

Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.

European Journal of Medicinal Chemistry 228 (2022) 114030

Contents lists available at ScienceDirect

ELSEVIER



Discovery of 9,10-dihydrophenanthrene derivatives as SARS-CoV-2 3CL^{pro} inhibitors for treating COVID-19



Jian-Wei Zhang ^{a, 1}, Yuan Xiong ^{b, 1}, Feng Wang ^a, Fu-Mao Zhang ^a, Xiaodi Yang ^a, Guo-Qiang Lin ^a, Ping Tian ^{a, **}, Guangbo Ge ^{b, ***}, Dingding Gao ^{a, *}

^a Shanghai Frontiers Science Center of TCM Chemical Biology, Innovation Research Institute of Traditional Chinese Medicine, Shanghai University of Traditional Chinese Medicine, Shanghai, 201203, China

^b Shanghai Frontiers Science Center of TCM Chemical Biology, Institute of Interdisciplinary Integrative Medicine Research, Shanghai University of Traditional Chinese Medicine, Shanghai, 201203, China

A R T I C L E I N F O

Article history: Received 10 October 2021 Received in revised form 22 November 2021 Accepted 27 November 2021 Available online 1 December 2021

Keywords: COVID-19 SARS-CoV-2 3CL^{pro} 9,10-Dihydrophenanthrenes Structure-activity relationships

ABSTRACT

The epidemic coronavirus disease 2019 (COVID-19) caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) has now spread worldwide and efficacious therapeutics are urgently needed. 3-Chymotrypsin-like cysteine protease ($3CL^{pro}$) is an indispensable protein in viral replication and represents an attractive drug target for fighting COVID-19. Herein, we report the discovery of 9,10-dihydrophenanthrene derivatives as non-peptidomimetic and non-covalent inhibitors of the SARS-CoV-2 $3CL^{pro}$. The structure-activity relationships of 9,10-dihydrophenanthrenes as SARS-CoV-2 $3CL^{pro}$ inhibitors have carefully been investigated and discussed in this study. Among all tested 9,10-dihydrophenanthrene derivatives, **C1** and **C2** display the most potent SARS-CoV-2 $3CL^{pro}$ inhibition activity, with IC₅₀ values of $1.55 \pm 0.21 \ \mu$ M and $1.81 \pm 0.17 \ \mu$ M, respectively. Further enzyme kinetics assays show that these two compounds dose-dependently inhibit SARS-CoV-2 $3CL^{pro} via$ a mixed-inhibition manner. Molecular docking simulations reveal the binding modes of **C1** in the dimer interface and substrate-binding pocket of the target. In addition, **C1** shows outstanding metabolic stability in the gastrointestinal tract, human plasma, and human liver microsome, suggesting that this agent has the potential to be developed as an orally administrated SARS-CoV-2 $3CL^{pro}$ inhibitor.

© 2021 Elsevier Masson SAS. All rights reserved.

1. Introduction

Since the emergence of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), the public around the world is plagued with Coronavirus Disease 2019 (COVID-19) [1,2]. Accordingly, many worldwide endeavors were made to effectively prevent or treat COVID-19, boosting several vaccines that were successfully administered applied to the public [3–5]. However, SARS-CoV-2 is constantly mutating for better survival in hosts [6]. For example, the B.1.617.2 (Delta) variant appears with a faster transmission capacity and stronger toxicity, and has become dominant in the current pandemic

E-mail addresses: tianping@shutcm.edu.cn (P. Tian), geguangbo@dicp.ac.cn (G. Ge), gaodingding@shutcm.edu.cn (D. Gao).

¹ These authors contributed equally to this work.

[7,8]. Nevertheless, the latest reports manifest that the effectiveness of COVID-19 vaccines is reduced against the Delta variant, and people who have been vaccinated are still facing the risk of being infected [9,10]. Until now, there are few effective antiviral medications to cure CoV-related diseases, such as COVID-19. We urgently need novel antiviral agents to fight against the ongoing SARS-CoV-2 and other unknown coronaviruses in the long term. At the same time, scientists are working against time to discover potential anti-SARS-CoV-2 drugs by targeting the key proteases to obstruct the viral life cycle [11,12]. 3-Chymotrypsin-like cysteine protease (3CL^{pro}), an indispensable tool for an intact coronaviral replication cycle, can convert the viral polyproteins into functional proteins [13]. Besides this important function, 3CL^{pros} share a highly conserved gene sequence among several pathogenic human coronaviruses (such as SARS-CoV-2, MERS-CoV, and SARS-CoV) [14]; accordingly, 3CL^{pro} is deemed to be an attractive target for developing broad-spectrum antiviral drugs. In addition, potent 3CL^{pro} inhibition can reduce the risk of mutation-mediated drug resistance [15].

^{*} Corresponding author.

^{**} Corresponding author.

^{***} Corresponding author.

Up to now, a variety of SARS-CoV-2 3CL^{pro} inhibitors have been developed using multiple drug discovery strategies, which can be divided into covalent inhibitors and non-covalent inhibitors according to their inhibitory mechanisms [16–18]. Covalent SARS-CoV-2 3CL^{pro} inhibitors, including peptidomimetics and small molecules, generally bear different covalent warheads and act through a two-step mechanism [19]. First, these agents bind to the active site and form a non-covalent complex with the target protease. Then, the warhead forms a covalent bond with the nucleophilic residues, especially catalytic Cys145 or other key cysteines (such as Cys300 [20] and Cys44 [20,21]), whose covalent modifications can further elicit the inactivation of SARS-CoV-2 3CL^{pro}. Several chemical warheads, such as Michael acceptor (compound **1**) [22], α-ketoamide (compound **2**) [23], aldehyde (compounds **3**) [24], α -hydroxy bisulfite (compound **4**, GC-376) [25], hydroxvmethyl ketone (compound 5, PF-00835231) [26], cyano (compound **6**, PF-07321332) [27], pyrogallol (compound **7**, myricetin) [20,28], quinone (compound 8, Vitamin K3) [29], and ester carbonyl group (compound **9**) have been explored by pharmaceutical companies or research groups (Fig. 1) [30]. Among these inhibitors, only compounds **5** and **6** from Pfizer Inc. entered the clinical trial stage. Additionally, some natural products or small molecules, such as sciadopitysin (compound 10) [31], 23R (compound 11) [32], CCF981 (compound 12) [33], and pyridine 13 [34], were found to have strong inhibitory activities against SARS-CoV-2 3CLpro via noncovalent modes. Candidates for use as anti-COVID-19 agents are expected to possess several positive traits, such as excellent pharmacokinetic property and good cell permeability, besides the superior biological activities. However, most of these peptidomimetics or natural products exhibit poor membrane permeability and poor oral bioavailability. Further, the potential offtarget issue of covalent inhibitors may lead to unpredictable toxic side effects clinically [19,35]. Thus, there is a strong desire for novel potent non-covalent SARS-CoV-2 3CL^{pro} inhibitors with drug-like properties.

Herein, we report the discovery of non-covalent SARS-CoV-2 3CL^{pro} inhibitors with 9,10-dihydrophenanthrene scaffold using an established fluorescence resonance energy transfer (FRET) biochemical assay. We also carefully carried out the structure-activity relationship (SAR) and inhibitory mechanism studies.

Furthermore, the *in vitro* metabolic stability in the gastrointestinal tract, human plasma, and liver microsome validated that the newly identified SARS-CoV-2 3CL^{pro} inhibitor **C1** has the potential to be developed into an orally administered agent.

2. Results and discussion

2.1. Screening and SARs analysis

To obtain potent SARS-CoV-2 3CL^{pro} inhibitors, we first screened our in-house compound library (compounds containing 9,10dihydrophenanthrene scaffold) using established fluorescence resonance energy transfer (FRET) assay at final concentrations of 1 μM, 10 μM, and 100 μM (Fig. 2) [31,36]. Briefly, the fluorescent substrate can be recognized and cleaved by 3CL^{pro}, and the products emit a strong fluorescence at 490 nm after being excited at 340 nm. Disulfiram (a reported SARS-CoV-2 3CL^{pro} inhibitor) was used as the positive control compound [22]. Of them, some compounds showed good inhibitory activities and were selected for further IC₅₀ values investigation. It was worth noting that the functional groups of our screened compounds varied at the positions R^1 , R^2 , and R^3 , which was because of the limitations of our invented synthetic methodology [37]. As shown in Table 1, we summarized the SARs of the tested 9.10-dihydrophenanthrene derivatives. For R¹ substituent, the activities against SARS-CoV-2 3CL^{pro} were improved along with an increasing volume of the substituted groups, such as methyl (A1, $IC_{50} = 61.15 \ \mu M$) to ethyl (A2, $IC_{50} = 33.06 \ \mu M$), isopropyl (A3, $IC_{50} = 29.46 \ \mu M$), cyclohexyl (**A4**, $IC_{50} = 9.06 \ \mu$ M), and 4-bromine phenyl (**A5**, $IC_{50} = 6.44 \ \mu$ M). When we kept increasing the size of the \mathbf{R}^1 group, such as 4- cyanophenyl (**A6**, $IC_{50} = 19.32 \ \mu M$) and benzyl (**A7**, $IC_{50} = 11.39 \ \mu M$), the inhibitory activities were decreased. For the \mathbf{R}^2 substituent, introducing methyl at C-2 or C-3 position generated compounds A13 and A14, which displayed inhibitory activities similar to A1 (A13, $IC_{50} = 57.67 \ \mu M$, A14, $IC_{50} = 53.81 \ \mu M$). The presence of substituent groups such as methyl (A8), methoxyl (A9), fluorine (A10), or bromine (A12) at the C-4 position led to a significant loss of potency. For the R³ substituent, the incorporation of 6-methyl (A15), 5-methyl (A16), or 4-fluorine (A18) at pyridyl group exhibited comparable inhibitory activities with A1. The incorporation of

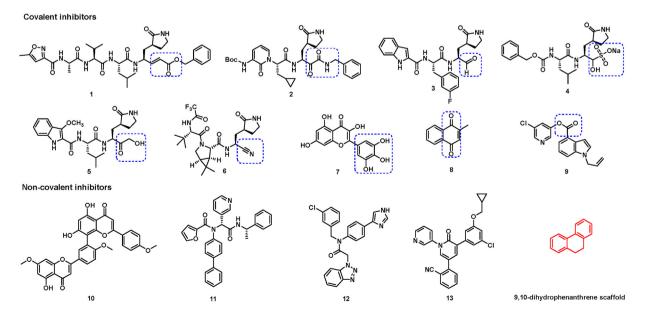


Fig. 1. Representative SARS-CoV-2 3CL^{pro} inhibitors. For covalent inhibitors, different covalent warheads are emphasized by dashed rectangles.

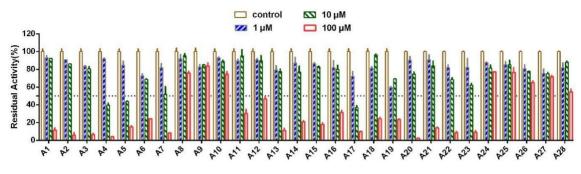


Fig. 2. Inhibitory effects of A1-A28 against SARS-CoV-2 3CL^{pro}.

4-chlorine (**A19**) or 4-bromine (**A20**) at pyridyl group or replacing pyridine with quinoline (**A21**, **A22**, **A23**) showed increased inhibitory activities. Compound **A17** with 5-phenyl at the pyridyl group displayed the best inhibitory activity with an IC₅₀ value of 5.65 μ M. In contrast, the substituents of pyrimidine (**A24**, **A25**), pyrazole (**A26**), and purine derivative (**A27**, **A28**) demonstrated no inhibitory activity against SARS-CoV-2 3CL^{pro}.

The abovementioned analyses suggested the preliminary SARs for the 9,10-dihydrophenanthrene derivatives that can be summarized as follows: (1) the installation of suitable bulkier groups at the R^1 position was preferred and, cyclohexyl and 4-Br phenyl displayed better inhibitory activities than others. (2) The conversion of the pyridine group into quinoline at the R^3 position was favorable, whereas pyrimidine, pyrazole, and purine derivatives at this position were unfavorable. (3) The incorporation of 5-phenyl at the pyridyl group was of great importance for the inhibition activity.

2.2. Rational design of SARS-CoV-2 3CL^{pro} inhibitors

The abovementioned structure–activity relationships showed that cyclohexyl and 4-Br phenyl groups at R¹ position (**A4** and **A5**) and 5-phenyl at the pyridyl group (**A17**) were optimal options for maintaining the SARS-CoV-2 3CL^{pro} inhibition activity. To improve the biological activity of the identified hit compounds, we maintained those privileged fragments and designed compounds **B1** and **B2** (as shown in Fig. 3). On the basis of the optimized 4-Br phenyl group, SARs of various substitutions at the pyridine ring were explored again (**B3–B20**). Furthermore, the ethyl ester group was hydrolyzed to investigate its effect on inhibitory activity (**C1–C7**).

2.3. Chemistry

The designed compounds were prepared according to our previously reported synthetic method, which is shown in Scheme 1 [37]. First, the substrates cyclohexadienone-containing 1,6-enynes **16** and **17** were prepared from commercially available 4-bromo-4'-hydroxybiphenyl and 4-cyclohexylphenol, respectively, using propargyl alcohol and phenyliodine (III) diacetate (PIDA). In addition, 2-arylazaarenes **18a-18v** were purchased directly from commercial companies or generated *via* a Suzuki cross-coupling reaction with the use of phenylboronic acid and substituted 2bromopyridine. Subsequently, an efficient and one-pot synthesis of 9,10-dihydrophenanthrenes from **16**, **17**, and **18a-18v** was conducted using rhodium (III)-catalyzed C–H activation and relay Diels–Alder reaction to obtain target compounds **B1–B20**. Subsequently, compounds **B2**, **B6**, **B10**, **B11**, **B12**, **B14**, and **B18** were hydrolyzed under 1 M NaOH and CH₃OH solution to afford **C1–C7**.

