

Review

In Silico Prediction of Steroids and Triterpenoids as Potential Regulators of Lipid Metabolism

Valery M. Dembitsky 

Centre for Applied Research, Innovation and Entrepreneurship, Lethbridge College, 3000 College Drive South, Lethbridge, AB T1K 1L6, Canada; valery.dembitsky@lethbridgecollege.ca; Fax: +1-888-858-8517

Abstract: This review focuses on a rare group of steroids and triterpenoids that share common properties as regulators of lipid metabolism. This group of compounds is divided by the type of chemical structure, and they represent: aromatic steroids, steroid phosphate esters, highly oxygenated steroids such as steroid endoperoxides and hydroperoxides, α,β -epoxy steroids, and secosteroids. In addition, subgroups of carbon-bridged steroids, neo steroids, miscellaneous steroids, as well as synthetic steroids containing heteroatoms S (*epithio steroids*), Se (*selenium steroids*), Te (*tellura steroids*), and At (*astato steroids*) were presented. Natural steroids and triterpenoids have been found and identified from various sources such as marine sponges, soft corals, starfish, and other marine invertebrates. In addition, this group of rare lipids is found in fungi, fungal endophytes, and plants. The pharmacological profile of the presented steroids and triterpenoids was determined using the well-known computer program PASS, which is currently available online for all interested scientists and pharmacologists and is currently used by research teams from more than 130 countries of the world. Our attention has been focused on the biological activities of steroids and triterpenoids associated with the regulation of cholesterol metabolism and related processes such as anti-hyperlipoproteinemic activity, as well as the treatment of atherosclerosis, lipoprotein disorders, or inhibitors of cholesterol synthesis. In addition, individual steroids and triterpenoids were identified that demonstrated rare or unique biological activities such as treating neurodegenerative diseases, Alzheimer's, and Parkinson's diseases with a high degree of certainty over 95 percent. For individual steroids or triterpenoids or a group of compounds, 3D drawings of their predicted biological activities are presented.

Keywords: steroids; triterpenoids; hormones; regulators; lipid metabolism; activity



Citation: Dembitsky, V.M. In Silico Prediction of Steroids and Triterpenoids as Potential Regulators of Lipid Metabolism. *Mar. Drugs* **2021**, *19*, 650. <https://doi.org/10.3390/md19110650>

Received: 25 October 2021

Accepted: 19 November 2021

Published: 22 November 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the author. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Lipids are a complex name for many metabolites of natural origin, and they are combined into various groups according to a common physical property, hydrophobicity, that is, insolubility in water [1–4]. However, this definition is currently not entirely correct, since some groups, phospholipids or sphingolipids and some others, manifest themselves as amphiphilic compounds; that is, they can dissolve both in polar substances and in non-polar solvents [5–8].

Lipids play an important role in the body in storing energy and are components of biological membranes, steroid hormones, bile acids, and vitamins. They come from food or de novo synthesis in the liver. Fatty acids, stored primarily as triglycerides, are the main source of energy for the muscles and heart. However, the overproduction and accumulation of triglycerides in adipose tissue and other tissues are closely associated with metabolic disorders in humans. Disorders of lipid metabolism lead to the development of many diseases, including atherosclerosis, which occurs because of a violation of cholesterol homeostasis and is closely associated with atherosclerosis [9,10]. In addition, estrogen and estrogen receptors are well-known regulators of several aspects of metabolism, including glucose and lipid metabolism, and impairment of estrogen signaling is associated with the development of metabolic diseases [11].

The regulation of lipid metabolism is of interest primarily in the context of the regulation of energy flow and the way of its integration with other energy sources in tissues. A special role in the regulation of lipid metabolism is played by hormones such as adrenaline and norepinephrine, glucagon, glucocorticoids, hormones of the anterior pituitary gland, as well as thyroxine and sex hormones [12–20].

This review is devoted to natural, semi-synthetic and synthetic steroids, and triterpenoids isolated from plants, fungi, marine invertebrates, and synthesized in various laboratories around the world. The presented class of molecules differs from many other steroids or triterpenoids that exhibit a wide range of biological activities aimed at lowering cholesterol by inhibiting cholesterol synthesis or other activities associated with these processes.

2. Aromatic Steroids Derived from Natural Sources

The most famous natural monoaromatic steroids that are of some practical interest in terms of regulators of lipid metabolism are estrone (1), estradiol (2), estriol (3), and equilin (4). The chemical structures of these aromatic steroids are shown in Figure 1. It is known that estrone is a minor female sex hormone, which was discovered in the 1920s from the urine of pregnant women independently by two groups of scientists from Germany and the United States, biochemist Adolph Butenandt and American scientists Edward Doisy and Edgar Allen, respectively [21–28]. Later, Adolf Frederik Johann Butenandt from the Institute of Chemistry in Götting (Germany) received the Nobel Prize in Chemistry in 1939 for the discovery of this hormone.

Estrone, or (8R,9S,13S,14S)-3-hydroxy-13-methyl-7,8,9,11,12,14,15,16-octahydro-6H-cyclopenta[a]phenanthren-17-one (1), is produced in vivo from androstenedione and/or testosterone via estradiol. The above-mentioned estrogens show estrogenic activity with various variations [29,30], although changes in the structure of estrogens D ring may also demonstrate anticancer activity [31]. The presence of female sex hormonal (1–4) estrogens was first detected in plants in 1926 by Dohrn and co-workers [32] and then, in the 1930s, simultaneously by Butenandt and Jacobi [26] and Skarzynski [33]. Recently, Janeczko and Skoczowski [34] published a survey in which they summarized the data on the presence of mammalian sex hormones and their physiological role in plants. These hormones, such as 3,17 β -dihydroxy-1,3,5(10)-estratriene, rosterone, testosterone, or progesterone, were present in 60–80% of the plant species investigated. Butenandt and Jacobi [26] isolated estrone (1) from seeds and pollen of *Hyphaene thebaica*, *Glossostemon bruguieri*, *Glycyrrhiza glabra*, *Malus pumila*, *Phoenix dactylifera*, *Ph. vulgaris*, *Punica granatum*, *Salix caprea*, and *Salix* sp. The chemical structures of steroids and triterpenoids are shown in Figure 1, and biological activity is shown in Table 1.

From the 1930s to the present, more than 15,000 articles have been devoted to various issues of estrone and its derivatives, and more than 200 reviews summarize the data on the activity of this hormone. Estrone (1) and estradiol (2) and their derivatives (3–10) are the main natural estrogens found in humans and are involved in estrogen metabolism [35–38]. Endogenous estrogens in humans demonstrate a wide range of biological activities and are involved in the regulation of lipid metabolism, which has been the subject of tens of thousands of articles, reviews, and books. Additionally, in this short paragraph, we will not list and discuss this topic, but in Table 1, we give a short list of the main biological activities that aromatic steroids show.

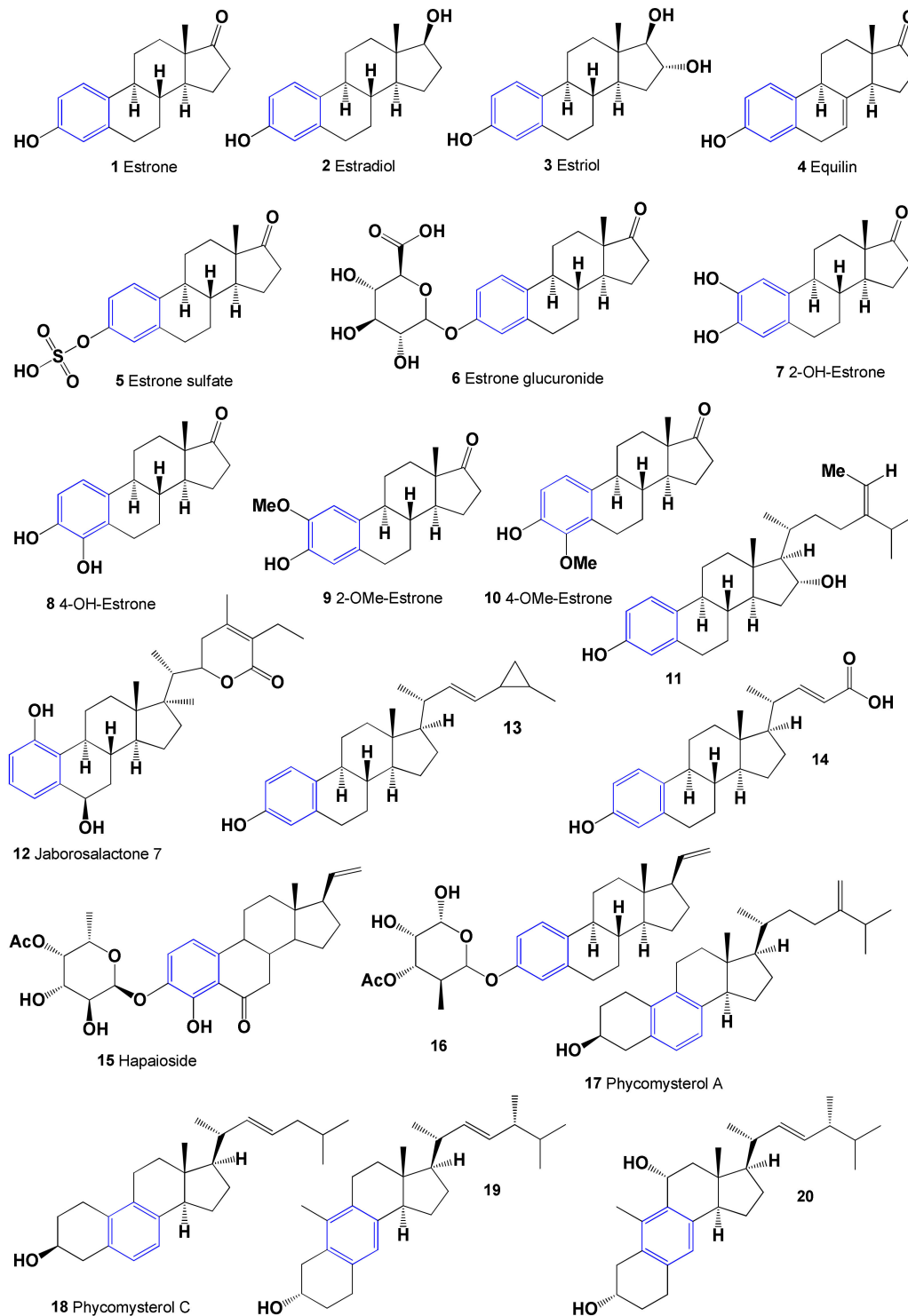


Figure 1. Bioactive aromatic steroids demonstrating with dominance of antihypercholesterolemia and antiovarulation activity.

Aromatic steroids found in animals and humans are also produced by some marine invertebrates and are found in the extracts of certain plants and fungi, as well as in marine sediments. Thus, the extract of bark from the main wooden rod of ketapang *Terminalia catappa* (family Combretaceae) contained estrone (1), estriol (3), equilin (4), equilin sulfate (5), and (11) [39]. The ethanol extract of leaves of *T. catappa* shows antimicrobial activity against *Escherichia coli*, *Staphylococcus aureus*, and *Candida albicans*, as well as antifungal activity against *Epidermophyton floccosum* and *C. albicans* [40–42]. In addition, extracts of leaves and roots and other parts have been studied for many medicinal properties and have shown

antidiabetic, wound healing, anti-inflammatory, anticancer, antimicrobial, hepatoprotective, and antioxidant activities [43]. Unusual withanolide (20S,22R)-1,6 β ,17 α ,27-tetrahydroxy-19-norwitha-1,3,5(10),24-tetraenolide, named jaborosalactone 7-(**12**), was isolated from the leaf extracts of *Juborosa leucotricha* [44].

The steroid 24,26-cyclo-19-norcholesta-1,3,5(10),22-tetraen-3-ol (**13**) is characterized by the presence of an aromatic A ring containing a cyclopropane ring in the side chain was isolated from the Hainan soft coral *Dendronephthya studeri* [45]. The ring A-aromatic bile acid 3-hydroxy-19-norchola-1,3,5(10),22-tetraen-24-oic acid (**14**) was isolated from methanol extract of the marine sponge *Sollasella moretonensis*, collected from the seabed of northern Queensland, Australia [46]. Commenting on the release of this unusual lipid, the authors point out that aromatic steroids are more typical for plants and less so for marine invertebrates or higher animals. A 4-hydroxy-6-oxopregnane-3-glycoside with an aromatic ring A containing the sugar 6'-deoxy-L- β -altropyranose-4'-acetate named hapaoside (**15**) was found in the extract of a Pohnpei, a tube sponge, *Cribrochalina olemda* from the family Niphatidae [47]. The activity of this glycoside has not been determined, although it is known that this marine sponge contains cyclic peptides called kapakahines, some of which show cytotoxicity against P388 murine leukemia cells [48]. Structurally similar to hapaoside, 3-(4-O-acetyl-6-deoxy- β -galactopyranosyloxy)-19-norpregna-1,3,5(10),20-tetraene (**16**) was isolated from bush coral *Alcyonium gracillimum* (syn. *Scleronephthya gracillimum*), which was collected from the Gulf of Sagami, Japan [49].

Monoaromatic B ring steroids are rare natural lipids found predominantly in the mushroom kingdom and are also present in marine sediments or oils. Thus, phycomysterol A (**17**) and C (**18**), which possess a unusual natural 19-norergostane skeleton with an aromatic B ring, are synthesized by a filamentous fungus *Phycomyces blakesleeanus* (order Mucorales, the phylum Zygomycota) [50]. In addition, phycomysterol A showed antiviral inhibition at a concentration of 0.64 μ g per well (200 μ L) and showed (IC₅₀ = 5.0 μ g/mL) against both mouse lymphomas (IC₅₀ = 10 μ g/mL) and against the three human cell lines, A549 (lung carcinoma), HT29 (colon carcinoma), and MEL28 (melanoma) human cell lines.

Pathogenic fungus *Gibberella zeae* (syn. *Fusarium roseum*) is a worldwide parasite that can produce a wide variety of steroids and synthesized unusual (22E,24R)-1(10 \rightarrow 6)-abeoergosta-5,7,9,22-tetraen-3 α -ol (**19**), which was isolated from the cultures of *Gibberella zeae*, an endophytic fungus isolated from the marine green alga *Codium fragile* [51]. This compound showed significant cytotoxicity toward murine colorectal CT26 and human leukemia K562 cancer cell lines. An aromatic B ring called topsentisterol E1 (**20**) was isolated from bioactive fractions of a marine sponge *Topsentia* sp. [52]. The presence of this unusual steroid may indicate that it is synthesized by the endophytic fungus, which is a symbiont in this sponge species.

Comparison of Biological Activities of Natural Aromatic Steroids

It is known that the chemical structure of both natural and synthetic molecules pre-determines biological activity, which makes it possible to analyze the structure–activity relationships (SAR). This idea was first proposed by Brown and Fraser more than 150 years ago, in 1868 [53], although, according to other sources, SAR originates from the field of toxicology, according to which Crois in 1863 determined the relationship between the toxicity of primary aliphatic alcohols and their solubility in water [54]. More than 30 years later, Richet in 1893 [55], Meyer in 1899 [56], and Overton in 1901 [57] separately found a linear correlation between lipophilicity and biological effects. By 1935, Hammett [58,59] presented a method of accounting for the effect of substituents on reaction mechanisms using an equation that considered two parameters, namely the substituent constant and the reaction constant. Complementing Hammett's model, Taft proposed in 1956 an approach for separating the polar, steric, and resonance effects of substituents in aliphatic compounds [60]. Combining all previous developments, Hansch and Fujita laid out the mechanistic basis for the development of the QSAR method [61], and the linear Hansch

equation and Hammett's electronic constants are detailed in the book by Hansch and Leo published in 1995 [62].

Some well-known computer programs can, with some degree of reliability, estimate the pharmacological activity of organic molecules isolated from natural sources or synthesized compounds [63–65]. It is known that classical SAR methods are based on the analysis of (quantitative) structure–activity relationships for one or more biological activities using organic compounds belonging to the same chemical series as the training set [66].

The computer program PASS, which has been continuously updating and improving for the past thirty years [67], is based on the analysis of a heterogeneous training set included information about more than 1.3 million known biologically active compounds with data on ca. 10,000 biological activities [68,69]. Chemical descriptors implemented in PASS, which reflect the peculiarities of ligand–target interactions and original realization of the Bayesian approach for 18 elucidations of structure–activity relationships, provide the average accuracy and predictivity for several thousand biological activities equal to about 96% [70]. In several comparative studies, it was shown that PASS outperforms in predictivity some other recently developed methods for estimation of biological activity profiles [71,72]. Freely available via the Internet, the PASS Online web service [73] is used by more than thirty thousand researchers from almost a hundred countries to determine the most promising biological activities for both natural and synthetic compounds [74–76]. To reveal the hidden pharmacological potential of the natural substances, we have successfully been using PASS for the past fifteen years [77–80].

Table 1. Biological activities of aromatic steroids with over 90% confidence.

No.	Discovered Activity, (Pa) *	Reported Activity	References
1	Ovulation inhibitor (0.942) Cardiovascular analeptic (0.924) Antihypercholesterolemic (0.871) Lipid metabolism regulator (0.730)	Inhibitor aromatase Sulfatase inhibitor Estrogenic Promotor breast cancer	[81–84]
2	Antihypercholesterolemic (0.904) Ovulation inhibitor (0.889) Neuroprotector (0.870) Anesthetic general (0.868) Acute neurologic disorders treatment (0.793) Prostate disorders treatment (0.729) Anti-inflammatory (0.713)	Antioxidant Anti-inflammatory Uterotrophic RNA polymerase Promoter of breast, ovarian and endometrial cancers Neuroprotective properties	[81,85–87]
3	Ovulation inhibitor (0.900) Acute neurologic disorders treatment (0.822) Antihypercholesterolemic (0.791)	Estrogenic Agonist of the ERs RNA polymerase	[88,89]
4	Antihypercholesterolemic (0.856) Ovulation inhibitor (0.847) Cardiovascular analeptic (0.842) Lipid metabolism regulator (0.788)	Estrogenic Estrogen agonist	[90–92]
5	Acute neurologic disorders treatment (0.912) Male reproductive dysfunction treatment (0.847) Ovulation inhibitor (0.786) Postmenopausal disorders treatment (0.643) Antihypercholesterolemic (0.640) Menopausal disorders treatment (0.579)	Inhibitor aromatase Sulfatase inhibitor Inhibitor of human breast cancer Concentration Cardiovascular agent Postmenopausal disorders	[93,94]
6	Antihypercholesterolemic (0.973) Acute neurologic disorders treatment (0.922) Lipid metabolism regulator (0.907) Antithrombotic (0.714) Ovulation inhibitor (0.662) Dementia treatment (0.616) Hypolipemic (0.613) Male reproductive dysfunction treatment (0.587) Menopausal disorders treatment (0.582)	Estrogenic Ovarian activity Urine production	[95]

Table 1. Cont.

No.	Discovered Activity, (Pa) *	Reported Activity	References			
7	Ovulation inhibitor (0.956)	Estrogenic Antiestrogenic effects Anticancer	[96–98]			
	Cardiovascular analeptic (0.927)					
	Antihypercholesterolemic (0.868)					
	Male reproductive dysfunction treatment (0.847)					
	Menopausal disorders treatment (0.842)					
	Acute neurologic disorders treatment (0.745)					
	Lipid metabolism regulator (0.701)					
8	Menstruation disorders treatment (0.639)	Estrogenic Strongest neuroprotective effect UDP-glucuronosyltransferase Anti-breast cancer	[99–101]			
	Postmenopausal disorders treatment (0.605)					
	Muscular dystrophy treatment (0.601)					
	Contraceptive female (0.570)					
	Anti-infertility, female (0.567)					
	Ovulation inhibitor (0.930)					
	Cardiovascular analeptic (0.925)					
9	Antihypercholesterolemic (0.855)	Antioxidant Estrogenic Anti-breast cancer	[102]			
	Male reproductive dysfunction treatment (0.843)					
	Acute neurologic disorders treatment (0.780)					
	Menopausal disorders treatment (0.747)					
	Lipid metabolism regulator (0.601)					
	Muscular dystrophy treatment (0.579)					
	Postmenopausal disorders treatment (0.561)					
10	Menstruation disorders treatment (0.533)	Estrogenic Proliferation of human breast cancer	[103–105]			
	Ovulation inhibitor (0.953)					
	Cardiovascular analeptic (0.928)					
	Antihypercholesterolemic (0.857)					
	Menopausal disorders treatment (0.807)					
	Male reproductive dysfunction treatment (0.805)					
	Acute neurologic disorders treatment (0.660)					
11	Contraceptive (0.655)	Activity not studied				
	Lipid metabolism regulator (0.645)					
	Contraceptive female (0.558)					
	Menstruation disorders treatment (0.542)					
	Anti-infertility, female (0.503)					
	Postmenopausal disorders treatment (0.500)					
	12			Ovulation inhibitor (0.925)	Activity not studied	
Cardiovascular analeptic (0.922)						
Antihypercholesterolemic (0.833)						
Male reproductive dysfunction treatment (0.821)						
Menopausal disorders treatment (0.712)						
Acute neurologic disorders treatment (0.662)						
Contraceptive (0.602)						
13	Lipid metabolism regulator (0.571)	Activity not studied				
	Muscular dystrophy treatment (0.548)					
	Antihypercholesterolemic (0.959)					
	Hypolipemic (0.808)					
	14			Lipid metabolism regulator (0.913)	Activity not studied	
				Antihypercholesterolemic (0.767)		
				Antihypercholesterolemic (0.961)		
Hypolipemic (0.711)						
15		Antihypercholesterolemic (0.953)	Activity not studied			
		Antihypercholesterolemic (0.946)				
		Antihypercholesterolemic (0.907)				
	Antihypercholesterolemic (0.929)					
	Antihypercholesterolemic (0.935)					
	Antihypercholesterolemic (0.950)					
	Antihypercholesterolemic (0.914)	Anticancer			[52]	

* Only activities with Pa > 0.5 are shown.

In the current study, we obtained PASS predictions for about one hundred steroids and triterpenoids produced by different living organisms. PASS estimates are presented as

Pa values, which correspond to the probability of belonging to a class of “actives” for each predicted biological activity. The higher the Pa value is, the higher the confidence that the experiment will confirm the predicted biological activity [78,80].

The study of the biological activity of aromatic steroids using the PASS program showed that all steroids presented in Table 1 can be divided into two groups. The first group includes steroids that show antiovarulation activity with a high degree of reliability, and the second group includes steroids for which antihypercholesterolemic activity is dominant. The first group includes aromatic steroids numbered 7, 9, 1, 8, and 10 with over 92.5 percent confidence, although the second group includes steroids 6, 11, 13, 14, 15, and 19 with over 94.6 percent confidence, according to PASS.

A 3D graph of the predicted and calculated activities of aromatic steroids belonging to the first group is shown in Figure 2. From the first group of steroids that are inhibitors of ovulation, 2-hydroxyestrone (7) can be distinguished, which has a confidence level of more than 95%, and its predicted activities are presented in Figure 3. Additionally, 3D graphs that demonstrate the predicted pharmacological activity were obtained using the Origin Pro 2021 graphical program. The program analyzes the data obtained by the PASS program and builds graphs that are given in this publication.

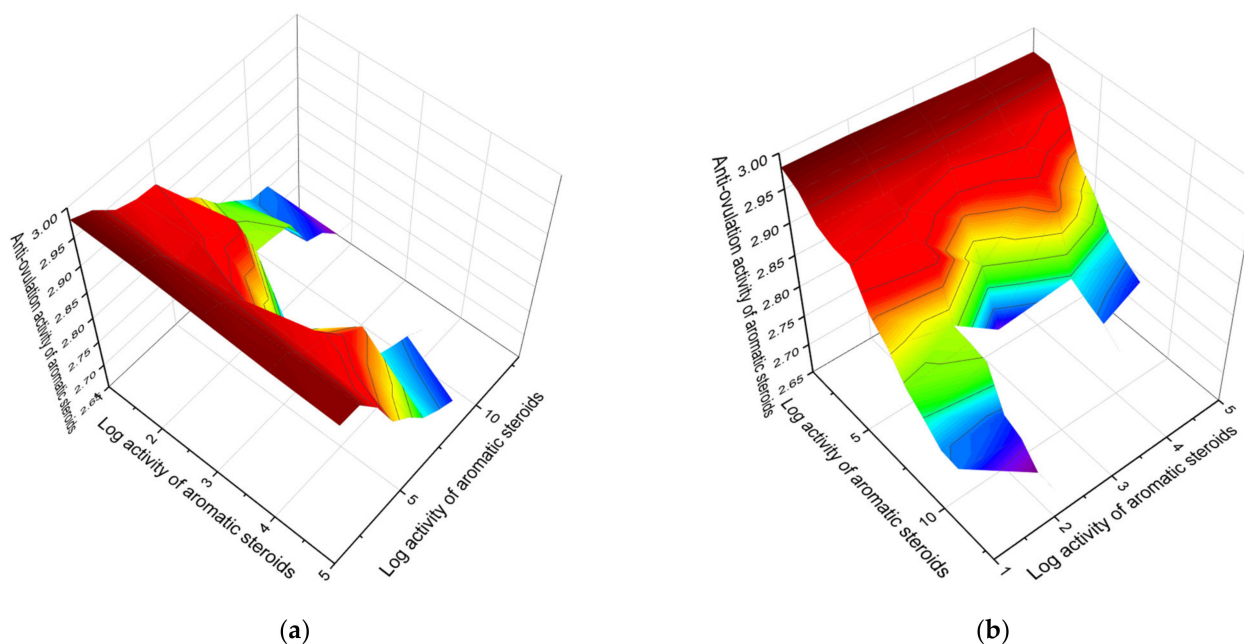


Figure 2. The 3D graph (X (a) and Y (b) views) shows the predicted and calculated antiovarulation activity of aromatic steroids (compound numbers: 7, 9, 1, 8, and 10) showing the highest degree of confidence, more than 92.5%. These steroids are derived from animals, including humans, as well as the extract of bark from the main wooden rod of ketapang *Terminalia catappa*, and can be used in clinical medicine as potential agents with strong ovulation inhibitors. The units of measurement of the x–y digits (Cos–Sin or Sin–Sin) are the relative dimensions that the Origin Pro 2021 graphics program chooses independently, depending on the data obtained by the PASS program. On the 3D graphs, the Origin Pro software in red indicates the maximum biological activity of an individual steroid, and blue indicates the minimum activity.

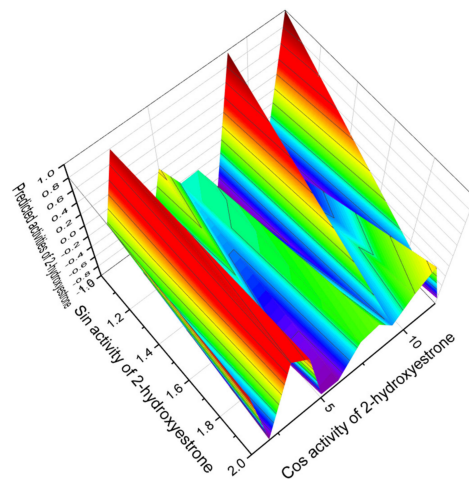


Figure 3. The 3D graph shows the predicted pharmacological activities of 2-hydroxyestrone (7). 2-Hydroxyestrone (2-OHE1) or 2,3-estracatechol, also known as *estra-1,3,5(10)-trien-2,3-diol-17-one*, is an endogenous, naturally occurring catechol estrogen that was first isolated from human urine more than 60 years ago [106], apparently as a product of estrone and estradiol metabolism. According to the PASS data, 2-hydroxyestrone is the strongest ovulation inhibitor among aromatic steroids. In addition, it is a cardiovascular analeptic and can be used as a drug for the treatment and prevention of male reproductive dysfunction, menopausal disorders in women, as well as treatment of muscular dystrophy.

Among the second group of aromatic steroids that demonstrate strong antihypercholesterolemic activity, and a 3D graph of predicted and calculated antihypercholesterolemic activity is shown in Figure 4; the strongest steroid is estrone glucuronide (6), and a 3D graph of its predicted pharmacological activities is shown in Figure 5.

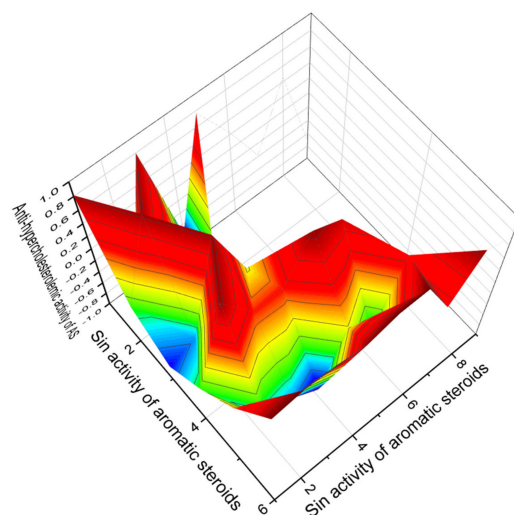


Figure 4. The 3D graph showing the predicted and calculated antihypercholesterolemic activity of aromatic steroids (compound numbers: 6, 13, 11, 14, 15, and 19) with the highest degree of confidence, more than 94.6%. These steroids have been obtained from a variety of natural sources, including plant, fungal, and marine invertebrate extracts, and appear to be useful in clinical medicine as potential agents with potent antihypercholesterolemic activity.

