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Optic Neuritis-Independent Retinal Atrophy in Neuromyelitis Optica Spectrum Disorder

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INTRODUCTION

Neuromyelitis optica spectrum disorder (NMOSD) is a rare immune-mediated disease of the central nervous system (CNS), associated, in the majority of cases, with autoantibodies directed against the astrocytic water channel aquaporin-4 (AQP4).(1) AQP4-IgG+ NMOSD presents predominantly with episodes of optic neuritis (ON) and longitudinally extensive

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transverse myelitis. Unlike other neuro-inflammatory diseases, such as multiple sclerosis (MS), where a progressive disease course has been well established, AQP4-IgG+ NMOSD is considered a primarily relapsing disorder and disease progression independent of attack-associated incidents is considered exceedingly rare.(1, 2) A limited number of studies have investigated the presence of ongoing disease activity independent of relapses in NMOSD and data are conflicting.

Cross-sectional studies utilizing retinal optical coherence tomography (OCT) have suggested that retinal ganglion cell damage may occur in NMOSD eyes in the absence of a history of ON, as evidenced by lower thickness of the inner retinal layers when compared to healthy controls (HC).(3–5) To date, few longitudinal studies have explored whether there is subclinical visual pathway involvement outside of ON incidents in NMOSD. While some studies have suggested that progressive visual evoked potentials (VEP) latency delay and progressive GCIPL atrophy may occur in NMOSD even in the absence of ON attacks, existing evidence is conflicting.(6–10) Utilizing OCT, the objective of our study was to examine whether AQP4-IgG+ NMOSD patients exhibit progressive retinal neuro-axonal loss, independently of ON attacks.

METHODS

Standard protocol approvals, registrations, and patient consents

The study was approved by the Johns Hopkins University Institutional Review Board. All participants provided written informed consent.

Study design and participants

Patients were recruited from the Johns Hopkins Neuromyelitis Optica Clinic between 2008 and 2020, and fulfilled the 2015 International Panel for Neuromyelitis Optica Diagnosis criteria for AQP4-IgG+ NMOSD (retroactively applied to patients recruited prior to 2015). (1) AQP4-IgG+ NMOSD patients underwent serial retinal imaging with OCT and visual function assessments at their clinical visits. HC were recruited by convenience sampling, and were invited for annual OCT scans.

Patients with a clinical history of ON less than six months prior to baseline were excluded, because acute ON events are expected to lead to significant inner retinal atrophy. Existing literature supports that the vast majority of GCIPL and pRNFL thinning, as well as maximal visual recovery, has already occurred by six months.(11–13) Data of subjects who developed ON during follow-up were censored at the last available scan prior to ON. Since chiasmal/ optic tract involvement is common in ON in AQP4-IgG+ NMOSD, both eyes were excluded or censored in the analyses, even in cases of apparent unilateral ON during follow-up.(1) Only eyes with at least six months of OCT follow-up were included.

Subjects with history of diabetes mellitus, uncontrolled hypertension, or other significant ophthalmological/neurological disorders were excluded from the study.

ОСТ

Retinal imaging was performed with spectral-domain OCT (Cirrus HD-OCT, Model 5000; Carl Zeiss Meditec, Dublin, CA), as previously described.(14) Briefly, peripapillary retinal nerve fiber layer (pRNFL) thicknesses were derived by the conventional Cirrus HD-OCT software. Macular ganglion cell+inner plexiform layer (GCIPL) was automatically segmented and thickness was calculated within an annulus centered at the fovea, with an internal diameter of 1mm and an external diameter of 5mm. The scans underwent rigorous quality control in accordance with the OSCAR-IB criteria; scans with signal strength below 7/10, with artifact, or with segmentation errors were excluded from the study.(15)

Visual Function

Monocular visual acuity (VA) was assessed using standardized retro-illuminated eye charts (Precision Vision, La Salle, IL) with habitual correction. High-contrast (100%) letter acuity (HCLA) was assessed with Early Treatment Diabetic Retinopathy Study (ETDRS) charts (at 4m), and low-contrast (2.5% and 1.25%) letter acuity (LCLA) with Sloan Letter charts (at 2m). The maximum score for each chart is 70 letters, corresponding to a Snellen visual acuity of 20/10.

