

Genetic Variability in Patients with HCV-Related Hepatocellular Carcinoma

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Background: The present paper evaluates the genetic variability of HCV in patients with hepatocellular carcinoma (HCC).

Methods: Amino acid substitutions (aas) in NS3, NS5A and core regions were analyzed in 17 patients with HCC (Cases) and 13 without HCC (Controls), all naïve to DAAs. For the Cases, a sample of neoplastic liver tissue, non-neoplastic liver tissue and a serum sample were collected; for the Controls, a sample of liver tissue was collected. Sanger sequencing of three regions was performed using homemade protocols.

Results: Phylogenetic trees showed that there was no difference in the virus populations in the three compartments analyzed for the three HCV regions in patients with HCC. Low variability and no difference between the Cases and Controls were observed in the core and NS5A regions; however, in the NS3 region, a higher variability was observed in the Cases. No difference was observed in the core region between Cases and Controls. In NS3, aa substitutions at positions 103 and 122 were more frequently found in Cases than Controls (in both cases 50% vs 9.1%, $p < 0.05$); moreover, aas in positions 32, 44 ($p = 0.035$ for both), 79 ($p = 0.008$) and 121 ($p = 0.018$) were observed in the Cases and absent in the Controls. Finally, considering the NS5A region, aa substitutions at positions 37 and 54 were more frequently identified in the Cases than the Controls, but without statistical significance.

Conclusion: These data may suggest a higher aa variability in patients with HCC than in those without, especially in the NS3 region.

Keywords: hepatocellular carcinoma, HCV, viral variability, chronic hepatitis C, chronic infection C, core region, NS5A region, NS3 region, aa substitutions, phylogenesis

Introduction

Hepatocellular carcinoma (HCC) is the sixth most frequent cancer worldwide and the fourth leading cause of cancer-related deaths and arises especially in cirrhotic patients.¹

The prevalence of HCC reflects the epidemiology of the etiological agents responsible for chronic liver diseases. In developing countries, liver cancer is associated with hepatitis B virus (HBV) and hepatitis C virus (HCV) chronic infection. In developed countries, the impact of HBV and HCV on the epidemiology of HCC shows a reduction with an increasing role of non-alcoholic fatty liver diseases (NAFLD).² In fact, the universal HBV vaccination has significantly reduced the incidence of HBV-related HCC, and the increasing use in recent years of directly acting antiviral agents significantly reduced, although perhaps not fully eliminate, the risk of developing HCC.^{3,4}

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Hepatocarcinogenesis is a complex multistep process and is influenced by many risk factors, including the oncogenic effect of HBV proteins, steatosis and insulin resistance, chronic inflammation and fibrosis, oxidative stress and chromosomal instability.^{5,6}

HCV is indirectly involved in hepatocarcinogenesis through the development of cirrhosis: HCV-related necrosis may promote cell proliferation and long-term hepatic inflammation that may contribute to the development of HCC with oxidative stress and DNA damage.^{7–9} However, recent reports hypothesized that HCV directly favored the development of HCC by different molecular mechanisms, such as the alteration of normal cellular signaling pathways to stimulate host cell growth and cell transformation. Moreover, some studies have recently shown that some amino acid substitutions (aas) in the core region of genotype 1b were associated with HCC; specifically, aas 70 and/or 91 seem to be predictors of hepatocarcinogenesis.^{10,11}

The present study evaluated the genetic variability of HCV in patients with HCC compared to those without by analyzing NS3, NS5A and core HCV regions in different specimens, serum, cancer liver tissue and non-cancer liver tissue.

Patients and Methods

Patients

Of the 68 consecutive patients who underwent a diagnostic liver biopsy for HCC at the liver unit of University of Campania, Naples, Southern Italy, from June 2013 to December 2014,^{12,13} 17 HCV-RNA positive were retrospectively enrolled in the present study according to the availability of liver tissue collected (Case group). HCC was diagnosed according to the EASL/EORTC criteria.¹⁴ To reach the aim of the study, the first 13 patients without HCC who underwent liver biopsy to stage their liver disease in June–July 2013 and for whom the liver specimens collected at -80°C were available were enrolled (Control group).

A physical examination, full liver function tests and serology for HBsAg, anti-HCV and anti-HIV were performed in each patient enrolled. The stage of HCC was assessed according to the BCLC (Barcelona Clinic Liver Cancer) criteria.¹⁵

For the 17 Cases, a sample of cancer tissue (CT), non-neoplastic liver tissue (non-CT) and a serum sample (S) were collected for the aims of the study. For the 13 Controls, a sample of liver tissue was collected. Both for

the Cases and Controls, fragments of nearly 3 mg were cut away from the two extremities of the liver biopsies and stored at -80°C in RNAlater solution (Qiagen GmbH, Hilden, Germany) for molecular analyses.

