RESEARCH Open Access

PAX7 gene polymorphism in muscular temporomandibular disorders as potentially related to muscle stem cells



Valquiria Quinelato^{1*}, Letícia Ladeira Bonato¹, Alexandre Rezende Vieira², José Mauro Granjeiro³, Karla Menezes⁴, Radovan Borojevic⁴, Priscila Ladeira Casado¹, Jose Albuquerque Calasans-Maia¹ and Ricardo Tesch⁴

Abstract

Background: Temporomandibular disorders (TMD) are a group of painful and debilitating disorders, involving the masticatory muscles and/or the temporomandibular joint (TMJ). Chronic TMD pain can be associated with genetic changes in the key muscle development genes.

Objective: To evaluate the association between polymorphisms in the *PAX7* (paired box 7) gene and masticatory myalgia in patients with temporomandibular disorders (TMD).

Materials and methods: This is a case-control study. Patients with TMD were divided into two groups: (a) presence of muscular TMD (n = 122) and (b) absence of muscular TMD (n = 49). Genomic DNA was obtained from saliva samples from all participants to allow for genotyping single nucleotide polymorphisms in *PAX7* (rs766325 and rs6659735). Over-representation of alleles was tested using chi-square or Fisher's exact tests. Values of p < 0.05 were considered to be statistically significant.

Results: Individuals without muscular TMD were less likely to have the *PAX7* rs6659735 GG genotype (p = 0.03). No associations were found for *PAX7* rs766325.

Conclusions: Alterations in *PAX7* may influence muscular pathophysiology and individuals with TMD and the rs6659735 homozygous genotype (GG) are seemingly associated with muscular involvement of the disorder. No associations were found in the region rs766325.

Keywords: Chronic pain, Temporomandibular joint dysfunction syndrome, Myalgia, Polymorphism genetic

Background

Temporomandibular disorders (TMD) are a group of painful and debilitating disorders, involving the masticatory muscles and/or the temporomandibular joint (TMJ). Among the pain-suffering TMD patients, 50 to 70% present the diagnosis of masticatory myalgia [1],

a heterogeneous pathologic condition established as a result of phenotypes, derived from a genetic mosaic, that interact with environmental factors [2].

Oral parafunction habits, such as clenching teeth during waking time, are the major clinical orofacial characteristics that predict TMD incidence [3]. During waking hours, the sustained tonic activity of the masseter muscle was significantly higher in the group of patients with orofacial pain history [4]. Experimental maximal voluntary clenching of the masseter was reported to induce nearly a five-fold reduction of oxygen saturation in the group with

Braga St, 28 – Center, Niterói, RJ 24020-140, Brazil Full list of author information is available at the end of the article

BMC

© The Author(s) 2021. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

^{*}Correspondence: valquiriaquinelato@yahoo.com.br 1 School of Dentistry, Universidade Federal Fluminense, Mario Santos

the high parafunction as compared to the low parafunction group, indicating that apparently healthy individuals at risk for TMD can have abnormalities in the masseter oxygenation [5]. Structural abnormalities in the masticatory muscles may be also involved, as reported in a study of two independent cohorts that compared chronic muscular TMD patients to controls [6]. Repair and regeneration of the attained tissues may be thus required, involving mobilization and activation of the resident muscle progenitor cells.

Recent studies on muscle functions have proposed that maintenance of the TMD chronic pain can also depend upon the patient's genetic profile and the presence of locally produced or circulating tissue factors, which may result in chronic inflammation of muscles [7]. There is direct evidence for increased levels of the proinflammatory cytokines in the skeletal muscle performing low force tasks, suggesting that chronic repetitive activities could initiate a chronic inflammatory response in the working muscles [8]. While acute inflammation can result in muscle edema, a chronic inflammation can result in muscle destruction [9]. Hypo-oxygenation of muscles suffering chronic inflammation may increase tissue damage, predisposing the muscle to further injury, thus minimizing the probability of adequate repair [10].

