

The Role of the Renin–Angiotensin System in the Cancer Stem Cell Niche

Ethan J. Kilmister and Swee T. Tan

Gillies McIndoe Research Institute, Wellington, New Zealand (EJK, STT); Wellington Regional Plastic, Maxillofacial and Burns Unit, Hutt Hospital, Wellington, New Zealand (STT); and Department of Surgery, The University of Melbourne, Parkville, Victoria, Australia (STT)

Summary

Cancer stem cells (CSCs) drive metastasis, treatment resistance, and tumor recurrence. CSCs reside within a *niche*, an anatomically distinct site within the tumor microenvironment (TME) that consists of malignant and non-malignant cells, including immune cells. The renin–angiotensin system (RAS), a critical regulator of stem cells and key developmental processes, plays a vital role in the TME. Non-malignant cells within the CSC *niche* and stem cell signaling pathways such as the Wnt, Hedgehog, and Notch pathways influence CSCs. Components of the RAS and cathepsins B and D that constitute bypass loops of the RAS are expressed on CSCs in many cancer types. There is extensive *in vitro* and *in vivo* evidence showing that RAS inhibition reduces tumor growth, cell proliferation, invasion, and metastasis. However, there is inconsistent epidemiological data on the effect of RAS inhibitors on cancer incidence and survival outcomes, attributed to different patient characteristics and methodologies used between studies. Further mechanistic studies are warranted to investigate the precise effects of the RAS on CSCs directly and/or the CSC *niche*. Targeting the RAS, its bypass loops, and convergent signaling pathways participating in the TME and other key stem cell pathways that regulate CSCs may be a novel approach to cancer treatment. (J Histochem Cytochem 69: 835–847, 2021)

Keywords

angiogenesis, cancer, cancer stem cell, cancer stem cell *niche*, drug repurposing, drug resistance, microenvironmental *niche*, renin–angiotensin system, stemness, tumor heterogeneity, tumor immune infiltrate, tumor microenvironment

Introduction

Cancer is the leading cause of death globally across all income levels, with its incidence predicted to rapidly rise. This is largely attributed to aging populations, and to a lesser extent, adoption of certain lifestyles.¹ The growing burden of cancer and the escalating cost of treatment² underscore an urgent need for more effective and affordable treatment.

There are two concepts guiding cancer research: (1) the stochastic model, also known as the clonal expansion model, and (2) the cancer stem cell (CSC) model, also known as the hierarchical model of cancer.³ The stochastic model proposes that cancer cells result from normal cells acquiring genetic mutations and oncogenic properties in a stepwise fashion. The high mutation rate increases the probability of cell

clones being formed that are adapted to the selection pressures of a given tumor site.³ However, the CSC model proposes CSCs as the origin of cancer and the driver of treatment resistance,⁴ cancer metastasis, and recurrence.⁵ CSCs divide asymmetrically giving rise to non-tumorigenic cancer cells which form the bulk of the tumor, and identical highly tumorigenic CSCs that sit atop the cellular hierarchy in an undifferentiated state.⁶ CSCs have several proposed origins, including non-malignant stem cells and/or

Received for publication March 29, 2021; accepted May 28, 2021.

Corresponding Author:

Swee T. Tan, Gillies McIndoe Research Institute, PO Box 7184, Newtown, Wellington 6242, New Zealand.
E-mail: swee.tan@gmri.org.nz

progenitor cells,⁵ mature cells, and from the fusion of stem cells and mutant cells.⁷

The stochastic model and the hierarchical model of cancer are not mutually exclusive, as the concept of cellular plasticity bridges the apparent dichotomy of the two concepts.⁸ Cancer cells are plastic and possess the ability to interconvert between differentiated and stem-like phenotypes, depending on a continuum of cell fates and the surrounding microenvironmental *niche*.⁸ It has also been observed that hierarchical cell populations contain transitory cells that change their position within the cellular hierarchy⁹ and that stochastic cell populations can generate hierarchically organized cell populations.¹⁰ Furthermore, cancer cells can dedifferentiate and enter the CSC pool to gain tumor repopulation capability. This may be inherited, as proposed in the hierarchical concept by which cancer cells are endowed with intrinsic stem cell properties, rather than acquiring them in a stepwise fashion. Alternatively, this capability may be acquired by gene mutations, as proposed by the stochastic model.⁸

Metastasis is the primary cause of cancer death.¹¹ Metastasis, resistance to chemotherapy, radiotherapy,¹² and immunotherapy¹³ have been attributed to the presence of CSCs, which have been identified in many cancer types^{14–17} using stemness-associated markers and functional investigations.¹⁷ Several strategies to target CSCs have emerged and they have been investigated in clinical trials, including (1) monoclonal antibodies against CSC-specific surface biomarkers; CSC-associated signaling pathways such as the Notch (by various methods), Hedgehog, Wnt, and mitochondrial oxidative phosphorylation pathways, (2) inhibiting C-X-C chemokine receptor type 4 (CXCR4), part of the CSC microenvironment, (3) CSC-directed immunotherapies, such as chimeric antigen receptor T-cell therapy, and (4) immune checkpoint blockers targeting the receptors programmed cell death protein 1 and cytotoxic T-lymphocyte-associated protein 4.¹⁸ The activity of human epidermal growth factor receptor 2–targeted therapies and CDK4/6 inhibitors against CSCs has been recently realized.^{19,20}

The tumor microenvironment (TME) is an important regulator of CSCs.²¹ The TME contains immune cells such as tumor-associated macrophages (TAMs), tumor blood vessels and lymphatics, cancer-associated fibroblasts (CAFs), pericytes, and adipocytes.²² Intercellular communication within the TME is achieved with cytokines, growth factors, chemokines, inflammatory mediators, and remodeling enzymes in the tissue.²² Targeting non-malignant cells and the mediators of intercellular communication, and other TME components, has been proposed as a treatment strategy for cancer.²²

Understanding the interactions between CSCs and the TME is crucial, as CSCs are a key mediator of treatment resistance, tumor metastasis, and recurrence.⁵ Key interactions between CSCs and immune cells result in evasion of immune detection and elimination and can create an anti-inflammatory and/or protumorigenic microenvironment *milieu*.⁵ CSCs also alter the balance between protumorigenic and antitumor immune cell activity within the TME.⁵ CSCs achieve successful immune evasion through reduction of antigenicity by downregulating components involved in antigen presentation and processing, such as transport associated with antigen processing, and/or the major histocompatibility complex.²³ The downregulation of these components facilitates immune evasion of CSCs from the adaptive immune system, and thus their survival.⁵

The renin–angiotensin system (RAS) plays an important role in the TME through its effects on tumor cells, non-malignant cells, hypoxia, angiogenesis, and the inflammatory response.²¹ This endocrine system regulates blood pressure, blood volume, and electrolyte homeostasis²⁴ and functions at both systemic and local levels.²⁵ Physiologically, angiotensinogen is synthesized and released by the liver and is converted to angiotensin I (ATI) by renin, released by the juxtaglomerular apparatus of the kidney. ATI is converted to angiotensin II (ATII) by angiotensin-converting enzyme (ACE). ATII is the main effector for the RAS that binds to ATII receptor 1 (AT₁R) and ATII receptor 2 (AT₂R) to exert its physiological effects.²⁵ Angiotensin 1–7 (Ang1–7) is a cleavage product of ATII that maintains cardiovascular homeostasis by binding to the G protein–coupled receptor MAS.²⁶ ATII is converted to angiotensin III by aminopeptidase A. Bypass loops which include the proteases cathepsins B, D, and G, and chymase provide redundancies to this system²⁶ (Fig. 1).

