [74] J. Lewandowska-Łańcucka, A. Gilarska, A. Buła, W. Horak, A. Łatkiewicz, and M. Nowakowska, "Genipin crosslinked bioactive collagen/chitosan/hyaluronic acid injectable hydrogels structurally amended via covalent attachment of surface-modified silica particles," *International Journal of Biological Macromolecules*, vol. 136, pp. 1196–1208, 2019.
- [75] A. R. Simão, V. H. Fragal, A. M. d. O. Lima et al., "pH-responsive hybrid hydrogels: chondroitin sulfate/casein trapped silica nanospheres for controlled drug release," *International Journal of Biological Macromolecules*, vol. 148, pp. 302–315, 2020.
- [76] A. Aslan, S. Elanthikkal, and A. Bozkurt, "Chitosan/hollow silica sphere nanocomposites for wound healing application," *Journal of Materials Research*, vol. 34, no. 2, pp. 231–239, 2019.
- [77] E. Robles, L. Csóka, and J. Labidi, "Effect of reaction conditions on the surface modification of cellulose nanofibrils with aminopropyl triethoxysilane," *Coatings*, vol. 8, no. 4, p. 139, 2018.
- [78] I. A. Rahman, M. Jafarzadeh, and C. S. Sipaut, "Synthesis of organo-functionalized nanosilica via a co-condensation modification using γ -aminopropyltriethoxysilane (APTES)," *Ceramics International*, vol. 35, no. 5, pp. 1883–1888, 2009.
- [79] M. V. Reyes-Peces, A. Pérez-Moreno, D. M. de-los-Santos et al., "Chitosan-GPTMS-silica hybrid mesoporous aerogels for bone tissue engineering," *Polymers*, vol. 12, no. 11, p. 2723, 2020.
- [80] L. S. Connell, F. Romer, M. Suárez et al., "Chemical characterisation and fabrication of chitosan-silica hybrid scaffolds with 3-glycidoxypropyl trimethoxysilane," *Journal of Materials Chemistry B*, vol. 2, no. 6, pp. 668–680, 2014.
- [81] D. Wang, F. Romer, L. Connell et al., "Highly flexible silica/chitosan hybrid scaffolds with oriented pores for tissue regeneration," *Journal of Materials Chemistry B*, vol. 3, no. 38, pp. 7560–7576, 2015.
- [82] T. J. Horrr and G. D. Reynolds, "The reactions of 3-glycidoxypropyltrimethoxysilane in acidic solutions on polymerization and in the presence of silica," *Journal of Adhesion Science and Technology*, vol. 11, no. 7, pp. 995–1009, 1997.
- [83] X. Bourges, P. Weiss, G. Daculsi, and G. Legeay, "Synthesis and general properties of silylated-hydroxypropyl methylcellulose in prospect of biomedical use," *Advances in Colloid and Interface Science*, vol. 99, no. 3, pp. 215–228, 2002.

- [84] N. Buchtová, A. D'Orlando, P. Judeinstein, O. Chauvet, P. Weiss, and J. Le Bideau, "Water dynamics in silanized hydroxypropyl methylcellulose based hydrogels designed for tissue engineering," *Carbohydrate Polymers*, vol. 202, pp. 404–408, 2018.
- [85] N. Buchtová, G. Réthoré, C. Boyer et al., "Nanocomposite hydrogels for cartilage tissue engineering: mesoporous silica nanofibers interlinked with siloxane derived polysaccharide," *Journal of Materials Science: Materials in Medicine*, vol. 24, no. 8, pp. 1875–1884, 2013.
- [86] L. S. Connell, L. Gabrielli, O. Mahony, L. Russo, L. Cipolla, and J. R. Jones, "Functionalizing natural polymers with alkoxy silane coupling agents: reacting 3-glycidoxypropyl trimethoxysilane with poly(γ -glutamic acid) and gelatin," *Polymer Chemistry*, vol. 8, no. 6, pp. 1095–1103, 2017.
- [87] O. Mahony, S. Yue, C. Turdean-Ionescu et al., "Silica-gelatin hybrids for tissue regeneration: inter-relationships between the process variables," *Journal of Sol-Gel Science and Technology*, vol. 69, no. 2, pp. 288–298, 2014.
- [88] O. Mahony, O. Tsigkou, C. Ionescu et al., "Silica-gelatin hybrids with tailorable degradation and mechanical properties for tissue regeneration," *Advanced Functional Materials*, vol. 20, no. 22, pp. 3835–3845, 2010.
- [89] C. K. Balavigneswaran, R. Venkatesan, P. S. Karuppiah et al., "Silica release from silane cross-linked gelatin based hybrid scaffold affects cell proliferation," *ACS Applied Bio Materials*, vol. 3, no. 1, pp. 197–207, 2020.
- [90] P. Innocenzi, C. Figus, T. Kidchob, M. Valentini, B. Alonso, and M. Takahashi, "Sol-gel reactions of 3-glycidoxypropyl-trimethoxysilane in a highly basic aqueous solution," *Dalton Transactions*, vol. 42, no. 42, pp. 9146–9152, 2009.
- [91] B. Menaa, M. Takahashi, P. Innocenzi, and T. Yoko, "Crystallization in hybrid Organic–Inorganic materials induced by self-organization in basic conditions," *Chemistry of Materials*, vol. 19, no. 8, pp. 1946–1953, 2007.