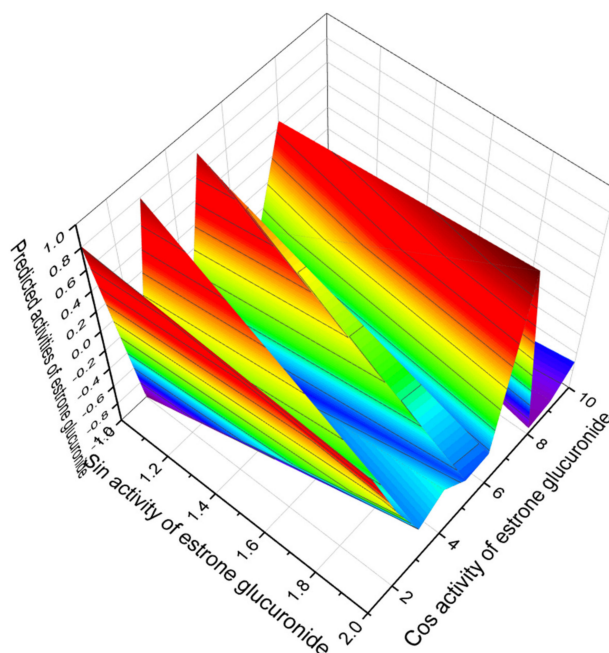


Figure 5. The 3D graph shows the predicted pharmacological activities of estrone glucuronide (6). The water-soluble conjugate of estrone, also known as estrone 3- β -D-glucosiduronic acid, is a naturally occurring steroidal estrogen containing β -D-glucopyranuronic acid that was first discovered and isolated from the urine of women in the late 1930s [107–109]. According to the PASS data, estrone glucuronide, in addition to the main antihypercholesterolemic activity (97.3%), is also a potent regulator of lipid metabolism with a reliability of 90.7% and can be used to treat acute neurological disorders (92.2%).

3. Natural, Semi-, and Synthetic Steroid Phosphate Esters

Steroid phosphate esters are rare lipid molecules that can form the building blocks of biological membranes and have been found more recently in starfish extracts. So, for the first time, steroid phosphates were discovered more than a quarter of a century ago by Italian scientists from the University of Federico II, in the city of Napoli. Unique steroids were isolated from the extract of the polar lipids of the starfish *Tremaster novaecaledoniae*, which was collected at a depth of 530 m of New Caledonia [110]. The isolated phosphated steroid glycosides were called tremasterols A and C, and their structures were identified as 3 β -O-sulphated, 6 α -O-phosphated, and 16 β -O-acetylated groupings on a steroidal skeleton (21, 22, and 23).

Synthetic steroids, such as testosterone 17 β -phosphate (24), cortisol 21-phosphate (25), and cholesterol 3 β -phosphate (26), were chosen by us for comparison of biological activity with the activity of steroids isolated from marine invertebrates [111]. Testosterone 17 β -phosphate (24) is an androgen and belongs to the class of anabolic steroids and is used for intramuscular injection, and it is a substrate for phosphatases in the phosphatase pool of the prostate [112]. Cortisol 21-phosphate (25) refers to the glucocorticoid class of hormones, and it functions to increase blood sugar levels through gluconeogenesis and to promote the metabolism of fats, proteins, and carbohydrates, and it is a substrate for alkaline phosphatase and is used for an enzyme immunoassay for human chorionic gonadotropin, human growth hormone, and α -fetoprotein and estradiol [113]. Cholesterol 3 β -phosphate (26) promotes normalization of blood pressure and plays an important role in atherogenesis [114,115].

Two novel cholesterol-lowering agents called sodium ascorbic campestanol phosphate (27) and sodium ascorbic sitostanol phosphate (28) were synthesized, and their properties were studied [116]. Using Western blot analysis of P-GP expression, it was shown that changes in *mdr-1* gene expression lead to correlating changes in P-GP protein expression. More recently, two steroid phosphate esters (29 and 30, structures see on Figure 6 and

activities on Table 2) have been synthesized, acting as inhibitors of cholesterol biosynthesis. Methods of treating or preventing various diseases, conditions, and disorders by administering these steroids or compositions are also provided [117]. Estradiol phosphate (31) is ester of estrogen with phosphoric acid and acts as a prodrug of estradiol in the human body. In medical practice, both drugs can be used to treat prostate cancer [118].

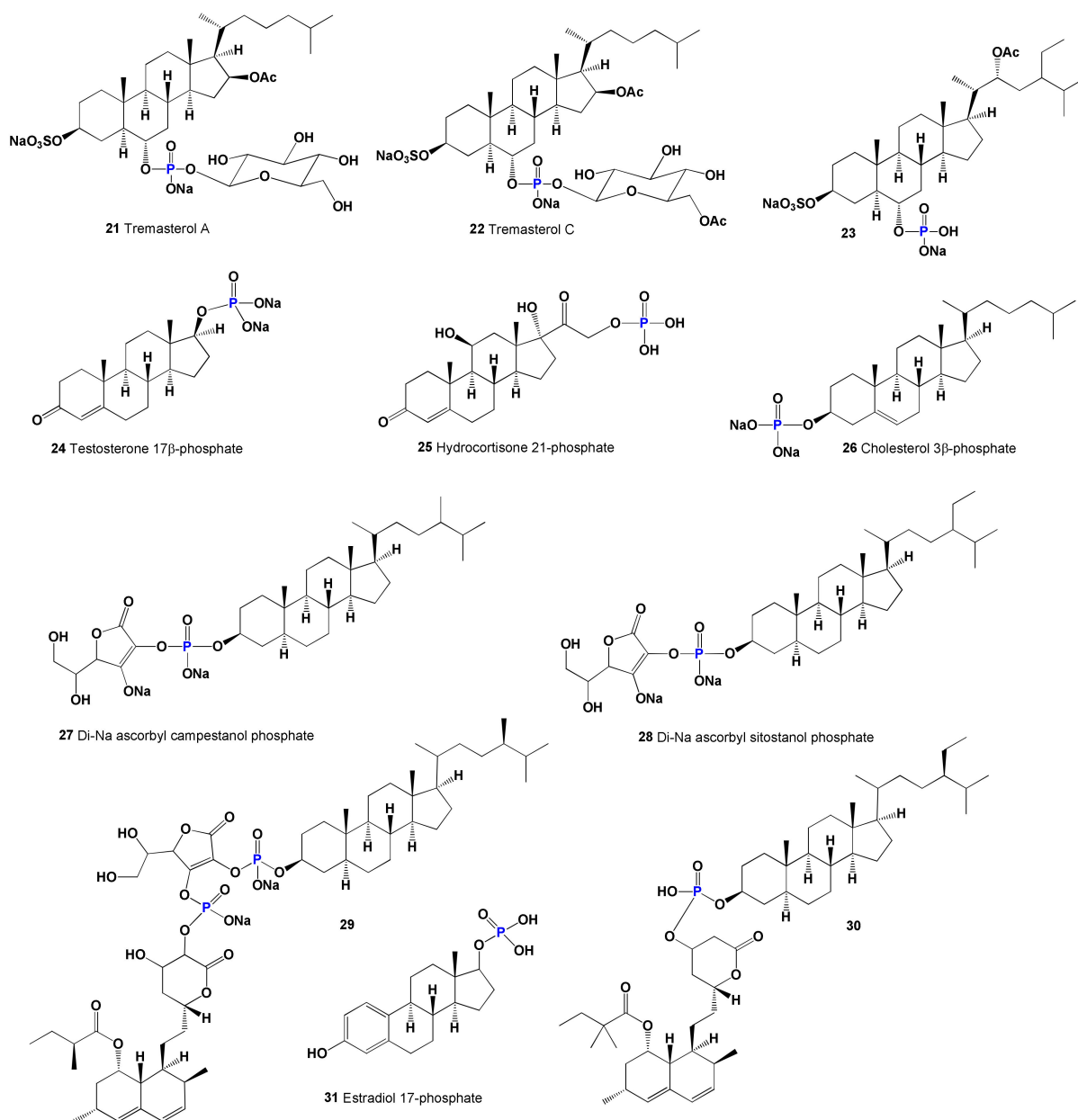


Figure 6. Bioactive natural, semi-synthetic, and synthetic steroid phosphate esters.

Table 2. Predicted biological activities of steroid phosphate esters.

No.	Lipid Metabolism Regulators, (Pa) *	Reported Activity	Ref.
21	Wound healing agent (0.975) Hepatoprotectant (0.961) Analeptic (0.952) Antihypercholesterolemic (0.926) Cholesterol synthesis inhibitor (0.799)	Activity not studied	
22	Wound healing agent (0.947) Analeptic (0.941) Hepatoprotectant (0.932) Anticarcinogenic (0.915) Antihypercholesterolemic (0.912) Cholesterol synthesis inhibitor (0.778)	Activity not studied	
23	Antihypercholesterolemic (0.900) Hepatoprotectant (0.853) Wound healing agent (0.844) Antineoplastic (0.816) Antiinflammatory (0.782) Cholesterol synthesis inhibitor (0.778) Atherosclerosis treatment (0.675)	Activity not studied	
24	Neuroprotector (0.987) Anesthetic general (0.959) Respiratory analeptic (0.944) Antihypercholesterolemic (0.909)	Substrate for phosphatases	[112]
25	Anesthetic general (0.991) Respiratory analeptic (0.990) Neuroprotector (0.976) Antiinflammatory (0.906) Antihypercholesterolemic (0.900)	Increases blood sugar levels	[113]
26	Respiratory analeptic (0.979) Anesthetic general (0.973) Neuroprotector (0.972) Antihypercholesterolemic (0.971) Wound healing agent (0.913) Antineoplastic (0.826) Cholesterol synthesis inhibitor (0.801)	Stabilizes blood pressure	[114]
27	Respiratory analeptic (0.995) Anesthetic general (0.948) Antihypercholesterolemic (0.945) Neuroprotector (0.932) Hemostatic (0.910) Wound healing agent (0.897) Cholesterol synthesis inhibitor (0.867) Acute neurologic disorders treatment (0.827)	Reduces cholesterol levels	[116]
28	Antihypercholesterolemic (0.967) Wound healing agent (0.921) Neuroprotector (0.909) Cholesterol synthesis inhibitor (0.872)	Reduces cholesterol levels	[116]
29	Antihypercholesterolemic (0.996) Cholesterol absorption inhibitor (0.976) Cholesterol synthesis inhibitor (0.952) Lipid metabolism regulator (0.952) Lipoprotein disorders treatment (0.893)	Cholesterol biosynthesis inhibitor	[117]
30	Antihypercholesterolemic (0.999) Antihyperlipoproteinemic (0.986) Hypolipemic (0.974) Cholesterol absorption inhibitor (0.957) Lipid metabolism regulator (0.954) Cholesterol synthesis inhibitor (0.916) Lipoprotein disorders treatment (0.782) Acute neurologic disorders treatment (0.751) Atherosclerosis treatment (0.729)	Cholesterol biosynthesis inhibitor	[117]
31	Neuroprotector (0.982) Anesthetic general (0.931) Antihypercholesterolemic (0.909) Acute neurologic disorders treatment (0.831) Prostate disorders treatment (0.640)	Remedy for the treatment of prostate cancer	[118]

* Only activities with Pa > 0.5 are shown.

Comparison of Biological Activities of Steroid Phosphate Esters

According to PASS, antihypercholesterolemic activity is characteristic of all steroid phosphate esters presented in Table 2 with varying degrees of confidence. In addition, all steroids are inhibitors of cholesterol synthesis, except for steroid 31. However, some lipid steroidal molecules such as 21–23 can be agents for wound healing and hepato-protectors

with a high degree of certainty, and steroids 23–28 and 31 also demonstrate the properties of a neuroprotective agent with a high degree of certainty from 90 to 98%.

Steroid phosphate esters are of great practical interest in medicine, as they are wound healing agents. The confidence level of steroid 21 is 97.5%. Figure 7 shows the 3D graph of biological activities with a dominant property as a wound healing agent.

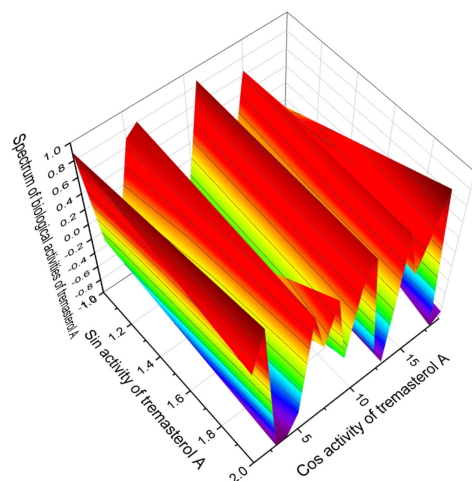


Figure 7. The 3D graph showing the predicted pharmacological activities of tremasterol A (21). Data from the PASS program show that phosphated steroid glycoside named tremasterol A exhibits 20 different biological activities, with dominant properties as a wound healing agent. Interestingly, tremasterol A is isolated from the starfish *Trenzaster novaecaledoniae*. This is a rare occasion when glycosides from starfish demonstrate such beneficial biological activities.

The sitostanol derivative (30) was synthesized by scientists from British Columbia (Canada) as a strong inhibitor of cholesterol synthesis, and as shown by the PASS data, this lipid molecule is in fact an excellent drug for the treatment of atherosclerosis. The wide range of biological activities of the sitostanol derivative (30) is shown in Figure 8.

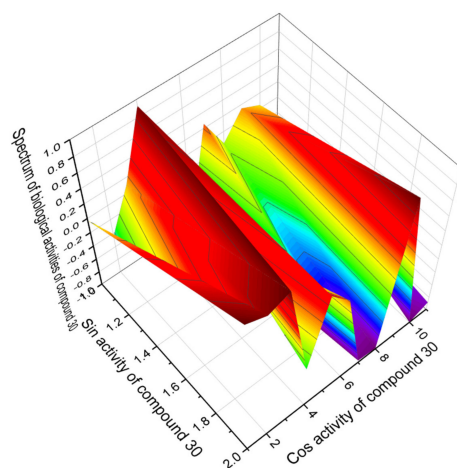


Figure 8. The 3D graph showing the predicted pharmacological activities of the sitostanol derivative (30). This drug has antihypercholesterolemic and antihyperlipoproteinemic properties that help lower cholesterol levels in the human body and reduce the risk of strokes and heart attacks. In addition, the drug is a strong cholesterol absorption inhibitor with a confidence level of 95.7% and a regulator of lipid metabolism with a confidence level of 95.4% and can be used to treat acute neurological disorders.

4. Highly Oxygenated Natural Steroids and Triterpenoids

Highly oxygenated steroids are a large group of steroids and triterpenoids found in plant, fungal, and invertebrate extracts, and many of them exhibit a wide range of biological activities. This group of lipids includes secosteroids, epoxy steroids and peroxy steroids, and triterpenoids, which demonstrate a high degree of activity as potential regulators of lipid metabolism [119–123].

4.1. Secosteroids Derived from Marine and Terrestrial Sources

Secosteroids are a large group of natural steroidal hormones with a so-called ‘broken’ ring by oxidation of rings A, B, C, or D. Typical representatives of secosteroids are fat-soluble vitamins of group D [124–127]. Secosteroids are found in plant and animal extracts, produced by fungi, and found in marine invertebrates and algae [121,128–130].

Two secosterols, 3 β -hydroxy-8 α ,9 α -oxido-8,9-secoergosta-7,9(11),22-triene (**32**) and 3 β -hydroxy-8 α ,9 α -oxido-8,9-secoergosta-7,22-dien-12-one (**33**), named tylopiol A and B, respectively, were isolated from the fresh fruit bodies of fungus *Tylopius plumbeoviolaceus* [131]. The other two steroids, named gloeophyllin J (**34**) and I (**35**), have been isolated from the solid cultures of North American wood-rotting fungi *Gloeophyllum abietinum*. Both compounds showed cytotoxic activity against human cancer cell lines K562 and HCT116 [132]. The chemical structures of steroids are shown in Figure 9 and the biological activity is shown in Table 3.

7-Oxasteroid (**36**) was isolated and characterized from the culture of *Aspergillus ochraceus* EN-31. This endophytic fungus is found and isolated from the marine brown alga *Sargassum kjellmanianum* (Dalian coastline, China) [133]. Compound **36** has been previously reported from a *Penicillium* sp. [134], and it displayed cytotoxic activity against NCI-H460, SMMC-7721, and SW1990 cell lines with IC₅₀ values of 12.1, 16.9, and 67.6 μ M, respectively [133].

Secosteroid (**37**) has been determined in extracts of Australian soft coral *Sinularia* sp. [135,136], and other octocoral *Sinularia leptoclados* are sources of bioactive 9,11-secosteroids and steroid 3,11-dihydroxy-9,11-secogorgost-5-en-9-one (**38**), which showed the highest peroxisome proliferator-activated receptor (PPAR) activity with an IC₅₀ value of 8.3 μ M for inhibiting human breast adenocarcinoma cell (MCF-7) growth. In addition, this steroid modulated the expression of various PPAR-regulated downstream biomarkers, including cyclin D1, cyclin-dependent kinase, B-cell lymphoma 2 (Bcl-2), p38, and extracellular-signal-regulated kinase [137,138].

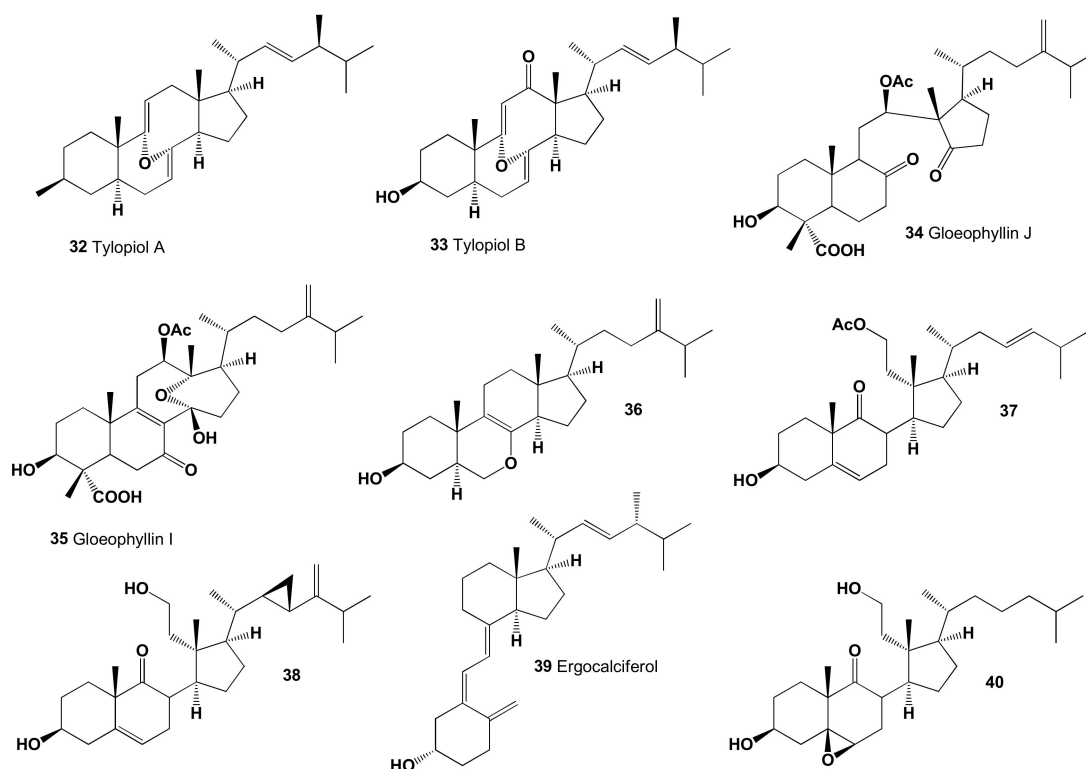


Figure 9. Secosteroids showing activity as lipid metabolism regulators.

Vitamin D₂, known as ergocalciferol or 9,10-seco-(5Z,7E)-5,7,10(19),22-ergostatetraene-3 β -ol (**39**), is the main one that is used in human nutrition [139]. All forms of vitamin D were found in mushrooms (brown Italian cremini, chanterelle, enoki, maitake, morel, shiitake, oyster, portobello, and white button mushrooms) and yeast [140–145]. Secosteroid, 3,11-dihydroxy-5,6-epoxy-9,11-secocholestan-9-one (**40**), was found and identified from extracts of the Taiwanese soft coral *Cespitularia taeniata* [146].

Table 3. Predicted biological activities of secosteroids.

No.	Discovered Activity, (Pa) *	Reported Activity	Ref.
32	Antihypercholesterolemic (0.912)	Activity not studied	
	Hypolipemic (0.802)		
	Atherosclerosis treatment (0.643)		
	Antiparkinsonian, rigidity relieving (0.562)		
33	Antihypercholesterolemic (0.905)	Activity not studied	
	Hypolipemic (0.753)		
	Atherosclerosis treatment (0.559)		
34	Antihypercholesterolemic (0.908)	Anticancer	[131]
	Antineoplastic (0.785)		
	Hypolipemic (0.764)		
	Apoptosis agonist (0.747)		
	Cholesterol synthesis inhibitor (0.744)		

Table 3. Cont.

No.	Discovered Activity, (Pa) *	Reported Activity	Ref.
35	Antihypercholesterolemic (0.916) Antineoplastic (0.833) Hypolipemic (0.827) Apoptosis agonist (0.771) Cholesterol synthesis inhibitor (0.685) Atherosclerosis treatment (0.633)	Anticancer	[131]
36	Antihypercholesterolemic (0.904) Hypolipemic (0.767) Antineoplastic (0.743) Apoptosis agonist (0.676) Proliferative diseases treatment (0.625) Atherosclerosis treatment (0.539)	Anticancer	[133]
37	Antihypercholesterolemic (0.915) Lipid metabolism regulator (0.768)	Activity not studied	
38	Antihypercholesterolemic (0.909) Hypolipemic (0.786)	Anticancer	[147]
39	Antiparkinsonian, rigidity relieving (0.960) Hyperparathyroidism treatment (0.892) Antihypercholesterolemic (0.845) Hypolipemic (0.790) Atherosclerosis treatment (0.628)	Calcium and phosphates metabolism regulator	[148]
40	Chemopreventive (0.989) Hepatoprotectant (0.986) Respiratory analeptic (0.978) Antihypercholesterolemic (0.977) Proliferative diseases treatment (0.969) Antimycobacterial (0.939) Neuroprotector (0.895) Antineoplastic (0.874) Antiprotozoal (Leishmania) (0.772) Atherosclerosis treatment (0.601) Neurodegenerative diseases treatment (0.590) Alzheimer's disease treatment (0.570)	Activity not studied previously	

* Only activities with Pa > 0.5 are shown.

Comparison of Biological Activities of Secosteroids

Among the secosteroids shown in Figure 9, which show activity as regulators of lipid metabolism, the most interesting is lipid molecule number 40, which has an antihypercholesterolemic activity with a confidence level of 97.7 percent. Figure 10 shows the 3D graph of the predicted pharmacological activities of this steroid. Ergocalciferol (39) or vitamin D2 is also of great interest, as PASS showed strong antiparkinson activity with a confidence level of 96.0%. The full spectrum of the predicted pharmacological activities of ergocalciferol is shown in the 3D graph in Figure 11.

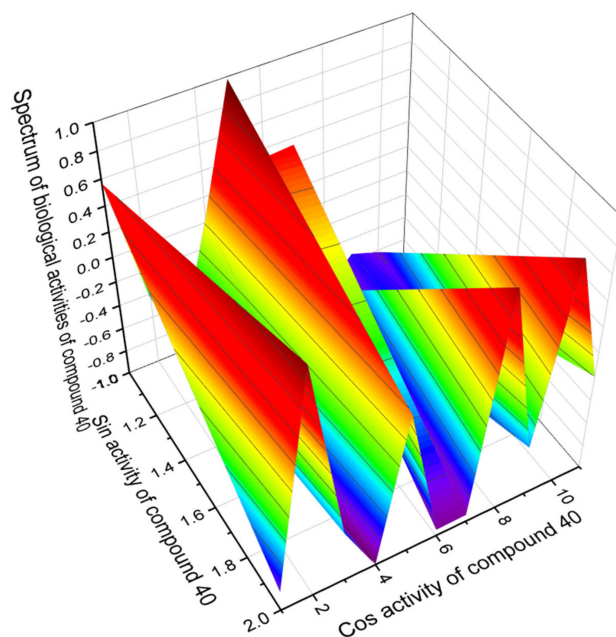


Figure 10. The 3D graph shows the predicted pharmacological activities of compound (40). This secosteroid is characterized by antihypercholesterolemic properties. In addition, it exhibits hepato-protective properties, and it is an inhibitor of cell proliferation, which de facto can prevent several pathological diseases such as atherosclerosis, rheumatoid arthritis, psoriasis, idiopathic pulmonary fibrosis, scleroderma, and liver cirrhosis.

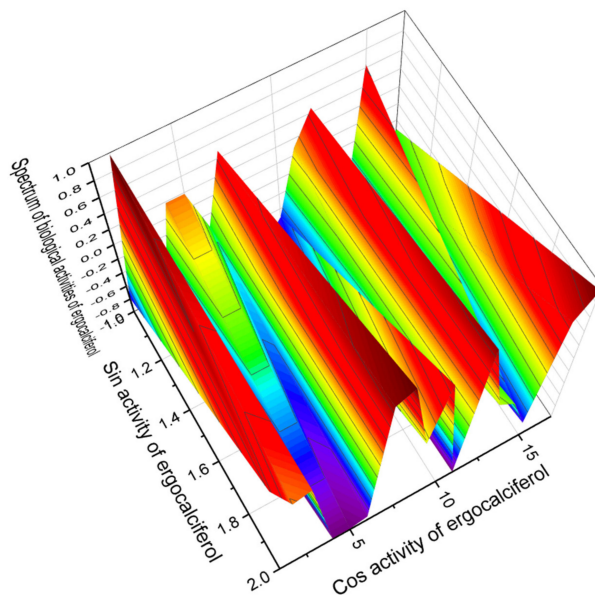


Figure 11. The 3D graph showing the predicted pharmacological activities of ergocalciferol (39). A secosteroid hormone called ergocalciferol was first isolated as a radiation product of 7-dehydrocholesterol and described in 1936 by Windaus and co-workers [149]. Ergocalciferol is an important drug and is recommended by the World Health Organization. In addition, it has anticancer, anti-inflammatory, and antieczema properties, and can also be recommended as an antihypercholesterolemic agent and an agent against Parkinson's disease. Certain foods, such as breakfast cereals and margarine, contain ergocalciferol in some countries, and it is found in the lichen *Cladonia arbuscula* and alfalfa (*Medicago sativa*) [150–154].

4.2. Natural Epoxy Steroids Derived from Marine Sources

α,α -epoxy- and/or β,β -steroids are found in lipid extracts of marine invertebrates, including sponges, soft corals, starfish, and nudibranchs (Mollusca) [123,155–159]. Two cytotoxic epoxysteroids, $5\alpha,6\alpha$ -epoxystigmasta-7,22-en- 3β -ol (41) and $5\alpha,6\alpha$ -epoxystigmasta-7-en- 3β -ol (42) were isolated from the ethanolic extract of the marine sponge *Ircinia aruensis* [160]. The chemical structures of steroids are shown in Figure 12 and the biological activity is shown in Table 4. Topsentisterol B4 (43), epoxy steroid with the β - and α -hydroxyl groups at position 3 and 7, respectively, were present in the extract of the far eastern sponge of *Topsentia* sp. [161]. (24E)- $5\alpha,6\alpha$ -epoxystigmasta-7,24(28)-dien- 3β -ol (44) was isolated from the South China Sea sponge *Phyllospongia foliascens* without studying the biological activity [162]. Triterpene glycoside, eryloside U (45) with be the 7,8-epoxide group was isolated from the sponge *Erylus goffrilleri* collected near Arresife-Seko Reef (Cuba) [163].

