



Review

Significance of GABA_A Receptor for Cognitive Function and Hippocampal Pathology

Yuya Sakimoto *, Paw Min-Thein Oo, Makoto Goshima, Itsuki Kanehisa, Yutaro Tsukada and Dai Mitsushima

Department of Physiology, Yamaguchi University Graduate School of Medicine, Ube 755-8505, Japan; pawmto@gmail.com (P.M.-T.O.); b905eb@yamaguchi-u.ac.jp (M.G.); i029eb@yamaguchi-u.ac.jp (I.K.); i057@yamaguchi-u.ac.jp (Y.T.); mitsu@yamaguchi-u.ac.jp (D.M.)

* Correspondence: ysaki@yamaguchi-u.ac.jp

Abstract: The hippocampus is a primary area for contextual memory, known to process spatiotemporal information within a specific episode. Long-term strengthening of glutamatergic transmission is a mechanism of contextual learning in the dorsal cornu ammonis 1 (CA1) area of the hippocampus. CA1-specific immobilization or blockade of α -amino-3-hydroxyl-5-methyl-4-isoxazole-propionate (AMPA) receptor delivery can impair learning performance, indicating a causal relationship between learning and receptor delivery into the synapse. Moreover, contextual learning also strengthens GABA_A (gamma-aminobutyric acid) receptor-mediated inhibitory synapses onto CA1 neurons. Recently we revealed that strengthening of GABA_A receptor-mediated inhibitory synapses preceded excitatory synaptic plasticity after contextual learning, resulting in a reduced synaptic excitatory/inhibitory (E/I) input balance that returned to pretraining levels within 10 min. The faster plasticity at inhibitory synapses may allow encoding a contextual memory and prevent cognitive dysfunction in various hippocampal pathologies. In this review, we focus on the dynamic changes of GABA_A receptor mediated-synaptic currents after contextual learning and the intracellular mechanism underlying rapid inhibitory synaptic plasticity. In addition, we discuss that several pathologies, such as Alzheimer's disease, autism spectrum disorders and epilepsy are characterized by alterations in GABA_A receptor trafficking, synaptic E/I imbalance and neuronal excitability.

Keywords: AMPA receptor; GABA_A receptor; contextual learning; synaptic plasticity



Citation: Sakimoto, Y.; Oo, P.M.-T.; Goshima, M.; Kanehisa, I.; Tsukada, Y.; Mitsushima, D. Significance of GABA_A Receptor for Cognitive Function and Hippocampal Pathology. *Int. J. Mol. Sci.* **2021**, *22*, 12456. <https://doi.org/10.3390/ijms222212456>

Academic Editor: Natalia V. Gulyaeva

Received: 29 October 2021

Accepted: 8 November 2021

Published: 18 November 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

The hippocampal CA1 region has a total number of 350,000 neurons within a range from 320,000 to 380,000 at postnatal day 30 in Wistar rats [1]. Gamma-aminobutyric acid (GABA)ergic interneurons contain a conservative estimate of ~38,500 inhibitory interneurons in the CA1 region [2,3]. According to their molecular signatures, GABAergic interneurons can be divided into five main groups: Parvalbumin, somatostatin, neuropeptide Y, vasoactive intestinal peptide and cholecystokinin interneuron [4,5]. A single cornu ammonia 1 (CA1) pyramidal neuron receives approximately 3000 excitatory [6] and 1700 GABAergic synapses on their dendrites, somata and proximal axons [6]. While excitatory inputs target a distal dendritic spine of a CA1 pyramidal neuron, inhibitory inputs are largely concentrated in the perisomatic region. From this distribution of excitatory and inhibitory inputs, a potent perisomatic inhibition is considered to control dendritic excitatory inputs and play an important role in the decision-making of pyramidal cell activation itself [6].

2. The GABAergic System

GABA is the main inhibitory neurotransmitter in the mature mammalian central nervous system. GABA is stocked in synaptic vesicles and released in the synaptic cleft after stimulation by presynaptic neuron depolarization. GABA diffuses across the cleft to target receptors on the postsynaptic region. There are three types of GABA receptors

in the central nervous system, namely, ionotropic GABA_A and GABA_C receptors and metabotropic GABA_B receptors [5,7,8].

The nature of contextual fear learning-induced pre- and post-synaptic plasticity is complicated by the fact that learning also affects GABA_A receptor-mediated inhibitory synapses in CA1 pyramidal neurons [9–11]. GABA_A receptors typically consist of 2 α and 2 β subunits, together with either an γ or δ subunit [12]. Pore opening allows Cl[−] influx to induce postsynaptic hyperpolarization upon GABA binding. Considering that each presynaptic vesicle contains ~2500 molecules of GABA [13,14], we also quantified miniature postsynaptic GABA_A receptor currents induced by single-synaptic GABA vesicles (miniature inhibitory postsynaptic current (mIPSC)).

The activity of GABA_A receptor is regulated by cross-talk with other receptors, such as NMDA receptor, dopamine D5 receptor and GABA_B receptor [15,16]. GABA_A receptors are co-localized with them in certain synapses and their neurotransmitters are simultaneously activated or co-released [15]. While co-activation of these receptors occurs with GABA_A receptor-suppressed GABAergic inhibition, sole GABA_A receptor activity inhibits the response of these receptors [15].

3. Contextual Fear Memory Triggers Rapid Synaptic Plasticity

Pharmacological manipulation of AMPA or GABA_A receptors in the CA1 suggested different roles of the receptors after training [10,17–22]. Microinjections of an AMPA receptor blocker (7-nitro-2, 3-dioxo-1, 4-dihydroquinoxaline-6-carbonitrile (CNQX)) into the CA1 impairs inhibitory avoidance (IA) task training immediately (0–5 min), but these effects are lost 30–60 min after training [17,18,21], whereas GABA_A receptor blocker microinjection improves performance if administered immediately after training [17,20–22]. While these studies suggested a critical period for plasticity immediately after training, the dynamic changes in learning-induced synaptic diversity were poorly understood. Recently, Sakimoto et al. [23] revealed a dynamic of synaptic plasticity for memory in hippocampal CA1. Contextual learning rapidly strengthened E/I synapses in various ways in individual CA1 neurons, producing a broad diversity of synaptic input across the CA1 neuronal population within 5 min after training.

While rapid plasticity of excitatory CA1 synapses is considered an initial step of memory encoding rather than retrieval [23,24], conclusive evidence for the dynamic change of synaptic current is still lacking. We found a rapid increase in mEPSC amplitude within 5 min after IA training, showing that memory encoding rather than retrieval strengthens AMPA receptor-mediated excitatory synapses. Using fluctuation analysis of CA1 pyramidal neurons, we recently confirmed that training increased postsynaptic AMPA receptor channels without changing the cation current per channel and increase in presynaptic glutamate release [25]. As to the causal relationship between learning and plasticity, we previously reported that bilateral expression of GluA1-containing AMPA receptor delivery blockers in CA1 neurons impairs IA learning [26]. Moreover, a chromophore-assisted light-inactivation technique demonstrated that optical inactivation of synaptic AMPA receptors can erase acquired memories [27]. From these results, rapid trafficking of AMPA receptors after IA training is essential for encoding contextual memories.

The plasticity at inhibitory synapses seems to be task dependent and region specific [9,10,28]. As for hippocampal-dependent contextual learning, IA training clearly increased mIPSC amplitudes, suggesting postsynaptic strengthening of GABA_A receptor-mediated plasticity [10]. In addition, the mIPSC frequency rapidly increased without an increase in GABA release probability, suggesting a rapid activation of inhibitory silent or subthreshold synapses to increase the number of overthreshold synapses. Many mIPSC events may be small and below the detection threshold (<10 pA) and increased postsynaptic responses may increase the amplitude of these small events above the detection level (>10 pA), resulting in an apparent increase in mIPSC frequency. Moreover, Sakimoto et al. [23] found a rapid increase in mIPSC amplitude immediately after training, indicating that memory encoding rather than retrieval strengthens GABA_A receptor-mediated inhibitory synapses. This was the first report showing a rapid phosphory-

lation of the Ser^{408–409} GABA_A receptor β_3 subunit (GABA_AR β_3) within 1 min after training, concerning sites necessary to attenuate clathrin-dependent endocytosis of synaptic receptors, leading to both increased mIPSC amplitude and frequency in cultured neurons (Figure 1) [29].