2.4. Biological assays

2.4.1. SARS-CoV-2 3CL^{pro} inhibition assays

To measure the inhibitory activity of our designed compounds against SARS-CoV-2 3CL^{pro}, a fluorescence resonance energy transfer (FRET) protease assay was performed as described previously [31,36]. As seen in Table 2, compound B2 with the privileged fragment 4-Br phenyl group at the R¹ position and 5-phenyl at the R^2 position could improve the activity as expected with an IC_{50} value of 2.46 µM. However, compound **B1** with a cyclohexyl at the R^1 position and 5-phenyl at the R^2 position failed to enhance the activity with an IC_{50} value of 8.54 $\mu M.$ Therefore, we fixed 4-Br phenyl group at the R¹ position and changed the group at pyridine region to examine the influence on the inhibitory activity. When the substituent groups, such as methyl (**B3**), chlorine (**B4**), cyano (B5), aldehyde (B6), N, N-dimethyl (B7), pyrrole (B8), and morpholine (**B9**), were located at the C-5 position of the pyridine ring, compounds B4, B6, and B9 displayed better inhibitory activities with IC₅₀ values of 4.79 μ M, 3.32 μ M, and 7.39 μ M, respectively. Other compounds exhibited moderate activities with IC₅₀ values more than 10 µM. Incorporation of the substituent groups, such as methyl (B10), fluorine (B11) chlorine (B12), bromine (B13), cyano (B14), aldehyde (B15), methylsulfonyl (B16), acetyl (B17), N, Ndimethyl (B18), phenyl (B19), and N-benzyl (B20) at the C-4 position, all the compounds demonstrated potent activities with IC₅₀ values ranging from 2.72 μ M to 8.89 μ M. To further explore the antiviral activity of the ethyl ester group, compounds **B2**, **B6**, **B10**, B11, B12, B14, and B18 with better activity described above were hydrolyzed to the provided compounds C1–C7, containing a typical hydroxyl pharmacophore. Interestingly, only compounds C1 and C2 showed a slight increase in the inhibitory activity with IC₅₀ values of 1.55 µM and 1.81 µM, respectively (Table 3). However, compounds C4, C5, and C7 led to slight decreases in the inhibitory activity, whereas C6 with a 4-cyano group resulted in a substantial decrease of the activity (IC₅₀ = 10.68 μ M). Taken together, compounds C1 and C2 showed better in vitro inhibitory activities against SARS-CoV-2 3CL^{pro} and were selected for further evaluation.

2.4.2. Inhibition kinetics assays of **C1** and **C2** against SARS-CoV-2 $3CL^{pro}$

To uncover the inhibitory mechanisms of two newly identified SARS-CoV-2 3CL^{pro} inhibitors (**C1** and **C2**), the inhibition kinetics assays of these two agents were carefully evaluated *in vitro*. As shown in Fig. 4, the Lineweaver–Burk plots showed that both **C1** and **C2** could dose-dependently inhibit SARS-CoV-2 3CL^{pro} *via* a mixed-inhibition manner, with K_i values of 6.09 µM and 7.64 µM, respectively. In addition, the corresponding second plot of the Lineweaver–Burk plots were presented in Fig. S4, showing good linearity ($\mathbb{R}^2 > 0.95$).

 Table 1

 Inhibitory activities of 9,10-dihydrophenanthrene derivatives against SARS-CoV-2 3CL^{pro}.



Compds	R ¹	R ²	R ³	$IC_{50} \pm SD (\mu M)^a$	
A1	CH3	Н	Ň	61.15 ± 3.60	
12	C_2H_5	Н		33.06 ± 8.58	
13	isopropyl	Н		29.46 ± 1.64	
14	cyclohexyl	Н	N N	9.06 ± 0.34	
15	4-Br Phenyl	Н	Ň	6.44 ± 0.64	
16	4-CN Phenyl	Н		19.32 ± 1.92	
47	benzyl	Н	Ň	11.39 ± 0.64	
18	CH ₃	4-CH ₃	Ň	>100	
19	CH₃	4-0CH ₃		>100	
410	CH ₃	4-F	Ň	>100	
111	CH ₃	4-Cl	Ň	85.58 ± 2.36	
112	CH ₃	4-Br	Ň	>100	
413	CH ₃	3-CH ₃		57.67 ± 2.78	
A14	CH ₃	2-CH ₃	× ×	53.81 ± 8.49	
15	CH ₃	Н	1)-6 2N_55	58.91 ± 4.58	
A16	CH ₃	Н	$2N \xrightarrow{1}_{3} \xrightarrow{6}_{4} 5$ $N \xrightarrow{1}_{3} \xrightarrow{6}_{4} 5$	68.16 ± 2.64	
417	CH ₃	Н		5.65 ± 1.40	
A18	CH ₃	Н		66.80 ± 2.74	

Table 1 (continued)

		$ \begin{array}{c} OH & 4 \\ OH & 4 \\ R^3 \\ R^3 \\ OAc \end{array} $		
Compds	R ¹	R ²	R ³	$IC_{50} \pm SD (\mu M)^a$
A19	CH ₃	Н	Ň	35.77 ± 2.96
A20	CH ₃	Н	N N	18.50 ± 0.76
A21	CH ₃	Н	Br	14.17 ± 1.30
A22	CH ₃	Н	Ň	11.97 ± 0.68
A23	CH3	н	Ň	13.82 ± 0.90
A24	CH ₃	Н		>100
A25	CH ₃	Н		>100
A26	CH ₃	Н	Ň	>100
A27	CH ₃	Н		>100
A28	CH ₃	Н		>100
disulfiram	_	_	— N N	1.04 ± 0.03

^a The inhibitory effects of these compounds against SARS-CoV-2 3CLpro were determined by the FRET assay. The data are the mean ± SD from at least three independent experiments.

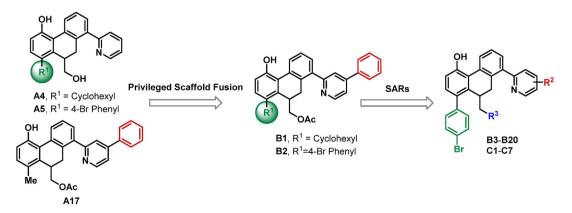
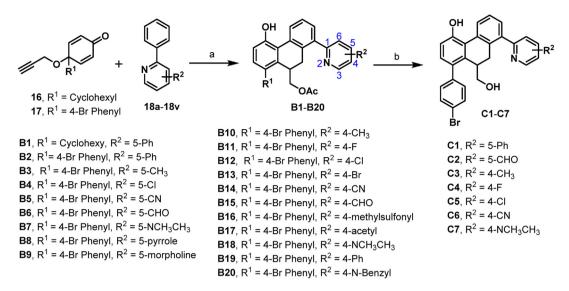
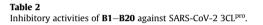


Fig. 3. Rational design of SARS-CoV-2 3CL^{pro} inhibitors by privileged scaffold fusion and structure-activity relationships study.



Scheme 1. Reagents and conditions: (a) [Cp*RhCl2]2, NaBArF4, CH3COOH, 100 °C, 8 h, 20.0%-87.0%; (b) 1 M NaOH (aq.), CH3OH, 50 °C, 2 h, 37.0%-99.0%.





Compds	R ¹	R ²	$IC_{50} \pm SD (\mu M)^a$
B1	Cyclohexyl	5-Ph	8.54 ± 0.94
B2	4-Br Phenyl	5-Ph	2.46 ± 0.14
B3	4-Br Phenyl	5-CH ₃	9.82 ± 1.16
B4	4-Br Phenyl	5-Cl	4.79 ± 0.22
B5	4-Br Phenyl	5-CN	10.26 ± 0.66
B6	4-Br Phenyl	5-CHO	3.32 ± 0.25
B7	4-Br Phenyl	5-NCH ₃ CH ₃	10.00 ± 0.47
B8	4-Br Phenyl	~h	11.23 ± 0.70
		5 N	
B9	4-Br Phenyl	5	7.39 ± 0.11
B10	4-Br Phenyl	4-CH ₃	5.25 ± 0.24
B11	4-Br Phenyl	4-F	4.29 ± 0.62
B12	4-Br Phenyl	4-Cl	2.72 ± 0.23
B13	4-Br Phenyl	4-Br	6.73 ± 1.29
B14	4-Br Phenyl	4-CN	2.78 ± 0.18
B15	4-Br Phenyl	4-CHO	8.89 ± 3.34
B16	4-Br Phenyl	4-methylsulfonyl	6.65 ± 0.26
B17	4-Br Phenyl	4-acetyl	5.83 ± 0.34
B18	4-Br Phenyl	4-NCH ₃ CH ₃	3.66 ± 0.17
B19	4-Br Phenyl	4-Ph	8.28 ± 1.88
B20	4-Br Phenyl	4-N-Benzyl	3.31 ± 0.36
disulfiram	_	-	1.04 ± 0.03

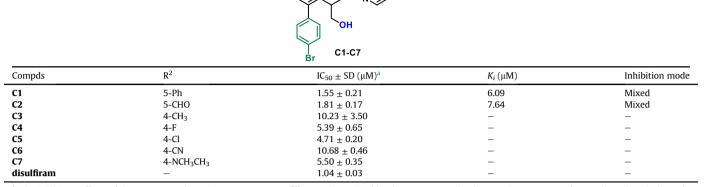
^a The inhibitory effects of these compounds against SARS-CoV-2 3CL^{pro} were determined by the FRET assay. The data are the mean ± SD from at least three independent experiments.

2.4.3. SARS-CoV 3CL^{pro} inhibition assays

To verify whether these series compounds have a broadspectrum inhibitory effect on $3CL^{pro}$, we obtained highly purified SARS-CoV $3CL^{pro}$ according to the previously reported expression and purification procedures of SARS-CoV-2 $3CL^{pro}$ [31]. Subsequently, 11 compounds that exhibited potent inhibitory effects ($IC_{50} < 5 \mu M$) against SARS-CoV-2 $3CL^{pro}$ were chosen for SARS-CoV 3CL^{pro} inhibition assay. It is clearly shown in Table 4 and Fig. S8 that most of the tested compounds show relatively strong inhibitory activities against SARS-CoV 3CL^{pro}, with IC₅₀ values less than 10 μ M. These results suggest that the 9,10-dihydrophenanthrene derivatives can easily be developed as broad-spectrum 3CL^{pro} inhibitors for the treatment of coronavirus-related diseases.

Table 3

Inhibitory activities of C1–C7 against SARS-CoV-2 3CL^{pro}.



∩н

^a The inhibitory effects of these compounds against SARS-CoV-2 3CL^{pro} were determined by the FRET assay. The data are the mean ± SD from at least three independent experiments.

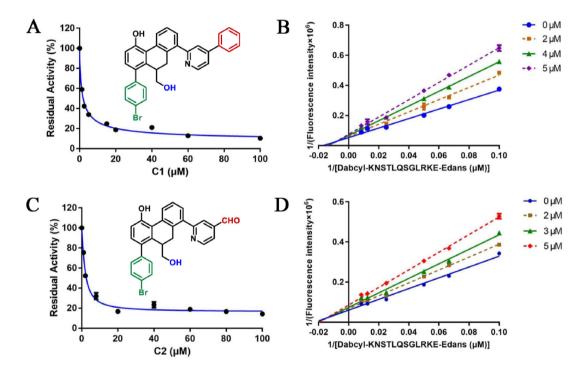


Fig. 4. The dose-dependent inhibition curves (left) and the second plot of Lineweaver-Burk plots (right) for C1 (A, B) and C2 (C, D) against SARS-CoV-2 3CL^{pro}.

2.5. Molecular docking simulations

As **C1** inhibits SARS-CoV-2 3CL^{pro} via mixed inhibition, which means this small molecule could bind at least two sites in the target protein. Recent studies indicated that the dimerization of SARS-CoV-2 3CL^{pro} was crucial to enzymatic activity, the monomers were catalytically inactive [38]. Therefore, there are at least two potential sites that can be employed to develop inhibitors against this enzyme: the substrate-binding pocket and the dimer interface of SARS-CoV-2 3CL^{pro}, both of whose key interactions with the inhibitors may affect the catalytic activity. After failing to obtain the crystal structure of **C1**-SARS-CoV-2-3CL^{pro} complex, the molecular docking study was conducted to predict the binding modes at the substrate binding pocket and the dimer interface (Fig. 5A).

For the dimer interface position (Fig. 5B), the phenolic hydroxyl group of **C1** formed a hydrogen bond with the crucial residue Glu290, which may disrupt the salt bridge interaction with Arg4 in another monomer. Furthermore, a benzene ring generated a cation- π interaction with Lys5. Meanwhile, the binding affinity of **C1** was enhanced through some hydrophobic interactions with Phe3 and Tyr126 residues. For the substrate-binding pocket (Fig. 5C and D), the 4-bromine phenyl moiety deeply inserted into the big S2 site, which was usually large enough to accommodate the larger hydrophobic fragment. The phenol group occupied the S1' site and was stacked with the imidazole ring of His41. Notably, this hydroxyl group formed two hydrogen bonds with Gly143 and Cys145. The 5-phenyl group at the pyridine ring fitted well into the S4 pocket and formed multiple hydrophobic interactions with the target. In

J.-W. Zhang, Y. Xiong, F. Wang et al.

Table 4

Compds	R ¹	R ²	$IC_{50}\pm SD~(\mu M)^a$
B2	4-Br Phenyl	5-Ph	17.66 ± 1.42
B4	4-Br Phenyl	5-Cl	5.44 ± 0.36
B6	4-Br Phenyl	5-CHO	3.20 ± 0.25
B11	4-Br Phenyl	4-F	7.81 ± 0.51
B12	4-Br Phenyl	4-Cl	4.06 ± 0.17
B14	4-Br Phenyl	4-CN	7.12 ± 0.32
B18	4-Br Phenyl	4-NCH ₃ CH ₃	5.92 ± 0.22
B20	4-Br Phenyl	4-Benzyl	5.79 ± 0.35
C1	4-Br Phenyl	5-Ph	5.35 ± 0.30
C2	4-Br Phenyl	5-CHO	5.32 ± 0.22
C5	4-Br Phenyl	4-Cl	10.49 ± 0.23
Disulfiram	_	_	6.72 ± 1.02

^a The inhibitory effects of these compounds against SARS-CoV 3CL^{pro} were determined by the FRET assay. The data are the mean \pm SD from at least three independent experiments.

addition, the hydroxymethyl group could form a 2.3 Å hydrogen bond interaction with Gln189, which was consistent with the observation that the hydroxyl group worked better than ethyl ester group (**B2** *vs* **C1**). In short, these interactions between **C1** and the target are responsible for its potent SARS-CoV-2 3CL^{pro} inhibitory activity.

2.6. In vitro gastrointestinal, plasma, and microsome stability assays

Next, the potential of compounds **C1** and **C2** as orally administered agents was investigated by performing gastrointestinal stability and metabolic stability assays. Firstly, the gastrointestinal stabilities of **C1** and **C2** in simulated gastric fluid (SGF) and simulated intestinal fluid (SIF) systems were assayed. As presented in Fig. 6, both **C1** and **C2** showed good gastrointestinal stabilities in 90 min. Moreover, the plasma stability of **C1** was also prominent,

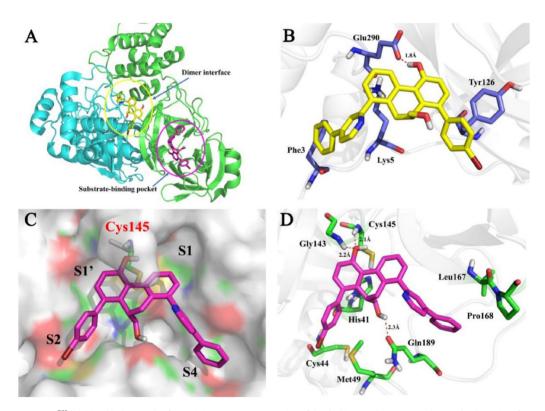


Fig. 5. Predicted SARS-CoV-2 3CL^{pro} inhibitor binding modes for C1. (A) Cartoon representation of the docking results. Compound C1 at the dimer interface position is shown as a yellow stick; Compound C1 at substrate-binding pocket is shown as a magenta stick; (B) Predicted interactions of C1 at the dimer interface position; (C) Surface representation of C1 in the substrate-binding pocket. Four subsites, S1', S1, S2, and S4, are labeled; (D) Predicted interactions of C1 at the substrate-binding pocket. Hydrogen bonds are indicated as dashed lines.

