

## RESEARCH ARTICLE

# Transcutaneous spinal cord stimulation and motor responses in individuals with spinal cord injury: A methodological review

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## Abstract

### Background

Transcutaneous spinal cord stimulation (tSCS) is a non-invasive modality in which electrodes can stimulate spinal circuitries and facilitate a motor response. This review aimed to evaluate the methodology of studies using tSCS to generate motor activity in persons with spinal cord injury (SCI) and to appraise the quality of included trials.

### Methods

A systematic search for studies published until May 2021 was made of the following databases: EMBASE, Medline (Ovid) and Web of Science. Two reviewers independently screened the studies, extracted the data, and evaluated the quality of included trials. The electrical characteristics of stimulation were summarised to allow for comparison across studies. In addition, the surface electromyography (EMG) recording methods were evaluated.

### Results

A total of 3753 articles were initially screened, of which 25 met the criteria for inclusion. Studies were divided into those using tSCS for neurophysiological investigations of reflex responses ( $n = 9$ ) and therapeutic investigations of motor recovery ( $n = 16$ ). The overall quality of evidence was deemed to be poor-to-fair ( $10.5 \pm 4.9$ ) based on the Downs and Black Quality Checklist criteria. The electrical characteristics were collated to establish the dosage range across stimulation trials. The methods employed by included studies relating to stimulation parameters and outcome measurement varied extensively, although some trends are beginning to appear in relation to electrode configuration and EMG outcomes.

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## Conclusion

This review outlines the parameters currently employed for tSCS of the cervicothoracic and thoracolumbar regions to produce motor responses. However, to establish standardised procedures for neurophysiological assessments and therapeutic investigations of tSCS, further high-quality investigations are required, ideally utilizing consistent electrophysiological recording methods, and reporting common characteristics of the electrical stimulation administered.

## Introduction

Transcutaneous spinal cord stimulation (tSCS) is a non-invasive form of neuromodulation in which electrodes are placed on the skin and used to stimulate the spinal circuitries via an electrical current [1–3]. It has been proposed that this tool could not only provide us with greater understanding of spinal inter-neuronal functioning but also enhance the rehabilitation potential for people with neurological disorders, such as spinal cord injury (SCI) [2, 4–6]. As this modality is under the relatively early stages of investigation with injured individuals, there is still much to learn about its implementation and clinical potential.

Modelling studies have demonstrated that electrical pulses delivered from spinal cord stimulation (SCS) preferentially depolarize sensory afferents in the posterior roots, which can elicit a motor reflex response [7, 8]. This response has been termed a posterior root-muscle (PRM) reflex [9], multisegmental monosynaptic response (MMR, [5]), or transspinal evoked potential (TEP, [10, 11]), amongst other nomenclature. As an alternative to the H-Reflex, the study of the PRM reflex allows us to expand the neurophysiological assessment of sensory-motor transmission of stimuli to more muscle groups and provide greater insights into the functioning of spinal circuitries [4, 12].

Spinal stimulation via transcutaneous input is believed to be distinguished from direct stimulation of motor efferents, such as in traditional nerve or muscle stimulation techniques, due to the transsynaptic transmission of motor responses via monosynaptic or oligosynaptic pathways [13]. Several studies have investigated the reflex nature of responses, using paired pulses to demonstrate post-activation depression (PAD), in which the amplitude of the second pulse of a pair is attenuated with respect to the first [14–22]. Additionally, the inhibition of tSCS evoked responses via tendon vibration is consistent with the stimulation of reflex responses from Ia afferents [5, 22]. Other studies have focused on alternative methods to demonstrate spinal neuromodulation of motor responses through outcomes such as increased response latencies [2, 23], differential muscle activation patterns [24], phase-dependent modulation of reflex responses [5, 14] and the alteration of amplitudes subsequent to afferent input [25] or interlimb conditioning [26].

It is also theorized that SCS can modulate inter-neuronal spinal excitability, and that this may account for the observed motor recovery when used in individuals with SCI [19, 27–29]. By activating networks such as central pattern generators (CPGs) and the propriospinal system (PSS), spinal excitability may be augmented and the threshold for motor impulse propagation lowered [30, 31]. A CPG is a spinal network of neurons believed to be capable of generating a co-ordinated rhythmic motor output such as locomotion in the absence of input from supraspinal centres and/or afferent feedback [32]. The PSS has been described as an interface between spinal segments that contributes to movement and rhythmic coordination [33, 34], as

well as providing a background of subthreshold excitation [30, 35]. The modulation of spinal networks and altered threshold for impulse propagation may explain the results of several studies using tonic spinal stimulation that have reported improved motor outcomes in chronically paralysed individuals [36–38].

Indeed, in the case of SCI, spinal neuromodulation may provide greater functional recovery beyond the capacity of currently available therapies, particularly after more severe or chronic injury [30, 39]. Thus far, a selection of studies investigating the effects of tSCS on motor rehabilitation in chronic SCI have published cases of improved lower limb [19, 40–42], trunk [37] and upper limb functioning [36, 43–45]. Despite these promising initial results, a recent review evaluating the therapeutic effects of tSCS on motor recovery in individuals with SCI reported that due to the small heterogeneous sample sizes, diverse range of outcome measures and low methodological quality of reviewed studies, no conclusions can be drawn on its effectiveness [46].

The most consistently employed measure across studies directly evaluating motor responses from spinal stimulation appears to be surface electromyography (EMG) [46, 47]. EMG can be used either to quantify evoked motor potentials and/or the degree of voluntary muscle activation facilitated by tSCS. In evaluating any clinical therapy, the effects may not be initially present at a functional level but could be present at the neuromuscular level, and these effects may be quantified by EMG [48, 49]. EMG can also estimate the real-time effects of tSCS parameters and application on spinal excitability [17, 22, 50, 51], potentially even providing physiological input for closed-loop control paradigms [52, 53]. While the use of EMG has many advantages, it is subject to limitations that must be carefully considered when recording, processing, and interpreting the data [54]. The use of EMG with neurologically impaired individuals presents many additional challenges [48], such as sub-optimal normalisation procedures [55] and signal contamination by involuntary tonic activity [56]. Furthermore, stimulation artifact can contaminate the EMG motor signals produced by tSCS [49].

Presently, there is a lack of consensus surrounding the standardised use of tSCS to facilitate motor responses in individuals with SCI [46]. Optimal stimulation parameters and experimental protocols remain unclear and there is much variability seen in outcome measurement. The extent of this methodological variability would benefit from a systematic evaluation in order to synthesize the information on currently employed parameters and provide recommendations to enhance the development of future studies investigating the properties and efficacy of tSCS. As such, the objective of this systematic review was to methodologically appraise studies which used tSCS to generate motor activity in persons with SCI. In doing so, this review sought to critique the quality of included trials, review intervention parameters employed and compare the methods of evaluating motor responses with surface EMG.

## Methods

A systematic review of the literature was undertaken using the methodology described by the Preferred Reporting Items for Systematic Review and Meta-Analysis (PRISMA) Protocols Statement [57].

## Search strategy

An extensive literature search was carried out using the following electronic databases: EMBASE, Medline (Ovid) and Web of Science. It included studies published until 31<sup>st</sup> May 2021. The initial search was kept broad to in an attempt to capture all possible spinal stimulation studies using varying nomenclature. The search was built with the help of a research librarian (DM) based on anchoring terms from the following categories: **spinal cord stimulation**, **spinal cord injury** and **motor responses**. Search terms were expanded using a vast list of

alternative terminologies, truncations, and abbreviations. The exact search algorithm and medical subject heading (MeSH) terms used with each engine are presented in [S1 Appendix](#). Additional relevant publications were also sought out by retrospectively completing a manual search of the bibliographies of all included studies and by manually searching for other publications from authors of tSCS studies that were identified in the search.

### Study selection procedure

Two independent reviewers (CT, CMcH) completed an initial title screen to remove any highly irrelevant papers. The eligibility criteria ([Table 1](#)) were designed based on the PICO model (Population, Intervention, Comparison, Outcome). Pilot testing of the exclusion criteria was conducted using a subset of 150 abstracts screened by both reviewers and the reasons for exclusion were documented. The reviewers then completed the abstract screening and a Cohen's Kappa of 0.88 was reached. This correlation was deemed sufficient. Finally, the full texts were reviewed for inclusion and all reasons for exclusion were recorded. If there was any uncertainty about inclusion, a third reviewer (NF) was consulted until a consensus was reached. The independent reviewers were not blinded to the study authors, institutes, or journal titles. As there were a small number of publications meeting the inclusion criteria, we did not require a minimum sample size. The literature search was last performed on the 31<sup>st</sup> May 2021.

### Quality appraisal

In order to appraise the quality of the included full texts, the Downs and Black (D&B) Checklist was employed [58]. This tool has been used to evaluate non-randomised controlled trials (RCTs) in other systematic reviews pertaining to populations with SCI [59–61] and its use is recommended by the SCIRE (Spinal Cord Injury Research Evidence) Research Team [62]. The D&B Checklist has also been recommended for use in assessing non-RCTs due to its psychometric properties [63, 64].

Two independent reviewers (CT, CMcH) conducted the quality appraisal, and any disagreement was discussed with a third reviewer (NF) until consensus was reached. The D&B

**Table 1. The eligibility criteria to determine suitable studies for inclusion in the full-text systematic review.**

	Inclusion	Exclusion
<b>Participants</b>	<ul style="list-style-type: none"> <li>• Aged &gt; 18 years</li> <li>• A primary diagnosis of spinal cord injury (any level, complete or incomplete).</li> </ul>	<ul style="list-style-type: none"> <li>• Animal studies</li> <li>• Aged &lt; 18 years</li> </ul>
<b>Intervention</b>	<ul style="list-style-type: none"> <li>• Transcutaneous spinal cord stimulation aimed at producing a motor response.</li> <li>• Pulsed and continuous electrical spinal stimulation protocols.</li> </ul>	<ul style="list-style-type: none"> <li>• Magnetic stimulation or direct current stimulation</li> <li>• Peripheral stimulation such as Functional Electrical Stimulation (FES) or Neuromuscular Electrical Stimulation (NMES)</li> <li>• Paired Associative Stimulation (PAS)</li> <li>• Epidural spinal cord stimulation (eSCS).</li> </ul>
<b>Comparison</b>	<ul style="list-style-type: none"> <li>• No intervention, sham intervention, or pre-post analysis</li> </ul>	
<b>Outcomes</b>	<ul style="list-style-type: none"> <li>• A measure of motor activity in a targeted muscle/muscle groups by EMG recordings</li> </ul>	<ul style="list-style-type: none"> <li>• The primary outcome selected and reported on measures pain, autonomic function, or spasticity</li> </ul>
<b>Data analysis</b>	<ul style="list-style-type: none"> <li>• Study must report details pertaining to the transcutaneous spinal cord stimulation parameters utilised</li> </ul>	<ul style="list-style-type: none"> <li>• Studies that fail to specify any stimulation parameters</li> </ul>
<b>Publication type</b>	<ul style="list-style-type: none"> <li>• Original primary data from a prospective interventional, quasi-experimental, or observational study</li> <li>• Published in peer reviewed journal until 31<sup>st</sup> May 2021</li> <li>• Published in English</li> </ul>	<ul style="list-style-type: none"> <li>• Review articles, conference proceedings, expert opinions, or any other secondary publication</li> <li>• Abstract or full text not available in English</li> </ul>

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Checklist is a 27-item list that evaluates methodological strengths and weaknesses of articles based on the categories of (1) Reporting, (2) Internal Validity (Bias), (3) Internal Validity (Confounding), (4) External Validity and (5) Power [58]. Power level calculations (1- $\beta$  error probability) for the checklist were made using the G\*Power Application [65] and analysis was derived from the statistical tests applied to the main study findings. The following marks were awarded: 1 point for a power level of 70%, 2 points for power level of 80%, 3 points for power level of 85%, 4 points for power level of 90%, 5 points for power level of 95%. The modified version of the D&B Checklist was not used, as the authors felt it important to adequately represent the sufficient powering of studies as per the original checklist. The following rounded cut-off points were used to categorize studies by quality [66]: excellent (91%–100%), good (71%–90%), fair (51%–70%), and poor (0%–50%).

## Data extraction

Results were generated from data extracted to standardised spreadsheets which included (i) study type, (ii) sample characteristics and clinical variables, (iii) intervention parameters, (iv) outcome measurements (v) EMG data collection and signal processing (vi) and safety/adverse events. Table results were pooled by two study members until consensus was reached, and disagreements were discussed with the third reviewer. Studies investigating similar objectives were grouped together for comparison, in particular, a distinction was made between neurophysiological experiments and therapeutic investigations seeking motor rehabilitation.

The electrical and timing characteristics of the stimulation signals used in tSCS vary widely, making comparisons between studies difficult. Moreover, there is a lack of consistency in the definition of these parameters. This study sought to clearly define key stimulation parameters and descriptors and, where possible, extract data from each publication according to these definitions. Fig 1 shows typical waveforms for constant current pulsed stimulation and identifies selected characteristics, while Table 2. defines the parameters that were used to characterise the tSCS administered.

The root mean square (RMS) current is useful for estimating average electrical power and therefore the heat generating capacity of a waveform,  $P_{avg} = i_{rms}^2 R$ . For a square wave such, as at Fig 1B, the RMS current calculation simplifies to:

$$i_{rms} = i \sqrt{\frac{t1}{T}}$$

Or, for a typical symmetric biphasic waveform like that at Fig 1A, the calculation would be:

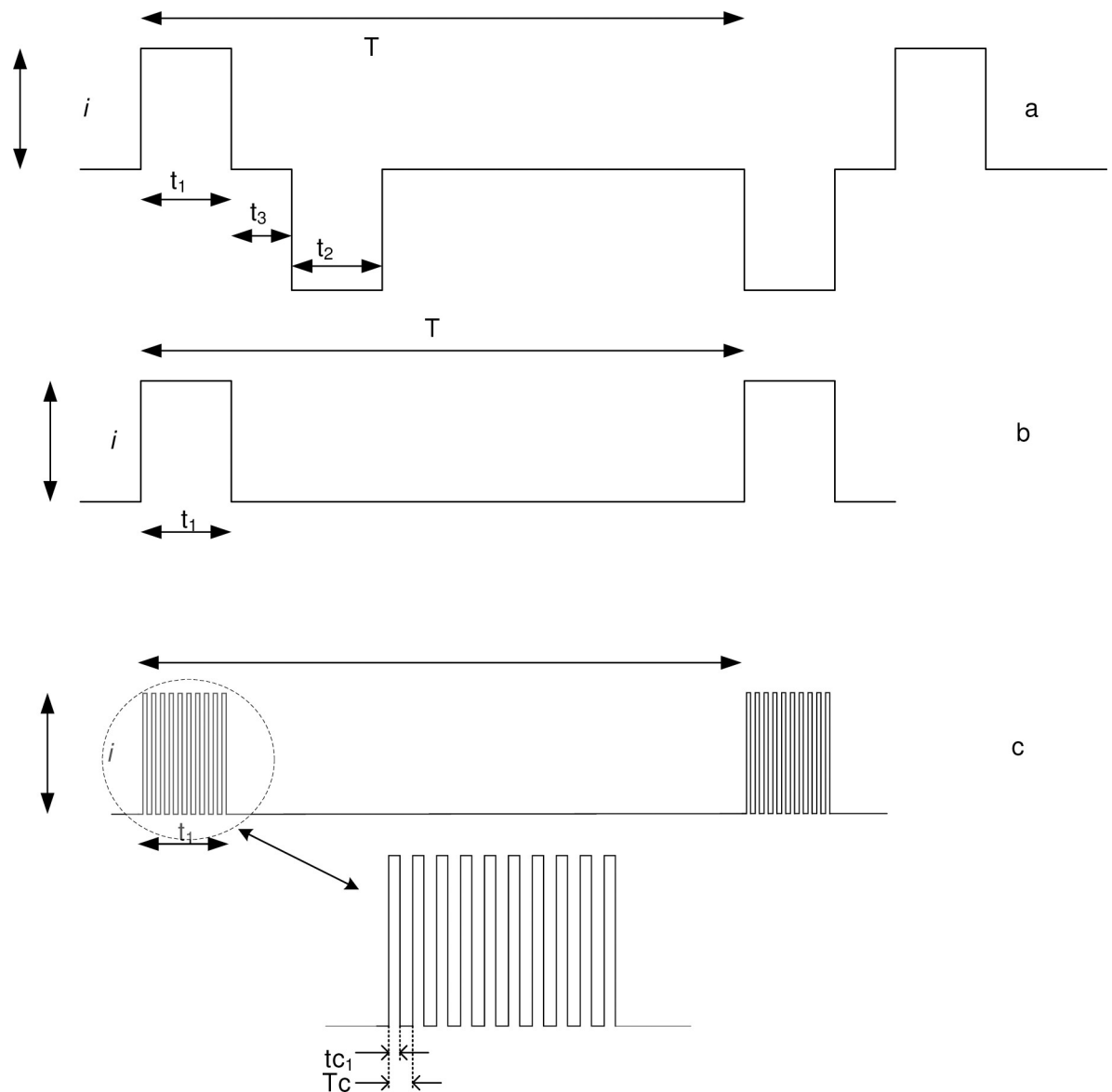
$$i_{rms} = i \sqrt{\frac{t1 + t2}{T}}$$

For the descriptions of other details of included studies, ranges are given with the mean  $\pm$  standard deviation. Due to the heterogeneity in the methods used to evaluate the outcomes and the diverse experimental methodologies, a meta-analysis was not possible, and a descriptive qualitative review was conducted.