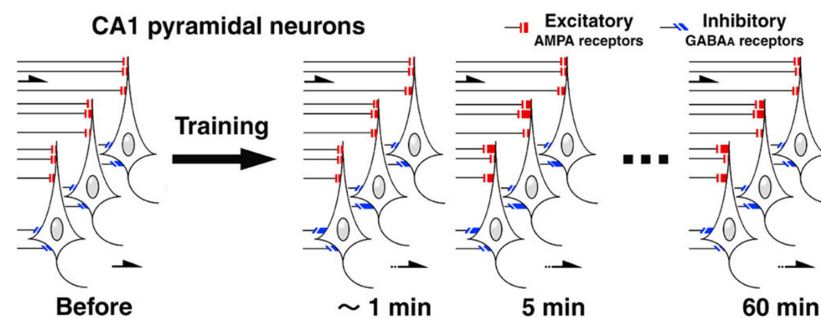


Figure 1. Schematic image of CA1 pyramidal neurons. IA training rapidly strengthened GABA_A receptor-mediated inhibitory synapses within 1 min, while the training strengthened AMPA receptor-mediated excitatory synapses within 5 min. CA1 pyramidal neurons exhibited broad diversity of excitatory/inhibitory synaptic currents within 5 min, and the neuron-specific synaptic diversity was sustained for more than 60 min.

A possible causal relationship between GABAergic plasticity and learning has been previously reported. Genetic deficiency of GABA_AR β_3 severely impairs the contextual freezing response without affecting pain perception [30], and phosphorylation in the cytoplasmic loop of the β_3 subunit (Ser^{408–409}) is known to play an essential role for PKA, PKB, PKC, Ca²⁺ and calmodulin-dependent protein kinase II-dependent plasticity [31], as phosphorylation can increase surface levels of GABA_A receptors containing β_3 subunits in cultured neurons (Figure 2) [32–35]. Not only the genetic deficiency of GABA_AR β_3 , but also prevention of GABA_A receptor-mediated plasticity in CA1 impairs contextual learning [10,30]. Optogenetic manipulation of CA1 neurons further proved the timing-specific causal relationship between GABAergic inputs and learning; optic inactivation of dendrite-targeting CA1 interneurons during aversive stimuli was sufficient to prevent fear teaching [11]. In a preliminary study, we found that microinjections of an interference peptide in Ser^{408–409} phosphorylation into the CA1 successfully blocked training-induced mIPSC strengthening. Moreover, bilateral peptide microinjections resulted in a drastic decrease in IA task-learning performance, suggesting further causal relationship between learning and Ser^{408–409} phosphorylation of the GABA_AR β_3 subunit.

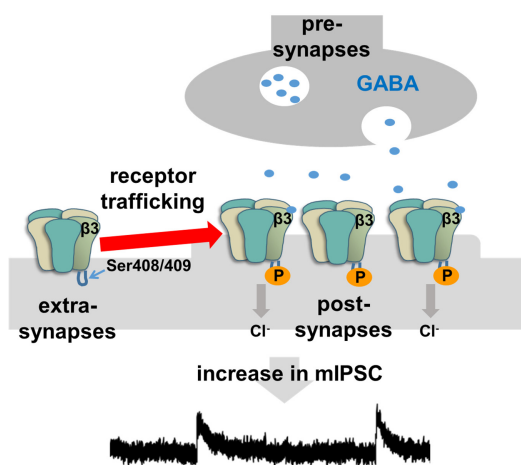


Figure 2. Schematic image of GABA_A receptor trafficking mechanisms. The phosphorylation in the β_3 subunit Ser^{408–409} increased levels of GABA_A receptors at post-synapses, resulting an increase in mIPSC. Since IA training facilitated the phosphorylation in the β_3 subunit (Ser^{408–409}) and GABA_A receptor-mediated current within 1 min, we suggested that the rapid inhibitory plasticity may contribute to maintaining memory function in hippocampus.

4. Intracellular Mechanism of Rapid Inhibitory Synaptic Plasticity

Questions arise as to how the training can increase GABA_A receptor-mediated currents so rapidly. GABA_A receptor mobility may be closely associated with the above issue, since removal from the postsynaptic membrane or lateral diffusion decreases the synaptic GABAergic current [36–38]. Recent single-particle tracking analysis further demonstrated quick diffusion of a single GABA_A receptor (0.07 μm²/s) in cultured hippocampal neurons; it can move rapidly between the two synapses within a few hundred milliseconds to a few seconds. Abundant GABA_A receptors heterosynaptically locate at glutamatergic synapses, and play a key role in the stimulus-dependent rapid changes in the postsynaptic number of receptors [39], probably because learning may rapidly recruit heterosynaptic GABA_A receptors to strengthen inhibitory synapses.

Once the receptor reaches the postsynaptic region through lateral diffusion [36,37], gephyrin seems to stabilize the synaptic receptors [31,40]. Gephyrin can bind to the major subunits of GABA_A receptors (α₁₋₃ and β₂₋₃) [41] and preventing its binding decreases mIPSC amplitudes [42]. Because phosphorylation of Ser^{408–409} GABA_ARβ3 is known to prevent clathrin adaptor protein 2-mediated GABA_A receptor internalization, training-induced Ser^{408–409} phosphorylation may help to stabilize surface receptors [43–45]. While training-induced Ser^{408–409} phosphorylation is rapid and transient, gephyrin may contribute to sustaining large mIPSC amplitude. Finally, using fluctuation analysis of CA1 pyramidal neurons, we recently confirmed that training increases the postsynaptic number of GABA_A receptor channels without changing the Cl[−] current per channel [24].

5. Alterations to GABA_ARβ3 in Cognitive Disease

Several pathologies, such as Alzheimer's disease (AD), autism spectrum disorders (ASDs), status epilepticus (SE) and posttraumatic stress disorder (PTSD), are characterized by synaptic E/I imbalance, neuronal hyperactivity and cognitive dysfunction [30,46–49]. In particular, alterations of GABA_ARβ3 have been observed in all these pathologies [50].

5.1. AD

AD is a progressive neurologic disorder characterized by a decrease in memory function and hippocampal alterations. Its early stage shows synaptic alterations and an increase in synaptic E/I balance and neuronal hyperactivity, resulting in induced neuron loss and reduction in hippocampal volume at late stages [51,52]. Amyloid β peptide 1–40 or 1–42 (Aβ_{1–40} or 1–42) is known as a major causative agent [53–56]. A biomarker study showed that Aβ_{1–42} accumulation signals the symptom onset of synaptic dysfunction, tau-mediated neuronal injury, brain structure, cognition and clinical function [57]. Soluble Aβ_{1–40} oligomers impair long-term potentiation and increased neuronal hyperactivity by glutamatergic/GABAergic imbalance in the hippocampus [51,52]. Long-term exposure to Aβ_{1–42} (1–3 d) impaired AMPA receptor trafficking by reducing the synaptic distribution of Ca²⁺ and calmodulin-dependent protein kinase II in cultured pyramidal neurons [58]. In contrast, the effect of soluble oligomeric assemblies of Aβ_{1–42} oligomer is more rapid, decreasing surface levels of AMPA receptors within 30 min [59].

While less is known about its toxic effects at inhibitory synapses, Aβ_{1–42} specifically binds to nicotinic α₇ receptors [60], impairing learning-induced plasticity at GABA_A receptor-mediated inhibitory synapses [10,61]. Bath application of Aβ_{1–42} weakens GABA_A receptor-mediated synaptic currents within 10 min through GABA_A receptor downregulation via receptor endocytosis in slice [62], while directly blocking nicotinic α₇ receptor-mediated cholinergic response within 3 min [63]. This result indicates that the disinhibited GABA_A receptor-mediated synaptic inhibition by Aβ leads to the hyperexcitability characteristic of AD, and might be partly related to the loss of functional GABA_A receptors in the AD brain [62,64]. Understanding the dynamic changes occurring during learning-promoted plasticity is necessary to identify a failure point in cognitive disorders.

GABA_A Receptor as Therapeutic Target in AD

Since A β weakened GABA_A receptor-mediated synaptic inhibition, GABA_A receptor agonists may improve either symptoms or progression of AD. A human AD patient showed several alterations in GABA_A receptor subunits including α_1 , α_2 , α_5 , β_2 , β_3 and γ_2 [65,66]. In cultured rat cortical neurons pre-treatment with muscimol, a GABA_A receptor agonist, ≥ 24 h prior to A β_{1-42} treatment inhibited A β_{1-42} -induced neuronal apoptosis and glutamate release [67]. Moreover, chronic administration of propofol to aged (18-months old) mice also decreased A β_{1-40} and A β_{1-42} levels [68]. However, baclofen, a GABA_A receptor and GABA_B receptor agonist, failed to inhibit A β_{1-42} induced neuronal death [67]. Thus, selective GABA_A receptor activation prevents A β 's adverse effects on neurons.