The RAS is implicated in cancer,²⁷ fibrotic conditions,^{28,29} cardiovascular,³⁰ renal,³¹ and lung³² diseases. Components of the RAS^{33–35} and cathepsins B, D, and G that constitute bypass loops of the RAS^{36–38} are expressed by CSCs in many cancer types. Targeting CSCs by inhibiting the RAS, its bypass loops, and converging signaling pathways has been proposed as a potential treatment strategy for cancer by repurposing existing low-cost medications.^{25,26}

CSCs are also regulated by crucial stem cell pathways, including the Notch, Wnt, Hedgehog, and Hippo⁵ pathways. In clinical trials, some of these CSC-associated pathways have been targeted to regulate and eliminate CSCs, which sustain and drive tumor development. Given the RAS is also an important stem cell pathway, it may also play a role in directly regulating CSCs.³⁹

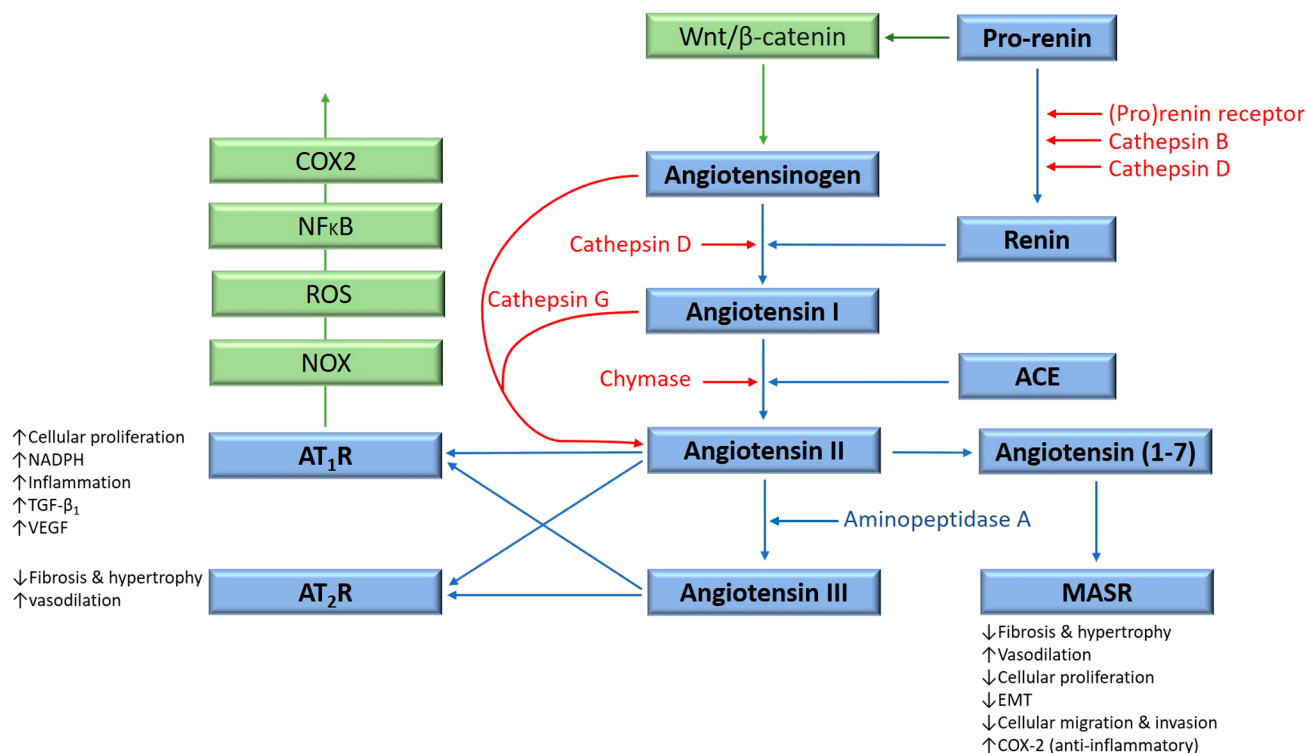


Figure 1. The renin–angiotensin system (RAS), its bypass loops, and converging signaling pathways. In the classical RAS (blue), angiotensinogen is converted to angiotensin I (ATI) by renin. ATI is converted to angiotensin II (ATII) by angiotensin-converting enzyme (ACE). ATII is the main effector for the RAS that binds to ATII receptor 1 (AT_1R) and ATII receptor 2 (AT_2R) to exert its physiological effects. Angiotensin 1–7 (Ang I–7) is a cleavage product of ATII which binds to the G protein–coupled receptor MAS receptor (MASR), which in turn exerts physiological effects. ATII is converted to angiotensin III (ATIII) by aminopeptidase A. ATII and ATIII bind to both AT_1R and AT_2R . Proteases including cathepsins B, D, and G, and chymase constitute bypass loops of the RAS (red), providing redundancies to this system. Other signaling pathways converge on the RAS (green) with the Wnt/ β -catenin signaling pathway being upstream of the RAS, activated by the prorenin receptor. Inflammatory signaling via the NOX-ROS-NF κ B-COX2 axis, downstream of the RAS, is induced by activation of AT_1R . Abbreviations: COX2, cyclooxygenase 2; ROS, reactive oxygen species; VEGF, vascular endothelial growth factor; TGF- β_1 , transforming growth factor β_1 ; EMT, epithelial-to-mesenchymal transition; NOX, NADPH oxidase; NF κ B, nuclear factor kappa B.

This article discusses the interaction between CSCs and the TME, and the role of the constituents of the TME, including immune cells and the RAS, in regulating the CSC *niche*. We propose a system-wide approach to targeting CSCs by exploiting the TME using inhibitors of the RAS, its bypass loops, and convergent signaling pathways.

Tumor Microenvironmental Niche and CSCs

CSCs reside in a *niche*, an anatomically distinct microenvironment within the overall TME. Cells within the *niche* produce factors that induce CSC self-renewal and angiogenesis. They also attract immune cells and other stromal cells that release growth factors, which promote tumor invasion and metastasis.^{8,40} Tumor development is influenced by a balance of protumorigenic factors and antitumor immune cell activity within

the TME. CSCs also influence this balance within the TME to drive and sustain tumorigenesis. CSCs and the TME are highly relevant clinically, as treatment resistance caused by CSCs^{12,13} ultimately leads to treatment failure and cancer death. Treatment resistance is partly attributed to quiescence by which CSCs are able to arrest the G0 phase of the cell cycle^{41,42}; upregulation of drug efflux pumps such as ATP-binding cassette (ABC) transporters, for example, ABCB1, ABCC1, and ABCG2⁴³; metabolic mediators⁴⁴; and activation of DNA repair mechanisms.⁴⁵

Interactions between CSCs and their microenvironmental *niche* also contribute to treatment resistance.⁴⁶ TME-induced conversion of non-CSCs into CSCs is one such interaction, underscoring the plasticity of non-CSCs that may contribute to therapy resistance.⁸ This highlights the capacity of cancer cells to acquire CSC characteristics under the influence of TME-related factors, for example, in ovarian cancer, in which

cancer cells acquire a CSC-like phenotype following cisplatin administration *in vitro*.⁴⁷ CSCs are sustained by several crucial stem cell signaling pathways, including the Notch, Wnt, Hedgehog, and Hippo pathways, which are often altered in CSCs,⁴⁸ supporting CSC traits such as maintenance and survival.⁴⁸ These pathways interact with oncogenic signaling pathways that function within the TME, including the nuclear factor kappa B (NF κ B), mitogen-activated protein kinase (MAPK), phosphatidylinositol-3 kinase, and epidermal growth factor receptor (EGFR) cascades,⁵ which are treatment targets for some cancers.⁴⁸ In addition, cathepsin-related signaling pathways operating within the TME may contribute to cancer growth and invasiveness. Cathepsins B, X, and K are involved in treatment resistance and tumor cell apoptosis,⁴⁹ while other cathepsins cause tumor suppression.⁵⁰ In particular, cathepsin K has been demonstrated in glioblastoma CSC *niches*.⁵¹ It is tightly regulated by cathepsin K activating proteases and other components of the TME, supporting a functional role of cathepsin K in the glioblastoma CSC *niche*.⁵¹ Cathepsin B is also highly active within glioblastoma CSC *niches*, which affects the interactions between glioblastoma cells and endothelial cells, and endothelial cell proliferation.⁵¹ Despite these findings, the precise effects of the functionally diverse cathepsins demonstrated in CSC *niches*, as in glioblastoma, remain unclear.

The Immune System and the TME

The ability of CSCs to evade immune detection and induce a protumorigenic environment⁵² is attributed to their interactions with immune cells within their *niche*. For example, TAMs induce proliferation of CSCs in hepatocellular carcinoma via interleukin 6 (IL-6)-induced activation of STAT3, which causes cytokine release, and a positive feedback loop that contributes to CSC self-renewal^{5,53} (Fig. 2). In breast cancer, the epithelial-to-mesenchymal transition (EMT) process upregulates EphA4 and the stem cell marker CD90, which mediate interactions between TAMs and CSCs by binding with their respective receptors.⁵⁴ Activation of the EphA4 receptor causes activation of Src and NF κ B.⁵⁴ NF κ B in CSCs induces the release of cytokines that sustain CSC properties.^{54,55} Myeloid-derived suppressor cells (MDSCs) are another immune cell type that support CSCs. In ovarian cancer, MDSCs promote CSC characteristics by causing microRNA-101 expression, inducing expression of stemness genes.⁵⁶

Glioma CSCs promote macrophage polarization toward the M2 phenotype, which is immunosuppressive and protumorigenic, by producing cytokines such

as transforming growth factor β and macrophage inhibitory cytokines.⁵⁷ Interestingly, CSCs can attenuate the anticancer properties of TAMs and dendritic cells.⁵ CSCs in malignant melanoma and glioblastoma inhibit proliferative T-cell responses and IL-2 production *in vitro*,^{58,59} and CSCs in head and neck squamous cell carcinoma cause an immunosuppressive and procancer effect through induced proliferation of regulatory T cells.⁵⁸ These interactions underscore the complex relationships between CSCs and their surrounding TME, and the many potential therapeutic targets that exist.