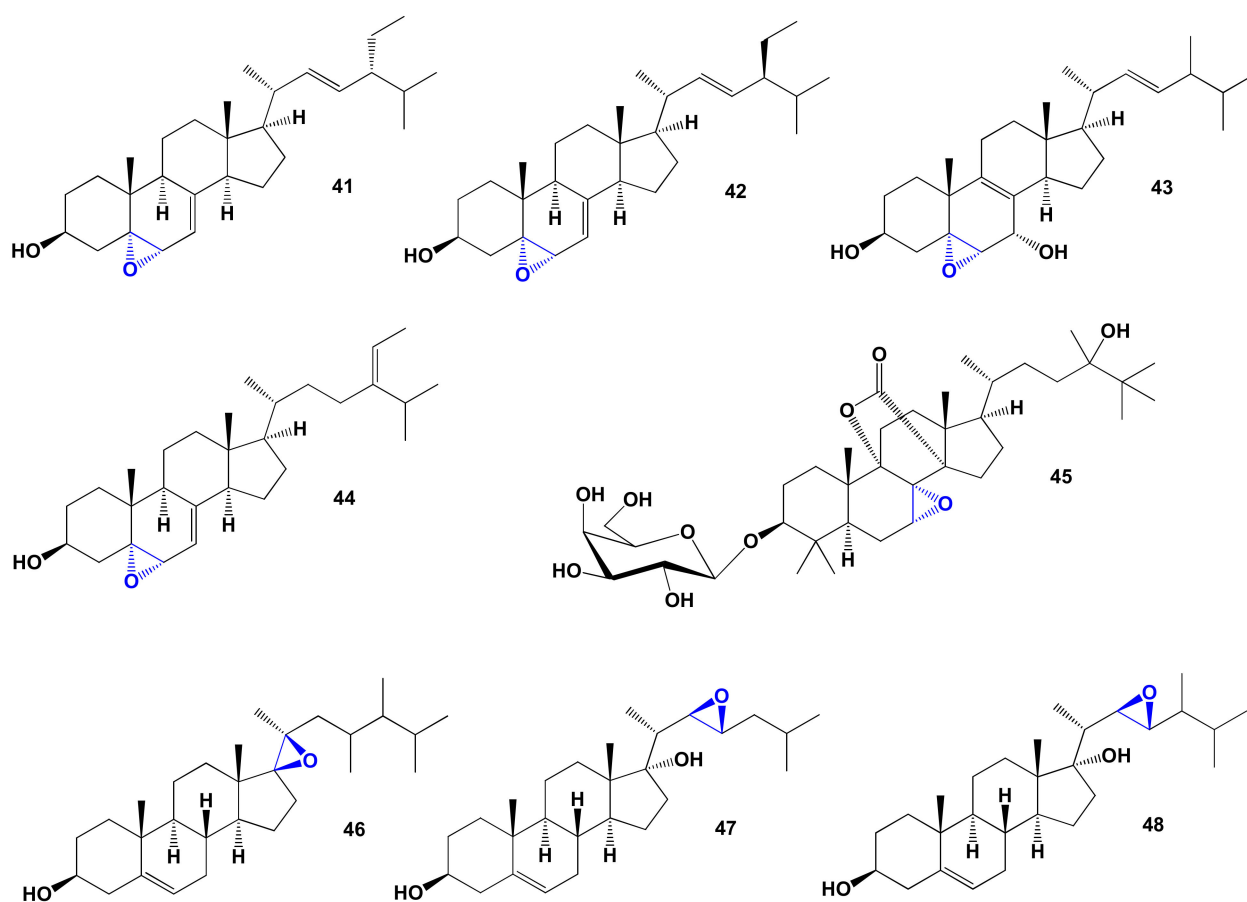


Figure 12. Bioactive α,β -epoxy steroids derived from marine sources.

Unusual $17\beta,20\beta$ -epoxy-23,24-dimethylcholest-5-ene- $3\beta,22$ -diol (46) was found in the Indian Ocean soft coral *Sarcophyton crassocaule* [164].

Table 4. Predicted biological activities of α,β -epoxy steroids.

No.	Discovered Activity, (Pa) *	Reported Activity	Ref.
41	Apoptosis agonist (0.950) Antihypercholesterolemic (0.931) Antineoplastic (0.886) Antieczematic (0.842) Atherosclerosis treatment (0.712)	Cytotoxic	[160]
42	Apoptosis agonist (0.950) Antihypercholesterolemic (0.931) Antineoplastic (0.886) Antieczematic (0.842) Atherosclerosis treatment (0.712)	Cytotoxic	[160]
43	Apoptosis agonist (0.954) Antineoplastic (0.914) Antihypercholesterolemic (0.906) Atherosclerosis treatment (0.741)	Activity not studied	
44	Antihypercholesterolemic (0.934) Apoptosis agonist (0.929) Hypolipemic (0.864) Antineoplastic (0.861)	Activity not studied	
45	Hepatoprotectant (0.994) Respiratory analeptic (0.990) Antihypercholesterolemic (0.897)	Activity not studied	
46	Antihypercholesterolemic (0.900) Neuroprotector (0.749) Cholesterol synthesis inhibitor (0.636)	Activity not studied	
47	Antihypercholesterolemic (0.901) Lipid metabolism regulator (0.833) Prostate disorders treatment (0.714)	Anticancer	[165]
48	Antihypercholesterolemic (0.924) Lipid metabolism regulator (0.820) Neuroprotector (0.728)	Anticancer	[165]

* Only activities with Pa > 0.5 are shown.

Two diol 22,23-epoxy steroids have been isolated from the marine sponge *Axinella* cf. *bidderi*, 17 α -hydroxy-22 β ,23 β -epoxycholest-5-en-3 β -ol (**47**) and 17 α -hydroxy-22 β ,23 β -epoxy-24-methylcholest-5-en-3 β -ol (**48**). Isolated steroids showed activity against prostate, ovary, pancreas, colon, and lung cell lines in vitro [165].

Comparison of Biological Activities of α,β -Epoxy Steroids Derived from Marine Sources

Comparing the data from the PASS obtained for the α,β -epoxy steroids presented in Table 4, it can be concluded that there are no outstanding steroids with lipid metabolism regulator properties; then, we present in Figure 13 a comparative 3D graph for this subgroup of steroids.

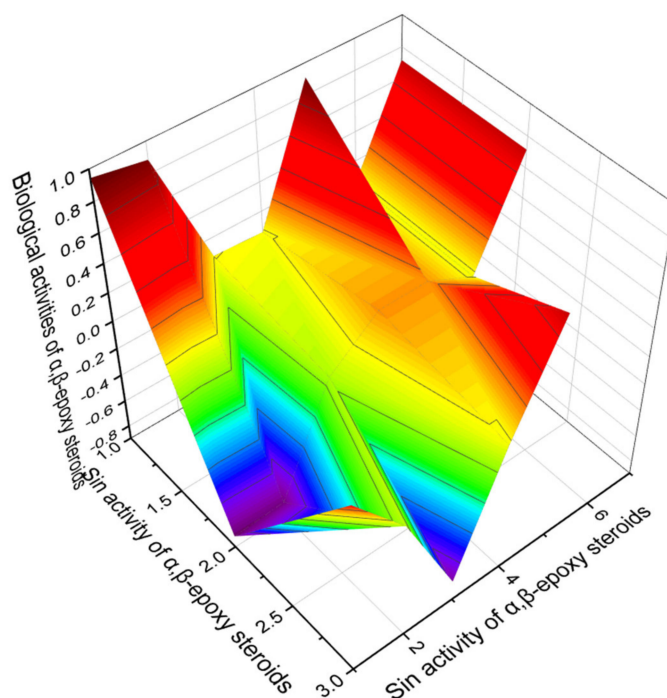


Figure 13. The 3D graph shows the predicted and calculated biological activity of α,β -epoxy steroids (compound numbers: 42, 44, and 48) showing the highest degree of confidence, more than 92.4%.

For a comparative graphic characterization of α,β -epoxy steroids, we selected steroids numbered 42, 44, and 48. All these lipids are characterized by the property of regulators of lipid metabolism with dominant antihypercholesterolemic activity. According to PASS data, steroids 42 and 44 also show a high level of anticancer activity, while steroid 48 is additionally characterized by neuroprotective properties.

4.3. Peroxy-Type Steroids Derived from Natural Sources

Natural and/or synthetic compounds containing a peroxy group (R-O-O-R) are called peroxides [166–172]. Natural peroxides represent a rather large group of compounds that many microorganisms produce, and they have also been found in plants, mushrooms, animals, and marine invertebrates [166–168]. Peroxy steroids are a small group of natural lipids, mainly found in leaves, roots, and bark of plants, and are produced by fungal endophytes and are found in mushrooms [166,167,173].

4.3.1. Steroid Endoperoxides

Astropecten polyacanthus starfish extract has significant cytotoxic effects and contains an unusual peroxy steroid called astropectenol B (49) was isolated from a methanol extract of this starfish [174].

Cytotoxic steroid, (3,5,8,24R,25R)-epidioxy-24,26-cyclocholesta-6,9(11)-dien-3-ol (50) was identified from marine sponge *Tethya* sp. [175]. (3,5,8,24R)-Epidioxy-24-methylcholesta-6-en-3-ol (51) was detected in MeOH extract of the marine sponge *Luffariella* cf. *variabilis* [176], and 22,23-dihydro-5,8-epidioxystigmast-6-en-3-ol (52) was found in the marine sponges *Luffariella* cf. *variabilis* and *Tethya* sp. and sea squirt *Dendrodia grossularia* [175–178].

Fuscoporianol D (53) was found in field-grown mycelia of fungus *Inonotus obliquus* (family Hymenochaetaceae) [179]. Ergosterol peroxide 3-O- β -D-glucopyranoside (54) produced by fungus *Tremella fuciformis* [180] and same compound was detected in the fruiting bodies of the Chinese toxic woodland mushroom *Naematoloma fasciculare* [181]. Endoperoxy steroid 55 was found in popular mushroom in Japan, *Buna shimeji* and in oyster fungus *Pleurotus ostreatus* [182]. Two endoperoxy glycosides (55 and 56) were found in ethanol extracts of the fungus *Lactarius volemus*, which demonstrated anticancer

activity [183,184]. The chemical structures of steroids are shown in Figure 14, and the biological activity is shown in Table 5.

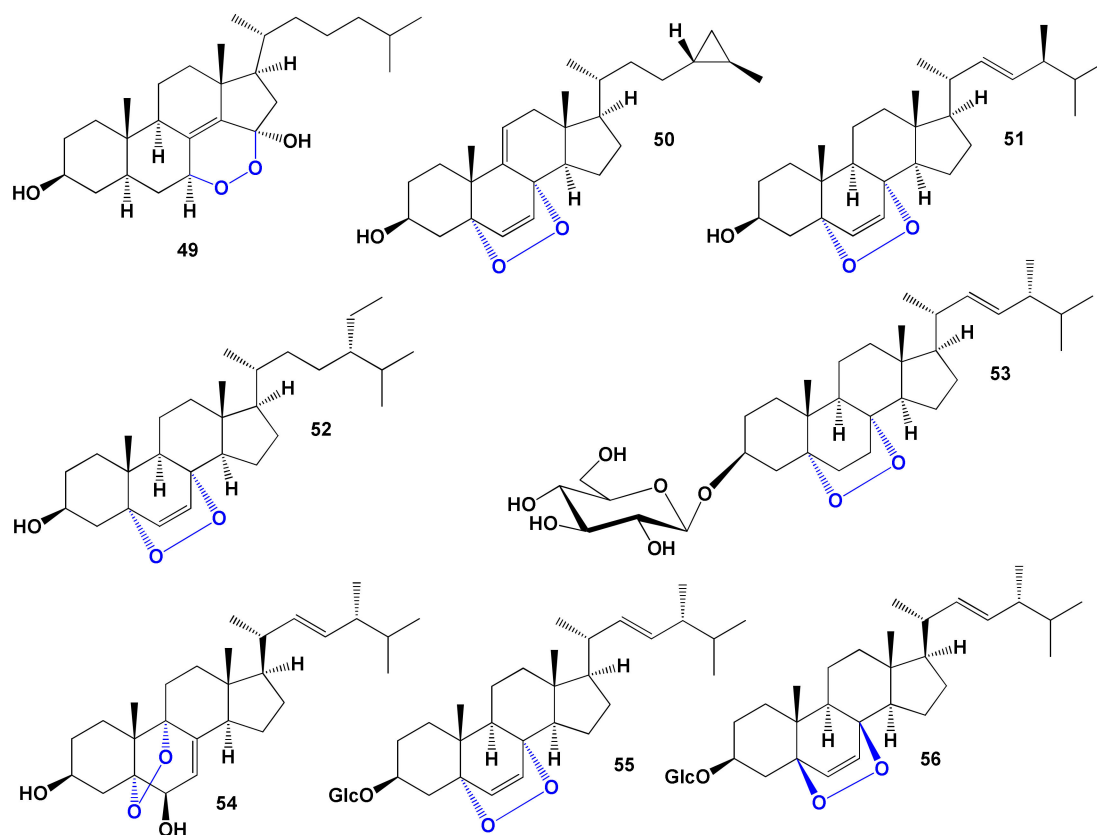


Figure 14. Bioactive steroid endoperoxides derived from marine sources and fungi.

Table 5. Biological activities of endoperoxy steroids.

No.	Lipid Metabolism Regulators, (Pa) *	Reported Activity	Ref.
49	Antihypercholesterolemic (0.914) Hypolipemic (0.635)	Activity not studied	
50	Atherosclerosis treatment (0.911) Hypolipemic (0.836) Lipoprotein disorders treatment (0.826) Antihypercholesterolemic (0.802)	Activity not studied	
51	Atherosclerosis treatment (0.907) Antihypercholesterolemic (0.788)	Activity not studied	
52	Atherosclerosis treatment (0.919) Hypolipemic (0.822) Lipoprotein disorders treatment (0.814)	Activity not studied	
53	Antihypercholesterolemic (0.926) Hypolipemic (0.800) Atherosclerosis treatment (0.709)	Activity not studied	
54	Antihypercholesterolemic (0.900) Hypolipemic (0.827) Atherosclerosis treatment (0.659) Hyperparathyroidism treatment (0.502)	Activity not studied	
55	Antihypercholesterolemic (0.917) Hypolipemic (0.786)	Anticancer	[183]
56	Antihypercholesterolemic (0.917) Atherosclerosis treatment (0.858) Hypolipemic (0.786)	Anticancer	[183]

* Only activities with Pa > 0.5 are shown.

4.3.2. Steroid and Triterpenoid Hydroperoxides

A flowering herbaceous perennial plant from the family Araceae, *Arum italicum*, also known as Italian arum and Italian lords-and-ladies, contains a suite of hydroperoxysterols, including two (57 and 58) that are interesting for lipid metabolism [185]. Two steroids (59 and 60), which showed a cytotoxic effect against several human cancer cell lines, have been isolated from the bark of the chinaberry tree, *Melia azedarach* [186]. The chemical structures of steroids are shown in Figure 15, and the biological activity is shown in Table 6.

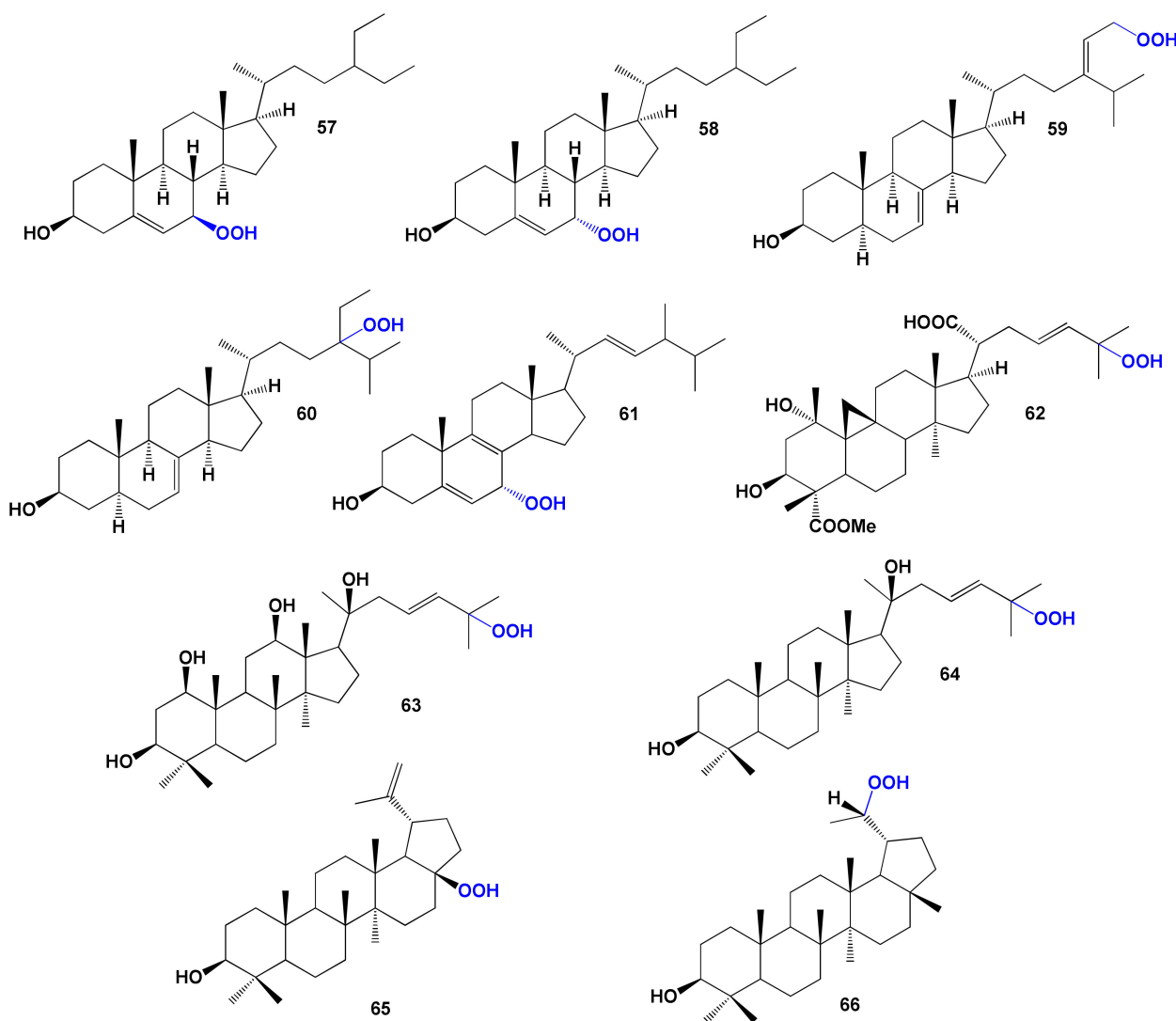


Figure 15. Bioactive steroid hydroperoxides derived from plants.

Ponce and co-workers obtained ergosterol 7-hydroperoxide (61) by the photo-oxidation of ergosterol with singlet oxygen *in vivo* and *in vitro* [187]. In addition, the yeast *Saccharomyces cerevisiae* with the singlet oxygen leads to rapid oxidation of ergosterol to ergosterol 7-hydroperoxide (61) [188].

A tree called Sakae Naa (*Combretum quadrangulare*) in Vietnam, Cambodia, Laos, Myanmar, and Thailand contained a cycloartane-type triterpene quadrangularic acid F (62), and the aqueous and EtOH extracts show antibacterial, anti-HIV, hepatoprotective, and cytotoxic activities [189–191].

The plant *Proboscidea louisiana* produced dammarane triterpenes known as probosciderol I (63) [192], and the stem bark of *Rhus javanica* contained isofouquierone peroxide (64) [193].

The leaves of *Melaleuca ericifolia* contained antiproliferative norlupane triterpene (65) [194], and the aerial roots of *Ficus microcarpa* afforded similar norlupane triterpene (66) [195].

Table 6. Predicted biological activities of hydroperoxy steroids.

No.	Discovered Activity, (Pa) *	Reported Activity	Ref.
57	Antihypercholesterolemic (0.905) Cholesterol synthesis inhibitor (0.799)	Activity not studied	
58	Antihypercholesterolemic (0.905) Hypolipemic (0.802) Cholesterol synthesis inhibitor (0.621) Atherosclerosis treatment (0.542)	Activity not studied	
59	Antihypercholesterolemic (0.933) Hypolipemic (0.877) Cholesterol synthesis inhibitor (0.644) Atherosclerosis treatment (0.675) Prostate disorders treatment (0.645)	Cytotoxic	[186]
60	Antihypercholesterolemic (0.933) Hypolipemic (0.877) Antineoplastic (0.835) Cholesterol synthesis inhibitor (0.650) Atherosclerosis treatment (0.554)	Cytotoxic	[186]
61	Antihypercholesterolemic (0.922) Hypolipemic (0.868) Atherosclerosis treatment (0.678) Antiparkinsonian, rigidity relieving (0.516)	Activity not studied	
62	Antihypercholesterolemic (0.913) Antineoplastic (0.802) Hypolipemic (0.795)	Hepatoprotective Cytotoxic	[191]
63	Antihypercholesterolemic (0.933) Hypolipemic (0.877) Cholesterol synthesis inhibitor (0.644)	Activity not studied	
64	Antihypercholesterolemic (0.923) Hypolipemic (0.774) Cholesterol synthesis inhibitor (0.604) Biliary tract disorders treatment (0.577)	Activity not studied	
65	Antihypercholesterolemic (0.918) Hypolipemic (0.779) Biliary tract disorders treatment (0.655)	Antiproliferative	[194]
66	Antihypercholesterolemic (0.930) Hypolipemic (0.767) Biliary tract disorders treatment (0.717) Atherosclerosis treatment (0.590)	Activity not studied	

* Only activities with Pa > 0.5 are shown.

4.3.3. Comparison of Biological Activities of Peroxy Steroids Derived from Natural Sources

According to published data, most natural peroxides isolated from both plants and marine invertebrates show predominantly antiprotozoal activity. Such compounds include diterpenoids, triterpenoids, and steroids [166–168,171,173,196–199].

Analysis of PASS data on peroxy steroids and triterpenoids such as endoperoxides and hydroperoxides showed that most of these lipids have a high confidence level of more than 90 percent, but nevertheless, only three of all peroxy steroids deserve attention; these

are steroids numbered 59, 63, and 66, which have a confidence level of over 93 percent. Figure 16 presents the 3D graph showing the predicted and calculated biological activity of steroid hydroperoxides.

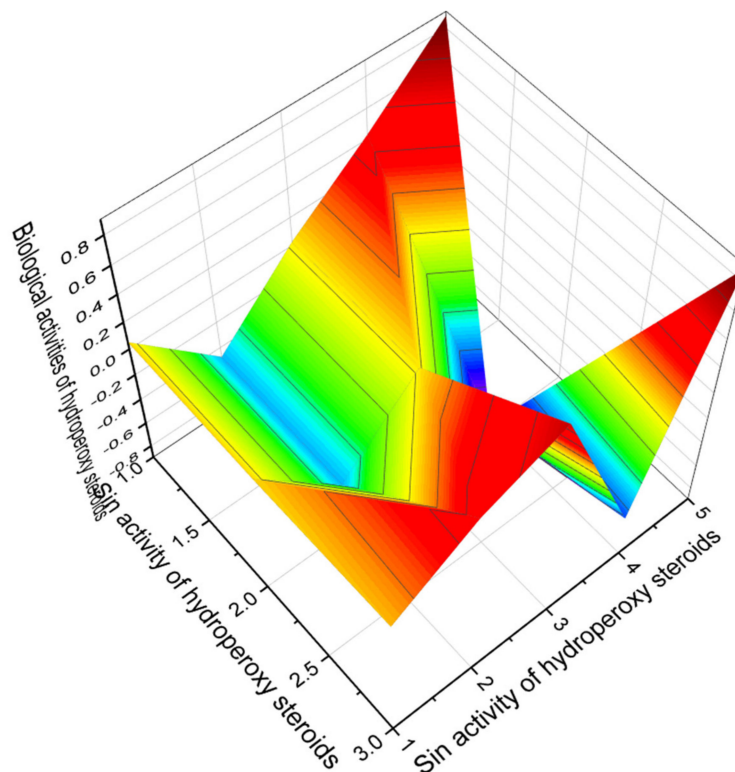


Figure 16. The 3D graph shows the predicted and calculated biological activity of steroid hydroperoxides (compound numbers: 59, 63, and 66) showing the highest degree of confidence, more than 93%.

5. Carbon-Bridged Steroids (CBS) and Triterpenoids

In both natural and synthetic steroids, when an additional ring is formed within the steroid skeleton, through a direct bond between any two carbon atoms (or more) of the steroid ring system or an attached side chain, such steroids (or triterpenoids) are called carbon-bridged steroids [13,200–202].

Of the more than 500 carbon-bridged steroids and triterpenoids studied, we found only twelve lipids that have a confidence level of more than 90 percent as potential lipid regulators. We give their description and their sources in nature below [13,203].

Studying the photoproducts obtained by photochemical processes of vitamin D, the cyclobutane containing derivative 67 was identified [204], and similar secosteroid named toxisterol (68), as a minor transformation product of vitamin D₂, has been found in various mushrooms [205]. The chemical structures of steroids and triterpenoids are shown in Figure 17, and the biological activity is shown in Table 7.

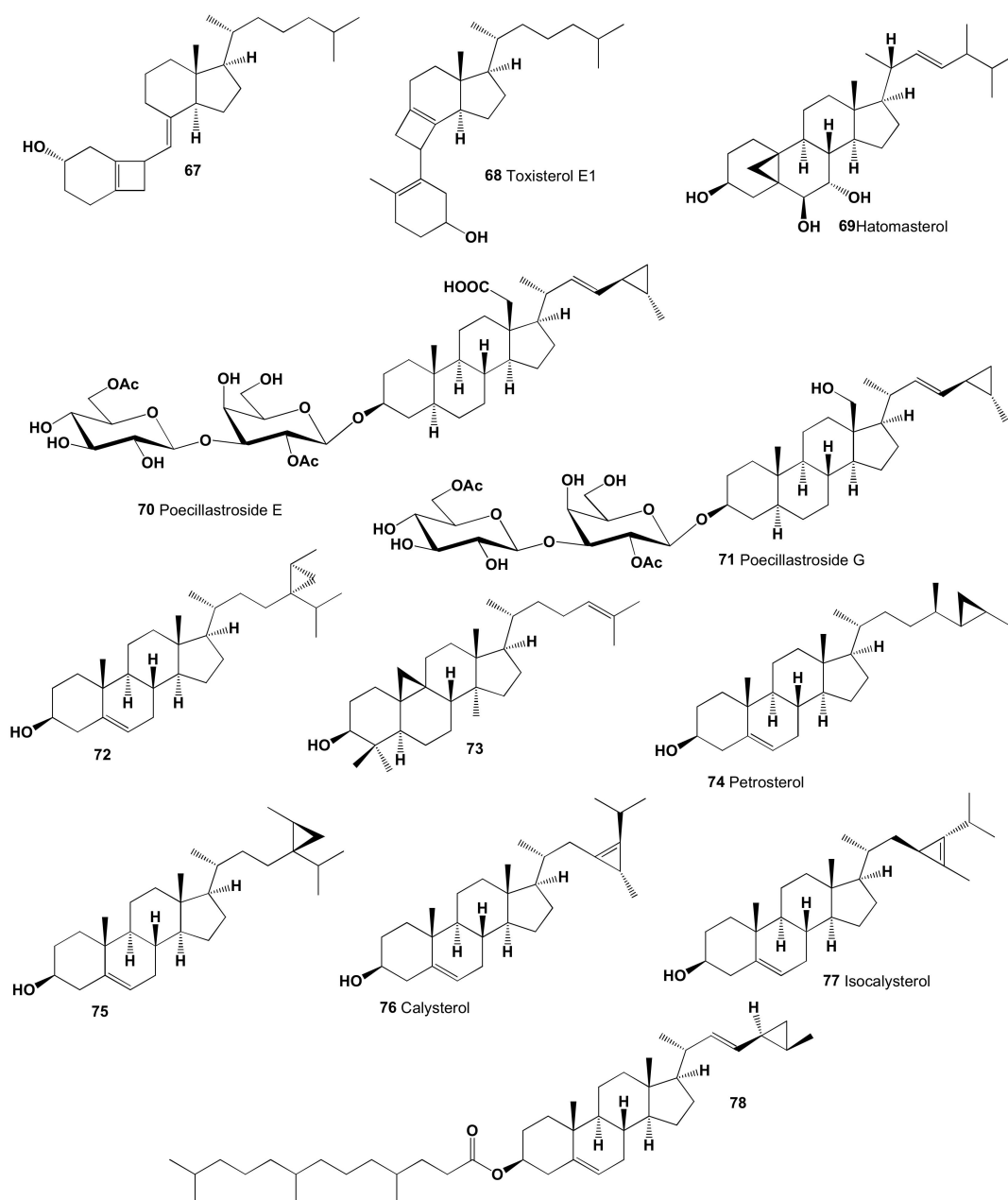


Figure 17. Bioactive CBS derived from fungi and marine sources.