While many cells are involved in chronic inflammation, macrophages, through their association with skeletal muscle stem cell, satellite cell (SC) activation, may be one of the major players in the control of injury and impaired muscle regeneration. An imbalance of M1 and M2 macrophages may lead to impaired SC activation in skeletal muscle regeneration after damage. Therefore, strategies reverting chronic inflammation into a pro-regenerative reaction appear to be appropriate. Recent studies suggest that muscular manual acupuncture produces a phenotypic switch in macrophages and increases IL-10 concentrations in muscle, reducing pain and inflammation [11]. Conversely, the manual massage facilitates membrane permeability, which can be associated with an increase in satellite cell number [12].

Satellite cells (SC), the stem cells of skeletal muscle, are required for skeletal muscle recovery. Their functionality is modulated by intrinsic signaling pathways as well as by their interactions with the stem cell niche [13]. Following muscle injury, SC are activated, proliferate extensively, and ultimately differentiate into myoblasts, fusing with existing fibers or other SC to form new myofibers [14]. In healthy individuals, expansion of the SC pool was identified in the cervical muscles after heavy resistance training [15], as well as low-force muscle loads [16]. Repetitive muscle overloading may also result in increased inflammatory and myogenic activity, and painful muscle fibers may require higher numbers of SC than muscle fibers not

exposed to this recruitment. Craniofacial muscles have a higher proportion of SC as compared to other skeletal muscles, which may increase their capacity for regeneration in response to tissue damage under normal conditions [17].

At the lesion site, growth factors are required for activating SC. Expression of the transcription factor *PAX7* (paired box 7) is fundamental to the control of muscle cell differentiation [18, 19]. Quiescent resident SC expressing *PAX7* can be induced to migrate into the lesion site, then becoming proliferative and expressing, in addition to *PAX7*, the myogenic regulatory factor, *MyoD* (myogenic differentiation). The final differentiation of these SC into myoblasts is marked by reduction of *PAX7* that is substituted progressively by an increased *MyoD* expression. The fusion of differentiated myoblasts generates new myofibrils or repairs the damaged ones [20].

Reduced *MyoD* expression and the maintenance of a high *PAX7* expression is required for the maintenance and self-renewal of SC [21]. When the SC population is activated, its fraction undergoes a terminal differentiation while the other one remains in the pool of quiescent satellite cells, which can be induced again to proliferate, to be activated and to participate in regeneration and control of inflammation. The size of this resident pool of SC and its capacity to attend the local demands may control the response to injury in qualitative and quantitative terms, and this is known to be regulated by the *PAX7* activity [22].

These data were the basis of the here-proposed hypothesis that polymorphisms in the *PAX7* gene could modulate the activity of this gene to influence the regenerative role played by SC in the muscular environment. We believe that this genetic pattern may influence the development and chronic persistence of muscle disorders, including the masticatory ones. However, more recent studies indicate that the involved mechanisms are almost exclusively attributed to facilitation, peripheral and central, in processing pain. As far as we are aware, the presence of polymorphisms in genes associated with muscle tissue embryogenesis and tissue regeneration processes have not yet been investigated with respect to the development of masticatory myalgia.

The objective of the present study was to test the hypothesis that polymorphisms in the *PAX7* gene are associated with the presence of masticatory myalgia in patients with TMD.

Materials and methods

The present clinical study is descriptive and cross-sectional, being evaluated and approved by the Research Ethics Committee of the Antônio Pedro University Hospital of the Fluminense Federal University on September

25, 2016 (protocol number 1.744.837). All of the subjects consented freely to participate in this project. Informed consent forms were received and signed by the participants before the research was conducted. All methods were carried out in accordance with relevant guidelines and regulations. The design development of this study followed the recommendations of *Strengthening the Reporting of Observational Studies in Epidemiology* (STROBE) [23].

The study included patients attended at the Fluminense Federal University (18 to 65 years) as previously described [24]. They were randomly selected over a period of 6 months. Exclusion criteria were the history of macro trauma and/or surgery in the temporomandibular region, diagnosis of rheumatoid arthritis, fibromyalgia, or other types of systemic musculoskeletal diseases, and/or previous treatment for TMD. The inclusion criteria were the presence of one or multiple TMD diagnoses.

