

CORRECTION OPEN



Correction: Smooth muscle 22 alpha protein inhibits VSMC foam cell formation by supporting normal LXRA signaling, ameliorating atherosclerosis

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The original version of this article unfortunately contained a mistake. Due to a typesetting error some of the figures were omitted and figure legends were misplaced. We sincerely apologize for the errors. The correct figures and legends can be found below.



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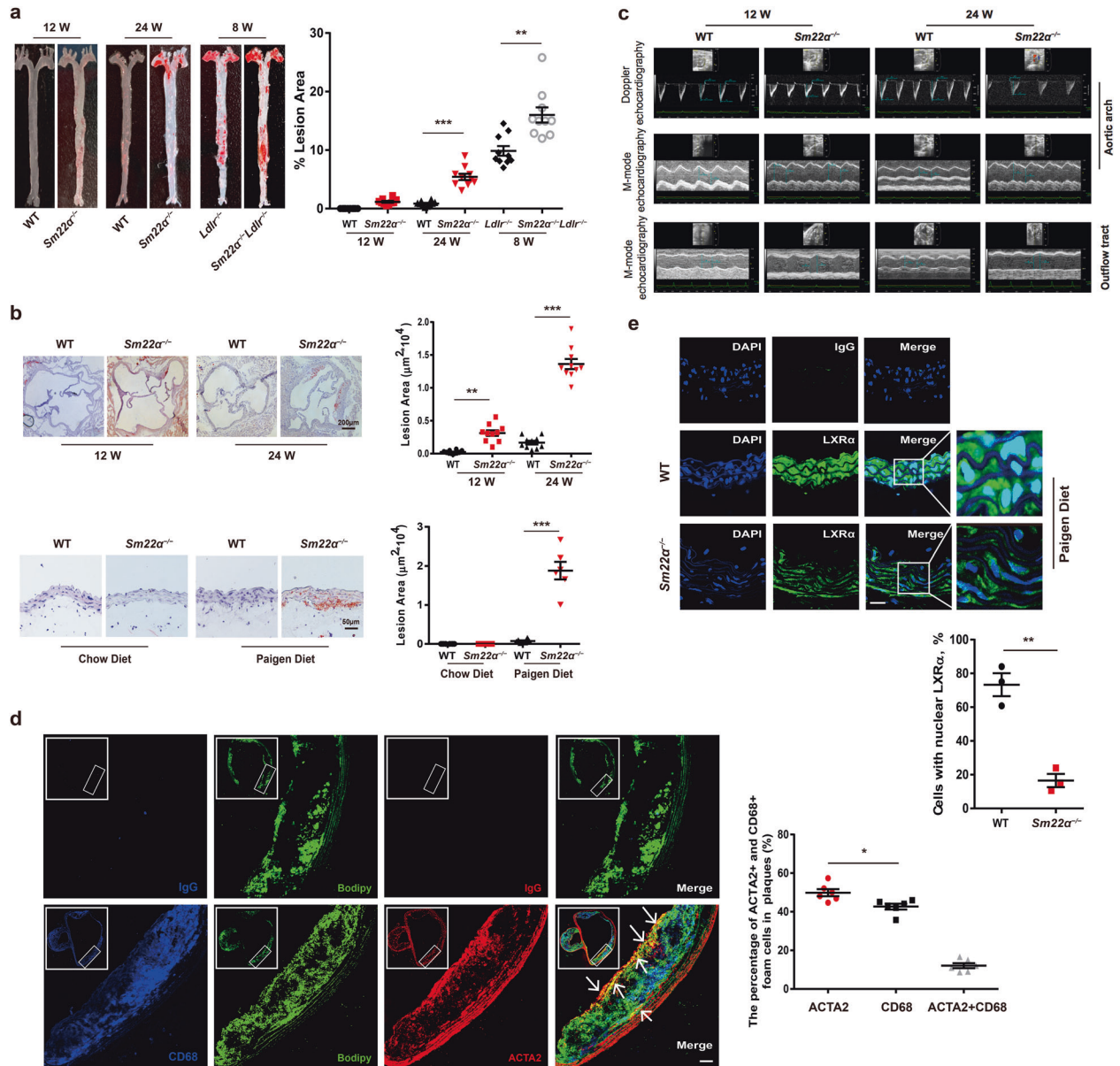
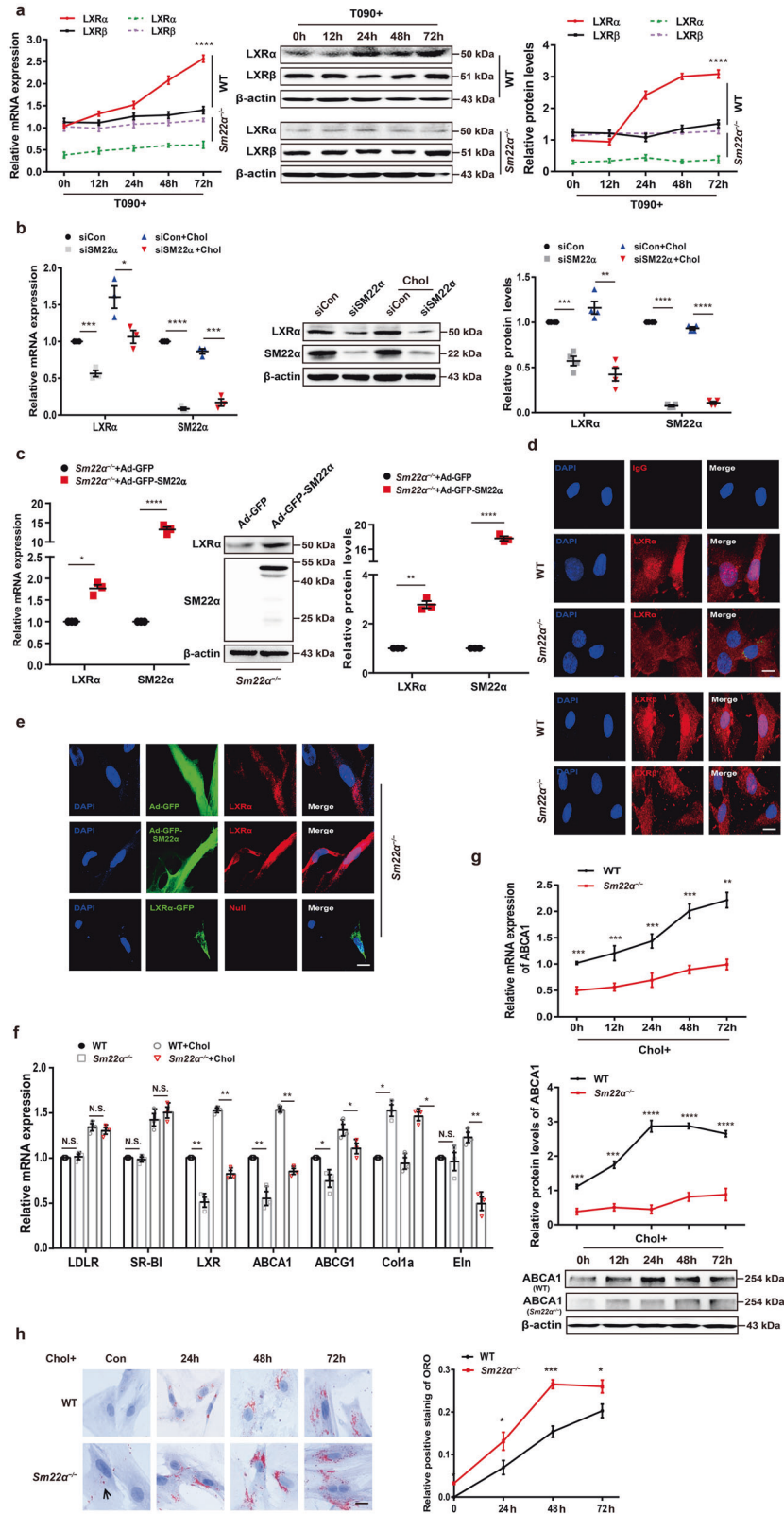