## Results

### Literature search and selection

Of the 3753 articles identified (Embase: 1960, Medline (Ovid): 1425, Web of Science:368), 2499 were taken to title and abstract screening after the duplicates were removed. After the removal of 2391 articles from title and abstract screening, 108 full texts were evaluated for



**Fig 1.**

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eligibility. Finally, 25 articles that assessed the ability of tSCS to generate motor responses in individuals with SCI were included in this review (*Fig 2*).

### Study characteristics

Studies were categorized as neurophysiological assessments if their objective was to investigate the properties, mechanisms, or effects of tSCS on outcomes related to nervous system functioning ( $n = 9$ ), whereas studies were labelled as therapeutic if they aimed to enhance motor rehabilitation and recovery in patients with SCI ( $n = 16$ ). In therapeutic investigations, tSCS was commonly combined with simultaneous rehabilitative interventions such as physical therapy, treadmill training, body weight support and the use of exoskeletons or pharmacological agents (*Table 3*). Of the 25 included studies, 7 were case reports, 6 were case series, 3 were

**Table 2. Summary of stimulation parameters and a detailed description of how they are defined.** Definitions and characteristics of stimulation parameters.

Parameter	Symbol	Unit	Description
Pulse interval	T	ms	The time interval between pulses of a sequence
Pulse frequency	f	Hz	The inverse of the pulse interval, $f = 1/T$ , is the number of pulses per second
Phase duration	$t_1$	ms	The duration of the leading phase
Pulse amplitude	i	mA	Current amplitude measured baseline to peak
Phase charge	$q_c$	$\mu\text{C}$	Total charge in the phase
Pulse duration	p	ms	The sum of $t_1+t_2+t_3$
Carrier frequency	$f_c$	Hz	Frequency of a carrier waveform which is modulated by the stimulation waveform
Carrier-on-time	$tc_1$	$\mu\text{s}$	Phase duration of carrier waveform
Carrier period	$T_c$	$\mu\text{s}$	Inverse of carrier frequency
Phase charge density	$q_d$	$\mu\text{C}/\text{cm}^2$	The phase charge per unit electrode area
Root mean square current	$i_{rms}$	mA	$i_{rms} = \sqrt{\frac{1}{T} \int_0^T i(t)^2 dt}$
Electrode current density	$j_e$	mA/ $\text{cm}^2$	$j_e = i_{rms}/A$
Electrode area (active)	A	$\text{cm}^2$	The area of electrical contact at the skin. (assumed uniform current distribution within electrode)

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crossover trials, 7 were quasi-experimental studies (non-equivalent control group or nonrandomised intervention design), one was a sample cohort study, and one was a non-randomised control trial.

## Participant demographics

A total of 173 participants with SCI were recruited across the 25 studies to receive tSCS and their characteristics are described in Table 3. Further analysis only includes data from participants with SCI due to the purposes of this review. The sample sizes in included studies were generally modest ( $n = 7 \pm 6$ ). Neurophysiological investigations tended to have larger samples ( $n = 10 \pm 5$ ) than therapeutic investigations ( $n = 5 \pm 5$ ). A large range of ages, from 18 to 70 (mean  $35 \pm 13$  years), injury classifications (from the level of C1—L2), and impairment levels (AIS A—D classifications), were represented across the included studies. Studies explored the effects of tSCS on different injury chronicity's, from one year to 43 years post-injury occurrence, however, no published studies investigated the use of tSCS at < 1-year post injury.

## Quality appraisal

The quality of included trials was evaluated using the D&B Checklist [58] and this tool deemed the overall evidence quality to be poor-to-fair, with results ranging from 3 to 19, out of a possible score of 32 (Tables 4 and 5). The mean score across all trials was  $10.5 \pm 4.9$ , with  $11.4 \pm 5.1$  for neurophysiological and  $10.0 \pm 5.1$  for therapeutic investigations. In particular, low scores were repeatedly awarded for external validity and selection bias, and the majority of studies were deemed insufficiently powered.

## Methodological evaluation

The methodologies of selected studies were reviewed to outline the common procedures for stimulation implementation and outcome evaluation. The stimulation variables selected

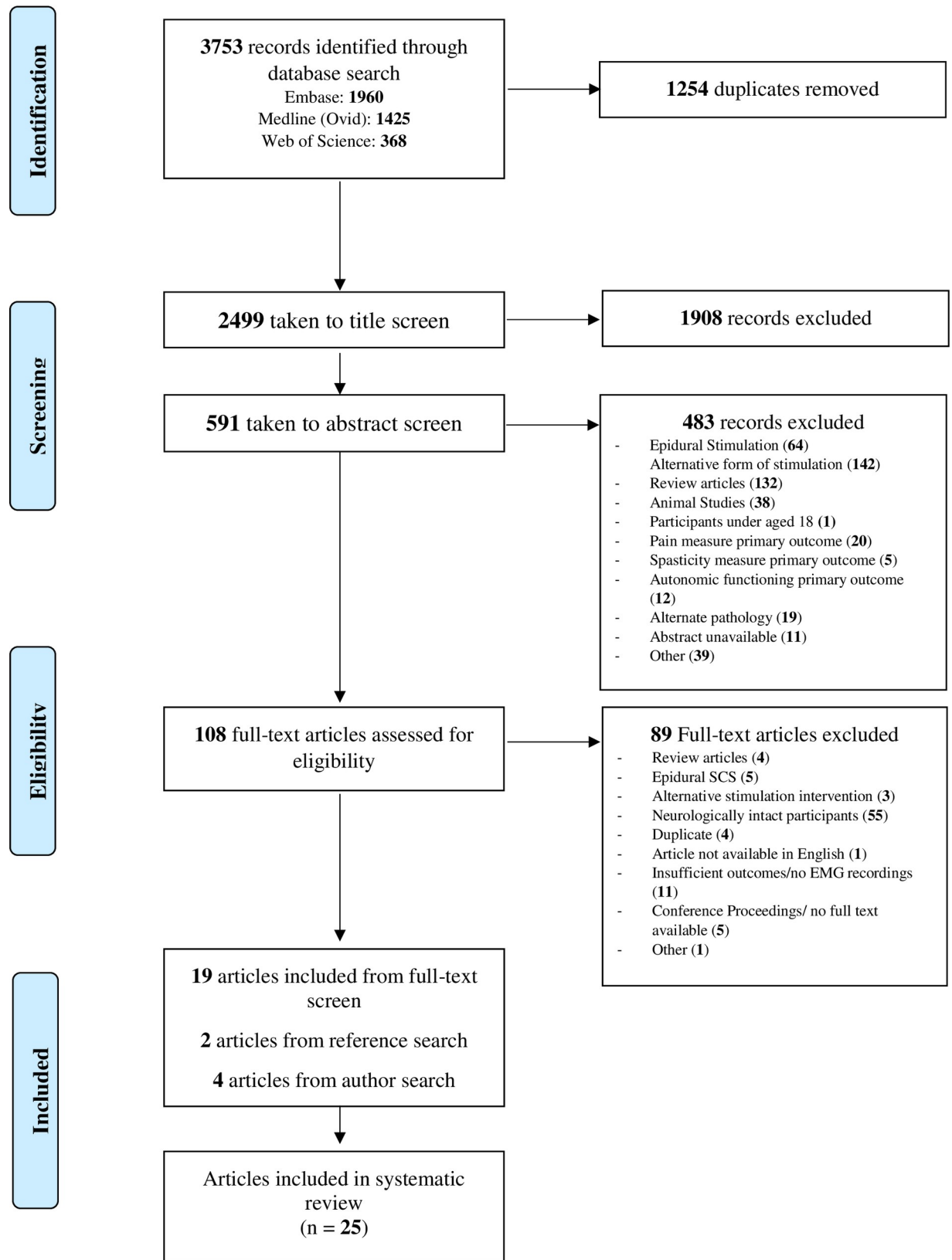


Fig 2. PRISMA flow diagram of screening and selection processes.

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**Table 3. Details of study characteristics and the demographics of included participants.** Study characteristics and participant demographics.

Study	Design	Simultaneous Interventions	Other Subjects (n)	Participants with SCI Demographics					
				Sample (n)	Age; Mean (SD)	Gender	AIS	Level	Years Since Injury; Mean (SD)
Dy <i>et al.</i> , 2010 [14]	NP, QE	Treadmill and BWS	9 NI	9	32.6 (9.2)	M = 9	A	C5-T7	6.4 (9.4)
Hofstoetter <i>et al.</i> , 2013 [6]	TP, CR	Treadmill	-	1	29	F = 1	D	T9	11
Gerasimenko <i>et al.</i> , 2015 [67]	TP, CS	Assisted movement, Buspirone	-	5	31.4 (16.8)	M = 5	B	C5-T4	3.2 (1.6)
Hofstoetter <i>et al.</i> , 2015 [18]	TP, CS	Treadmill	-	3	32.7 (5.0)	M = 2, F = 1	D	C5, T9	10.7(1.5)
Bedi, 2016 [68]	TP, CR	-	-	1	25	M = 1	C	T12	-
Emeliannikov <i>et al.</i> , 2016 [15]	NP, CS	Seated gait device, pharmacology	-	10	39.1 (11.3)	M = 7, F = 3	A-D	T5-L2	4.8 (4.2)
Minassian <i>et al.</i> , 2016 [19]	TP, CS	RDGO, Treadmill and BWS	-	4	39.5 (17.1)	M = 3, F = 1	A	C8-T8	2.8 (1.4)
Gad <i>et al.</i> , 2017 [41]	TP, CR	Exoskeleton, Buspirone	-	1	40	M = 1	A	T9	4
Freyvert <i>et al.</i> , 2018 [44]	TP, CrT	Buspirone, hand grip exercises	-	6	19.2 (1.3)	M = 4, F = 2	B	C5-C8	2.4 (0.9)
Gad <i>et al.</i> , 2018 [43]	TP, CS	Hand grip exercises	-	6	40.2 (16.6)	M = 5, F = 1	B, C	C4-C8	8.0 (7.7)
Hofstoetter <i>et al.</i> , 2018 [16]	NP, QE	PT	7 SCI eSCS	10	39.7 (20.1)	M = 7, F = 3	A, C, D	C4-T7	4.5 (2.8)
Inanici <i>et al.</i> , 2018 [36]	TP, CR	Activity-based PT	-	1	62	M	D	C3	2
Rath <i>et al.</i> , 2018 [37]	TP, RCrT	-	-	8	29.4 (7.7)	M = 7, F = 1	A, C	C4-T9	7.3 (3.3)
Hofstoetter <i>et al.</i> , 2019 [17]	NP, QE	-	10 NI	10	40.1 (18)	M = 8, F = 2	A, C, D	C4-T7	9.7 (12.5)
Murray and Knikou, 2019 [20]	NP, QE	-	10 NI	10	36.3 (11.2)	M = 7, F = 3	A-D	C6-T12	8.8 (8.1)
Sayenko <i>et al.</i> , 2019 [42]	TP, RCrT	Stand Training	-	15	31.2 (8.7)	M = 12, F = 3	A-C	C4-T12	6 (3.2)
Al'Joboori <i>et al.</i> , 2020 [69]	Coh	BWS sit-to-stand training, standing exercises	2 SCI Control	5	36.8 (2.5)	M = 3, F = 2	A, C, D	C6/7 – T10	3 (3.4)
Alam <i>et al.</i> , 2020 [40]	TP, CR	Stand Training and Treadmill	-	1	48	F = 1	D	C7	21
Atkinson <i>et al.</i> , 2020 [26]	NP, QE	-	15 NI	18	29.4 (7.3)	M = 16, F = 2	A-D	C2-T6	4.6 (3.1)
Meyer <i>et al.</i> , 2020 [70]	TP, CS	Overground walking with BWS, ankle movement	-	10	25.4 (12.4)	M = 9, F = 1	D	C3-T10	11.6 (10.2)
Militskova <i>et al.</i> , 2020 [25]	NP, CR	PT, treadmill and BWS	-	1	21	F = 1	A	T11	1
Shapkova <i>et al.</i> , 2020 [38]	TP, NRCT	Exoskeleton	16 SCI Control	19	31.2 (8.6)	M = 15, F = 4	A-C	C8-L2	4.6 (3.3)
Wu <i>et al.</i> , 2020 [23]	NP, QE	-	14 NI, 4 ALS	13	45.9 (13.7)	M = 10, F = 3	B-D	C2-C8	10.8 (5.9)
Zhang <i>et al.</i> , 2020 [71]	TP, CR	Task specific hand training	-	1	38	M = 1	A	C5	15
Islam <i>et al.</i> , 2021 [72]	NP, QE	Treadmill walking and robotic gait orthosis BWS	13 NI	5	43.8 (11.4)	M = 9, F = 1	B-D	C1- T11	13.4 (9.0)

Abbreviations: AIS; ASIA impairment scale, ALS; amyotrophic lateral sclerosis, BWS; Body weight support, Coh; cohort study, CR; case report, CrT; crossover trial, CS; case series, eSCS; epidural spinal cord stimulation, F; female, M; male, NI; neurologically intact, NP; neurophysiological investigation, NRCT; non-randomised controlled trial, PT; physical therapy, QE; quasi-experimental study, RDGO; Robotic driven gait orthosis, RCrT; randomised crossover trial, SCI; spinal cord injury, SD; standard deviation, TP; therapeutic investigation.

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**Table 4. Results from the quality appraisal of neurophysiological investigations using the Down's and Black Checklist.** Quality of Evidence Assessment–Neurophysiological Investigations.

Study	Reporting	External Validity	Internal Validity (Bias)	Internal Validity (Selection Bias)	Power	Quality Score	Percentage	Evidence Category
Dy <i>et al.</i> , 2010 [14]	6	0	4	0	0	10	31%	Poor
Emeliannikov <i>et al.</i> , 2016 [15]	2	0	2	0	0	4	12%	Poor
Hofstoetter <i>et al.</i> , 2018 [16]	7	0	4	1	0	12	37%	Poor
Hofstoetter <i>et al.</i> , 2019 [17]	8	0	5	1	5	19	59%	Fair
Murray and Knikou, 2019 [20]	7	0	3	0	1	11	34%	Poor
Atkinson <i>et al.</i> , 2020 [26]	6	0	4	0	0	10	31%	Poor
Militskova <i>et al.</i> , 2020 [25]	4	0	1	0	0	5	16%	Poor
Wu <i>et al.</i> , 2020 [23]	9	0	3	1	5	18	56%	Fair
Islam <i>et al.</i> , 2021 [72]	6	0	3	0	5	14	44%	Poor
	/11	/3	/7	/6	/5	/32	/100%	

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determine the electrical field generated and subsequent motor responses, and the outcomes used to evaluate these responses are essential in understanding the utility of the parameters selected and the overall effectiveness of tSCS. Factors such as safety and adverse events are also critical to a methodological review.