Moreover, AD patient hippocampus showed decreased GABA_AR β_3 expression [64,65]. Phosphorylation in β_3 subunit Ser⁴⁰⁸⁻⁴⁰⁹ facilitated synaptic trafficking of GABA_A receptor and prevented the receptor internalization, resulting in an increase in GABA_A receptor-mediated postsynaptic currents [29]. Recently, we reported that contextual learning rapidly strengthened GABA_A receptor-mediated postsynaptic currents and Ser⁴⁰⁸⁻⁴⁰⁹ phosphorylation in the β_3 subunit, suggesting that phosphorylation underlies rapid inhibitory synaptic plasticity and contextual memory encoding [23]. While A β_{1-42} treatment decreased GABA_A receptor-mediated postsynaptic currents via receptor internalization, inhibiting GABA_A receptor endocytosis prevented its adverse effects [62]. Thus, controlling GABA_A receptor trafficking may provide a new therapeutic target in AD.

A benzodiazepine (BZD) binding site is located in the extracellular domain of the GABA_A receptor, at the $\alpha+$ / $\gamma-$ interface, which modulates the GABA-induced Ch⁻ ion current [69]. AD patients show a reduction in the abundance of BZD sites in the hippocampus [70]. Baicalein (a positive allosteric modulator of the BZD site) significantly reduced A β production, improved cognitive function and decreased pathological features in an eight-week-old AD mouse model [68]. Moreover, our preliminary data shows that A β_{1-42} oligomers significantly impair the single channel current but not the number of channels in postsynaptic GABA_A receptors by using non-stationary fluctuation analysis, suggesting that A β_{1-42} oligomers act as a negative allosteric modulator [71]. Flumazenil, a silent or neutral allosteric modulator, was shown to prevent positive/negative allosteric modulator for the occupation of a binding site [72]. The hippocampus of AD patients showed a decrease in flumazenil binding, being positively correlated with hippocampal volume and memory function [73]. Thus, silent or neutral allosteric modulators may prevent adverse A β_{1-42} oligomer effects, improving hippocampal function at early stages of AD.

5.2. ASD

ASDs are a group of complex neurodevelopmental disorders characterized by repetitive behaviors and deficit of social cognitive and synaptic E/I imbalance [74]. They result from a complex interaction between genetics and the environment, with heritability estimates ranging from 40 to 80% [75,76]. Genetic studies have reported a few hundred genes linked to ASD, some encoding GABA_A receptor subunits, namely *GABRB3*, *GABRA5* and *GABRG3*, encoding for β_3 , α_5 and γ_3 subunits, respectively [74,76,77]. In particular, *GABRB3* (rs2081648 and rs1426217) presented a single-nucleotide polymorphism associated with ASD regardless of age or sex [74,78]. A deficiency of GABA_AR β_3 (*Gabrb3*) in mice reduces GABA_A receptor expression and enhances seizure susceptibility and autistic-like cognitive and motor deficits [30,79,80]. Indeed, ASD patients showed decreased expression of GABA_AR β_3 s in the parietal cortex and the cerebellum [81].

While the hippocampus of ASD patients has a larger volume than that from healthy persons from childhood to adolescence [82], few studies have examined hippocampal dysfunction in ASD. Recently, an ASD patient showed a deficit in hippocampal-dependent memory, including cognitive maps or episodic memory [83,84]. In addition, the hippocampal CA2 region plays an essential role for social recognition memory [85]. Recently, there

had been increasing interest is hippocampal dysfunction, synaptic alternation and relating cognition in ASD.

GABA_A Receptor as Therapeutical Target in ASD

Since genetic animal models for ASD have often shown a reduction in inhibitory neurotransmission, GABA agonists have been used as therapy [86]. *PX-RICS*^{-/-} mice (loss-of-*PX-RICS* function) exhibit ASD-like behaviors, and have reduced GABA_A receptor surface expression and lower mIPSC amplitude but not frequency [87]. A GABA_A receptor agonist (clonazepam, a positive allosteric modulator of the BZD site) improved some of its ASD-like behavioral phenotypes [87]. Other ASD mouse models (BTBR mice: Idiopathic autism; *Scn1a*^{+/-} mice: A monogenic model of ASDs [88]) also showed a reduced GABA_A receptor-mediated inhibition; treatment with positive allosteric modulators, either BZD or clonazepam, led to improved social and cognitive deficits [88,89]. Interestingly, a selective positive allosteric modulator of GABA_A receptor α_2 and/or α_3 subunits, L-838,417, also improved behavioral deficits in both BTBR and *Scn1a*^{+/-} mice [89]. Accordingly, clinical trials using α_2/α_3 selective positive allosteric modulators of GABA_A receptors have been developed by AstraZeneca and the National Institutes of Health [86].

In addition, a recent study reported an alteration of synaptic trafficking via phosphorylation in ASD [50,90]. The sodium valproate-induced rat ASD model shows impaired spatial memory, limited exploration, increased anxiety and reduced sociability [90], and reduced GABA_AR β_3 expression at different postnatal developmental stages, as well as downregulation of the phosphorylated form of the receptor subunit. This reduction facilitates receptor internalization, resulting in blocked inhibitory plasticity [50]. Thus, GABA_AR β_3 phosphorylation may prevent a decrease in GABA_A receptor expression and allow recovery from synaptic alterations and cognitive dysfunction in ASD.

5.3. SE

Epilepsy is group of neurological disorders characterized by a striking synaptic E/I imbalance [87]. SE is defined as seizure lasting >30 min or occurrence of ≥ 2 seizures without recovery of consciousness [91]. Inactivation of GABA_A receptors with bicuculline or picrotoxin results in epileptic seizures [92]. In addition, mutant animals lacking the β_3 (*Gabrb3*) subunit showed neuronal hyperactivity and seizures, which led to pathologies such as AD, ASD and Angelman syndrome [30,74,93–95].

Interestingly, epileptic patients consistently show cognitive deficits, but their underlying basis is yet to be determined [96]. In animal studies, kainic acid-induced SE impairs hippocampal-dependent short- and long-term spatial teaching [97], suggesting there are adverse effects of SE on hippocampal cognitive function. In the hippocampus, SE induced by pilocarpine, a non-selective mAChR agonist, decreased PKC-mediated phosphorylation of β_3 subunit Ser^{408–409} and increased binding to AP2 and GABA_A receptor endocytosis via dephosphorylation [98]. An acute stressor, such as foot shock or restrain, increased the performance in hippocampal-dependent tasks [99,100] and hippocampal BDNF concentration [100], while decreasing seizure susceptibility [101]. Recently, we found rapidly strengthened GABA_A receptor mediated synapses and phosphorylation in β_3 subunit Ser^{408–409} immediately after a foot shock in an IA task training. Thus, we suggested that the rapid inhibitory plasticity produced by the exposure to a stressful episode might contribute to reducing seizure vulnerability and maintaining cognitive function in the hippocampus.

GABA_A Receptor as Therapeutical Target in SE

The GABA_A receptor is a major target of antiseizure drugs [102–104]. In particular, BZDs, positive allosteric modulators, are effective in improving and blocking seizures [102,103]. Temporal lobe epilepsy has been linked to a significant loss of BZD binding sites [102], and the activation of GABA_A receptors by various allosteric ligands is crucial for the prevention of seizures [105]. Indeed, current SE treatment guidelines recommend a stepwise anti-seizure

medication treatment with up to two BZD doses within the first 5–10 min of SE onset, followed by non-BZD ASM after 10 min [106,107].

In addition, new drugs have focused on GABA_ARβ3 phosphorylation. Loreclezole, a subtype-selective positive allosteric modulator, increased the seizure threshold caused by a strongly potentiated recombinant GABA_A receptor containing a β₂ or β₃ subunit but not β₁-containing receptors. Moreover, phosphorylating β₃ subunit Ser^{408–409} by PDBu (a PKC activator) increases GABA_A receptor cell surface expression levels and recovers synaptic inhibition in SE [98]. Thus, rapid GABA_A mediated inhibitory plasticity via phosphorylation of β₃ subunit Ser^{408–409} may prevent seizure vulnerability and improve memory function in SE patients.

5.4. PTSD

PTSD is an anxiety disorder that occurs following exposure to severe trauma. The lifetime prevalence of PTSD is about 10–12% in women and 5–6% in men [108]. In human and animal studies, early traumatic experience, such as maternal separation, postnatal neglect and abuse, significantly increase abnormal behavioral reaction, alternation of brain morphology and synaptic plasticity in adulthood [109,110]. PTSD is characterized by deficits in GABAergic transmission and cognitive function in the brain, in particular the hippocampus [111]. Juvenile traumatic stress induced chronic anxiety, hippocampal-dependent memory loss and alternation in some subunit expression of GABA_A receptor in the hippocampus [110]. In addition, several studies reported that hippocampal GABAergic dysfunction attenuated juvenile stress induced an increase in risk factor of PTSD and cognitive and synaptic plasticity impairments [112–116]. Recently, Torrisi et al. [117] found an impaired hippocampal synaptic plasticity specifically at CA3–CA1 synapses of trauma susceptible mice showing long-lasting PTSD-like phenotypes.