All participants were clinically examined by the same evaluator (co-author L.L.B.), using the Research Diagnostic Criteria for Temporomandibular Disorders (RDC/TMD) – Axis I [25], validated for the physical diagnosis of TMD, and enabling classification of participants in one of the following diagnostic subgroups: (0) absence of TMD; (1) myofascial pain; (2) changes in the position of the articular disc; (3) painful and/or degenerative TMJ conditions. This process occurred in a non-mutually exclusive manner, allowing each participant to belong to more than one diagnostic sub-group. Participants were then grouped into three groups: (a) control (absence of TMD); (b) with muscular TMD; (c) without muscular TMD.

The decision of including patients with absence of TMD as a control group assumed that they were probably not strongly subjected to the common risk factors for the development of these disorders, such as muscular and joint overloading by oral parafunctional habits. This is not a study of TMD risk, but a study of muscular involvement predisposition in patients with TMD.

Genomic DNA was obtained from saliva samples from all participants, as previously described [26]. The concentration and purity of the DNA were analyzed using the NanoDrop® spectrophotometer (Thermo Scientific, Wilmington, DE, USA). All samples had to present an A260 nm / A280 nm ratio higher than 1.9.

Two polymorphisms of a single nucleotide (SNP) in the *PAX7* gene (rs766325, rs6659735) were selected, considering the linkage disequilibrium and gene structure relationships. These SNPs were previously identified and included in the database of the National Center for Biotechnology Information (http://www.ncbi.nlm.nih.gov/SNP/), with the lowest allele frequency having to be > 012. All procedures followed the recommendations of

the Strengthening the Reporting of Genetic Association Studies (STREGA) [27].

For a better understanding, the methodology steps are illustrated in Fig. 1.

Data processing and statistical analyses were done using STATA 12.0 (Stata Corp., College Station, TX, USA). The sample size included the spontaneous demand of patients over 6 months, respecting the inclusion criteria. Using data from a previous study under conditions of a 95% confidence interval and 90% power of test, the sample was calculated considering a 40% prevalence of TMD [28]. Considering a loss of 5%, the estimated number to produce adequate statistical power was 200 individuals. Since the main inclusion criterion of the sample was the presence of TMD regardless of the type, we were able to obtain enough individuals. However, when the sample was stratified into different TMD subgroups, the number of individuals in each group was below the size of the calculated sample, therefore the differences between the groups were considered to be significant only after the statistical analysis using Fisher's exact test with an alpha of 0.05.

Differences between groups in the frequency of genotypes and alleles were analyzed using the chi-square test after assembly for the Hardy-Weinberg equilibrium. Statistical differences between groups were calculated using the chi-square or Fisher's exact tests. To calculate linkage disequilibrium and diplotypes, the ARLEQUIN software program was used (http://cmpg.unibe.ch/software/arlequin3/). Values of p < 0.05 were considered to be statistically significant, and the risks associated with individual alleles and genotypes were calculated as odds ratio (OR) with a 95% confidence interval (CI).

Results

From the 337 volunteers evaluated over the six-month period, after the adoption of the exclusion criteria, 325 were included in the study. A total of 154 (47.3%) did not have a TMD diagnosis, 122 (37.5%) patients had muscular TMD while 49 (15%) had only non-muscular TMD.

The group with muscular TMD consisted of 85 women and 37 men, and the group without muscular TMD consisted of 32 women and 17 men. The mean age found was 45.1 ± 12.3 years. There was no significant difference between gender and the risk of developing muscular $(p\!=\!0.17)$ or other type disorders $(p\!=\!0.71)$. The characteristics of the two polymorphisms studied in the PAX7 gene are presented in Table 1.

The integrity rate of the genomic DNA was 92.3%, with a high correspondence rate between replicate samples, indicating a highlevel of confidence in accurate and unbiased genotyping. The frequencies of the genotypes and alleles are shown in Table 2.