**Table 5. Results from the quality appraisal of therapeutic investigations using the Down's and Black Checklist.** Quality of Evidence Assessment–Therapeutic Investigations.

Study	Reporting	External Validity	Internal Validity (Bias)	Internal Validity (Selection Bias)	Power	Quality Score	Percentage	Evidence Category
Hofstoetter <i>et al.</i> , 2013 [6]	1	0	2	0	0	3	9%	Poor
Gerasimenko <i>et al.</i> , 2015 [67]	4	0	2	0	0	6	19%	Poor
Hofstoetter <i>et al.</i> , 2015 [18]	2	0	2	0	0	4	12%	Poor
Bedi, 2016 [68]	4	0	3	0	0	7	22%	Poor
Minassian <i>et al.</i> , 2016 [19]	5	0	4	0	0	9	28%	Poor
Gad <i>et al.</i> , 2017 [41]	3	0	0	0	0	3	9%	Poor
Freyvert <i>et al.</i> , 2018 [44]	7	0	5	2	0	14	44%	Poor
Gad <i>et al.</i> , 2018 [43]	9	0	2	0	0	11	34%	Poor
Inanici <i>et al.</i> , 2018 [36]	5	0	3	1	0	9	28%	Poor
Rath <i>et al.</i> , 2018 [37]	7	0	4	1	0	12	37%	Poor
Sayenko <i>et al.</i> , 2019 [42]	6	0	3	4	4	17	53%	Fair
Al'Joboori <i>et al.</i> , 2020 [69]	8	1	4	2	0	15	47%	Poor
Alam <i>et al.</i> , 2020 [40]	3	0	2	4	0	9	28%	Poor
Meyer <i>et al.</i> , 2020 [70]	8	0	4	3	1	16	50%	Poor
Shapkova <i>et al.</i> , 2020 [38]	8	1	5	0	3	17	53%	Fair
Zhang <i>et al.</i> , 2020 [71]	5	0	3	0	0	8	25%	Poor
Total Maximum Score	/11	/3	/7	/6	/5	/32	/100%	

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## Electrode configurations

The electrode configurations with regard to position and location varied substantially across experiments (Tables 6 and 7). The cathode was positioned dorsally over the vertebral column in the majority of studies, otherwise electrodes were used that alternated polarity within a biphasic pulse [16–18, 23, 40, 69, 70]. Some studies specified a paravertebral dorsal electrode orientation [16–19, 68], whereas most others placed the electrode in the midline over the vertebral column [14, 20, 23, 25, 26, 36, 37, 40–44, 67, 69–72]. Many studies targeted a single site, however, 8 out of 14 therapeutic investigations favoured the stimulation of multiple sites simultaneously [36, 37, 40–43, 67, 71].

The most common vertebral level stimulated for targeting lower limb motor activity was T11–T12 and/or L1–L2. Two studies placed electrodes within the range of T9–L2, but adjusted the exact positions based on evoked EMG motor responses [25, 26]. An additional secondary stimulating electrode was also placed on the coccygeal bone during two experiments [41, 67]. For the upper limb responses, the cathode site varied substantially across the four studies and was placed on C5 [44], T2–T4 [23], or C3–C4 simultaneously with C6–C7 or C7–T1 [36, 43, 71].

The anode location selected for experiments targeting lower limb motor responses varied between the anterior superior iliac spine (ASIS) and iliac crests ( $n = 6$ ) or para-umbilically over the anterior abdomen ( $n = 8$ ), with two studies recording the use of both locations depending on patient comfort [20, 72]. Conversely, one study described placing both the anode and the cathode in the midline over the vertebral column [69]. In studies of upper limb responses, the iliac crests or ASIS were chosen by four investigations [36, 43, 44, 71] with only Wu *et al.*, [23] placing the anode on the anterior neck.

## Electrical dosage

Clear differences in dosage arose between neurophysiological and therapeutic investigations. In neurophysiological investigations, tSCS was typically delivered using isolated single or paired pulses with long refractory periods allowing a return of resting membrane potential and frequencies, when outlined, were typically low. A notable exception to this was a recent investigation by Islam *et al.* [72] who administered high frequency pulse trains at 333Hz, the rationale for which was unclear. Delivered current in neurophysiological investigations ranged between 24.7 [14] to  $248 \pm 87.06$  mA [72] with only two studies exceeding a maximum of 100mA [20, 72]. A variety of criteria were used to determine the stimulation intensities, for example, the point at which threshold responses were observed in some [20, 23] or all muscles [14, 16, 17], maximum tolerance [25], response magnitude plateau [26], equivalent amplitude to H-Reflex soleus response [17, 20, 72], or the lowest amplitude that completely suppressed the second stimulus of a pair [15]. The majority of neurophysiological experiments reported using a square or rectangular monophasic current waveform with 1ms pulse width, with just two studies using biphasic pulses [16, 17], and one which trialled both [23].

In contrast, therapeutic investigations typically reported the application of continuous pulse trains, with a burst frequency of 5–30 Hz and an intra-pulse carrier frequency of 2.5–10 kHz [36, 37, 42, 43, 67, 68]. The use of this intra-pulse carrier frequency is poorly justified and appears to be for analgesic purposes, although no evaluation of this could be identified. Other therapeutic experiments selected simplified phase characteristics with either biphasic or monophasic rectangular-waves with a frequency ranging from 1–90 Hz, with 20–30 Hz the most commonly occurring selection [6, 18, 19, 38, 40, 44, 69, 71]. The duration of therapeutic stimulation varied from bouts of <5 mins [37, 40, 67] to > 45 mins [36, 38, 68] and was generally paired with concomitant rehabilitative activities. Recorded current ranges in therapeutic experiments reached a maximum of 180mA in the thoracolumbar region [67] and 210mA in

**Table 6. Parameters selected by neurophysiological assessments investigating the effects of tSCS on spinal cord functioning in individuals with SCI.** Stimulation parameters selected by studies carrying out neurophysiological assessments into the properties of spinal cord stimulation with SCI participants.

Study	Patient position	ELECTRODES			STIMULATION PROTOCOL			ELECTRICAL CHARACTERISTICS	
		Size/shape [Area]	Polarity	Location	Description	Frequency	Intensity	Phase Charge (μC)	Max Phase Charge Density (μC/cm <sup>2</sup> )
<i>Lower limb responses</i>									
Dy <i>et al.</i> , 2010 [14]	Lying prone, BWS standing, BWS stepping	ø 2.5 cm [4.9 cm <sup>2</sup> ]	Cathode	T11-T12	Single, t <sub>1</sub> = 1ms, mono square wave pulses	1) Prone/ Standing: 0.5 Hz	24.7 - 83mA	25–83	16.9
		Pair 5.0 x 10.2 cm	Anodes	Iliac crests		2) Stepping: 0.25–0.33 Hz	Set to where consistent responses observed in all measured muscles in standing		
Emeliannikov <i>et al.</i> , 2016 [15]	Seated	-	-	T11-T12	t <sub>1</sub> = 1ms paired pulses (50ms inter-pulse interval)	0.3 Hz for H-Reflex	30–80 mA	30–80	-
		-	-	-			Lowest amplitude to completely suppress the second stimulus of a pair		
Hofstoetter <i>et al.</i> , 2018 [16]	Lying supine	Pair ø 5 cm [2 x 19.6 cm <sup>2</sup> ]	Alternating (anode first pulse, cathode second)	T11-T12 paravertebrally	Charge balanced, symmetric biph rectangular t <sub>1</sub> = 1ms	-	32–86 mA Adjusted to reach target threshold >100uV in all muscle groups studied	32–86	2.2
		8 x 13 cm	Alternating	Para-umbilically lower abdomen					
Hofstoetter <i>et al.</i> , 2019 [17]	Lying supine	5 x 9 cm [45 cm <sup>2</sup> ]	Alternating (anode first pulse, cathode second)	T11–T12 paravertebrally	Charge balanced, symmetric biph rectangular t <sub>1</sub> = 1ms	-	Adjusted to elicit control-PRM reflexes in the right soleus with amplitudes that best matched the control-H reflexes and to elicit PRM reflexes in other muscles studied	-	-
		Pair 8 x 13 cm	Alternating	Lower abdomen					
Murray and Knikou, 2019 [20]	Lying supine	10.2 x 5.1 cm [52 cm <sup>2</sup> ]	Cathode	T10—L1/L2	1) Intervention: alternating suprathreshold and subthreshold stimulation (60 mins/session), mono square wave t <sub>1</sub> = 1ms	1) 0.2Hz	Selected based on threshold to produce right soleus evoked potential (96.9 ± 24 mA). Treatment sessions ranged from 0.4–4.3 times this resting threshold	97	1.9
		Connected pair 10.2 x 5.1 cm <sup>2</sup>	Anode	Para-umbilically/ iliac crests	2) Assessment: mono square wave t <sub>1</sub> = 1ms	2) 0.1, 0.125, 0.2, 0.33, 1.0 Hz		From below motor threshold until plateau reached	417
Atkinson <i>et al.</i> , 2020 [26]	Lying supine	ø 1.8 cm [2.6 cm <sup>2</sup> ]	Cathode	Midline T9-T10 (n = 1), T10-T11 (n = 7), T11-T12 (n = 6), T12-L1 (n = 1)	Single, mono square wave pulses, t <sub>1</sub> = 1ms	-	0-100mA or until response magnitude plateaued	0 to 100	39.3
		Pair 5 x 9 cm	Anode	Anterior superior iliac spines					

(Continued)

Table 6. (Continued)

Study	Patient position	ELECTRODES			STIMULATION PROTOCOL			ELECTRICAL CHARACTERISTICS	
		Size/shape [Area]	Polarity	Location	Description	Frequency	Intensity	Phase Charge (μC)	Max Phase Charge Density (μC/cm <sup>2</sup> )
Militskova <i>et al.</i> , 2020 [25]	Lying supine, BWS standing	ø 2.5 cm [4.9 cm <sup>2</sup> ]	Stimulating	Midline T9-T10, T10-T11, T11-T12, T12-L1, and L1-L2	Mono rectangular pulses t <sub>1</sub> = 1ms	-	30-100mA or maximum tolerated	30 to 100	20.4
		Pair 4 x 2 cm	Reference	Lower abdomen					
Islam <i>et al.</i> , 2021 [72]	Lying supine, BWS stepping	10.2 x 5.1 cm [52 cm <sup>2</sup> ]	Cathode	Longitudinally between T10-L1 vertebrae	1) Single mono pulses t <sub>1</sub> = 1ms		248 ± 87.06 mA for single pulses,	248 ± 87.06	4.8
		pair 10.2 x 5.1 cm	Anode	Iliac crests or either side of abdominal muscles	2) Pulse train of 12 1 ms pulses with a total duration of 33 ms randomly across the step cycle	333.3 Hz	57 to 160 mA for pulse trains Intensity set for Sol TEPs to be equivalent to the Sol H-reflex	57 to 160	18.8 to 53

**Upper limb responses**

Wu <i>et al.</i> , 2020 [23]	Seated	5 x 10cm [50 cm <sup>2</sup> ]	1) Alternating polarity	4cm caudal to C7 (T2-T4)	1) Anode posterior 2ms, t <sub>1</sub> = 1ms biph,	0.2Hz	80–175% of RMT, (RMT = 5.5–51 ma)	89 μc mono	1.8
			2) Cathode posterior for majority		2) Cathode posterior 2ms, t <sub>1</sub> = 1ms biph,			89 μc biph	1.8
		5 x 10cm	1) Alternating polarity	1-2cm above sternal notch (C4-C5 levels anteriorly)	3) Cathode posterior t <sub>1</sub> = 0.5ms biph,		Threshold calculated as enough to elicit > 50uv in 5/10 reps		
			2) Anode anterior for majority		4) Cathode posterior t <sub>1</sub> = 1ms mono				

**Abbreviations:** biph; biphasic, mono; monophasic, PRM; posterior root-muscle, RMT; resting motor threshold, Sol; soleus, TEP; transpinal evoked potential. Where more than one test protocol existed within a given publication, the protocols were detailed using numerical listing: 1) 2) 3) etc. Phase charge density is given in terms of upper limits.

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the cervical region [43]. Intensity criteria was not always explicitly specified. Some studies note that it was based off sufficient levels to reach desired muscle responses [43, 67], perceived sensory thresholds [68] or the amplitude at which reflex threshold was reached [19]. The pulse width was between 0.5 to 1 ms per phase with rectangular waveforms and the majority applied monophasic pulses, with the exception of several studies that selected biphasic pulses [6, 18, 40, 69, 70] and two studies that used both [36, 43]. Two studies used voltage pulses [6, 18] and the resulting current amplitude was not available.

**Electrical characteristics**

The variances in electrode sizes and configurations along with differences in dosage parameters such as amplitude, frequency and pulse duration make it difficult to compare the electrical characteristics of stimulation across studies. We have therefore attempted to calculate common characteristics that were gleaned from the available data. The pulse charge was reasonably

**Table 7. Parameters selected for therapeutic stimulation investigating the effects of tSCS on motor rehabilitation.** Stimulation parameters selected by therapeutic studies investigating the effects of transcutaneous spinal cord stimulation for motor recovery.

Study	Position/ Action	ELECTRODES			STIMULATION PROTOCOL				ELECTRICAL CHARACTERISTICS		
		Size/shape [Area]	Polarity	Location	Description	Frequency	Intensity	Duration	Pulse Charge ( $\mu\text{C}$ )	Current RMS (mA)	Current Density (mA/ $\text{cm}^2$ )
Hofstoetter <i>et al.</i> , 2013 [6]	Upright/ stepping on treadmill	Pair $\phi$ 5cm [2 x 19.6 $\text{cm}^2$ ]	-	Sgl: T11/T12	Sub-motor threshold, charge balanced, symmetric, biph rectangular pulses of $t_1 = 1\text{ms}$	30Hz	18V	-	-	-	-
		Pair 8 x 13cm	-	Lower anterior abdomen							
Gerasimenko <i>et al.</i> , 2015 [67]	Side-lying/ gravity-neutral stepping	$\phi$ 2.5cm [4.9 $\text{cm}^2$ ]	Cathode	Mult: T11-T12 and coccyx 1 (Co1)	Monopolar rectangular stimuli, $t_1 = 1\text{ms}$	T11: 30Hz (+10kHz <i>cf</i> )	80-180mA, Stepping motor threshold	3 x 3 mins (T11, Co1, both)	40-90	9.8-22	2.0-4.5
		Pair 5 x 10.2 $\text{cm}^2$	Anode	Iliac crests		Co1: 5Hz (+10kHz <i>cf</i> )			40-90	4.0-9.0	0.8-1.8
Hofstoetter <i>et al.</i> , 2015 [18]	Standing/ treadmill stepping	Pair $\phi$ 5cm [2 x 19.5 $\text{cm}^2$ ]	Alternating (anode 1 <sup>st</sup> phase, cathode 2 <sup>nd</sup> )	Sgl: T11/T12 paravertebrally	Charge-balanced, symmetric, biph rectangular $t_1 = 1\text{ms}$	30Hz	18-27 V, 86% of reflex threshold (P1), 71% (P2), 80% (P3).	-	-	-	-
		Pair 8 x 13cm	Alternating	Para-umbilically							
Bedi, 2016 [68]	Voluntary and passive movement	Pair 4.5 x 9 cm [2 x 41 $\text{cm}^2$ ]	-	Sgl: T10-L1 paravertebrally	-	30, 50, 70, 90 Hz (+2.5 kHz <i>cf</i> )	Raised to sensory threshold	45 mins per frequency	-	-	-
Minassian <i>et al.</i> , 2016 [19]	Supine, Standing/ assisted treadmill stepping	Pair $\phi$ 5 cm [2 x 19.6 $\text{cm}^2$ ]	Cathode	Sgl: T11- T12, paravertebrally, 1cm apart	Rectangular mono $t_1 = 1\text{ms}$	30Hz	P1: 140mA	10 gait cycles	P1: 140	24.25	0.62
							P2: 100 mA		P2: 100	17.32	0.44
							P3: 170 mA		P3: 170	29.44	0.75
							P4: 125 mA		P4: 125	21.65	0.55
		Pair 8 x 13cm	Anode	Abdomen			Increments of 5ma until reflex threshold				
Gad <i>et al.</i> , 2017 [41]	Standing, supine/ exoskeleton stepping, voluntary movement	$\phi$ 2.5cm [4.9 $\text{cm}^2$ ]	Cathode	Mult: T11-T12 and coccyx 1 (Co1)	-	T11: 30H	-	3 x 20 mins/ session	-	-	-
						Co1: 5Hz					
		5 x 10.2 $\text{cm}^2$	Anode	Iliac crests		Tc = 100 $\mu\text{s}$ <i>cf</i>					
Sayenko <i>et al.</i> , 2019 [42]	Standing/ standing balance exercises, sit-to-stand	$\phi$ 3.2cm diameter [8 $\text{cm}^2$ ]	Cathode	Mult + sgl:	Mono $t_1 = 1\text{ms}$ pulses	1) T11/L1: 5, 15, 25 Hz Tc = 100 $\mu\text{s}$ <i>cf</i>	1) T11/L1: Up to 150 mA	-	75	16.7 (@ 25 Hz)	2.1
				1) T11 and/or L1							
		Pair 7.5 x 13 cm	Anode	Iliac crests		2) L1: 15Hz	2) L1: Up to 100 mA		75	8.7 (@15 Hz)	1.1
Al'Joboori <i>et al.</i> , 2020 [69]	Standing/ sit-to-stand training and standing exercises	5x5 cm [25 $\text{cm}^2$ ]	Cathode	T10/11	2) Biph $t_1 = 1\text{ms}$	2) 30 Hz	Below motor threshold	60 mins per session	110	26.9	1.1
		5x5 cm	Anode	T12/L1			40-110 mA				