The RAS and the TME

The RAS participates in the TME to facilitate hallmarks of cancer, such as angiogenesis, tumor cell proliferation, and hypoxia.⁶⁰ This is largely mediated by binding of ATII to AT₁R, which increases cellular proliferation and vascular endothelial growth factor (VEGF) production, causing angiogenesis, inflammation, and oxidative stress.²¹ The local (paracrine) RAS can act independently of or synergistically with the endocrine RAS. Administration of ATII increases the ability of cells with a CSC phenotype to form spheroids, and growth in a 3D culture Matrigel-based assay *in vitro*.⁶¹ ATII also improves cell survival via an anchorage-independent process, and their expression of the stemness-associated marker CD133 is significantly upregulated.⁶¹ This shows the RAS is not only a critical regulator of stem cells, but also increases the stemness of cancer cells via the action of ATII.⁶¹ This and the fact that CSCs in multiple cancer types express components of the RAS and its bypass loops^{60,62–65} suggest the RAS and its convergent signaling pathways play an important role in regulating CSCs within the TME.²⁶

ATII causes cellular proliferation by activating MAPK-STAT3⁶⁶ and phosphatidylinositol signaling via AT₁R, to increase cytosolic Ca²⁺ which in turn affects mitogenesis⁶⁷ (Fig. 2). ATII also affects cell growth by modulating tyrosine kinases⁶⁸ and stimulates migration of fibroblasts via activation of AT₁R in an EGFR activation-dependent process.^{21,69} In a breast cancer model, AT₁R signaling promotes tumor cell contraction, migration, and invasion, by enhancing the expression of CXCR4—a chemokine receptor involved in the attraction of tumor cells to lymph nodes⁷⁰ (Fig. 2). AT₁R signaling also increases expression of focal adhesion kinase (FAK)/RhoA signaling molecules and their downstream effectors ROCK1, ROCK2, and MLC, which cause cell contraction and pseudopodia formation, allowing cell migration and invasion. This is relevant as FAK/RhoA signaling molecules are key

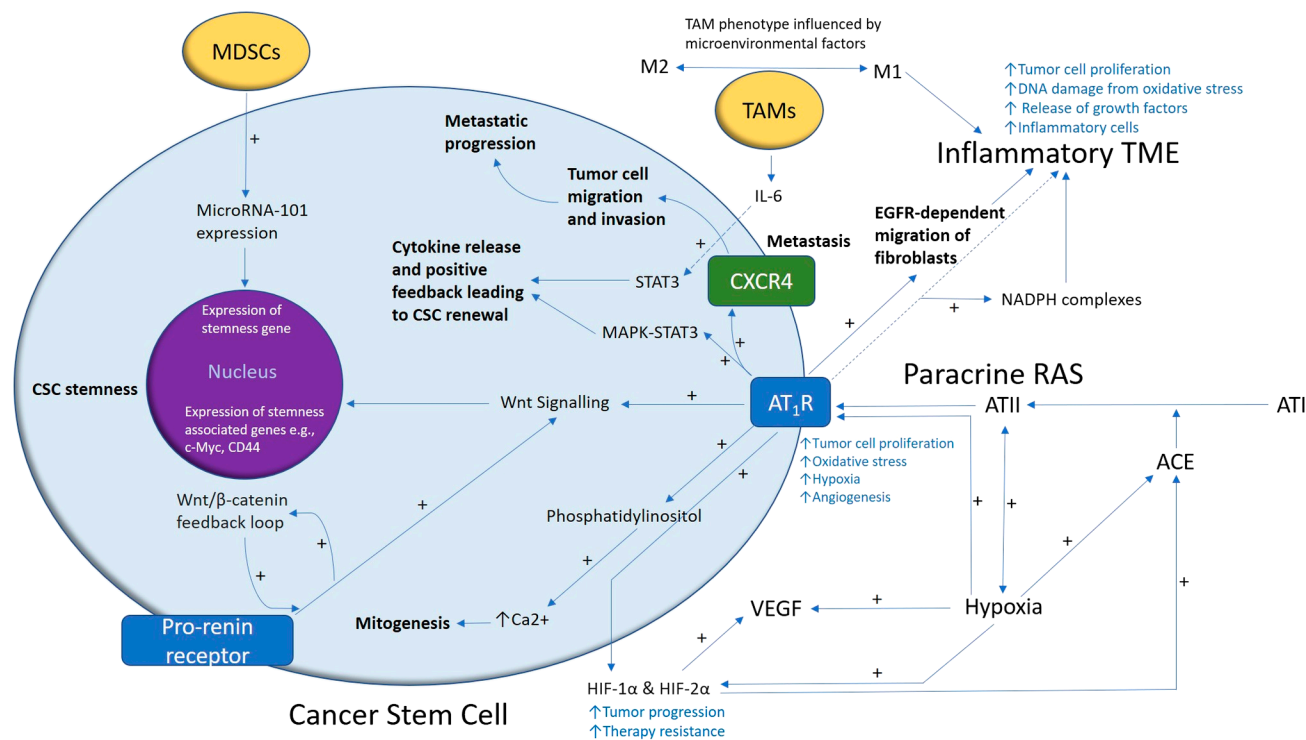


Figure 2. A proposed model demonstrating the role of the renin–angiotensin system (RAS) in cancer stem cell (CSC) niche. A CSC (with the cytoplasm in light blue and the nucleus in purple) residing within the tumor microenvironment (TME). Angiotensin II (ATII), the physiologically active end-product of the paracrine RAS, activates ATII receptor I (AT₁R) leading to increased tumor cell proliferation, oxidative stress, hypoxia and angiogenesis, and inflammation — the hallmarks of cancer. This contributes to an inflammatory TME by increasing the number of inflammatory cells, partly by increasing the number of NADPH complexes, leading to tumor cell proliferation, DNA damage from oxidative stress, and release of growth factors. AT₁R also activates phosphatidylinositol signaling which increases cytosolic Ca²⁺ to promote mitogenesis. Hypoxia increases paracrine RAS activity by upregulating angiotensin-converting enzyme (ACE) and the expression of hypoxia-inducible factor 1α (HIF-1α) and HIF-2α, which increase tumor progression and treatment resistance. HIF-1α, HIF-2α, and hypoxia increase the expression of vascular endothelial growth factor (VEGF). VEGF increases angiogenesis. Binding of AT₁R to C-X-C chemokine receptor type 4 (CXCR4) promotes tumor cell migration and invasion, leading to metastatic progression. AT₁R, via MAPK-STAT3 signaling, contributes to a cytokine release that leads to CSC renewal. AT₁R signaling also contributes to the migration of fibroblasts in an epidermal growth factor receptor (EGFR)-dependent fashion. AT₁R signaling and the prorenin receptor, which act in a feedback loop with Wnt/β-catenin, increase Wnt signaling which promotes CSC stemness by upregulating stemness-associated markers. Myeloid-derived suppressor cells (MDSCs) promote CSC characteristics by increasing microRNA-101 expression that induces expression of stemness-related genes in CSCs. Under the influence of the TME, polarization of tumor-associated macrophages (TAMs) — immune cells that are located within the TME — changes from the M1 to M2 phenotype. M2 TAMs induce the proliferation of CSCs via interleukin 6 (IL-6)-induced activation of STAT3, leading to cytokine release and positive feedback contributing to CSC renewal. Abbreviations: ATI, angiotensin I; AT₂R, ATII receptor 2; AngI–7, angiotensin I–7; ATIII, angiotensin III; MAPK, mitogen-activated protein kinase.

promoters of directional cell movement.⁷⁰ CXCR4 signaling is crucial for AT₁R-induced lymph node metastasis in breast cancer.⁷⁰ These observations support the use of RAS modulating agents to mitigate cancer and metastasis.