Inclusion criteria:

Volunteers of both sexes, between 18 and 65 years of age. (n=337)

Exclusion criteria:

No history of macro trauma of the TMJs, joint surgery, diagnosis of rheumatoid arthritis, systemic lupus erythematosus, fibromyalgia, other types of systemic joint diseases, and/or previous treatment for TMD.

(n=325)

Clinical diagnosis of TMD:

RDC/TMD - Axis I was applied and three groups were formed:

- control (no TMD diagnosed) (n=154)
- with muscle disorders (n=122)
- without muscle disorders (n=49)

Genotyping

Genomic DNA was obtained from saliva samples from all participants. All samples had to present an A260 nm / A280 nm ratio greater than 1.9.

Single nucleotide (SNP) in the PAX7 gene:

- rs766325
- rs6659735)

Fig. 1 Flow chart showing the inclusion of patients, groups and SNP in this study

Table 1 Characteristics of the PAX7 gene polymorphisms studied

Gene symbol	Gene name	SNP	Chromosome	Base pair position*	SNP type	Base change ^a		MAFb
						Major	Minor	
PAX7	paired box 7	rs766325	1	18,629,964	Upstream	А	G	0.48
		rs6659735		18,657,203	Intragenic	Α	G	0.26

SNP Single nucleotide polymorphism. / a Base change according to Applied Biosystems. / b MAF: minor allele frequency according to GenBank

The analysis of the *PAX7* rs6659735 marker showed a lower-representation of the GG genotype among individuals with non-muscular TMD, when compared to the muscular TMD group (p = 0.03) (Table 2).

Discussion

The present clinical study analyzed the possible associations of the polymorphisms observed in the gene coding *PAX7* transcription-factor with the presence and with clinical aspects of the muscular TMD. The results showed that the patients not bearing the GG genotype of

rs6659735 were apparently protected from the muscular involvement in the TMD disorders.

Being concerned with multiple testing, we avoided applying the strict Bonferroni correction, increasing the Type II error. Using this correction, we would have lowered the alpha to 0.025 (0.05/2). It has been previously shown that the known associations are missed when correction for multiple testing is implemented [29].

The *PAX7* rs766325 SNP was associated with craniofacial skeletal variations in patients with malocclusions [30]. This polymorphism was described in two studies,

Table 2 Distribution of the genotypes and alleles of the PAX7 gene

Gene	SNP	Genotypes	Control N = 154	With Muscular TMD N = 122	Without Muscular TMD N = 49	P-value* (OR;CI)		
						Control x With Muscular TMD	With Muscular TMD x Without Muscular TMD	
PAX7	rs766325	AA-AG-GG	40-53-52	32–34-42	16-14-17	0.7	0.86	
		AG + GG	105	76	31	0.82 (0.9 (0.5–1.5)	0.71 (0.8 (0.3-1.6)	
		Α	133	98	46	0.49 (0.9 (0.6–1.3)	0.64 (1.1 (0.7-1.8)	
		G	157	118	48			
	rs6659735	AA-AG-GG	73-58-11	64-42-8	32-14-0	0.75	0.03	
		AG + GG	69	50	14	0.53 (0.82 (0.5-1.3)	0.16 (0.5 (0.2-1.1)	
		А	204	170	78	0.55 (1.1 (0.7–1.7)	0.06 (1.9 (1-3.6)	
		G	80	58	14			

No significant associations were found in the gene haplotype analysis (Table 3)

Table 3 Analysis of diplotypes

Gene	Diplotype	Frequency		<i>P</i> -value		
		Control N = 154	With Muscular TMD N = 122	Without Muscular TMD N = 49	Control x With Muscular TMD	With Muscular TMD x Without Muscular TMD
PAX7- Ch1 (rs766325; rs6659735)	AA	0.43	0.43	0.47	=	-
	GA	0.27	0.31	0.36	0.53	0.5
	GG	0.26	0.23	0.14	0.58	0.08
	AG	0.02	0.01	0.01	0.76	0.75

which investigated their relationship with palate and lip clefts [31, 32]. The SNP is located upstream of the gene and does not influence formation of the protein. Conversely, the less common homozygous genotype of rs6659735 showed a suggestive effect on the muscular TMD.