Fig. 1 Impaired SM22 α expression is associated with development of atherosclerosis. **a, b** WT ($n = 10$) and *Sm22 α ^{-/-}* ($n = 10$) mice with or without *Ldlr*^{-/-} background ($n = 10$) fed Paigen diet for 8, 12, and 24 weeks, respectively. Representative images of *en face* ORO-stained aortas (**a**), aortic sinus, aortic cross sections (**b**), and quantification of lesion areas are shown. **c** M-mode and Doppler echocardiography images obtained from aortic arch and outflow tract of WT ($n = 15$) and *Sm22 α ^{-/-}* ($n = 15$) mice fed Paigen diet for 12 and 24 weeks. A_s: outflow tract and aortic diameter in systole; A_d: outflow tract and aortic diameter in diastole. **d** Identification of SMC-derived foam cells within atherosclerotic lesion of *Sm22 α ^{-/-}* mice ($n = 6$) by CD68 (blue), ACTA2 (red), and Bodipy (green). Scale bar, 20 μ m. Arrows indicated foam cells, which were VSMCs-derived. **e** Representative immunofluorescence of LXR α (red) and quantification of cells with nuclear LXR α in the aortic sections from WT ($n = 3$) and *Sm22 α ^{-/-}* ($n = 3$) mice. Scale bar, 15 μ m. Data and images are representative of at least three independent experiments. Data in **a** and **b** were analyzed by two-way and one-way ANOVA, respectively. Data in **d** and **e** were analyzed by unpaired t-test. * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$.