(Continued)

Table 7. (Continued)

Study	Position/ Action	ELECTRODES			STIMULATION PROTOCOL				ELECTRICAL CHARACTERISTICS										
		Size/shape [Area]	Polarity	Location	Description	Frequency	Intensity	Duration	Pulse Charge (μC)	Current RMS (mA)	Current Density (mA/cm <sup>2</sup> )								
Alam <i>et al.</i> , 2020 [40]	Standing, sitting, supine/standing, stepping and voluntary movement training	Pair ø 3.2cm [2 x 8 cm <sup>2</sup> ]	Alternating	Mult: T11 and L1	1) Biphasic with	1) 20 Hz /100 μs	1) T11: 105ma, L1: 100 mA	3 x ~10 mins and 3 x 2–3 mins/ session	5.3	3.8	0.3								
					Tc <sub>1</sub> = 50μsec														
					T <sub>1</sub> = 100 μs														
		Pair 6x9 cm		Iliac crests	2) Tonic biphasic stimulation with	2) 30 Hz /100 μs	2) T11: 95ma, L1: 90 mA		4.8	4.7	0.3								
					Tc <sub>1</sub> = 50μs														
					T <sub>1</sub> = 100 μs														
			3) Tonic biphasic stimulation with	3) 20–30 Hz/1ms	3) T11: 20–120ma, L1: 20–120 mA		60	14.7 (@30Hz)	0.9										
			Tc <sub>1</sub> = 50μs t <sub>1</sub> = 1ms																
Meyer <i>et al.</i> , 2020 [70]	Supine, standing/stepping and voluntary movement training	5 x 9 cm [45 cm <sup>2</sup> ]	Alternating (anode for 1 <sup>st</sup> phase, cathode 2 <sup>nd</sup> )	T11-t12	Charge-balanced, symmetric, biphasic rectangular t <sub>1</sub> = 1ms	15 hz, 30 Hz, and 50 Hz	0.8–1.0 x PRM-reflex threshold, 1) Ankle movements 26.4 ± 17.3 mA	1) Ankle control and 2) walk test: max 5 min	26.4	4.6 (@15 Hz)	0.1								
												pair 7.5 x 13 cm	Alternating	Abdomen, paraumbilically	2) Walk tests: 34.8 ± 13.1 ma	3) Spinal reflex assessment: max 15 min	34.8	6.0 (@15 Hz)	0.13
															3) Spinal reflex: 28.8 ± 14.7 mA				
		supine, standing/exoskeleton walk training	Pair 3 x 4 cm [2 x 12 cm <sup>2</sup> ]	Cathode		Sgl: T12 vertebrae	T <sub>1</sub> = 0.5ms mono square wave	G1: 1 Hz, G2: 3 Hz, G3: 67 Hz	1.3–1.4 x motor threshold	~41–53 mins/ session	50	G1: 2.2	0.1						
												G2: 3.9		0.2					
												G3: 618.3			0.8				
	Pair 3 x 4 cm	Anode	Central abdomen			5–100 mA													
<b>Upper limb responses</b>																			
Freyvert <i>et al.</i> , 2018 [44]	Voluntary hand contractions	-	Cathode	Sgl: C5		5-30Hz for 15–30 mins	20–100 mA	15–30 mins/ session	-	-	-								
			Grounding	ASIS															
Gad <i>et al.</i> , 2018 [43]	Voluntary hand contractions	ø 2cm diameter [3.1 cm <sup>2</sup> ]	Cathode	Mult: C3-C4 and C6-C7	1) Pulsed mono t <sub>1</sub> = 1ms	1) 1Hz	1) 10–200 mA	-	100	4.5 (1 Hz)	1.4								
		Pair 5.0 x 10cm <sup>2</sup> , rectangular	Anode	Iliac crests	2) Continuous biphasic or mono t <sub>1</sub> = 1ms	2) 30 Hz + (Tc = 100μs cf)	2) 70–210 mA			25.7 (@30Hz, bi)	8.2								
										36.4 (@30Hz mono)		11.6							

(Continued)

Table 7. (Continued)

Study	Position/ Action	ELECTRODES			STIMULATION PROTOCOL				ELECTRICAL CHARACTERISTICS		
		Size/shape [Area]	Polarity	Location	Description	Frequency	Intensity	Duration	Pulse Charge ( $\mu\text{C}$ )	Current RMS (mA)	Current Density (mA/cm <sup>2</sup> )
Inanici <i>et al.</i> , 2018 [36]	Upper limb activity-based PT	Pair x $\phi$ 2.5cm [2 x 8 cm <sup>2</sup> ]	Cathode	Mult: midline C3-C4 and C6-C7	1) Continuous 60 $\pm$ 20 mins biph $t_1 = 1$ ms	1) 30 Hz + (Tc = 100 $\mu\text{s}$ cf)	1) 80–120 mA	1) 60 $\pm$ 20 mins/session	60	14.7 (30Hz)	1.5
		Pair x 5 x 10cm	Anodes	Iliac crests	2) Pulsed mono rectangular $t_1 = 1$ ms bursts	2) 1Hz	2) 10–120 mA at 10 mA intervals	2) Pulsed	60	2.7 (1Hz)	0.3
Zhang <i>et al.</i> , 2020 [71]	Seated/ Upper limb activity-based PT	Pair x round electrodes 8 x 13cm	Cathode	Mult: midline C3-C4, C7-T1	Pulsed mono rectangular $t_1 = 1$ ms bursts	30 Hz + (Tc = 100 $\mu\text{s}$ cf)	0–80 mA at 5 mA increments	Pulsed assessment 60 mins/session	40	9.8 @ 30Hz	0.6
			Anode	Iliac crests			C3-4: 50 mA				
				C7-T1: 15 mA,							
				Regularly adjusted							
<b>Trunk responses</b>											
Rath <i>et al.</i> , 2018 [37]	Sitting/ seated balance tasks	2 x $\phi$ 3.2cm [2 x 8 cm <sup>2</sup> ]	Cathode	Mult: T11 and L1	Mono, rectangular 1ms pulses	T11: 30Hz (Tc = 100 $\mu\text{s}$ cf)	1) 10–150 mA to detect motor threshold	3–4 x ~1–2 mins/session	50	12.25 (T11)	1.52
		Pair 7.5 x 13cm	Anode	Iliac crests			L1: 15Hz (Tc = 100 $\mu\text{s}$ cf)				
				T11: 25–100 mA							
				L1: 5–80 mA							

Abbreviations: ASIS; Anterior Superior iliac Spine, biph; biphasic, cf; carrier frequency, Co1; coccyx 1, G; group, mono; monophasic, mult; multiple stimulation levels, P; participant, PT; physical therapy, sgl; single stimulation level.

Where more than one test protocol existed within a given publication, the protocols were detailed using numerical listing: 1) 2) 3) etc.

Electrical characteristics such as current RMS and current density include the upper limits.

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consistent in the neurophysiological investigations, in the range 30 to 100  $\mu\text{C}$ , although two studies did exceed this value [20, 72]. The resulting maximal charge density at the spinal electrode varied enormously between studies because of the range of electrode sizes used, 1.8 to 53  $\mu\text{C}/\text{cm}^2$ .

The therapeutic investigations used sustained trains of pulses and the resulting current and current density was compared. Root-mean-square current was in the range 2.2 to 36.4 mA, with most studies below 20 mA. Once again, the variation in electrode area led to a wide range of current densities between studies, 0.1 to 4.5 mA/cm<sup>2</sup>, with one study exceeding this reaching 11.6 mA/cm<sup>2</sup> due to a high current combined with a small electrode area [43].



## Participant positioning

The majority of neurophysiological investigations were conducted with subject in either supine [16, 17, 20, 26] or seated [15, 23] positions. A small number of studies were carried out with participants standing [14, 72] and prone [14]. One investigation compared a number of varying positions to investigate positional effects [25]. In contrast, the majority of therapeutic interventions targeting lower limb responses were conducted in an upright standing position [6, 18, 19, 38, 40–42, 69] and/or while supine [19, 38, 40, 41]. Stimulation targeting trunk control and sitting balance was carried out in a seated position [37] and seated positioning was only outlined in one study of upper limb functioning [71].

## The reflex nature of tSCS responses

Only four therapeutic investigations [18, 19, 69, 70] assessed the nature of motor responses generated by tSCS, while in contrast, all neurophysiological investigations recorded the reflex origin of evoked responses. Primarily the transynaptic modulation of responses was demonstrated using the paired pulse paradigm in which two pulses were delivered with a short conditioning-test interval (CTI) to demonstrate PAD of the second response. Interstimulus intervals between 30–50ms were generally selected to demonstrate PAD [15, 16, 23, 25, 69, 70], with a loss of amplitude attenuation of the second pulse occurring at intervals greater than 100ms [17, 19]. Other than the paired pulse paradigm, response latencies were also used to indirectly evaluate stimulation of dorsal afferents to trigger a reflex response [23], along with the use of vibration to demonstrate pre-synaptic inhibition of motor responses [14].

## Outcome measurement

There were a large variety of outcome measures employed by therapeutic investigations to evaluate motor performance, with 30 different measures used across 16 investigations (Table 8). A total of 10 studies measured joint kinematics, 7 studies assessed functional outcomes and 5 studies each assessed gait parameters and force production. Only two therapeutic investigations evaluated the effects of tSCS on subjective quality of life outcomes [36, 69]. Apart from the recording of EMG data, the most frequently employed objective outcomes in therapeutic studies were an evaluation of AIS scoring ( $n = 6$ ), goniometer data of joint angles ( $n = 5$ ), and centre of pressure/foot loading data ( $n = 5$ ).

Neurophysiological investigations focused primarily on objectively evaluating the amplitude of EMG responses evoked from tSCS, although some studies additionally looked at the conditioning effects of tSCS on spinal excitability as measured by H-reflex and M-wave amplitude [15, 17, 20, 26, 72]. Temporal/phasic modulation of responses evoked by tSCS during gait were also assessed by two studies [14, 72].

## Surface electromyography

Surface EMG recordings from 28 different muscle locations on the lower limb ( $n = 11$ ), upper limb ( $n = 13$ ), and trunk ( $n = 4$ ) were described. An overview of the recording, processing and presentation of EMG signals are presented in Tables 9 and 10 for neurophysiological and therapeutic studies, respectively. Only 8 studies provided adequate details of the preparation including skin preparation, electrode type, shape, composition, and inter-electrode distance [6, 14, 16–19, 68, 72], each experiment recorded the use of round silver-silver chloride electrodes with an interelectrode distance of 1.7, 2 or 3 cm. Sampling frequencies ranged from 1,000 to 10,000 Hz.

**Table 8. Outcomes measures employed by therapeutic studies investigating the outcomes of tSCS on motor rehabilitation in individuals with SCI.** Outcomes selected in the included studies evaluating the effects of therapeutic transcutaneous spinal cord stimulation in spinal cord injured individuals.

Study	Force	Kinematics	Gait	Function	Other
Hofstoetter <i>et al.</i> , 2013 [6]	-	joint angles (goniometer)	Stride length, cycle duration (pressure switches)	-	-
Gerasimenko <i>et al.</i> , 2015 [67]	-	joint angles (goniometer)	-	-	-
Hofstoetter <i>et al.</i> , 2015 [18]	-	joint angles (goniometer)	Swing/stance phase duration, cycle duration (foot sensor)	-	-
Bedi, 2016 [68]	-	-	-	-	-
Minassian <i>et al.</i> , 2016 [19]	-	-	-	-	-
Gad <i>et al.</i> , 2017 [41]	-	joint angles (goniometer and EKSO position sensors)	cycle duration (EKSO device)	-	Self-scoring: muscle tone, sensation, perspiration, coordination, level of robotic assistance, mean HR/BP during training
Freyvert <i>et al.</i> , 2018 [44]	Handgrip force measurement	-	-	UEMS (AIS), ARAT	Spasticity (MAS)
Gad <i>et al.</i> , 2018 [43]	Handgrip force measurement (transducer)	-	-	Motor and sensory scores (AIS)	Self-report QoL
Inanici <i>et al.</i> , 2018 [36]	Pinch strength (pinch gauge)	-	-	AIS scoring, GRASSP	QoL questionnaires (WHO Quality of Life—BREF, SF-Qualiveen, SCIM III)
Rath <i>et al.</i> , 2018 [37]	-	Video and 3D kinematic recordings (Xbox One Kinect), centre of pressure (force plate system)	-	-	-
Sayenko <i>et al.</i> , 2019 [42]	Knee assistance (force sensing resistor)	Centre of pressure (force plate)	-	-	Qualitative level of assistance, time spent standing
Al'Joboori <i>et al.</i> , 2020 [69]	-	Ankle and knee joint ROM (electro-goniometers), STS leg loading (force plates)	-	Motor and Sensory scores (AIS)	BMCA, SF-36 health survey, SCIM III
Alam <i>et al.</i> , 2020 [40]	-	joint angles and body positions (integrated motion capture system), Sit-to-stand transitions (force plate)	-	AIS scoring	-
Meyer <i>et al.</i> , 2020 [70]	-	Ankle ROM and gait kinematics (motion capture system)	gait speed	-	-
Shapkova <i>et al.</i> , 2020 [38]	-	Joint angles and body position (ExoAtlet Global exoskeleton), foot loading (force plates and F-Scan sensors)	Hauser Ambulation Index, maximum nonstop walk duration (ExoAtlet Global exoskeleton), Asymmetry Index (ASI)	AIS scoring	Spasticity (MAS), spinal excitability (H-Reflex amplitude)
Zhang <i>et al.</i> , 2020 [71]	Handgrip force, lateral pinch force, elbow flexion torque (dynamometer)	-	-	GRASSP, NRS	spinal motor evoked potentials

**Abbreviations:** AIS; ASIA Impairment Scale, ARAT; Action Research Arm Test, ASI; asymmetry index, BMCA; Brain Motor Control Assessment, EKSO; Ekso Bionics, EMG; electromyography, GRASSP; Graded and Redefined Assessment of Strength, Sensibility and Prehension, MAS; Modified Ashworth Scale, NRS; Neuromuscular Recovery Scale, QoL; quality of life, SCIM III; Spinal Cord Independence Measure Version III, SF; short form, STS; sit-to-stand, UEMS; upper extremity motor score, WHO; World Health Organisation.