The major functions of the PAX7 gene in skeletal muscle are control of the basal reserve population of resident muscular SC, and of their mobilization when attending to a need for regeneration after injury, as well as in the case of muscular volume increase in response to sustained efforts. These SC reside in the sub-fascial and perivascular niches within muscles, remaining in the phase zero of the cell cycle. They can be activated rapidly in response to injury [33]. In this case, SC reenter the cell cycle upon activation, migrate from their niche into the regenerating region and engage in formation and maintenance through a controlled renewal of the proliferating myoblast pool. These cells are subsequently engaged in their terminal differentiation into myocytes, which merge into preexisting damaged muscle fibers or fuse with other cells to generate new muscle fibers [21]. The cell regeneration requires the progressive interruption of the *PAX7* gene expression and its substitution by expression of *MyoD*, which is required for the myocyte differentiation.

Alternatively, depending upon intensity and duration of injuries, a portion of activated SC can re-enter into quiescence, return to their niche and again express PAX7. This is a self-renewal of the muscular SC pool, and it is required for a long-term maintenance of the basal SC pool that remains available for the new waves of a regenerative supply of myoblasts, when required. The PAX7 gene activity is thus in charge of the delicate controls within the cells that supply new myoblasts that engage into the myocyte terminal differentiation, and resident cells that remain available for subsequent waves of regenerative production of new myoblasts [34]. In chronic or repetitive aggressions of muscle tissue, the equilibrated PAX7 activity has to grant both the immediate and efficient tissue regeneration and the long-term maintenance of the resident SC pool providing regeneration for the future demands.

Our study does not supply information on the properties and activities of the *PAX7* gene in the context of the GG genotype polymorphism. Since it has clearly a long-term effect on the muscular involvement in the TMD disorder, we may raise the hypotheses of either decreased or increased responses of the *PAX7* gene activity to the tissue environment in the context of TMD, as compared to controls.

In the former case, the decreased operational activity of PAX7 in muscular SC populations, before or along their engagement into myoblast differentiation, will potentially lead to an increased or accelerated supply of PAX7-negative and MyoD-positive myocytes to be engaged in the muscle fiber repair. In acute lesions this can lead to a more rapid and efficient tissue response and regeneration. In chronic and repetitive lesions, we can expect an association with a low reposition or exhaustion of the resident SC pool, which requires a high level of PAX7 activity. Conversely, in the case of increased total activity of PAX7 within the muscular SC population, we can expect a somewhat delayed substitution of PAX7 by the MyoD expression, with a potential delay in myocyte differentiation. However, this can lead to a permanent increase of the resident SC pool, with an increased long-term capacity to respond to chronic or repetitive injuries, and an increased capacity for repair and regeneration.

The similar rationale may be applied to the question of the relationship with the inflammatory processes that are associated with TMD that can be one of the causes of the associated pain. The PAX-7 activity has limited direct relationship with mobilization and activity of inflammatory cell populations. However, muscle SC can interfere with maturing macrophages by insulin-like growth factor-2, inducing M1-M2 macrophage conversion and an overall anti-inflammatory protection [35]. A more robust resident SC population can thus have an increased basal anti-inflammatory capacity. The PAX-7 controls of the resident muscle SC population can potentially influence a broad set of tissue reactions to stress. The proposed hypotheses should be addressed in experimental in vitro and in vivo models and are the object of ongoing studies.

Conclusion

Alterations in the *PAX7* may influence muscular pathophysiology since individuals with TMD and the rs6659735 homozygous genotype (GG) seem to be associated with muscular involvement of the disorder. This result leads to the proposal of an unexplored stem cell related genetic predisposition to muscular TMD.

Abbreviations

PAX7: Transcription factor *PAX7* (paired box 7); TMD: Temporomandibular disorders; TMJ: Temporomandibular joint; SC: Satellite cells; SNP: Single nucleotide polymorphism.