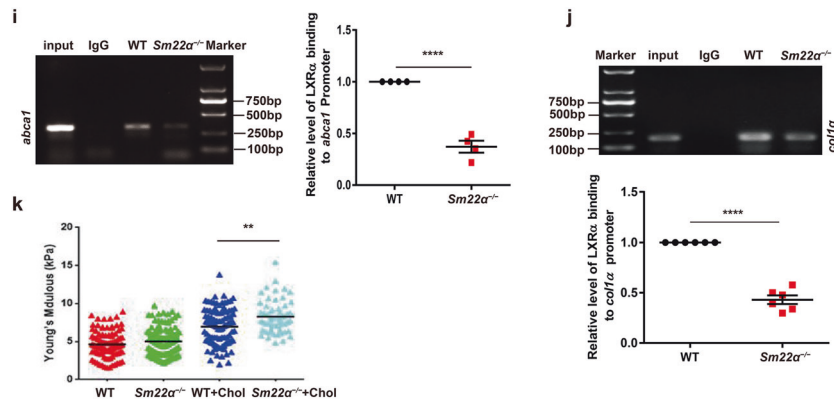
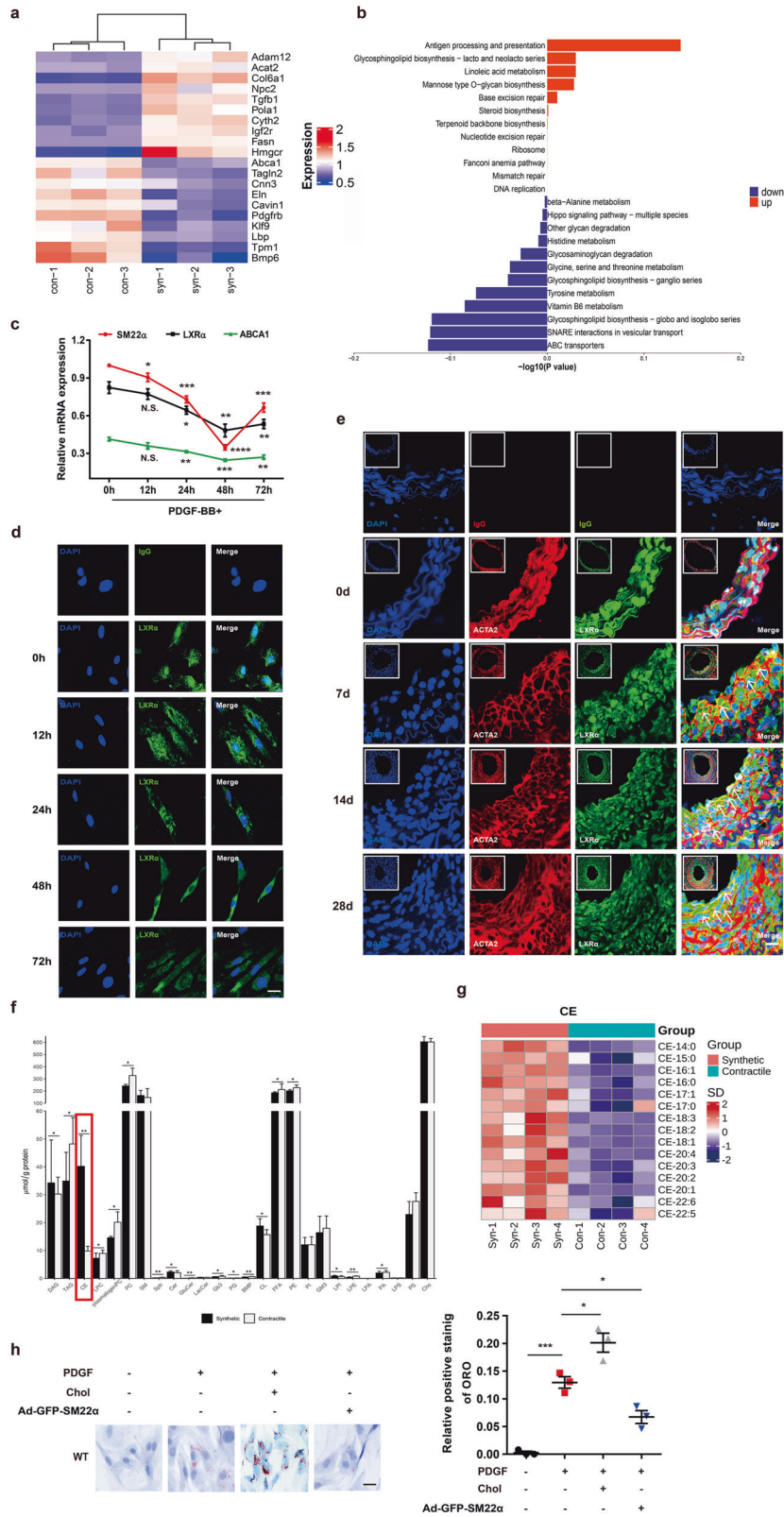


Fig. 2 Expression and activity of LXR α is abnormal in *Sm22 α ^{-/-}* VSMCs. **a** qRT-PCR and Western blot analysis of LXR α and LXR β in WT and *Sm22 α ^{-/-}* VSMCs treated with LXRs agonist T090 for 0, 12, 24, 48, and 72 h, respectively, ($n = 3$). **b** qRT-PCR and western blot analysis of LXR α in WT VSMCs with or without cholesterol loading following knockdown of SM22 α ($n = 3$). **c** qRT-PCR and western blot analysis of LXR α and SM22 α expression in *Sm22 α ^{-/-}* VSMCs transduced with Ad-GFP and Ad-GFP-SM22 α for 24 h ($n = 3$). **d** Confocal microscopy images of LXR α and LXR β distribution in WT and *Sm22 α ^{-/-}* VSMCs. Scale bar, 10 μ m. **e** Immunofluorescence staining for endogenous LXR α and LXR α -GFP in *Sm22 α ^{-/-}* VSMCs transduced with Ad-GFP and Ad-GFP-SM22 α or not. Scale bar, 10 μ m. **f** qRT-PCR analysis of cholesterol intake (LDLR, SR-BI), efflux genes (ABCA1, ABCG1) and sclerosis related genes (Col1 α , Eln) in WT and *Sm22 α ^{-/-}* VSMCs incubated with or without cholesterol ($n = 3$). **g** The mRNA and protein levels of ABCA1 in WT and *Sm22 α ^{-/-}* VSMCs treated with cholesterol for 0, 12, 24, 48, and 72 h, respectively ($n = 3$). **h** ORO staining of WT and *Sm22 α ^{-/-}* VSMCs stimulated with cholesterol for 0, 24, 48, and 72 h, respectively, and quantification of positive ORO staining. Scale bar, 20 μ m. **i** The binding activity of LXR α to the promoter of *abca1* gene was decreased in *Sm22 α ^{-/-}* VSMCs ($n = 4$). **j** ChIP and RT-PCR detected LXR α binding to *col1a* promoter in WT and *Sm22 α ^{-/-}* VSMCs ($n = 6$). **k** The Young's modulus of WT and *Sm22 α ^{-/-}* VSMCs treated with or without cholesterol ($n = 120$). Data and images are representative of at least three independent experiments. Data in **a**, **g**, and **h** were analyzed by Kruskal–Wallis rank sum test and two-way ANOVA. Data in **b**, **c**, **f**, **i**, and **j** were analyzed by unpaired *t*-test. N.S. not significantly different; * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$; **** $p < 0.0001$.