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Several studies explicitly reported filters for stimulus artefact such as bandpass [67], Butterworth [6, 14, 41] or linear adaptive filters [37, 42], whereas others attempted to quantify stimulation artifact by placing electrodes on alternative trunk muscles that were not directly

**Table 9. A summary of evoked surface EMG data collection, recording and signal processing.** EMG recording and signal processing for studies carrying out neuro-physiological assessments.

Study	PREPARATION/RECORDING		SIGNAL PROCESSING			RESULTS	
	Muscles [Preparation Described]	Recording Device [Sampling Frequency]	Filter Passband [Stim artefact filtering]	Rectification	Cycle averaging	Amplitude Normalization	Output Presented
Dy <i>et al.</i> , 2010 [14]	Sol, MG, TA, med hams, VL	Hard wired A/D board and customized labVIEW software	20-1000Hz for resting and standing	P2P amplitude for resting standing. Full wave rectified and peak for stepping.	12 MMR responses	Mean MMR for each muscle was normalized to sol responses as stim electrode placement was determined by optimization of sol response	Quantitative comparison between NI and SCI for MMR resting and standing, and phase-dependant MMR during stepping
	[✓]	[200Hz]	40-500Hz for stepping				
			[✓]				
Emeliannikov <i>et al.</i> , 2016 [15]	RF, BF, TA, and LG	Viasys Viking Select	-	P2P amplitude	-	-	Comparison of MMR, H-Reflex and M-Wave at rest.
	[X]	[NS]	[X]				
Hofstoetter <i>et al.</i> , 2018 [16]	RF, BF, TA and TS	DasyLab 11.0	1. 10-500Hz	P2P amplitude	-	-	Quantitative comparison of 1 <sup>st</sup> and 2 <sup>nd</sup> MMR amplitude for TSS and ESS. Onset offset and duration of 1 <sup>st</sup> and 2 <sup>nd</sup> MMR responses. Normalized response thresholds for TSS and ESS
	[✓]	Codas ADC system	2. 30-700Hz with add. 500Hz low-pass				
		[2048 and 2002Hz]	[✓]				
Hofstoetter <i>et al.</i> , 2019 [17]	RF, BF, TA and Sol	Phoenix multichannel EMG system	10-1000Hz	P2P amplitude	10	Response amplitude of 2 <sup>nd</sup> stimulus in each pair was normalized to the respective 1 <sup>st</sup> for increasing inter-pulse interval (20-5000ms)	Quantitative comparison between NI and SCI for recovery of 2 <sup>nd</sup> PRM as inter-pulse interval increased.
	[✓]	[2048Hz]	[✓]				
Murray and Knikou, 2019 [20]	Sol, MG, PL, TA, med hams, lat ham, RF, and GRC	1401 Plus System	10-1000Hz	Full wave rectified AUC for each TEP response.	15	Responses at increasing intensities were normalized to the associated max response for recruitment curve.	Quantitative comparison of recruitment curve sigmoid parameters, PAD and HD, before and after 60min TSS.
	[X]	[2000Hz]	[X]			Responses at increasing frequency (0.1–1.0 Hz) normalized to response at 0.1Hz for HD.	
						Response amplitude of 2 <sup>nd</sup> stimulus of a pair was normalized to the respective 1 <sup>st</sup> for PAD	
Atkinson <i>et al.</i> , 2020 [26]	RF, VL, med ham, TA, MG, Sol.	MA300 EMG System	-	P2P amplitude	10	Recruitment responses normalized to P2P amplitude at the maximum rate of recruitment (RRmax) within each muscle. CTI: 2 <sup>nd</sup> stimulus in each pair was normalized to the respective 1 <sup>st</sup> for increasing inter-pulse interval (40-160ms)	Quantitative comparison of interlimb conditioning between NI and SCI.
	[X]	[5000Hz]	[X]				

(Continued)

Table 9. (Continued)

Study	PREPARATION/RECORDING		SIGNAL PROCESSING				RESULTS
	Muscles [Preparation Described]	Recording Device [Sampling Frequency]	Filter Passband [Stim artefact filtering]	Rectification	Cycle averaging	Amplitude Normalization	Output Presented
Militskova <i>et al.</i> , 2020 [25]	RF, med ham, TA, sol	Neuro MEP- (Neurosoft, Ivanovo, Russia)	-	-	10	-	Quantitative comparison of SEP latency, threshold and amplitude across (A) 3 stim sites, (B) lying supine vs. standing and (C) pre- post-rehab
	[X]	[5000Hz]	[X]				
Wu <i>et al.</i> , 2020 [23]	APB, ADM, FCR, BB	Customized LabVIEW software (National Instruments USB-6363)	15-2000Hz	-	10	Response amplitude of 2 <sup>nd</sup> stimulus of a pair was normalized to the respective 1 <sup>st</sup> (PAD)	Quantitative comparison recruitment curves across stim configuration. Quantitative comparison of PAD across stim intensity between NI and SCI.
	[X]	[5000Hz]	[X]				
Islam <i>et al.</i> , 2021 [72]	Sol, MG, TA, PL [✓]	Data acquisition card (National Instruments, Austin, TX, USA) [2000Hz]	20–500 Hz [X]	Full wave rectification, linear enveloping via 20Hz low pass filter. Average RMS for entire step cycle.	-	Maximal stepping EMG without stimulation	Quantitative comparison between stim off, stim on (pulse train or single pulses) and post stim of RMS EMG amplitude, mean power frequency, inter/intra limb coordination and h-reflex modulation during stepping in individuals with and without SCI
				P2P amplitude for h-reflex		M-wave for h-reflex	

Abbreviations: ADM; abductor digiti minimi, APB; abductor pollicis brevis, BB; biceps brachii, AUC; area under curve, BR; brachioradialis, CTI; conditioning-test interval, Delt; deltoid, DGO; ED; extensor digitorum, FCR; flexor carpi radialis, FD; flexor digitorum, GRC; gracilis, ham; hamstrings, HD; homosynaptic depression, lat ham; lateral hamstrings, LG; lateral gastrocnemius, med ham; medial hamstrings, MG; medial gastrocnemius, MMR; multisegmental monosynaptic response, P2P; peak-to-peak, PAD; post-activation depression, PL; peroneus longus, Q; quadriceps, RA; rectus abdominis, RF; rectus femoris, RMS; root mean square, Sol; soleus, SEP; spinally evoked potential, TA; Tibialis Anterior, TB; triceps brachii, TFL; tensor fascia lata, TP; tibialis posterior, TS; triceps surae/calf, VL; vastus lateralis.

\*Preparation described refers to a clear description of preparation of the skin before surface electrode application, recording electrode type, orientation, shape and composition as well as interelectrode distance.

†Artifact filtering refers to an attempt made by the authors to account for and remove artifacts contaminating or obscuring the recorded EMG signals such as with the use of a filter.

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stimulated and using this data to then inverse filter surface EMG signal channels [16, 18, 19]. The most popular methods for EMG amplitude processing were the use of full-wave rectification [20, 37, 42], the root mean square [6, 19, 68] and integrated EMG value [36, 41, 43, 69]. Several studies chose only to present raw dynamic EMG data [6, 40, 67].

Only a select number of studies normalized EMG amplitude, four of which were therapeutic investigations [18, 42, 43, 70] and seven of which were neurophysiological investigations [14, 16, 17, 20, 23, 26, 72]. Evoked responses were typically normalized to maximal response at a specific stimulus intensity [20, 26, 42] or when evaluating PAD, the amplitude of the second stimulus of a pair was normalized relative the first [17, 20, 23]. A single investigation recording evoked potentials [14], temporally normalized the responses of pulsed stimulus via a foot switch in an attempt to evaluate if the spinal cord could modulate the evoked responses based on the phase of gait. Using a similar methodology, a more recent paper [72] examined the

**Table 10. A summary of dynamic surface EMG data collection, recording and signal processing.** EMG recording and signal processing for therapeutic studies.

Study	PREPARATION/RECORDING		SIGNAL PROCESSING				RESULTS
	Muscles [Preparation Described]	Recording Device [Sampling Frequency]	Filter Passband [Stim artefact filtration]	Rectification	Cycle averaging	Amplitude Normalization	Output Presented
Hofstoetter <i>et al.</i> , 2013 [6]	Q, Ham, TA, TS	Wired EMS Handels system	10–500 Hz	Raw EMG	-	-	Exemplary raw EMG traces during stepping. Qualitative comparison of stim on/off
	[✓]	[2048Hz]	[X]				
Gerasimenko <i>et al.</i> , 2015 [67]	Sol, MG, TA, med ham, VL	Wired A/D board and customized labVIEW software	10–10,000Hz	-	-	-	Exemplary raw EMG during voluntary movement. Scatter-plot of antagonistic muscle activity patterns. Qualitative description of EMG change during stim.
	[X]	[10,000Hz]	[✓]				
Hofstoetter <i>et al.</i> , 2015 [18]	Q, Ham, TA, TS	EMS-Handels system	10–500 Hz	RMS	10 gait cycles	EMG during stance and swing phase normalized to muscle activity with stim off	Exemplary raw EMG during stepping. Radar chart of RMS during stance and swing. Qualitative comparison of stim on/off
	[✓]	[2048Hz]	[✓]				
Bedi, 2016 [68]	Q, Ham, TA, TP	Neurostim Medicad System	10–500 Hz	RMS	3 repetitions per side	-	Tables of RMS data during voluntary movement before and after stim
	[✓]	[NS]	[X]				
Minassian <i>et al.</i> , 2016 [19]	Q, Ham, TA, TS, TFL	Wired Poly-EMG system (EMS-Handels)	20–500 Hz	RMS	10 gait cycles	-	Within subject quantitative comparison (ANOVA) of RMS data for treadmill speed x hip extension. Exemplary raw EMG during stepping and standing.
	[✓]	[2048Hz]	[✓]				
Gad <i>et al.</i> , 2017 [41]	Sol, MG, TA, med ham, VL	Wired Powerlab and LabChart. Delsys EMG System also mentioned.	10–1000 Hz	Integrated	30 steps	-	Exemplary raw EMG and iEMG during stepping and voluntary movement. Exemplary evoked responses. Qualitative comparison of EMG during passive and active stepping during the intervention.
	[✓]	[10,000Hz]	[✓]		5 evoked responses		
Freyvert <i>et al.</i> , 2018 [44]	FD, ED, BR, BB, TB	Konigsberg EMG system	-	-	9 x 3.5 sec hand-grip Repetition	-	Quantitative comparison of raw EMG amplitude across each test phase Exemplary raw EMG during voluntary hand grip tasks.
	[X]	[NS]	[X]				
Rath <i>et al.</i> , 2018 [37]	RA, Obl, E-T7, E-L3, RF, delt	Wired Powerlab 16/35	10–2000 Hz	Full wave rectification	-	-	Exemplary rectified EMG during trunk movement. Quantitative comparison of mean EMG with stim on/off.
	[X]	[2000Hz]	[✓]				
Gad <i>et al.</i> , 2018 [43]	BB, FD, ED	Wired Powerlab	10–10,000Hz, 60Hz Notch	Integrated	5 evoked responses per intensity	Evoked responses normalized to baseline.	Exemplary raw and iEMG of evoked responses. Exemplary raw EMG traces during voluntary contractions. Quantitative comparison of iEMG response to stim on/off and pre- post-intervention
	[X]	[10,000Hz]	[X]				
Inanici <i>et al.</i> , 2018 [36]	Delt, TB, BB, BR, ED, FD, ADM, thenar muscles	Wired Delsys Bagnoli system	-	Raw EMG	-	-	Exemplary trace of evoked response in thenar muscles compared every 2 weeks across intervention.
	[X]	[1000Hz]	[X]				

(Continued)

Table 10. (Continued)

Study	PREPARATION/RECORDING		SIGNAL PROCESSING				RESULTS
	Muscles [Preparation Described]	Recording Device [Sampling Frequency]	Filter Passband [Stim artefact filtration]	Rectification	Cycle averaging	Amplitude Normalization	Output Presented
Sayenko <i>et al.</i> , 2019 [42]	Sol, TA, VL, med ham	Wired Powerlab	10–2000 Hz	Full wave rectification and RMS	6 evoked responses	EMG during anterior/posterior weight shift normalized to activity during initial position	Exemplary raw EMG during transition and standing. Quantitative comparison of mean EMG between stim on/off, sitting and standing and Quantitative comparison of RMS EMG and motor thresholds across training sessions.
	[X]	[4000Hz]	[✓]				
Al'Joboori <i>et al.</i> , 2020 [69]	Q, TA, ham, gastric	Wired CED Power 1401	10–200 Hz	RMS (0.4s window). Integrated to average level of rectified signal.	3 trials with flex/ext phases	-	Integrated EMG activity pre and post trial temporally normalized to phase of movement
	[X]	[2000Hz]	[X]				
Alam <i>et al.</i> , 2020 [40]	Q, TA, ham, gastric	Wireless, BTS Telemg	[X]	P2P amplitude of MEPs.	-	-	Exemplary raw EMG during sit-to-stand and stand-to-sit task. Intensity response curves for each muscle.
	[X]	[2000Hz]					
Meyer <i>et al.</i> , 2020 [70]	1) RF, VM, ST, TA, MG	1) Wireless Aktos System	1) 10–500 Hz	Noise corrected RMS	1) 20 cycles of each rep ankle movements	Tonic stim responses normalized to the respective values in the tSCS-off condition using z-scores	Exemplary data comparing stim off vs. on - 15Hz/30Hz/50Hz
		[2000Hz]	2) 20 Hz—3000 Hz				
	2) Spinal reflex protocol: bilateral TA	2) Dantec Keypoint Focus Workstation	[✓]		[X]		phase of gait muscle activity
		[6000Hz]					
Shapkova <i>et al.</i> , 2020 [38]	RF, BF, GL, TA	Viasys Viking Select	-	-	-	-	Exemplary trace of H-Reflex and MMR reponse.
	[X]	[2000Hz]	[X]				
Zhang <i>et al.</i> , 2020 [71]	Trap, delt, BB, TB, BR, ECU, FCR, ECR	Wired MA-400 Motion Lab Systems	10–1000 Hz 60Hz Notch	Integrated	3 evoked responses	-	Evoked potential amplitudes pre- and post-stimulation intervention
	[X]	[10200 Hz]	[X]				

**Abbreviations:** BB; biceps brachii, BR; brachioradialis, delt; deltoid, ED; extensor digitorum, E-L3; erector spinae at level of L3, EMG; electromyography, E-T7; erector spinae at level of T7, ECU; extensor carpi ulnaris, ECR; extensor carpi radialis, FCU; flexor carpi ulnaris, FD; flexor digitorum, Ham; hamstrings, iEMG; integrated EMG, med ham; medial hamstrings, MG; medial gastrocnemius, obl; external oblique, P2P; peak-to-peak, PL; Peroneus Longus, Q; quadriceps, RA; rectus abdominis, RF; rectus femoris, RMS; root mean square, sol; soleus, ST; semitendinosus, TA; Tibialis Anterior, TB; triceps brachii, TFL; tensor fascia lata, TP; tibialis posterior, Trap; trapezius, TB; triceps brachii, TS; triceps surae, VL; vastus lateralis.

\* Recording electrodes described refers to a clear description of preparation of the skin before surface electrode application, recording electrode type, orientation, shape and composition as well as interelectrode distance.

† Artifact filtering refers to an attempt made by the authors to account for and remove artifacts contaminating or obscuring the recorded EMG signals such as with the use of a filter or reference EMG electrodes for artefact cancellation.