GABA_A Receptor as Therapeutic Target in PTSD

During the juvenile period, exposure of traumatic stress induced alternation in some α subunit (α₁, α₂, and α₅) expression of GABA_A receptor in the hippocampus [110,118]. The α subunit of the GABA_A receptor is associated with various pharmacological properties of BZD [119]. PET (positron emission tomography) scan showed significantly reduced flumazenil binding through the cortex, hippocampus and thalamus in PTSD patients [120]. Treatment of BZD strengthened inhibitory neurotransmission by binding to the BZD site of the GABA_A receptor, resulting in improving anxiety and sleep disturbances in PTSD [110]. Recently, another study showed an increase in GABA_A receptor α₁ subunit expression in CA1 after juvenile traumatic stress [118]. They also reported that enriched environment exposure during juvenility prevented stress-associated increase of the α₁ subunit [118]. Thus, dysfunction of the GABA_A receptor α₁ subunit may improve some PTSD symptoms. On the other hand, while the relation of the GABA_ARβ3 subunit gene (GABRB3) to PTSD patients has been known [121], few studies have examined a therapeutic efficacy of the β₃ subunit. Thus, understanding of GABA_ARβ3 subunit in relation to PTSD will lead to the development of novel therapeutic agents.

6. Conclusions

Contextual learning not only induces synaptic delivery of AMPA receptors but also strengthens GABA_A receptor-mediated inhibitory synapses onto CA1 neurons. Several pathologies, such as AD, ASD and SE, are characterized by neuronal hyperactivity, down-regulation of inhibitory neurotransmission and alterations in GABA_ARβ3 trafficking and phosphorylation [50,64–66]. Indeed, GABA_A receptor agonist or positive allosteric modulator can help improve some symptoms of these pathologies. However, other GABA_A receptor subunits (e.g., α₁, α₂, α₅, β₂, β₃, and γ₂) are also consistently altered in these pathologies. While the complexity of these alterations is not compatible with a simple compensatory mechanism [65,66], we believe that understanding GABA_A receptor trafficking would provide new therapeutic targets for these pathologies.

Author Contributions: D.M. and Y.S. wrote the manuscript. P.M.-T.O., M.G., Y.T. and I.K. provided critical preliminary data for writing the manuscript. All authors have read and agreed to the published version of the manuscript.

Funding: This project was funded by Grants-in-Aid for Scientific Research B Grant Number 19H03402 (D.M.), Scientific Research C Grant Number 20K07276 (Y.S., D.M.), and Scientific Research in Innovative Areas Grant Number 26115518 (D.M.), from the Ministry of Education, Culture, Sports, Science, and Technology of Japan.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

- West, M.J.; Slomianka, L.; Gundersen, H.J. Unbiased stereological estimation of the total number of neurons in the subdivisions of the rat hippocampus using the optical fractionator. *Anat. Rec.* **1991**, *231*, 482–497. [[CrossRef](#)] [[PubMed](#)]
- Bezaire, M.J.; Soltesz, I. Quantitative assessment of CA1 local circuits: Knowledge base for interneuron-pyramidal cell connectivity. *Hippocampus* **2013**, *23*, 751–785. [[CrossRef](#)] [[PubMed](#)]
- Pelkey, K.A.; Chittajallu, R.; Craig, M.T.; Tricoire, L.; Wester, J.C.; McBain, C.J. Hippocampal GABAergic inhibitory interneurons. *Physiol. Rev.* **2017**, *97*, 1619–1747. [[CrossRef](#)]
- DeFelipe, J.; López-Cruz, P.L.; Benavides-Piccione, R.; Bielza, C.; Larranaga, P.; Anderson, S.; Burkhalter, A.; Cauli, B.; Fairen, A.; Feldmeyer, D.; et al. New insights into the classification and nomenclature of cortical GABAergic interneurons. *Nat. Rev. Neurosci.* **2013**, *14*, 202–216. [[CrossRef](#)]
- Li, J.; Chen, L.; Guo, F.; Han, X. The effects of GABAergic system under cerebral ischemia: Spotlight on cognitive function. *Neural Plast.* **2020**, *2020*, 8856722. [[CrossRef](#)]
- Megias, M.; Emri, Z.; Freund, T.F.; Gulyas, A.I. Total number and distribution of inhibitory and excitatory synapses on hippocampal CA1 pyramidal cells. *Neuroscience* **2001**, *102*, 527–540. [[CrossRef](#)]
- Bowery, N.G.; Hudson, A.L.; Price, G.W. GABA_A and GABA_B receptor site distribution in the rat central nervous system. *Neuroscience* **1987**, *20*, 365–383. [[CrossRef](#)]
- Bormann, J.; Feigenspan, A. GABA_C receptor. *Trends Neurosci.* **1995**, *18*, 515–519. [[CrossRef](#)]
- Cui, Y.; Costa, R.M.; Murphy, G.G.; Elgersma, Y.; Zhu, Y.; Gutmann, D.H.; Parada, L.F.; Mody, I.; Silva, A.J. Neurofibromin regulation of ERK signaling modulates GABA release and learning. *Cell* **2008**, *135*, 549–560. [[CrossRef](#)]
- Mitsushima, D.; Sano, A.; Takahashi, T. A cholinergic trigger drives learning-induced plasticity at hippocampal synapses. *Nat. Commun.* **2013**, *4*, 2760. [[CrossRef](#)]
- Lovett-Barron, M.; Kaifosh, P.; Kheirbek, M.A.; Danielson, N.; Zaremba, J.D.; Reardon, T.R.; Turi, G.F.; Hen, R.; Zemelman, B.V.; Losonczy, A. Dendritic inhibition in the hippocampus supports fear learning. *Science* **2014**, *343*, 857–863. [[CrossRef](#)]
- Sallard, E.; Letourneur, D.; Legendre, P. Electrophysiology of ionotropic GABA receptors. *Cell Mol. Life Sci.* **2021**, *78*, 5341–5370. [[CrossRef](#)] [[PubMed](#)]
- Telgkamp, P.; Padgett, D.E.; Ledoux, V.A.; Woolley, C.S.; Raman, I.M. Maintenance of high-frequency transmission at Purkinje to cerebellar nuclear synapses by spillover from boutons with multiple release sites. *Neuron* **2004**, *41*, 113–126. [[CrossRef](#)]
- Pugh, J.R.; Raman, I.M. GABA_A receptor kinetics in the cerebellar nuclei: Evidence for detection of transmitter from distant release sites. *Biophys. J.* **2005**, *88*, 1740–1754. [[CrossRef](#)] [[PubMed](#)]
- Shrivastava, A.N.; Triller, A.; Sieghart, W. GABA_A receptors: Post-synaptic co-localization and cross-talk with other receptors. *Front. Cell Neurosci.* **2011**, *5*, 7. [[CrossRef](#)] [[PubMed](#)]
- Maingret, F.; Groc, L. Characterization of the functional cross-talk between surface GABA_A and dopamine D5 receptors. *Int. J. Mol. Sci.* **2021**, *22*, 4867. [[CrossRef](#)] [[PubMed](#)]
- Castellano, C.; McGaugh, J.L. Effects of post-training bicuculline and muscimol on retention: Lack of state dependency. *Behav. Neural Biol.* **1990**, *54*, 156–164. [[CrossRef](#)]
- Jerusalinsky, D.; Ferreira, M.B.C.; Walz, R.; Da Silva, R.C.; Bianchin, M.; Ruschel, A.C.; Zanatta, M.S.; Medina, J.H.; Izquierdo, I. Amnesia by post-training infusion of glutamate receptor antagonists into the amygdala, hippocampus and entorhinal cortex. *Behav. Neural Biol.* **1992**, *58*, 76–80. [[CrossRef](#)]
- Bonini, J.S.; Rodrigues, L.; Kerr, D.S.; Bevilaqua, L.R.; Cammarota, M.; Izquierdo, I. AMPA/kainate and group-I metabotropic receptor antagonists infused into different brain areas impair memory formation of inhibitory avoidance in rats. *Behav. Pharmacol.* **2003**, *14*, 161–166. [[CrossRef](#)]
- Luft, T.; Pereira, G.S.; Cammarota, M.; Izquierdo, I. Different time course for the memory facilitating effect of bicuculline in hippocampus, entorhinal cortex, and posterior parietal cortex of rats. *Neurobiol. Learn. Mem.* **2004**, *82*, 52–56. [[CrossRef](#)]
- Izquierdo, I.; Bevilaqua, L.R.M.; Rossato, J.I.; Bonini, J.S.; Medina, J.H.; Cammarota, M. Different molecular cascades in different sites of the brain control consolidation. *Trends Neurosci.* **2006**, *28*, 496–505. [[CrossRef](#)]