Acknowledgements

The authors are indebted to all of the patients who agreed to participate in this project, and gratefully acknowledge the assistance of the Department of Oral Biology at the University of Pittsburgh School of Dental Medicine, and of the Clinical Research Unit of the University Hospital Antônio Pedro, Fluminense Federal University, Niterói.

Conflict of interest

All authors disclose no actual or potential conflict of interest including any financial, personal, or other relationships with other people or organizations that could inappropriately influence (bias) their work.

Informed consent

All participants included in this study provided informed consent.

Authors' contributions

All authors have made substantial contributions to all of the following: the conception and design of the study or acquisition of data (LLB, RT and VQ), analysis and interpretation of data (RT, JACM and PLC), drafting the article (LLB, KM and RB), revising it critically for important intellectual content (ARV and JMG), final approval of the version to be submitted (all authors).

Funding

Financial support of this work was provided by School of Medicine of Petropolis.

Availability of data and materials

The datasets generated or analyzed during the current study are available from the corresponding author upon reasonable request; SNPs were previously identified and included in the database of the National Center for Biotechnology Information (http://www.ncbi.nlm.nih.gov/SNP/), with the lowest allele frequency having to be >012.

Declarations

Ethics approval and consent to participate

This study was conducted according to the recommendations of the Research Ethics Committee of Antônio PedroUniversity Hospital of Fluminense Federal University (registration number 1.744.837).

Consent for publication

Not applicable.

Competing interests

All authors declare that they have no competing interests.

Author details

¹School of Dentistry, Universidade Federal Fluminense, Mario Santos Braga St, 28 – Center, Niterói, RJ 24020-140, Brazil. ²University of Pittsburgh School of Dental Medicine, Pittsburgh, USA. ³National Institute of Metrology, Quality and Technology, Rio de Janeiro, Brazil. ⁴School of Medicine of Petropolis, Petropolis, Brazil.

Received: 13 May 2021 Accepted: 7 October 2021 Published online: 17 November 2021

References

- Cairns BE. Pathophysiology of TMD pain-basic mechanisms and their implications for pharmacotherapy. J Oral Rehabil. 2010;37:391–410. https://doi.org/10.1111/j.1365-2842.2010.02074.x.
- Smith SB, Mir E, Bair E, Slade GD, Dubner R, Fillingim RB, et al. Genetic variants associated with development of TMD and its intermediate