Clinically significant VA worsening was defined as a decrease in monocular letter acuity 5 letters for HCLA, and 7 letters for LCLA, in accordance with prior studies in MS.(16, 17)

Statistical Methods

Statistical analyses were performed using Stata version 16 (StataCorp) and R version 4.0.2. Baseline comparisons were performed with mixed-effects linear regression models with subject-specific random intercepts. Rates of change of retinal layer thicknesses and letter acuity scores were analyzed using mixed-effects linear regression models, with subject-specific and eye-specific random intercepts and random slopes in time, using time from baseline visit as a continuous variable. All analyses were adjusted for age, sex, and race. Statistical significance was defined as p<0.05.

RESULTS

Cohort

32 AQP4-IgG+ NMOSD patients (61 eyes) met inclusion criteria (Figure E1) and we also included 48 HC (95 eyes), approximately matched to the NMOSD group for age, sex, and race. Demographics and clinical characteristics are summarized in Table 1. Median OCT follow-up was 4.3 years in NMOSD and 4.0 years in HC.

For a subset of NMOSD patients (n=30) and HC (n=42), VA assessments were available cross-sectionally. Additionally, 20 NMOSD patients (63%) had available serial VA assessments (median follow-up 3.0 years, IQR: 1.0 - 7.6).

Baseline OCT analysis

At baseline, GCIPL and pRNFL thicknesses were markedly lower in NMOSD eyes with a prior ON history (NMOSD-ON) compared to HC (adjusted mean differences — GCIPL: -20.29μ m; pRNFL: -27.80μ m; p<0.001 for both; Table 2).

NMOSD eyes without a prior ON history (NMOSD-nonON) also exhibited significantly lower GCIPL thickness, albeit less pronounced relative to NMOSD-ON eyes, as compared to HC (adjusted mean difference: -5.38μ m, p<0.001). pRNFL thickness was also lower in NMOSD-nonON eyes compared to HC, but this difference did not attain statistical significance (adjusted mean difference: -4.06μ m, p=0.09; Table 2).

Longitudinal OCT analysis

Mean annualized rates of change in GCIPL and pRNFL thicknesses in NMOSD and HC participants are reported in Table 3 and individual eye trajectories are shown in Figure 1. NMOSD patients exhibited faster rates of pRNFL thinning compared to HC (-0.45μ m/ year vs. -0.21μ m/year, β = -0.25μ m/year, 95%CI:-0.45 to -0.05, p=0.014). Similarly, GCIPL thinning was also accelerated in NMOSD patients (-0.29μ m/year vs. -0.20μ m/year, β = -0.09μ m/year, 95%CI:-0.17 to 0.00, p=0.05). These differences correspond to 119% faster pRNFL and 45% faster GCIPL thinning in NMOSD compared to HC eyes.

This was driven by faster pRNFL and GCIPL thinning in NMOSD-nonON eyes compared to HC (pRNFL: -0.64μ m/year vs. -0.21μ m/year, β = -0.43μ m/year, 95%CI:-0.67 to -0.19, p<0.001; GCIPL: -0.35μ m/year vs. -0.20μ m/year, β = -0.15μ m/year, 95%CI:-0.25 to -0.05, p=0.005). In contrast, NMOSD-ON eyes did not demonstrate significant differences in rates of pRNFL or GCIPL thinning in comparison to HC (pRNFL: -0.28μ m/year vs. -0.21μ m/year, β = -0.07μ m/year, 95%CI:-0.31 to 0.16, p=0.53; GCIPL: -0.21μ m/year vs. -0.20μ m/year, β = -0.01μ m/year, 95%CI:-0.11 to 0.10, p=0.90). However, when restricting analyses to NMOSD-ON eyes with baseline pRNFL thickness 70µm (lower limit of pRNFL thickness range in the HC cohort; n=12 NMOSD-ON eyes), we found faster rates of pRNFL thinning as compared to HC (-0.57μ m/year vs. -0.21μ m/year, β = -0.36μ m/year, 95%CI:-0.57 to -0.21, p=0.034), consistent with a potential floor effect. The rate of GCIPL thinning was also faster in NMOSD-ON eyes with baseline pRNFL thickness 70µm compared to HC, although this finding was not statistically significant (-0.21μ m/year vs. -0.32μ m/year, β = -0.11μ m/year, 95%CI: -0.27 to 0.05, p=0.16).