22. Kim, D.H.; Kim, J.M.; Park, S.J.; Cai, M.; Liu, X.; Lee, S.; Shin, C.Y.; Ryu, J.H. GABA_A receptor blockade enhances memory consolidation by increasing hippocampal BDNF levels. *Neuropsychopharmacology* **2012**, *37*, 422–433. [[CrossRef](#)] [[PubMed](#)]
23. Sakimoto, Y.; Kida, H.; Mitsushima, D. Temporal dynamics of learning-promoted synaptic diversity in CA1 pyramidal neurons. *FASEB J.* **2019**, *33*, 14382–14393. [[CrossRef](#)] [[PubMed](#)]
24. Miyashita, T.; Kubik, S.; Haghighi, N.; Steward, O.; Guzowski, J.F. Rapid activation of plasticity-associated gene transcription in hippocampal neurons provides a mechanism for encoding of one-trial experience. *J. Neurosci.* **2009**, *29*, 898–906. [[CrossRef](#)] [[PubMed](#)]
25. Sakimoto, Y.; Mizuno, J.; Kida, H.; Kamiya, Y.; Ono, Y.; Mitsushima, D. Learning promotes subfield-specific synaptic diversity in hippocampal CA1 neurons. *Cereb. Cortex* **2019**, *29*, 2183–2195. [[CrossRef](#)] [[PubMed](#)]
26. Mitsushima, D.; Ishihara, K.; Sano, A.; Kessels, H.W.; Takahashi, T. Contextual learning requires synaptic AMPA receptor delivery in the hippocampus. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 12503–12508. [[CrossRef](#)] [[PubMed](#)]
27. Takemoto, K.; Iwanari, H.; Tada, H.; Suyama, K.; Sano, A.; Nagai, T.; Hamakubo, T.; Takahashi, T. Optical inactivation of synaptic AMPA receptors erases fear memory. *Nat. Biotechnol.* **2017**, *35*, 38–47. [[CrossRef](#)] [[PubMed](#)]
28. Kida, H.; Tsuda, Y.; Ito, N.; Yamamoto, Y.; Owada, Y.; Kamiya, Y.; Mitsushima, D. Motor training promotes both synaptic and intrinsic plasticity of layer II/III pyramidal neurons in the primary motor cortex. *Cereb. Cortex* **2016**, *26*, 3494–3507. [[CrossRef](#)] [[PubMed](#)]
29. Kittler, J.T.; Chen, G.; Honing, S.; Bogdanov, Y.; McAinsh, K.; Arancibia-Carcamo, I.L.; Jovanovic, J.N.; Pangalos, M.N.; Hauche, V.; Yan, Z.; et al. Phospho-dependent binding of the clathrin AP2 adaptor complex to GABA_A receptors regulates the efficacy of inhibitory synaptic transmission. *Proc. Natl. Acad. Sci. USA* **2005**, *102*, 14871–14876. [[CrossRef](#)]
30. DeLorey, T.M.; Handforth, A.; Anagnostaras, S.G.; Homanics, G.E.; Minassian, B.A.; Asatourian, A.; Ellison, G.D.; Olsen, R.W. Mice lacking the β_3 subunit of the GABA_A receptor have the epilepsy phenotype and many of the behavioral characteristics of Angelman syndrome. *J. Neurosci.* **1998**, *18*, 8505–8514. [[CrossRef](#)] [[PubMed](#)]
31. Luscher, B.; Fuchs, T.; Kilpatrick, C.L. GABA_A receptor trafficking-mediated plasticity of inhibitory synapses. *Neuron* **2011**, *70*, 385–409. [[CrossRef](#)]
32. McDonald, B.J.; Moss, S.J. Differential phosphorylation of intracellular domains of gamma-aminobutyric acid type A receptor subunits by calcium/calmodulin type 2-dependent protein kinase and cGMP-dependent protein kinase. *J. Biol. Chem.* **1994**, *269*, 18111–18117. [[CrossRef](#)]
33. McDonald, B.J.; Amato, A.; Connolly, C.N.; Benke, D.; Moss, S.J.; Smart, T.G. Adjacent phosphorylation sites on GABA_A receptor beta subunits determine regulation by cAMP-dependent protein kinase. *Nat. Neurosci.* **1998**, *1*, 23–28. [[CrossRef](#)] [[PubMed](#)]
34. Brandon, N.J.; Delmas, P.; Kittler, J.T.; McDonald, B.J.; Sieghart, W.; Brown, D.A.; Smart, T.G.; Moss, S.J. GABA_A receptor phosphorylation and functional modulation in cortical neurons by a protein kinase C-dependent pathway. *J. Biol. Chem.* **2000**, *275*, 38856–38862. [[CrossRef](#)] [[PubMed](#)]
35. Brandon, N.J.; Jovanovic, J.N.; Smart, T.G.; Moss, S.J. Receptor for activated C kinase-1 facilitates protein kinase C-dependent phosphorylation and functional modulation of GABA_A receptors with the activation of G-protein-coupled receptors. *J. Neurosci.* **2002**, *22*, 6353–6361. [[CrossRef](#)] [[PubMed](#)]
36. Thomas, P.; Mortensen, M.; Hosie, A.M.; Smart, T.G. Dynamic mobility of functional GABA_A receptors at inhibitory synapses. *Nat. Neurosci.* **2005**, *8*, 889–897. [[CrossRef](#)] [[PubMed](#)]
37. Bogdanov, Y.; Michels, G.; Armstrong-Gold, C.; Haydon, P.G.; Lindstrom, J.; Pangalos, M. Synaptic GABA_A receptors are directly recruited from their extrasynaptic counterparts. *EMBO J.* **2006**, *25*, 4381–4389. [[CrossRef](#)]
38. Mele, M.; Leal, G.; Buarte, C.B. Role of GABA_AR trafficking in the plasticity of inhibitory synapses. *J. Neurochem.* **2016**, *139*, 997–1018. [[CrossRef](#)] [[PubMed](#)]
39. De Luca, E.; Ravasenga, T.; Petrini, E.M.; Polenghi, A.; Nieuws, T.; Guazzi, S.; Barberis, A. Inter-synaptic lateral diffusion of GABA_A receptors shapes inhibitory synaptic currents. *Neuron* **2017**, *95*, 63–69. [[CrossRef](#)] [[PubMed](#)]
40. Tretter, V.; Mukherjee, J.; Maric, H.M.; Schindelin, H.; Sieghart, W.; Moss, S.J. Gephyrin, the enigmatic organizer at GABAergic synapses. *Front. Cell Neurosci.* **2012**, *6*, 23. [[CrossRef](#)] [[PubMed](#)]
41. Kowalczyk, S.; Winkelmann, A.; Smolinsky, B.; Förstera, B.; Neundorff, I.; Schwarz, G.; Meier, J.C. Direct binding of GABA_A receptor β_2 and β_3 subunits to gephyrin. *Eur. J. Neurosci.* **2013**, *37*, 544–554. [[CrossRef](#)] [[PubMed](#)]
42. Mukherjee, J.; Kretschmannova, K.; Gouzer, G.; Maric, H.M.; Ramsden, S.; Tretter, V.; Harvey, K.; Davies, P.A.; Triller, A.; Schindelin, H.; et al. The residence time of GABA_ARs at inhibitory synapses is determined by direct binding of the receptor $\alpha 1$ subunit to gephyrin. *J. Neurosci.* **2011**, *31*, 14677–14687. [[CrossRef](#)] [[PubMed](#)]
43. Jovanovic, J.N.; Thomas, P.; Kittler, J.T.; Smart, T.G.; Moss, S.J. Brain-derived neurotrophic factor modulates fast synaptic inhibition by regulating GABA_A receptor phosphorylation, activity and cell-surface stability. *J. Neurosci.* **2004**, *24*, 522–530. [[CrossRef](#)]
44. Lu, H.; Cheng, P.L.; Lim, B.K.; Khoshnevisrad, N.; Poo, M.M. Elevated BDNF after cocaine withdrawal facilitates LTP in medial prefrontal cortex by suppressing GABA inhibition. *Neuron* **2010**, *67*, 821–833. [[CrossRef](#)] [[PubMed](#)]
45. Petrini, E.M.; Barberis, A. Diffusion dynamics of synaptic molecules during inhibitory postsynaptic plasticity. *Front. Cell Neurosci.* **2014**, *8*, 300. [[CrossRef](#)] [[PubMed](#)]
46. Thompson-Vest, N.M.; Waldvogel, H.J.; Rees, M.I.; Faull, R.L. GABA_A receptor subunit and gephyrin protein changes differ in the globus pallidus in Huntington's diseased brain. *Brain Res.* **2003**, *994*, 265–270. [[CrossRef](#)] [[PubMed](#)]