The TME is often hypoxic, which promotes tumor progression and treatment resistance, largely via the effects of hypoxia-inducible factor 1α (HIF-1α) and HIF-2α²¹ (Fig. 2). RAS activation in the TME contributes to hypoxia by ATII-mediated production of reactive oxygen species (ROS)⁵² which create a proinflammatory environment containing proangiogenic cytokines.^{27,52}

Hypoxia upregulates ATII, ACE, and AT₁R.^{71,72} ATII activates HIF-1α via AT₁R activation, leading to upregulated expression of both VEGF and ACE. This increases ATII formation that binds to and increases AT₁R signaling.^{69,73} The effect of the RAS in increasing the expression of VEGFα is supported by the reduction of VEGF-A mRNA and protein, and reduced tumor growth in lung tumors following ACE inhibitor (ACEI) administration in vivo.⁷⁴ Furthermore, administration of an AT₁R antagonist prevents an ATII-mediated increase in VEGF-A mRNA and protein in vivo.⁷⁴

Inflammation contributes to the TME through various mechanisms, notably the higher number of inflammatory cells, increased ROS, DNA damage, growth factors, and other inflammatory *milieu*.²¹ The RAS influences inflammation⁷⁵ by activation of AT₁R by ATII. This causes oxidant signaling via NADPH oxidase (NOX) complexes that play a role in inflammation.^{27,76} Components of the RAS are expressed by CSCs and non-CSCs, and stromal cells such as TAMs and CAFs. The expression of the RAS by TAMs is relevant, as these cells can either increase or decrease inflammation depending on their polarization. M1 macrophages produce a large amount of proinflammatory cytokines.⁷⁷ The TME can polarize M1 macrophages to form M2 macrophages, which reduce inflammation, and increase angiogenesis and tissue repair.⁷⁷ The RAS may also contribute to an immunosuppressive TME by reducing infiltration of TAMs.⁷⁸

EMT is a critical process in carcinogenesis and metastasis.⁷⁹ The involvement of the RAS in EMT and cellular differentiation is demonstrated in kidney disease and metastatic colorectal cancer to the liver.⁸⁰ Furthermore, high levels of ATII induce EMT in tubular cells toward a myfibroblastic phenotype in chronic nephropathy; this causes renal fibrosis, a process prevented by ACE inhibitor (ACEI) and angiotensin receptor blocker (ARB) administration.⁸¹ Treatment of human colorectal cancer with ARBs decreases the EMT markers ZEB1, vimentin, and MM-9.⁸⁰ The effects of RAS inhibition in tumor models and cell lines have been demonstrated in many cancer types, for example, inhibition of tumor cell growth, proliferation and migration, metastasis, and prolonged survival in tumor-bearing animals.²⁶ Administration of losartan, an AT₁R inhibitor, improves delivery of chemotherapy agents to breast cancer and pancreatic cancer xenografts, via a tumor stroma-mediated reduction in matrix stiffness and decompression of tumor microvessels.⁸² The finding of reduced tumor growth upon transplantation of melanoma cells into Agtr1^{-/-} mice further supports the critical role of the RAS in the TME.⁸³ Candesartan, an ARB, significantly reduces angiogenesis and metastases in a mouse lung cancer model⁸⁴ and prostate⁸⁵ and ovarian⁸⁶ cancer xenograft models. Reduction in tumor growth rates by losartan has also been observed in multiple cancer types.^{82,87} Irbesartan, another ARB, reduces tumor growth and angiogenesis in metastases of colorectal cancer to the liver in an *in vivo* model.⁸⁸

The RAS has multiple effects on the TME and cancer cells, although it is not clear whether this critical endocrine cascade also directly affects the stemness of CSCs. The RAS is important for the maintenance and differentiation of stem cells²⁶ and has been shown

to drive differentiation of mesenchymal stem cells into adipocytes.⁸⁹ ACE is required for hemangioblast expansion and modulation of AT₁R and AT₂R signaling which directly affects differentiation of the blasts toward a hematopoietic or endothelial lineage.⁹⁰ The RAS also plays an important role in other developmental processes involving stem cells^{91–93} including vasculogenesis, myeloid differentiation, and erythropoiesis.^{91–94} The RAS acts in a feedback loop with the Wnt/ β -catenin pathway, as prorenin can induce Wnt/ β -catenin (Fig. 2). This is important for embryonic development, as it induces differentiation of pluripotent stem cells.⁹⁵ This is relevant, as CSCs express stemness-associated markers such as the Yamanaka factors⁹⁶ and because components of the RAS are targets of Wnt/ β -catenin. Wnt signaling is implicated in cancer, and its end targets include the stemness-associated markers CD44 and c-MYC.⁹⁷ It could be that CSCs require activation of Wnt signaling, as seen in leukemia stem cells in acute myeloid leukemia.⁹⁸ This suggests that RAS modulators could be used to inhibit Wnt signaling and its downstream effects on CD44 and c-MYC, and possibly other stemness-associated markers, expressed by CSCs.^{26,99,100} Animals treated with losartan develop tumors containing more differentiated cells compared with untreated tumors which contain less differentiated cells.¹⁰¹ This supports the possible influence of the RAS on cancer cell differentiation.¹⁰¹ Furthermore, 20% of animals in the losartan treatment group are tumor free, compared with control animals, by the end of the study at 100 days.¹⁰¹ This suggests that AT₁R signaling is important for tumor progression, the loss of differentiation in tumor cells, and possibly the stemness of CSCs. Further research is required to determine the precise role of the Wnt/ β -catenin pathway within the RAS, in regulating CSCs within the CSC *niche*. These findings call for further investigation of RAS inhibition as a potential therapeutic option for treating cancer, with clinical trials.

RAS Inhibitors and Cancer

Hypertension is common in cancer patients, and given the widespread use of RAS modulators for cardiovascular and renal diseases, their effects on cancer incidence and outcomes have been investigated in many observational studies.⁹⁴ Numerous studies have shown that RAS inhibitors (RASIs), especially ACEIs and ARBs, used for the treatment of hypertension, improve not only cardiovascular disease outcomes but also survival outcomes for cancer patients (Table 1). For example, RASIs have been associated with improved survival of patients with non-small cell lung cancer in some studies^{107,108} although not in others.¹⁰⁹

Table 1. Studies Investigating the Association Between RASIs and Cancer Incidence and Outcomes.

Authors	Year	Study Type	Patient Number	RASI(s) Used	Outcome
Jung et al. ¹⁰²	2021	Cohort study	293,962	ARB	Decreased overall risk of cancer compared with patients who use ACEIs (HR = 0.76, 95% CI: 0.72–0.80).
Sun et al. ¹⁰³	2017	Meta-analysis	55 studies ^a	ACEIs and ARBs	Significant improvement in overall survival (HR = 0.82; 95% CI: 0.77–0.88; $p < 0.001$), progression free survival (HR = 0.74; 95% CI: 0.66–0.84; $p < 0.001$), and disease-free survival (HR = 0.80; 95% CI: 0.67–0.95; $p = 0.01$) compared with non-users.
Sipahi et al. ¹⁰⁴	2011	Meta-analysis	61,774	ACEI	No association with cancer occurrence or cancer-related deaths.
Collaboration AT ¹⁰⁵	2011	Meta-analysis	138,769	ARB	No effect on overall incidence of lung, breast, and prostate cancer.
Sipahi et al. ¹⁰⁶	2010	Meta-analysis	161,917	ARB	Increased risk of lung cancer (RR = 1.08, 95% CI: 1.01–1.18, $p = 0.001$).

Abbreviations: HR, hazard ratio; ARB, angiotensin receptor blocker; ACEI, angiotensin-converting enzyme inhibitor; RASI, renin–angiotensin system inhibitor; RR, risk ratio; CI, confidence interval.

^aTotal participant number not stated.

ACEI administration has not been shown to change the incidence of cancer in several studies.^{104,106,110} A meta-analysis of 161,917 patients from 13 studies reports an increased risk of lung cancer in patients who were administered ARB.¹⁰⁶ Another meta-analysis of 61,774 patients from 14 studies concludes that ACEI therapy does not affect cancer occurrence or cancer-related deaths.¹⁰⁴ The incidence of certain cancers increases in organ transplant recipients, with immunosuppressive therapy being a major risk factor. For example, cutaneous squamous and basal cell carcinomas are common in renal transplantation patients. ACEI or ARB use has been associated with an approximate 2-fold risk reduction of skin cancers in renal transplantation patients compared with non-users.¹¹¹ This association is supported by functional data, which show ATII blockade in an SCID-beige mouse pulmonary metastasis model effectively blocks cyclosporin-associated tumor progression in vivo.¹¹² A cohort study demonstrates ARB use is associated with a decreased overall risk of cancer compared with patients who use ACEI [hazard ratio (HR) = 0.76, 95% confidence interval (CI): 0.72–0.80] with a gradual dose–response relationship also being observed, and the cancer risk decreasing in those on prolonged ARB regimes compared with ACEI use.¹⁰²

Other studies show no improvement of outcomes of cancer patients administered with RASIs (Table 1). For example, a meta-analysis of 138,769 patients concludes that ARB administration does not affect the overall incidence of cancer including lung, breast, and prostate cancer or overall cancer deaths

in those treated with ARBs.¹⁰⁵ In contrast, a recent meta-analysis of 55 studies demonstrates a significant improvement in overall survival (HR = 0.82; 95% CI: 0.77–0.88; $p < 0.001$), progression-free survival (HR = 0.74; 95% CI: 0.66–0.84; $p < 0.001$), as well as disease-free survival (HR = 0.80; 95% CI: 0.67–0.95; $p = 0.01$) for cancer patients using RASIs compared with non-users. This study also shows the effect of RASIs is dependent on the cancer type, with differences demonstrated between hepatocellular carcinoma, renal cell carcinoma, gastric cancer, and bladder cancer.¹⁰³ These conflicting findings from observational studies highlight the need for further investigation with well-designed studies, including double-blinded randomized controlled trials to determine the effect of RASIs on the incidence and outcomes of cancer patients.