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impact of tSCS on phase dependant modulation of the H-Reflex during gait. The majority of therapeutic studies recorded dynamic EMG during voluntary movements. However, these signals remained for the most part un-normalized, often with the presentation of exemplary un-rectified EMG traces.

## Safety and adverse events

Of all 25 studies included in this review, only 7 explicitly reported on the presence or absence of adverse events [20, 23, 36, 42, 43, 69, 71]. While some studies made comments on stimulation tolerability and pain levels [18, 38, 40, 44], there were insufficient details to rule out all potential safety issues or complications. Four studies reported the complete absence of adverse events while monitoring vital signs throughout, [20, 36, 43, 71]. The associated recorded events from the other studies included: a modest increase in tone in the 24hrs post treatment, unintentional activation of the micturition reflex and voiding during standing, skin breakage and transient redness [42], as well as discomfort during stimulation at high intensities, asymptomatic variations to heart rate and blood pressure and mild side effects possibly related to cervical stimulation including incidents of light headedness, feeling flushed, nausea, a metallic taste, a sensation of 'sharp' breathing, neck pain, and throat discomfort [23]. None of the adverse events recorded were reported to be consistent across treatment sessions, serious, or long-lasting.

## Discussion

### Summary of findings

This review separated studies utilising tSCS into two broad categories: studies evaluating neurophysiological properties of stimulation at a spinal level and those using tSCS as a therapeutic modality for motor recovery. While publications in both categories have grown in number, the quality of the current evidence base is limited, and a large degree of methodological heterogeneity exists between studies. In particular, extensive variability in stimulation parameters and inconsistent processing and/or presentation of EMG signals make it difficult to draw meaningful conclusions about the effects of tSCS on motor engagement. Efforts should be made in future studies to standardise reporting of muscle activity as well as the electrical parameters of tSCS being administered including electrode dimensions and location, charge polarity, phase duration and stimulation frequency.

### A comparison between neurophysiological and therapeutic investigations

Thus far, neurophysiological investigations have focused on the production of evoked motor potentials and the properties of these responses, such as factors affecting response modulation [14, 25, 26] and the characteristics and reflex contributions of stimulation responses [16, 17, 20]. In neurophysiological investigations, stimulation is generally applied with individual or paired pulses at low frequencies to evaluate an evoked response or attain a motor threshold. As such, eliciting specific PRM reflexes is the likely target of these investigations and the stimulation parameters are selected accordingly. Indeed, all neurophysiological investigations recorded the reflex origin of evoked responses.

In contrast, therapeutic investigations aim to exploit tSCS in order to neuromodulate the spinal cord and augment motor responses produced by individuals with SCI [46]. As a result, tSCS is generally applied for a longer duration in combination with rehabilitative activities. It is likely that these aims are considered in the selection of stimulation parameters and may explain why several therapeutic studies have chosen multiple stimulation sites [36, 37, 40–43, 67, 71], as opposed to targeting specific sites as is typical in neurophysiological assessments. Only four therapeutic investigations [18, 19, 69, 70] attempted to determine the reflex nature of motor responses generated by tSCS.

### Stimulation parameters

**Electrode configuration.** The electrode configuration (size, polarity, location) plays an important role in the electrical field produced by tSCS, and consequently the structures

targeted. One of the criticisms of tSCS, when compared to the epidural alternative, is failure to create a localised electrical field thereby limiting activation selectivity [16, 73]. This review found a lack of consensus regarding electrode configuration, particularly in the cervical region, and limited rationale for selected parameters throughout.

All included studies either placed the cathode posteriorly over the spinal column or used biphasic current with alternating polarity. Other studies investigating use of tSCS on neurologically intact individuals placed the cathode anteriorly with the anode posterior [6, 16–18, 23, 36, 43], but this has not been tested in subjects with SCI. Studies describe using monophasic current (n = 10), biphasic current (n = 7) or both (n = 3). Biphasic current has been noted to reduce risk of tissue damage [74], and Hofstoetter *et al.* [16] found that evoked responses were initiated by the abrupt change of polarity of the biphasic stimulation pulses. Only one included study by Wu *et al.* [23] reported directly comparing different polarities and found biphasic 2-ms or monophasic 1-ms pulses with the cathode posterior elicited larger responses at lower intensities.

Electrode location varies throughout included studies, with regard to both rostral-caudal and mediolateral alignment. The cathodes were positioned paravertebrally (n = 7) or centrally (n = 16) but no conclusions have been drawn on the effects of these different positions. One study in uninjured individuals demonstrated that lateralisation of motor responses in lower limbs (right/left differentiation) can be achieved from the placement of stimulating electrodes ~2cm laterally from the lumbar spinous process [75].

With regard to the spinal levels selected, activation specificity of muscle responses has previously been demonstrated along the rostral-caudal axis of the lumbosacral enlargement in neurologically intact [76, 77] and injured [25] individuals. One study tested for optimal evoked responses at different spinal levels prior to commencing the experimental protocol in order to account for inter-individual variability [26]. The consideration of participant-specific parameter selection could better account for anatomical variation between individuals [26, 78], such as conus medullaris termination level [79] or injury scar tissue, thereby providing more targeted treatment. Several therapeutic studies found superior effects from stimulating the coccygeal level along with a lumbar stimulation site, however the rationale for stimulating a region overlying the termination of the cauda equina remains unclear. Indeed, Roy *et al.* [80] found, using a paired pulse stimulation test, that spinal reflexes were optimally elicited with tSCS when the cathode was over the upper-lumbar vertebrae (L1-L3), and M-waves were optimally elicited with tSCS when the cathode was placed more caudally (L5, S1). If the proposed mechanism for tSCS involves activation of spinal reflex pathways to lower threshold for CPG or voluntary movement, then it would seem important that therapists confirm that the stimulus is transspinally modulated and not just acting as surrogate FES, as may be the case over the coccyx.

For the stimulation of lower limb responses, the anode was placed over the lower abdomen [6, 16–19, 25, 38, 70] and/or the ASIS/iliac crests [14, 20, 26, 40, 41, 67, 72]. This is consistent with previous investigations testing neurologically intact participants [21, 77, 81, 82]. Only one study placed both the cathode and anode in the same plane over the vertebrae [69]. In the cervical region, there was greater variance as anodes were placed superiorly above the sternal notch [23] or inferiorly on the ASIS/iliac crests [36, 41, 44]. Similarly, in cervical tSCS studies of uninjured individuals, anode locations vary between the left acromion [83], upper trapezius and mid clavicle [84], and anterior neck [22]. A previous study investigating the effects of anode position has shown that it is critical for inducing spinal reflexes [85]. Limited human research has further explored the effects of different anode-cathode configurations as a determinant for stimulation outcomes.

**Electrical characteristics.** The voltage that builds up on a skin electrode during a pulse depends on the charge density, i.e., the accumulated charge divided by the electrode area.



Large electrodes have lower charge density and therefore lower pulse voltages for the same current. Vargas Luna *et al.* [86] define a charge density threshold beyond which electro-osmotic effects become significant in the skin conduction mechanism. This may have implications for skin comfort and irritation. The charge density in the reviewed studies ranged between 1.8 to  $53 \mu\text{C}/\text{cm}^2$ . Large electrodes also disperse the current in the underlying tissues which may reduce the likelihood of reaching stimulation thresholds at target neurons while increasing the probability of unwanted collateral stimulation.

For sustained trains of pulses lasting several seconds the accumulated direct current, stimulation of pain receptors and heating effect in the skin must be considered. Monophasic pulse trains produce direct current which can give rise to unwanted electrochemical effects at the electrode site leading to skin irritation and even damage. DC levels higher than  $0.5 \text{ mA}/\text{cm}^2$  at the cathode, and  $1.0 \text{ mA}/\text{cm}^2$  at the anode, are potentially harmful [87]. Even for balanced biphasic waveforms, safety standards such as IEC 60601-2-10 require that the user be advised when the skin current density exceeds  $2 \text{ mA}/\text{cm}^2$ . Neurologically intact subjects with normal skin sensation will usually find this current density quite uncomfortable, and even further rigour must be taken by researchers when working with participants with impaired sensation. FDA guidance documents advise against power densities greater than  $0.25 \text{ W}/\text{cm}^2$  due to the potential heat damage to tissue [88].

The use of carrier frequency in therapeutic studies seems poorly explored or justified at present. The selection of a relatively high frequency carrier waveform in the stimulation pulse is believed to give more efficient signal transfer because the skin-electrode interface impedance has an area-dependent capacitive component which presents less electrical impedance at higher frequency [89–91]. Many studies use a 1 ms pulse with a 10 kHz carrier that has the effect of chopping the pulse into 10 x 50  $\mu\text{s}$  sub-pulses, which reduces the net charge by 50%. It is difficult to compare stimulation effectiveness between studies because of the wide variation in cathode electrode areas used. The included data does not suggest that the current threshold is less when using a carrier, if anything, the opposite may be the case. Meyer *et al.* [69] report a mean current amplitude of 35 mA for a 1 ms biphasic pulse (no carrier) whereas most of the studies using 1 ms monophasic pulses with 10 kHz carriers report much higher peak currents, which may be necessary to compensate for the reduced net charge (Table 7). Modern stimulators are current controlled and automatically adjust their output voltage to ensure the pre-set current is delivered. This ensures electrical losses in the skin do not affect the current that is delivered, perhaps obviating the need for carriers.

**Position of participants.** Different testing conditions may alter tSCS motor responses and studies have demonstrated that spinally-evoked muscle response amplitudes are facilitated or suppressed depending on positional factors and activity phases [5, 14, 92]. A case report by Militskova *et al.* [25] found that spinally evoked response amplitudes were highest in standing, compared to supine, in an individual with SCI. Conversely, a study of 10 healthy participants by Danner *et al.* [93] found that response amplitudes were higher in supine than standing, but that response thresholds were lowest in standing. These studies suggest that results could be affected by position-dependent changes in the electrical field distribution or afferent input altering spinal excitability. Additionally, body position alters the location of the spinal cord within the vertebral canal [94]. Future studies must consider the effects of activity and body positioning and explore conditions that mimic potential clinical scenarios.

## Interpretation of EMG data

The included studies used EMG signals either to quantify the evoked responses at rest, or the level of muscle activity recorded during voluntary movement. Reported methods were

frequently lacking detail, and in some cases not reflective of best practice for the recording, processing, or presentation of surface EMG [55, 95].

In terms of evaluating the magnitude of evoked responses, the majority of studies reported peak-to-peak amplitude of the unrectified EMG signal during a specific time-window after a tSCS pulse was administered. Exceptions to this included Murray and Knikou [20] who quantified the area under the curve of the rectified waveforms, and Dy *et al.* [14] who evaluated phase-dependant modulation of the evoked response during stepping. The reflex nature of evoked responses was most commonly evaluated by quantifying PAD. Other neurophysiological indices calculated from these evoked EMG waveforms included latency and motor threshold.

A notable methodological consideration for EMG recorded during spinal stimulation, and to a greater extent transcutaneously, is the presence of considerable stimulus artefact within the signal. This review identified several filtering approaches including employing high order band-pass filtering of 30-200Hz [67], lower order 6<sup>th</sup> order Butterworth filters with a passband of 30-1000Hz [41] and the implementation of blanking intervals based on stimulus artefact recorded from trunk musculature [18, 19]. Of note was the detailed description by Rath *et al.* [37] of a multi-stage “linear adaptive filter” process which was subsequently utilised by Sayenko *et al.* [42]. However, the efficacy of one approach relative to another for optimising signal: noise ratio has yet to be fully elucidated. Unfortunately, the majority of therapeutic studies (n = 8) did not explicitly report any attempt at filtering non-physiological noise associated with tSCS. A recent study proposed a novel filtration method, Artefact Component Specific Rejection, in order to address this technical challenge and dissociate the net muscle activity from stimulation artefact [49]. This filtration method was employed with data from stroke survivors and has not yet been utilised in other neurologically impaired populations [42].

A recent consensus statement on EMG signal normalisation highlighted its importance for comparing muscle activity between measurement sessions and/or experimental conditions [55]. Despite its well-documented importance, we found only two therapeutic studies that attempted to normalize dynamic EMG signals between sessions or experimental conditions. Many of the therapeutic studies present un-normalized and/or un-rectified exemplary EMG traces, providing some limited qualitative evidence of motor engagement during gravity neutral leg movements [67], assisted robotic stepping, sit-to-stand movement [40] or voluntary handgrip task [44]. Other studies attempted to statistically compare un-normalized EMG signals recorded intermittently over several months [41, 44]. In either case, no meaningful conclusions regarding the efficacy of tSCS to alter muscle activity in patients with SCI can be drawn from this data. Future studies attempting to examine the effect of tSCS therapy on muscle activity during dynamic movements are recommended to present EMG envelopes which have been normalized to an appropriate reference maxima [55] and averaged across multiple cycles or repetitions. Examples of this approach can be seen in the detailed qualitative [96] and quantitative [97] comparisons of EMG recorded from patients with SCI during stepping in the presence or absence of eSCS.

### Quality of included trials

Research investigating the effects of tSCS is an emerging field that predominantly consists of exploratory clinical trials with small sample sizes, and studies were unsurprisingly found to be of poor-to-fair quality using the D&B Checklist. All studies scored poorly on external validity due to a lack of balanced protocols and reporting on recruitment methods. Research in this field is difficult to extrapolate to the population as a whole, as people with SCI differ markedly,

even within the same clinical classifications [98]. Despite this challenge, some investigations with this population have attempted to employ balanced protocols with respect to variables such as AIS classification [69, 98]. Moreover, studies lacked comprehensive accounts of their recruitment protocols and only 7 detailed eligibility criteria, [20, 23, 38, 42–44, 69]. Regardless of inherent recruitment challenges, greater transparency is needed.

Studies also scored poorly for internal validity and there was limited use of randomisation, blinding or sham stimulation. The use of non-randomised designs is common in studying individuals with chronic conditions such as SCI due to inherent methodological, ethical, and practical considerations [99, 100]. Despite this, two studies did employ randomisation using a crossover design [37, 42]. In only three studies, assessors were blinded to the intervention [38, 42, 44]. Sayenko *et al.* [42] was the only study that attempted to use a placebo in the form of two sham stimulation conditions; one on a different location on the spinal cord that did not project to the motor pools assessed and another designed to give the sensation of stimulation without targeting motor responses. While these forms of sham stimulation may not be completely inert in their effects, it demonstrates the only attempt to account for the potential placebo effect of stimulation.

### Limitations of this review

We acknowledge that this review is subject to several potential limitations. Due to the variance in terminology in this field and the lack of standardised nomenclature, it is possible that relevant studies may have been missed by our search strategy. Additionally, our eligibility criteria included studies using EMG outcomes and therefore other studies detailing the tSCS parameters may have been excluded. Finally, study outcomes were not possible to pool due to the heterogeneity of included experiments, and therefore conclusions regarding the optimal stimulation parameters and study protocols cannot be drawn.

### Recommendations and future directions

To fully exploit the capacity of tSCS to facilitate motor activity, future research must directly explore the effects of different parameters to determine the optimal conditions for desired motor outcomes. Greater justification for the selection of therapeutic stimulation parameters needs to be provided by experiments that bridge the gap in our understanding of parameter optimisation, clinical application, and the mechanisms that promote motor recovery. The quality of future trials would be improved with better reporting of recruitment methods and intervention protocols and with the application of techniques such as randomisation and sham-stimulation. The presence or absence of adverse events must be explicitly detailed to provide a larger evidence base supporting the safety and feasibility. Finally, studies must also include improve the methodological rigour for data collection, processing and reporting in particular of EMG data.

### Conclusions

The results of this systematic review indicate that studies investigating the effects of tSCS interventions for individuals with SCI face both methodological and measurement deficiencies. While initial investigations have improved our understanding of the neurophysiological impact of this technology and demonstrated its feasibility in motor rehabilitation, greater homogeneity in the reporting of stimulation parameters and outcome measurement will be required to pool cumulative outcomes from small sample sizes. A higher quality of studies will be needed to demonstrate conclusive evidence on the standardised application and uses of tSCS.