47. Rudolph, U.; Möhler, H. Analysis of GABA_A receptor function and dissection of the pharmacology of benzodiazepines and general anesthetics through mouse genetics. *Annu. Rev. Pharmacol. Toxicol.* **2004**, *44*, 475–498. [[CrossRef](#)] [[PubMed](#)]
48. Rojas-Charry, L.; Nardi, L.; Methner, A.; Schmeisser, M.J. Abnormalities of synaptic mitochondria in autism spectrum disorder and related neurodevelopmental disorders. *J. Mol. Med.* **2020**, *99*, 161–178. [[CrossRef](#)] [[PubMed](#)]
49. Kang, J.O. Epileptic mechanisms shared by Alzheimer's disease: Viewed via the unique lens of genetic epilepsy. *Int. J. Mol. Sci.* **2021**, *22*, 7133. [[CrossRef](#)] [[PubMed](#)]
50. Mele, M.; Costa, R.O.; Duarte, C.B. Alterations in GABA_A-receptor trafficking and synaptic dysfunction in brain disorders. *Front. Cell Neurosci.* **2019**, *13*, 77. [[CrossRef](#)]
51. Lei, M.; Xu, H.; Li, Z.; Wang, Z.; O'Malley, T.T.; Zhang, D.; Walsh, D.M.; Xu, P.; Selkoe, D.J.; Li, S. Soluble A β oligomers impair hippocampal LTP by disrupting glutamatergic/GABAergic balance. *Neurobiol. Dis.* **2016**, *85*, 111–121. [[CrossRef](#)]
52. Vyas, Y.; Montgomery, J.M.; Cheyne, J.E. Hippocampal deficits in amyloid β related rodent models of Alzheimer's disease. *Front. Neurosci.* **2020**, *14*, 266. [[CrossRef](#)] [[PubMed](#)]
53. Shankar, G.M.; Li, S.; Mehta, T.H.; Garcia-Munoz, A.; Shepardson, N.E.; Smith, I.; Brett, F.M.; Farrell, M.A.; Rowan, M.J.; Lemere, C.A.; et al. Amyloid-beta protein dimers isolated directly from Alzheimer's brains impair synaptic plasticity and memory. *Nat. Med.* **2008**, *14*, 837–842. [[CrossRef](#)]
54. Querfurth, H.W.; LaFerla, F.M. Alzheimer's disease. *N. Engl. J. Med.* **2010**, *362*, 329–344. [[CrossRef](#)] [[PubMed](#)]
55. Penzes, P.; Cahill, M.E.; Jones, K.A.; VanLeeuwen, J.E.; Woolfrey, K.M. Dendritic spine pathology in neuropsychiatric disorders. *Nat. Neurosci.* **2011**, *14*, 285–293. [[CrossRef](#)] [[PubMed](#)]
56. Sevigny, J.; Chiao, P.; Bussiere, T.; Weinreb, P.H.; Maier, M.; Dunstan, R.; Salloway, S.; Chen, T.; Ling, Y.; O'Gorman, J.; et al. The antibody aducanumab reduces A β plaques in Alzheimer's disease. *Nature* **2016**, *537*, 50–56. [[CrossRef](#)]
57. Counts, S.E.; Ikonovic, M.D.; Mercado, N.; Vega, I.; Mufson, E.J. Biomarkers for the early detection and progression of Alzheimer's disease. *Neurotherapeutics* **2017**, *14*, 35–53. [[CrossRef](#)]
58. Gu, Z.; Liu, W.; Yan, Z. β -amyloid impairs AMPA receptor trafficking and function by reducing Ca²⁺/calmodulin-dependent protein kinase II synaptic distribution. *J. Biol. Chem.* **2009**, *284*, 10639–10649. [[CrossRef](#)]
59. Zhao, W.Q.; Santini, F.; Breese, R.; Ross, D.; Zhang, X.D.; Stone, D.J.; Ferrer, M.; Townsend, M.; Wolfe, A.L.; Seager, M.A.; et al. Inhibition of calcineurin-mediated endocytosis and AMPA receptor prevent amyloid β oligomer-induced synaptic disruption. *J. Biol. Chem.* **2010**, *285*, 7619–7632. [[CrossRef](#)]
60. Wang, H.Y.; Lee, D.H.; D'Andrea, M.R.; Peterson, P.A.; Shank, R.P.; Reitz, A.B. β -Amyloid₁₋₄₂ binds to α_7 nicotinic acetylcholine receptor with high affinity. Implications for Alzheimer's disease pathology. *J. Biol. Chem.* **2000**, *275*, 5626–5632. [[CrossRef](#)] [[PubMed](#)]
61. Townsend, M.; Whyment, A.; Walczak, J.S.; Jeggo, R.; van den Top, M.; Flood, D.G.; Leventhal, L.; Patzke, H.; Koenig, G. α_7 -nAChR agonist enhances neural plasticity in the hippocampus via a GABAergic circuit. *J. Neurophysiol.* **2016**, *116*, 2663–2675. [[CrossRef](#)] [[PubMed](#)]
62. Ulrich, D. Amyloid- β impairs synaptic inhibition via GABA_A receptor endocytosis. *J. Neurosci.* **2015**, *35*, 9205–9210. [[CrossRef](#)] [[PubMed](#)]
63. Liu, Q.; Kawai, H.; Berg, D.K. β -Amyloid peptide blocks the response of α_7 -containing nicotinic receptors on hippocampal neurons. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 4734–4739. [[CrossRef](#)] [[PubMed](#)]
64. Limon, A.; Reyes-Ruiz, J.M.; Mileti, R. Loss of functional GABA_A receptors in the Alzheimer diseased brain. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 10071–10076. [[CrossRef](#)] [[PubMed](#)]
65. Kwakowsky, A.; Calvo-Flores Guzmán, B.; Pandya, M.; Turner, C.; Waldvogel, H.J.; Faull, R.L. GABA_A receptor subunit expression changes in the human Alzheimer's disease hippocampus, subiculum, entorhinal cortex and superior temporal gyrus. *J. Neurochem.* **2018**, *145*, 374–392. [[CrossRef](#)] [[PubMed](#)]
66. Kwakowsky, A.; Calvo-Flores, G.B.; Govindpani, K.; Waldvogel, H.J.; Faull, R.L. Gamma-aminobutyric acid A receptors in Alzheimer's disease: Highly localized remodeling of a complex and diverse signaling pathway. *Neural Regen. Res.* **2018**, *13*, 1362–1363. [[CrossRef](#)] [[PubMed](#)]
67. Lee, B.Y.; Ban, J.Y.; Seong, Y.H. Chronic stimulation of GABA_A receptor with muscimol reduces amyloid beta protein (23-35)-induced neurotoxicity in cultured rat cortical cells. *Neurosci. Res.* **2005**, *52*, 347–356. [[CrossRef](#)] [[PubMed](#)]
68. Zhang, S.Q.; Obregon, D.; Ehrhart, J.; Deng, J.; Tian, J.; Hou, H.; Giunta, B.; Swamiller, D.; Tan, J.S.-Q.; Obregon, D.; et al. Baicalein reduces β -amyloid and promotes nonamyloidogenic amyloid precursor protein processing in an Alzheimer's disease transgenic mouse model. *J. Neurosci. Res.* **2013**, *91*, 1239–1246. [[CrossRef](#)] [[PubMed](#)]
69. Crocetti, L.; Guerrini, G. GABA_A receptor subtype modulators in medicinal chemistry: An updated patent review (2014-present). *Expert Opin. Ther. Pat.* **2020**, *30*, 409–432. [[CrossRef](#)]
70. Shimohama, T.; Taniguchi, T.; Fujiwara, M.; Kemeyama, M. Changes in benzodiazepine receptors in Alzheimer-type dementia. *Ann. Neurol.* **1998**, *23*, 404–406. [[CrossRef](#)] [[PubMed](#)]
71. Sakimoto, Y.; Tsukada, Y.; Kimura, R.; Kida, H.; Mitsushima, D. Adverse effects of A β ₁₋₄₂ oligomers: Contextual learning and GABA_A synapses in CA1 pyramidal neurons. *J. Physiol. Sci.* **2021**, *71* (Suppl. 1), 24.
72. Alanis, B.A.V.; Iorio, M.T.; Silva, L.L.; Bampali, K.; Ernst, M.; Schnurch, M.; Mihovilovic, M.D. Allosteric GABA_A receptor modulators—A review on the most recent heterocyclic chemotypes and their synthetic accessibility. *Molecules* **2020**, *25*, 999. [[CrossRef](#)]