In order to exclude effects due to chiasmal or optic tract involvement in patients with a prior history of contralateral ON, we compared rates of retinal atrophy between NMOSD patients who had never experienced ON in either eye (n=11 patients) and HC and the results were similar (pRNFL: -0.62μ m/year vs. -0.21μ m/year, β = -0.41μ m/year, 95%CI:-0.68 to -0.15, p=0.002; GCIPL: -0.31μ m/year vs. -0.20μ m/year, β = -0.11μ m/year, 95%CI:-0.22 to 0.00, p=0.05).

Finally, we explored the effects of non-ON relapses during follow-up on rates of pRNFL and GCIPL thinning. Ten patients had relapses during follow-up (9 transverse myelitis, 1 area postrema syndrome). Patients with relapses did not exhibit significant differences in rates of GCIPL (-0.27μ m/year vs. -0.32μ m/year, $\beta=0.05\mu$ m/year, 95% CI:-0.10 to 0.20, p=0.51)

or pRNFL thinning (-0.44 μ m/year vs. -0.51 μ m/year, β =0.08 μ m/year, 95%CI:-0.28 to 0.43; p=0.67), compared to those who were clinically stable.

Baseline VA analysis

At baseline, as expected, HCLA and LCLA were markedly worse in NMOSD-ON eyes compared to HC (adjusted mean differences — HCLA: -23.00 letters; 2.5% LCLA: -23.95 letters; 1.25% LCLA: -16.97 letters, p<0.001 for all; Table 2).

HCLA did not differ between NMOSD-nonON and HC eyes (adjusted mean difference: -2.42 letters, p=0.10), however NMOSD-nonON eyes exhibited worse 2.5% and 1.25% LCLA compared to HC (adjusted mean differences: 2.5% LCLA: -8.15 letters; 1.25% LCLA: -8.66 letters, p<0.001 for both; Table 2).

Longitudinal VA analysis

Of the 61 NMOSD eyes included in this study, 40 eyes had serial visual function assessments. Of these 40 eyes, 6 eyes (15%) exhibited clinically significant HCLA worsening by the end of the follow-up period, 4 eyes (10%) exhibited clinically significant 2.5% LCLA worsening and 5 eyes (13%) exhibited clinically significant 1.25% LCLA worsening. Nine eyes (23%) exhibited HCLA and/or LCLA worsening.

Rates of GCIPL and pRNFL thinning were compared between eyes with clinically significant VA worsening by the end of the follow-up period and eyes with stable VA. We did not find significant differences in these rates between eyes with VA worsening compared to those with stable VA (GCIPL: -0.28μ m/year vs. -0.33μ m/year, β =0.05 μ m/year, 95%CI:-0.11 to 0.22, p=0.51; pRNFL: -0.63μ m/year vs. -0.51μ m/year, β = -0.12μ m/year, 95%CI:-0.55 to 0.32, p=0.6).

We also calculated the annualized rates of change of HCLA and LCLA eyes in NMOSD eyes, and did not observe a significant change in visual function over time (Table 4). There was no difference in the rate of change of HCLA and LCLA between NMOSD-ON and NMOSD-nonON eyes.

DISCUSSION

In this longitudinal study, we observed accelerated pRNFL and GCIPL thinning in AQP4-IgG+ NMOSD eyes clinically unaffected by ON. This finding suggests that subclinical progressive retinal neuro-axonal loss occurs in AQP4-IgG+ NMOSD. This has important implications for our understanding of the disease mechanisms of NMOSD, since it provides evidence that there is ongoing insidious disease activity independent of clinical relapses. Importantly, our results remained unaltered in sensitivity analyses that were restricted to patients who had never experienced ON in either eye, suggesting that our findings are not the consequence of a symptomatic ON event but they likely reflect a separate pathophysiologic process.

Our findings are in agreement with prior work by Oertel et al. reporting faster GCIPL thinning in AQP4-IgG+ NMOSD eyes, independently of ON attacks.(6) Furthermore, in

a multi-center longitudinal VEP study, an increase of P100 latencies and a decline of amplitudes was observed in AQP4-IgG+ NMOSD eyes without ON during follow-up.(7) Together, these findings support that the visual pathway may be a site of subclinical involvement in AQP4-IgG+ NMOSD. In our study, these phenomena appeared to be independent, not only of ON attacks, but also of other non-ON attacks, since pRNFL and GCIPL atrophy rates did not differ between patients who experienced relapses during follow-up and patients who were clinically stable. In a cohort of 27 NMOSD patients, Pisa et al. reported accelerated pRNFL thinning in patients with disease activity during follow-up, but not in stable patients.(18) In contrast, in the study by Oertel et al, patients who experienced attacks during follow-up had pRNFL thickening.(6) Further larger studies are likely needed to investigate the retinal phenomena that occur during inflammatory attacks and reconcile these conflicting results.