There is, however, substantial evidence demonstrating that RASIs decrease tumor growth and progression, angiogenesis, and metastasis.^{26,94} Existing in vitro and in vivo functional data consistently show an anticancer effect of RASIs, and the overall epidemiological data suggest RASIs could improve the survival of cancer patients with the degree depending on the cancer type and the type of RASI used. Further functional work using more sophisticated model systems, such as organoid systems, may advance our understanding of the mechanisms underpinning the relationship between CSCs and the TME, and particularly the RAS. Randomized controlled trials are warranted to establish the precise role of RASIs on cancer incidence and outcomes.

Therapeutic Targeting of the CSCs: Future Directions

Despite the emergence of new targeted therapies for cancer such as small-molecule inhibitors, immunotherapy, and biochemotherapy (combination of chemotherapy and immunotherapy), treatment resistance and treatment failure remain an unsolved clinical problem. For example, treatment of BRAF V600E mutant malignant melanoma with a BRAF inhibitor, or in combination with an MEK inhibitor, results mostly in an initial partial treatment response due to genetic and epigenetic mechanisms determined by the TME, as well as immunogenic and transcriptomic alterations.^{113,114} These changes may contribute to reactivation of MAPK signaling and subsequent treatment resistance.¹¹⁵ Furthermore, targeted therapy is not always successful due to the high intratumor genetic and epigenetic heterogeneity, and cellular plasticity. This increases the possibility of switching a cancer cell into a slow-cycle state, which causes treatment resistance. Non-coding RNAs also contribute to treatment resistance by becoming resistant to anti-BRAF treatment, influenced by dynamic deregulation of microRNA. In turn, this leads to alteration of cell survival pathways and enhancement of proinflammatory and proangiogenic cues.¹¹⁶ These resistance mechanisms underscore the need for a holistic system-wide approach that affects the tumor-forming capacity and stemness of CSCs, the proposed origin of cancer and metastasis.

The RAS plays a critical role in the CSC *niche* that regulates CSCs. In addition to the presence of bypass loops that provide redundancies, there are several inflammatory and developmental signaling pathways that converge on the RAS. For example, prorenin receptor induces Wnt/ β -catenin signaling within a feedback loop. AT₁R signaling can also cause inflammatory signaling via the NOX-ROS-NF κ B-COX2 (cyclooxygenase 2) axis²⁶ (Fig. 1). Each primary step of this axis could be targeted using NOX inhibitors, ROS inhibitors, metformin, and nonsteroidal anti-inflammatory drugs, respectively (Fig. 1). Wnt/ β -catenin can also be targeted by its respective inhibitor.²⁶ These axes and many steps of the RAS and its bypass loops can be inhibited using existing, low-cost medications. For example, renin secretion can be inhibited by β -blockers such as propranolol,¹¹⁷ ACE can be targeted with ACEIs, and AT₁R with ARBs. Enzymes that constitute bypass loops of the RAS such as cathepsins B, D, and G, and chymase can be blocked by their respective inhibitors. The MAS receptor has been demonstrated to have anticancer effects, making it a potential treatment target. Aminopeptidase A may be targeted with aminopeptidase A inhibitors.

This holistic system-wide approach of targeting the RAS, its bypass loops expressed on CSCs, and convergent signaling pathways, to regulate the TME that affects the tumor-forming capacity and stemness of CSCs, warrants further functional work with in vitro and in vivo models such as organoid systems. This treatment approach is currently being tested in a phase I/II clinical trial on glioblastoma and metastatic melanoma.²⁵

Competing Interests

The author(s) declared the following potential conflicts of interest with respect to the research, authorship, and/or publication of this article: S.T. is an inventor of the patents Cancer Diagnosis and Therapy (PCT/NZ2015/050108), Cancer Therapeutic (PCT/NZ2018/050006), Novel Pharmaceutical Compositions for Cancer Therapy (US/62/711709), and Cancer diagnosis and therapy (United States Patent No. 10281472).

Author Contributions

EJK drafted the manuscript and STT critically reviewed and commented on the manuscript. The authors approved the final manuscript.

Funding

The author(s) received no financial support for the research, authorship, and/or publication of this article.

Literature Cited

1. Torre LA, Siegel RL, Ward EM, Jemal A. Global cancer incidence and mortality rates and trends—an update. *Cancer Epidemiology Biomarkers Prev.* 2016;25(1):16–27.
2. Pfister DG. The just price of cancer drugs and the growing cost of cancer care: oncologists need to be part of the solution. *J Clin Oncol.* 2013;31(28):3487–9.
3. Aponte PM, Caicedo A. Stemness in cancer: stem cells, cancer stem cells, and their microenvironment. *Stem Cells Int.* 2017;2017:5619472.
4. Gurel C, Inetas G, Hortu I, Tunc E, Kuscu GC, Dindaroglu FC, Sahin O, Buhur A, Oktem G. Cancer and cancer stem cells: new molecular perspectives. *Crit Rev Oncog.* 2019;24(1):99–104.
5. Clara JA, Monge C, Yang Y, Takebe N. Targeting signalling pathways and the immune microenvironment of cancer stem cells—a clinical update. *Nat Rev Clin Oncol.* 2020;17(4):204–32.
6. Bradshaw A, Wickremesekera A, Brasch HD, Chibnall AM, Davis PF, Tan ST, Itinteang T. Cancer stem cells in glioblastoma multiforme. *Front Surg.* 2016;3:48.
7. Eun K, Ham SW, Kim H. Cancer stem cell heterogeneity: origin and new perspectives on CSC targeting. *BMB Rep.* 2017;50(3):117–25.