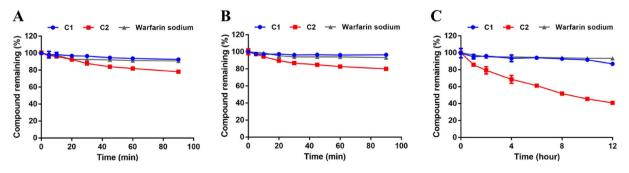


Fig. 6. (A-B) The gastrointestinal stabilities of C1 and C2 in SGF (A) and SIF (B); (C) The plasma stabilities of C1 and C2 in human blood plasma.

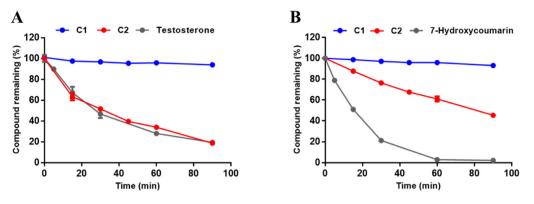


Fig. 7. Metabolic stabilities of C1 and C2 in phase I (A) and phase II (B) metabolic system in human liver microsomes.

Table 5

The half-life of C1 and C2 in phase I or phase II metabolic system in human liver microsomes.

Compound	$T_{1/2}$ (min)	
	phase I	phase II
C1	>90	>90
C2	32.09	81.88
Testosterone	27.62	ND
7-Hydroxycoumarin	ND	15.53
	C1 C2 Testosterone	C1 >90 C2 32.09 Testosterone 27.62

^a ND = Not detected.

whereas the half-life of C2 in human plasma was 8.97 h.

Meanwhile, the in vitro metabolic stability assays for compounds C1 and C2 in human liver microsome were also investigated and the results are shown in Fig. 7 and Table 5. Notably, C1 was hardly metabolized by the human cytochrome P450 enzymes (hCYPs) or the human hepatic UDP-Glucuronosyltransferases (hUGTs) in the presence of sugar donor UDPGA (Uridine-Diphosphate-Glucuronic Acid trisodium salt). In contrast, C2 could be readily metabolized by hCYPs and moderately metabolized by hUGTs, with the in vitro half-life of 32.09 min (in the phase I metabolism system) and 81.88 min (in the phase II metabolism system). Moreover, as shown in Fig. S11, C1 showed moderate inhibitory effects against most CYPs except for CYP2C8 at 100 µM. These results indicate that C1 possessed excellent plasma stability and good gastrointestinal and metabolic stability, suggesting that this agent has a great potential as a novel orally administered agent to fight against COVID-19 via targeting the key SARS-CoV-2-3CL^{pro}.

3. Conclusions

In summary, this study reported the discovery of 9.10dihydrophenanthrene derivatives as potent noncovalent SARS-CoV-2 3CL^{pro} inhibitors for the treatment of COVID-19. Systematic SAR explorations resulted in the discovery of two efficacious lead compounds (C1 and C2) for further development. Enzyme kinetic analyses revealed that both C1 and C2 dose-dependently inhibited SARS-CoV-2 3CL^{pro} through a mixed-inhibition manner. Molecular docking simulations elucidated the possible binding mode of **C1** in the substrate-binding pocket and the dimer interface of the target. Furthermore, compound C1 showed good metabolic stability in the human gastrointestinal tract, plasma, and liver microsomes, which was a great advantage compared with covalent peptides or peptidomimetics. We hope that this work can provide more structural reference for the development of SARS-CoV-2 3CL^{pro} inhibitors for combating this rapidly evolving virus and further in-depth research will be reported in due course.

4. Experimental section

4.1. Chemistry

The reagents and solvents were purchased from commercial companies, such as Adamas-beta®, Bide pharmatech etc., and used without further purification. The nonaqueous reactions were performed under nitrogen atmosphere. The reactions were monitored by Shimadzu 8040 quadrupole LC/MS system or TLC and terminated as judged by the consumption of the starting material. Flash chromatography was performed on silica gel (200–300 mesh) using Sepa-Bean® machine and visualized under UV light monitor. ¹HNMR (600 MHz) and ¹³CNMR (150 MHz) spectra were measured on Bruker 600 MHz spectrometer using TMS as internal standard at ambient temperature. The chemical shifts (δ) were described as parts per million (ppm) downfield and coupling constants (*J*) values were expressed as hertz (Hz). High-resolution mass spectra (HRMS) data were reported by Agilent 6545 Accurate-Mass QTOF LC/MS system.

4.1.1. General procedures for the synthesis of compounds **18a-18e**, **18j**, **18k**, **18m-18p**, **18r**, **18s** [39]

Reactions were carried out with Phenylboronic acid (146 mg, 1.2 mmol), 2-bromo-pyridine derivatives (234 mg, 1.0 mmol, 2bromo-4-phenylpyridine for 18a; 172 mg, 1.0 mmol, 2-bromo-4methylpyridine for 18b; 192 mg, 1.0 mmol, 2-bromo-4chloropyridine for 18c; 183 mg, 1.0 mmol, 2-bromoisonicotinonitrile for 18d; 186 mg, 1.0 mmol, 2-bromoisonicotinaldehyde for 18e; 176 mg, 1.0 mmol, 2-bromo-5-fluoropyridine for 18j; 192 mg, 1.0 mmol, 2-bromo-5-chloropyridine for 18k; 183 mg, 1.0 mmol, 6bromonicotinonitrile for 18m; 186 mg, 1.0 mmol, 6bromonicotinaldehyde for 18n; 236 mg, 1.0 mmol, 2-bromo-5-(methylsulfonyl)pyridine for 180, 200 mg, 1.0 mmol, 1-(6bromopyridin-3-yl)ethan-1-one for 18p, 234 mg, 1.0 mmol, 2bromo-5-phenylpyridine for 18r; 263 mg, 1.0 mmol, N-benzyl-6bromopyridin-3-amine for 18s), Pd(PPh₃)₂Cl₂ (3.5 mg, 0.5 mol %), K₂CO₃ (387 mg, 2.8 mmol) in DME/H₂O (8 mL, 3:1) under nitrogen atmosphere. The mixture was stirred at 80 °C for about 14 h and the completion of the reaction was monitored by TLC. After cooling to room temperature, the mixture was extracted with ethyl acetate. The combined organic layers were washed with brine and the organic phase was dried with anhydrous sodium sulfate and concentrated in vacuo. The crude compound was purified by silica gel column chromatography to give corresponding intermediates.

4.1.1.1 2,4-*diphenylpyridine* (**18***a*). Colorless oil, 57.0% yield. ¹H NMR (600 MHz, CDCl₃) δ 8.71 (d, *J* = 5.1 Hz, 1H), 8.04 (d, *J* = 7.5 Hz, 2H), 7.90 (s, 1H), 7.65 (d, *J* = 7.5 Hz, 2H), 7.49–7.45 (m, 4H), 7.43 (d, *J* = 7.4 Hz, 1H), 7.40 (d, *J* = 6.8 Hz, 2H). ESI-HRMS [M + H]⁺ calcd for C₁₇H₁₄N: 232.1121, found: 232.1131.

4.1.1.2. 4-methyl-2-phenylpyridine (**18b**). White solid, 42.0% yield. ¹H NMR (600 MHz, CDCl₃) δ 8.55 (d, J = 5.0 Hz, 1H), 7.97 (d, J = 7.4 Hz, 2H), 7.55 (s, 1H), 7.47 (t, J = 7.6 Hz, 2H), 7.40 (t, J = 7.3 Hz, 1H), 7.06 (d, J = 4.9 Hz, 1H), 2.42 (s, 3H). ESI-HRMS [M + H]⁺ calcd for C₁₂H₁₂N: 170.0964, found: 170.0965.

4.1.1.3. 4-*chloro-2-phenylpyridine* (**18***c*). Colorless oil, 54.0% yield. ¹H NMR (600 MHz, CDCl₃) δ 8.57 (d, J = 5.1 Hz, 1H), 7.96 (d, J = 7.3 Hz, 2H), 7.72 (s, 1H), 7.49–7.42 (m, 3H), 7.23 (d, J = 3.6 Hz, 1H). ESI-HRMS [M + H]⁺ calcd for C₁₁H₉ClN: 190.0418, found: 190.0421.

4.1.1.4. 2-Phenylisonicotinonitrile (**18d**). White solid, 77.8% yield. ¹H NMR (600 MHz, CDCl₃) δ 8.78 (d, J = 4.3 Hz, 1H), 7.92 (dd, J = 8.1, 1.4 Hz, 2H), 7.86 (s, 1H), 7.45–7.39 (m, 3H), 7.37 (dd, J = 4.9, 1.3 Hz, 1H). ESI-HRMS [M + H]⁺ calcd for C₁₂H₉N₂: 181.0760, found: 181.0772.

4.1.1.5. 2-Phenylisonicotinaldehyde (**18e**). Colorless oil, 95.0% yield. ¹H NMR (600 MHz, CDCl₃) δ 10.02 (s, 1H), 8.83 (d, J = 4.8 Hz, 1H), 8.02 (s, 1H), 7.96 (d, J = 7.3 Hz, 2H), 7.52 (dd, J = 4.8, 1.1 Hz, 1H), 7.41 (t, J = 7.4 Hz, 2H), 7.37 (d, J = 7.2 Hz, 1H). ESI-HRMS [M + H]⁺ calcd for C₁₂H₁₀NO: 184.0757, found: 184.0758.

4.1.1.6. 4-fluoro-2-phenylpyridine (**18***j*). White solid, 82.9% yield. ¹H NMR (600 MHz, CDCl₃) δ 8.46 (d, J = 2.9 Hz, 1H), 7.85 (d, J = 7.4 Hz, 2H), 7.63 (dd, J = 8.7, 4.3 Hz, 1H), 7.42–7.36 (m, 3H), 7.33 (t, J = 7.3 Hz, 1H). ESI-HRMS [M + H]⁺ calcd for C₁₁H₉FN: 174.0714, found: 174.0716.

4.1.1.7. 5-*chloro-2-phenylpyridine* (**18***k*). Colorless oil, 82.0% yield. ¹H NMR (600 MHz, CDCl₃) δ 8.56 (d, J = 2.4 Hz, 1H), 7.89–7.86 (m, 2H), 7.64 (dd, J = 8.5, 2.5 Hz, 1H), 7.59 (d, J = 8.5 Hz, 1H), 7.40 (t, J = 7.5 Hz, 2H), 7.36–7.33 (m, 1H). ESI-HRMS [M + H]⁺ calcd for C₁₁H₉NCl: 190.0418, found: 190.0420.

4.1.1.8. 5-*Phenylnicotinonitrile* (**18m**). White solid, 93.4% yield. ¹H NMR (600 MHz, CDCl₃) δ 8.94 (dd, J = 2.1, 0.7 Hz, 1H), 8.04 (dd, J = 7.9, 1.7 Hz, 2H), 8.01 (dd, J = 8.3, 2.2 Hz, 1H), 7.85 (dd, J = 8.3, 0.8 Hz, 1H), 7.53–7.50 (m, 3H). ESI-HRMS [M + H]⁺ calcd for C₁₂H₉N₂: 181.0760, found: 181.0763.

4.1.1.9. 5-*Phenylnicotinaldehyde* (**18n**). White solid, 79.1% yield. ¹H NMR (600 MHz, CDCl₃) δ 10.16 (s, 1H), 9.15 (d, *J* = 1.9 Hz, 1H), 8.26 (dd, *J* = 8.2, 2.2 Hz, 1H), 8.11 (dd, *J* = 8.0, 1.5 Hz, 2H), 7.93 (d, *J* = 8.2 Hz, 1H), 7.56–7.51 (m, 3H). ESI-HRMS [M + H]⁺ calcd for C₁₂H₁₀NO: 184.0757, found: 184.0765.

4.1.1.10. 4-(methylsulfonyl)-2-phenylpyridine (**180**). White solid, 99.0% yield. ¹H NMR (600 MHz, CDCl₃) δ 9.20 (d, J = 2.3 Hz, 1H), 8.27 (dd, J = 8.4, 2.4 Hz, 1H), 8.07 (dd, J = 7.7, 1.9 Hz, 2H), 7.92 (d, J = 8.4 Hz, 1H), 7.55–7.50 (m, 3H), 3.15 (s, 3H). ESI-HRMS [M + H]⁺ calcd for C₁₂H₁₂NO₂S: 234.0583, found: 234.0591.

4.1.1.11. 1-(6-phenylpyridin-3-yl)ethan-1-one (**18p**). White solid, 75.0% yield. ¹H NMR (600 MHz, CDCl₃) δ 9.26 (d, *J* = 1.9 Hz, 1H), 8.32 (dd, *J* = 8.3, 2.3 Hz, 1H), 8.09 (d, *J* = 7.0 Hz, 2H), 7.87 (d, *J* = 8.3 Hz, 1H), 7.55–7.49 (m, 3H), 2.69 (s, 3H). ESI-HRMS [M + H]⁺ calcd for C₁₃H₁₂NO: 198.0913, found: 198.0915.

4.1.1.12. 2,5-*diphenylpyridine* (**18***r*). Colorless oil, 44.2% yield. ¹H NMR (600 MHz, CDCl₃) δ 8.86 (d, *J* = 2.3 Hz, 1H), 7.97 (d, *J* = 7.1 Hz, 2H), 7.88 (dd, *J* = 8.2, 2.4 Hz, 1H), 7.73 (dd, *J* = 8.2, 0.6 Hz, 1H), 7.56 (d, *J* = 7.1 Hz, 2H), 7.42 (t, *J* = 7.6 Hz, 4H), 7.37–7.32 (m, 2H). ESI-HRMS [M + H]⁺ calcd for C₁₇H₁₄N: 232.1121, found: 232.1172.

4.1.1.13. *N*-benzyl-6-phenylpyridin-3-amine (**18s**). Brown oil, 48.0% yield. ¹H NMR (600 MHz, CDCl₃) δ 8.16 (d, J = 2.8 Hz, 1H), 7.89 (d, J = 7.2 Hz, 2H), 7.54 (d, J = 8.6 Hz, 1H), 7.42 (t, J = 7.7 Hz, 2H), 7.40–7.35 (m, 4H), 7.33–7.29 (m, 2H), 6.95 (dd, J = 8.6, 2.9 Hz, 1H), 4.39 (s, 2H). ESI-HRMS [M + Na]⁺ calcd for C₁₈H₁₆N₂Na: 283.1206, found: 283.1209.