Interestingly, we observed progressive pRNFL and GCIPL atrophy in AQP4-IgG+ NMOSD eyes without a prior history of ON, but not in ON eyes. This is presumably due to a floor effect, since ON associated with AQP4-IgG+ NMOSD results in marked retinal ganglion cell and axonal loss which may limit detection of further decreases in pRNFL or GCIPL thicknesses.(19, 20) Notably, subgroup analyses restricted to NMOSD-ON eyes with baseline pRNFL thickness 70µm revealed faster rates of pRNFL thinning compared to HC, consistent with this hypothesis.

Even though the pathoetiology of the observed progressive retinal neuro-axonal loss could not be established in the current study, several hypotheses need to be considered. AQP4 is a water channel protein that is highly expressed in the retina by astrocytes and more abundantly by a unique type of glial cell, Müller cells. Müller cells regulate vital retinal homeostatic mechanisms, including release of neurotropic factors and degradation of glutamate. It is therefore conceivable that the observed retinal changes are a consequence of direct retinal damage due to anti-AQP4 antibody activity.(21) Interestingly, it has been previously reported that Müller cells are richly expressed in the foveal region, located in the center of the macula, and foveal thinning has been reported in AQP4-IgG+ eyes without a history of ON; it has been postulated that this may represent the occurrence of a primary retinal astrocytopathy.(21, 22) In a pathological retinal study, AQP4-IgG+ NMOSD was characterized by loss of AQP4 immunoreactivity on Müller cells; it is likely that alterations in the dynamics of astrocyte and Müller cell function may be responsible for subsequent neurodegeneration.(23) Alternatively, our findings may be the result of subclinical involvement of the anterior or posterior visual pathway. A study by Hokari et al. reported pathological evidence of optic nerve involvement in AQP4-IgG+ NMOSD eyes with no clinical ON attacks.(23) Moreover, gadolinium enhancement of the optic nerve has been reported at the time of attacks involving other anatomical locations, in the absence of visual symptoms.(24) Retrograde trans-synaptic degeneration due to involvement of the posterior visual pathway is another consideration. This is a less likely explanation however, since optic radiation (OR) lesions are not as common in NMOSD relative to MS.(25) Moreover, development of new asymptomatic brain lesions in clinically stable AQP4-IgG+ NMOSD patients appears to be rare.(26) Furthermore, we did not observe any cases of homonymous GCIPL thinning (which would be expected to be observed in unilateral posterior visual pathway involvement), although this may be difficult to assess

in the presence of pre-existing optic neuropathy and/or bilateral posterior visual pathway involvement.

Our VA findings are also of interest, since we found that NMOSD-nonON eyes had worse performance in LCLA charts compared to HC. This suggests that some level of vision impairment may be present in AQP4-IgG+ NMOSD eyes, even without a clinical history of ON, and while the etiology of this finding is not clear, considerations include prior subclinical ON and/or primary retinal astrocytopathy, as outlined above. We also found that 23% of NMOSD eyes had worsening visual function during follow-up, based on cut-offs that have been previously proposed for MS eyes.(16) However, rates of retinal thinning did not differ between eyes with VA worsening compared to those with stable VA and, on a group level, we did not observe a significant change in visual function over time. Even though this finding may suggest that there is no direct correlation between OCT measures and functional outcomes, this negative finding should be interpreted with caution, given our relatively small sample size and the fact that VA assessments were available for only a subset of our cohort. Alternatively, this result may suggest that OCT is a more sensitive indicator of anterior visual pathway involvement and retinal neuro-axonal loss may precede visual deficits.

One limitation of our study is the relatively small sample size, which is expected given the rarity of the studied condition. Furthermore, longitudinal visual acuity data were not available for the majority of the healthy control participants. It is likely that our study was underpowered to detect potential associations between neuro-axonal loss and vision loss, due to insufficient follow-up time and/or sample size. Moreover, our study was retrospective and brain and optic nerve MRI scans were not routinely performed in all patients. Therefore, we could not assess for the presence of lesions along the visual pathway. Finally, the vast majority of participants were on treatment with rituximab during follow-up, and we were therefore unable to investigate any potential associations between different disease-modifying treatments and OCT measures. To conclude, our findings support that subclinical progressive retinal axonal loss may occur in AQP4-IgG+ NMOSD. Further studies are warranted to validate these findings and to elucidate their clinical relevance, since the presence of insidious pathology of the visual system has important implications for our understanding of the disease mechanisms of NMOSD and for monitoring treatment response.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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Jerry Prince is a founder of Sonovex, Inc. and serves on its Board of Directors. He has received consulting fees from JuneBrain LLC and is PI on research grants to Johns Hopkins from Biogen.