8. Plaks V, Kong N, Werb Z. The cancer stem cell niche: how essential is the niche in regulating stemness of tumor cells? *Cell Stem Cell*. 2015;16(3):225–38.
9. Quail DF, Taylor MJ, Postovit L-M. Microenvironmental regulation of cancer stem cell phenotypes. *Curr Stem Cell Res Ther*. 2012;7(3):197–216.
10. Chaffer CL, Weinberg RA. How does multistep tumorigenesis really proceed? *Cancer Discov*. 2015;5(1):22–4.
11. Chaffer CL, Weinberg RA. A perspective on cancer cell metastasis. *Science*. 2011;331(6024):1559–64.
12. Chang JC. Cancer stem cells: role in tumor growth, recurrence, metastasis, and treatment resistance. *Medicine*. 2016;95(suppl. 1):S20–5.
13. Shen C, Yang C, Xia B, You M. Long non-coding RNAs: emerging regulators for chemo/immunotherapy resistance in cancer stem cells. *Cancer Lett*. 2020;500:244–52.
14. Kilmister EJ, Patel J, van Schaijik B, Bockett N, Brasch HD, Paterson E, Sim D, Davis PF, Roth IM, Itinteang T, Tan ST. Cancer stem cell subpopulations are present within metastatic head and neck cutaneous squamous cell carcinoma. *Front Oncol*. 2020;10:1091.
15. Yoganandarajah V, Patel J, van Schaijik B, Bockett N, Brasch HD, Paterson E, Sim D, Davis PF, Roth IM, Itinteang T, Tan ST. Identification of cancer stem cell subpopulations in head and neck metastatic malignant melanoma. *Cells*. 2020;9(2):324.
16. Korkaya H, Liu S, Wicha MS. Breast cancer stem cells, cytokine networks, and the tumor microenvironment. *J Clin Invest*. 2011;121(10):3804–3809.
17. Abbaszadegan MR, Bagheri V, Razavi MS, Momtazi AA, Sahebkar A, Gholamin M. Isolation, identification, and characterization of cancer stem cells: a review. *J Cell Physiol*. 2017;232(8):2008–18.
18. Yang L, Shi P, Zhao G, Xu J, Peng W, Zhang J, Zhang G, Wang X, Dong Z, Chen F, Cui H. Targeting cancer stem cell pathways for cancer therapy. *Signal Transduct Target Ther*. 2020;5(1):8.
19. Korkaya H, Paulson A, Iovino F, Wicha MS. HER2 regulates the mammary stem/progenitor cell population driving tumorigenesis and invasion. *Oncogene*. 2008;27(47):6120–30.
20. Dai M, Zhang C, Ali A, Hong X, Tian J, Lo C, Fils-Aimé Burgos SA, Ali S, Lebrun J-J. CDK4 regulates cancer stemness and is a novel therapeutic target for triple-negative breast cancer. *Sci Rep*. 2016;6(1):35383.
21. Catarata MJ, Ribeiro R, Oliveira MJ, Robalo Cordeiro C, Medeiros R. Renin-angiotensin system in lung tumor and microenvironment interactions. *Cancers*. 2020;12(6):1457.
22. Balkwill FR, Capasso M, Hagemann T. The tumor microenvironment at a glance. *J Cell Sci*. 2012;125(23):5591–6.
23. Zhang D, Tang DG, Rycaj K. Cancer stem cells: regulation programs, immunological properties and immunotherapy. *Semin Cancer Biol*. 2018;52(Pt 2):94–106.
24. Fyhrquist F, Saijonmaa O. Renin-angiotensin system revisited. *J Intern Med*. 2008;264(3):224–36.
25. Tan DC, Roth IM, Wickremesekera AC, Davis PF, Kaye AH, Mantamadiotis T, Stylli SS, Tan ST. Therapeutic targeting of cancer stem cells in human glioblastoma by manipulating the renin-angiotensin system. *Cells*. 2019;8(11):1364.
26. Roth I, Wickremesekera AC, Wickremesekera SK, Davis PF, Tan ST. Therapeutic targeting of cancer stem cells via modulation of the renin-angiotensin system. *Front Oncol*. 2019;9:745.
27. George AJ, Thomas WG, Hannan RD. The renin-angiotensin system and cancer: old dog, new tricks. *Nat Rev Cancer*. 2010;10(11):745–59.
28. Kilmister EJ, Paterson C, Brasch H, Davis P, Tan S. The role of the renin-angiotensin system and vitamin D in keloid disorder—a review. *Front Surg*. 2019;6:67.
29. Murphy AM, Wong AL, Bezuhyly M. Modulation of angiotensin II signaling in the prevention of fibrosis. *Fibrogenesis Tissue Repair*. 2015;8(1):7.
30. Kobori H, Nangaku M, Navar LG, Nishiyama A. The intrarenal renin-angiotensin system: from physiology to the pathobiology of hypertension and kidney disease. *Pharmacol Rev*. 2007;59(3):251–87.
31. Brewster UC, Perazella MA. The renin-angiotensin-aldosterone system and the kidney: effects on kidney disease. *Am J Med*. 2004;116(4):263–72.
32. Shi Y, Liu T, Yao L, Xing Y, Zhao X, Fu J, Xue X. Chronic vitamin D deficiency induces lung fibrosis through activation of the renin-angiotensin system. *Sci Rep*. 2017;7(1):3312.
33. Munro MJ, Wickremesekera AC, Davis PF, Marsh R, Tan ST, Itinteang T. Renin-angiotensin system and cancer: a review. *Integr Cancer Sci Ther*. 2017;4(2):1–6.
34. Itinteang T, Dunne JC, Chibnall AM, Brasch HD, Davis PF, Tan ST. Cancer stem cells in moderately differentiated oral tongue squamous cell carcinoma express components of the renin-angiotensin system. *J Clin Pathol*. 2016;69(10):942–5.
35. Siljee S, Pilkington T, Brasch HD, Bockett N, Patel J, Paterson E, Davis PF, Tan ST. Cancer stem cells in head and neck metastatic malignant melanoma express components of the renin-angiotensin system. *Life*. 2020;10(11):268.
36. Featherston T, Marsh RW, van Schaijik B, Brasch HD, Tan ST, Itinteang T. Expression and localization of cathepsins B, D, and G in two cancer stem cell subpopulations in moderately differentiated oral tongue squamous cell carcinoma. *Front Med*. 2017;4:100.
37. Mehrotra S, Wickremesekera SK, Brasch HD, Van Schaijik B, Marsh RW, Tan ST, Itinteang T. Expression and localization of cathepsins B, D and G in cancer stem cells in liver metastasis from colon adenocarcinoma. *Front Surg*. 2018;5:40.
38. Koh SP, Wickremesekera AC, Brasch HD, Marsh R, Tan ST, Itinteang T. Expression of cathepsins B, D, and G in isocitrate dehydrogenase-wildtype glioblastoma. *Front Surg*. 2017;4:28.

39. Sadik NA-h Metwally NS, Shaker OG, Soliman MS, Mohamed AA, Abdelmoaty MM. Local renin-angiotensin system regulates the differentiation of mesenchymal stem cells into insulin-producing cells through angiotensin type 2 receptor. *Biochimie*. 2017;137:132–8.
40. Oskarsson T, Batlle E, Massagué J. Metastatic stem cells: sources, niches, and vital pathways. *Cell Stem Cell*. 2014;14(3):306–21.
41. Bao S, Wu Q, McLendon RE, Hao Y, Shi Q, Hjelmeland AB, Dewhirst MW, Bigner DD, Rich JN. Glioma stem cells promote radioresistance by preferential activation of the DNA damage response. *Nature*. 2006;444(7120):756–60.
42. Chen W, Dong J, Haiech J, Kilhoffer MC, Zeniou M. Cancer stem cell quiescence and plasticity as major challenges in cancer therapy. *Stem Cells Int*. 2016;2016:1740936.
43. Gottesman MM, Fojo T, Bates SE. Multidrug resistance in cancer: role of ATP-dependent transporters. *Nature Rev Cancer*. 2002;2(1):48–58.
44. Elyer CE, Rich JN. Survival of the fittest: cancer stem cells in therapeutic resistance and angiogenesis. *J Clin Oncol*. 2008;26(17):2839–45.
45. Diehn M, Cho RW, Lobo NA, Kalisky T, Dorie MJ, Kulp AN, Qian D, Lam JS, Ailles LE, Wong M, Joshua B, Kaplan MJ, Wapnir I, Dirbas FM, Somlo G, Garberoglio C, Paz B, Shen J, Lau SK, Quake SR, Brown JM, Weissman IL, Clarke MF. Association of reactive oxygen species levels and radioresistance in cancer stem cells. *Nature*. 2009;458(7239):780–3.
46. Schulenburg A, Blatt K, Cerny-Reiterer S, Sadovnik I, Herrmann H, Marian B, Grunt TW, Zielinski CC, Valent P. Cancer stem cells in basic science and in translational oncology: can we translate into clinical application? *J Hematol Oncol*. 2015;8:16.
47. Wiechert A, Saygin C, Thiagarajan PS, Rao VS, Hale JS, Gupta N, Hitomi M, Nagaraj AB, DiFeo A, Lathia JD, Reizes O. Cisplatin induces stemness in ovarian cancer. *Oncotarget*. 2016;7(21):30511–22.
48. Saygin C, Matei D, Majeti R, Reizes O, Lathia JD. Targeting cancer stemness in the clinic: from hype to hope. *Cell Stem Cell*. 2019;24(1):25–40.
49. Olson OC, Joyce JA. Cystein cathepsin proteases: regulators of cancer progression and therapeutic response. *Nat Rev Cancer*. 2015;15(12):712–29.
50. Lah TT, Durán Alonso MB, Van Noorden CJ. Antiproteases therapy in cancer: hot or not? *Expert Opin Biol Ther*. 2006;6(3):257–79.
51. Breznik B, Limebaeck Stokin C, Kos J, Khurshed M, Hira VVV, Bošnjak R, Lah TT, Van Noorden CJF. Cysteine cathepsins B, X and K expression in peri-arteriolar glioblastoma stem cell niches. *J Mol Histol*. 2018;49(5):481–97.
52. Pagé EL, Robitaille GA, Pouysségur J, Richard DE. Induction of hypoxia-inducible factor-1alpha by transcriptional and translational mechanisms. *J Biol Chem*. 2002;277(50):48403–48409.
53. Wan S, Zhao E, Kryczek I, Vatan L, Sadovskaya A, Ludema G, Simeone DN, Zou W, Welling TH. Tumor-associated macrophages produce interleukin 6 and signal via STAT3 to promote expansion of human hepatocellular carcinoma stem cells. *Gastroenterology*. 2014;147(6):1393–1404.
54. Lu H, Clauser KR, Tam WL, Fröse J, Ye X, Eaton EN, Reinhardt F, Donnenberg VS, Bhargava R, Carr SA, Weinburg RA. A breast cancer stem cell niche supported by juxtacrine signalling from monocytes and macrophages. *Nat Cell Biol*. 2014;16(11):1105–17.
55. Korkaya H, Liu S, Wicha MS. Regulation of cancer stem cells by cytokine networks: attacking cancer's inflammatory roots. *Clin Cancer Res*. 2011;17(19):6125–9.
56. Cui TX, Kryczek I, Zhao L, Zhao E, Kuick R, Roh MH, Vatan L, Szeliga W, Mao Y, Thomas DG, Kotarski J, Tarkowski R, Wicha M, Cho K, Giordano T, Liu R, Zou W. Myeloid-derived suppressor cells enhance stemness of cancer cells by inducing microRNA101 and suppressing the corepressor CtBP2. *Immunity*. 2013;39(3):611–21.
57. Jinushi M. Role of cancer stem cell-associated inflammation in creating pro-inflammatory tumorigenic micro-environments. *Oncoimmunology*. 2014;3:e28862.
58. Schatton T, Schütte U, Frank NY, Zhan Q, Hoerning A, Robles SC, Zhou J, Hodi FS, Spagnoli GC, Murphy GF, Frank MH. Modulation of T-cell activation by malignant melanoma initiating cells. *Cancer Res*. 2010;70(2):697–708.
59. Di Tomaso T, Mazzoleni S, Wang E, Sovena G, Clavenna D, Franzin A, Mortini P, Ferrone S, Doglioni C, Marincola FM, Galli R, Parmiani G, Maccalli C. Immunobiological characterization of cancer stem cells isolated from glioblastoma patients. *Clin Cancer Res*. 2010;16(3):800–13.
60. Wegman-Ostrosky T, Soto-Reyes E, Vidal-Millán S, Sánchez-Corona J. The renin-angiotensin system meets the hallmarks of cancer. *J Renin Angiotensin Aldosterone Syst*. 2015;16(2):227–33.
61. Tawinwung S, Ninsontia C, Chanvorachote P. Angiotensin II increases cancer stem cell-like phenotype in lung cancer cells. *Anticancer Res*. 2015;35(9):4789–97.
62. Bradshaw AR, Wickremesekera AC, Brasch HD, Chibnall AM, Davis PF, Tan ST, Itinteang T. Glioblastoma multiforme cancer stem cells express components of the renin-angiotensin system. *Front Surg*. 2016;3:51.
63. Wickremesekera AC, Brasch HD, Lee VM, Davis PF, Parker A, Koeck H, Itinteang T, Tan ST. Cancer stem cell subpopulations in metastatic melanoma to the brain express components of the renin-angiotensin system. *J Cancer Metastasis*. 2019;5:62.
64. Siljee SBO, Bocket N, van Shaijik B, Patel J, Brasch HD, Sim D, Davis PF, Itinteang T, Tan ST. Cancer stem cells in metastatic cutaneous head and neck squamous cell carcinoma express components of the renin-angiotensin system. *Cell*. 2021;10(2):243.