4.1.2. General procedures for the synthesis of compounds **18f-18h**, **18q** [40]

Reactions were carried out with Phenylboronic acid (146 mg, 1.2 mmol), 2-Bromo-pyridine derivatives (201 mg, 1.0 mmol, 2-bromo-N,N-dimethylpyridin-4-amine for **18f**; 227 mg, 1.0 mmol, 2-bromo-4-(pyrrolidin-1-yl)pyridine for **18g**, 243 mg, 1.0 mmol, 4-(2-bromopyridin-4-yl)morpholine for **18h**, 201 mg, 1.0 mmol, 4-(2-bromopyridin-4-yl)morpholine for **18h**, 201 mg, 1.0 mmol, 201 mg, 1.0 mmol, 6-bromo-N,N-dimethylpyridin-3-amine for **18q**), Pd(PPh₃)₄ (12 mg, 1.0 mol %), K₂CO₃ (276 mg, 2.0 mmol) in C₂H₅OH (5 mL) under nitrogen atmosphere. The mixture was stirred at 75 °C for about 15 h and the completion of the reaction was monitored by TLC. After cooling to room temperature, the mixture was extracted with ethyl acetate. The combined organic layers were washed with brine and the organic phase was dried with anhydrous sodium sulfate and concentrated *in vacuo*. The crude compound was purified by silica gel column chromatography to give corresponding intermediates.

4.1.2.1. *N*, *N*-dimethyl-2-phenylpyridin-4-amine (**18f**). White solid, 83.0% yield. ¹H NMR (600 MHz, CDCl₃) δ 8.25 (d, *J* = 6.0 Hz, 1H), 7.89 (d, *J* = 8.1 Hz, 2H), 7.40 (t, *J* = 7.5 Hz, 2H), 7.35 (d, *J* = 7.2 Hz, 1H), 6.81 (s, 1H), 6.38 (d, *J* = 5.9 Hz, 1H), 2.93 (s, 6H). ESI-HRMS [M + H]⁺ calcd for C₁₃H₁₅N₂: 199.1230, found: 199.1231.

4.1.2.2. 2-phenyl-4-(pyrrolidin-1-yl)pyridine (**18g**). White solid, 75% yield. ¹H NMR (600 MHz, CDCl₃) δ 8.31 (d, J = 5.9 Hz, 1H), 7.93 (d, J = 7.6 Hz, 2H), 7.45 (t, J = 7.5 Hz, 2H), 7.40 (t, J = 7.3 Hz, 1H), 6.76 (d, J = 2.0 Hz, 1H), 6.38 (dd, J = 5.9, 2.1 Hz, 1H), 3.39 (t, J = 6.5 Hz, 4H), 2.06 (t, J = 6.6 Hz, 4H). ESI-HRMS [M + H]⁺ calcd for C₁₅H₁₈N₂: 225.1386, found: 225.1388.

4.1.2.3. 4-(2-phenylpyridin-4-yl)morpholine (**18h**). Colorless oil, 61.0% yield.¹H NMR (600 MHz, CDCl₃) δ 8.40 (d, J = 5.9 Hz, 1H), 7.91 (d, J = 7.6 Hz, 2H), 7.45 (t, J = 7.5 Hz, 2H), 7.39 (t, J = 7.3 Hz, 1H), 7.08 (d, J = 2.4 Hz, 1H), 6.65 (dd, J = 5.9, 2.5 Hz, 1H), 3.90–3.85 (m, 4H), 3.39–3.33 (m, 4H). ESI-HRMS [M + H]⁺ calcd for C₁₅H₁₇N₂O: 241.1335, found: 241.1338.

4.1.2.4. *N*, *N*-dimethyl-6-phenylpyridin-3-amine (**18q**). White solid, 83.0% yield. ¹H NMR (600 MHz, Acetone-d₆) δ 8.08 (s, 1H), 7.91–7.84 (m, 2H), 7.60 (d, *J* = 8.8 Hz, 1H), 7.28–7.25 (m, 2H), 7.15 (t, *J* = 6.6 Hz, 1H), 7.04 (dd, *J* = 9.6, 2.3 Hz, 1H), 2.90 (s, 6H). ESI-HRMS [M + H]⁺ calcd for C₁₃H₁₅N₂: 199.1230, found: 199.1231.

4.1.3. General procedures for the synthesis of B1–B20

The synthesis of the compounds **B1–B20** was previously reported in ref. 37.

Reactions were carried out with Phenylpyridine analogs (0.4 mmol), 1,6-enynes (60 mg, 0.2 mmol), $[Cp*RhCl_2]_2$ (8 mg, 5 mol %), NaBArF₄ (55 mg, 24 mol %) in CH₃COOH (1.0 mL) under nitrogen atmosphere, The mixture was stirred at 100 °C for about 8 h and the completion of the reaction was monitored by TLC. After cooling to room temperature, the NaHCO₃ aqueous solution was added to adjust the pH to weak alkaline. The mixture was extracted with ethyl acetate. The combined organic layers were washed with brine and the organic phase was dried with anhydrous sodium sulfate and concentrated *in vacuo*. The crude compound was purified by silica gel. column chromatography to give corresponding products.

4.1.3.1. (8-cyclohexyl-5-hydroxy-1-(4-phenylpyridin-2-yl)-9,10dihydrophenanthren-9-yl)methyl acetate (**B1**). White solid, yield: 45.0%. ¹H NMR (600 MHz, CDCl₃) δ 8.79 (d, J = 5.2 Hz, 1H), 8.16 (d, J = 7.7 Hz, 1H), 7.71 (d, J = 7.2 Hz, 2H), 7.67 (d, J = 1.0 Hz, 1H), 7.56 (dd, J = 5.2, 1.8 Hz, 1H), 7.56 (dd, J = 5.2, 1.8 Hz, 1H), 7.51 (t, J = 7.4 Hz, 2H),7.47 (d, J = 7.2 Hz, 1H), 7.32 (dd, J = 7.5, 0.8 Hz, 1H), 7.24 (d, J = 7.7 Hz, 1H), 7.00 (d, J = 8.6 Hz, 1H), 6.74 (d, J = 8.5 Hz, 1H), 8.87 (dd, J = 10.8, 5.7 Hz, 1H), 3.48 (t, J = 10.5 Hz, 1H), 3.43–3.37 (m, 1H), 2.92 (dd, J = 15.5, 2.0 Hz, 1H), 2.78–2.69 (m, 2H), 1.86–1.67 (m, 8H), 1.57–1.51 (m, 1H), 1.44–1.33 (m, 2H), 1.29–1.22 (m, 2H). ¹³C NMR (150 MHz, CDCl₃) δ 170.78, 160.42, 151.69, 149.05, 148.99, 139.79, 137.99, 137.05, 135.64, 133.24, 132.79, 129.31, 129.23, 128.08, 128.01, 127.10, 126.07, 125.92, 122.61, 121.71, 119.87, 116.46, 63.65, 38.97, 35.90, 33.62, 33.08, 28.15, 27.10, 26.25, 20.51. ESI-HRMS [M + H]⁺ calcd for C₃₄H₃₄NO₃: 504.2533, found: 504.2540.

4.1.3.2. (8-(4-bromophenyl)-5-hydroxy-1-(4-phenylpyridin-2-yl)-9,10-dihydrophenanthren-9-yl)methyl acetate (**B2**). White solid, yield: 68.5%. ¹H NMR (600 MHz, CDCl₃) δ 8.78 (d, J = 5.3 Hz, 1H), 8.26 (d, J = 7.7 Hz, 1H), 7.70 (d, J = 7.2 Hz, 2H), 7.63 (s, 1H), 7.58 (dd, J = 5.3, 1.6 Hz, 1H), 7.52–7.45 (m, 6H), 7.24 (d, J = 7.7 Hz, 1H), 7.11 (d, J = 8.3 Hz, 2H), 6.86 (d, J = 8.3 Hz, 1H), 6.71 (d, J = 8.3 Hz, 1H), 3.65 (dd, J = 10.8, 5.6 Hz, 1H), 3.49 (t, J = 10.5 Hz, 1H), 3.13–3.09 (m, 1H), 2.88 (dd, J = 15.6, 4.5 Hz, 1H), 2.77 (dd, J = 15.6, 2.2 Hz, 1H), 1.50 (s, 3H).¹³C NMR (150 MHz, CDCl₃) δ 170.42, 153.43, 148.86, 140.48, 137.81, 136.27, 133.13, 132.78, 132.44, 131.23, 131.19, 129.71, 129.42, 129.26, 128.48, 128.09, 127.07, 125.91, 122.56, 121.89, 120.97, 120.01, 118.12, 116.25, 63.46, 34.14, 28.30, 20.42. ESI-HRMS [M + H]⁺ calcd for C₃₄H₂₇BrNO₃: 576.1169, found: 576.1186.

4.1.3.3. (8-(4-bromophenyl)-5-hydroxy-1-(4-methylpyridin-2-yl)-9,10-dihydrophenanthren-9-yl)methyl acetate (**B3**). White solid, yield: 40.9%, ¹H NMR (600 MHz, DMSO- d_6) δ 10.12 (s, 1H), 8.56 (d, J = 5.0 Hz, 1H), 8.49 (d, J = 7.8 Hz, 1H), 7.64 (d, J = 8.4 Hz, 2H), 7.42 (t, J = 7.7 Hz, 1H), 7.37 (dd, J = 7.6, 1.2 Hz, 1H), 7.35 (s, 1H), 7.33 (d, J = 8.4 Hz, 2H), 7.26 (d, J = 5.0 Hz, 1H), 7.04 (q, J = 8.4 Hz, 2H), 3.54 (dd, J = 10.8, 6.2 Hz, 1H), 3.44 (t, J = 10.2 Hz, 1H), 3.24–3.19 (m, 1H), 2.92–2.84 (m, 2H), 2.43 (s, 3H), 1.64 (s, 3H).¹³C NMR (150 MHz, DMSO- d_6) δ 170.00, 159.24, 155.01, 149.13, 147.43, 140.93, 140.65, 136.57, 133.26, 132.51, 132.13, 131.62, 131.49, 130.28, 128.80, 128.60, 125.96, 125.36, 123.26, 121.12, 120.61, 115.99, 63.25, 33.82, 28.25, 21.00, 20.60. ESI-HRMS [M + H]⁺ calcd for C₂₉H₂₅BrNO₃: 514.1012, found; 514.1020.

4.1.3.4. (8-(4-bromophenyl)-1-(4-chloropyridin-2-yl)-5-hydroxy-9,10-dihydrophenanthren-9-yl)methyl acetate (**B4**). White solid, yield: 27.3%, ¹H NMR (600 MHz, Acetone- d_6) δ 9.03 (s, 1H), 8.66 (d, J = 5.4 Hz, 1H), 8.59 (dd, J = 5.4, 3.8 Hz, 1H), 7.62 (d, J = 8.3 Hz, 2H), 7.60 (d, J = 1.8 Hz, 1H), 7.48 (dd, J = 5.3, 1.9 Hz, 1H), 7.42 (d, J = 1.8 Hz, 1H), 7.41 (s, 1H), 7.33 (d, J = 8.3 Hz, 2H), 7.06 (q, J = 8.3 Hz, 2H), 3.65 (dd, J = 10.9, 5.8 Hz, 1H), 3.52 (t, J = 10.5 Hz, 1H), 3.31–3.26 (m, 1H), 2.98 (dd, J = 15.7, 2.3 Hz, 1H), 2.91–2.88 (m, 1H), 1.66 (s, 3H)·¹³C NMR (150 MHz, CDCl₃) δ 170.42, 153.43, 140.48, 137.81, 137.73, 136.27, 136.23, 133.13, 132.78, 132.44, 132.33, 131.19, 129.71, 129.26, 128.48, 127.07, 125.91, 122.56, 121.89, 120.97, 120.01, 116.25, 63.46, 34.14, 28.30, 20.42. ESI-HRMS [M + H]⁺ calcd for C₂₈H₂₂BrClNO₃: 534.0466, found: 534.0475.

4.1.3.5. (8-(4-bromophenyl)-1-(4-cyanopyridin-2-yl)-5-hydroxy-9,10-dihydrophenanthren-9-yl)methyl acetate (**B5**). White solid, yield: 69.9%, ¹H NMR (600 MHz, Acetone- d_6) δ 9.05 (d, J = 16.6 Hz, 1H), 8.61 (dd, J = 7.4, 1.4 Hz, 1H), 8.33 (dd, J = 8.2, 2.1 Hz, 1H), 7.78 (d, J = 8.2 Hz, 1H), 7.61 (d, J = 8.3 Hz, 2H), 7.46–7.42 (m, 2H), 7.32 (d, J = 8.3 Hz, 2H), 7.08–7.02 (m, 2H), 3.64 (dd, J = 10.9, 5.9 Hz, 1H), 3.49 (t, J = 10.4 Hz, 1H), 3.30–3.25 (m, 1H), 3.00 (dd, J = 15.7, 2.2 Hz, 1H), 2.94 (dd, J = 13.5,4.5 Hz, 1H), 1.62 (s, 3H). ¹³C NMR (150 MHz, Acetone- d_6) δ 171.11, 164.81, 156.25, 153.61, 142.61, 141.50, 140.93, 138.50, 135.36, 134.69, 134.00, 133.43, 132.98, 131.92, 131.61, 130.30, 127.59, 126.25, 122.92, 122.32, 118.60, 117.44, 109.53, 64.54, 35.90, 29.80, 21.33. ESI-HRMS [M + Na]⁺ calcd for C₂₉H₂₁BrN₂O₃Na: 547.0628, found: 547.0635.

4.1.3.6. (8-(4-bromophenyl)-1-(4-formylpyridin-2-yl)-5-hydroxy-9,10-dihydrophenanthren-9-yl)methyl acetate (**B6**). Yellow solid, yield: 35.8%, ¹H NMR (600 MHz, Acetone- d_6) δ 10.23 (s, 1H), 9.03 (s, 1H), 8.97 (d, *J* = 4.9 Hz, 1H), 8.60 (d, *J* = 7.7 Hz, 1H), 7.96 (s, 1H), 7.80 (d, *J* = 4.8 Hz, 1H), 7.60 (d, *J* = 8.1 Hz, 2H), 7.48–7.40 (m, 2H), 7.31 (d, *J* = 8.1 Hz, 2H), 7.04 (q, *J* = 8.3 Hz, 2H), 3.64 (dd, *J* = 10.8, 5.9 Hz, 1H), 3.51 (t, *J* = 10.4 Hz, 1H), 3.31–3.23 (m, 1H), 3.00 (d, *J* = 15.6 Hz, 1H), 2.94–2.90 (m, 1H), 1.57 (s, 3H). ¹³C NMR (150 MHz, Acetone- d_6) δ 192.34, 169.32, 161.24, 154.49, 150.52, 142.34, 140.88, 139.77, 136.73, 133.43, 132.75, 132.16, 131.65, 131.17, 130.01, 129.30, 128.48, 125.69, 122.84, 121.30, 120.48, 119.83, 115.63, 62.80, 48.89, 34.15, 19.49. ESI-HRMS [M + Na]⁺ calcd for C₂₉H₂₂BrNO₄Na: 550.0624, found: 550.0631.