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Shiv Saidha has received consulting fees from Medical Logix for the development of CME programs in neurology and has served on scientific advisory boards for Biogen, Genzyme, Genentech Corporation, EMD Serono, and Celgene. He is the PI of investigator-initiated studies funded by Genentech Corporation and Biogen, and received support from the Race to Erase MS foundation. He has received equity compensation for consulting from JuneBrain LLC, a retinal imaging device developer. He is also the site investigator of a trial sponsored by MedDay Pharmaceuticals.

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Figure 1.

Longitudinal changes in pRNFL and GCIPL thickness

Longitudinal trajectories of GCIPL and pRNFL thickness for individual eyes are shown, separately for healthy controls, AQP4-IgG+ NMOSD eyes without a history of ON (NMOSD-nonON) and with a history of ON (NMOSD-ON). The thicker lines represent the average trajectory, as fitted by mixed-effects models. The y-axis range differs for the NMOSD-ON group (given lower GCIPL and pRNFL thicknesses), but the slopes are comparable across panels since the scales of the y-axes identical.

NMOSD: neuromyelitis optica spectrum disorder; ON: optic neuritis; GCIPL: ganglion cell

+ inner plexiform layer; pRNFL: peri-papillary retinal nerve fiber layer

Table 1.

Demographic and clinical characteristics at baseline

	нс	AQP4-IgG+ NMOSD	P-values
Participants, n	48	32	
Age in years, mean (SD)	40.5 (11.3)	45.5 (12.3)	0.063
Female, n (%)	38 (79%)	27 (84%)	0.77
Race, n (%)			0.50
Caucasian-American	29 (60%)	15 (47%)	
African-American	18 (38%)	16 (50%)	
Asian-American	1 (2%)	1 (3%)	
Relapse history, n (%)	-		-
ON & TM		13 (41%)	
ON & TM & Brainstem attack		6 (19%)	
Isolated TM		8 (25%)	
Isolated ON		2 (6%)	
Brainstem attack & TM		3 (9%)	
Treatment during follow-up, patient-years (%)	-		-
No treatment		11.3 (8.7%)	
Rituximab		86.3 (66.7%)	
Mycophenolate Mofetil		15.9 (12.3%)	
Azathioprine		4.7 (3.6%)	
Eculizumab		3.8 (2.9%)	
Eculizumab + Mycophenolate Mofetil		6.0 (4.6%)	
Inebilizumab		1.4 (1.1%)	
Eyes with ON history, n (%)	-	34 (55%) ^a	-
Disease duration; median (IQR)	-	6 (1–9)	-
Follow-up, median (IQR)	4.0 (1.8 - 7.5)	4.3 (2.6 - 7.5)	0.89

^{*a*}8 NMOSD-ON eyes had microcystic macular pathology (MMP)

HC: healthy controls; AQP4-IgG+ NMOSD: aquaporin-4-IgG seropositive Neuromyelitis Optica Spectrum Disorder; SD: standard deviation; TM: transverse myelitis; ON: optic neuritis; IQR: interquartile range

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Table 2.

Cross-sectional OCT measures and visual acuity scores in AQP4-IgG+ NMOSD and HC eyes and comparisons between groups

				HC vs. NMOSD-nor	NOI	HC vs. NMOSD-O	z
	HC (n=95 eyes)	NMOSD-nonON (n=30 eyes)	NMOSD-ON (n=31 eyes)	Difference (95% CI) ^a	P-value	Difference (95% CI) ^a	P-value
OCT measures, µm, mean (SD)							
GCIPL	76.9 (5.2)	71.6 (4.8)	56.7 (8.2)	-5.38 (-8.16 to -2.60)	<0.001	-20.29 (-23.37 to -17.20)	<0.001
pRNFL	93.9 (9.2)	91.2 (8.2)	67.3 (13.1)	-4.06 (-8.78 to 0.66)	0.09	-27.80 (-32.62 to -22.98)	<0.001
Letter acuity scores b , median (IQR)							
100% contrast (HCLA)	61.5 (59–65)	59 (51.5–63)	44.5 (7–54)	-2.42 (-5.24 to 0.40)	0.10	-23.00 (-28.92 to -17.07)	<0.001
2.5% contrast (LCLA)	34 (29–38)	24 (16–33)	0 (0-16.5)	-8.15 (-12.15 to -4.15)	<0.001	-23.95 (-27.93 to -19.96)	<0.001
1.25% contrast (LCLA)	20 (14–25)	6.5 (5–18)	0 (0–1.5)	-8.66 (-13.24 to -4.08)	<0.001	-16.97 (-20.90 to -13.05)	<0.001
2							