A unique steroid containing a 5,19-cycloergostane skeleton, (3 β ,5 β ,6 β ,7 α ,22 E ,24 ζ)-5,19-cycloergost-22-ene-3,6,7-triol, named hatomasterol (69), was found in the extracts of the Okinawan sponge *Stylissa* sp., and this compound demonstrated cytotoxicity against HeLa cells in vitro [206]. Steroidal saponins named poecillastrosides E (70) and G (71), an oxidized methyl at C-18, into a primary alcohol or a carboxylic acid, have been found in extracts of the Mediterranean deep-sea sponge *Poecillastra compressa*. Poecillastroside E bearing a carboxylic acid at C-18 showed antifungal activity against *Aspergillus fumigatus* [207], and other cyclopropyl containing steroids (72) and (75) were found in the methanol extract of the marine sponge *Petrosia weinbergi* [208]. Cycloart-24-en-3-ol (73) was detected in ethanol extract of marine green alga *Cladophora fascicularis* [209].

A cytotoxic sterol named petrosterol (74) showed cytotoxic activities on A549, HL-60, MCF-7, SK-OV-3, and U937 cancer cell lines, and was present in extracts of several marine sponges such as Vietnamese sponge *Ianthella* sp., *Petrosia spheroida* from the Indian

Ocean, *Halichondria* cf. *panicea* of the Japanese island Iriomote, and Japanese marine sponge *Strongylophora corticate* [210–213].

Table 7. Predicted biological activities of CBS steroids.

No.	Discovered Activity, (Pa) *	Reported Activity	Ref.
67	Antihypercholesterolemic (0.902) Hypolipemic (0.721) Cholesterol synthesis inhibitor (0.534)	Activity not studied	
68	Antihypercholesterolemic (0.932) Hypolipemic (0.695) Cholesterol synthesis inhibitor (0.588)	Activity not studied	
69	Antineoplastic (0.915) Antihypercholesterolemic (0.900) Hypolipemic (0.897) Apoptosis agonist (0.892) Antineoplastic (liver cancer) (0.822) Chemopreventive (0.776) Atherosclerosis treatment (0.690) Cytoprotectant (0.611) Prostate cancer treatment (0.557) Antimetastatic (0.528)	Cytotoxic	[206]
70	Antihypercholesterolemic (0.953) Hypolipemic (0.758) Lipid metabolism regulator (0.674) Atherosclerosis treatment (0.513)	Antifungal	[207]
71	Antihypercholesterolemic (0.939) Hypolipemic (0.746) Lipid metabolism regulator (0.599)	No activity detected	[207]
72	Antihypercholesterolemic (0.923) Hypolipemic (0.732) Atherosclerosis treatment (0.643) Cholesterol synthesis inhibitor (0.640)	Activity not studied	
73	Hypolipemic (0.900) Atherosclerosis treatment (0.689) Cholesterol synthesis inhibitor (0.671) Antihypercholesterolemic (0.662) Lipid metabolism regulator (0.529)	Activity not studied	
74	Antihypercholesterolemic (0.964) Hypolipemic (0.849) Antineoplastic (0.849) Antihyperlipoproteinemic (0.801) Cholesterol synthesis inhibitor (0.671) Atherosclerosis treatment (0.610) Prostate cancer treatment (0.601)	Cytotoxic Anticancer	[211–214]
75	Antihypercholesterolemic (0.923) Hypolipemic (0.732) Atherosclerosis treatment (0.643) Cholesterol synthesis inhibitor (0.640)	Activity not studied	
76	Antihypercholesterolemic (0.935) Hypolipemic (0.731) Antihyperlipoproteinemic (0.689) Cholesterol synthesis inhibitor (0.600)	Activity not studied	
77	Antihypercholesterolemic (0.908) Hypolipemic (0.726) Cholesterol synthesis inhibitor (0.589) Antihyperlipoproteinemic (0.587)	Activity not studied	
78	Antihypercholesterolemic (0.969) Hypolipemic (0.810) Lipid metabolism regulator (0.716) Cholesterol synthesis inhibitor (0.707) Atherosclerosis treatment (0.586)	Activity not studied	

* Only activities with Pa > 0.5 are shown.

A rare steroid named calysterol (76), the minor sterol component of the sponge *Calyx niceaensis* and *Petrosia ficiformis*, possessing the unique feature of a cyclopropene ring bridging C23,24, and isocalysterol (77), was detected in the same sponge [214–218]. Sterol ester, 24,26-cyclo-5 α -cholest-(22E)-en-3 β -4',8',12'-trimethyltridecanoate (78), has been isolated from a deep-water marine sponge, *Xestospongia* sp. [218].

Comparison of Biological Activities of CBS Steroids and Triterpenoids

Carbon-bridged steroids (CBS) and triterpenoids belong to a rare group of natural hormones found in various natural sources such as green, yellow-green, and red algae, sea sponges, soft corals, ascidians, starfish, and other marine invertebrates. In addition, this group of rare lipids is found in amoebas, fungi, fungal endophytes, and plants [13,203].

We have isolated carbon-bridged steroids presented in Figure 18, which, according to the PASS data, have a confidence level of more than 90 percent. Among this group of lipids, we identified three, numbered 70, 74, and 78, the activity of which most clearly reflects their regulatory functions of lipid metabolism, with antihypercholesterolemic properties dominating. In addition, these steroids can be used as inhibitors of cholesterol synthesis and as drugs for the treatment of atherosclerosis. The 3D graph demonstrating the predicted and calculated biological activity of carbon-bridged steroids is shown in Figure 18.

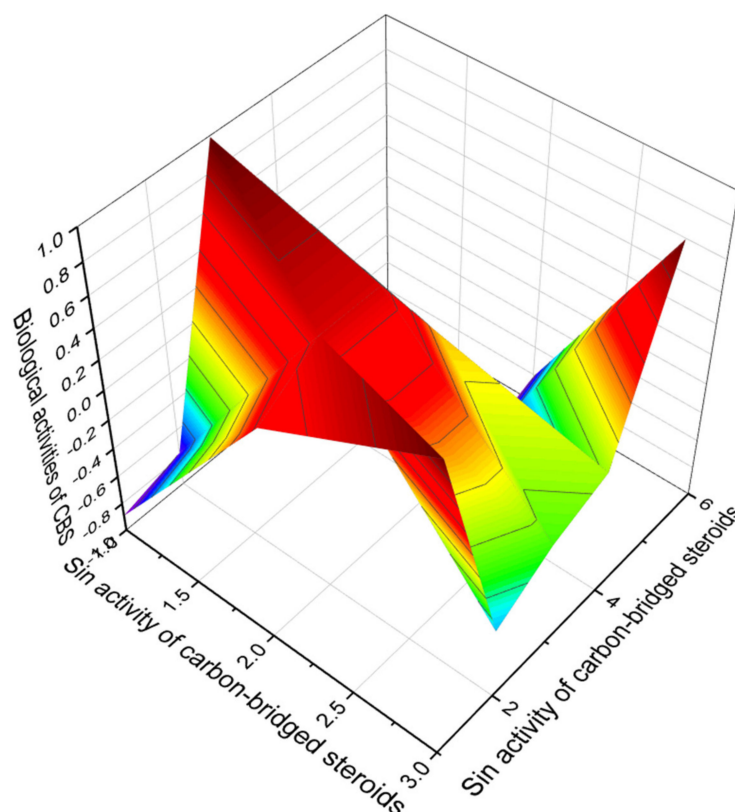


Figure 18. The 3D graph shows the predicted and calculated biological activity of carbon-bridged steroids (compound numbers: 70, 74, and 78) showing the highest degree of confidence, more than 95%.

6. Neo Steroids Derived from Terrestrial and Marine Sources

Secondary metabolites containing a tertiary butyl group (or tertbutyl unit) are rather rare compounds found in cyanobacteria, plant leaves, fungi, marine invertebrates, and algae [12,219–221].

Neo steroids are a small group of lipids that are synthesized by yeast and fungi and are found in various parts of plants. In recent years, with the improvement of steroid anal-

ysis methods, they have been found in seaweeds, marine sponges (class Demospongiae), anemones (class Anthozoa), and cucumbers (class Holothuroidea) [12,219].

Two neo steroids (79 and 85) were present in leaves and stems and the pericarp of the fruit and roots of a plant from the family Cucurbitaceae [222], and another sterol, 24-methylene-25-methyl-lathosterol (80), was isolated from aerial parts of the herbaceous plant, *Sicyos angulatus* [223]. The chemical structures of steroids and triterpenoids are shown in Figure 19, and the biological activity is shown in Table 8.

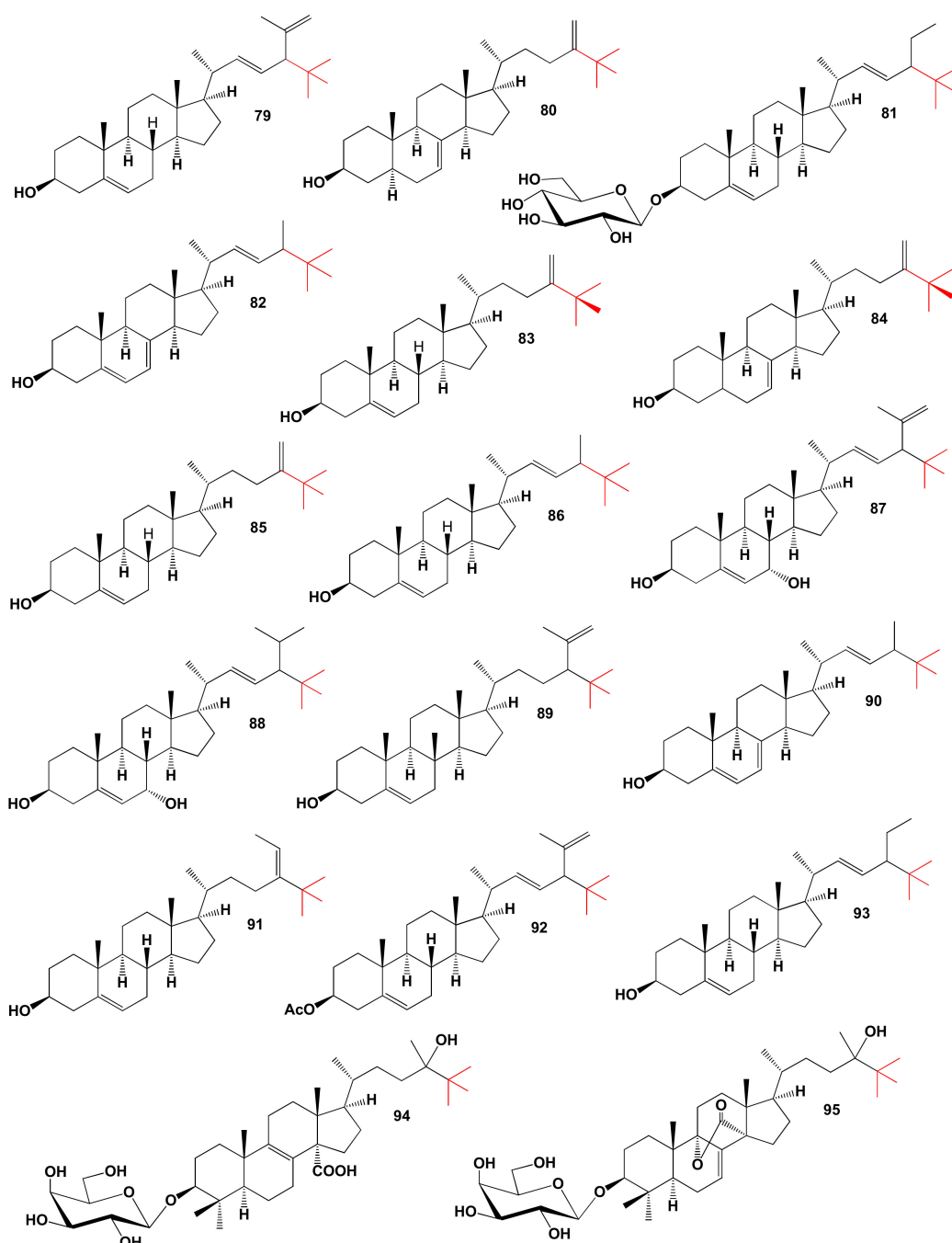


Figure 19. Bioactive neo steroids derived from plants and marine sources.

Aerial parts of the strongly aromatic herb *Ocimum basilicum* from the Labiatae family, such as seeds, flowers, and roots, are widely used as medicines [224–226]. The leaves and flowers of this plant are used in folk medicine as a tonic and anthelmintic [227]. Leaf tea is used to treat flatulence and dysentery, while the plant's oil may be useful for relieving

mental fatigue, colds, cramps, rhinitis, and as a first aid for treating wasp stings and snake bites [227,228]. A species of this plant from Pakistan contains (22E)-24 ξ -ethyl-25-methylcholesta-5,22-diene-3 β -ol-3-O-D-gluco-pyranoside (**81**) [229].

Three neo steroids (**82**, **83** and **86**) have been isolated from this auxotroph mutant *Saccharomyces cerevisiae* strain GL7 using appropriate substrates for biosynthesis [230]. The sterol C24-methyl transferase from *Trypanosoma brucei* TbSMT1 produces 24-methyl sterols that serve as substrates for 24-dimethyl sterols that contain a Δ 25(27)-bond, and the neo steroid (**84**) was isolated from the extract *S. cerevisiae* [231]. Topsentinols C (**87**) and E (**88**) contain a tertiary butyl group in steroids, and these neo steroids were obtained from the Okinawan marine sponge *Topsentia* sp. [232,233]. (3 β ,24E)-25-Methylstigmasta-5,24(28)-dien-3-ol (**89**) and axinyssasterol (**90**) were identified from a marine sponge *Pseudoaxinyssa* sp. [234].

Neo steroid (3 β ,22E,24 ξ)-28,28-dimethyl-stigmasta-5,22,25-trien-3-ol acetate (**91**) and (3 β ,22E)-25-methyl-stigmasta-5,22-dien-3-ol (**92**) were obtained from an ethanol extract of the sponge *Halichondria* sp. [235,236], and the compound (**93**) was also found in specimens of the sponge *T. aplysinoides* from the inshore waters of Sri Lanka [236]. The antimicrobial halistanol (**93**) was isolated from the Okinawan sponge *Halichondria* cf. *moorei* more than 25 years ago [237].

Table 8. Biological activities of neo steroids.

No.	Discovered Activity, (Pa) *	Reported Activity	Ref.
79	Antihypercholesterolemic (0.964) Atherosclerosis treatment (0.717)	Activity not studied	
80	Antihypercholesterolemic (0.956)	Activity not studied	
81	Antihypercholesterolemic (0.989) Hypolipemic (0.808)	Activity not studied	
82	Antihypercholesterolemic (0.960) Atherosclerosis treatment (0.683)	Activity not studied	
83	Antihypercholesterolemic (0.961) Cholesterol synthesis inhibitor (0.745)	Activity not studied	
84	Antihypercholesterolemic (0.956) Cholesterol synthesis inhibitor (0.747)	Activity not studied	
85	Antihypercholesterolemic (0.964)	Activity not studied	
86	Antihypercholesterolemic (0.969)	Activity not studied	
87	Antihypercholesterolemic (0.952) Atherosclerosis treatment (0.710)	Activity not studied	
88	Antihypercholesterolemic (0.965) Atherosclerosis treatment (0.677)	Activity not studied	
89	Antihypercholesterolemic (0.969)	Activity not studied	
90	Antihypercholesterolemic (0.965) Atherosclerosis treatment (0.700)	Activity not studied	
91	Antihypercholesterolemic (0.969)	Activity not studied	
92	Antihypercholesterolemic (0.962) Atherosclerosis treatment (0.704) Lipid metabolism regulator (0.702)	Activity not studied	

Table 8. Cont.

No.	Discovered Activity, (Pa) *	Reported Activity	Ref.
93	Antihypercholesterolemic (0.974) Antimicrobial treatment (0.717)	Antimicrobial	[237]
94	Antihypercholesterolemic (0.961) Antineoplastic (0.863) Anticarcinogenic (0.828) Lipid metabolism regulator (0.747) Antimetastatic (0.590)	Anticancer	[238,239]
95	Antihypercholesterolemic (0.936) Antineoplastic (0.870) Anticarcinogenic (0.807) Antimetastatic (0.598)	Anticancer	[238,239]

* Only activities with Pa > 0.5 are shown.

The Atlantic tropical sponge *Erylus goffrilleri* contains unusual lanostane glycosides, erylosides R (94) and T (95), and the same glycosides were isolated from the Caribbean sponge *E. goffrilleri*. Both eryloside glycosides R and T exhibit cytotoxic activities against Ehrlich carcinoma tumor cells [238,239].

Comparison of Biological Activities of Neo Steroids

Neo steroids are a rare group of naturally occurring lipid molecules that exhibit high levels and a wide range of activities. The chemical structures shown in Figure 19 have been found in plant and marine invertebrate extracts. According to PASS data, neo steroids show a high confidence level of up to 98.9 percent, with antihypercholesterolemic activity being dominant. In addition, virtually all neo steroids are cholesterol synthesis inhibitors and can be used to treat atherosclerosis and related diseases.

Of the seventeen neo steroids, we have selected four that show a high confidence level of 96.9 to 98.9 percent. Figure 20 demonstrates the 3D graph and shows the predicted and calculated biological activity of neo steroids (compound numbers: 81, 86, 89, and 93) showing the highest degree of confidence, more than 96.9%.

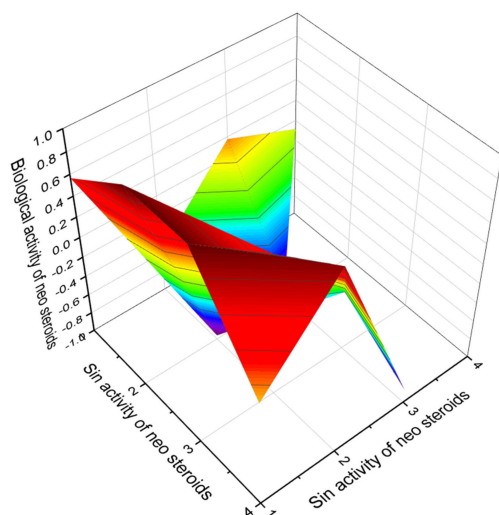


Figure 20. The 3D graph shows the predicted and calculated biological activity of neo steroids (compound numbers: 81, 86, 89, and 93) showing the highest degree of confidence, more than 96.9%.

7. Miscellaneous Steroids and Triterpenoids Derived from Marine Sources

In this section, we have collected steroids that do not belong to the first six groups of triterpenoids but show high antihypercholesterolemic activity. Mostly, the steroids shown in Figure 20 are found in soft corals collected in various regions of the world's oceans.

An interesting question is why from coral? The fact is that, by studying the activities of various marine organisms, we concluded that corals or their fungal endophytes synthesize many biologically active metabolites. It is well known that corals are associated with many microscopic fungi, so a reasonable question arises: what synthesizes bioactive molecules in corals? Whether they do so themselves or their fungal endophytes or bacteria is a question that remains open [240–242].

Cytotoxic steroid called stereosteroid G (96) was isolated from the methylene chloride extract of the Formosan soft coral *Stereonephthya crystalliana*. The extract of this coral showed significant cytotoxicity against A549, HT-29 and P-388 cancer cells in vitro [243]. The chemical structures of steroids and triterpenoids are shown in Figure 21, and the biological activity is shown in Table 9.

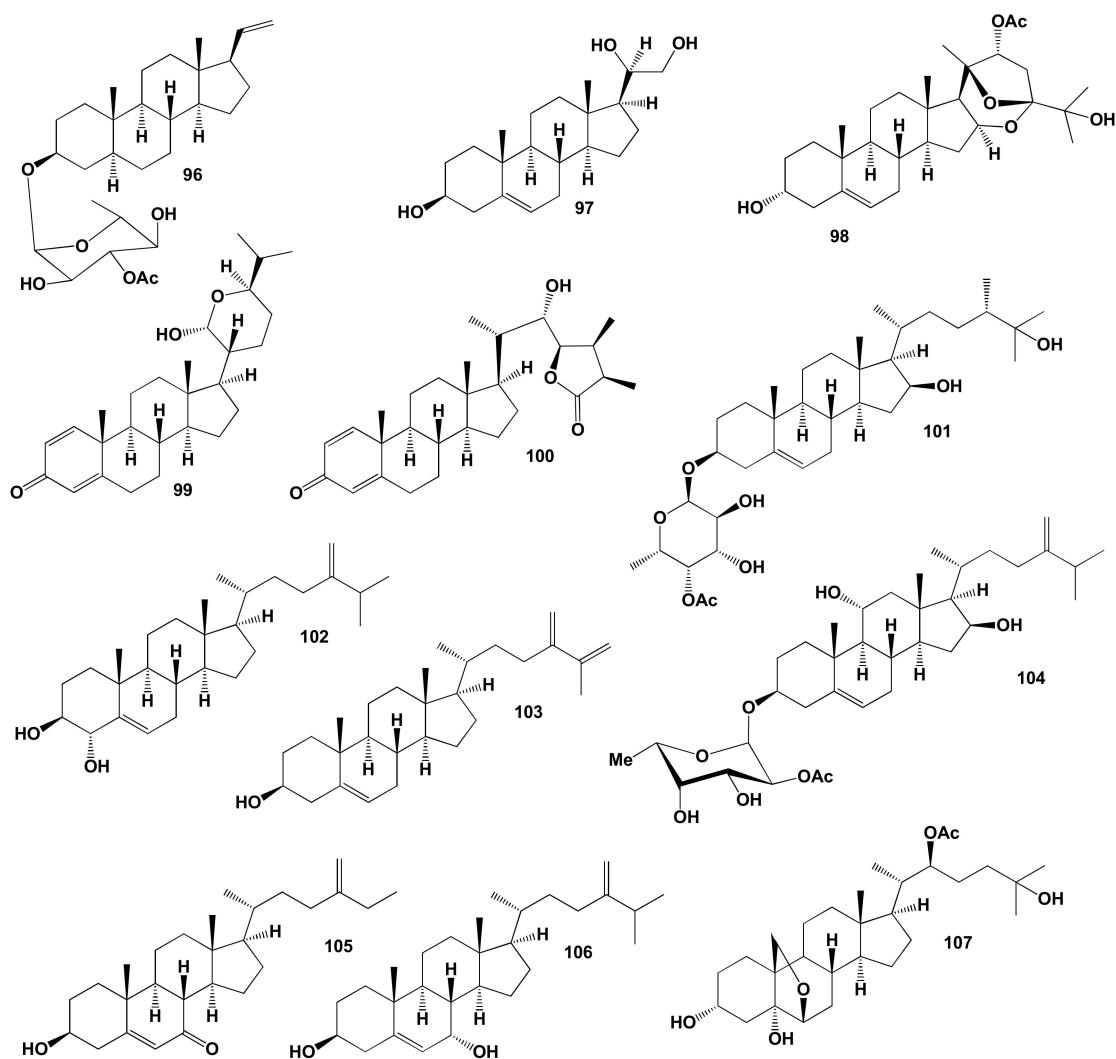


Figure 21. Bioactive steroids derived from soft corals.

Trihydroxy sterol, pregna-5-ene-3,20,21-triol (97), has been isolated from the Gulf of California gorgonian *Muricea cf. austera* [244], and a rare spiroketal steroid, 22-acetoxy-3,25-dihydroxy-16,24,20-bisepoxy-(3,16,20S,22R,24S)-cholest-5-ene (98) was found in extracts of the Indian Ocean gorgonian, *Gorgonella umbraculum* [245]. The cholestane class steroidal hemiacetals named anastomosacetal A (99) was obtained from the gorgonian coral *Euplexaura anastomosans*, collected off the shore of Keomun Island, South Sea Korea [246], and petasitosterone B (100) was isolated from a Formosan marine soft coral *Umbellulifera petasites* [247]. Petasitosterone B (100) displayed inhibitory activity against the proliferation

of a limited panel of cancer MOLT-4 and DLD-1 cell line. Steroidal glycoside (**101**) was isolated from water–methanol solutions of the soft coral *Sinularia gibberosa* [248], and steroid **102** was found in the methanol extract of the Vietnamese soft coral *Sinularia nanolobata* [249]. The minor sterol **103** was isolated from the soft coral extract of the genus *Sinularia*, and it was synthesized [250]. Steroid named crassarosteroside A (**104**) was obtained from *Sinularia granosa* and *S. crassa* soft coral extracts [251,252], compound **105** has been isolated from *Sinularia conferta* and *S. nanolobata* [253,254], and metabolite **106** was detected in MeOH extract of the soft coral *S. cruciata* [255]. Oxysterol (**107**) was detected in extracts of octocoral of the genus *Gorgonia* from the eastern Pacific, Panama [256].

Table 9. Biological activities of soft coral steroids.

No.	Discovered Activity, (Pa) *	Reported Activity	Ref.
96	Neuroprotector (0.983)	Anticancer	[243]
	Antihypercholesterolemic (0.919)		
	Acute neurologic disorders treatment (0.636)		
	Hypolipemic (0.626)		
97	Antihypercholesterolemic (0.954)	Activity not studied	
	Neuroprotector (0.754)		
	Hypolipemic (0.704)		
98	Antihypercholesterolemic (0.927)	Activity not studied	
	Neuroprotector (0.773)		
	Hypolipemic (0.584)		
99	Antihypercholesterolemic (0.962)	Activity not studied	
100	Antihypercholesterolemic (0.918)	Anti-inflammatory Anticancer	[247]
	Anti-inflammatory (0.903)		
	Antineoplastic (0.875)		
	Proliferative diseases treatment (0.856)		
	Atherosclerosis treatment (0.618)		
101	Antihypercholesterolemic (0.976)	Activity not studied	
	Hypolipemic (0.735)		
	Lipid metabolism regulator (0.707)		
102	Antihypercholesterolemic (0.920)	Activity not studied	
	Hypolipemic (0.798)		
	Atherosclerosis treatment (0.637)		
	Hyperparathyroidism treatment (0.523)		
103	Antihypercholesterolemic (0.955)	Activity not studied	
	Atherosclerosis treatment (0.596)		
104	Proliferative diseases treatment (0.967)	Anticancer	[251]
	Chemopreventive (0.958)		
	Antihypercholesterolemic (0.947)		
	Anticarcinogenic (0.907)		
	Antineoplastic (0.902)		
	Hypolipemic (0.781)		
Atherosclerosis treatment (0.609)			
105	Antihypercholesterolemic (0.957)	Activity not studied	
	Hypolipemic (0.809)		
	Atherosclerosis treatment (0.592)		
106	Antihypercholesterolemic (0.955)	Activity not studied	
	Hypolipemic (0.878)		
	Atherosclerosis treatment (0.658)		
107	Neuroprotector (0.938)	Activity not studied	
	Antihypercholesterolemic (0.912)		
	Atherosclerosis treatment (0.548)		

* Only activities with Pa > 0.5 are shown.

Comparison of Biological Activities of Soft Coral Steroids

The chemical and structural diversity of soft corals sterols and triterpenoids is well known and has been documented in several review articles in the literature [128,130,257–261]. PASS analysis shows that the steroids in Figure 21 synthesized by soft corals are indeed of interest due to their high anticholesterol activity. Both the biological activities found using PASS and those

obtained experimentally coincide. In addition, we have identified three steroids, numbered 99, 101, and 104, which demonstrate antihypercholesterolemic activity with a high degree of certainty. Figure 22 shows a 3D graph of predicted and calculated biological activity of these steroids.

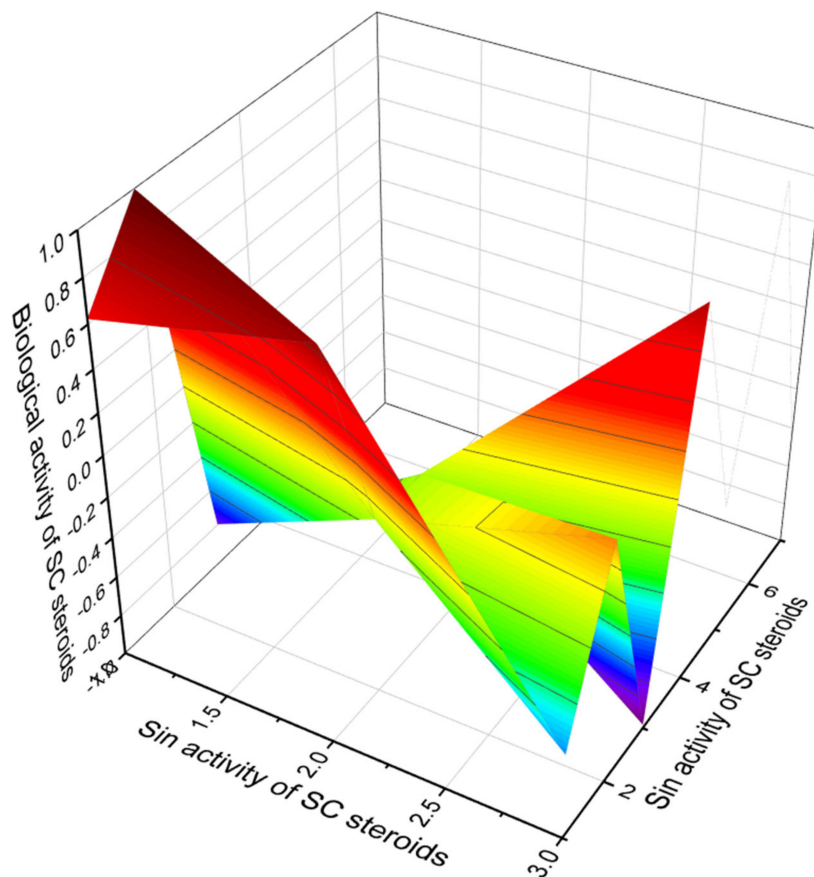


Figure 22. The 3D graph shows the predicted and calculated biological activity of soft coral steroids (compound numbers: 99, 101 and 104) showing the highest degree of confidence, more than 96.2%.