73. Pascual, B.; Prieto, E.; Arbizu, J.; Marti-Climent, J.M.; Penuelas, I.; Quincoces, G.; Zarauza, R.; Pappata, S.; Masdeu, J.C. Decreased carbon-11-flumazenil binding in early Alzheimer's disease. *Brain* **2012**, *135*, 2817–2825. [[CrossRef](#)] [[PubMed](#)]
74. Menzikov, S.A.; Morozov, S.G.; Kubatiev, A.A. Intricacies of GABA_A receptor function: The Critical Role of the β_3 subunit in norm and pathology. *Int. J. Mol. Sci.* **2021**, *22*, 1457. [[CrossRef](#)]
75. Chaste, P.; Leboyer, M. Autism risk factors: Genes, environment, and gene-environment interactions. *Dialogues Clin. Neurosci.* **2012**, *14*, 281–292. [[PubMed](#)]
76. Rylaarsdam, L.; Guemez-Gamboa, A. Genetic causes and modifiers of autism spectrum disorder. *Front. Cell Neurosci.* **2019**, *13*, 385. [[CrossRef](#)] [[PubMed](#)]
77. Coghlan, S.; Horder, J.; Inkster, B.; Mendez, M.A.; Murphy, D.G.; Nutt, D. GABA system dysfunction in autism and related disorders: From synapse to symptoms. *Neurosci. Biobehav. Rev.* **2012**, *36*, 2033–2055. [[CrossRef](#)] [[PubMed](#)]
78. Rubenstein, J.L.; Merzenich, M.M. Model of autism: Increased ratio of excitation/inhibition in key neural systems. *Genes Brain Behav.* **2003**, *2*, 255–267. [[CrossRef](#)]
79. Homanics, G.E.; DeLorey, T.M.; Firestone, L.L.; Quinlan, J.J.; Handforth, A.; Harrison, N.L.; Krasowski, M.D.; Rick, C.E.M.; Korpi, E.R.; Makela, R.; et al. Mice devoid of γ -aminobutyrate type_A receptor β_3 subunit have epilepsy, cleft palate, and hypersensitive behavior. *Proc. Natl. Acad. Sci. USA* **1997**, *94*, 4143–4148. [[CrossRef](#)]
80. Lee, E.; Lee, J.; Kim, E. Excitation/inhibition imbalance in animal models of autism spectrum disorders. *Biol. Psychiatry* **2017**, *81*, 838–847. [[CrossRef](#)] [[PubMed](#)]
81. Fatemi, S.H.; Reutiman, T.J.; Folsom, T.D.; Thuras, P.D. GABA_A receptor downregulation in brains of subjects with autism. *J. Autism Dev. Disord.* **2009**, *39*, 223–230. [[CrossRef](#)]
82. Barnea-Goraly, N.; Frazier, T.W.; Piacenza, L.; Minshew, N.J.; Keshavan, M.S.; Reiss, A.L.; Hardan, A.Y. A preliminary longitudinal volumetric MRI study of amygdala and hippocampal volumes in autism. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **2014**, *48*, 124–128. [[CrossRef](#)] [[PubMed](#)]
83. Bangerter, A.; Ness, S.; Aman, M.G.; Esbensen, A.J.; Goodwin, M.S.; Dawson, G.; Hendren, R.; Leventhal, B.; Khan, A.; Opler, M.; et al. Autism behavior inventory: A novel tool for assessing core and associated symptoms of autism spectrum disorder. *J. Child Adolesc. Psychopharmacol.* **2017**, *27*, 814–822. [[CrossRef](#)] [[PubMed](#)]
84. Banker, S.M.; Gu, X.; Schiller, D.; Foss-Feig, J.H. Hippocampal contributions to social and cognitive deficits in autism spectrum disorder. *Trends Neurosci.* **2021**, *44*, 793–807. [[CrossRef](#)] [[PubMed](#)]
85. Culotta, L.; Penzes, P. Exploring the mechanisms underlying excitation/inhibition imbalance in human iPSC-derived models of ASD. *Mol. Autism* **2020**, *11*, 32. [[CrossRef](#)]
86. Cellot, G.; Cherubini, E. GABAergic signaling as therapeutic target for autism spectrum disorders. *Front. Pediatr.* **2014**, *2*, 70. [[CrossRef](#)] [[PubMed](#)]
87. Nakamura, Y.; Darnieder, L.M.; Deeb, T.Z.; Moss, S.J. Regulation of GABA_ARs by phosphorylation. *Adv. Pharmacol.* **2015**, *72*, 97–146. [[PubMed](#)]
88. Han, S.; Tai, C.; Westenbroek, R.E.; Yu, F.H.; Cheah, C.S.; Potter, G.B.; Rubenstein, J.L.; Scheuer, T.; de la Iglesia, H.O.; Catterall, W.A. Autistic-like behaviour in *Scn1a*^{+/-} mice and rescue by enhanced GABA-mediated neurotransmission. *Nature* **2012**, *489*, 385–390. [[CrossRef](#)] [[PubMed](#)]
89. Han, S.; Tai, C.; Jones, C.J.; Scheuer, T.; Catterall, W.A. Enhancement of inhibitory neurotransmission by GABA_A receptors having $\alpha 2,3$ -subunits ameliorates behavioral deficits in a mouse model of autism. *Neuron* **2014**, *81*, 1282–1289. [[CrossRef](#)] [[PubMed](#)]
90. Li, Y.; Sun, H.; Chen, Z.; Xu, H.; Bu, G.; Zheng, H. Implications of GABAergic Neurotransmission in Alzheimer's Disease. *Front. Aging Neurosci.* **2016**, *8*, 31. [[CrossRef](#)] [[PubMed](#)]
91. Cherian, A.; Thomas, S.V. Status epilepticus. *Ann. Indian Acad. Neurol.* **2009**, *12*, 140–153.
92. Asada, H.; Kawamura, Y.; Maruyama, K.; Kume, H.; Ding, R.; Ji, F.Y.; Kanbara, N.; Kuzume, H.; Sanbo, M.; Yagi, T.; et al. Mice lacking the 65 kDa isoform of glutamic acid decarboxylase (GAD65) maintain normal levels of GAD67 and GABA in their brains but are susceptible to seizures. *Biochem. Biophys. Res. Commun.* **1996**, *229*, 891–895. [[CrossRef](#)]
93. Janve, V.S.; Hernandez, C.C.; Verdier, K.M.; Hu, N.; Macdonald, R.L. Epileptic encephalopathy de novo GABRB mutations impair γ -aminobutyric acid type A receptor function. *Ann. Neurol.* **2016**, *79*, 806–825. [[CrossRef](#)] [[PubMed](#)]
94. Møller, R.S.; Wuttke, T.V.; Helbig, I.; Marini, C.; Johannesen, K.M.; Brilstra, E.H.; Vaher, U.; Borggraefe, I.; Talvik, I.; Talvik, T.; et al. Mutations in GABRB3: From febrile seizures to epileptic encephalopathies. *Neurology* **2017**, *88*, 483–492. [[CrossRef](#)]
95. Absalom, N.L.; Ahring, P.K.; Liao, V.W.; Balle, T.; Jiang, T.; Anderson, L.L.; Arnold, J.C.; McGregor, I.S.; Bowen, M.T.; Chebib, M. Functional genomics of epilepsy-associated mutations in the GABA_A receptor subunits reveal that one mutation impairs function and two are catastrophic. *J. Biol. Chem.* **2019**, *294*, 6157–6171. [[CrossRef](#)] [[PubMed](#)]
96. Bernard, C. Alterations in synaptic function in epilepsy. In *Jasper's Basic Mechanisms of the Epilepsies*, 4th ed.; Noebels, J.N., Avoli, M., Rogawski, M., Olsen, R., Delgado-Escueta, A., Eds.; Oxford University Press: New York, NJ, USA, 2012.
97. Sayin, U.; Sutula, T.P.; Stafstrom, C.E. Seizures in the developing brain cause adverse long-term effects on spatial learning and anxiety. *Epilepsia* **2004**, *45*, 1539–1548. [[CrossRef](#)] [[PubMed](#)]
98. Terunuma, M.; Xu, J.; Vithlani, M.; Sieghart, W.; Kittler, J.; Pangalos, M.; Haydon, P.G.; Coulter, D.A.; Moss, S.J. Deficits in phosphorylation of GABA_A receptors by intimately associated protein kinase C activity underlie compromised synaptic inhibition during status epilepticus. *J. Neurosci.* **2008**, *28*, 376–384. [[CrossRef](#)] [[PubMed](#)]