The study was approved by the Ethics Committee of the Azienda Ospedaliera Universitaria of the Second University of Naples (n°349/2013). All procedures performed in this study were in accordance with the ethics standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethics standards. Informed consent was obtained from all participants included in the study.

Routine Methods

Anti-HCV was performed using a third-generation commercial immunoenzymatic assay (Ortho Diagnostic Systems, Neckargemund, Germany). Liver biochemistry and routine analyses were performed by routine methods in a Cobas Modular 6000 automated analyzer using c501 biochemistry modules (Roche Diagnostics Ltd, Rotkreuz, Switzerland).

HCV Molecular Methods

Viral RNA was extracted from 200 μL of serum samples and from HCC (CT) and non-HCC (non-CT) tissues using a microspin column (QIAamp RNA viral kit and QIAamp DNA/RNA mini-kit, respectively, Qiagen GmbH, Hilden, Germany). Serum HCV-RNA was quantified by a real-time PCR method in a Light cycler 1.5 (Roche Diagnostics, Branchburg, NJ, USA) with a sensibility of less than 40 IU/mL, as previously reported.¹⁶

HCV genotypes were identified by HCV genotype Lipa assay (Bayer, France).

Sanger sequencing of NS3, NS5A and core regions was performed on samples applying home-made protocols.¹⁷ Briefly, the NS3 (181aa), NS5A (140aa) and Core (100 aa) regions were amplified by primers as shown in [Supplementary Table 1](#) and sequenced using ABI 3500 Genetic Analyzer (Applied Biosystems).

The sequences were compared with reference sequences recognized in the literature and taken from Los Alamos Database and other databases (GenBank-accession-numbers: HCV-1a: H77 NC_004102, HCV-1b: D90208, HCV-2a/2c: D50409, HCV-3a: D28917, HCV-3h: JF735126, HCV-4: DQ418786). The phylogenetic trees were made using the Mega 10 program. By the comparison with the reference sequence, it was possible to establish the

genotypes and possible contaminations of the samples. The mutations and quasi-species were identified by seqscape program software (Applied Biosystems) with a >20% tolerance for improper sequencing. Finally, we interpreted the amino acid (aa) substitutions based on the reference sources.

Statistical Analysis

Continuous variables were summarized as mean and standard deviation, and categorical variables as absolute and relative frequencies. For continuous variables, the differences were evaluated by the Student's *t*-test; categorical variables were compared by the chi-square test, using exact procedures if needed. Differences were considered statistically significant at $p < 0.05$.

Results

The demographic and biochemical characteristics of Cases and Controls are shown in Table 1. The Cases were older (median age 71 years, range 66–79) than the Controls (median age 60, range 42–71), (<0.00001) and mostly males (58.8% vs 46.2, a difference not significant to the statistical analysis.) (Table 1). Compared with the Controls, the Cases showed lower transaminase serum values [ALT, mean \times normal value \pm standard deviation (SD): 1.51 ± 0.99 vs 3.052 ± 1.58 , $p = 0.16$; AST: 1.59 ± 0.97

vs 1.87 ± 1.07 , $p = 0.71$] (Table 1). On the contrary, total bilirubin value was higher in the Cases than Controls (1.64 ± 1.17 vs 0.99 ± 0.50 mg/dL, $p = 0.10$.) (Table 1). The serum HCV-RNA viral load was higher in the Controls (Table 1). The most representative HCV genotype was 1b in both groups: 14 (82%) patients in the Cases and 11 (85%) in the Controls; 3 Cases harbored HCV genotype 2c and 2 Controls genotype 2a (Table 1). All patients were naïve to DAAs.

The sequence analyzed in the different HCV regions, core, NS3 and NS5A, for the Cases and Controls are shown in Supplementary Tables 2 and 3, respectively. The sequences of patients with HCC (Case group) obtained in the three compartments (cancer tissue, non-cancer tissue and serum) were used to create phylogenetic trees (Figure 1A–C). Phylogenetic trees showed that there was no difference between the virus populations in the three compartments analyzed in the patients with HCC (Figure 1).

Table 2 shows the HCV variability in the three viral regions analyzed in the Cases and Controls. Low variability and no difference between the Cases and the Controls were observed in the core and NS5A regions; however, in the NS3 region a higher variability was observed in the Cases. In fact, compared with the patients without, those with HCC showed in all three compartments a higher number of aa substitutions in the NS3 region: median (IQR) aa substitutions, 9 (9.5) in cancer tissue, 7.5 (12.75) in non-cancer tissue and 8 (24) in serum in the Cases vs 5 (4) in liver tissue in the Controls.