## Supporting information

**S1 Checklist. PRISMA 2009 checklist.**

(DOCX)

**S1 Appendix. Search strategy and terminology.**

(DOCX)

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## References

1. Gerasimenko Y, Gorodnichev R, Moshonkina T, Sayenko D, Gad P, Reggie Edgerton V. Transcutaneous electrical spinal-cord stimulation in humans. *Annals of Physical & Rehabilitation Medicine*. 2015; 58(4):225–31.
2. Minassian K, Persy I, Rattay F, Dimitrijevic MR, Hofer C, Kern H. Posterior root-muscle reflexes elicited by transcutaneous stimulation of the human lumbosacral cord. *Muscle and Nerve*. 2007; 35(3):327–36. <https://doi.org/10.1002/mus.20700> PMID: 17117411
3. Minassian K, Persy I, Rattay F, Pinter MM, Kern H, Dimitrijevic MR. Human lumbar cord circuitries can be activated by extrinsic tonic input to generate locomotor-like activity. *Human Movement Science*. 2007; 26(2):275–95. <https://doi.org/10.1016/j.humov.2007.01.005> PMID: 17343947
4. Hofstoetter US, Minassian K, Hofer C, Mayr W, Rattay F, Dimitrijevic MR. Modification of reflex responses to lumbar posterior root stimulation by motor tasks in healthy subjects. *Artif Organs*. 2008; 32(8):644–8. <https://doi.org/10.1111/j.1525-1594.2008.00616.x> PMID: 18782137
5. Courtine G, Harkema SJ, Dy CJ, Gerasimenko YP, Dyhre-Poulsen P. Modulation of multisegmental monosynaptic responses in a variety of leg muscles during walking and running in humans. *J Physiol*. 2007; 582(Pt 3):1125–39. <https://doi.org/10.1113/jphysiol.2007.128447> PMID: 17446226
6. Hofstoetter US, Hofer C, Kern H, Danner SM, Mayr W, Dimitrijevic MR, et al. Effects of transcutaneous spinal cord stimulation on voluntary locomotor activity in an incomplete spinal cord injured individual. *Biomed Tech (Berl)*. 2013; 58 Suppl 1.
7. Danner SM, Hofstoetter US, Ladenbauer J, Rattay F, Minassian K. Can the human lumbar posterior columns be stimulated by transcutaneous spinal cord stimulation? A modeling study. *Artif Organs*. 2011; 35(3):257–62. <https://doi.org/10.1111/j.1525-1594.2011.01213.x> PMID: 21401670
8. Ladenbauer J, Minassian K, Hofstoetter U, Dimitrijevic M, Rattay F. Stimulation of the Human Lumbar Spinal Cord With Implanted and Surface Electrodes: A Computer Simulation Study. *Neural Systems and Rehabilitation Engineering, IEEE Transactions on*. 2010; 18:637–45. <https://doi.org/10.1109/TNSRE.2010.2054112> PMID: 21138794

9. Minassian K, Persy I, Rattay F, Dimitrijevic MR, Hofer C, Kern H. Posterior root-muscle reflexes elicited by transcutaneous stimulation of the human lumbosacral cord. *Muscle & nerve*. 2007; 35(3):327–36. <https://doi.org/10.1002/mus.20700> PMID: 17117411
10. Knikou M. Neurophysiological characterization of transspinal evoked potentials in human leg muscles. *Bioelectromagnetics*. 2013; 34(8):630–40. <https://doi.org/10.1002/bem.21808> PMID: 24115026
11. Einhorn J, Li A, Hazan R, Knikou M. Cervicothoracic multisegmental transspinal evoked potentials in humans. *PLoS ONE*. 2013; 8(10):e76940. <https://doi.org/10.1371/journal.pone.0076940> PMID: 24282479
12. Krenn M, Hofstoetter US, Danner SM, Minassian K, Mayr W. Multi-Electrode Array for Transcutaneous Lumbar Posterior Root Stimulation. *Artificial organs*. 2015; 39(10):834–40. <https://doi.org/10.1111/aor.12616> PMID: 26471133
13. Sayenko DG, Angeli C, Harkema SJ, Edgerton VR, Gerasimenko YP. Neuromodulation of evoked muscle potentials induced by epidural spinal-cord stimulation in paralyzed individuals. *J Neurophysiol*. 2014; 111(5):1088–99. <https://doi.org/10.1152/jn.00489.2013> PMID: 24335213
14. Dy CJ, Gerasimenko YP, Edgerton VR, Dyhre-Poulsen P, Courtine G, Harkema SJ. Phase-dependent modulation of percutaneously elicited multisegmental muscle responses after spinal cord injury. *J Neurophysiol*. 2010; 103(5):2808–20. <https://doi.org/10.1152/jn.00316.2009> PMID: 20357075
15. Emeliannikov DV, Shapkova EY, Moshonkina TR, Gerasimenko YP. Evaluation of motor neuron excitability in lumbosacral spinal cord: Transcutaneous spinal cord stimulation as compared to H-reflex. *Fiziologija Cheloveka*. 2016; 42(3):32–6. PMID: 29446609
16. Hofstoetter US, Freundl B, Binder H, Minassian K. Common neural structures activated by epidural and transcutaneous lumbar spinal cord stimulation: Elicitation of posterior root-muscle reflexes. *PloS one*. 2018; 13(1):e0192013. <https://doi.org/10.1371/journal.pone.0192013> PMID: 29381748
17. Hofstoetter US, Freundl B, Binder H, Minassian K. Recovery cycles of posterior root-muscle reflexes evoked by transcutaneous spinal cord stimulation and of the H reflex in individuals with intact and injured spinal cord. *PLoS ONE*. 2019; 14(12):e0227057. <https://doi.org/10.1371/journal.pone.0227057> PMID: 31877192
18. Hofstoetter US, Krenn M, Danner SM, Hofer C, Kern H, McKay WB, et al. Augmentation of Voluntary Locomotor Activity by Transcutaneous Spinal Cord Stimulation in Motor-Incomplete Spinal Cord-Injured Individuals. *Artif Organs*. 2015; 39(10):E176–86. <https://doi.org/10.1111/aor.12615> PMID: 26450344
19. Minassian K, Hofstoetter US, Danner SM, Mayr W, Bruce JA, McKay WB, et al. Spinal Rhythm Generation by Step-Induced Feedback and Transcutaneous Posterior Root Stimulation in Complete Spinal Cord-Injured Individuals. *Neurorehabil Neural Repair*. 2016; 30(3):233–43. <https://doi.org/10.1177/1545968315591706> PMID: 26089308
20. Murray LM, Knikou M. Transspinal stimulation increases motoneuron output of multiple segments in human spinal cord injury. *PloS one*. 2019; 14(3):e0213696. <https://doi.org/10.1371/journal.pone.0213696> PMID: 30845251
21. Andrews JC, Stein RB, Roy FD. Post-activation depression in the human soleus muscle using peripheral nerve and transcutaneous spinal stimulation. *Neuroscience Letters*. 2015; 589:144–9. <https://doi.org/10.1016/j.neulet.2015.01.041> PMID: 25600855
22. Milosevic M, Masugi Y, Sasaki A, Sayenko DG, Nakazawa K. On the reflex mechanisms of cervical transcutaneous spinal cord stimulation in human subjects. *J Neurophysiol*. 2019; 121(5):1672–9. <https://doi.org/10.1152/jn.00802.2018> PMID: 30840527
23. Wu YK, Levine JM, Wecht JR, Maher MT, LiMonta JM, Saeed S, et al. Posteroanterior cervical transcutaneous spinal stimulation targets ventral and dorsal nerve roots. *Clin Neurophysiol*. 2020; 131(2):451–60. <https://doi.org/10.1016/j.clinph.2019.11.056> PMID: 31887616
24. Zheng Y, Hu X. Muscle activation pattern elicited through transcutaneous stimulation near the cervical spinal cord. *J Neural Eng*. 2020; 17(1):016064. <https://doi.org/10.1088/1741-2552/ab5e09> PMID: 31791027
25. Militskova A, Mukhametova E, Fatykhova E, Sharifullin S, Cuellar CA, Calvert JS, et al. Supraspinal and Afferent Signaling Facilitate Spinal Sensorimotor Network Excitability After Discomplete Spinal Cord Injury: A Case Report. *Front Neurosci*. 2020; 14:552. <https://doi.org/10.3389/fnins.2020.00552> PMID: 32655351
26. Atkinson DA, Sayenko DG, D'Amico JM, Mink A, Lorenz DJ, Gerasimenko YP, et al. Interlimb conditioning of lumbosacral spinally evoked motor responses after spinal cord injury. *Clin Neurophysiol*. 2020; 131(7):1519–32. <https://doi.org/10.1016/j.clinph.2020.03.021> PMID: 32403065
27. Angeli CA, Edgerton VR, Gerasimenko YP, Harkema SJ. Altering spinal cord excitability enables voluntary movements after chronic complete paralysis in humans. *Brain*. 2014; 137(Pt 5):1394–409. <https://doi.org/10.1093/brain/awu038> PMID: 24713270

28. Young W. Electrical stimulation and motor recovery. *Cell transplantation*. 2015; 24(3):429–46. <https://doi.org/10.3727/096368915X686904> PMID: 25646771
29. Islam MA, Zaaya M, Comiskey E, Demetrio J, O'Keefe A, Palazzo N, et al. Modulation of soleus H-reflex excitability following cervical transspinal conditioning stimulation in humans. *Neurosci Lett*. 2020; 732:135052. <https://doi.org/10.1016/j.neulet.2020.135052> PMID: 32439478
30. Taccola G, Sayenko D, Gad P, Gerasimenko Y, Edgerton VR. And yet it moves: Recovery of volitional control after spinal cord injury. *Progress in neurobiology*. 2018; 160:64–81. <https://doi.org/10.1016/j.pneurobio.2017.10.004> PMID: 29102670
31. Hofstoetter US, Knikou M, Guertin PA, Minassian K. Probing the Human Spinal Locomotor Circuits by Phasic Step-Induced Feedback and by Tonic Electrical and Pharmacological Neuromodulation. *Curr Pharm Des*. 2017; 23(12):1805–20. <https://doi.org/10.2174/1381612822666161214144655> PMID: 27981912
32. Minassian K, Hofstoetter US, Dzeladini F, Guertin PA, Ijspeert A. The Human Central Pattern Generator for Locomotion: Does It Exist and Contribute to Walking? *Neuroscientist*. 2017; 23(6):649–63. <https://doi.org/10.1177/1073858417699790> PMID: 28351197
33. Dietz V, Fouad K, Bastiaanse CM. Neuronal coordination of arm and leg movements during human locomotion. *Eur J Neurosci*. 2001; 14(11):1906–14. <https://doi.org/10.1046/j.0953-816x.2001.01813.x> PMID: 11860485
34. Laliberte AM, Goltash S, Lalonde NR, Bui TV. Propriospinal Neurons: Essential Elements of Locomotor Control in the Intact and Possibly the Injured Spinal Cord. *Frontiers in Cellular Neuroscience*. 2019; 13(512). <https://doi.org/10.3389/fncel.2019.00512> PMID: 31798419
35. Eblen-Zajjur AA, Sandkühler J. Synchronicity of nociceptive and non-nociceptive adjacent neurons in the spinal dorsal horn of the rat: stimulus-induced plasticity. *Neuroscience*. 1997; 76(1):39–54. [https://doi.org/10.1016/s0306-4522\(96\)00286-2](https://doi.org/10.1016/s0306-4522(96)00286-2) PMID: 8971757
36. Inanici F, Samejima S, Gad P, Edgerton VR, Hofstetter CP, Moritz CT. Transcutaneous Electrical Spinal Stimulation Promotes Long-Term Recovery of Upper Extremity Function in Chronic Tetraplegia. *IEEE Trans Neural Syst Rehabil Eng*. 2018; 26(6):1272–8. <https://doi.org/10.1109/TNSRE.2018.2834339> PMID: 29877852
37. Rath M, Vette AH, Ramasubramaniam S, Li K, Burdick J, Edgerton VR, et al. Trunk Stability Enabled by Noninvasive Spinal Electrical Stimulation after Spinal Cord Injury. *J Neurotrauma*. 2018; 35(21):2540–53. <https://doi.org/10.1089/neu.2017.5584> PMID: 29786465
38. Shapkova EY, Pismennaya EV, Emelyannikov DV, Ivanenko Y. Exoskeleton Walk Training in Paralyzed Individuals Benefits From Transcutaneous Lumbar Cord Tonic Electrical Stimulation. *Front Neurosci*. 2020; 14:416. <https://doi.org/10.3389/fnins.2020.00416> PMID: 32528238
39. van den Brand R, Mignardot JB, von Zitzewitz J, Le Goff C, Fumeaux N, Wagner F, et al. Neuroprosthetic technologies to augment the impact of neurorehabilitation after spinal cord injury. *Annals of physical and rehabilitation medicine*. 2015; 58(4):232–7. <https://doi.org/10.1016/j.rehab.2015.04.003> PMID: 26100230
40. Alam M, Ling YT, Wong AYL, Zhong H, Edgerton VR, Zheng Y-P. Reversing 21 years of chronic paralysis via non-invasive spinal cord neuromodulation: a case study. *Annals of clinical and translational neurology*. 2020; 7(5):829–38. <https://doi.org/10.1002/acn3.51051> PMID: 32436278
41. Gad P, Gerasimenko Y, Zdunowski S, Turner A, Sayenko D, Lu DC, et al. Weight Bearing Over-ground Stepping in an Exoskeleton with Non-invasive Spinal Cord Neuromodulation after Motor Complete Paraplegia. *Front Neurosci*. 2017; 11:333. <https://doi.org/10.3389/fnins.2017.00333> PMID: 28642680
42. Sayenko DG, Rath M, Ferguson AR, Burdick JW, Havton LA, Edgerton VR, et al. Self-Assisted Standing Enabled by Non-Invasive Spinal Stimulation after Spinal Cord Injury. *J Neurotrauma*. 2019; 36(9):1435–50. <https://doi.org/10.1089/neu.2018.5956> PMID: 30362876
43. Gad P, Lee S, Terrafranca N, Zhong H, Turner A, Gerasimenko Y, et al. Non-Invasive Activation of Cervical Spinal Networks after Severe Paralysis. *J Neurotrauma*. 2018; 35(18):2145–58. <https://doi.org/10.1089/neu.2017.5461> PMID: 29649928
44. Freyvert Y, Yong NA, Morikawa E, Zdunowski S, Sarino ME, Gerasimenko Y, et al. Engaging cervical spinal circuitry with non-invasive spinal stimulation and buspirone to restore hand function in chronic motor complete patients. *Sci Rep*. 2018; 8(1):15546. <https://doi.org/10.1038/s41598-018-33123-5> PMID: 30341390
45. Inanici F, Brighton LN, Samejima S, Hofstetter CP, Moritz CT. Transcutaneous Spinal Cord Stimulation Restores Hand and Arm Function after Spinal Cord Injury. *IEEE Transactions on Neural Systems and Rehabilitation Engineering*. 2021; 29:310–9. <https://doi.org/10.1109/TNSRE.2021.3049133> PMID: 33400652