99. Aguayo, F.I.; Tejos-Bravo, M.; Diaz-Veliz, G.; Pacheco, A.; Garcia-Rogo, G.; Corrales, W.; Olave, F.A.; Aliaga, E.; Ulloa, J.; Avalos, A.M.; et al. Hippocampal memory recovery after acute stress: Behavioral, morphological and molecular study. *Front. Mol. Neurosci.* **2018**, *11*, 283. [[CrossRef](#)]
100. Uysal, N.; Sisman, A.R.; Dayi, A.; Ozbal, S.; Cetin, F.; Baykara, B.; Aksu, I.; Tas, A.; Cavus, S.A.; Gonenc-Arda, S.; et al. Acute foot-shock-stress increases spatial learning-memory and correlates to increased hippocampal BDNF and VEGF and cell numbers in adolescent male and female rats. *Neurosci. Lett.* **2012**, *514*, 141–146. [[CrossRef](#)] [[PubMed](#)]
101. Shizadian, A.; Ostadhadi, S.; Hassanipour, M.; Shafaroodi, H.; Khoshnoodi, M.; Haj-Mirzaian, A.; Sharifzadeh, M.; Amiri, S.; Ghasemi, M.; Dehpour, A.R. Acute foot-shock stress decreased seizure susceptibility against pentylentetrazole-induced seizures in mice: Interaction between endogenous. *Epilepsy Behav.* **2018**, *87*, 25–31. [[CrossRef](#)] [[PubMed](#)]
102. Greenfield, L.J., Jr. Molecular mechanisms of antiseizure drug activity at GABA_A receptor. *Seizure* **2013**, *22*, 589–600. [[CrossRef](#)] [[PubMed](#)]
103. Jankovic, S.M.; Djesevic, M.; Jankovic, S.V. Experimental GABA_A receptor agonists and allosteric modulators for the treatment of focal epilepsy. *J. Exp. Pharmacol.* **2021**, *13*, 235–244. [[CrossRef](#)] [[PubMed](#)]
104. Fritschy, J.M.; Kiener, T.; Bouillere, V.; Loup, F. GABAergic neurons and GABA_A-receptors in temporal lobe epilepsy. *Neurochem. Int.* **1999**, *34*, 435–445. [[CrossRef](#)]
105. Stamboulian-Platel, S.; Legendre, A.; Chabrol, T.; Platel, J.C.; Pernot, F.; Duveau, V.; Roucard, C.; Baudry, M.; Depaulis, A. Activation of GABA_A receptors controls mesiotemporal lobe epilepsy despite changes in chloride transporters expression: In vivo and in silico approach. *Exp. Neurol.* **2016**, *284* (Pt A), 11–28. [[CrossRef](#)]
106. Brophy, G.M.; Bell, R.; Claassen, J.; Alldredge, B.; Bleck, T.P.; Glauser, T.; Laroche, S.M.; Riviello, J., Jr.; Shutter, L.; Sperling, M.R.; et al. Guidelines for the evaluation and management of status epilepticus. *Neurocrit Care.* **2012**, *17*, 3–23. [[CrossRef](#)]
107. Wilkes, R.; Tasker, R.C. Pediatric intensive care treatment of uncontrolled status epilepticus. *Crit. Care Clin.* **2013**, *29*, 239–257. [[CrossRef](#)]
108. Olf, M. Sex and gender differences in post-traumatic stress disorder: An update. *Eur. J. Psychotraumatol.* **2017**, *8*, 1351204. [[CrossRef](#)]
109. Teicher, M.; Samson, J.A.; Anderson, C.M.; Ohashi, K. The effects of childhood maltreatment on brain structure, function and connectivity. *Nat. Rev. Neurosci.* **2016**, *17*, 652–666. [[CrossRef](#)]
110. Lu, C.Y.; Liu, D.X.; Jiang, H.; Pan, F.; Ho, C.S.H.; Ho, R.C.M. Effects of traumatic stress induced in the juvenile period on the expression of GABA_A receptor subunits in adult rat brain. *Neural Plast.* **2017**, *2017*, 5715816. [[CrossRef](#)] [[PubMed](#)]
111. Girgenti, M.J.; Wang, J.; Ji, D.; Cruz, D.A.; Traumatic Stress Brain Research Group; Stein, M.B.; Gelernter, J.; Young, K.A.; Huber, B.R.; Williamson, D.E.; et al. Transcriptomic organization of the human brain in post-traumatic stress disorder. *Nat. Neurosci.* **2021**, *24*, 24–33. [[CrossRef](#)] [[PubMed](#)]
112. Kavushansky, A.; Vouimba, R.M.; Choen, H.; Richter-Levin, G. Activity and plasticity in the CA1, the dentate gyrus, and the amygdala following controllable vs. uncontrollable water stress. *Hippocampus* **2006**, *16*, 35–42. [[CrossRef](#)] [[PubMed](#)]
113. Sharvit, A.; Segal, M.; Kehat, O.; Stork, O.; Richter-Levin, G. Differential modulation of synaptic plasticity and local circuit activity in the dentate gyrus and CA1 regions of the rat hippocampus by corticosterone. *Stress* **2015**, *18*, 319–327. [[CrossRef](#)]
114. Zhou, H.; Xiong, G.-J.; Jing, L.; Song, N.-N.; Pu, D.-L.; Tang, X.; He, X.-B.; Xu, F.-Q.; Huang, J.-F.; Li, L.-J.; et al. The interhemispheric CA1 circuit governs rapid generalization but not fear memory. *Nat. Commun.* **2017**, *8*, 2190. [[CrossRef](#)] [[PubMed](#)]
115. Vouimba, R.M.; Anunu, R.; Richter-Levin, G. GABAergic transmission in the basolateral amygdala differentially modulates plasticity in the dentate gyrus and the CA1 areas. *Int. J. Mol. Sci.* **2020**, *21*, 3786. [[CrossRef](#)] [[PubMed](#)]
116. Tripathi, K.; Demiray, Y.E.; Kliche, S.; Jing, L.; Hazra, S.; Hazra, J.D.; Richter-Levin, G.; Stork, O. Reducing glutamic acid decarboxylase in the dorsal dentate gyrus attenuates juvenile stress induced emotional and cognitive deficits. *Neurobiol. Stress* **2021**, *15*, 100350. [[CrossRef](#)] [[PubMed](#)]
117. Torrisi, S.A.; Lavanco, G.; Maurel, O.M.; Gulisano, W.; Laudani, S.; Geraci, F.; Grasso, M.; Barbagallo, C.; Caraci, F.; Bucolo, C.; et al. A novel arousal-based individual screening reveals susceptibility and resilience to PTSD-like phenotypes in mice. *Neurobiol. Stress* **2020**, *14*, 100286. [[CrossRef](#)] [[PubMed](#)]
118. Ardi, Z.; Richter-Levin, A.; Xu, L.; Cao, X.; Volkmer, H.; Stork, O.; Richter-Levin, G. The role of GABA_A receptor α 1 subunit in the ventral hippocampus in stress resilience. *Sci. Rep.* **2019**, *9*, 13513. [[CrossRef](#)] [[PubMed](#)]
119. Barnard, E.A.; Skolnick, P.; Olsen, R.W.; Mohler, H.; Sieghart, W.; Biggio, G.; Braestrup, C.; Bateson, A.N.; Langer, S.A. International union of pharmacology. XV. Subtypes of GABA_A receptor: Classification on the basis of subunit structure and receptor function. *Pharmacol. Rev.* **1998**, *50*, 291–314.
120. Geuze, E.; van Berchel, B.N.M.; Lammertsma, A.A.; Boellaard, R.; de Kloet, C.S.; Vermetten, E.; Westenberg, H.G.M. Reduced GABA_A benzodiazepine receptor binding in veterans with post-traumatic stress disorder. *Mol. Psychiatry* **2008**, *13*, 74–83. [[CrossRef](#)]
121. Feusner, J.; Ritchie, T.; Lawford, B.; Young, R.; Kann, B.; Noble, E.P. GABA_A receptor β 3 subunit gene and psychiatric morbidity in a post-traumatic stress disorder population. *Psychiatry Res.* **2001**, *104*, 109–117. [[CrossRef](#)]