Tables 3–5 show, for each HCV region analyzed, the positions in which there was more frequent amino acid substitutions (in at least 25% of the subjects).

Table 6 shows the positions of amino acid (aa) substitutions in NS3, NS5A and core region, amino acid substitutions identified and the compartments in which these were found.

Considering the HCV core region, no differences in the aa substitutions were found in the Cases and Controls (Table 3).

Considering the NS3 region, aa substitutions at positions 103 and 122 were more frequently found in the Cases than the Controls (in both cases 50% vs 9.1%, $p < 0.05$; Table 4). Amino acid substitutions in positions 30, 32, 41, 44, 52, 54, 57, 79, 80, 107, 112, 117, 121 and 130 were also observed in the Cases, but were absent in the Controls, with a difference significant to the statistical analysis for position 32 and 44 ($p = 0.035$ for both), for

Table 1 Demographic and Clinical Characteristics in the Cases and Controls

	Cases	Controls	P
Patients	17	13	
Median age (range)	71 (66–79)	60 (42–71)	<0.00001
Males (%)	10 (58.8)	6 (46.2)	0.490644
ALT/mL \times normal value (media \pm SD)	1.51 ± 0.99	3.02 ± 1.58	0.166052
AST/mL \times normal value (media \pm SD)	1.59 ± 0.97	1.87 ± 1.07	0.712249
Total bilirubin mg/dL (media \pm SD)	1.64 ± 1.17	0.99 ± 0.50	0.108374
Median HCV-RNA UI/ML (range)	3.4×10^5 (1.3×10^3 – 6.9×10^6)	2.5×10^5 (4.5×10^5 – 2.8×10^7)	0.032062
N° (%) of patients with			
HCV genotype 1b	14 (82.4)	11 (84.6)	0.869125
HCV genotype 2a	0	2 (15.4)	0.0941
HCV genotype 2c	3 (17.6)	0	0.1104

position 79 ($p=0.008$) and 121 ($p=0.018$) (Table 4). Interestingly, 2 aa substitutions (S122G and D168E) conferring resistance to anti-NS3 inhibitors were identified: 4 Cases and 1 Control harbored S122G and 1 Case D168E (Table 4).

Finally, considering the NS5A region, aa substitutions at position 37 and 54 were more frequently identified in the Cases than in the Controls, but due to the small number of patients, without statistical significance (90% vs 55.6% and 80% vs 33%, respectively, Table 5). Moreover, substitutions in positions 101, 120 and 126 were present in the Cases but absent in the Controls

(Table 5). Interestingly, in both groups of patients aa substitutions conferring resistance to DAAs were identified, Y93H in two Cases and in 1 Control, L31M in 1 Control (Table 5).

Discussion

In the present study, analyzing the HCV sequences in core, NS3 and NS5A regions in serum, non-cancer liver tissue and in cancer liver tissue of 17 patients with HCC and in the liver tissue of 13 patients without, we tested the variability of HCV genome as the cause of hepatocarcinogenesis.