46. Megía García A, Serrano-Muñoz D, Taylor J, Avendaño-Coy J, Gómez-Soriano J. Transcutaneous Spinal Cord Stimulation and Motor Rehabilitation in Spinal Cord Injury: A Systematic Review. *Neurorehabil Neural Repair*. 2020; 34(1):3–12. <https://doi.org/10.1177/1545968319893298> PMID: 31858871
47. McHugh C, Taylor C, Mockler D, Fleming N. Epidural spinal cord stimulation for motor recovery in spinal cord injury: A systematic review. *NeuroRehabilitation*. 2021. <https://doi.org/10.3233/NRE-210093> Epub ahead of print. PMID: 33967072.
48. Pilkar R, Momeni K, Ramanujam A, Ravi M, Garbarini E, Forrest GF. Use of Surface EMG in Clinical Rehabilitation of Individuals With SCI: Barriers and Future Considerations. *Frontiers in neurology*. 2020; 11:578559–. <https://doi.org/10.3389/fneur.2020.578559> PMID: 33408680
49. Kim M, Moon Y, Hunt J, McKenzie KA, Horin A, McGuire M, et al. A Novel Technique to Reject Artifact Components for Surface EMG Signals Recorded During Walking With Transcutaneous Spinal Cord Stimulation: A Pilot Study. *Frontiers in Human Neuroscience*. 2021; 15(283):660583. <https://doi.org/10.3389/fnhum.2021.660583> PMID: 34149379
50. Barss TS, Parhizi B, Mushahwar VK. Transcutaneous spinal cord stimulation of the cervical cord modulates lumbar networks. *J Neurophysiol*. 2020; 123(1):158–66. <https://doi.org/10.1152/jn.00433.2019> PMID: 31747338
51. Benavides FD, Jo HJ, Lundell H, Edgerton VR, Gerasimenko Y, Perez MA. Cortical and Subcortical Effects of Transcutaneous Spinal Cord Stimulation in Humans with Tetraplegia. *J Neurosci*. 2020; 40(13):2633–43. <https://doi.org/10.1523/JNEUROSCI.2374-19.2020> PMID: 31996455
52. Taccola G, Gad P, Culaclii S, Ichiyama RM, Liu W, Reggie Edgerton V. Using EMG to deliver lumbar dynamic electrical stimulation to facilitate cortico-spinal excitability. *Brain Stimulation*. 2020; 13(1):20–34. <https://doi.org/10.1016/j.brs.2019.09.013> PMID: 31585723
53. Wagner FB, Mignardot JB, Le Goff-Mignardot CG, Demesmaeker R, Komi S, Capogrosso M, et al. Targeted neurotechnology restores walking in humans with spinal cord injury. *Nature*. 2018; 563(7729):65–71. <https://doi.org/10.1038/s41586-018-0649-2> PMID: 30382197
54. De Luca CJ. The use of surface electromyography in biomechanics. *Journal of applied biomechanics*. 1997; 13(2):135–63.
55. Besomi M, Hodges PW, Clancy EA, Van Dieën J, Hug F, Lowery M, et al. Consensus for experimental design in electromyography (CEDE) project: Amplitude normalization matrix. *Journal of Electromyography and Kinesiology*. 2020; 53:102438. <https://doi.org/10.1016/j.jelekin.2020.102438> PMID: 32569878
56. McKay WB, Ovechkin AV, Vitaz TW, Terson de Paleville DGL, Harkema SJ. Long-lasting involuntary motor activity after spinal cord injury. *Spinal cord*. 2011; 49(1):87–93. <https://doi.org/10.1038/sc.2010.73> PMID: 20585326
57. Moher D, Shamseer L, Clarke M, Ghersi D, Liberati A, Petticrew M, et al. Preferred reporting items for systematic review and meta-analysis protocols (PRISMA-P) 2015 statement. *Syst Rev*. 2015; 4(1):1. <https://doi.org/10.1186/2046-4053-4-1> PMID: 25554246
58. Downs SH, Black N. The feasibility of creating a checklist for the assessment of the methodological quality both of randomised and non-randomised studies of health care interventions. *J Epidemiol Community Health*. 1998; 52(6):377–84. <https://doi.org/10.1136/jech.52.6.377> PMID: 9764259
59. Lam T, Eng JJ, Wolfe DL, Hsieh JT, Whittaker M. A systematic review of the efficacy of gait rehabilitation strategies for spinal cord injury. *Top Spinal Cord Inj Rehabil*. 2007; 13(1):32–57. <https://doi.org/10.1310/sci1301-32> PMID: 22915835
60. Ibitoye MO, Hamzaid NA, Hasnan N, Abdul Wahab AK, Davis GM. Strategies for Rapid Muscle Fatigue Reduction during FES Exercise in Individuals with Spinal Cord Injury: A Systematic Review. *PloS one*. 2016; 11(2):e0149024. <https://doi.org/10.1371/journal.pone.0149024> PMID: 26859296
61. Silverman SR, Schertz LA, Yuen HK, Lowman JD, Bickel CS. Systematic review of the methodological quality and outcome measures utilized in exercise interventions for adults with spinal cord injury. *Spinal Cord*. 2012; 50(10):718–27. <https://doi.org/10.1038/sc.2012.78> PMID: 22777488
62. Eng JJ, Teasell R, Miller WC, Wolfe DL, Townson AF, Aubut JA, et al. Spinal Cord Injury Rehabilitation Evidence: Methods of the SCIRE Systematic Review. *Top Spinal Cord Inj Rehabil*. 2007; 13(1):1–10. <https://doi.org/10.1310/sci1301-1> PMID: 22767989
63. Saunders LD, Soomro GM, Buckingham J, Jamtvedt G, Raina P. Assessing the methodological quality of nonrandomized intervention studies. *West J Nurs Res*. 2003; 25(2):223–37. <https://doi.org/10.1177/0193945902250039> PMID: 12666645
64. Deeks JJ, Dinnes J, D'Amico R, Sowden AJ, Sakaravitch C, Song F, et al. Evaluating non-randomised intervention studies. *Health Technol Assess*. 2003; 7(27):iii–x, 1–173. <https://doi.org/10.3310/hta7270> PMID: 14499048

65. Faul F, Erdfelder E, Lang A-G, Buchner A. G\*Power 3: A flexible statistical power analysis program for the social, behavioral, and biomedical sciences. *Behavior Research Methods*. 2007; 39(2):175–91. <https://doi.org/10.3758/bf03193146> PMID: 17695343
66. Hamdan PNF, Hamzaid NA, Razak NA, Hasnan N. Contributions of the Cyathlon championship to the literature on functional electrical stimulation cycling among individuals with spinal cord injury: A bibliometric review. *Journal of Sport and Health Science*. 2020. <https://doi.org/10.1016/j.jshs.2020.10.002> Epub ahead of print. PMID: 33068748.
67. Gerasimenko YP, Lu DC, Modaber M, Zdunowski S, Gad P, Sayenko DG, et al. Noninvasive Reactivation of Motor Descending Control after Paralysis. *J Neurotrauma*. 2015; 32(24):1968–80. <https://doi.org/10.1089/neu.2015.4008> PMID: 26077679
68. Bedi P. Tapping the Neural Circuitry: Surface Spinal Stimulation in Spinal Cord Injury: a Case Report. *Journal of Exercise Science and Physiotherapy*. 2016;12.
69. Al'Joboori Y, Massey SJ, Knight SL, Donaldson NDN, Duffell LD. The Effects of Adding Transcutaneous Spinal Cord Stimulation (tSCS) to Sit-To-Stand Training in People with Spinal Cord Injury: A Pilot Study. *Journal of Clinical Medicine*. 2020; 9(9):2765 <https://doi.org/10.3390/jcm9092765> PMID: 32858977
70. Meyer C, Hofstoetter US, Hubli M, Hassani RH, Rinaldo C, Curt A, et al. Immediate Effects of Transcutaneous Spinal Cord Stimulation on Motor Function in Chronic, Sensorimotor Incomplete Spinal Cord Injury. *Journal of clinical medicine*. 2020; 9(11):3541. <https://doi.org/10.3390/jcm9113541> PMID: 33147884
71. Zhang F, Momeni K, Ramanujam A, Ravi M, Carnahan J, Kirshblum S, et al. Cervical Spinal Cord Transcutaneous Stimulation Improves Upper Extremity and Hand Function in People With Complete Tetraplegia: A Case Study. *IEEE Trans Neural Syst Rehabil Eng*. 2020; 28(12):3167–74. <https://doi.org/10.1109/TNSRE.2020.3048592> PMID: 33382659
72. Islam MA, Pulverenti TS, Knikou M. Neuronal Actions of Transspinal Stimulation on Locomotor Networks and Reflex Excitability During Walking in Humans With and Without Spinal Cord Injury. *Front Hum Neurosci*. 2021; 15:620414. <https://doi.org/10.3389/fnhum.2021.620414> PMID: 33679347
73. Duffell LD, Donaldson NdN. A Comparison of FES and SCS for Neuroplastic Recovery After SCI: Historical Perspectives and Future Directions. *Frontiers in neurology*. 2020; 11:607–. <https://doi.org/10.3389/fneur.2020.00607> PMID: 32714270
74. Hofmann L, Ebert M, Tass P, Hauptmann C. Modified Pulse Shapes for Effective Neural Stimulation. *Frontiers in Neuroengineering*. 2011; 4(9). <https://doi.org/10.3389/fneng.2011.00009> PMID: 22007167
75. Calvert JS, Manson GA, Grahn PJ, Sayenko DG. Preferential activation of spinal sensorimotor networks via lateralized transcutaneous spinal stimulation in neurologically intact humans. *J Neurophysiol*. 2019; 122(5):2111–8. <https://doi.org/10.1152/jn.00454.2019> PMID: 31553681
76. Krenn M, Toth A, Danner SM, Hofstoetter US, Minassian K, Mayr W. Selectivity of transcutaneous stimulation of lumbar posterior roots at different spinal levels in humans. *Biomed Tech (Berl)*. 2013; 58 Suppl 1. <https://doi.org/10.1515/bmt-2013-4010> PMID: 24042628
77. Sayenko DG, Atkinson DA, Dy CJ, Gurley KM, Smith VL, Angeli C, et al. Spinal segment-specific transcutaneous stimulation differentially shapes activation pattern among motor pools in humans. *Journal of Applied Physiology*. 2015; 118(11):1364–74. <https://doi.org/10.1152/jappphysiol.01128.2014> PMID: 25814642
78. Troni W, Di Sapio A, Berra E, Duca S, Merola A, Sperli F, et al. A methodological reappraisal of non invasive high voltage electrical stimulation of lumbosacral nerve roots. *Clinical Neurophysiology*. 2011; 122(10):2071–80. <https://doi.org/10.1016/j.clinph.2011.03.002> PMID: 21458371
79. Saifuddin A, Burnett SJ, White J. The variation of position of the conus medullaris in an adult population. A magnetic resonance imaging study. *Spine (Phila Pa 1976)*. 1998; 23(13):1452–6.
80. Roy RR, Harkema SJ, Edgerton VR. Basic concepts of activity-based interventions for improved recovery of motor function after spinal cord injury. *Arch Phys Med Rehabil*. 2012; 93(9):1487–97. <https://doi.org/10.1016/j.apmr.2012.04.034> PMID: 22920448
81. Kaneko N, Masugi Y, Usuda N, Yokoyama H, Nakazawa K. Muscle-Specific Modulation of Spinal Reflexes in Lower-Limb Muscles during Action Observation with and without Motor Imagery of Walking. *Brain Sciences*. 2019; 9(12):21.
82. Kitano K, Koceja DM. Spinal reflex in human lower leg muscles evoked by transcutaneous spinal cord stimulation. *J Neurosci Methods*. 2009; 180(1):11–5. <https://doi.org/10.1016/j.jneumeth.2009.03.006> PMID: 19427537
83. Sabbahi MA, Sengul YS. Cervical multisegmental motor responses in healthy subjects. *Spinal Cord*. 2012; 50(6):432–9. <https://doi.org/10.1038/sc.2011.166> PMID: 22249322



84. Sabbahi MA, Uzun S, Bittar FO, Sengul Y. Similarities and differences in cervical and thoracolumbar multisegmental motor responses and the combined use for testing spinal circuitries. *Journal of Spinal Cord Medicine*. 2014; 37(4):401–13. <https://doi.org/10.1179/2045772313Y.0000000157> PMID: 24621020
85. Masugi Y, Obata H, Nakazawa K. Effects of anode position on the responses elicited by transcutaneous spinal cord stimulation. *Conf Proc IEEE Eng Med Biol Soc*. 2017; 2017:1114–7. <https://doi.org/10.1109/EMBC.2017.8037024> PMID: 29060070
86. Vargas Luna JL, Krenn M, Cortés Ramírez JA, Mayr W. Dynamic Impedance Model of the Skin-Electrode Interface for Transcutaneous Electrical Stimulation. *PLOS ONE*. <https://doi.org/10.1371/journal.pone.0125609>; 10(5):e0125609. PMID: 25942010
87. Bélanger A. *Therapeutic Electrophysical Agents: Evidence Behind Practice*: Wolters Kluwer Health/Lippincott Williams & Wilkins; 2010.
88. Administration UFaD. *Guidance document for powered muscle stimulator 501 (k) s*. In: FDA, editor. Washington 1999.
89. Currier DP, Nelson RM, Hayes KW. *Clinical Electrotherapy*: Appleton & Lange; 1999.
90. Manson GA, Calvert JS, Ling J, Tychon B, Ali A, Sayenko DG. The relationship between maximum tolerance and motor activation during transcutaneous spinal stimulation is unaffected by the carrier frequency or vibration. *Physiological Reports*. 2020; 8(5):e14397. <https://doi.org/10.14814/phy2.14397> PMID: 32170844
91. Ward AR, Robertson VJ, Makowski RJ. Optimal frequencies for electric stimulation using medium-frequency alternating current. *Arch Phys Med Rehabil*. 2002; 83(7):1024–7. <https://doi.org/10.1053/apmr.2002.33116> PMID: 12098166
92. Masugi Y, Kawashima N, Inoue D, Nakazawa K. Effects of movement-related afferent inputs on spinal reflexes evoked by transcutaneous spinal cord stimulation during robot-assisted passive stepping. *Neuroscience Letters*. 2016; 627:100–6. <https://doi.org/10.1016/j.neulet.2016.05.047> PMID: 27235576
93. Danner SM, Krenn M, Hofstoetter US, Toth A, Mayr W, Minassian K. Body position influences which neural structures are recruited by lumbar transcutaneous spinal cord stimulation. *PLoS ONE*. 2016; 11(1). <https://doi.org/10.1371/journal.pone.0147479> PMID: 26797502
94. Ranger MR, Irwin GJ, Bunbury KM, Peutrell JM. Changing body position alters the location of the spinal cord within the vertebral canal: a magnetic resonance imaging study. *Br J Anaesth*. 2008; 101(6):804–9. <https://doi.org/10.1093/bja/aen295> PMID: 18936040
95. Hermens HJ, Freriks B, Disselhorst-Klug C, Rau G. Development of recommendations for SEMG sensors and sensor placement procedures. *J Electromyogr Kinesiol*. 2000; 10(5):361–74. [https://doi.org/10.1016/s1050-6411\(00\)00027-4](https://doi.org/10.1016/s1050-6411(00)00027-4) PMID: 11018445
96. Huang H, He J, Herman R, Carhart MR. Modulation effects of epidural spinal cord stimulation on muscle activities during walking. *IEEE Trans Neural Syst Rehabil Eng*. 2006; 14(1):14–23. <https://doi.org/10.1109/TNSRE.2005.862694> PMID: 16562627
97. Gill ML, Grahn PJ, Calvert JS, Linde MB, Lavrov IA, Strommen JA, et al. Neuromodulation of lumbosacral spinal networks enables independent stepping after complete paraplegia. *Nature Medicine*. 2018; 24(11):1677–82. <https://doi.org/10.1038/s41591-018-0175-7> PMID: 30250140
98. Sherwood AM, McKay WB, Dimitrijević MR. Motor control after spinal cord injury: assessment using surface EMG. *Muscle Nerve*. 1996; 19(8):966–79. [https://doi.org/10.1002/\(SICI\)1097-4598\(199608\)19:8<966::AID-MUS5>3.0.CO;2-6](https://doi.org/10.1002/(SICI)1097-4598(199608)19:8<966::AID-MUS5>3.0.CO;2-6) PMID: 8756162
99. Ginis KA, Hicks AL. Exercise research issues in the spinal cord injured population. *Exerc Sport Sci Rev*. 2005; 33(1):49–53. PMID: 15640721
100. Wilbanks S, Schertz LA, Yuen H, Lowman J, Bickel C. Systematic review of the methodological quality and outcome measures utilized in exercise interventions for adults with spinal cord injury. *Spinal cord*. 2012; 50:718–27. <https://doi.org/10.1038/sc.2012.78> PMID: 22777488