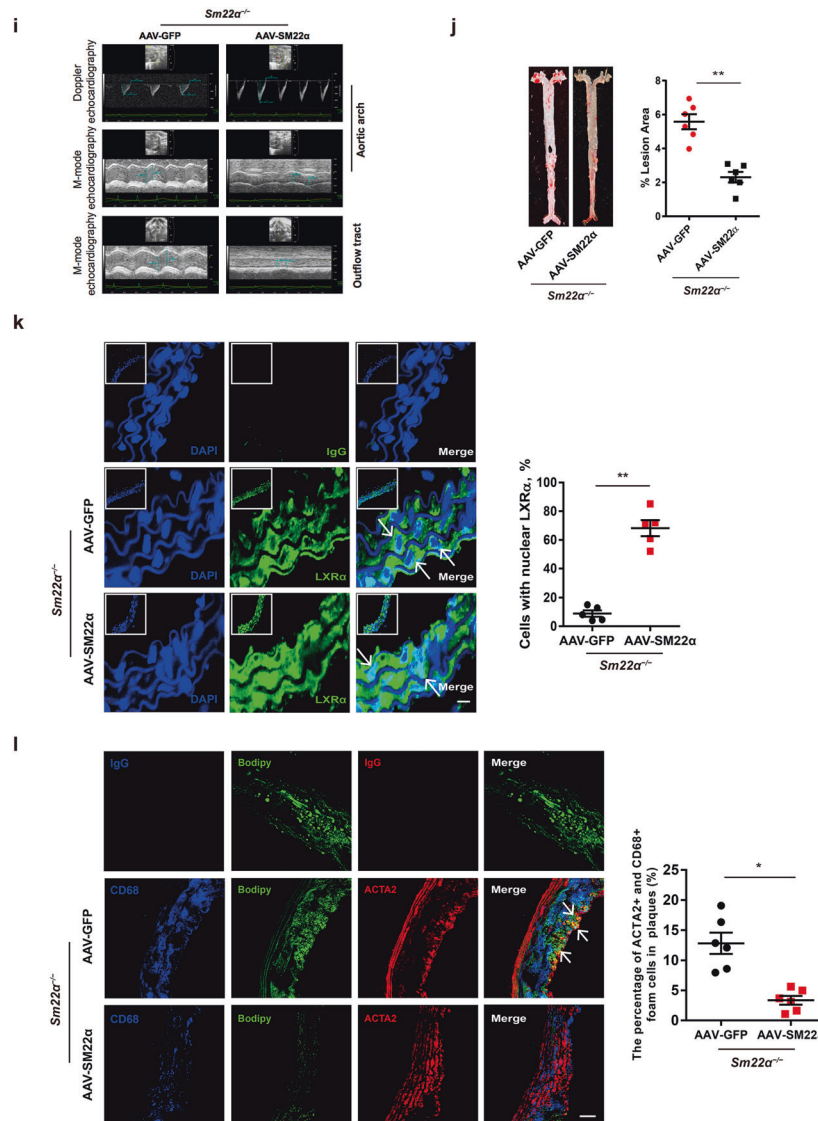


Fig. 3 Function of LXR α -ABCA1 axis is impaired in phenotypically switched VSMCs. **a** Heatmap of proteomic analysis between synthetic and contractile VSMCs. **b** Analysis of KEGG pathway enriched by differentially expressed genes of proteomic analysis between synthetic and contractile VSMCs. **c** The mRNA expression of SM22 α , LXR α , and ABCA1 in WT VSMCs treated with PDGF-BB for 0, 12, 24, and 48 h, respectively ($n = 3$). **d** Confocal microscopy images of LXR α distribution in WT VSMCs incubated with PDGF-BB for 0, 12, 24, and 48 h, respectively. Scale bar, 10 μ m. **e** Confocal microscopy images of LXR α and ACTA2 in arterial walls of WT mice after ligation for 0, 7, 14, and 28 days. Scale bar, 20 μ m. **f** Quantification of each lipid class in synthetic and contractile VSMCs. Lipid