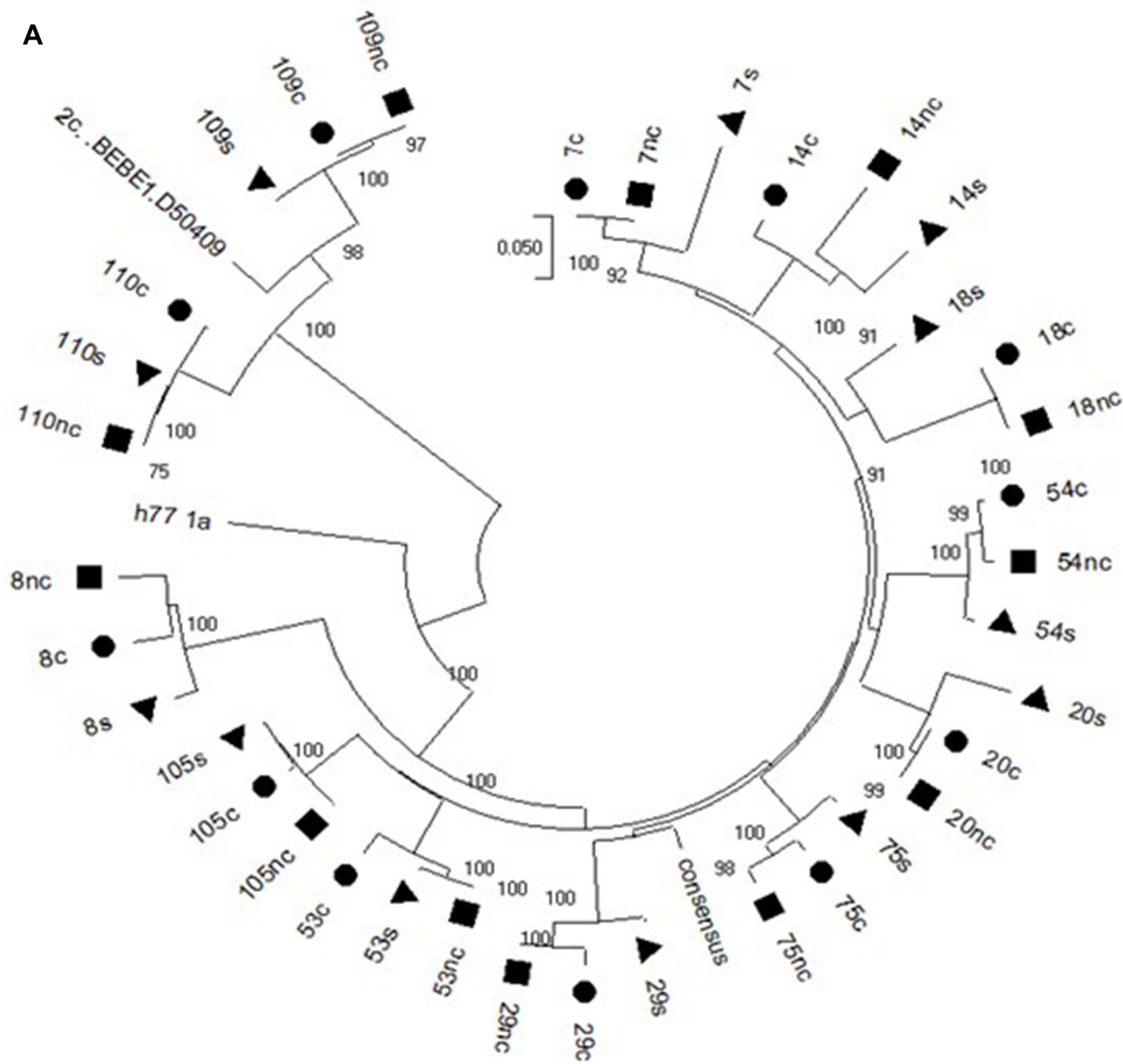


Figure 1 Continued.

Table 2 Number of aa Substitutions in Core, NS3 and NS5A Regions for the Cases and Controls

	Compartments	NS3		NS5A		Core	
		Samples	Median (IQR)	Samples	Median (IQR)	Samples	Median (IQR)
Cases	Cancer tissue	16	9 (9.5)	10	6 (3.5)	15	3 (3)
	Non-cancer tissue	16	7.5 (12.75)	10	5.5 (4.5)	13	3 (4)
	Serum	13	8 (24)	10	6 (4)	11	3 (3)
Controls	Liver tissue	11	5 (4)	9	5 (2.5)	11	3 (3)

Table 3 Prevalence of Patients with More Frequent* Amino Acid Substitutions at Different Positions of the Core Region

Core Region			ρ
Position	Cases (16)	Controls (11)	
70	4 (25%)	4 (36.4%)	0.5252
75	7 (43.8%)	5 (45.5%)	0.9302
91	4 (25%)	1 (9.1%)	0.2957
110	4 (25%)	3 (27.3%)	0.8947

Note: *Amino acid substitutions present in at least 25% of the subjects.

cancer tissue and that from non-cancer liver tissue and serum.

Moreover, by the analysis of the mutation patterns of the NS3, NS5A and core regions between patients with or without HCC, a higher aa variability in the NS3 region was observed in the Cases. In addition, we observed that in NS3 and NS5A regions there were some mutations that were repeated more frequently in the patients with HCC than in the Controls, such as in positions 32, 44, 79, 103, 121 and 122 in NS3 and in positions 37 and 54 in the NS5A region. These data may suggest a higher aa variability in patients with HCC than in those without, especially in NS3.

This greater genetic variability found in the NS3 and NS5A regions of the HCV genome of subjects with HCC in our opinion could have a link with the development of liver cancer, since literature studies have shown that the NS3 and NS5A regions are involved in hepatocarcinogenesis. In fact, the NS3 and NS5A proteins may promote hepatocarcinogenesis by its binding to some cellular proteins, such as p21 and p53^{24,25} and the NS5A protein modulates genes that regulate the cell cycle, promote cell growth and induce aberrant mitosis and chromosomal instability leading to HCC.^{26–29}

However, the patients with HCC were older than those without; thus, these data may be due to a longer infection time that caused a higher aa variability.