- phenotypes: the genetic architecture of TMD in the OPPERA prospective cohort study. J Pain. 2013;14:T91–T101. https://doi.org/10.1016/j.jpain. 2013.09.004.
- Ohrbach R, Bair E, Fillingim RB, Gonzalez Y, Gordon SM, Lim PF, et al. Clinical orofacial characteristics associated with risk of first-onset TMD: the
 OPPERA prospective cohort study. J Pain. 2013;14:T33–50. https://doi.org/10.1016/i.jpain.2013.07.018.
- Mude AH, Kawakami S, Kato S, Minagi S. Properties of tonic episodes of masseter during waking and sleep in subjects with/without history of orofacial pain. J Prosthodont Res. 2018;62:234–8. https://doi.org/10. 1016/j.jpor.2017.09.003.
- Shah N, Melo L, Reid WD, Cioffi I. Masseter Deoxygenation in adults at risk for Temporomandibular disorders. J Dent Res. 2019;98:666–72. https:// doi.org/10.1177/0022034519837249.
- Moayedi M, Krishnamoorthy G, He PYT, Agur A, Weissman-Fogel I, Tenenbaum HC, et al. Structural abnormalities in the temporalis musculoaponeurotic complex in chronic muscular temporomandibular disorders. Pain. 2020;161:1787–97. https://doi.org/10.1097/j.pain.0000000000 001864.
- Jounger SL, Christidis N, Svensson P, List T, Ernberg M. Increased levels of intramuscular cytokines in patients with jaw muscle pain. J Headache Pain. 2017;18:30. https://doi.org/10.1186/s10194-017-0737-y.
- Elliott MB, Barr AE, Clark BD, Wade CK, Barbe MF. Performance of a repetitive task by aged rats leads to median neuropathy and spinal cord inflammation with associated sensorimotor declines. Neuroscience. 2010;170:929–41. https://doi.org/10.1016/j.neuroscience.2010.07.041.
- Cook JL, Purdam CR. Is tendon pathology a continuum? A pathology model to explain the clinical presentation of load-induced tendinopathy. Br J Sports Med. 2009;43:409–16. https://doi.org/10.1136/bjsm.2008. 051193.
- Fiorentino NM, Blemker SS. Musculotendon variability influences tissue strains experienced by the biceps femoris long head muscle during highspeed running. J Biomech. 2014;47:3325–33. https://doi.org/10.1016/j. ibiomech.2014.08.010.
- Da Silva MD, Bobinski F, Sato KL, Kolker SJ, Sluka KA, Santos AR. IL-10 cytokine released from M2 macrophages is crucial for analgesic and anti-inflammatory effects of acupuncture in a model of inflammatory muscle pain. Mol Neurobiol. 2015;51:19–31. https://doi.org/10.1007/ s12035-014-8790-x.
- Hunt ER, Confides AL, Abshire SM, Dupont-Versteegden EE, Butterfield TA. Massage increases satellite cell number independent of the age-associated alterations in sarcolemma permeability. Physiol Rep. 2019;7:e14200. https://doi.org/10.14814/phy2.14200.
- Schmidt M, Schüler SC, Hüttner SS, von Eyss B, von Maltzahn J. Adult stem cells at work: regenerating skeletal muscle. Cell Mol Life Sci. 2019;76:2559–70. https://doi.org/10.1007/s00018-019-03093-6.
- Hawke TJ. Garry DJ (2001) myogenic satellite cells: physiology to molecular biology. J Appl Physiol. 1985;91:534–51. https://doi.org/10.1152/jappl. 2001.91.2.534.
- Kadi F, Eriksson A, Holmner S, Butler-Browne GS, Thornell LE. Cellular adaptation of the trapezius muscle in strength-trained athletes. Histochem Cell Biol. 1999;111:189–95. https://doi.org/10.1007/s004180050348.
- Mackey AL, Holm L, Reitelseder S, Pedersen TG, Doessing S, Kadi F, et al. Myogenic response of human skeletal muscle to 12 weeks of resistance training at light loading intensity. Scand J Med Sci Sports. 2011;21:773– 82. https://doi.org/10.1111/j.1600-0838.2010.01178.x.
- McLoon LK, Thorstenson KM, Solomon A, Lewis MP. Myogenic precursor cells in craniofacial muscles. Oral Dis. 2007;13:134–40. https://doi.org/10. 1111/j.1601-0825.2006.01353.x.
- Seale P, Sabourin LA, Girgis-Gabardo A, Mansouri A, Gruss P, Rudnicki MA. Pax7 is required for the specification of myogenic satellite cells. Cell. 2000;102:777–86. https://doi.org/10.1016/s0092-8674(00)00066-0.
- Buckingham M, Relaix F. The role of Pax genes in the development of tissues and organs: Pax3 and Pax7 regulate muscle progenitor cell functions. Annu Rev Cell Dev Biol. 2007;23:645–73. https://doi.org/10.1146/ annurev.cellbio.23.090506.123438.
- Tedesco FS, Dellavalle A, Diaz-Manera J, Messina G, Cossu G. Repairing skeletal muscle: regenerative potential of skeletal muscle stem cells. J Clin Invest. 2010;120:11–9. https://doi.org/10.1172/JCI40373.