8. Synthetic and Semi-Synthetic Steroids and Triterpenoids and Comparison of Their Biological Activities

Steroid hormones and triterpenoids belong to the group of physiologically active substances (sex hormones, corticosteroids, etc.) that regulate vital processes in vertebrates and many species of invertebrates and humans [262–265]. They are regulators of the fundamental vital processes of a multicellular organism—coordinated growth, differentiation, reproduction, adaptation, and behavior [266–268]. For more than 90 years, steroid hormones and triterpenoids have been the subject of close attention of chemists since synthetic analogues have long replaced natural steroids and triterpenoids [269–273]. It is known that semi-synthetic or synthetic steroid hormones have properties that steroids do not have, isolated from natural sources. In this section, we present synthetic and semi-synthetic steroids and triterpenoids that contain heteroatoms and do not exist in nature but demonstrate activities that are necessary for pharmacologists and physicians.

8.1. Bioactive Epithio Steroids and Triterpenoids

Anabolic steroids are pharmacological drugs that mimic the effect of the male sex hormone testosterone and its derivatives [274–276]. Anabolic steroids accelerate the synthesis of protein within cells, which leads to a pronounced hypertrophy of the muscle tissue, because of which, they have found wide application in sports medicine and bodybuilding [277–279].

Semi- and/or synthetic epithio steroids represent a rare group of bioactive lipids, since they are hydrophobic molecules insoluble in water, which are not found in nature. Epithio steroids have been reported to possess a variety of cytotoxic activities, and they are widely used as anticancer agents. The thiirane group is an important substance and shows some promising biological activities. Steroids containing an epithio group in positions 2 and 3 are anabolic steroids and are widely known and used in sports medicine. Representatives of this group of steroids are of great interest for pharmaceutical chemistry and medicine [280–286]. The most widely known are such epithio steroids that are used in sports pharmacology and medicine: epistane (108, 2 α ,3 α -epithio-17 α -methyl-5 α -androstan-17 β -ol), epitio stanol (109, 2 α ,3 α -epithio-5 α -androstan-17 β -ol), a known potent antiestrogenic and antitumor agent, and hemapolin (110, 2 β ,3 β -epithio-17 α -methyl-5 α -androstan-17 β -ol) [282,283,287–291]. The chemical structures of epithio steroids are shown in Figure 23 and the biological activities are shown in Table 10. Presented in Figure 24, the anabolic 2 α ,3 α -epithio chlostan (111) shown dominant anticancer activity, and steroid (112) demonstrated dominant properties as an antisecretory agent with 96.7% confidence and acts as an estrogen antagonist with 94.6% confidence.

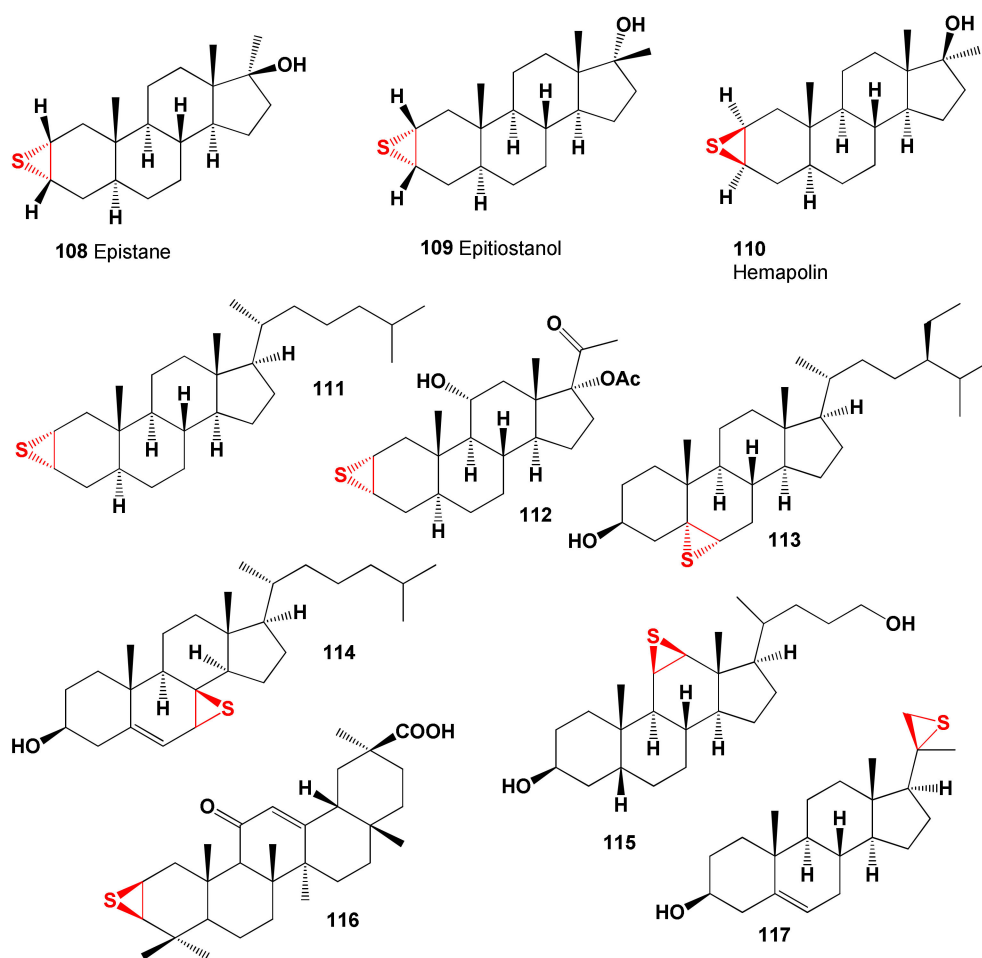


Figure 23. Bioactive epithio steroids and triterpenoid containing the thiirane group in a variety of steroid backbones.

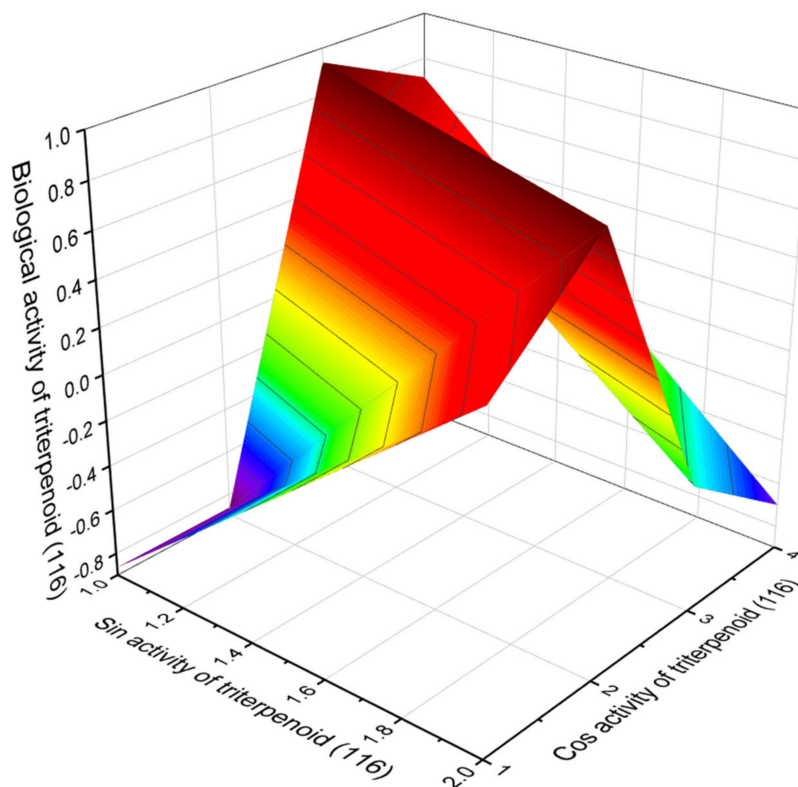


Figure 24. The 3D graph shows the predicted and calculated biological activity of triterpenoid (116), which was synthesized from a natural sample of 18β-glycyrrhetic acid, showing the highest degree of confidence as lipid metabolism regulator properties, more than 95.4%. 18β-Glycyrrhetic acid is a β-amyrin-type pentacyclic triterpenoid derived from the herb licorice (*Glycyrrhiza glabra*). It is known and used as a flavoring agent and masks the bitter taste of drugs such as aloe and quinine; it is also effective in the treatment of peptic ulcer disease and has some additional pharmacological properties with possible antiviral, antifungal, antiprotozoal, and antibacterial effects.

Epithio steroids (113–115 and 117) and are cholesterol antagonists, and the anticancer triterpenoid (116), which was synthesized from a natural sample of 18β-glycyrrhetic acid [292,293], shows lipid metabolism regulator properties.

Table 10. Biological activities of epithio steroids.

No.	Discovered Activity, (Pa) *	Reported Activity	Ref.
108	Antineoplastic (0.964)	Anti-breast cancer Estrogen receptor antagonist	[294]
	Antisecretoric (0.948)		
	Estrogen antagonist (0.860)		
	Cardiotonic (0.729)		
	Prostate disorders treatment (0.709)		
	Neuroprotector (0.723)		
	Bone diseases treatment (0.693)		
Antineoplastic (breast cancer) (0.598)			
109	Antineoplastic (0.966)	Estrogen antagonist	[294]
	Antisecretoric (0.952)		
	Estrogen antagonist (0.832)		
	Anti-inflammatory (0.754)		
	Prostate disorders treatment (0.736)		
Prostatic (benign) hyperplasia treatment (0.673)			

Table 10. Cont.

No.	Discovered Activity, (Pa) *	Reported Activity	Ref.
110	Antineoplastic (0.966) Antisecretoric (0.952) Estrogen antagonist (0.832) Anti-inflammatory (0.754) Prostate disorders treatment (0.736) Prostatic (benign) hyperplasia treatment (0.673)	Estrogen antagonist	[294]
111	Antineoplastic (0.932) Antihypercholesterolemic (0.759) Bone diseases treatment (0.729) Hypolipemic (0.676) Estrogen antagonist (0.660)	Anabolic	[294]
112	Antisecretoric (0.967) Estrogen antagonist (0.946) Antineoplastic (0.939) Anabolic (0.823)	Anabolic	[284–286]
113	Cholesterol antagonist (0.933) Antihypercholesterolemic (0.929) Hypolipemic (0.818) Estrogen antagonist (0.443)	Antiseptic, Germicidal, Fungicidal	[294]
114	Cholesterol antagonist (0.946) Antihypercholesterolemic (0.930) Hypolipemic (0.781) Estrogen antagonist (0.465)	Antiseptic, Germicidal, Fungicidal	[294]
115	Cholesterol antagonist (0.932) Antihypercholesterolemic (0.900) Cardiotonic (0.886) Choleretic (0.871) Atherosclerosis treatment (0.838) Antineoplastic (0.775) Hypolipemic (0.746) Estrogen antagonist (0.611)	DOCA inhibitor	[294]
116	Lipid metabolism regulator (0.954) Antineoplastic (0.924) Apoptosis agonist (0.869) Estrogen antagonist (0.505)	Anticancer	[292,295]
117	Cholesterol antagonist (0.916) Antihypercholesterolemic (0.836) Hypolipemic (0.801)	Activity not studied	

* Only activities with Pa > 0.5 are shown.

8.2. Bioactive Seleno Steroids

Selenium is an essential metalloid, and it is one of the most necessary trace elements for humans [296]. Selenium occupies an important place in the regulation of metabolism in humans, and therefore, it is necessary to monitor its presence in consumed foods [297,298]. The Allium and Brassica families as well as Brazil nuts, mushrooms (shiitake and white mushrooms), beans, chia seeds, brown rice, sunflower, sesame and flax seeds, and cabbage and spinach contain high enough selenium and organoselenium concentrations [299,300].

There are also many excellent reviews in the literature, which are devoted to the biological role and functions of organoselenium compounds [301–304]. Apparently, seleno steroids are the main group of the essential metalloids that have been synthesized over the past 50 years, and approximately 300 have been synthesized [304–309].

The seleno steroids numbered 118, 119, 124, 125, 126, and 127 show dominant antihypercholesterolemic activity with a low degree of confidence from 90.5 to 91.2%, although

for steroid number 128, the confidence was 95.3%. For another group, the selena steroids numbered 120, 121, 122, and 123, the dominant properties are hyperlipemia, treatment of atherosclerosis, and treatment of lipoprotein disorders with a strong degree of confidence up to 99.6%. The chemical structures of steroids are shown in Figure 25, and the biological activity is shown in Table 11. Figure 26 shows the 3D graph the predicted and calculated biological activity of the selena steroids numbered 120, 121, and 122 with dominant properties as hyperlipemia and treatment of atherosclerosis.

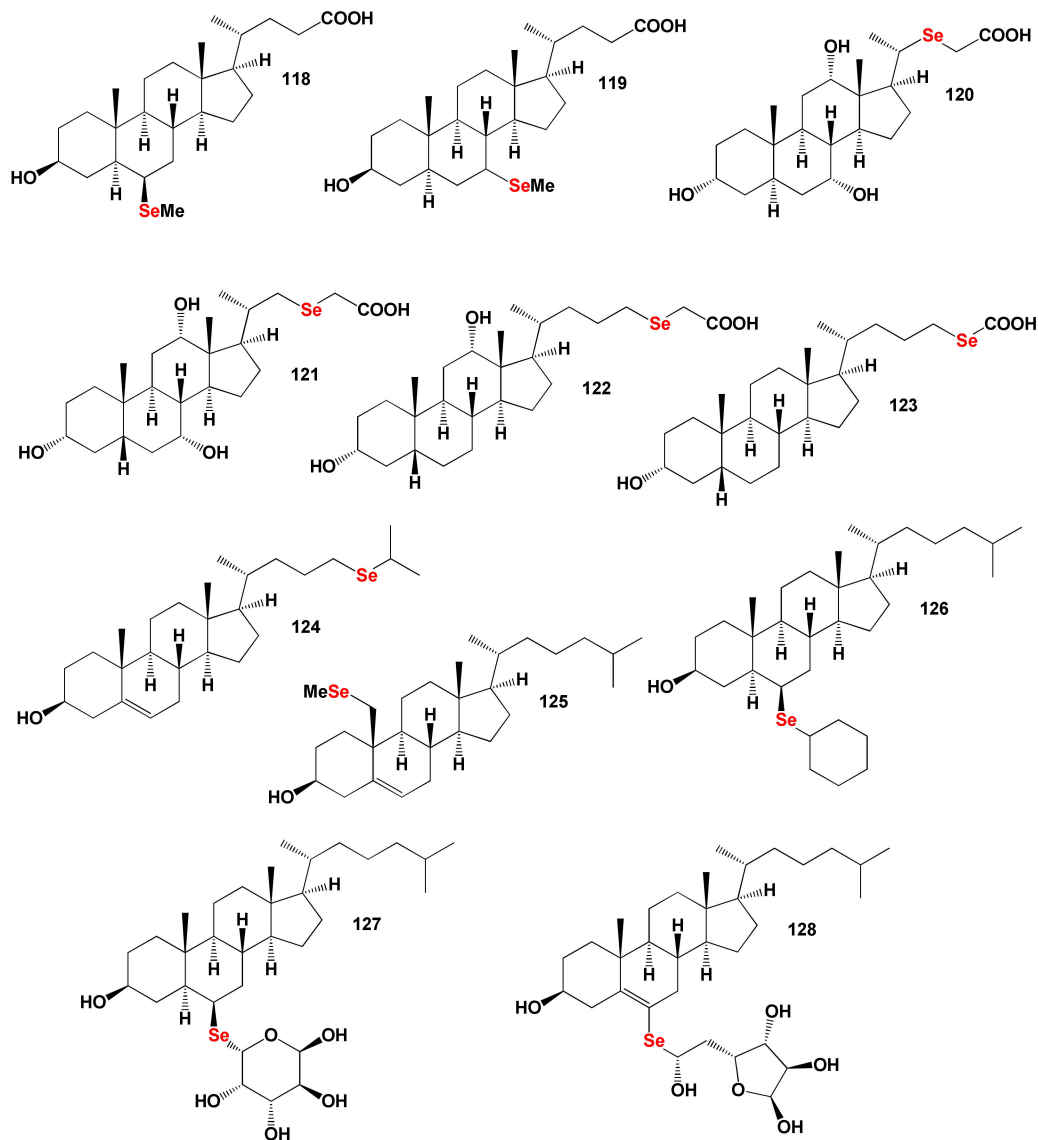


Figure 25. Bioactive synthetic selena steroids.

Table 11. Biological activities of the selena steroids.

No.	Discovered Activity, (Pa) *	Reported Activity	Ref.
118	Antihypercholesterolemic (0.905)	Activity not studied	
119	Choleretic (0.909) Antihypercholesterolemic (0.905)	Activity not studied	
120	Hypolipemic (0.995) Atherosclerosis treatment (0.991) Lipoprotein disorders treatment (0.982) Antioxidant (0.973) Erythropoiesis stimulant (0.823) Biliary tract disorders treatment (0.808) Laxative (0.709)	Activity not studied	
121	Hypolipemic (0.995) Atherosclerosis treatment (0.989) Lipoprotein disorders treatment (0.980) Antioxidant (0.970) Biliary tract disorders treatment (0.808) Erythropoiesis stimulant (0.730) Laxative (0.683)	Activity not studied	
122	Hypolipemic (0.996) Atherosclerosis treatment (0.995) Lipoprotein disorders treatment (0.991) Antioxidant (0.978) Erythropoiesis stimulant (0.756)	Activity not studied	
123	Hypolipemic (0.913) Antihypercholesterolemic (0.884) Atherosclerosis treatment (0.822)	Activity not studied	
124	Antihypercholesterolemic (0.902) Hypolipemic (0.899) Atherosclerosis treatment (0.791)	Activity not studied	
125	Antihypercholesterolemic (0.905)	Activity not studied	
126	Antihypercholesterolemic (0.908) Bone diseases treatment (0.772) Hypolipemic (0.769)	Activity not studied	
127	Antihypercholesterolemic (0.912) Antineoplastic (0.884) Anticarcinogenic (0.753)	Cytotoxic activity Agent for Alzheimer's disease	[303,310]
128	Antihypercholesterolemic (0.953) Antineoplastic (0.856) Anticarcinogenic (0.804)	Cytotoxic activity Agent for Alzheimer's disease	[303,310]

* Only activities with Pa > 0.5 are shown.

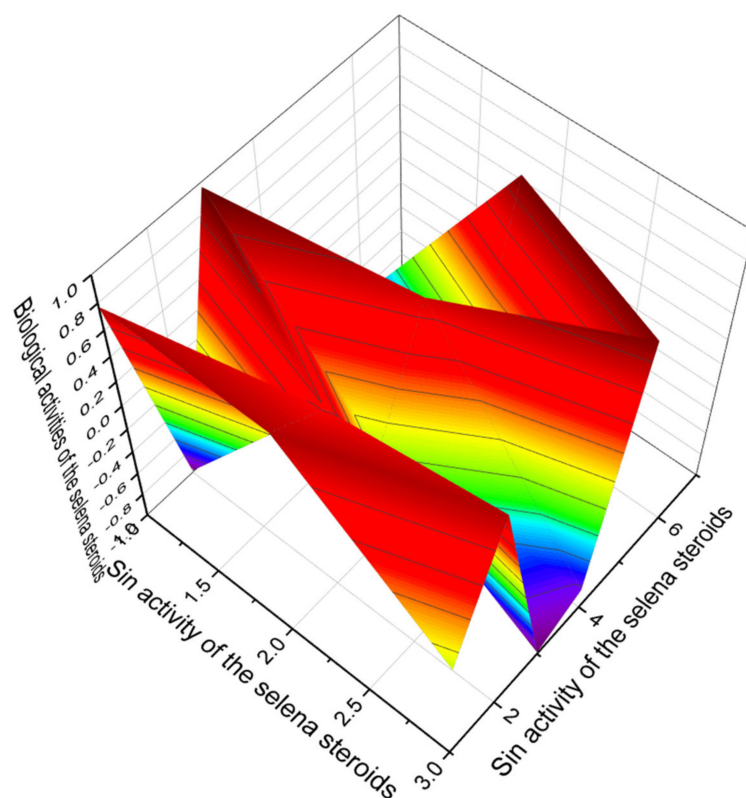


Figure 26. The 3D graph shows the predicted and calculated biological activity of the selena steroids (compound numbers: 120, 121 and 122) showing the highest degree of confidence, more than 99.5%.

8.3. Bioactive Tellura Steroids

Tellura steroids are a rare group of organic synthetic compounds whose biological activity is of great interest for medicine, pharmacology, and the pharmaceutical industry [304,306,308,309,311]. The chemical structures of steroids and triterpenoids are shown in Figure 27, and the biological activity is shown in Table 12.

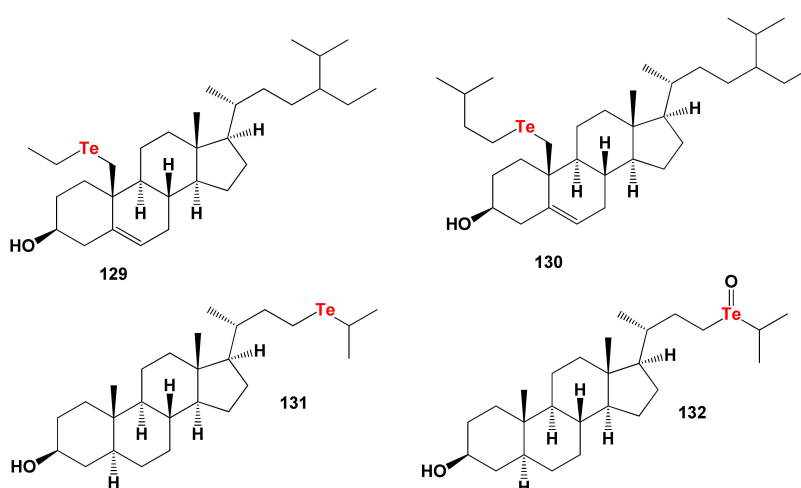


Figure 27. Bioactive synthetic tellura steroids.

Table 12. Biological activities of tellura steroids.

No.	Discovered Activity, (Pa) *	Reported Activity
129	Atherosclerosis treatment (0.977)	Activity not studied
	Antioxidant (0.963)	
	Antihypercholesterolemic (0.956)	
	Antiparkinsonian (0.955)	
	Neurodegenerative diseases treatment (0.954)	
	Alzheimer's disease treatment (0.940)	
Antihyperlipoproteinemic (0.811)		
130	Antihypercholesterolemic (0.958)	Activity not studied
	Atherosclerosis treatment (0.890)	
	Alzheimer's disease treatment (0.838)	
	Antioxidant (0.807)	
131	Neurodegenerative diseases treatment (0.801)	Activity not studied
	Antioxidant (0.922)	
	Atherosclerosis treatment (0.908)	
	Neurodegenerative diseases treatment (0.877)	
	Antihypercholesterolemic (0.869)	
132	Alzheimer's disease treatment (0.868)	Activity not studied
	Antiparkinsonian (0.848)	
	Antihypercholesterolemic (0.909)	
	Atherosclerosis treatment (0.876)	
	Alzheimer's disease treatment (0.828)	
132	Neurodegenerative diseases treatment (0.808)	Activity not studied
	Biliary tract disorders treatment (0.807)	

* Only activities with Pa > 0.5 are shown.

We found only four tellura steroids, which exhibit properties as regulators of lipid metabolism, dominated by antihypercholesterolemic activity. However, the most interesting from the point of view of pharmacological values is tellura steroid with number **129**. In addition to its antihypercholesterolemic activity, it is worth pointing out that this steroid has also been shown to be used as an agent for the treatment of neurodegenerative diseases Alzheimer's and Parkinson's with strong confidence, over 94 percent. The 3D graph demonstrating the predicted and calculated biological activity of the tellura steroid (**129**) is shown in Figure 28.

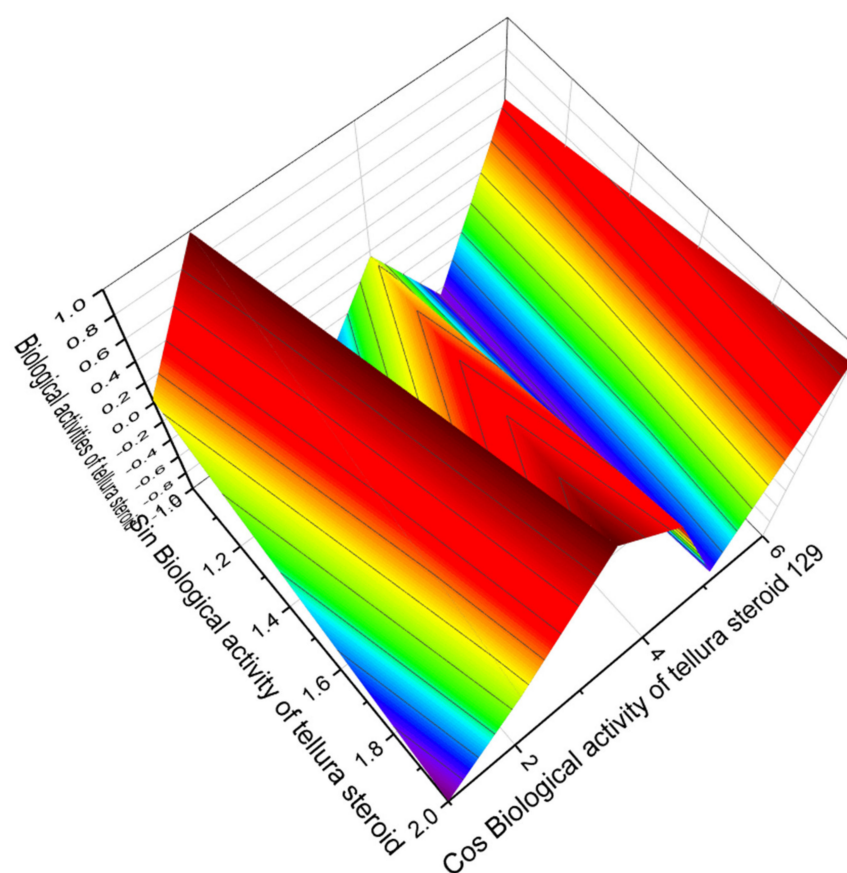


Figure 28. The 3D graph shows the predicted and calculated biological activity of the tellura steroid (129) showing the highest degree of confidence, more than 99.5%. This tellura steroid is interesting, in that it is a rare case when any chemical compound shows simultaneously such activities as prevention and treatment of neurodegenerative diseases Alzheimer’s and Parkinson’s with strong confidence, over 94 percent. In addition, this steroid demonstrated antioxidant and antihypercholesterolemic activities, and can also be used as a potential drug for the treatment of atherosclerosis. The maximum values of various biological activities are shown in red.