classes were expressed as μ mol per g protein. **g** Heatmap of CEs between synthetic and contractile VSMCs. **h** ORO staining of WT VSMCs transfected with or without Ad-GFP-SM22 α following with PDGF-BB and/or cholesterol treatment and quantification of positive ORO staining. Scale bar, 20 μ m ($n = 3$). **i** M-mode and Doppler echocardiography images obtained from aortic arch and outflow tract of *Sm22 α ^{-/-}* mice transfected with AAV-GFP ($n = 10$) and AAV-SM22 α ($n = 10$) fed Paigen diet for 12 weeks. A_s : outflow tract and aortic diameter in systole; A_d : outflow tract and aortic diameter in diastole. **j** Representative images of en face ORO-stained aortas and quantification of lesion areas ($n = 6$). **k** Representative immunofluorescence of LXR α (green) and quantification of cells with nuclear LXR α in the aortic sections from *Sm22 α ^{-/-}* mice transfected with AAV-GFP ($n = 4$) and AAV-SM22 α ($n = 4$) fed Paigen diet for 24 weeks. Scale bar, 10 μ m. Arrows indicated the distribution of LXR α . **l** Identification of SMC-derived foam cells within atherosclerotic lesion of *Sm22 α ^{-/-}* mice infected with AAV-GFP ($n = 4$) and AAV-SM22 α ($n = 4$) fed Paigen diet for 24 weeks by CD68 (blue), ACTA2 (red), and Bodipy (green). Scale bar, 25 μ m. Arrows indicated foam cells which were VSMCs-derived. Data and images are representative of at least three independent experiments. Data in **c** were analyzed by two-way ANOVA. Data in **f**, **h**, **j**, **k**, and **l** were analyzed by unpaired *t*-test. * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$; **** $p < 0.0001$.

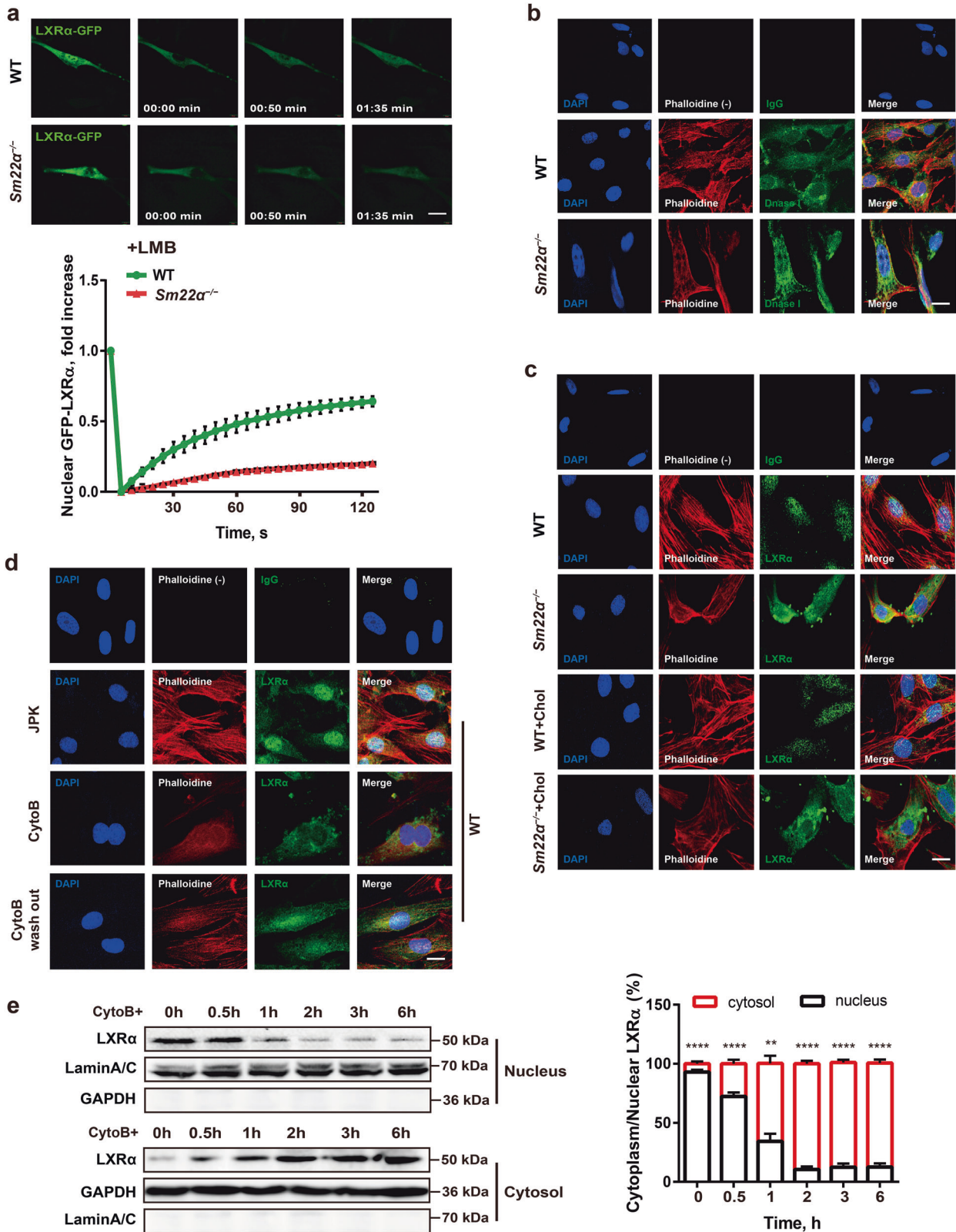
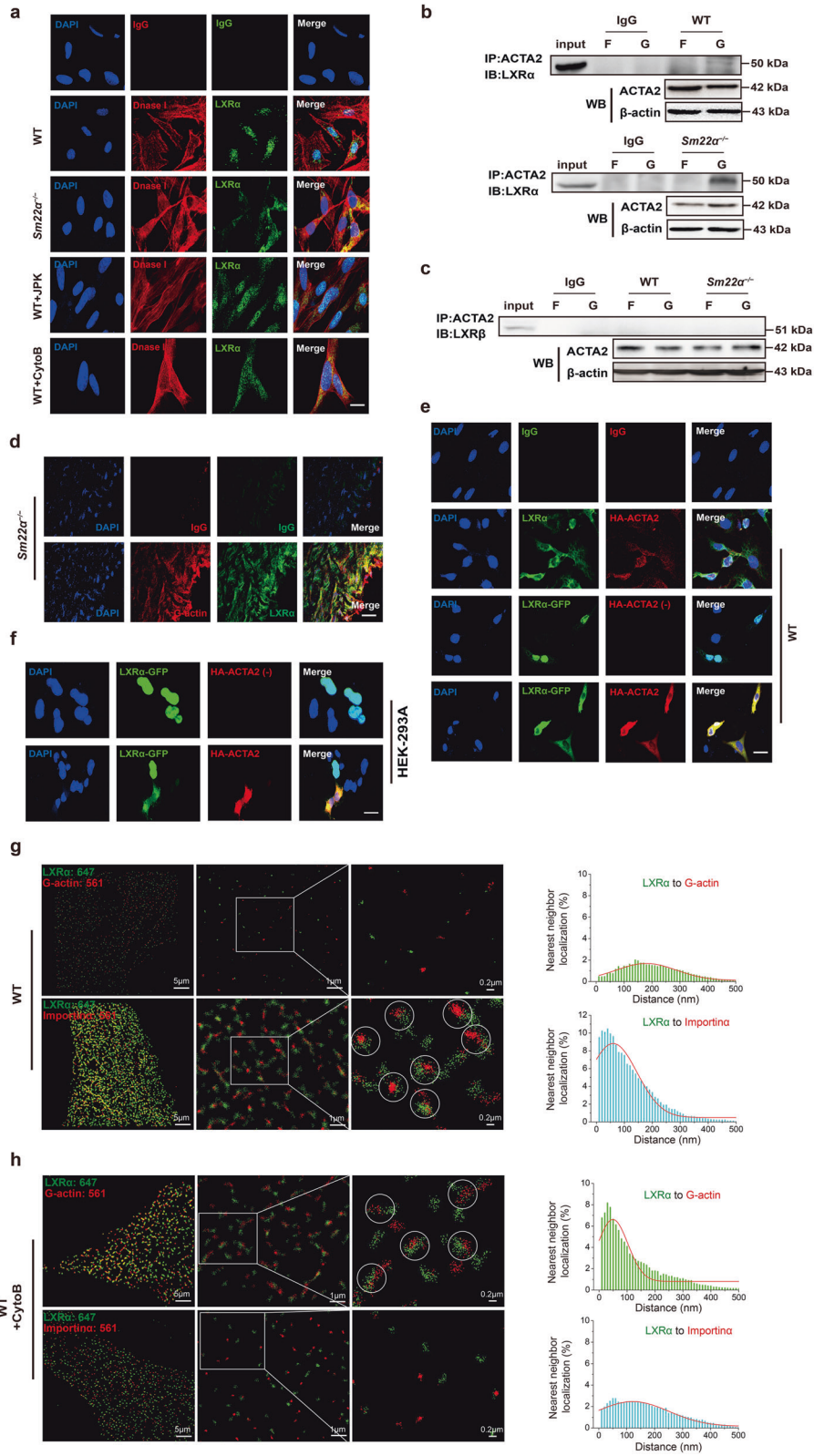