4.1.3.7. (8-(4-bromophenyl)-1-(4-(dimethylamino)pyridin-2-yl)-5hydroxy-9,10-dihydrophenanthren-9-yl)methyl acetate (**B7**). White solid, yield: 36.0%, ¹H NMR (600 MHz, CDCl₃) δ 8.30 (s, 1H), 8.26 (d, *J* = 5.8 Hz, 1H), 7.46 (d, *J* = 7.7 Hz, 2H), 7.09 (d, *J* = 7.7 Hz, 2H), 7.00 (d, *J* = 3.7 Hz, 2H), 6.77 (s, 2H), 6.60 (d, *J* = 4.4 Hz, 1H), 6.54 (s, 1H), 3.60 (dd, *J* = 10.6, 5.7 Hz, 1H), 3.46 (t, *J* = 9.8 Hz, 1H), 3.10 (s, 7H), 2.88 (d, *J* = 14.9 Hz, 1H), 2.56 (d, *J* = 15.1 Hz, 1H), 1.63 (s, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 170.37, 155.76, 155.00, 141.05, 135.58, 133.32, 132.70, 131.39, 131.23, 131.04, 129.46, 127.48, 125.32, 121.54, 120.62, 116.97, 107.22, 105.24, 64.07, 39.55, 33.98, 29.71, 28.44, 20.64. ESI-HRMS [M + H]⁺ calcd for C₃₀H₂₈BrN₂O₃: 543.1278, found: 543.1288.

4.1.3.8. (8-(4-bromophenyl)-5-hydroxy-1-(4-(pyrrolidin-1-yl)pyridin-2-yl)-9,10-dihydrophenanthren-9-yl)methyl acetate (**B8**). White solid, yield: 68.5%, ¹H NMR (600 MHz, CDCl₃) δ 8.41–8.36 (m, 1H), 8.11 (d, *J* = 6.0 Hz, 1H), 7.46 (d, *J* = 8.3 Hz, 2H), 7.09 (d, *J* = 8.3 Hz, 2H), 7.07 (d, *J* = 4.1 Hz, 2H), 7.03 (d, *J* = 8.0 Hz, 1H), 6.82 (d, *J* = 8.3 Hz, 1H), 6.55 (dd, *J* = 6.8, 2.2 Hz, 1H), 6.43 (d, *J* = 2.2 Hz, 1H), 3.60 (dd, *J* = 10.9, 5.9 Hz, 1H), 3.46 (d, *J* = 10.5 Hz, 5H), 3.12–3.16 (m, 1H), 3.02 (d, *J* = 13.6 Hz, 1H), 2.55 (d, *J* = 14.3 Hz, 1H), 2.09 (s, 4H), 1.62 (s, 3H)·¹³C NMR (150 MHz, CDCl₃) δ 170.36, 154.85, 153.92, 140.77, 135.56, 133.76, 132.74, 131.47, 131.38, 131.12, 130.62, 130.04, 127.64, 126.05, 120.96, 120.74, 116.92, 107.77, 105.99, 64.05, 48.14, 33.98, 28.62, 25.24, 20.68. ESI-HRMS [M + Na]⁺ calcd for C₃₂H₂₉BrN₂O₃Na: 591.1254, found: 591.1259.

4.1.3.9. (8-(4-bromophenyl)-5-hydroxy-1-(4-morpholinopyridin-2-yl)-9,10-dihydrophenanthren-9-yl)methyl acetate (**B9**). White solid, yield: 23.0%, ¹H NMR (600 MHz, CDCl₃) δ 8.41 (d, J = 6.0 Hz, 1H), 8.26 (d, J = 7.8 Hz, 1H), 7.47 (d, J = 8.3 Hz, 2H), 7.09 (d, J = 8.3 Hz, 2H), 7.05–6.98 (m, 2H), 6.76–6.71 (m, 3H), 6.63 (d, J = 8.2 Hz, 1H), 3.87–3.82 (m, 4H), 3.56 (dd, J = 10.8, 6.0 Hz, 1H), 3.49 (t, J = 10.1 Hz, 1H), 3.41–3.35 (m, 4H), 3.06 (s, 1H), 2.86 (dd, J = 15.5, 4.0 Hz, 1H), 2.59 (d, J = 14.1 Hz, 1H), 1.65 (s, 3H)·¹³C NMR (150 MHz, CDCl₃) δ 170.35, 156.05, 154.52, 141.03, 135.76, 132.89, 131.52, 131.35, 131.06, 129.25, 127.62, 125.09, 121.87, 120.67, 116.75, 108.66, 106.49, 66.33, 64.09, 46.10, 34.01, 28.39, 20.71. ESI-HRMS [M + H]⁺ calcd for C₃₂H₃₀BrN₂O₄: 585.1383, found: 585.1396.

4.1.3.10. (8-(4-bromophenyl)-5-hydroxy-1-(5-methylpyridin-2-yl)-9,10-dihydrophenanthren-9-yl)methyl acetate (**B10**). White solid, yield: 44.6%, ¹H NMR (600 MHz, CDCl₃) δ 8.57 (s, 1H), 8.20 (t, J = 4.6 Hz, 1H), 7.62 (d, J = 7.8 Hz, 1H), 7.48 (d, J = 8.3 Hz, 2H), 7.31 (d, *J* = 7.9 Hz, 1H), 7.16 (d, *J* = 4.4 Hz, 2H), 7.09 (d, *J* = 8.3 Hz, 2H), 6.82 (d, *J* = 8.3 Hz, 1H), 6.67 (d, *J* = 8.3 Hz, 1H), 3.64 (dd, *J* = 10.9, 5.9 Hz, 1H), 3.44 (t, *J* = 10.4 Hz, 1H), 3.12−3.07 (m, 1H), 2.82 (dd, *J* = 15.5, 4.5 Hz, 1H), 2.70 (dd, *J* = 15.5, 2.0 Hz, 1H), 2.44 (s, 3H), 1.66 (s, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 170.39, 153.44, 148.78, 140.54, 137.28, 136.31, 133.06, 132.69, 132.33, 131.66, 131.26, 131.17, 129.63, 128.51, 127.78, 125.80, 123.98, 121.91, 120.92, 116.18, 63.66, 34.08, 28.35, 20.63, 18.25. ESI-HRMS [M + H]⁺ calcd for C₂₉H₂₅BrNO₃: 514.1012, found: 514.1020.

4.1.3.11. (8-(4-bromophenyl)-1-(5-fluoropyridin-2-yl)-5-hydroxy-9,10-dihydrophenanthren-9-yl)methyl acetate (**B11**). White solid, yield: 36.0%, ¹H NMR (600 MHz, CDCl₃) δ 8.57 (d, J = 2.6 Hz, 1H), 8.21 (dd, J = 6.8, 2.0 Hz, 1H), 7.50 (d, J = 8.1 Hz, 3H), 7.42 (dd, J = 8.6, 4.4 Hz, 1H), 7.33–7.29 (m, 2H), 7.13 (d, J = 8.2 Hz, 2H), 6.94 (d, J = 8.3 Hz, 1H), 6.77 (d, J = 8.3 Hz, 1H), 3.69 (dd, J = 10.9, 5.8 Hz, 1H), 3.48 (t, J = 10.4 Hz, 1H), 3.17 (s, 1H), 2.88–2.78 (m, 2H), 1.68 (s, 3H)·¹³C NMR (150 MHz, CDCl₃) δ 170.47, 155.90, 153.06, 140.27, 139.16, 137.11, 136.96, 136.42, 133.05, 132.81, 131.28, 131.22, 129.96, 128.71, 128.02, 126.19, 125.32, 123.39, 123.26, 121.69, 121.11, 115.99, 63.50, 34.13, 28.34, 20.64. ¹⁹F NMR (565 MHz, CDCl₃) δ –129.13. ESI-HRMS [M + Na]⁺ calcd for C₂₈H₂₁BrFNO₃Na: 540.0581, found: 540.0590.

4.1.3.12. (8-(4-bromophenyl)-1-(5-chloropyridin-2-yl)-5-hydroxy-9,10-dihydrophenanthren-9-yl)methyl acetate (**B12**). White solid, yield: 25.5%, ¹H NMR (600 MHz, CDCl₃) δ 8.67 (d, J = 2.1 Hz, 1H), 8.22 (dd, J = 6.9, 1.7 Hz, 1H), 7.76 (dd, J = 8.3, 2.4 Hz, 1H), 7.51 (d, J = 8.2 Hz, 2H), 7.38 (d, J = 8.3 Hz, 1H), 7.34–7.29 (m, 2H), 7.13 (d, J = 8.2 Hz, 2H), 6.94 (d, J = 8.3 Hz, 1H), 6.77 (d, J = 8.3 Hz, 1H), 6.71 (s, 1H), 3.69 (dd, J = 11.0, 5.8 Hz, 1H), 3.47 (t, J = 10.4 Hz, 1H), 3.17 (s, 1H), 2.88–2.80 (m, 2H), 1.68 (s, 3H)·¹³C NMR (150 MHz, CDCl₃) δ 170.44, 157.72, 152.96, 147.78, 140.21, 139.07, 136.45, 136.17, 133.04, 132.84, 132.79, 131.27, 131.19, 130.64, 130.01, 128.67, 128.11, 126.28, 125.12, 121.61, 121.11, 115.96, 63.47, 34.11, 28.36, 20.64. ESI-HRMS [M + H]⁺ calcd for C₂₈H₂₂BrClNO₃: 534.0466, found: 534.0475.

4.1.3.13. (8-(4-bromophenyl)-1-(5-bromopyridin-2-yl)-5-hydroxy-9,10-dihydrophenanthren-9-yl)methyl acetate (**B13**). Yellow solid, yield: 20.0%, ¹H NMR (600 MHz, CDCl₃) δ 8.77 (d, J = 2.0 Hz, 1H), 8.22 (dd, J = 7.2, 1.6 Hz, 1H), 7.90 (d, J = 8.3 Hz, 1H), 7.51 (d, J = 8.3 Hz, 2H), 7.33 (t, J = 8.7 Hz, 3H), 7.14 (d, J = 8.2 Hz, 2H), 6.96 (dd, J = 8.3, 1.8 Hz, 1H), 6.80 (dd, J = 8.3, 2.5 Hz, 1H), 6.40 (s, 1H), 3.69 (dd, J = 10.9, 5.7 Hz, 1H), 3.48 (t, J = 10.4 Hz, 1H), 3.19 (s, 1H), 2.86 (d, J = 3.3 Hz, 2H), 1.69 (s, 3H)·¹³C NMR (150 MHz, CDCl₃) δ 170.41, 157.98, 152.86, 150.05, 140.16, 138.96, 136.53, 133.04, 132.90, 132.84, 131.29, 131.19, 130.09, 128.69, 128.00, 127.13, 126.40, 125.59, 121.59, 121.14, 119.26, 115.93, 63.44, 34.13, 28.38, 20.65. ESI-HRMS [M + Na]⁺ calcd for C₂₈H₂₁Br₂NO₃Na: 599.9780, found: 599.9788.

4.1.3.14. (8-(4-bromophenyl)-1-(5-cyanopyridin-2-yl)-5-hydroxy-9,10-dihydrophenanthren-9-yl)methyl acetate (**B14**). White solid, yield: 26.9%, ¹H NMR (600 MHz, Acetone- d_6) δ 9.05 (s, 1H), 8.94 (d, J = 5.0 Hz, 1H), 8.61 (d, J = 7.6 Hz, 1H), 7.92 (s, 1H), 7.75 (dd, J = 5.0, 1.3 Hz, 1H), 7.61 (d, J = 8.3 Hz, 2H), 7.46–7.41 (m, 2H), 7.32 (d, J = 8.3 Hz, 2H), 7.07–7.03 (m, 2H), 3.66 (dd, J = 10.9, 5.8 Hz, 1H), 3.50 (t, J = 10.4 Hz, 1H), 3.30–3.25 (m, 1H), 2.99–2.90 (m, 2H), 1.63 (s, 3H)·¹³C NMR (150 MHz, Acetone- d_6) δ 169.34, 160.87, 154.52, 150.32, 140.84, 138.93, 136.66, 133.50, 132.80, 132.15, 131.65, 131.18, 130.09, 129.62, 128.50, 125.88, 125.76, 123.30, 121.16, 120.50, 120.35, 116.61, 115.65, 62.78, 34.13, 27.89, 19.55. ESI-HRMS [M + H]⁺ calcd for C₂₉H₂₂BrN₂O₃: 525.0808, found: 525.0822. 4.1.3.15. (8-(4-bromophenyl)-1-(5-formylpyridin-2-yl)-5-hydroxy-9,10-dihydrophenanthren-9-yl)methyl acetate (**B15**). White solid, yield: 28.3%, ¹H NMR (600 MHz, CDCl₃) δ 10.18 (s, 1H), 9.17 (s, 1H), 8.29–8.26 (m, 2H), 7.62 (d, *J* = 8.0 Hz, 1H), 7.51 (d, *J* = 8.0 Hz, 2H), 7.43–7.36 (m, 2H), 7.14 (d, *J* = 7.9 Hz, 2H), 6.97 (d, *J* = 8.2 Hz, 1H), 6.81 (d, *J* = 8.2 Hz, 1H), 6.40 (s, 1H), 3.71 (dd, *J* = 10.8, 5.7 Hz, 1H), 3.49 (t, *J* = 10.4 Hz, 1H), 3.19 (s, 1H), 2.94–2.86 (m, 2H), 1.66 (s, 3H)·¹³C NMR (150 MHz, CDCl₃) δ 190.36, 170.36, 164.75, 152.84, 151.49, 140.07, 139.26, 136.54, 136.09, 133.25, 133.04, 132.98, 131.32, 131.17, 130.20, 129.67, 128.87, 128.75, 126.51, 124.86, 121.46, 121.20, 115.96, 63.40, 34.11, 28.51, 20.62. ESI-HRMS [M + Na]⁺ calcd for C₂₉H₂₂BrNO₄Na: 550.0624, found: 500.0632.

4.1.3.16. (8-(4-bromophenyl)-5-hydroxy-1-(5-(methylsulfonyl)pyridin-2-yl)-9,10-dihydrophenanthren-9-yl)methyl acetate (**B16**). White solid, yield: 39.0%, ¹H NMR (600 MHz, Acetone- d_6) δ 9.17 (s, 1H), 9.04 (s, 1H), 8.62 (d, *J* = 7.6 Hz, 1H), 8.40 (dd, *J* = 8.2, 2.2 Hz, 1H), 7.81 (d, *J* = 8.2 Hz, 1H), 7.60 (d, *J* = 8.2 Hz, 2H), 7.47-7.42 (m, 2H), 7.31 (d, *J* = 8.3 Hz, 2H), 7.07-7.03 (m, 2H), 3.64 (dd, *J* = 10.9, 5.9 Hz, 1H), 3.50 (t, *J* = 10.4 Hz, 1H), 3.29 (s, 4H), 3.00 (d, *J* = 15.7 Hz, 1H), 2.93 (dd, *J* = 15.7, 4.5 Hz, 1H), 1.61 (s, 3H). ¹³C NMR (150 MHz, Acetone- d_6) δ 170.29, 165.03, 155.44, 148.65, 141.80, 140.11, 137.69, 136.53, 136.50, 134.52, 133.92, 133.17, 132.61, 132.15, 131.08, 130.72, 129.51, 126.75, 125.50, 122.11, 121.48, 116.63, 63.79, 44.73, 35.06, 29.01, 20.54. ESI-HRMS [M + H]⁺ calcd for C₂₉H₂₅BrNO₅S: 578.0631, found: 578.0636.