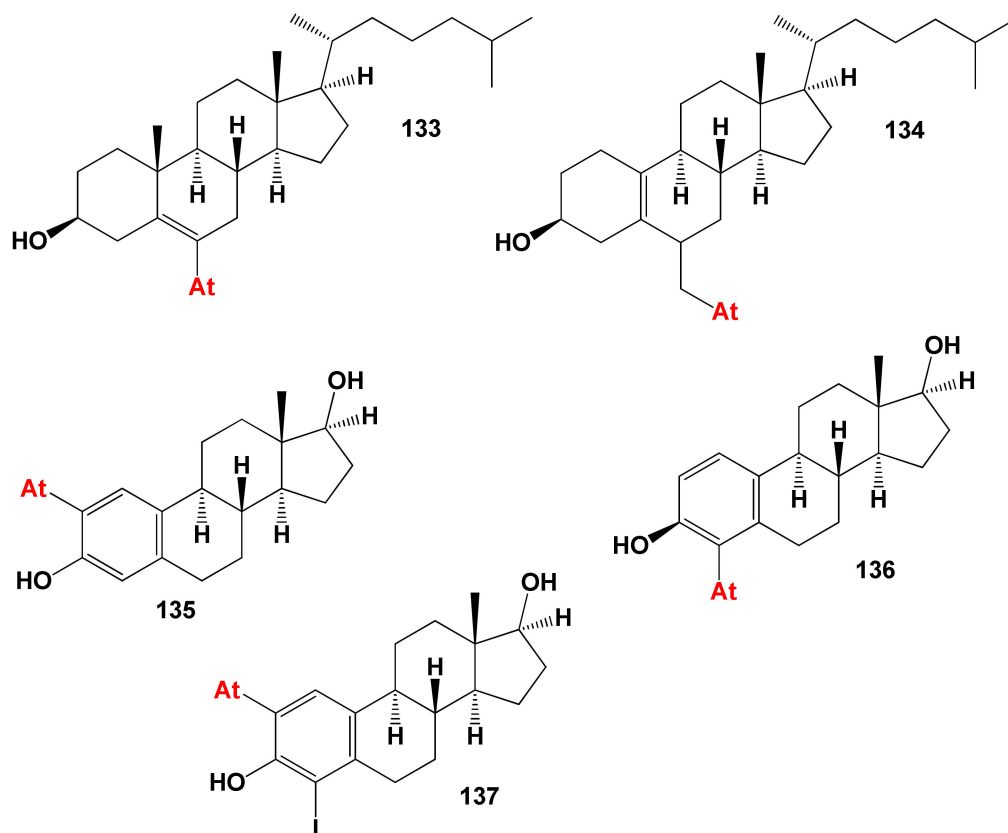
8.4. Bioactive Astatosteroids

Astatine is natural radioelement that has short-lived isotopes, and synthetic organic astatine compounds are commonly used for radiotherapy [312–314]. Steroids containing astatine, which are called astatosteroids, were first synthesized approximately 40 years ago [315]. Some astatosteroids (2- and 4-astatoestradiol and 6-At-cholesterol, **135**, **136**, and **137**) have been synthesized in high radiochemical yields by the reaction of $^{211}\text{At}/\text{I}_2$ and the corresponding chloromercury compounds. The stability in vitro was determined under different conditions in comparison with the analogous iodo compounds [313]. More recently, 6-astatomethyl-19-norcholest-5(10)-en-3 β -ol (**134**) was synthesized at a yield of 60–70% [316]. The biological activity of these compounds has not been determined. The predicted biological activity of astatosteroids is presented in Table 13. The most characteristic biological properties for these steroids were antineoplastic, antiseborrheic, antisecretoric, and antihypercholesterolemic activities. The chemical structures of steroids are shown in Figure 29 and the biological activity is shown in Table 13. For all astatosteroids shown in Figure 29, antihypercholesterolemic activity is dominated. In addition, all steroids of this group, as shown by PASS, have properties as a treatment for bone diseases. This is a rare property for steroids. Figure 30 shows the 3D graph the predicted and calculated biological activity of the astatosteroid (**133**).

Table 13. Biological activities of astatosteroids.

No.	Discovered Activity, (Pa) *	Reported Activity
133	Antihypercholesterolemic (0.967)	Activity not studied
	Antineoplastic (0.824)	
	Bone diseases treatment (0.796)	
	Hypolipemic (0.785)	
	Neuroprotector (0.758)	
	Antipsoriatic (0.739)	
	Anti-inflammatory (0.728)	
	Apoptosis agonist (0.724)	
Prostate disorders treatment (0.719)		
134	Antihypercholesterolemic (0.927)	Activity not studied
	Bone diseases treatment (0.784)	
	Hypolipemic (0.740)	
135	Antihypercholesterolemic (0.920)	Activity not studied
	Growth stimulant (0.805)	
	Bone diseases treatment (0.743)	
136	Antihypercholesterolemic (0.901)	Activity not studied
	Bone diseases treatment (0.720)	
	Growth stimulant (0.703)	
137	Antihypercholesterolemic (0.912)	Activity not studied
	Bone diseases treatment (0.777)	

* Only activities with Pa > 0.5 are shown.

**Figure 29.** Biological active synthetic astatosteroids.

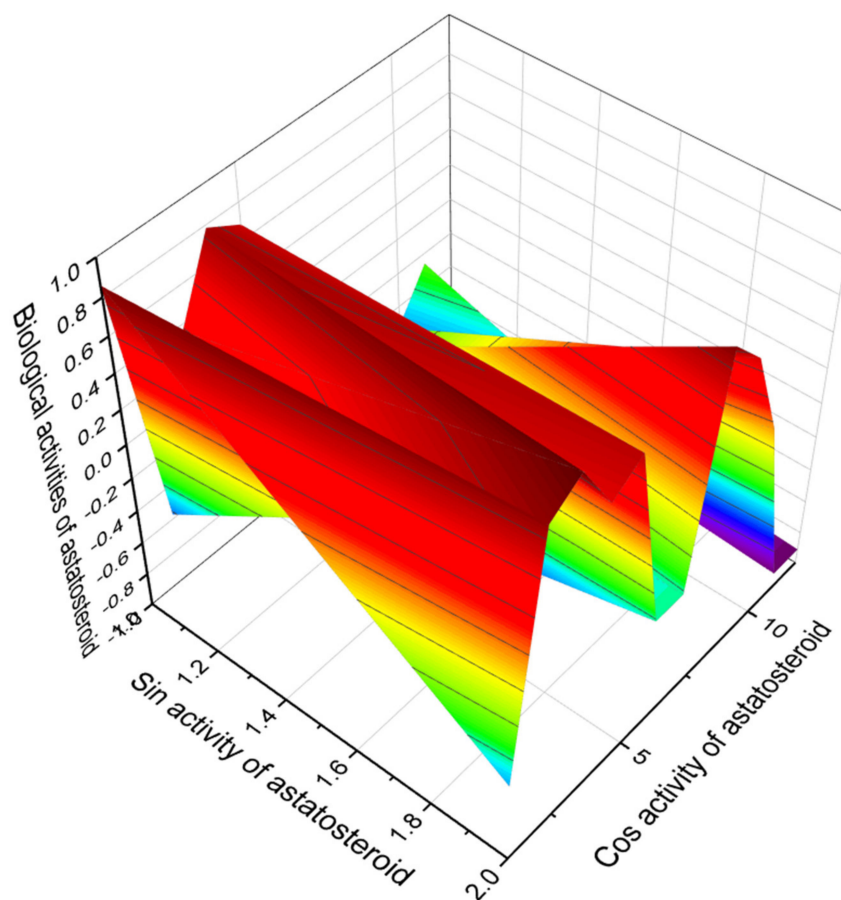


Figure 30. The 3D graph shows the predicted and calculated biological activity of the astatosteroid (133) showing the highest degree of confidence, more than 97.6%. Moreover, it can also be used as a potential drug for the treatment of bone diseases.

9. Conclusions

This review focuses on the intriguing topic of lipid metabolism regulation. The literature does not fully describe the means that regulate lipid metabolism. Steroids and triterpenoids presented in this review are of great interest for medicine, and some of them may be potential regulators of lipid metabolism. However, experimental work is required to confirm this thesis. In the world in general, and in North America in particular, the study of biological activities using computer programs is gaining popularity. This is due to the fact that the number of isolated natural and synthetic compounds has long exceeded 20 million; there is no technical possibility of determining biological activity experimentally. Using the PASS program for the last fifteen years has shown that we are on the right track. During this time, we have investigated over 10,000 compounds and identified their potential biological activities. Based on early studies, we have selected a group of steroids and triterpenoids that are presented in this review and correspond to the name of the topic, as potential regulators of lipid metabolism.

Funding: This work did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Acknowledgments: The author is grateful to Anne Kathrin Bendt (Life Sciences Institute, National University of Singapore, Singapore) who initiated this review, and grateful to Tatyana A. Glorizova (Institute of Biomedical Chemistry, Moscow, Russia) for prompt help in determining the biological activity of the steroids and triterpenoids presented in the article.

Conflicts of Interest: The author declares that he has no known competing financial interests or personal relationships that could affect the work described in this article.

References

1. Cheng, X.; Smith, J.C. Biological membrane organization and cellular signaling. *Chem. Rev.* **2019**, *119*, 5849–5880. [[CrossRef](#)]
2. Jensen, M.Ø.; Mouritsen, O.G. Lipids do influence protein function—The hydrophobic matching hypothesis revisited. *Biochim. Biophys. Acta Biomembr.* **2004**, *1666*, 205–226. [[CrossRef](#)] [[PubMed](#)]
3. MacCallum, J.L.; Tieleman, D.P. Hydrophobicity scales: A thermodynamic looking glass into lipid–protein interactions. *Trends Biochem. Sci.* **2011**, *36*, 653–662. [[CrossRef](#)] [[PubMed](#)]
4. Lundstedt, E.; Kahne, D.; Ruiz, N. Assembly, and maintenance of lipids at the bacterial outer membrane. *Chem. Rev.* **2021**, *121*, 5098–5123. [[CrossRef](#)] [[PubMed](#)]
5. Suzumura, M. Phospholipids in marine environments: A review. *Talanta* **2005**, *66*, 422–434. [[CrossRef](#)]
6. Li, J.; Wang, X.; Zhang, T.; Wang, C.; Huang, Z.; Luo, X.; Deng, Y. A review on phospholipids and their main applications in drug delivery systems. *Asian J. Pharm. Sci.* **2015**, *10*, 81–98. [[CrossRef](#)]
7. Dowhan, W. Molecular basis for membrane phospholipid diversity: Why are there so many lipids? *Ann. Rev. Biochem.* **1997**, *66*, 199–232. [[CrossRef](#)] [[PubMed](#)]
8. Berne, B.J.; Weeks, J.D.; Zhou, R. Dewetting and hydrophobic interaction in physical and biological systems. *Ann. Rev. Phys. Chem.* **2009**, *60*, 85–103. [[CrossRef](#)]
9. Gisterå, A.; Hansson, G. The immunology of atherosclerosis. *Nat. Rev. Nephrol.* **2017**, *13*, 368–380. [[CrossRef](#)]
10. Alexopoulos, N.; Raggi, P. Calcification in atherosclerosis. *Nat. Rev. Cardiol.* **2009**, *6*, 681–688. [[CrossRef](#)]
11. Faulds, M.H.; Zhao, C.; Dahlman-Wright, K.; Gustafsson, J.Å. The diversity of sex steroid action: Regulation of metabolism by estrogen signaling. *J. Endocrinol.* **2011**, *212*, 3–12. [[CrossRef](#)]
12. Dembitsky, V.M. Antitumor and hepatoprotective activity of natural and synthetic neo steroids. *Prog. Lipid Res.* **2020**, *79*, 101048. [[CrossRef](#)]
13. Dembitsky, V.M.; Glorizova, T.A.; Poroikov, V.V. Antitumor profile of carbon-bridged steroids (CBS) and triterpenoids. *Mar. Drugs* **2021**, *19*, 324. [[CrossRef](#)]
14. Pounina, T.A.; Glorizova, T.A.; Savidov, N.; Dembitsky, V.M. Sulfated and sulfur-containing steroids and their pharmacological profile. *Mar. Drugs* **2021**, *19*, 240. [[CrossRef](#)]
15. Zagoskin, P.P.; Erlykina, E.I. Bile acids as a new type of steroid hormones regulating nonspecific energy expenditure of the body (review). *Sovremen. Tehmol. Med.* **2020**, *12*, 114. [[CrossRef](#)]
16. Ko, C.W.; Qu, J.; Black, D.D.; Tso, P. Regulation of intestinal lipid metabolism: Current concepts and relevance to disease. *Nat. Rev. Gastroenterol. Hepatol.* **2020**, *17*, 169–183. [[CrossRef](#)]
17. De Bose-Boyd, R.A. Significance, and regulation of lipid metabolism. *Semin. Cell Dev. Biol.* **2018**, *81*, 97. [[CrossRef](#)]
18. Luo, J.; Yang, H.; Song, B.L. Mechanisms, and regulation of cholesterol homeostasis. *Nat. Rev. Mol. Cell Biol.* **2020**, *21*, 225–245. [[CrossRef](#)]
19. Schoeler, M.; Caesar, R. Dietary lipids, gut microbiota, and lipid metabolism. *Rev. Endocr. Metab. Disord.* **2019**, *20*, 461–472. [[CrossRef](#)]
20. Matsushita, Y.; Nakagawa, H.; Koike, K. Lipid metabolism in oncology: Why it matters, how to research, and How to treat. *Cancers* **2021**, *13*, 474. [[CrossRef](#)]
21. Edgar, A.; Doisy, E.A. An ovarian hormone: Preliminary report on its localization, extraction and partial purification, and action in test animals. *J. Am. Med. Assoc.* **1923**, *81*, 819–821.
22. Doisy, E.A.; Clement, D.V.; Sidney, T. Folliculin from urine of pregnant women. *Am. J. Phys.* **1929**, *90*, 329–330.
23. Butenandt, A.B. Über Progynonein krystallisiertes weibliches Sexualhormon. *Die Naturwissenschaften* **1929**, *17*, 879. [[CrossRef](#)]
24. Butenandt, A. Über physikalische und chemische Eigenschaften des krystallisierten Follikelhormons. Untersuchungen über das weibliche Sexualhormon. *Hoppe-Seyler's Zeit. Physiol. Chem.* **1930**, *191*, 140–156. [[CrossRef](#)]
25. Butenandt, A. Über die chemische Untersuchung der Sexualhormone. *Zeit Angew. Chem.* **1931**, *44*, 905–908. [[CrossRef](#)]
26. Butenandt, A.; Jacobi, H. Über die Darstellung eines krystallisierten pflanzlichen Tokokinins (Thelykinins) und seine Identifizierung mit dem α -Follikelhormon. Untersuchungen über das weibliche Sexualhormon. *Hoppe Seyler's Z. Physiol. Chem.* **1933**, *218*, 104–112. [[CrossRef](#)]
27. Ramirez, M.P.; Haas, S. Hormone replacement therapy for women: The benefits, risks, and considerations for use in 2003. *Curr. Opinion Endocrin. Diabet.* **2003**, *10*, 400–418. [[CrossRef](#)]
28. Fluhmann, C.F. Estrogenic hormones: Their clinical usage. *Cal. West Med.* **1938**, *49*, 362–366.
29. Younglai, E.V.; Solomon, S. Formation of estra-1,3,5(10)-triene-3,15a,16a,17b-tetrol (estetrol) and estra-1,3,5(10)-triene-3,15a,17btriol from neutral precursors. *J. Clin. Endocrinol. Metab.* **1968**, *28*, 1611–1617. [[CrossRef](#)]
30. Raeside, J.I. A brief account of the discovery of the fetal/placental unit for estrogen production in equine and human pregnancies: Relation to human medicine. *Yale J. Biol. Med.* **2017**, *90*, 449–461.
31. Trifunović, J.; Borčić, V.; Vukmirović, S.; Mikov, M. Structural insights into anticancer activity of D-ring modified estrone derivatives using their lipophilicity in estimation of SAR and molecular docking studies. *Drug Test Anal.* **2017**, *9*, 1650. [[CrossRef](#)]
32. Dohrn, M.; Faure, W.; Poll, H.; Blotevogel, W. Tokokinine, Stoff mit sexualhormonartiger Wirkung aus Pflanzenzellen. *Med. Klin.* **1926**, *22*, 1417–1419.
33. Skarzynski, B. An oestrogenic substance from plant material. *Nature* **1933**, *131*, 766.
34. Janeczko, A.; Skoczowski, A. Mammalian sex hormones in plants. *Folia Histochem. Cytobiol.* **2005**, *43*, 71–79.

35. Jailer, J.W. The metabolism of the estrogens: A review. *J. Clin. Endocrinol.* **1949**, *9*, 557–572. [[CrossRef](#)]
36. Xu, Y.; López, M. Central regulation of energy metabolism by estrogens. *Mol. Metabol.* **2018**, *15*, 104–115. [[CrossRef](#)]
37. Ventura-Clapier, R.; Piquereau, J.; Veksler, V.; Garnier, A. Estrogens, estrogen receptors effects on cardiac and skeletal muscle mitochondria. *Front. Endocrinol.* **2019**, *10*, 557. [[CrossRef](#)]
38. Le, J.; Thomas, N.; Gurvich, C. Cognition, the menstrual cycle, and premenstrual disorders: A Review. *Brain Sci.* **2020**, *10*, 198. [[CrossRef](#)]
39. Su, Z.; Yuan, W.; Wang, P.; Li, S. Ethnobotany, phytochemistry, and biological activities of *Taxodium Rich.* *Pharm. Crops* **2013**, *4*, 1–14.
40. Sukandar, E.Y.; Suganda, A.G.; Pertiwi, G.U. Aktivitas sediaan yang mengandung ekstrak daun ketapang pada kulit kelinci yang diinfeksi dengan ephidermophyton floccosum dan *Candida albicans*. *Acta Pharm. Ind.* **2007**, *32*, 45–49.
41. Suganda, A.G.; Sukandar, E.Y.; Hardhiko, R.S. Aktivitas antimikroba ekstrak etanol daun yang dipetik dan ekstrak air daun gugur pohon ketapang (*Terminalia catappa* L.). *Acta Pharm. Ind.* **2004**, *29*, 129–133.
42. Suganda, A.G.; Sukandar, E.Y.; Ratna, L. Aktivitas antimikroba ekstrak etanol daun dua belas jenis *Tumbuhan marga terminalia* (Combretaceae). *Acta Pharm. Ind.* **2006**, *31*, 18–23.
43. Pranjali, C.; Lokesh, R. A review on medicinal potential of *Terminalia catappa*. *Int. J. Green Pharm.* **2020**, *14*, 229–234.
44. Misico, R.I.; Veleiro, A.S.; Burton, G.; Oberti, J.C. Withanolides from *Jaborosa leucotricha*. *Phytochemistry* **1997**, *45*, 1045–1048. [[CrossRef](#)]
45. Yan, X.-H.; Liu, H.-L.; Huang, H.; Li, X.-B.; Guo, Y.-W. Steroids with aromatic A rings from the Hainan soft coral *Dendronephthya stuederi* Ridley. *J. Nat. Prod.* **2011**, *74*, 175–180. [[CrossRef](#)]
46. Lu, Z.; Van Wagoner, R.M.; Harper, M.K.; Hooper, J.N.A.; Ireland, C.M. Two ring-A aromatized bile acids from the marine sponge *Sollasella moretonensis*. *Nat. Prod. Commun.* **2010**, *5*, 1571–1574. [[CrossRef](#)]
47. Yeung, B.K.S.; Hamann, M.T.; Scheuer, P.J.; Kelly-Borges, M. Hapaioside: A 19-norpregnane glycoside from the sponge *Cribrorchalina olemda*. *Tetrahedron* **1994**, *50*, 12593–12598. [[CrossRef](#)]
48. Nakao, Y.; Kuo, J.; Yoshida, W.Y.; Kelly, M.; Scheuer, P.J. More kapakahines from the marine sponge *Cribrorchalina olemda*. *Org. Lett.* **2003**, *5*, 1387–1390. [[CrossRef](#)]
49. Tomono, Y.; Hirota, H.; Imahara, Y.; Fusetani, N. Four new steroids from two octocorals. *J. Nat. Prod.* **1999**, *62*, 1538–1541. [[CrossRef](#)] [[PubMed](#)]
50. Barrero, A.F.; Oltra, J.E.; Poyatos, J.A.; Jiménez, D.; Oliver, E. Phycomysterols and other sterols from the fungus *Phycomyces blakesleeanus*. *J. Nat. Prod.* **1998**, *61*, 1491–1496. [[CrossRef](#)]
51. Liu, X.H.; Tang, X.Z.; Miao, F.P.; Ji, N.Y. A new pyrrolidine derivative and steroids from an algicolous *Gibberella zeae* strain. *Nat. Prod. Commun.* **2011**, *6*, 1243–1246. [[CrossRef](#)] [[PubMed](#)]
52. Luo, X.; Li, F.; Shinde, P.B.; Hong, J.; Lee, C.-O.; Im, K.S.; Jung, J.H. 26,27-Cyclosterols and other polyoxygenated sterols from marine sponge *Topsentia* sp. *J. Nat. Prod.* **2006**, *69*, 1760–1768. [[CrossRef](#)] [[PubMed](#)]
53. Brown, A.C.; Fraser, T.R. The connection of chemical constitution and physiological action. *Trans. Roy. Soc. Edinb.* **1868**, *25*, 224–242.
54. Cros, A.F.A. Action de L'alcohol Amylique Sur L'organisme. Ph.D. Thesis, University of Strasbourg, Strasbourg, France, 1863.
55. Richet, M.C. Note sur le rapport entre la toxicité et les propriétés physiques des corps. *Compt. Rend. Soc. Biol.* **1893**, *45*, 775–776.
56. Meyer, H. Zur Theorie der Alkoholnarkose. *Arch. Exp. Path. Pharm.* **1899**, *42*, 109–118. [[CrossRef](#)]
57. Overton, C.E. *Studien über Die Narkose*; Fischer: Jena, Germany, 1901.
58. Hammett, L.P. Some relations between reaction rates and equilibrium constants. *Chem. Rev.* **1935**, *17*, 125–136. [[CrossRef](#)]
59. Hammett, L.P. The effect of structure upon the reactions of organic compounds. Benzene derivatives. *J. Am. Chem. Soc.* **1937**, *59*, 96–103. [[CrossRef](#)]
60. Taft, R.W. Separation of polar, steric and resonance effects in reactivity. In *Steric Effects in Organic Chemistry*; Newman, M.S., Ed.; Wiley: Hoboken, NJ, USA, 1956; pp. 556–675.
61. Hansch, C.; Fujita, T. π - τ - π Analysis. A method for the correlation of biological activity and chemical structure. *J. Am. Chem. Soc.* **1964**, *86*, 1616–1626. [[CrossRef](#)]
62. Hansch, C.; Leo, A. *Exploring QSAR*; American Chemical Society: Washington, DC, USA, 1995.
63. Sliwoski, G.; Kothiwale, S.; Meiler, J.; Lowe, E.W., Jr. Computational methods in drug discovery. *Pharm. Rev.* **2014**, *66*, 334–395. [[CrossRef](#)]
64. Leelananda, S.P.; Lindert, S. Computational methods in drug discovery. *Beilstein J. Org. Chem.* **2016**, *12*, 2694–2718. [[CrossRef](#)]
65. Kokh, D.B.; Amaral, M.; Bomke, J.; Grädler, U.; Musil, D. Estimation of drug-target residence times by τ -random acceleration molecular dynamics simulations. *J. Chem. Theor. Comput.* **2018**, *14*, 3859–3869. [[CrossRef](#)]
66. Cherkasov, A.M.; Muratov, E.N.; Fourches, D.; Varnek, A.; Baskin, I.I. QSAR modeling: Where have you been? Where are you going to? *J. Med. Chem.* **2014**, *57*, 4977–5010. [[CrossRef](#)]
67. Burov, Y.V.; Poroikov, V.V.; Korolchenko, L.V. National system for registration and biological testing of chemical compounds: Facilities for new drugs search. *Bull. Natl. Cent. Biol. Act. Comp.* **1990**, *1*, 4–25.
68. Muratov, E.N.; Bajorath, J.; Sheridan, R.P.; Tetko, I.; Filimonov, D.; Poroikov, V.; Oprea, T. QSAR without borders. *Chem. Soc. Rev.* **2020**, *49*, 3525–3564. [[CrossRef](#)]

69. Poroikov, V.V.; Filimonov, D.A.; Glorizova, T.A.; Lagunin, A.A.; Druzhilovskiy, D.S.; Rudik, A.V. Computer-aided prediction of biological activity spectra for organic compounds: The possibilities and limitations. *Russ. Chem. Bull.* **2019**, *68*, 2143–2154. [CrossRef]
70. Filimonov, D.A.; Druzhilovskiy, D.S.; Lagunin, A.A.; Glorizova, T.A.; Rudik, A.V.; Dmitriev, P.V.; Poroikov, V.V. Computer-aided prediction of biological activity spectra for chemical compounds: Opportunities and limitations. *Biom. Chem. Res. Method.* **2018**, *1*, e00004. [CrossRef]
71. Anusevicius, K.; Mickevicius, V.; Stasevych, M.; Zvarych, V.; Komarowska-Porokhnyavets, O.; Novikov, V.; Tarasova, O.; Glorizova, T.; Poroikov, V. Design, synthesis, in vitro antimicrobial activity evaluation and computational studies of new *N*-(4-iodophenyl)-Alanine derivatives. *Res. Chem. Intermed.* **2015**, *41*, 7517–7540. [CrossRef]
72. Murtazalieva, K.A.; Druzhilovskiy, D.S.; Goel, R.K.; Sastry, G.N.; Poroikov, V.V. How good are publicly available web services that predict bioactivity profiles for drug repurposing? *SAR QSAR Environ. Res.* **2017**, *28*, 843–862. [CrossRef]
73. PASS Online. Available online: <http://www.way2drug.com/passonline/> (accessed on 4 June 2021).
74. Lagunin, A.A.; Goel, R.K.; Gawande, D.Y.; Priynka, P.; Glorizova, T.A.; Dmitriev, A.V.; Ivanov, S.M.; Rudik, A.V.; Konova, V.I.; Pogodin, P.V. Chemo- and bioinformatics resources for in silico drug discovery from medicinal plants beyond their traditional use: A critical review. *Nat. Prod. Rep.* **2014**, *31*, 1585–1611. [CrossRef]
75. Goel, R.K.; Poroikov, V.; Gawande, D.; Lagunin, A.; Randhawa, P.; Mishra, A. Revealing medicinal plants useful for comprehensive management of epilepsies and associated co-morbidities through in silico mining of their phytochemical diversity. *Planta Med.* **2015**, *81*, 495–506.
76. Dembitsky, V.M.; Glorizova, T.A.; Poroikov, V.V. Naturally occurring plant isoquinoline *N*-oxide alkaloids: Their pharmacological and SAR activities. *Phytomedicine* **2015**, *22*, 183–202. [CrossRef] [PubMed]
77. Gawande, D.Y.; Druzhilovskiy, D.; Gupta, R.C.; Poroikov, V.; Goel, R.K. Anticonvulsant activity and acute neurotoxic profile of *Achyranthes aspera* Linn. *J. Ethnopharmacol.* **2017**, *202*, 97–102. [CrossRef] [PubMed]
78. Dembitsky, V.M.; Dzhemileva, L.; Glorizova, T.; D'yakonov, V. Natural and synthetic drugs used for the treatment of the dementia. *Biochem. Biophys. Res. Commun.* **2020**, *524*, 772–783. [CrossRef] [PubMed]
79. Lagunin, A.; Povydysh, M.; Ivkin, D.; Luzhanin, V.; Krasnova, M. Antihypoxic action of *Panax japonicus*, *Tribulus terrestris* and *Dioscorea deltoidea* cell cultures: In silico and animal studies. *Mol. Inform.* **2020**, *39*, 2000093. [CrossRef]
80. Dembitsky, V.M.; Ermolenko, E.; Savidov, N.; Glorizova, T.A.; Poroikov, V.V. Antiprotozoal and antitumor activity of natural polycyclic endoperoxides: Origin, structures, and biological activity. *Molecules* **2021**, *19*, 686. [CrossRef]
81. Qureshi, R.; Picon-Ruiz, M.; Aurrekoetxea-Rodriguez, I.; Kesmodel, S.; del Mar Vivanco, M.; Slinger, J.M. The major pre- and postmenopausal estrogens play opposing roles in obesity-driven mammary inflammation and breast cancer development. *Cell Metab.* **2020**, *31*, 1154–1172. [CrossRef]
82. Ahmed, S.; Owen, C.P.; James, K.; Sampson, L.; Patel, C.K. Review of estrone sulfatase and its inhibitors—An important new target against hormone dependent breast cancer. *Curr. Med. Chem.* **2002**, *9*, 263–273. [CrossRef]
83. Neilson, H.K.; Friedenreich, C.M.; Brockton, N.T.; Millikan, R.C. Physical activity and postmenopausal breast cancer: Proposed biologic mechanisms and areas for future research. *Cancer Epidemiol. Biomark. Prev.* **2009**, *18*, 11–27. [CrossRef]
84. Bassol, S.; Garza-Flores, J. Review of ovulation return upon discontinuation of once-a-month injectable contraceptives. *Contraception* **1994**, *49*, 441–453. [CrossRef]
85. McCarthy, M.M. Estradiol, and the developing brain. *Physiol. Rev.* **2008**, *88*, 91–134. [CrossRef]
86. Soltysik, K.; Czekaj, P. Membrane estrogen receptors—Is it an alternative way of estrogen action? *J. Physiol. Pharmacol.* **2013**, *64*, 129–142.
87. Mauvais-Jarvis, F.; Klein, S.L.; Levin, E.R. Estradiol, progesterone, immunomodulation, and COVID-19 outcomes. *Endocrinology* **2020**, *161*, 127. [CrossRef]
88. Lappano, R.; Rosano, C.; De Marco, P.; De Francesco, E.M.; Pezzi, V.; Maggiolini, M. Estriol acts as a GPR30 antagonist in estrogen receptor-negative breast cancer cells. *Mol. Cell. Endocrinol.* **2010**, *320*, 162–170. [CrossRef]
89. Ali, E.S.; Mangold, C.; Peiris, A.N. Estriol: Emerging clinical benefits. *Menopause* **2017**, *24*, 1081–1085. [CrossRef]
90. Li, M.; Scott, R.Y. A review on structural elucidation of metabolites of environmental steroid hormones via liquid chromatography–mass spectrometry. *Trends Anal. Chem.* **2018**, *109*, 142–153.
91. Brandán, S.A. Structural and vibrational studies of equilenin, equilin and estrone steroids. *Biointerface Res. Appl. Chem.* **2019**, *9*, 4502–4516.
92. Bhavnani, B.R.; Stanczyk, F.Z. Pharmacology of conjugated equine estrogens: Efficacy, safety and mechanism of action. *J. Steroid Biochem. Mol. Biol.* **2014**, *142*, 16–29. [CrossRef]
93. Ruder, H.J.; Loriaux, L.; Lipsett, M.B. Lipsett, Estrone sulfate: Production rate and metabolism in man. *J. Clin. Investig.* **1972**, *51*, 1020–1033. [CrossRef]
94. Banjare, L.; Jain, A.K.; Thareja, A. Dual aromatase-sulphatase inhibitors (DASIs) for the treatment of hormone dependent breast cancer. *Mini Rev. Med. Chem.* **2021**, *21*, 10. [CrossRef]
95. Blackwell, L.F.; Cooke, D.G.; Brown, S. The use of estrone-3-glucuronide and pregnanediol-3-glucuronide excretion rates to navigate the continuum of ovarian activity. *Front. Public Health* **2018**, *6*, 153. [CrossRef]
96. Katayama, S.; Fishman, J. 2-Hydroxyestrone suppresses and 2-methoxyestrone augments the preovulatory prolactin surge in the cycling rat. *Endocrinology* **1982**, *110*, 1448–1450. [CrossRef]