Fig. 4 Nuclear import of LXR α is regulated by actin dynamics. **a** Fluorescence recovery after photobleaching (FRAP) studies with LXR α -GFP to measure nuclear import. Cells were pretreated with LMB. Decreased accumulation of nuclear fluorescence indicates a lower rate of nuclear import of LXR α -GFP in *Sm22 α ^{-/-}* VSMCs relative to WT controls ($n = 25$). **b** Representative images of F-actin (phalloidin, red) and G-actin (Dnase I, green) in WT and *Sm22 α ^{-/-}* VSMCs. Scale bar, 10 μ m. **c, d** Representative images for F-actin (phalloidin, red) and LXR α (green) in WT and *Sm22 α ^{-/-}* VSMCs with cholesterol loading or not (**c**) and in WT VSMCs treated with JPK, CytoB and after CytoB washout (**d**). Scale bars, 10 μ m. **e** Western blot analysis of cytoplasmic and nuclear LXR α in WT VSMCs treated with CytoB at different time points ($n = 6$). Data and images represent at least three independent experiments. Statistical analyses, unpaired t -test and Kruskal–Wallis rank sum test. ** $p < 0.01$; *** $p < 0.001$; **** $p < 0.0001$.



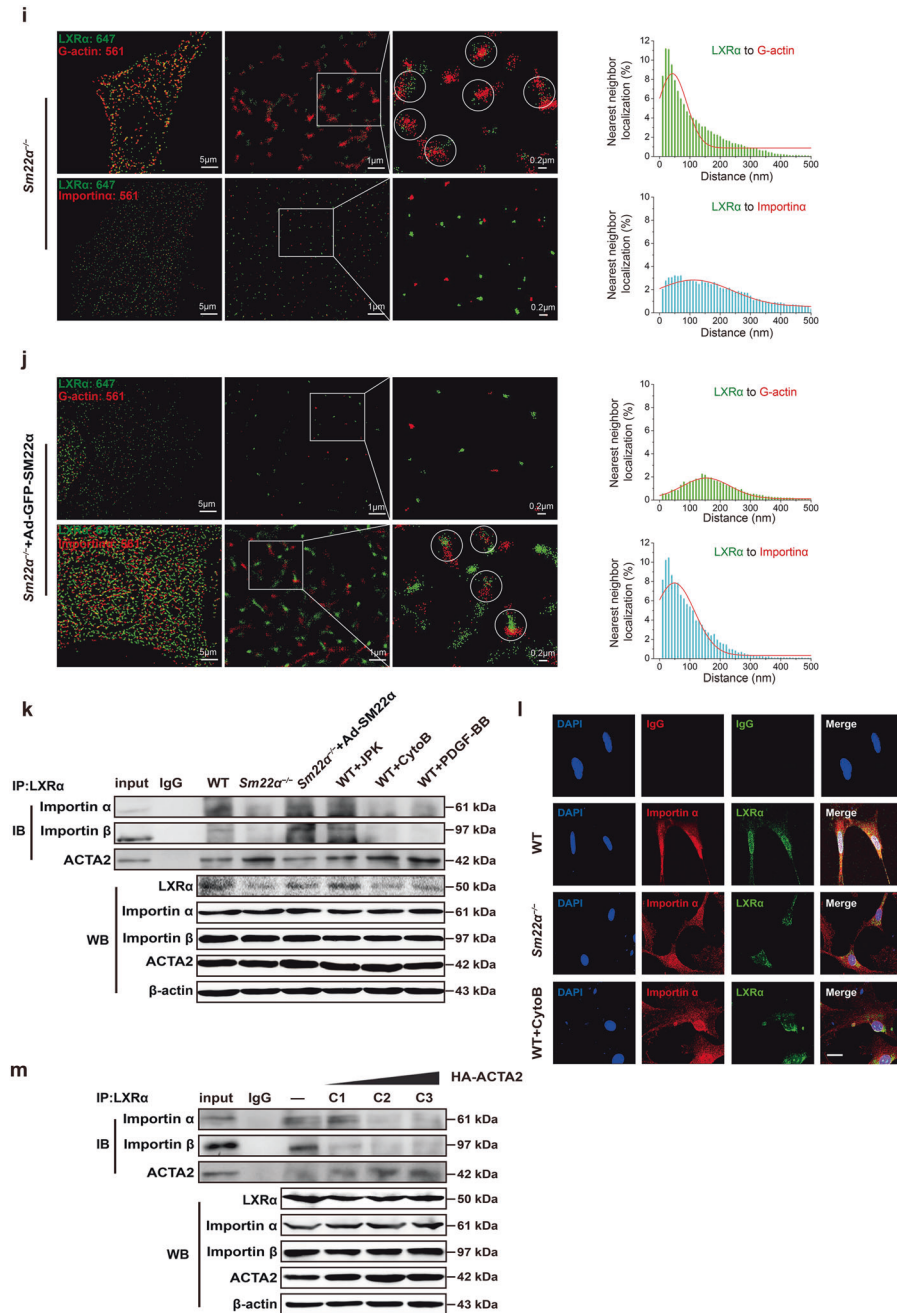


Fig. 5 G-actin interacts with and retains LXRα in the cytoplasm, blocking LXRα binding to Importin α. **a** Double immunofluorescence staining for G-actin (Dnase1, red) and LXRα (green) in WT VSMCs accompanied with treatment of JPK or CytoB and also in *Sm22α*^{-/-} VSMCs. Scale bar, 10 μm. **b, c** Co-immunoprecipitation of ACTA2 and LXRα (**b**) and LXRβ (**c**), respectively, in F- and G-actin fractions of WT and *Sm22α*^{-/-} VSMCs (*n* = 3). **d** Double immunofluorescence staining of G-actin (Dnase1, red) and LXRα (green) or IgG in atherosclerotic lesion in aortic wall of *Sm22α*^{-/-} mice. Scale bar, 20 μm. **e** Representative immunofluorescence staining for endogenous LXRα (green) and LXRα-GFP (green) in WT VSMCs transfected with HA-ACTA2 (red, stained by anti-HA antibody) or not. Scale bar, 15 μm. **f** Representative immunofluorescence staining for LXRα-GFP (green) and HA-ACTA2 (red, stained by anti-HA antibody) in HEK-293A cells. Scale bar, 10 μm. **g–j** Two-color STORM images and quantification of the co-localization degree between LXRα and G-actin as well as Importin α in WT VSMCs with (**h**) or without (**g**) CytoB treatment and *Sm22α*^{-/-} VSMCs with (**j**) or without (**i**) Ad-GFP-SM22α infection (*n* > 10). **k** Co-immunoprecipitation of LXRα and Importin α, Importin β or ACTA2 in WT and *Sm22α*^{-/-} VSMCs with or without JPK, CytoB, PDGF-BB, and Ad-GFP-SM22α treatment (*n* = 3). **l** Double immunofluorescence staining for Importin α (red) and LXRα (green) in WT and *Sm22α*^{-/-} VSMCs as well as CytoB-treated WT VSMCs. Scale bar, 15 μm. **m** Co-immunoprecipitation of LXRα and Importin α, Importin β or ACTA2 in WT VSMCs transfected with HA-ACTA2 of different concentration (*n* = 3). Data and images represent at least three independent experiments.