4.1.3.17. (1-(5-acetylpyridin-2-yl)-8-(4-bromophenyl)-5-hydroxy-9,10-dihydrophenanthren-9-yl)methyl acetate (**B17**). Yellow solid, yield: 33.8%, ¹H NMR (600 MHz, CDCl₃) δ 9.27 (s, 1H), 8.33 (d, J = 5.9 Hz, 1H), 8.28 (d, J = 6.5 Hz, 1H), 7.56 (d, J = 8.1 Hz, 1H), 7.50 (d, J = 8.3 Hz, 2H), 7.31 (q, J = 4.7 Hz, 2H), 7.11 (d, J = 8.3 Hz, 2H), 6.91 (d, J = 8.3 Hz, 1H), 6.73 (d, J = 8.3 Hz, 1H), 3.68 (dd, J = 11.0, 5.8 Hz, 1H), 3.47 (t, J = 10.4 Hz, 1H), 3.19–3.14 (m, 1H), 2.91 (dd, J = 15.6, 4.5 Hz, 1H), 2.81 (d, J = 15.6 Hz, 1H), 2.70 (s, 3H), 1.66 (s, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 196.40, 170.45, 163.70, 153.22, 149.11, 140.25, 138.99, 136.36, 136.08, 133.06, 133.04, 132.62, 131.27, 131.19, 130.55, 129.95, 128.88, 128.57, 126.18, 124.46, 121.53, 121.09, 116.04, 63.53, 34.05, 28.46, 26.82, 20.65. ESI-HRMS [M + H]⁺ calcd for C₃₀H₂₅BrNO₄: 542.0961, found: 542.0979.

4.1.3.18. (8-(4-bromophenyl)-1-(5-(dimethylamino)pyridin-2-yl)-5-hydroxy-9,10-dihydrophenanthren-9-yl)methyl acetate (**B18**). White solid, yield: 48.1%, ¹H NMR (600 MHz, CDCl₃) δ 8.24 (s, 1H), 8.18 (d, *J* = 6.5 Hz, 1H), 7.46 (d, *J* = 6.5 Hz, 2H), 7.24 (s, 1H), 7.14–7.04 (m, 5H), 6.75 (d, *J* = 8.2 Hz, 1H), 6.59 (d, *J* = 8.2 Hz, 1H), 3.61–3.59 (m, 1H), 3.44 (t, *J* = 10.2 Hz, 1H), 3.06 (s, 7H), 2.85 (d, *J* = 13.0 Hz, 1H), 2.70 (d, *J* = 15.5 Hz, 1H), 1.66 (s, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 170.48, 154.12, 147.61, 145.06, 140.97, 138.80, 136.05, 133.11, 132.78, 132.51, 131.70, 131.34, 131.06, 129.20, 128.22, 127.88, 125.21, 124.50, 122.18, 120.69, 119.67, 116.45, 64.00, 40.19, 34.10, 28.38, 20.69. ESI-HRMS [M + H]⁺ calcd for C₃₀H₂₈BrN₂O₃: 543.1278, found: 543.1290.

4.1.3.19. (8-(4-bromophenyl)-5-hydroxy-1-(5-phenylpyridin-2-yl)-9,10-dihydrophenanthren-9-yl)methyl acetate (**B19**). White solid, yield:43.8%, ¹H NMR (600 MHz, CDCl₃) δ 8.99 (s, 1H), 8.25 (d, J = 7.7 Hz, 1H), 8.02 (d, J = 7.6 Hz, 1H), 7.79 (s, 1H), 7.67 (d, J = 7.3 Hz, 2H), 7.56–7.44 (m, 6H), 7.34–7.27 (m, 2H), 7.12 (d, J = 7.5 Hz, 2H), 6.89 (d, J = 8.2 Hz, 1H), 6.75 (d, J = 8.2 Hz, 1H), 3.71–3.65 (m, 1H), 3.49 (t, J = 10.3 Hz, 1H), 3.16 (s, 1H), 2.88 (dd, J = 44.9, 15.4 Hz, 2H), 1.67 (s, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 170.42, 153.45, 140.45, 136.31, 135.19, 133.10, 132.90, 132.42, 131.25, 131.20, 129.77, 129.28, 128.56, 128.45, 128.34, 127.10, 126.02, 124.62, 121.78, 120.97, 116.22, 63.63, 34.10, 28.45, 20.66. ESI-HRMS [M + H]⁺ calcd for C₃₄H₂₇BrNO₃: 576.1169, found: 576.1186. 4.1.3.20. (1-(5-(benzylamino)pyridin-2-yl)-8-(4-bromophenyl)-5hydroxy-9,10-dihydrophenanthren-9-yl)methyl acetate (**B20**). Yellow solid, yield:87.0%, ¹H NMR (600 MHz, MeOD) δ 8.45 (d, *J* = 7.9 Hz, 1H), 7.96 (d, *J* = 2.7 Hz, 1H), 7.53 (d, *J* = 8.2 Hz, 2H), 7.38 (d, J = 7.5 Hz, 2H), 7.31 (t, J = 7.7 Hz, 3H), 7.25-7.21 (m, 2H),7.20–7.16 (m, 3H), 7.07 (dd, J = 8.5, 2.8 Hz, 1H), 6.96 (d, J = 8.3 Hz, 1H), 6.90 (d, I = 8.3 Hz, 1H), 4.40 (d, I = 2.0 Hz, 2H), 3.63 (dd, *J* = 10.8, 5.4 Hz, 1H), 3.40 (t, *J* = 10.6 Hz, 1H), 3.17–3.12 (m, 1H), 2.79 (s, 2H), 1.56 (s, 3H). ¹³C NMR (150 MHz, MeOD) δ 170.80, 154.53, 146.72, 143.93, 140.78, 139.63, 139.11, 136.15, 133.22, 132.28, 131.86, 131.19, 130.91, 129.54, 128.31, 128.23, 127.87, 126.87, 126.73, 125.28, 124.88, 121.46, 120.50, 119.49, 115.06, 62.88, 46.55, 34.31, 27.66, 19.16. ESI-HRMS $[M + H]^+$ calcd for $C_{35}H_{30}BrN_2O_3$: 605.1434, found: 605.1443.

4.1.4. General procedures for the synthesis of C1-C7

NaOH (1 M) water solution (3 mL, 3.00 mmol) and CH₃OH (3 mL) were added to the 0.2 mmol corresponding intermediate **B2, B6, B10, B11, B12, B14** or **B18** and stirred for 2 h. Then the methanol was removed under vacuum, and the residue was acidified to pH = 2 or below with HCl (1 M). Then the solution was extracted with ethyl acetate (20 mL × 3) and the combined organic solvents were dried with sodium sulfate and concentrated *in vacuo*. The crude compound was purified by silica gel column chromatography to give the desired compounds **C1–C7**.

4.1.4.1. 1-(4-bromophenyl)-10-(hydroxymethyl)-8-(4-phenylpyridin-2-yl)-9,10-dihydrophenanthren-4-ol (**C1**). White solid, yield:78.8%, ¹H NMR (600 MHz, MeOD) δ 8.59 (d, J = 5.3 Hz, 1H), 8.56 (d, J = 7.8 Hz, 1H), 7.89 (s, 1H), 7.80 (d, J = 7.3 Hz, 2H), 7.68 (dd, J = 5.3, 1.6 Hz, 1H), 7.54 (d, J = 8.3 Hz, 2H), 7.51 (t, J = 7.4 Hz, 2H), 7.46 (d, J = 7.1 Hz, 1H), 7.38 (t, J = 7.7 Hz, 1H), 7.35 (d, J = 7.5 Hz, 1H), 7.21 (d, J = 8.3 Hz, 2H), 6.95 (d, J = 8.3 Hz, 1H), 6.88 (d, J = 8.3 Hz, 1H), 4.60 (s, 1H), 3.29–3.27 (m, 1H), 3.21 (d, J = 14.3 Hz, 1H), 3.11–3.03 (m, 2H), 2.72 (dd, J = 15.1, 3.4 Hz, 1H). ¹³C NMR (150 MHz, MeOD) δ 160.19, 154.61, 149.82, 148.39, 141.03, 139.66, 137.59, 137.47, 133.96, 132.52, 131.79, 131.25, 130.88, 129.60, 129.11, 128.91, 128.01, 126.85, 125.37, 122.50, 121.16, 120.44, 119.88, 114.69, 60.29, 37.69, 26.77. ESI-HRMS [M + H]⁺ calcd for C₃₂H₂₅BrNO₂, 534.1063, found, 534.1078.

4.1.4.2. 2-(8-(4-bromophenyl)-5-hydroxy-9-(hydroxymethyl)-9,10dihydrophenanthren-1-yl)isonicotinaldehyde (**C2**). Colorless oil, yield:37.0%, ¹H NMR (600 MHz, CDCl₃) δ 10.16 (s, 1H), 8.82 (d, J = 5.0 Hz, 1H), 8.38 (d, J = 7.9 Hz, 1H), 7.96 (s, 1H), 7.72 (d, J = 5.0 Hz, 1H), 7.51 (d, J = 8.2 Hz, 2H), 7.44 (t, J = 7.8 Hz, 1H), 7.32 (d, J = 7.6 Hz, 1H), 7.17 (d, J = 8.2 Hz, 2H), 6.99 (d, J = 8.2 Hz, 1H), 6.83 (d, J = 8.2 Hz, 1H), 6.17 (s, 1H), 3.47–3.40 (m, 2H), 3.27 (t, J = 11.4 Hz, 1H), 3.13–3.11 (m, 1H), 2.55 (dd, J = 14.7, 3.4 Hz, 1H).¹³C NMR (150 MHz, CDCl₃) δ 191.09, 161.55, 152.96, 149.62, 143.08, 140.11, 138.63, 134.09, 133.22, 133.07, 131.33, 131.12, 130.25, 129.15, 128.31, 126.69, 123.09, 121.55, 121.30, 120.78, 115.30, 60.91, 37.91, 26.88. ESI-HRMS [M + Na]⁺ calcd for C₂₇H₂₀BrNO₃Na: 508.0519, found: 508.0534.

4.1.4.3. $1-(4-Bromophenyl)-10-(hydroxymethyl)-8-(5-methylpyridin-2-yl)phenanthren-4-ol (C3). White solid, yield:99.0%,¹H NMR (600 MHz, CDCl₃) <math>\delta$ 8.37 (s, 1H), 8.28 (d, J = 7.9 Hz, 1H), 7.66 (dd, J = 7.9, 1.5 Hz, 1H), 7.50 (d, J = 8.2 Hz, 2H), 7.44 (d, J = 7.9 Hz, 1H), 7.39 (t, J = 7.8 Hz, 1H), 7.25 (s, 1H), 7.17 (d, J = 8.2 Hz, 2H), 6.97 (d, J = 8.2 Hz, 1H), 6.82 (d, J = 8.2 Hz, 1H), 6.26 (s, 1H), 3.52 (dd, J = 14.6, 2.6 Hz, 1H), 3.41 (dd, J = 11.6, 5.1 Hz, 1H), 3.27 (t, J = 11.4 Hz, 1H), 3.12–3.07 (m, 1H), 2.50 (dd, J = 14.6, 3.4 Hz, 1H), 2.37 (s, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 156.68, 152.91, 148.35, 140.22, 139.70, 138.89, 138.34, 133.82, 133.18, 133.15, 131.82, 131.26,

131.14, 130.06, 129.11, 127.19, 126.43, 123.76, 121.78, 121.20, 115.17, 60.75, 38.11, 26.96, 18.19. ESI-HRMS $[M\ +\ H]^+$ calcd for $C_{27}H_{23}BrNO_2^+$ 472.0907, Found 472.0914.

4.1.4.4. 1-(4-bromophenyl)-8-(5-fluoropyridin-2-yl)-10-(hydroxymethyl)-9,10-dihydrophenanthren-4-ol (C4). White solid, yield:83.0%, ¹H NMR (600 MHz, CDCl₃) δ 8.42 (d, J = 2.3 Hz, 1H), 8.30 (d, J = 7.9 Hz, 1H), 7.60-7.53 (m, 2H), 7.52 (d, J = 8.2 Hz, 2H), 7.41 (t, J = 7.8 Hz, 1H), 7.24 (s, 1H), 7.16 (d, J = 8.2 Hz, 2H), 7.00 (d, J = 8.2 Hz, 1H), 6.84 (d, J = 8.2 Hz, 1H), 5.73 (s, 1H), 3.46-3.37 (m, 2H), 3.23 (t, J = 11.4 Hz, 1H), 3.10 (dd, J = 7.0, 3.8 Hz, 1H), 2.52 (dd, J = 14.7, 3.4 Hz, 1H). ¹³C NMR (150 MHz, CDCl₃) δ 152.65, 140.09, 138.81, 136.62, 136.46, 133.83, 133.39, 133.23, 131.33, 131.11, 130.20, 129.14, 127.53, 126.55, 125.38, 124.67, 121.65, 121.31, 115.26, 60.86, 37.99, 26.80. ¹⁹F NMR (565 MHz, CDCl₃) δ -128.27. ESI-HRMS [M + H]⁺ calcd for C₂₆H₂₀BrFNO₂: 476.0656, found: 476.0667.

4.1.4.5. 1-(4-bromophenyl)-8-(5-chloropyridin-2-yl)-10-(hydroxymethyl)-9,10-dihydrophenanthren-4-ol (C5). White solid, yield:84.4%, ¹H NMR (600 MHz, CDCl₃) δ 8.52 (d, J = 2.3 Hz, 1H), 8.32 (d, J = 7.9 Hz, 1H), 7.83 (dd, J = 8.3, 2.5 Hz, 1H), 7.50 (t, J = 8.2 Hz, 3H), 7.41 (t, J = 7.8 Hz, 1H), 7.25 (d, J = 8.1 Hz, 1H), 7.16 (d, J = 8.3 Hz, 2H), 6.99 (d, J = 8.2 Hz, 1H), 6.82 (d, J = 8.3 Hz, 1H), 6.02 (s, 1H), 5.27 (s, 1H), 3.45 (dd, J = 14.8, 2.7 Hz, 1H), 3.39 (s, 1H), 3.24 (t, J = 11.4 Hz, 1H), 3.13–3.09 (m, 1H), 2.53 (dd, J = 14.8, 2.5 Hz, 1H). ¹³C NMR (150 MHz, CDCl₃) δ 157.82, 152.83, 147.15, 140.10, 138.71, 138.58, 137.54, 133.93, 133.27, 133.11, 131.32, 131.11, 130.81, 130.23, 129.03, 127.89, 126.59, 125.20, 121.59, 121.29, 115.26, 60.88, 37.93, 26.84. ESI-HRMS [M + H]⁺ calcd for C₂₆H₂₀BrClNO₂: 492.0360, found: 492.0373.