65. Narayanan A, Wickremesekera SK, van Schaijik B, Marsh RW, Brasch HD, Tan ST, Itinteang T. Cancer stem cells in liver metastasis from colon adenocarcinoma express components of the renin-angiotensin system. *J Cancer Metastasis Treat.* 2019;5:36.
66. Hashemzahi M, Beheshti F, Hassanian SM, Ferns GA, Khazaei M, Avan A. Therapeutic potential of renin angiotensin system inhibitors in cancer cells metastasis. *Pathol Res Pract.* 2020;216(7):153010.
67. Yu C, Tang W, Wang Y, Shen Q, Wang B, Cai C, Meng X, Zou F. Downregulation of ACE2/ang-(1-7)/mas axis promotes breast cancer metastasis by enhancing store-operated calcium entry. *Cancer Lett.* 2016;376(2):268–77.
68. Lewandowska U, Lachowicz-Ochędalska A, Domińska K, Kaszewska D, Rębas E. Angiotensin II as a factor modulating protein tyrosine kinase activity in two breast cancer lines—MCF-7 and MDA-MB-231. *Endokrynol Pol.* 2011;62(2):151–8.
69. Yahata Y, Shirakata Y, Tokumaru S, Yang L, Dai X, Tohyama M, Tsuda T, Sayama K, Iwai M, Horiuchi M, Hashimoto K. A novel function of angiotensin II in skin wound healing. Induction of fibroblast and keratinocyte migration by angiotensin II via heparin-binding epidermal growth factor (EGF)-like growth factor-mediated EGF receptor transactivation. *J Biol Chem.* 2006;281(19):13209–16.
70. Ma Y, Xia Z, Ye C, Lu C, Zhou S, Pan J, Liu C, Zhang J, Liu T, Hu T, Xie L, Wu G, Zhao Y. AGTR1 promotes lymph node metastasis in breast cancer by upregulating CXCR4/SDF-1 α and inducing cell migration and invasion. *Aging.* 2019;11(12):3969–92.
71. Fung ML, Lam SY, Dong X, Chen Y, Leung PS. Postnatal hypoxemia increases angiotensin II sensitivity and up-regulates AT1a angiotensin receptors in rat carotid body chemoreceptors. *J Endocrinol.* 2002;173(2):305–13.
72. Zhang R, Wu Y, Zhao M, Liu C, Zhou L, Shen S, Liao S, Yang K, Li Q, Wan H. Role of HIF-1 α in the regulation ACE and ACE2 expression in hypoxic human pulmonary artery smooth muscle cells. *Am J Physiol Lung Cell Mol Physiol.* 2009;297(4):L631–40.
73. Liu C, Zhang J-W, Hu L, Song Y-C, Zhou L, Fan Y, Zhu H-Y, Wang Y, Li Q-P. Activation of the AT1R/HIF-1 α /ACE axis mediates angiotensin II-induced VEGF synthesis in mesenchymal stem cells. *BioMed Res Int.* 2014;2014:627380.
74. Imai N, Hashimoto T, Kihara M, Yoshida S-i Kawana I, Yazawa T, Kitamura H, Umemura S. Roles for host and tumor angiotensin II type 1 receptor in tumor growth and tumor-associated angiogenesis. *Lab Invest.* 2007;87(2):189–98.
75. Husain K, Hernandez W, Ansari RA, Ferder L. Inflammation, oxidative stress and renin angiotensin system in atherosclerosis. *World J Biol Chem.* 2015;6(3):209–17.
76. Nakamura K, Yaguchi T, Ohmura G, Kobayashi A, Kawamura N, Iwata T, Kiniwa Y, Okuyama R, Kawakami Y. Involvement of local renin-angiotensin system in immunosuppression of tumor microenvironment. *Cancer Sci.* 2018;109(1):54–64.
77. Mantovani A, Sozzani S, Locati M, Allavena P, Sica A. Macrophage polarization: tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes. *Trends Immunol.* 2002;23(11):549–55.
78. Pinter M, Jain RK. Targeting the renin-angiotensin system to improve cancer treatment: implications for immunotherapy. *Sci Transl Med.* 2017;9(410):eaan5616.
79. Biddle A, Mackenzie IC. Cancer stem cells and EMT in carcinoma. *Cancer Metastasis Rev.* 2012;31(1):285–93.
80. Nguyen L, Ager EI, Neo J, Christophi C. Regulation of colorectal cancer cell epithelial to mesenchymal transition by the renin angiotensin system. *J Gastroenterol Hepatol.* 2016;31(10):1773–82.
81. Burns WC, Velkoska E, Dean R, Burrell LM, Thomas MC. Angiotensin II mediates epithelial-to-mesenchymal transformation in tubular cells by ANG 1–7/MAS-1-dependent pathways. *Am J Physiol Renal Physiol.* 2010;299(3):F585–93.
82. Chauhan VP, Martin JD, Liu H, Lacorre DA, Jain SR, Kozin SV, Stylianopoulos T, Mousa AS, Han X, Adstamongkonkul P, Popović Z, Huang P, Bawendi MG, Boucher Y, Jain RK. Angiotensin inhibition enhances drug delivery and potentiates chemotherapy by decompressing tumour blood vessels. *Nat Commun.* 2013;4:2516.
83. Egami K, Murohara T, Shimada T, Sasaki KI, Shintani S, Sugaya T, Ishii M, Akagi T, Ikeda H, Matsuishi T, Imaizumi T. Role of host angiotensin II type 1 receptor in tumor angiogenesis and growth. *J Clin Invest.* 2003;112(1):67–75.
84. Fujita M, Hayashi I, Yamashina S, Itoman M, Majima M. Blockade of angiotensin AT1a receptor signaling reduces tumor growth, angiogenesis, and metastasis. *Biochem Biophys Res Commun.* 2002;294(2):441–7.
85. Kosaka T, Miyajima A, Takayama E, Kikuchi E, Nakashima J, Ohigashi T, Asano T, Sakamoto M, Okita H, Murai M, Hayakawa M. Angiotensin II type 1 receptor antagonist as an angiogenic inhibitor in prostate cancer. *Prostate.* 2007;67(1):41–9.
86. Sukanuma T, Ino K, Shibata K, Kajiyama H, Nagasaka T, Mizutani S, Kikkawa F. Functional expression of the angiotensin II type 1 receptor in human ovarian carcinoma cells and its blockade-therapy resulting in suppression of tumor invasion, angiogenesis, and peritoneal dissemination. *Clin Cancer Res.* 2005;11(7):2686–94.
87. Rhodes DR, Ateeq B, Cao Q, Tomlins SA, Mehra R, Laxman B, Kalyana-Sundaram S, Lonigro RJ, Helgeson BE, Bhojani MS, Rehmtulla A, Kleer CG, Hayes DF, Lucas PC, Varambally S, Chinnaiyan