- Olguín HC, Pisconti A. Marking the tempo for myogenesis: Pax7 and the regulation of muscle stem cell fate decisions. J Cell Mol Med. 2012;16:1013–25. https://doi.org/10.1111/j.1582-4934.2011.01348.x.
- Crameri RM, Aagaard P, Qvortrup K, Langberg H, Olesen J, Kjær M. Myofbre damage in human skeletal muscle: effects of electrical stimulation versus voluntary contraction. J Physiol. 2007;583:365–80. https://doi.org/10.1113/jphysiol.2007.128827.
- Von Elm E, Altman DG, Egger M, Pocock SJ, Gotzsche PC, Vandenbroucke JP. The strengthening the reporting of observational studies in epidemiology (STROBE) statement: guidelines for reporting observational studies. Int J Surg. 2014;12:1495–9. https://doi.org/10.1016/j.ijsu.2014.07.013.
- Tesch RS, Bonato LL, Quinelato V, Casado PL, Vieira AR, Granjeiro JM, et al. Evaluation of genetic risk related to catechol-O-methyltransferase (COMT) and β2-adrenergic receptor (ADRB2) activity in different diagnostic subgroups of temporomandibular disorder in Brazilian patients. Int J Oral Maxillofac Surg. 2019;49:237–43. https://doi.org/10.1016/j.ijom.2019. 06027.
- Dworkin SF, LeResche L. Research diagnostic criteria for temporomandibular disorders: review, criteria, examinations and specifications, critique. J Craniomand Disord. 1992;6:301–55.
- Kuchler EC, Tannure PN, Falagan-Lotsch P, Lopes TS, Granjeiro JM, Amorim LM. Buccal cells DNA extraction to obtain high quality human genomic DNA suitable for polymorphism genotyping by PCRRFLP and real-time PCR. J Appl Oral Sci. 2012;20:467–71. https://doi.org/10.1590/s1678-77572012000400013.
- Little J, Higgins JP, Ioannidis JP, Moher D, Gagnon F, Von Elm E, et al. Strengthening the reporting of genetic association studies (STREGA): an extension of the STROBE statement. Hum Genet. 2009;125:131–51. https://doi.org/10.1007/s00439-008-0592-7.
- Plesh O, Adams SH, Gansky SA. Temporomandibular joint and muscle disorder (TMJMD)-type pain and co-morbid pains in a national US sample. J Orofacial Pain. 2011;25:190–8.
- Vieira AR, Mchenry TG, Daack-Hirsch S, Murray JC, Ml M. Candidate gene/ loci studies in cleft lip/palate and dental anomalies finds novel susceptibility genes for clefts. Genet Med. 2008;10:668–74. https://doi.org/10. 1097/qim.0b013e3181833793.
- da Fontoura CG, Miller SF, Wehby GL, Amendt BA, Holton NE, Southard TE, et al. Candidate gene analyses of skeletal variation in malocclusion. J Dent Res. 2015;94:913–20. https://doi.org/10.1177/0022034515581643.
- Sull JW, Liang KY, Hetmanski JB, Fallin MD, Ingersoll RG, Park J, et al. Maternal transmission effects of the PAX genes among cleft case–parent trios from four populations. Eur J Hum Genet. 2009;17:831–9. https://doi.org/10.1038/ejhg.2008.250.
- Guo Q, Li D, Meng X, Liu T, Shi J, Hao Y, et al. Association between PAX7 and NTN1 gene polymorphisms and nonsyndromic orofacial clefts in a northern Chinese population. Medicine (Baltimore). 2017;96:e6724. https://doi.org/10.1097/MD.000000000000724.
- Jones NC, Tyner KJ, Nibarger L, Stanley HM, Cornelison DDW, Fedorov YV, et al. The p38alpha/beta MAPK functions as a molecular switch to activate the quiescent satellite cell. J Cell Biol. 2005;169:105–16. https:// doi.org/10.1083/jcb.200408066.
- Zammit PS, Golding JP, Nagata Y, Hudon V, Partidge TS, Beauchamp JR. Muscle satellite cells adopt duverf=gent fates: a mechanism fir self-renewal? J Cell Biol. 2004;166:347–57. https://doi.org/10.1083/jcb.20031 2007
- Fang J, Zhang S, Liu Z, Pan Y, Cao L, Hou P, et al. Skeletal muscle cells confer maturing macrophages anti-inflammatory properties through insulin-like growth factor-2. Stem Cells Transl Med. 2020;9:1–13. https:// doi.org/10.1002/sctm.19-0447.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.