4.1.4.6. 6-(8-(4-bromophenyl)-5-hydroxy-9-(hydroxymethyl)-9,10dihydrophenanthren-1-yl)nicotinonitrile (**C6**). White solid, yield:80.0%, ¹H NMR (600 MHz, MeOD) δ 8.93 (s, 1H), 8.58 (d, J = 7.7 Hz, 1H), 8.23 (d, J = 8.1 Hz, 1H), 7.79 (d, J = 8.2 Hz, 1H), 7.54 (d, J = 8.0 Hz, 2H), 7.42–7.34 (m, 2H), 7.21 (d, J = 8.0 Hz, 2H), 6.96 (d, J = 8.3 Hz, 1H), 6.88 (d, J = 8.3 Hz, 1H), 3.25 (dd, J = 10.3, 4.1 Hz, 1H), 3.09 (d, J = 15.3 Hz, 1H), 3.02 (s, 1H), 2.97 (t, J = 10.6 Hz, 1H), 2.76 (dd, J = 15.3, 3.7 Hz, 1H) ppm. ¹³C NMR (150 MHz, MeOD) δ 163.20, 154.58, 151.25, 140.95, 140.17, 138.47, 137.36, 134.16, 132.81, 131.76, 131.23, 130.90, 129.75, 127.97, 125.48, 124.84, 120.84, 120.47, 116.28, 114.71, 107.97, 60.28, 37.61, 26.88.ESI-HRMS [M + H]⁺ calcd for C₂₇H₂₀BrN₂O₂: 483.0703, found: 483.0724.

4.1.4.7. 1-(4-bromophenyl)-8-(5-(dimethylamino)pyridin-2-yl)-10-(hydroxymethyl)-9,10-dihydrophenanthren-4-ol (**C7**). White solid, yield:81.3%, ¹H NMR (600 MHz, CDCl₃) δ 8.21 (d, J = 7.8 Hz, 1H), 8.04 (d, J = 2.9 Hz, 1H), 7.50 (d, J = 8.2 Hz, 2H), 7.37 (dd, J = 18.0, 8.3 Hz, 2H), 7.25 (d, J = 7.7 Hz, 1H), 7.17 (d, J = 8.2 Hz, 2H), 7.13 (dd, J = 8.7, 2.9 Hz, 1H), 6.97 (d, J = 8.2 Hz, 1H), 6.84 (d, J = 8.2 Hz, 1H), 3.57 (d, J = 14.2 Hz, 1H), 3.40 (dd, J = 11.6, 5.1 Hz, 1H), 3.26 (t, J = 11.4 Hz, 1H), 3.13–3.07 (m, 1H), 2.99 (s, 6H), 2.49 (dd, J = 14.5, 3.3 Hz, 1H). ¹³C NMR (150 MHz, CDCl₃) δ 152.85, 144.97, 140.30, 139.02, 133.71, 133.30, 133.19, 131.24, 131.16, 129.94, 129.09, 126.37, 124.07, 121.95, 121.17, 115.18, 60.73, 40.06, 38.23, 27.07 ESI-HRMS [M + H]⁺ calcd for C₂₈H₂₆BrN₂O₂: 501.1172, found: 501.1180.

4.2. Biological evaluations

4.2.1. Chemicals and reagents

SARS-CoV-2 3CL^{pro} and SARS-CoV 3CL^{pro} were expressed and purified by ourselves. All tested compounds were dissolved in DMSO and stored at 4 °C. Dabcyl-KNSTLQSGLRKE-Edans is the fluorescent probe for SARS-CoV-2 3CL^{pro} inhibitory assessments [31,36], which was ordered from Shanghai Sangon Biological Engineering & Technology and Service Co. Ltd. (Shanghai, China). Ethylene Diamine Tetraacetic Acid (EDTA), disulfiram (the positive inhibitor), β-NADP⁺, D-glucose-6-phosphate (G-6-P), glucose-6phosphate dehydrogenase (G-6-PDH), testosterone, phenacetin, coumarin, paclitaxel, omeprazole, dextromethorphan, chlorzoxazone, pancreatin, as well as pepsin were provided by Dalian Meilun Biotechnology Co. LTD. (Dalian, China). Diclofenac was obtained from Ark Pharm (Wuhan, China). Lansoprazole was purchased from Hairong (Sichuan, China). KH₂PO₄, MgCl₂, NaOH and HCl were bought from Sinopharm Chemical Reagent Co., Ltd. (Shanghai, China). Uridine-50-diphosphoglucuronic acid (UDPGA), tris mefenamic acid and dodecyl polyglycol ether were purchased from Sigma-Aldrich (St. Louis, MO, USA). 7-hydroxycoumarin was gained from AlfaAesar (Shanghai, China). Warfarin sodium was purchased from J&K Scientific (Beijing, China). The pooled human liver microsomes (Lot No. H0610) from 50 individual donors were supplied by XenoTech (USA). The pooled human plasma samples were provided by Putuo Hospital, Shanghai University of Traditional Chinese Medicine (Shanghai, China), with the reference number of RTEC-A-2020-59-1. The stock solution of the fluorescent substrate, PBS buffer, Tris-HCl buffer were prepared by Millipore water (Millipore, Bedford, USA) and stored at 4 °C for further use. HPLC grade DMSO (Tedia, USA) was used throughout.

4.2.2. 3CL^{pro} inhibitory assays

The SARS-CoV-2 3CL^{pro} and SARS-CoV 3CL^{pro} inhibitory assays were conducted according to the reported procedure [31,36], which were established by the 3CL^{pro}-mediated Dabcyl-KNSTLQSGLRKE-Edans hydrolytic reaction. In brief, the tested compound, SARS-CoV-2 3CL^{pro} (4 µg/mL, final concentration) or SARS-CoV 3CL^{pro} (30 µg/mL, final concentration) were added into a black 96-well plate that filled with PBS buffer (0.1 M, pH 7.4, 1 mM EDTA) to co-incubate at 37 °C for 30 min. After the peptide-like substrate (20 µM or 40 µM, final concentration) was added to initiate the reaction, the produced fluorescent values were continuously detected by a microplate reader (SpectraMax® iD3, Molecular Devices, Austria) for 20 min at the excitation wavelength of 340 nm and emission wavelength of 490 nm.

4.2.3. Determination of inhibition constants of C1 and C2

The inhibition constants (K_i) and inhibition modes for C1 and C2 against SARS-CoV-2-3CL^{pro}-mediated-Dabcyl-KNSTLQSGLRKE-Edans cleavage were determined by inhibition kinetics. According to the previously reported methodology [41,42], increasing concentrations of C1 and C2 were separately co-incubated with various concentrations of Dabcyl-KNSTLQSGLRKE-Edans, and the generated fluorescence signals were analyzed to determine the inhibition modes and the K_i values.

4.2.4. Gastrointestinal stability assays

The simulated intestinal fluid (SIF) was confected by pancreatin (400 mg), KH₂PO₄ (272 mg), as well as NaOH (0.4%) in 40 mL water, with the pH value of 6.8. The simulated gastric fluid (SGF) was prepared by water (40 mL) that contained pepsin (400 mg) and HCI (656 μ L, 1 M). The gastrointestinal metabolic system (100 μ L) included SIF or SGF, and the analyte (20 μ M). After the reaction was proceed for 0 min, 5 min, 10 min, 15 min, 20 min, 30 min, 45 min, 60 min, 90 min at 37 °C, equal volume of pre-cooling acetonitrile (100 μ L) was added to quench the reaction. Then the samples were centrifuged at 20,000×g, 4 °C for 30 min, 100 μ L of which were transferred for LC-UV analysis.

4.2.5. Plasma stability assays

The stabilities of **C1** and **C2** were proceed in PBS buffer (100 mM, pH 7.4) at 37 $^{\circ}$ C. After the co-incubation was proceed for 0 min,

15 min, 30 min, 1 h, 2 h, 4 h, 6 h, 8 h, 10 h, 12 h, the pre-cooling acetonitrile (300 μ L) was added to precipitate the proteins. Then the samples were centrifuged at 20,000×g, 4 °C for 30 min, 100 μ L of which were transferred for LC-UV analysis [43].

4.2.6. Metabolic stability assays

The phase I metabolic assessments were proceed in PBS buffer (100 mM, pH 7.4) that comprised with C1 or C2 or testosterone (20 µM), HLMs (0.2 µg/mL), as well as specific cofactors (4 mM MgCl₂, 10 mM G-6-P, 1 unit/mL G-6-PDH). After equilibrated at 37 °C for 3 min, the samples were initiated by β -NADP⁺ (1 mM). While the phase II metabolic assessments were conducted in Tris-HCl buffer (50 mM, pH 7.4), which contained C1 or C2 or 7-Hydroxycoumarin (20 µM), HLMs (0.2 mg/mL), as well as MgCl₂ (5 mM). The reaction mixture was incubated at 37 °C for 3 min, then added with UDPGA (2 mM) to initiate the reaction. The reactions of phase I and phase II metabolic reactions were quenched by precooling acetonitrile (100 μ L) after the reactions proceed for different times (0 min, 5 min, 15 min, 30 min, 45 min, 60 min, 90 min). All the quenched mixtures were centrifuged at $20,000 \times g$, 4 °C for 30 min, the supernatant was transferred for LC-UV analysis [42].

4.3. Molecular docking

Molecular docking study was conducted using Schrodinger software package. Firstly, the SARS-CoV-2 3CL^{pro} was downloaded from Protein Data Bank (PDB code: 7BE7) [44] and prepared with the Protein Preparation Wizard model including the removal of waters, the addition of missing hydrogen atoms, the generation of heteroatom states using Epik: pH 7.0 \pm 2.0 and the assignment of bond order. Then, a restrained minimization to relieve static clashes was conducted using the OPLS-2005 force field (converge heavy atoms to RMSD 0.30 Å). For the docking study at the dimer interface position, the chain B should be deleted except for residues 1–8. Then, the receptor grid was generated at the centroid of residues 1–8 and the grid box size was set to 20 Å \times 20 Å \times 20 Å. For the docking study at the substrate-binding pocket, the receptor grid was generated at the centroid of ligand and the grid box size was set to 20 Å \times 20 Å \times 20 Å. After preparing ligands using LigPrep module, ligand docking was carried out using the standard precision (SP) with the default settings. Finally, the pictures were generated using pymol software.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgement

Financial support was generously provided by the National Natural Science Foundation of China (Nos.: 81903423, 21871184, 22071155, 81922070 and 81973286), the Shanghai Sailing Program (No: 19YF1449300), the Shanghai Municipal Education Commission (No: 2019-01-07-00-10-E00072), the Science and Technology Commission of Shanghai Municipality (Nos.: 18401933500 and 20400750300), Shanghai Science and Technology Innovation Action Plan biomedical technology support special project (No: 21S21900600), the Innovation Team and Talents Cultivation Program of National Administration of Traditional Chinese Medicine (No: ZYYCXTD-D-202004).

Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.ejmech.2021.114030.

References

- [1] S.P. Otto, T. Day, J. Arino, C. Colijn, J. Dushoff, M. Li, S. Mechai, G.V. Domselaar, J.H. Wu, D.J.D. Earn, N.H. Ogden, The origins and potential future of SARS-CoV-2 variants of concern in the evolving COVID-19 pandemic, Curr. Biol. 31 (2021) R918–R929.
- [2] A. Sharma, I. Ahmad Farouk, S.K. Lal, COVID-19: a review on the novel coronavirus disease evolution, transmission, detection, control and prevention, Viruses 13 (2021) 202.
- [3] T. Wu, S.T. Kang, W.Y. Peng, C.Z. Zuo, Y.H. Zhu, L.Y. Pan, K.Y. Fu, Y.X. You, X.Y. Yang, X. Luo, L.P. Jiang, M.C. Deng, Original hosts, clinical features, transmission routes, and vaccine development for coronavirus disease (COVID-19), Front. Med. 8 (2021) 702066.
- [4] A. Awadasseid, Y. Wu, Y. Tanaka, W. Zhang, Current advances in the development of SARS-CoV-2 vaccines, Int. J. Biol. Sci. 17 (2021) 8–19.
- [5] P. Cihan, Forecasting fully vaccinated people against COVID-19 and examining future vaccination rate for herd immunity in the US, Asia, Europe, Africa, South America, and the World, Appl. Soft Comput. 111 (2021) 107708.
- [6] C.L.D.C. Badua, K.A.T. Baldo, P.M. B Medina, Genomic and proteomic mutation landscapes of SARS-CoV-2, J. Med. Virol. 93 (2021) 1702–1721.
- [7] M. Scudellari, How the coronavirus infects cells and why Delta is so dangerous, Nature 595 (2021) 640-644.
- [8] J. Singh, S.A. Rahman, N.Z. Ehtesham, S. Hira, S.E. Hasnain, SARS-CoV-2 variants of concern are emerging in India, Nat. Med. 7 (2021) 1131–1133.
- [9] C. Liu, H.M. Ginn, W. Dejnirattisai, P. Supasa, B. Wang, A. Tuekprakhon, R. Nutalai, D. Zhou, A.J. Mentzer, Y.G. Zhao, H.M.E. Duyvesteyn, C. López-Camacho, J. Slon-Campos, T.S. Walter, D. Skelly, S.A. Johnson, T.G. Ritter, C. Mason, S.A.C. Clemens, F.G. Naveca, V. Nascimento, F. Nascimento, C.F.D. Costa, P.C. Resende, A. Pauvolid-Correa, M.M. Siqueira, C. Dold, N. Temperton, T. Dong, A.J. Pollard, J.C. Knight, D. Crook, T. Lambe, E. Clutterbuck, S. Bibi, A. Flaxman, M. Bittaye, S. Belij-Rammerstorfer, S.C. Gilbert, T. Malik, M.W. Carroll, P. Klenerman, E. Barnes, S.J. Dunachie, V. Baillie, N. Serafin, Z. Ditse, K.D. Silva, N.G. Paterson, M.A. Williams, D.R. Hall, S. Madhi, M.C. Nunes, P. Goulder, E.E. Fry, J. Mongkolsapaya, J. Ren, D.I. Stuart, G.R. Screaton, Reduced neutralization of SARS-CoV-2 B.1.617 by vaccine and convalescent serum, Cell 184 (2021) 4220–4236.
- [10] J. Lopez Bernal, N. Andrews, C. Gower, E. Gallagher, R. Simmons, S. Thelwall, J. Stowe, E. Tessier, N. Groves, G. Dabrera, R. Myers, C.N.J. Campbell, G. Amirthalingam, M. Edmunds, M. Zambon, K.E. Brown, S. Hopkins, M. Chand, M. Ramsay, Effectiveness of Covid-19 vaccines against the B.1.617.2 (Delta) varian, N. Engl. J. Med. 358 (2021) 585–594.
- [11] A. Shamsi, T. Mohammad, S. Anwar, S. Amani, M.S. Khan, F.M. Husain, M.T. Rehman, A. Islam, M.I. Hassan, Potential drug targets of SARS-CoV-2: from genomics to therapeutics, Int. J. Biol. Macromol. 177 (2021) 1–9.
- [12] T. Pillaiyar, S. Meenakshisundaram, M. Manickam, Recent discovery and development of inhibitors targeting coronaviruses, Drug Discov, Today Off. 25 (2020) 668–688.
- [13] Q. Li, C. Kang, Progress in developing inhibitors of SARS-CoV-2 3C-Like protease, Microorganisms 8 (2020) 1250.
- [14] R.J. Lu, X. Zhao, J. Li, P.H. Niu, B. Yang, H.L. Wu, W.L. Wang, H. Song, B.Y. Huang, N. Zhu, Y.H. Bi, X.J. Ma, F.X. Zhan, L. Wang, T. Hu, H. Zhou, Z.H. Hu, W.M. Zhou, L. Zhao, J. Chen, Y. Meng, J. Wang, Y. Lin, J.Y. Yuan, Z.H. Xie, J.M. Ma, W.J. Liu, D.Y. Wang, W.B. Xu, E.C. Holmes, G.F. Gao, G. Wu, W.J. Chen, W.F. Shi, W.J. Tan, Genomic characterisation and epidemiology of 2019 novel coronivrus: implications for virus origins and receptor binding, Lancet 395 (2020) 565–574.
- [15] M.G. Deshmukh, J.A. Ippolito, C.H. Zhang, E.A. Stone, R.A. Reilly, S.J. Miller, W.L. Jorgensen, K.S. Anderson, Structure-guided design of a perampanelderived pharmacophore targeting the SARS-CoV-2 main protease, Structure 8 (2021) 823–833.
- [16] R. Cannalire, C. Cerchia, A.R. Beccari, F.S.D. Leva, V. Summa, Targeting SARS-CoV-2 proteases and polymerase for COVID-19 treatment: state of the art and future opportunities, J. Med. Chem. doi: 10.1021/acs.jmedchem.0c01140.
- [17] I. Ahmad Shagufta, The race to treat COVID-19: potential therapeutic agents for the prevention and treatment of SARS-CoV-2, Eur. J. Med. Chem. 213 (2021) 113157.
- [18] Y.Z. Liu, C.Y. Liang, L. Xin, X.D. Ren, L. Tian, X.K. Ju, H. Li, Y.B. Wang, Q.Q. Zhao, H. Liu, W.Q. Cao, X.L. Xie, D.Z. Zhang, Y. Wang, Y.L. Jian, The development of Coronavirus 3C-Like protease (3CL^{pro}) inhibitors from 2010 to 2020, Eur. J. Med. Chem. 206 (2020) 112711, https://doi.org/10.1016/ j.ejmech.2020.112711.
- [19] E.D. Vita, 10 years into the resurgence of covalent drugs, Future Med. Chem. 13 (2021) 193–210.
- [20] Y. Xiong, G.H. Zhu, Y.N. Zhang, Q. Hu, H.N. Wang, H.N. Yu, X.Y. Qin, X.Q. Guan, Y.W. Xiang, H. Tang, G.B. Ge, Flavonoids in *Ampelopsis grossedentata* as covalent inhibitors of SARS-CoV-2 3CLpro: inhibition potentials, covalent binding sites and inhibitory mechanisms, Int. J. Biol. Macromol. 187 (2021) 976–987.
- [21] N. Verma, J.A. Henderson, J. Shen, Proton-coupled conformational activation of