- AM. AGTR1 overexpression defines a subset of breast cancer and confers sensitivity to losartan, an AGTR1 antagonist. *Proc Natl Acad Sci USA*. 2009;106(25):10284.
88. Neo JH, Malcontenti-Wilson C, Muralidharan V, Christophi C. Effect of ACE inhibitors and angiotensin II receptor antagonists in a mouse model of colorectal cancer liver metastases. *J Gastroenterol Hepatol*. 2007;22(4):577–84.
 89. Matsushita K, Wu Y, Okamoto Y, Pratt RE, Dzau VJ. Local renin angiotensin expression regulates human mesenchymal stem cell differentiation to adipocytes. *Hypertension*. 2006;48(6):1095–1102.
 90. Zambidis ET, Soon Park T, Yu W, Tam A, Levine M, Yuan X, Pryzhkova M, Péault B. Expression of angiotensin-converting enzyme (CD143) identifies and regulates primitive hemangioblasts derived from human pluripotent stem cells. *Blood*. 2008;112(9):3601–14.
 91. Walter J. Does captopril decrease the effect of human recombinant erythropoietin in haemodialysis patients? *Nephrol Dial Transplant*. 1993;8(12):1428.
 92. Khakoo AY, Sidman RL, Pasqualini R, Arap W. Does the renin-angiotensin system participate in regulation of human vasculogenesis and angiogenesis? *Cancer Res*. 2008;68(22):9112–5.
 93. Hubert C, Savary K, Gasc JM, Corvol P. The hematopoietic system: a new niche for the renin-angiotensin system. *Nat Clin Pract Cardiovasc Med*. 2006;3(2):80–5.
 94. Perini MV, Dmello RS, Nero TL, Chand AL. Evaluating the benefits of renin-angiotensin system inhibitors as cancer treatments. *Pharmacol Ther*. 2020;211:107527.
 95. Nusse R. Wnt signaling and stem cell control. *Cell Res*. 2008;18(5):523–7.
 96. Takahashi K, Tanabe K, Ohnuki M, Narita M, Ichisaka T, Tomoda K, Yamanaka S. Induction of pluripotent stem cells from adult human fibroblasts by defined factors. *Cell*. 2007;131(5):861–72.
 97. Kim J-H, Park S-Y, Jun Y, Kim J-Y, Nam J-S. Roles of Wnt target genes in the journey of cancer stem cells. *Int J Mol Sci*. 2017;18(8):1604.
 98. Wang Y, Krivtsov AV, Sinha AU, North TE, Goessling W, Feng Z, Zon LI, Armstrong SA. The Wnt/beta-catenin pathway is required for the development of leukemia stem cells in AML. *Science*. 2010;327(5973):1650–3.
 99. Bradshaw A, Wickremsekera A, Tan ST, Peng L, Davis PF, Itinteang T. Cancer stem cell hierarchy in glioblastoma multiforme. *Front Surg*. 2016;3:21.
 100. van Schaijik B, Davis PF, Wickremsekera AC, Tan ST, Itinteang T. Subcellular localisation of the stem cell markers OCT4, SOX2, NANOG, KLF4 and c-MYC in cancer: a review. *J Clin Pathol*. 2018;71(1):88–91.
 101. Coulson R, Liew SH, Connelly AA, Yee NS, Deb S, Kumar B, Vargas AC, O'Toole SA, Parslow AC, Poh A, Putoczki T, Morrow RJ, Alorro M, Lazarus KA, Yeap EFW, Walton KL, Harrison CA, Hannan NJ, George AJ, Clyne CD, Ernst M, Allen AM, Chand AL. The angiotensin receptor blocker, Losartan, inhibits mammary tumor development and progression to invasive carcinoma. *Oncotarget*. 2017;8(12):18640–56.
 102. Jung MH, Lee JH, Lee CJ, Shin JH, Kang SH, Kwon CH, Kim DH, Kim WH, Kim HL, Kim HM, Cho IJ, Cho I, Hwang J, Ryu S, Kang C, Lee HY, Chung WJ, Ihm SH, Kim KI, Cho EJ, Sohn IS, Park S, Shin J, Ryu SK, Rhee MY, Kang SM, Pyun WB, Cho MC, Sung KC. Effect of angiotensin receptor blockers on the development of cancer: a nationwide cohort study in Korea. *J Clin Hypertens*. 2021;23:879–87.
 103. Sun H, Li T, Zhuang R, Cai W, Zheng Y. Do renin-angiotensin system inhibitors influence the recurrence, metastasis, and survival in cancer patients? Evidence from a meta-analysis including 55 studies. *Medicine*. 2017;96(13):e6394.
 104. Sipahi I, Chou J, Mishra P, Debanne SM, Simon DI, Fang JC. Meta-analysis of randomized controlled trials on effect of angiotensin-converting enzyme inhibitors on cancer risk. *Am J Cardiol*. 2011;108(2):294–301.
 105. Collaboration AT. Effects of telmisartan, irbesartan, valsartan, candesartan, and losartan on cancers in 15 trials enrolling 138,769 individuals. *J Hypertension*. 2011;29(4):623–35.
 106. Sipahi I, Debanne SM, Rowland DY, Simon DI, Fang JC. Angiotensin-receptor blockade and risk of cancer: meta-analysis of randomised controlled trials. *Lancet Oncol*. 2010;11(7):627–36.
 107. Menter AR, Carroll NM, Sakoda LC, Delate T, Hornbrook MC, Jain RK, Kushi LH, Quinn VP, Ritzwoller DP. Effect of angiotensin system inhibitors on survival in patients receiving chemotherapy for advanced non-small-cell lung cancer. *Clin Lung Cancer*. 2017;18(2):189–97.e3.
 108. Wilop S, von Hobe S, Crysandt M, Esser A, Osieka R, Jost E. Impact of angiotensin I converting enzyme inhibitors and angiotensin II type 1 receptor blockers on survival in patients with advanced non-small-cell lung cancer undergoing first-line platinum-based chemotherapy. *J Cancer Res Clin Oncol*. 2009;135(10):1429–35.
 109. Aydiner A, Ciftci R, Sen F. Renin-Angiotensin system blockers may prolong survival of metastatic non-small cell lung cancer patients receiving erlotinib. *Medicine*. 2015;94(22):e887.
 110. Bangalore S, Kumar S, Kjeldsen SE, Makani H, Grossman E, Wetterslev J, Gupta AK, Sever PS, Gluud C, Messerli FH. Antihypertensive drugs and risk of cancer: network meta-analyses and trial sequential analyses of 324,168 participants from randomised trials. *Lancet Oncol*. 2011;12(1):65–82.
 111. Moscarelli L, Zanazzi M, Mancini G, Rossi E, Caroti L, Rosso G, Bertoni E, Salvadori M.

- Keratinocyte cancer prevention with ACE inhibitors, angiotensin receptor blockers or their combination in renal transplant recipients. *Clin Nephrol.* 2010;73(6):439–45.
112. Maluccio M, Sharma V, Lagman M, Konijn G, Suthanthiran M. Angiotensin II receptor blockade: a novel strategy to prevent immunosuppressant-associated cancer progression. *Transplant Proc.* 2001;33(1):1820–1.
113. Song C, Piva M, Sun L, Hong A, Moriceau G, Kong X, Zhang H, Lomeli S, Qian J, Yu CC, Damoiseaux R, Kelley MC, Dahlman KB, Scumpia PO, Sosman JA, Johnson DB, Ribas A, Hugo W, Lo RS. Recurrent tumor cell–intrinsic and–extrinsic alterations during MAPKi-induced melanoma regression and early adaptation. *Cancer Discov.* 2017;7(11):1248–65.
114. Doherty MR, Smigiel JM, Junk DJ, Jackson MW. Cancer stem cell plasticity drives therapeutic resistance. *Cancers.* 2016;8(1):8.
115. Shi H, Hugo W, Kong X, Hong A, Koya RC, Moriceau G, Chodon T, Guo R, Johnson DB, Dahlman KB, Kelly MC, Kefford RF, Chmielowski B, Glapsy JA, Sosman JA, van Baren N, Long GV, Ribas A, Lo RS. Acquired resistance and clonal evolution in melanoma during BRAF inhibitor therapy. *Cancer Discov.* 2014;4(1):80–93.
116. Fattore L, Ruggiero CF, Pisanu ME, Liguoro D, Cerri A, Costantini S, Capone F, Acunzo M, Romano G, Nigita G, Mallardo D, Ragone C, Carriero MV, Budillon A, Botti G, Ascierto PA, Mancini R, Ciliberto G. Reprogramming miRNAs global expression orchestrates development of drug resistance in BRAF mutated melanoma. *Cell Death Differ.* 2019;26(7):1267–82.
117. Bühler FR, Laragh JH, Baer L, Vaughan ED, Brunner HR. Propranolol inhibition of renin secretion. A specific approach to diagnosis and treatment of renin-dependent hypertensive diseases. *N Engl J Med.* 1972;287(24):1209–14.