97. Gupta, M.; McDougal, A.; Safe, S. Estrogenic and antiestrogenic activities of 16 α - and 2-hydroxy metabolites of 17 β -estradiol in MCF-7 and T47D human breast cancer cells. *J. Steroid Biochem. Mol. Biol.* **1998**, *67*, 413–419. [[CrossRef](#)]
98. Bradlow, H.L.; Telang, N.T.; Sepkovic, D.W.; Osborne, M.P. 2-Hydroxyestrone: The good estrogen. *J. Endocrinol.* **1996**, *150*, S259–S265.
99. Williams, J.G.; Longcope, C.; Williams, K.I.H. 4-Hydroxyestrone: A new metabolite of estradiol-17 β from humans. *Steroids* **1974**, *24*, 687–701. [[CrossRef](#)]
100. Emons, G.; Hoppen, H.O.; Ball, P.; Knuppen, R. 4-Hydroxyestrone, isolation and identification in human urine. *Steroids* **1980**, *36*, 73–79. [[CrossRef](#)]
101. Choi, H.J.; Lee, A.J.; Kang, K.S. 4-Hydroxyestrone, an endogenous estrogen metabolite. Can strongly protect neuronal cells against oxidative damage. *Sci. Rep.* **2020**, *10*, 7283. [[CrossRef](#)]
102. Mueck, A.O.; Seeger, H.; Lippert, T.H. Estradiol metabolism and malignant disease. *Maturitas* **2002**, *43*, 1–10. [[CrossRef](#)]
103. Anh, N.H.; Long, N.P.; Kim, S.J.; Min, J.E. Steroidomics for the prevention, assessment, and management of cancers: A systematic review and functional analysis. *Metabolites* **2019**, *9*, 199. [[CrossRef](#)]
104. Seeger, H.; Deuringer, F.-U.; Wallwiener, D.; Mueck, A.O. Breast cancer risk during HRT: Influence of estradiol metabolites on breast cancer and endothelial cell proliferation. *Maturitas* **2004**, *49*, 235–240. [[CrossRef](#)]
105. Fuhrman, B.J.; Schairer, C.; Gail, M.H.; Boyd-Morin, J. Estrogen metabolism and risk of breast cancer in postmenopausal women. *J. Nat. Cancer Inst.* **2012**, *104*, 326–339. [[CrossRef](#)]
106. Fishman, J.; Cox, R.I.; Gallagher, T.F. 2-Hydroxyestrone: A new metabolite of estradiol in man. *Archiv. Biochem. Biophys.* **1960**, *90*, 318–319. [[CrossRef](#)]
107. Taylor, H.C. The present status of gynecologic endocrine therapy. *Bull. N. Y. Acad. Med.* **1938**, *14*, 608–634. [[PubMed](#)]
108. Marrian, G.F. The conjugated estrogens. *Cold Spring Harb. Symp. Quant. Biol.* **1937**, *5*, 16–24. [[CrossRef](#)]
109. Dembitsky, V.M.; Savidov, N.; Poroikov, V.V.; Glorizova, T.A.; Imbs, A.B. Naturally occurring aromatic steroids and their biological activities. *Appl. Microbiol. Biotechnol.* **2018**, *102*, 4663–4674. [[CrossRef](#)]
110. De Riccardis, F.; Minale, L.; Riccio, R.; Giovannitti, B.; Iorizzi, M.; Debitus, C. Phosphated and sulfated marine polyhydroxylated steroids from the starfish *Tremaster novaecaledoniae*. *Gazz. Chim. Ital.* **1993**, *123*, 79–86.
111. Dembitsky, V.M.; Glorizova, T.A.; Savidov, N. Steroid phosphate esters and phosphonosteroids and their biological activities. *Appl. Microbiol. Biotechnol.* **2018**, *102*, 7679–7692. [[CrossRef](#)]
112. Delrio, G.; Botte, V. Testosterone 17-phosphate and 19-nortestosterone 17-phosphate as substrate for rabbit prostate phosphatases. *Biochim. Biophys. Acta* **1970**, *218*, 327–332. [[CrossRef](#)]
113. Kokado, A.; Tsuji, A.; Maeda, M. Chemiluminescence assay of alkaline phosphatase using cortisol-21-phosphate as substrate and its application to enzyme immunoassays. *Anal. Chim. Acta* **1997**, *337*, 335–340. [[CrossRef](#)]
114. Ellam, T.J.; Chico, T.J. Phosphate: The new cholesterol? The role of the phosphate axis in non-uremic vascular disease. *Atherosclerosis* **2012**, *220*, 310–318. [[CrossRef](#)]
115. Davis, S.C.; Szoka, F.C., Jr. Cholesterol phosphate derivatives: Synthesis and incorporation into a phosphatase and calcium-sensitive triggered release liposome. *Bioconjug. Chem.* **1998**, *9*, 783–792. [[CrossRef](#)]
116. Sachs-Barrable, K.; Darlington, J.W.; Wasan, K.M. The effect of two novel cholesterol-lowering agents, disodium ascorbyl phytostanol phosphate and nanostructured aluminosilicate on the expression and activity of P-glycoprotein within Caco-2 cells. *Lipids Health Dis.* **2014**, *13*, 153–163. [[CrossRef](#)]
117. Kutney, J.P.; Pritchard, H.P.; Lukic, T. Novel Compounds and Compositions Comprising Sterols and/or Stanols and Cholesterol Biosynthesis Inhibitors and Use Thereof in Treating or Preventing a Variety of Diseases and Conditions. Europe Patent EP1644399A2, 7 September 2003.
118. Gunnarsson, P.O.; Norlén, B.J. Clinical pharmacology of polyestradiol phosphate. *Prostate* **1988**, *13*, 299–304. [[CrossRef](#)]
119. Vil, V.; Glorizova, T.A.; Terentev, A.O.; Zhukova, N.V.; Dembitsky, V.M. Highly oxygenated isoprenoid lipids derived from terrestrial and aquatic sources: Origin, structures, and biological activities. *Vietnam J. Chem.* **2019**, *57*, 1–15. [[CrossRef](#)]
120. Vil, V.A.; Terentev, A.O.; Savidov, N.; Glorizova, T.A.; Poroikov, V.V.; Pounina, T.A.; Dembitsky, V.M. Hydroperoxy steroids and triterpenoids derived from plant and fungi: Origin, structures and biological activities. *J. Steroid Biochem. Mol. Biol.* **2019**, *190*, 76–87. [[CrossRef](#)]
121. Savidov, N.; Glorizova, T.A.; Poroikov, V.V.; Dembitsky, V.M. Highly oxygenated isoprenoid lipids derived from fungi and fungal endophytes: Origin and biological activities. *Steroids* **2018**, *140*, 114–124. [[CrossRef](#)]
122. Vil, V.A.; Glorizova, T.A.; Terentev, A.O.; Savidov, N.; Dembitsky, V.M. Hydroperoxides derived from marine sources: Origin and biological activities. *Appl. Microbiol. Biotechnol.* **2019**, *103*, 1627–1642. [[CrossRef](#)]
123. Dembitsky, V.M.; Glorizova, T.A.; Poroikov, V.V. Naturally occurring marine α,β -epoxy steroids: Origin and biological activities. *Vietnam J. Chem.* **2018**, *56*, 409–433. [[CrossRef](#)]
124. Rashidi, B.; Hoseini, Z.; Sahebkar, A.; Mirzaei, H. Anti-atherosclerotic effects of vitamins D and E in suppression of atherogenesis. *J. Cell. Physiol.* **2017**, *232*, 2968–2976. [[CrossRef](#)]
125. Kutner, A.; Brown, G. Vitamins D: Relationship between structure and biological activity. *Int. J. Mol. Sci.* **2018**, *19*, 2119. [[CrossRef](#)]
126. Dusso, A.S.; Brown, A.J.; Slatopolsky, E. Vitamin D. *Am. J. Physiol. Renal. Physiol.* **2005**, *289*, F8–F28. [[CrossRef](#)]
127. Perez-Lopez, F.R. Vitamin D: The secosteroid hormone and human reproduction. *Gynecolog. Endocrinol.* **2007**, *23*, 13–24. [[CrossRef](#)] [[PubMed](#)]

128. Ermolenko, E.V.; Imbs, A.B.; Glorizova, T.A.; Poroikov, V.V.; Sikorskaya, T.V.; Dembitsky, V.M. Chemical diversity of soft coral steroids and their pharmacological activities. *Mar. Drugs* **2020**, *20*, 613. [[CrossRef](#)] [[PubMed](#)]
129. Grishko, V.V.; Galaiko, N.V. Structural diversity, natural sources and pharmacological potential of naturally occurring A-seco-triterpenoids studies. *Nat. Prod. Chem.* **2016**, *51*, 51–149.
130. Sica, D.; Musumeci, D. Secosteroids of marine origin. *Steroids* **2004**, *69*, 743–756. [[CrossRef](#)] [[PubMed](#)]
131. Wu, S.H.; Luo, X.D.; Ma, Y.B.; Liu, Y.K.; Wu, D.G.; Zhao, B.; Lu, Y.; Zheng, Q.T. Two novel secoergosterols from the fungus *Tylophilus plumbeoviolaceus*. *J. Nat. Prod.* **2000**, *63*, 534–536. [[CrossRef](#)] [[PubMed](#)]
132. Zhao, Z.Z.; Chen, H.P.; Huang, Y.; Zhang, S.B.; Li, Z.H.; Feng, T.; Liu, J.K. Bioactive polyketides and 8,14-seco-ergosterol from fruiting bodies of the ascomycete *Daldinia childiae*. *Phytochemistry* **2017**, *142*, 68–75. [[CrossRef](#)] [[PubMed](#)]
133. Cui, C.M.; Li, X.M.; Meng, L.; Li, C.S.; Huang, C.G.; Wang, B.G. 7-Nor-ergosterolide, a pentalactone-containing norsteroid and related steroids from the marine-derived endophytic *Aspergillus ochraceus* EN-31. *J. Nat. Prod.* **2010**, *73*, 1780–1784. [[CrossRef](#)]
134. Onodera, H.; Ichimura, M.; Baba, K.; Agatsuma, T.; Sasho, S.; Suzuki, M.; Iwamoto, S.; Kakita, S. PCT Int. Appl. WO 2009096445, Nerve Trunk Cell Propagation Accelerator, 06.08.2009; Kyowa Hakko Kirin Co., Ltd.: Tokyo, Japan.
135. Kazlauskas, R.; Murphy, P.T.; Ravi, B.N.; Sanders, R.L.; Wells, R.J. Spermidine derivatives and 9,11-secosteroids from a soft coral (*Sinularia* sp.). *Austral. J. Chem.* **1982**, *35*, 69–75. [[CrossRef](#)]
136. Bonini, C.; Cooper, C.B.; Kazlauskas, R.; Wells, R.J.; Djerassi, C. Minor and trace sterols in marine invertebrates. 41. Structure and stereochemistry of naturally occurring 9,11-seco sterols. *J. Org. Chem.* **1983**, *48*, 2108–2111. [[CrossRef](#)]
137. Weng, J.R.; Chiu, C.F.; Sheu, J.H. A sterol from soft coral induces apoptosis and autophagy in MCF-7 breast cancer cells. *Mar. Drugs* **2018**, *16*, 238. [[CrossRef](#)]
138. Chang, Y.C.; Lai, K.H.; Kumar, S.; Chen, P.J.; Wu, Y.H.; Lai, C.L. 1H-NMR-Based Isolation of Anti-Inflammatory 9,11-secosteroids from the octocoral *Sinularia leptoclados*. *Mar. Drugs* **2020**, *18*, 271. [[CrossRef](#)]
139. Hirsch, A.L. Industrial aspects of vitamin D. In *Vitamin, D.*, 3rd ed.; Feldman, D., Pike, J.W., Adams, J.S., Eds.; Academic Press: San Diego, CA, USA, 2011; pp. 73–93.
140. Teichmann, A.; Dutta, P.C.; Staffas, A.; Jagerst ad, M. Sterol and vitamin D2 concentrations in cultivated and wild grown mushrooms: Effect of UV radiation. *LWT Food Sci. Technol.* **2007**, *40*, 815–822. [[CrossRef](#)]
141. Koyyalamudi, S.R.; Jeong, S.C.; Song, C.H.; Cho, K.Y.; Pang, G. Vitamin D2 formation and bioavailability from *Agaricus bisporus* button mushrooms treated with ultraviolet irradiation. *J. Agric. Food Chem.* **2009**, *57*, 3351–3355. [[CrossRef](#)]
142. Koyyalamudi, S.R.; Jeong, S.C.; Pang, G.; Teal, A.; Biggs, T. Concentration of vitamin D2 in white button mushrooms (*Agaricus bisporus*) exposed to pulsed UV light. *J. Food Comp. Anal.* **2011**, *24*, 976–979. [[CrossRef](#)]
143. Phillips, K.M.; Ruggio, D.M.; Horst, R.L.; Minor, B.; Simon, R.R.; Feeney, M.J.; Byrdwell, W.C.; Haytowitz, D.B. Vitamin D and sterol composition of 10 types of mushrooms from retail suppliers in the United States. *J. Agric. Food Chem.* **2011**, *59*, 7841–7853. [[CrossRef](#)]
144. Jasinghe, V.J.; Perera, C.O. Ultraviolet irradiation: The generator of vitamin D2 in edible mushrooms. *Food Chem.* **2006**, *95*, 638–643. [[CrossRef](#)]
145. Roberts, J.S.; Teichert, A.; McHugh, T.H. Vitamin D2 formation from post-harvest UV-B treatment of mushrooms (*Agaricus bisporus*) and retention during storage. *J. Agric. Food Chem.* **2008**, *56*, 4541–4544. [[CrossRef](#)]
146. Shen, Y.C.; Cheng, Y.B.; Kobayashi, J.; Kubota, T.; Takahashi, Y.; Mikami, Y.; Ito, J.; Lin, Y.S. Nitrogen-containing verticillene diterpenoids from the Taiwanese soft coral *Cespitularia taeniata*. *J. Nat. Prod.* **2007**, *70*, 1961–1965. [[CrossRef](#)]
147. Lakshmi, V.; Kumar, R. Metabolites from *Sinularia* species. *Nat. Prod. Res.* **2009**, *23*, 801–850. [[CrossRef](#)]
148. Kulda, V. Vitamin D metabolism. *Vnitř Lek.* **2012**, *58*, 400–404.
149. Windaus, A.; Schenck, F.R.; Werder, F.V. The anti-rachitically active irradiation product from 7-dehydro-cholesterol. *Hoppe-Seyler's Zeitsch. Physiol. Chem.* **1936**, *241*, 100–103. [[CrossRef](#)]
150. Wanga, T.; Bengtsson, G.; K arnefeldt, I.; Bj orn, L.O. Provitamins and vitamins D2 and D3 in *Cladina* spp. over a latitudinal gradient: Possible correlation with UV levels. *J. Photochem. Photobiol.* **2001**, *62B*, 118–122. [[CrossRef](#)]
151. Horst, R.L.; Reinhardt, T.A.; Russell, J.R.; Napoli, J.L. The isolation and identification of vitamin D2 and vitamin D3 from *Medicago sativa* (Alfalfa plant). *Arch. Biochem. Biophys.* **1984**, *231*, 67–71. [[CrossRef](#)]
152. World Health Organization. *World Health Organization Model List of Essential Medicines: 21st List 2019*; World Health Organization: Geneva, Switzerland, 2019.
153. Trump, D.L.; Aragon-Ching, J.B. Vitamin D in prostate cancer. *Asian J. Androl.* **2018**, *20*, 244–252. [[CrossRef](#)]
154. Laaksi, I. Vitamins, infectious and chronic disease during adulthood and aging Vitamin D and respiratory infection in adults. *Proceed. Nutr. Soc.* **2012**, *71*, 90–97. [[CrossRef](#)]
155. Saikia, S.; Kolita, B.; Dutta, P.P.; Dutta, D.J.; Neipihoi, S.; Nath, S. Marine steroids as potential anticancer drug candidates: In silico investigation in search of inhibitors of Bcl-2 and CDK-4/Cyclin D1. *Steroids* **2015**, *102*, 7–16. [[CrossRef](#)]
156. Dembitsky, V.M. Anticancer activity of natural and synthetic acetylenic lipids. *Lipids* **2006**, *41*, 883–924. [[CrossRef](#)]
157. Zhang, H.; Zhao, Z.; Wang, H. Cytotoxic natural products from marine sponge-derived microorganisms. *Mar. Drugs* **2017**, *15*, 68. [[CrossRef](#)] [[PubMed](#)]
158. Mioso, R.; Marante, F.J.T.; Bezerra, R.D.S.; Borges, F.V.P.; Santos, B.V.; Laguna, I.H.B.D. Cytotoxic compounds derived from marine sponges, A review (2010–2012). *Molecules* **2017**, *22*, 208. [[CrossRef](#)] [[PubMed](#)]

159. Dembitsky, V.M.; Rezanka, T.; Srebnik, M. Lipid compounds of freshwater sponges: Family Spongillidae, class Demospongiae. *Chem. Phys. Lipids* **2003**, *123*, 117–155. [[CrossRef](#)]
160. Xu, S.; Liao, X.; Du, B.; Zhou, X.; Huang, Q.; Wu, C. A series of new 5,6-epoxysterols from a Chinese sponge *Ircinia aruensis*. *Steroids* **2008**, *73*, 568–573.
161. Li, T.; Wang, N.; Zhang, T.; Zhang, B.; Sajeevan, T.P.; Valsamma, J.; Armstrong, L.; He, S.; Yan, X.; Naman, C.B. A systematic review of recently reported marine derived natural product kinase inhibitors. *Mar. Drugs* **2019**, *17*, 493. [[CrossRef](#)]
162. Zhang, H.J.; Yi, Y.H.; Yang, F.; Chen, W.S.; Lin, H.W. Sesterterpenes and a new sterol from the marine sponge *Phyllospongia foliascens*. *Molecules* **2010**, *15*, 834–841.
163. Afiyatullo, S.S.; Kalinovsky, A.I.; Antonov, A.S.; Ponomarenko, L.P. Isolation and structures of erylosides from the Caribbean sponge *Erylus goffrilleri*. *J. Nat. Prod.* **2007**, *70*, 1871–1877. [[CrossRef](#)]
164. Anjaneyulu, A.S.R.; Krishna Murthy, M.V.R.; Gowri, P.M. Novel epoxy steroids from the Indian ocean soft coral *Sarcophyton crassocaule*. *J. Nat. Prod.* **2000**, *63*, 112–118.
165. Funel, C.; Berru , F.; Roussakis, C.; Fernandez Rodriguez, R.; Amade, P. New cytotoxic steroids from the Indian ocean sponge *Axinella* cf. *bidderi*. *J. Nat. Prod.* **2004**, *67*, 491–494.
166. Dembitsky, V.M. Bioactive fungal endoperoxides. *Med. Mycol.* **2015**, *1*, 1–7.
167. Dembitsky, V.M. Astonishing diversity of natural peroxides as potential therapeutic agents. *J. Mol. Genet. Med.* **2015**, *9*, 1000163.
168. Dembitsky, V.M. Bioactive peroxides as potential therapeutic agents. *Eur. J. Med. Chem.* **2008**, *43*, 223–251.
169. Kyasa, S.K. New Methods for Synthesis of Organic Peroxides and Application of Peroxide Electrophiles to Synthesis of Functionalized Ethers. Ph.D. Thesis, University of Nebraska-Lincoln, Lincoln, NE, USA, 2005.
170. Klussmann, M. Alkenyl and aryl peroxides. *Chemistry* **2018**, *24*, 4480–4496.
171. Dembitsky, V.M.; Gloriovova, T.A.; Poroikov, V.V. Natural peroxy anticancer agents. *Mini-Rev. Med. Chem.* **2007**, *7*, 571–589. [[CrossRef](#)] [[PubMed](#)]
172. Dembitsky, V.; Shkrob, I.; Hanus, L.O. Ascaridole and related peroxides from the genus *Chenopodium*. *Biomed. Pap. Med. Fac. Univ. Palacky Olomouc Czech Repub.* **2008**, *152*, 209–215. [[CrossRef](#)] [[PubMed](#)]
173. Liu, D.Z.; Liu, J.K. Peroxy natural products. *Nat. Prod. Bioprospect.* **2013**, *3*, 161–206. [[CrossRef](#)]
174. Thao, N.P.; Cuong, N.X.; Luyen, B.T.T.; Nam, N.H. Steroidal constituents from the starfish *Astropecten polyacanthus* and their anticancer effects. *Chem. Pharm. Bull.* **2013**, *61*, 1044–1051. [[CrossRef](#)]
175. Seo, Y.W.; Rho, J.R.; Cho, K.W.; Sim, C.J.; Shin, J.H. Isolation of epidioxysteroids from a sponge of the genus *Tethya*. *Bull. Korean Chem. Soc.* **1997**, *18*, 631–635.
176. Gunatilaka, A.A.L.; Gopichand, Y.; Schmitz, F.J.; Djerassi, C. Minor and trace sterols in marine invertebrates. 26. Isolation and structure elucidation of nine new 5,8-epidoxysterols from four marine organisms. *J. Org. Chem.* **1981**, *46*, 3860–3866. [[CrossRef](#)]
177. Gauvin, A.; Smadja, J.; Akin, M.; Faure, R.; Gaydou, E.M. Isolation of bioactive 5 α ,8 α -epidioxysterols from the marine sponge *Luffariella* cf. *variabilis*. *Can. J. Chem.* **2000**, *78*, 986–992. [[CrossRef](#)]
178. Sheikh, Y.M.; Djerassi, C. Steroids from sponges. *Tetrahedron* **1974**, *30*, 4095–4103. [[CrossRef](#)]
179. Zheng, W.; Liu, T.; Xiang, X.; Gu, Q. Sterol composition in field-grown and cultured mycelia of *Inonotus obliquus*. *Yaoxue Xuebao* **2007**, *42*, 750–756.
180. Zhang, Y.; Pei, L.; Gao, L.; Huang, Q.; Qi, J. A neurotogenic compound from *Tremella fuciformis*. *Zhongguo Zhong Yao Za Zhi* **2011**, *36*, 2358–2360.
181. Shi, X.W.; Li, X.J.; Gao, J.M.; Zhang, X.C. Fasciculols H and I, two lanostane derivatives from Chinese mushroom *Naematoloma fasciculare*. *Chem. Biodivers.* **2011**, *8*, 1864–1870. [[CrossRef](#)] [[PubMed](#)]
182. Yaoita, Y.; Amemiya, K.; Ohnuma, H.; Furumura, K.; Masaki, A.; Matasuki, T.; Kikuchi, M. Sterol constituents from five edible mushrooms. *Chem. Pharm. Bull.* **1998**, *46*, 944–950. [[CrossRef](#)]
183. Yue, J.M.; Chen, C.N.; Lin, Z.W.; Sun, H.D. Sterols from the fungus *Lactarium volenus*. *Phytochemistry* **2001**, *56*, 801–806. [[CrossRef](#)]
184. Zang, M.; Ying, J.Z. *Economic Fungi in the South West of China*; Scientific Press: Beijing, China, 1994.
185. Greca, M.D.; Fiorentino, A.; Molinaro, A.; Monaco, P.; Previtera, L. Hydroperoxysterols in *Arum italicum*. *Nat. Prod. Lett.* **1994**, *5*, 7–14. [[CrossRef](#)]
186. Wu, S.B.; Bao, Q.Y.; Wang, W.X.; Zhao, Y.; Xia, G. Cytotoxic triterpenoids and steroids from the bark of *Melia azedarach*. *Planta Med.* **2011**, *77*, 922–928. [[CrossRef](#)]
187. Ponce, M.A.; Ramirez, J.A.; Galagovsky, L.R.; Gros, E.G.; Erra-Balsells, R. A new look into the reaction between ergosterol and singlet oxygen in vitro. *Photochem. Photobiol. Sci.* **2002**, *1*, 749–756. [[CrossRef](#)]
188. Bocking, T.; Barrow, K.D.; Netting, A.G.; Chilcott, T.C.; Coster, H.G.L.; Hofer, M. Effects of singlet oxygen on membrane sterols in the yeast *Saccharomyces cerevisiae*. *Eur. J. Biochem.* **2000**, *267*, 1607–1618. [[CrossRef](#)]
189. Banskota, A.H.; Tezuka, Y.; Phung, L.K.; Tran, K.Q.; Saiki, I. Cytotoxic cycloartane-type triterpenes from *Combretum quadrangulare*. *Bioorg. Med. Chem. Lett.* **1998**, *8*, 3519–3524. [[CrossRef](#)]
190. Bankota, A.H.; Tezuka, Y.; Tran, K.Q.; Tanaka, K.; Saiki, I.; Kadota, S. Thirteen novel cycloartane-type triterpenes from *Combretum quadrangulare*. *J. Nat. Prod.* **2000**, *63*, 57–64. [[CrossRef](#)]
191. Roy, R.; Raj, K.; Singh, K.; Jash, S.K.; Sarkar, A.; Gorai, D. *Combretum quadrangulare* (Combretaceae): Phytochemical constituents and biological activity. *Indo Amer. J. Pharm. Res.* **2014**, *4*, 3416–3427.