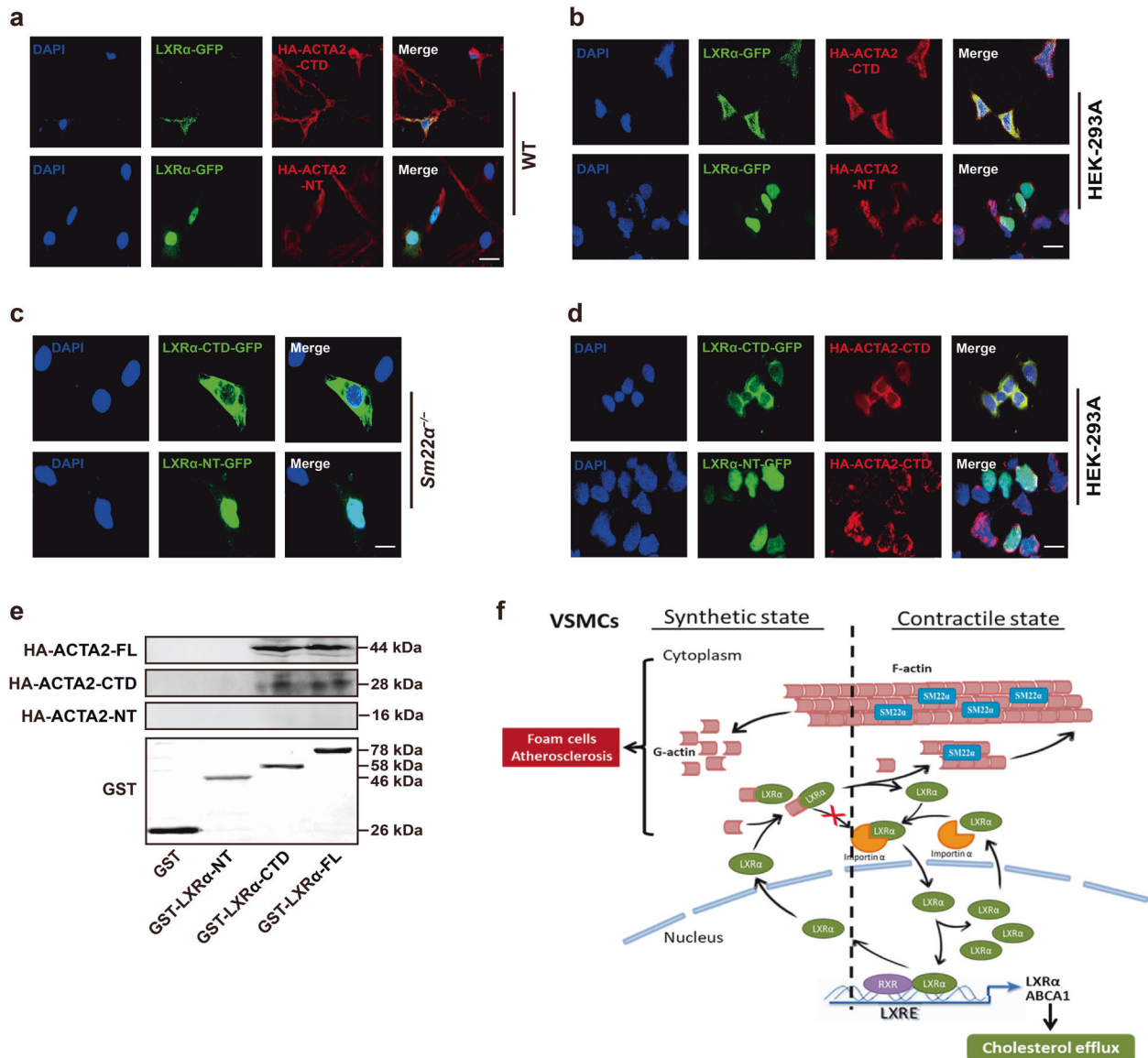


Fig. 6 The C-terminal domain mediates interaction between G-actin and LXRα. **a** Representative immunofluorescence staining for LXRα-GFP (green) in WT VSMCs transfected with HA-ACTA2-CTD (red) or HA-ACTA2-NT (red). Scale bar, 15 μm. **b** Representative immunofluorescence staining for LXRα-GFP (green) and HA-ACTA2-CTD (red) or HA-ACTA2-NT (red) in HEK-293A cells. Scale bar, 15 μm. **c** LXRα-CTD-GFP (green) or LXRα-NT-GFP (green) was transfected into *Sm22α*^{-/-} VSMCs. Scale bar, 10 μm. **d** LXRα (-CTD, -NT)-GFP (green) and HA-ACTA2-CTD (red) were co-expressed in HEK-293A cells. Scale bar, 10 μm. **e** Interaction of HA-ACTA2 (-FL, -CTD, -NT) and GST-LXRα (-FL, -CTD, -NT) proteins analyzed by *in vitro* pull-down assay ($n = 3$). **f** Schematic representation of a working model which SM22α inhibits VSMC-derived foam cell formation by blocking actin-LXRα signaling ameliorating atherosclerosis. Data and images represent at least three independent experiments.