SARS coronavirus main proteases and opportunity for designing smallmolecule broad-spectrum targeted covalent inhibitors, J. Am. Chem. Soc. 142 (2020) 21883–21890.

- [22] Z. Jin, X. Du, Y. Xu, Y. Deng, M. Liu, Y. Zhao, B. Zhang, X. Li, L. Zhang, C. Peng, Y. Duan, J. Yu, L. Wang, K. Yang, F. Liu, R. Jiang, X. Yang, T. You, X. Liu, X. Yang, F. Bai, H. Liu, X. Liu, L.W. Guddat, W. Xu, G. Xiao, C. Qin, Z. Shi, H. Jiang, Z. Rao, H. Yang, Structure of Mpro from COVID-19 virus and discovery of its inhibitors, Nature 582 (2020) 289–293.
- [23] L.L. Zhang, D.Z. Lin, X.Y.Y. Sun, U. Curth, C. Drosten, L. Sauerhering, S. Becker, K. Rox, R. Hilgenfeld, Crystal structure of SARS-CoV-2 main protease provides a basis for design of improved alpha-ketoamide inhibitors, Science 368 (2020) 409–412.
- [24] (a) W.H. Dai, B. Zhang, X.M. Jiang, H.X. Su, J. Li, Y. Zhao, X. Xie, Z.M. Jin, J.J. Peng, F.J. Liu, C.P. Li, Y. Li, F. Bai, H.F. Wang, X. Cheng, X.B. Cen, S.L. Hu, X.N. Yang, J. Wang, X. Liu, G.F. Xiao, H.L. Jiang, Z.H. Rao, L.K. Zhang, Y.C. Xu, H.T. Yang, H. Liu, Structure-based design of antiviral drug candidates targeting the SARS-CoV-2 main protease, Science 368 (2020) 1331–1335;
 (b) J.X. Qiao, Y.S. Li, R. Zeng, F.L. Liu, R.H. Luo, C. Huang, Y.F. Wang, J. Zhang, B.X. Quan, C.J. Shen, X. Mao, X.L. Liu, W.N. Sun, W. Yang, X.C. Ni, K. Wang, L. Xu, Z.L. Duan, Q.C. Zou, H.L. Zhang, W. Qu, Y.H.P. Long, M.H. Li, R.C. Yang, X.L. Liu, J. You, Y.L. Zhou, R. Yao, W.P. Li, J.M. Liu, P. Chen, Y. Liu, G.F. Lin, X. Yang, J. Zou, L.L Li, Y.G. Hu, G.W. Lu, W.M. Li, Y.Q. Wei, Y.T. Zheng, J. Lei, S.Y. Yang, SARS-CoV-2 Mpro inhibitors with antiviral activity in a transgenic mouse model, Science 371 (2021) 1374–1378.
 [25] C. Ma, M.D. Sacco, B. Hurst, I.A. Townsend, Y.M. Hu, T. Szeto, X.J. Zhang, J. Zou, J. Li, J. Sou, S. Cov, J. Mang, J. Mang, J. Mang, J. Zou, J. Science 371 (2021) 1374–1378.
- B. Tarbet, M.T. Marty, Y. Chen, J. Wang, Boceprevir, GC-376, and calpain inhibitors II, XII inhibit SARS-CoV-2 viral replication by targeting the viral main protease, Cell Res. 30 (2020) 678–692.
- [26] B. Boras, R.M. Jones, B.J. Anson, D. Arenson, L. Aschenbrenner, M.A. Bakowski, N. Beutler, J. Binder, E. Chen, H. Eng, H. Hammond, J. Hammond, R.E. Haupt, R. Hoffman, E.P. Kadar, R. Kania, E. Kimoto, M.G. Kirkpatrick, L. Lanyon, E.K. Lendy, J.R. Lillis, J. Logue, S.A. Luthra, C. Ma, S.W. Mason, M.E. McGrath, S. Noell, R.S. Obach, M.N. O'Brien, R. O'Connor, K. Ogilvie, D. Owen, M. Pettersson, M.R. Reese, T.F. Rogers, M.I. Rossulek, J.G. Sathish, N. Shirai, C. Steppan, M. Ticehurst, L.W. Updyke, S. Weston, Y. Zhu, J. Wang, A.K. Chatterjee, A.D. Mesecar, M.B. Frieman, A.S. Anderson, C. Allerton, Discovery of a novel inhibitor of coronavirus 3CL protease as a clinical candidate for the potential treatment of COVID-19, BioRxiv (2020), https://doi.org/ 10.1101/2020.09.12.293498.
- [27] Pfizer Unveils its Oral SARS-CoV-2 Inhibitor, 2021. Available from: https://cen. acs.org/acs-news/acs-meeting-news/Pfizerunveils-oral-SARS-CoV/99/i13.
- [28] H.X. Su, S. Yao, W.F. Zhao, Y.M. Zhang, J. Liu, Q. Shao, Q.X. Wang, M.J. Li, H. Xie, W.J. Shang, C.Q. Ke, L. Feng, X.R. Jiang, J.S. Shen, G.F. Xiao, H.L. Jiang, L.K. Zhang, Y. Ye, Y.C. Xu, Identification of pyrogallol as a warhead in design of covalent inhibitors for the SARS-CoV-2 3CL protease, Nat. Commun. 12 (2021) 3623.
- [29] R.Y. Wang, Q. Hu, H.N. Wang, G.H. Zhu, M.G. Wang, Q. Zhang, Y.S. Zhao, C.Y. Li, Y.N. Zhang, G.B. Ge, H.Z. Chen, L.L. Chen, Identification of Vitamin K3 and its analogues as covalent inhibitors of SARS-CoV-2 3CL 183 (2021) 182–192.
- [30] A.K. Ghosh, J. Raghavaiah, D. Shahabi, M. Yadav, B.J. Anson, E.K. Lendy, S. Hattori, N. Higashi-Kuwata, H. Mitsuya, A.D. Mesecar, Indole chloropyridinyl ester-derived SARS-CoV-2 3CLpro inhibitors: enzyme inhibition, antiviral efficacy, structure-activity relationship, and X-ray structural studies, J. Med. Chem. doi:10.1021/acs.jmedchem.1c01214.
- [31] Y. Xiong, G.H. Zhu, H.N. Wang, Q. Hu, L.L. Chen, X.Q. Guan, H.L. Li, H.Z. Chen, H. Tang, G.B. Ge, Discovery of naturally occurring inhibitors against SARS-CoV-2 3CL from *Ginkgo biloba* leaves via large-scale screening, Fitoterapia 152 (2021) 104909.
- [32] N. Kitamura, M.D. Sacco, C. Ma, Y.M. Hu, J.A. Townsend, X.Z. Meng, F.S. Zhang, X.J. Zhang, M. Ba, T. Szeto, A. Kukuljac, M.T. Marty, D. Schultz, S. Cherry, Y. Xiang, Y. Chen, J. Wang, Expedited approach toward the rational design of noncovalent SARS-CoV-2 main protease inhibitors, J. Med. Chem.. doi: 10.1021/acs.jmedchem. 1C00509.
- [33] S.H. Han, C. M Goins, T. Arya, W.J. Shin, J. Maw, A. Hooper, D.P. Sonawane, M.R. Porter, B.E. Bannister, R.D. Crouch, A.A. Lindsey, G. Lakatos, S.R. Martinez, J. Alvarado, W.S. Akers, N.S. Wang, J.U. Jung, J.D. Macdonald, S.R. Stauffer, Structure-based optimization of ML300-derived, noncovalent inhibitors targeting the severe acute respiratory syndrome coronavirus 3CL protease (SARS-CoV-2 3CL^{pro}), J. Med. Chem. (2021), https://doi.org/10.1021/ acs.jmedchem.1c00598.
- [34] C.H. Zhang, E.A. Stone, M. Deshmukh, J.A. Ippolito, M.M. Ghahremanpour, J. Tirado-Rives, K.A. Spasov, S. Zhang, Y. Takeo, S.N. Kudalkar, Z. Liang, F. Isaacs, B. Lindenbach, S.J. Miller, K.S. Anderson, W.L. Jorgensen, Potent noncovalent inhibitors of the main protease of SARS-CoV-2 from molecular sculpting of the drug perampanel guided by free energy perturbation calculations, ACS Cent. Sci. 7 (2021) 467–475.
- [35] M. Erak, K. Bellmann-Sickert, S. Els-Heindl, A.G. Beck-Sickinger, Peptide chemistry toolbox – transforming natural peptides into peptide therapeutics, Bioorg. Med. Chem. 26 (2018) 2759–2765.
- [36] L.L. Chen, S. Chen, C. Gui, J. Shen, X. Shen, H. Jiang, Discovering severe acute respiratory syndrome coronavirus 3CL protease inhibitors: virtual screening, surface plasmon resonance, and fluorescence resonance energy transfer assays, J. Biomol. Screen 118 (2006) 915–921.
- [37] D.D. Gao, X.Y. Liu, H. Xu, Y.X. Tan, Q. Liao, Q.H. Li, X.D. Yang, G.Q. Lin, P. Tian, One-pot preparation of 9,10-dihydrophenanthrenes initiated by rhodium (III)-catalyzed C–H activation and relay Diels-Alder reaction, Org. Lett. 22

J.-W. Zhang, Y. Xiong, F. Wang et al.

(2020) 4300-4305.

- [38] B. Goyal, D. Goyal, Targeting the dimerization of the main protease of coronaviruses: a potential broad-spectrum therapeutic strategy, ACS Comb. Sci. 22 (2020) 297–305.
- [39] A.J. Paterson, C.J. Heron, C.L. McMullin, M.F. Mahon, N.J. Pressc, C.G. Frost, α-Halo carbonyls enable meta selective primary, secondary and tertiary C-H alkylations by ruthenium catalysis, Org. Biomol. Chem. 15 (2017) 5993–6000.
 [40] F. Brunner, N. Marinakis, C. Wobill, Modular synthesis of simple cyclo-
- [40] F. Brunner, N. Marinakis, C. Wobill, Modular synthesis of simple cycloruthenated complexes with state-of-the-art performance in p-type DSCs, J. Mater. Chem. C. 4 (2016) 9823–9833.
- [41] Y.Q. Song, Z.M. Weng, M. Finel, Y.Q. Wang, L.L. Ding, Q. Jin, D.D. Wang, S.Q. Fang, Y.F. Cao, J. Hou, G.B. Ge, Inhibition of human carboxylesterases by

European Journal of Medicinal Chemistry 228 (2022) 114030

magnolol: kinetic analyses and mechanism, Chem. Biol. Interact. 308 (2019) 339–349.

- [42] P.C. Huo, X.Q. Guan, P. Liu, Y.Q. Song, M.R. Sun, R.J. He, L.W. Zou, L.J. Xue, J.H. Shi, N. Zhang, Z.G. Liu, G.B. Ge, Design, synthesis and biological evaluation of indanone-chalcone hybrids as potent and selective hCES2A inhibitors, Eur. J. Med. Chem. 209 (2021) 112856.
- [43] T.D.Y. Chung, D.B. Terry, L.H. Smith, In Vitro and in Vivo Assessment of ADME and PK Properties during Lead Selection and Lead Optimization – Guidelines, Benchmarks and Rules of Thumb, 2004, pp. 1307–1320. Bethesda (MD), https://www.ncbi.nlm.nih.gov/books/NBK326710/.
- [44] Crystal structure of MG-132 covalently bound to the main protease (3CLpro/ Mpro) of SARS-CoV-2, Available from, https://www.rcsb.org/structure/7BE7.