192. Asai, T.; Hara, N.; Fujimoto, Y. Fatty acid derivatives and dammarane triterpenes from the glandular trichome exudates of *Ibicella lutea* and *Proboscidea louisiana*. *Phytochemistry* **2010**, *71*, 877–894. [[CrossRef](#)]
193. Lee, I.S.; Oh, S.R.; Ahn, K.S.; Lee, H.K. Semialactone, isofouquierone peroxide and fouquierone, three new dammarane triterpenes from *Rhus javanica*. *Chem. Pharm. Bull.* **2001**, *49*, 1024–1029. [[CrossRef](#)]
194. Abdel Bar, F.M.; Zaghoul, A.M.; Bachawal, S.V.; Sylvester, P.W.; Ahmad, K.F.; El Sayed, K.A. Antiproliferative triterpenes from *Melaleuca ericifolia*. *J. Nat. Prod.* **2008**, *71*, 1787–1790. [[CrossRef](#)]
195. Chiang, Y.M.; Kuo, Y.H. New peroxy triterpenes from the aerial roots of *Ficus macrocarpa*. *J. Nat. Prod.* **2001**, *64*, 436–439. [[CrossRef](#)]
196. Sikorsky, T.V.; Ermolenko, E.V.; Glorizova, T.A.; Dembitsky, V.M. Mini Review: Anticancer activity of diterpenoid peroxides. *Vietnam J. Chem.* **2020**, *58*, 273–280. [[CrossRef](#)]
197. Dembitsky, V.M.; Yaremenko, I.A. Stable and unstable 1,2-dioxolanes: Origin, synthesis, and biological activities. *Sci. Synth. Knowl. Updates* **2020**, *38*, 277–321. [[CrossRef](#)]
198. Dembitsky, V.M.; Vil, V.A. Medicinal chemistry of stable and unstable 1,2-dioxetanes: Origin, formation, and biological activities. *Sci. Synth. Knowl. Updates* **2020**, *38*, 333–381. [[CrossRef](#)]
199. Phillipson, J.D.; Wright, C.W. Antiprotozoal agents from plant sources. *Planta Med.* **1991**, *57*, S53–S59. [[CrossRef](#)] [[PubMed](#)]
200. Moss, G.P. The nomenclature of steroids. *Eur. J. Biochem.* **1989**, *186*, 429–458.
201. Burger, A. Cyclopropane compounds of biological interest. *Prog. Drug Res.* **1971**, *15*, 227–270.
202. Schoenheimer, R.; Evans, E.A., Jr. The chemistry of the steroids. *Ann. Rev. Biochem.* **1937**, *6*, 139–162. [[CrossRef](#)]
203. Dembitsky, V.M.; Glorizova, T.A. Astonishing diversity of carbon-bridged steroids and their biological activities: A brief review. *Eur. J. Biotechnol. Biosci.* **2018**, *6*, 6–23.
204. Jacobs, H.J.C. Photochemistry of conjugated trienes: Vitamin D revisited. *Pure Appl. Chem.* **1995**, *67*, 63–70. [[CrossRef](#)]
205. Kalaras, M.D. Production of Ergocalciferol (Vitamin D2) and Related Sterols in Mushrooms with Exposure to Pulsed Ultraviolet Light. Ph.D. Thesis, Pennsylvania State University, State College, PA, USA, January 2012.
206. Mitome, H.; Shirato, N.; Hoshino, A.; Miyaoka, H.; Yamada, Y.; Yamada, Y.; Van Soest, R.W.M. New polyhydroxylated sterols stylisterols A–C and a novel 5, 19-cyclosterol hatomasterol from the Okinawan marine sponge *Stylissa* sp. *Steroids* **2005**, *70*, 63–70. [[CrossRef](#)]
207. Calabro, K.; Kalahroodi, E.L.; Rodrigues, D.; Diaz, C.; Cruz, M.D.L.; Cautain, B.; Laville, R. Poecillastrosides, steroidal saponins from the Mediterranean deep-sea sponge *Poecillastra compressa* (Bowerbank, 1866). *Mar. Drugs* **2017**, *15*, 199. [[CrossRef](#)]
208. Giner, J.-L.; Gunasekera, S.P.; Pomponi, S.A. Sterols of the marine sponge *Petrosia weinbergi*: Implications for the absolute configurations of the antiviral orthoesterols and weinbersterols. *Steroids* **1999**, *64*, 820–824. [[CrossRef](#)]
209. HXinping, H.; Xiaobin, Z.; Liping, D.; Zhiwei, D.; Wenhan, L. Cycloartane triterpenes from marine green alga *Cladophora fascicularis*. *Chin. J. Ocean. Limnol.* **2006**, *24*, 443–448. [[CrossRef](#)]
210. Tung, N.H.; Van Minh, C.; Ha, T.T.; Van Kiem, P.; Huong, H.T. C29-Sterols with a cyclopropane ring at C-25 and 26 from the Vietnamese marine sponge *Ianthella* sp. and their anticancer properties. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 4584–4588. [[CrossRef](#)]
211. Gauvin, A.; Smadja, J.; Akin, M.; Gaydou, E.M. Cyclopropane-containing sterols in the marine sponge *Petrosia spheroida*. *Comp. Biochem. Physiol.* **1998**, *121B*, 451–456. [[CrossRef](#)]
212. Abdjul, D.B.; Yamazaki, H.; Takahashi, O.; Kirikoshi, R.; Ukai, K.; Namikoshi, M. Isopetrosynol, a new protein tyrosine phosphatase 1B inhibitor, from the marine sponge *Halichondria* cf. *panicea* collected at Iriomote Island. *Chem. Pharm. Bull.* **2016**, *64*, 733–736. [[CrossRef](#)]
213. Umeyama, A.; Ito, S.; Yoshigaki, A.; Arihara, S. Two new 26,27-cyclosterols from the marine sponge *Strongylophora corticate*. *J. Nat. Prod.* **2000**, *63*, 1540–1542. [[CrossRef](#)]
214. Minale, L.; Riccio, R.; Scalona, O.; Sodano, G.; Fattorusso, E.; Magno, S.; Mayol, L.; Santacrose, C. Metabolism in Porifera. VII. Conversion of [7,7-3H₂]-fucosterol into calysterol by the sponge *Calyx niceaensis*. *Experientia* **1977**, *33*, 1550–1552. [[CrossRef](#)]
215. Fattorusso, E.; Magno, S.; Mayol, L.; Santacrose, C.; Sioa, D. Calysterol: A C-29 cyclopropene-containing marine sterol from the sponge *Calyx niceaensis*. *Tetrahedron* **1975**, *31*, 1715–1716. [[CrossRef](#)]
216. O'Connor, J.M.; Pu, L.; Chadha, R.K. Metallacycle annelation: Reaction of a metallacycle alpha-substituent and a vinylidene ligand to give a bicyclic metallalactone complex. *J. Am. Chem. Soc.* **1990**, *112*, 9627–9628. [[CrossRef](#)]
217. Li, L.N.; Li, H.T.; Lang, R.W.; Itoh, T.; Sica, D.; Djerassi, C. Minor and trace sterols in marine invertebrates. 31. Isolation and structure elucidation of 23H-isocalysterol, a naturally occurring cyclopropene. Some comparative observations on the course of hydrogenolytic ring opening of steroidal cyclopropenes and cyclopropanes. *J. Am. Chem. Soc.* **1982**, *104*, 6726–6732.
218. Gunasekera, S.P.; Cranick, S.; Pomponi, S.A. Pomponi, New sterol ester from a deep-water marine sponge, *Xestospongia* sp. *J. Nat. Prod.* **1991**, *54*, 1119–1122. [[CrossRef](#)]
219. Dembitsky, V.M. Natural neo acids and neo alkanes: Their analogs and derivatives. *Lipids* **2006**, *41*, 309–340. [[CrossRef](#)]
220. Bisel, P.; Al-Momani, L.; Müller, M. The tert-butyl group in chemistry and biology. *Org. Biomol. Chem.* **2008**, *6*, 2655–2665. [[CrossRef](#)]
221. Dembitsky, V.M. Alkoxy lipids of the Organic World. Chemistry and Biology. Ph.D. Thesis, Lomonosov University of Fine Chemical Technology, Moscow, Russia, 1996.
222. Akihisa, A.; Inada, Y.; Ghosh, P. Compositions of triterpene alcohols of seeds and mature plants of family Cucurbitaceae. *J. Am. Oil Chem. Soc.* **1988**, *65*, 607–610. [[CrossRef](#)]

223. Akihisa, T.; Tamura, T.; Matsumoto, T. 24-Methylene-25-methylthosterol: A sterol from *Sicyos angulatus*. *Phytochemistry* **1987**, *26*, 575–577. [[CrossRef](#)]
224. Rahimi, R.; Shams-Ardekani, M.R.; Abdollahi, M. A review of the efficacy of traditional Iranian medicine for inflammatory bowel disease. *World J. Gastroenterol.* **2010**, *16*, 4504–4514. [[CrossRef](#)]
225. Bojić, M.; Maleš, Ž.; Antolić, A.; Babić, I.; Tomičić, M. Antithrombotic activity of flavonoids and polyphenols rich plant species. *Acta Pharma.* **2019**, *69*, 483–495. [[CrossRef](#)]
226. Sestili, P.; Ismail, T.; Calcabrini, C.; Guescini, M.; Catanzaro, E.; Turrini, E. The potential effects of *Ocimum basilicum* on health: A review of pharmacological and toxicological studies. *Expert Opin. Drug Metab. Toxicol.* **2018**, *14*, 679–692. [[CrossRef](#)]
227. Devi, P.U. Radioprotective, anticarcinogenic and antioxidant properties of the Indian holy basil, *Ocimum sanctum* (Tulasi). *Indian J. Exp. Biol.* **2001**, *39*, 185–190.
228. Ch, M.A.; Naz, S.B.; Sharif, A.; Akram, M.; Saeed, M.A. Biological and pharmacological properties of the sweet basil (*Ocimum basilicum*). *Br. J. Pharm Res.* **2015**, *7*, 330–339. [[CrossRef](#)]
229. Siddiqui, B.S.; Aslam, H.; Ali, S.T. Two new triterpenoids and a steroidal glycoside from the aerial parts of *Ocimum basilicum*. *Chem. Pharm Bull.* **2007**, *55*, 516–519. [[CrossRef](#)]
230. Li, Y. Stereo Chemical Studies on the Metabolism of Sterols by *Saccharomyces Cerevisiae* Strain GL7. Ph.D. Thesis, Texas Tech University, Lubbock, TX, USA, 1996.
231. Liu, J.; Nes, W.D. Steroidal triterpenes: Design of substrate-based inhibitors of ergosterol and sitosterol synthesis. *Molecules* **2009**, *14*, 4690–4706. [[CrossRef](#)]
232. Ishibashi, M.; Yamagishi, E.; Kobayashi, J. Topsentinols A–J, new sterols with highly branched side chains from marine sponge *Topsentia* sp. *Chem. Pharm Bull.* **1997**, *45*, 1435–1438. [[CrossRef](#)]
233. Nojo, R.; Echigo, S.; Hara, N.; Fujimoto, Y. C-24 stereochemistry of marine sterols: (22E)-25,28-dimethylstigmasta-5,22,28-trien-3 β -ol and 25,28-dimethylstigmasta-5,28-dien-3 β -ol. *Nat. Prod. Commun.* **2014**, *9*, 1699–1704. [[CrossRef](#)]
234. Li, X.; Djerassi, C. Minor and trace sterols in marine invertebrates 40. Structure and synthesis of axinyssasterol, 25-methylfucosterol and 24-ethyl-24-methylcholesterol—Novel sponge sterols with highly branched side chains. *Tetrahedron Lett.* **1983**, *24*, 665–668. [[CrossRef](#)]
235. Shubina, L.K.; Makar'eva, T.N.; Stonik, V.A. Steroidal compounds of marine sponges. III. 24-Ethyl-25-methylcholesta-5,22-dien-3 β -ol—A novel marine sterol from the sponge *Halichondria* sp. *Khim. Prir. Soedin.* **1984**, *4*, 464–467.
236. Shubina, L.K.; Makar'eva, T.N.; Stonik, V.A. Steroidal compounds of marine sponges. VI. Sterols and their derivatives from *Trachyopsis aplysinooides*. *Khim. Prir. Soedin.* **1985**, *5*, 715–716.
237. Fusetani, N.; Matsunaga, S.; Konosu, S. Bioactive marine metabolites. II. Halistanol sulfate, an antimicrobial novel steroid sulfate from the marine sponge *Halichondria* cf. *moorei* Bergquist. *Tetrahedron Lett.* **1981**, *22*, 1985–1988. [[CrossRef](#)]
238. Gulavita, N.K.; Wright, A.E.; Kelly-Borges, M.; Longley, R.E. Eryloside E from an Atlantic sponge *Erylus goffrilleri*. *Tetrahedron Lett.* **1994**, *35*, 4299–4302. [[CrossRef](#)]
239. Ebada, S.S.; Lin, W.H.; Proksch, P. Bioactive sesterterpenes and triterpenes from marine sponges: Occurrence and pharmacological significance. *Marine Drugs* **2010**, *8*, 313–346. [[CrossRef](#)]
240. Morrison-Gardiner, S. Dominant fungi from Australian reefs. *Fungal Divers.* **2002**, *9*, 105–121.
241. Jones, E.B.G.; Sakayaroj, J.; Suetrong, S.; Somrithipol, S.; Pang, K.L. Classification of marine Ascomycota, anamorphic taxa and Basidiomycota. *Fungal Divers.* **2009**, *35*, 1–187.
242. El-Bondkly, E.A.M.; El-Bondkly, A.A.M.; El-Bondkly, A.A.M. Marine endophytic fungal metabolites: A whole new world of pharmaceutical therapy exploration. *Heliyon* **2021**, *7*, e06362. [[CrossRef](#)]
243. Wang, S.K.; Dai, C.F.; Duh, C.Y. Cytotoxic pregnane steroids from the Formosan Soft Coral *Stereonephthya crystalliana*. *J. Nat. Prod.* **2006**, *69*, 103–106. [[CrossRef](#)]
244. Murillo-Alvarez, J.; Encarnacion-Dimayuga, R. New bioactive pregnadiene-derived glycosides from the gulf of California gorgonian *Muricea* cf. *austera*. *Pharm. Biol.* **2003**, *41*, 531–535. [[CrossRef](#)]
245. Anjaneyulu, A.S.R.; Rao, V.L.; Sastry, V.G. A new spiroketal steroid from *Gorgonella umbraculum*. *Nat. Prod. Res.* **2003**, *17*, 149–152. [[CrossRef](#)] [[PubMed](#)]
246. Seo, Y.; Rho, J.R.; Cho, K.W.; Shin, J. Isolation of new steroidal hemiacetals from the gorgonian *Euplexaura anastomosans*. *J. Nat. Prod.* **1996**, *59*, 1196–1199. [[CrossRef](#)]
247. Huang, C.Y.; Chang, C.W.; Sheu, J.H. Bioactive steroids from the Formosan soft coral *Umbellulifera petasites*. *Mar. Drugs* **2016**, *14*, 180. [[CrossRef](#)]
248. Blunt, J.W.; Copp, B.R.; Munro, M.H.G.; Northcote, P.T.; Prinsep, M.R. Marine natural products. *Nat. Prod. Rep.* **2005**, *22*, 15–61. [[CrossRef](#)]
249. Ngoc, N.T.; Huong, P.T.M.; Thanh, N.V.; Cuong, N.X. Steroid constituents from the soft coral *Simularia nanolobata*. *Chem. Pharm. Bull.* **2016**, *64*, 1417–1419. [[CrossRef](#)] [[PubMed](#)]
250. Gebreyesus, T.; Stoilov, I.; Luo, F.T.; Djerassi, C. Minor and trace sterols in marine invertebrates 55. The isolation, structure elucidation and synthesis of ergosta-5,24(28),25-trien-3-ol. *Steroids* **1985**, *45*, 447–452. [[CrossRef](#)]
251. Chao, C.H.; Chou, K.J.; Huang, C.Y.; Wen, Z.H.; Hsu, C.H. Steroids from the soft coral *Simularia crassa*. *Mar. Drugs* **2012**, *10*, 439–450. [[CrossRef](#)]

252. Mariottini, G.L. The role of cnidaria in drug discovery. In *The Cnidaria, Past, Present and Future*; Goredó, S., Dubinsky, Z., Eds.; Springer: Cham, Switzerland, 2016.
253. İlhan, H.A.; Pulat, C.C. Cytotoxic and antitumor compounds from marine invertebrates. *Encycl. Mar. Biotechnol.* **2020**, *4*, 2529–2584. [[CrossRef](#)]
254. Ngoc, N.T.; Huong, P.T.M.; Van Thanh, N.; Chia, N.T.P.C. Cytotoxic steroids from the Vietnamese soft coral *Simularia conferta*. *Chem. Pharm. Bull.* **2017**, *65*, 300–305. [[CrossRef](#)]
255. Cuing, N.X.; Nhiem, N.X.; Thanh, N.V. Review of chemistry and biological activity studies some marine species in Vietnam in the period 2013–2017. *Vietnam J. Chem.* **2018**, *56*, 1–19.
256. Cardoso-Martínez, F.; de la Rosa, J.M.; Díaz-Marrero, A.R.; Darias, J. Oxysterols from an octocoral of the genus *Gorgonia* from the eastern Pacific of Panama. *J. RSC Adv.* **2013**, *6*, 38579–38591. [[CrossRef](#)]
257. Yan, X.; Liu, J.; Leng, X.; Ouyang, H. Chemical diversity and biological activity of secondary metabolites from soft coral genus *Simularia* since 2013. *Mar. Drugs* **2021**, *19*, 335. [[CrossRef](#)]
258. Radhika, P. Chemical constituents and biological activities of the soft corals of genus *Cladiella*: A review. *Biochem. Syst. Ecol.* **2006**, *34*, 781–789. [[CrossRef](#)]
259. Zubair, M.S.; Al-Footy, K.O.; Ayyad, S.-E.N.; Al-Lihaibi, S.S.; Alarif, W.M. A review of steroids from *Sarcophyton* species. *Nat. Prod. Res.* **2016**, *30*, 869–879. [[CrossRef](#)]
260. Amir, F.; Koay, Y.C.; Yam, W.S. Chemical constituents, and biological properties of the marine soft coral *Nephthea*: A review (Part 1). *Trop. J. Pharm. Res.* **2012**, *11*, 485–498.
261. Amir, F.; Koay, Y.C.; Yam, W.S. Chemical constituents, and biological properties of the marine soft coral *Nephthea*: A review (Part 2). *Trop. J. Pharm. Res.* **2012**, *11*, 499–517.
262. Baulieu, E.E. Steroid hormones in the brain: Several mechanisms? Steroid hormone regulation of the brain. In Proceedings of the International Symposium Held at the Wenner-Gren Center, Stockholm, Sweden, 27–28 October 1980; pp. 3–14.
263. Vest, R.S.; Pike, C.J. Gender, sex steroid hormones, and Alzheimer’s disease. *Horm Behav.* **2013**, *63*, 301–307. [[CrossRef](#)]
264. Lenz, K.M.; McCarthy, M.M. Organized for sex–steroid hormones and the developing hypothalamus. *Eur. J. Neurosci.* **2010**, *32*, 2096–2104. [[CrossRef](#)]
265. Tchernof, A.; Després, J.P. Sex steroid hormones, sex hormone-binding globulin, and obesity in men and women. *Horm. Metab. Res.* **2000**, *32*, 526–536. [[CrossRef](#)]
266. Luine, V.N. Sex steroids and cognitive function. *J. Neuroendocrinol.* **2008**, *20*, 866–872. [[CrossRef](#)]
267. Rubinow, K.B. An intracrine view of sex steroids, immunity, and metabolic regulation. *Mol. Metab.* **2018**, *15*, 92–103. [[CrossRef](#)] [[PubMed](#)]
268. Owens, S.J.; Murphy, C.E.; Purves-Tyson, T.D.; Weickert, T.W.; Weickert, S.C. Considering the role of adolescent sex steroids in schizophrenia. *J. Neuroendocrinol.* **2018**, *30*, e12538. [[CrossRef](#)] [[PubMed](#)]
269. Wang, S.; Dong, G.; Sheng, C. Structural simplification of natural products. *Chem. Rev.* **2019**, *119*, 4180–4220. [[CrossRef](#)] [[PubMed](#)]
270. Nahar, L.; Sarker, S.D. A review on steroid dimers: 2011–2019. *Steroids* **2020**, *164*, 108736. [[CrossRef](#)]
271. Sahu, P.; Gidwani, B.; Dhongade, H.J. Pharmacological activities of dehydroepiandrosterone: A review. *Steroids* **2020**, *153*, 108507. [[CrossRef](#)]
272. Biellmann, J.-F. Enantiomeric steroids: synthesis, physical, and biological properties. *Chem. Rev.* **2003**, *103*, 2019–2034. [[CrossRef](#)]
273. Skoda-Földes, R.; Kollár, L. Transition-metal-catalyzed reactions in steroid synthesis. *Chem. Rev.* **2003**, *103*, 4095–4130. [[CrossRef](#)]
274. Albano, G.D.; Amico, F.; Cocimano, G.; Liberto, A.; Maglietta, F. Adverse effects of anabolic-androgenic steroids: A literature review. *Healthcare* **2021**, *9*, 97. [[CrossRef](#)]
275. Pope, H.G., Jr.; Kanayama, G.; Hudson, J.I.; Kaufman, M.J. Anabolic-androgenic steroids, violence, and crime: Two cases and literature review. *Am. J. Addict.* **2021**, *30*, 423–432. [[CrossRef](#)]
276. Hearne, E.; Wazaify, M.; Van Hout, M.C.; Atkinson, A.; McVeigh, J. Anabolic-androgenic steroid use in the Eastern Mediterranean region: A scoping review of extant empirical literature. *Int. J. Ment. Health Addict.* **2021**, *19*, 1162–1189. [[CrossRef](#)]
277. Shamsaei, N.; Ahmadian, N. Investigation of psychological relationship between the level of religiosity and doping susceptibility among bodybuilding athletes. *Relig. Health Spring Summer* **2020**, *8*, 30–38.
278. Hoseini, M.; Yousefi, B.; Khazaei, A. The prevalence of anabolic-androgenic steroids abuse, knowledge and attitude of their side effects, and attitude toward them among the female bodybuilding athletes in kermanshah. *J. Fasa Univ. Med. Sci.* **2020**, *10*, 2436–2446.
279. Murtha, R.; Heffernan, C.; Hunt, T. Definition diets and deteriorating masculinity? Bodybuilding diets in Mid-Century America. *Glob. Food Hist.* **2021**, *7*, 71–91. [[CrossRef](#)]
280. Okano, M.; Sato, M.; Ikekita, A. Analysis of non-ketotic steroids 17 α -methyl-epithiostanol and desoxymethyl-testosterone in dietary supplements. *Drug Test. Anal.* **2009**, *1*, 518–525.
281. Akram, O.N.; Bursill, C.; Desai, R.; Heather, A.K.; Kazlauskas, R.; Handelsman, D.J.; Lambert, G. Evaluation of androgenic activity of nutraceutical-derived steroids using mammalian and yeast in vitro androgen bioassays. *Anal. Chem.* **2011**, *83*, 2065–2072. [[CrossRef](#)]
282. Díaz, F.C.; Sáez-González, E.; Benlloch, S.; Álvarez-Sotomayor, D. Albumin dialysis with MARS for the treatment of anabolic steroid-induced cholestasis. *Ann. Hepatol.* **2016**, *15*, 939–943.

283. Okuno, Y.; Nakabou, Y.; Suzuki, S.; Ichiba, S.; Sugiyama, H. Complete remission by mepitiostane in hypoplastic leukemia. *Rinsho Ketsueki* **1989**, *30*, 1280–1283.
284. Komeno, T. Steromal 2,3-Diol Cyclic Trithiocarbonate. U.S. Patent 3,139,128, 30 June 1964.
285. Korneno, T.; Kawanami, E. 11,12-Epithio Steroids of Pregnane Series. U.S. Patent 3160627, 25 December 1964.
286. Komeno, T. 2,3-Epithio-Steroids and Production Thereof. U.S. Patent 3,230,215, 18 January 1966.
287. Klimstra, P.D. Optionally 17-Hydrocarbon (Substituted), 17-Oxygenated-2,3-Epithio-5 α -Androstanes. U.S. Patent 3,405,124, 25 December 1968.
288. Hirata, M. Process for Stabilization of a Composition of 2,3-Epithio-Androstanes and Composition Obtained Thereby. U.S. Patent 3,670,080, 13 June 1972.
289. Miyake, T.; Uchida, K.; Kakushi, H.; Nomura, Y.; Kadowaki, M. 2 α ,3 α -epithio-5 α -androstan-17 β -yl 1-methoxycyclopentyl ether (10361-S), a new orally active anabolic-androgenic steroid. *Jpn. J. Pharmacol.* **1974**, *24*, 551–558. [[CrossRef](#)]
290. Kurachi, K.; Aono, T.; Tomoyama, J.; Matsumoto, K.; Nakasima, A. Effects of 2,3-epithio-5-androstan-17-ol (epitiostanol) on hypothalamo-pituitary-gonadal axis in humans. *Acta Obstet. Gynaecol. Jpn.* **1975**, *22*, 42–48.
291. Li, X.; Rhee, D.K.; Malhotra, R.; Mayeur, C.; Hurst, L.A.; Ager, E. Progesterone receptor membrane component-1 regulates hepcidin biosynthesis. *J. Clin. Investig.* **2016**, *126*, 389–401. [[CrossRef](#)]
292. Kang, L.; Li, X.Q.; Chen, C.X.; Wang, F.R. Research progress on structure modification and biological activity of 18 β -glycyrrhetic acid. *Curr. Opin. Compl. Alternat. Med.* **2014**, *1*, e00008.
293. Wu, S.; Wang, W.; Dou, J.H. Research progress on the protective effects of licorice-derived 18 β -glycyrrhetic acid against liver injury. *Acta Pharmacol. Sin.* **2021**, *42*, 18–26. [[CrossRef](#)]
294. Dembitsky, V.M.; Glorizova, T.A.; Poroikov, V.V. Pharmacological activities of epithio steroids. *J. Pharm. Res. Int.* **2017**, *18*, 1–19. [[CrossRef](#)]
295. Huang, M.; Xie, X.; Gong, P.; Wei, Y.; Du, H.; Xu, Y. A 18 β -glycyrrhetic acid conjugate with Vorinostat degrades HDAC3 and HDAC6 with improved antitumor effects. *Eur. J. Med. Chem.* **2020**, *188*, 111991. [[CrossRef](#)]
296. Conor, R. *Selenium in Food and Health*; Springer: Boston, MA, USA, 2006.
297. Kang, D.; Lee, J.; Wu, C. The role of selenium metabolism and selenoproteins in cartilage homeostasis and arthropathies. *Exp. Mol. Med.* **2020**, *52*, 1198–1208. [[CrossRef](#)]
298. Terry, N.; Zayed, A.M.; De Souza, M.P.; Tarun, A.S. Selenium in higher plants. *Ann. Rev. Plant Physiol. Plant Mol. Biol.* **2000**, *51*, 401–432. [[CrossRef](#)]
299. Pyrzynska, K.; Sentkowska, A. Selenium in plant foods: Speciation analysis, bioavailability, and factors affecting composition. *Crit. Rev. Food Sci. Nutrit.* **2021**, *61*, 1340–1352. [[CrossRef](#)]
300. Pilon-Smits, E.A.H. Selenium in plants. *Prog. Bot.* **2015**, *76*, 93–106.
301. Nogueira, C.W.; Barbosa, N.V.; Rocha, J.B.T. Toxicology and pharmacology of synthetic organoselenium compounds: An update. *Arch. Toxicol* **2021**, *95*, 1179–1226. [[CrossRef](#)]
302. Ranu, B.C.; Banerjee, B. *Organoselenium Chemistry*; De Gruyter: Berlin, Germany; Boston, MA, USA, 2020.
303. Li, Q.S.; Wu, D.M.; Zhu, B.C.; Wang, Y.G. Organic selenium resin in solid phase synthesis and its application in constructing medicinally relevant small organic molecules. *Mini Rev. Med. Chem.* **2013**, *13*, 854–869. [[CrossRef](#)] [[PubMed](#)]
304. Dembitsky, V.M.; Glorizova, T.A.; Poroikov, V.V. Biological activities of organometalloid (As, At, B, Ge, Si, Se, Te) steroids. *J. App. Pharm. Sci.* **2017**, *7*, 184–202.
305. Wirth, T. *Organoselenium Chemistry: Synthesis and Reactions*; John Wiley & Sons, Inc.: Weinheim, Germany, 2011.
306. Ibrahim-Ouali, M. Total synthesis of steroids and heterosteroids from BISTRO. *Steroids* **2015**, *98*, 9–28. [[CrossRef](#)] [[PubMed](#)]
307. Ibrahim-Ouali, M. First total synthesis of 11-selena steroids. *Tetrahedron Lett.* **2009**, *50*, 1607–1609. [[CrossRef](#)]
308. Ibrahim-Ouali, M. First total synthesis of 11-tellura steroids. *Tetrahedron Lett.* **2010**, *51*, 3610–3612. [[CrossRef](#)]
309. Ibrahim-Ouali, M.; Romero, E.; Bouleghlem, H. First total syntheses of (\pm)-3-aza-11-selena and (\pm)-3-aza-11-tellura steroids. *Tetrahedron* **2011**, *67*, 3668–3676. [[CrossRef](#)]
310. Santi, C. *Organoselenium Chemistry: Between Synthesis and Biochemistry*; Bentham Science Publishers: Sharjah, United Arab Emirates, 2014.
311. Knapp, F.F. The synthesis of 123Te-labeled 17 β -hydroxy-2-tellura-A-nor-5 α -androstane. *J. Label. Comp. Radiopharm.* **1980**, *17*, 81–91. [[CrossRef](#)]
312. Leimbach, D.; Karls, J.; Guo, Y.; Ahmed, R.; Ballof, J.; Bengtsson, L.; Pamies, F.B.; Borshevsky, A. The electron affinity of astatine. *Nat. Commun.* **2020**, *11*, 3824. [[CrossRef](#)]
313. Kugler, H.K.; Keller, C. *At Astatine*; Springer: Berlin/Heidelberg, Germany, 1985.
314. Eychenne, R.; Bouvry, C.; Bourgeois, M.; Loyer, P.; Benoist, E.; Lepareur, N. Overview of radiolabeled somatostatin analogs for cancer imaging and therapy. *Molecules* **2020**, *25*, 4012. [[CrossRef](#)]
315. Visser, G.W.M.; Diemer, E.L.; Kaspersen, F.M. The preparation of aromatic astatine compounds through aromatic mercury compound's part II: Astatination of pyrimidines and steroids. *J. Label. Comp. Radiopharm.* **1981**, *18*, 799–807. [[CrossRef](#)]
316. Liu, B.L.; Jin, Y.T.; Liu, Z.H.; Luo, C.; Kojima, M.; Maeda, M. Halogen exchanges using crown ethers: Synthesis and preliminary biodistribution of 6-(211At)-astatomethyl-19-norcholest-5(10)-en-3 β -ol. *Int. J. Appl. Radiat. Isot.* **1985**, *36*, 561–563.