Derived from mixed effects linear regression models, including age, sex and race

b Available for 42 HC and 30 NMOSD patients

OCT: optical coherence tomography; AQP4: aquaporin-4; NMOSD: Neuromyelitis Optica Spectrum Disorder; HC: healthy controls; ON: optic neuritis; CI: confidence interval; SD: standard deviation; GCIPL: ganglion cell + inner plexiform layer; pRNFL: peri-papillary retinal nerve fiber layer; IQR: interquartile range; HCLA: high-contrast letter acuity; LCLA: low-contrast letter acuity Author Manuscript

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		Annualized Rate	, of Change, μm/year		HC vs. NMOSI		HC vs. NMOSD-no	NOn	HC vs. NMOSD-	NO
	HC (n=95 eyes)	NMOSD (n=61 eyes)	NMOSD-nonON (n=30 eyes)	NMOSD-ON (n=31 eyes)	Difference in Annualized Rate of Change $(95\% \text{ CI})^{a}$, $\mu m/year$	P-value	Difference in Annualized Rate of Change (95% CI) ^{<i>a</i>} , µm/year	P-value	Difference in Annualized Rate of Change (95% CI) ^d , μm/year	P-value
GCIPL	-0.20 (-0.25 to -0.15)	-0.29 (-0.36 to -0.22)	-0.35 (-0.44 to -0.26)	-0.21 (-0.30 to -0.12)	-0.09 (-0.17 to 0.00)	0.05	-0.15 (-0.25 to -0.05)	0.005	-0.01 (-0.11 to 0.10)	06'0
pRNFL	-0.21 (-0.33 to -0.08)	-0.45 (-0.61 to -0.30)	-0.64 (-0.86 to -0.43)	-0.28 (-0.48 to -0.07)	-0.25 (-0.45 to -0.05)	0.014	-0.43 (-0.67 to -0.19)	<0.001	-0.07 (-0.31 to 0.16)	0.53
a a										

^dDerived from mixed effects linear regression models, including age, sex and race

AQP4: aquaporin-4; NMOSD: Neuromyelitis Optica Spectrum Disorder; HC: healthy controls; ON: optic neuritis; CI: confidence interval; GCIPL: ganglion cell + inner plexiform layer; pRNFL: peri-papillary retinal nerve fiber layer

Table 4.

Annualized change in visual acuity in AQP4-IgG+ NMOSD eyes

	Annu	alized Rate of Change, lett	ers/year	NMOSD-nonON vs. NMOS	D-ON
	NMOSD (n=40 eyes)	NMOSD-nonON (n=20 eyes)	NMOSD-ON n=20 eyes)	Difference in Annualized Rate of Change 95% CIJ ^a , letters/year	P-value
100% contrast (HCLA)	-0.04 (-0.38 to 0.29)	-0.10 (-0.54 to 0.33)	0.03 (-0.51 to 0.57)	0.13 (-0.56 to 0.83)	0.71
2.5% contrast (LCLA)	0.08 (-0.39 to 0.55)	-0.12 (-0.71 to 0.48)	0.20 (-0.51 to 0.91)	0.31 (-0.62 to 1.24)	0.51
1.25% contrast (LCLA)	0.02 (-0.31 to 0.34)	0.12 (-0.30 to 0.54)	-0.08 (-0.59 to 0.42)	-0.20 (-0.86 to 0.45)	0.55

 $^{a}\mathrm{Derived}$ from mixed effects linear regression models, including age, sex and race

AQP4: aquaporin-4; NMOSD: Neuromyelitis Optica Spectrum Disorder; ON: optic neuritis; CI: confidence interval; HCLA: high-contrast letter acuity; LCLA: low-contrast letter acuity