Table 4 Prevalence of Patients with More Frequent* Amino Acid Substitutions at Different Positions of the NS3 Region

NS3 Region			ρ
Position	Cases (16 Patients)	Controls (11 Patients)	
7	4 (25%)	5 (45.5%)	0.2679
48	6 (37.5%)	7 (63.6%)	0.1817
61	7 (43.8%)	4 (36.4%)	0.7011
65	4 (25%)	1 (9.1%)	0.2957
71	6 (37.5%)	2 (18.2%)	0.2801
72	4 (25%)	4 (36.4%)	0.5252
86	4 (25%)	3 (27.3%)	0.8947
87	5 (31.3%)	3 (27.3%)	0.8240
103	8 (50%)	1 (9.1%)	0.0267
122	8 (50%)	1 (9.1%)	0.0267
132	4 (25%)	5 (45.5%)	0.2679
170	5 (31.3%)	4 (36.4%)	0.7818
30	6 (37.5%)	0	0.06
32	7 (43.8%)	0	0.035
41	4 (25%)	0	0.21
44	7 (43.8%)	0	0.035
52	5 (31.3%)	0	0.12
54	4 (25%)	0	0.21
57	6 (37.5%)	0	0.06
79	9 (56.3%)	0	0.008
80	4 (25%)	0	0.21
107	4 (25%)	0	0.21
112	6 (37.5%)	0	0.06
117	6 (37.5%)	0	0.06
121	8 (50%)	0	0.018
130	5 (31.3%)	0	0.12

Note: *Amino acid substitutions present in at least 25% of the subjects.

There are few studies that analyze the genetic and aa variability of HCV in the cancerous and non-cancerous tissue of HCC patients, with equivocal results.^{30–34} In the study by Perez et al,³¹ core and E2 regions were amplified, cloned and sequenced in four patients with HCC who underwent liver transplantation, showing in all the patients a compartmentalization in the three compartments

Table 5 Prevalence of Patients with More Frequent* Amino Acid Substitutions at Different Positions of the NS5A Region

NS5A Region			ρ
Position	Cases (10)	Controls (9)	
34	3(30%)	1(11.1%)	0.3133
37	9(90%)	5(55.6%)	0.0887
54	8(80%)	3(33.3%)	0.0397
75	5(50%)	2(22.2%)	0.2101
78	4(40%)	2(22.2%)	0.4052
79	3(30%)	1(11.1%)	0.3133
101	3(30%)	0	0.25
120	4(40%)	0	0.11
126	3(30%)	0	0.25

Note: *Amino acid substitutions present in at least 25% of the subjects.

analyzed (tumoral, non-tumoral liver tissue and serum) for both viral regions tested. Harouaka et al³² evaluating 12 patients with HCV-associated HCC ($n^{\circ} = 8$) or non-HCC cirrhosis ($n^{\circ} = 4$) showed that HCV genetic diversity was significantly higher in livers containing HCC compared with the control non-HCC cirrhotic livers.

Interesting are the results of Sorbo et al³⁵ who investigated the prevalence of resistance-associated aa substitutions (RASs) in plasma, liver tumoral and non-tumoral tissue samples in 18 patients undergoing liver transplant or hepatic resection due to HCC and/or cirrhosis: 6 of 18 patients had at least one RAS in at least one of the compartments analyzed, and nucleotide variability assessed by genetic distance analysis showed higher median values for NS3 and NS5A sequences compared to NS5B in the tissue compartments.

The limit of the present study is the small sample size of the groups. However, we underline that the analysis done, sequencing of NS3, NS5A and core regions both in liver cancer and in non-cancer tissue and in serum of 17 patients with HCC, such as in liver tissue in 13 patients without HCC, although they did not allow to draw unequivocal conclusions, seem to be adequate to a preliminary evaluation of this topic.

In conclusion, from what has been said so far it would seem that what the previous studies have in common with ours is the identification of a greater genetic variability of HCV in tumor tissue compared to non-tumor tissue. Unfortunately, what unites them is also the small size of the sample, so further studies on a wide cohort of patients may better clarify this topic and identify other amino acid substitutions possibly associated with the development of HCC.

Table 6 Positions of Amino Acid (aa) Substitutions in NS3, NS5A and Core Region, Amino Acid Substitutions Identified and the Compartments in Which These Were Found

	Positions	Amino Acid Substitutions	Compartments
NS3	7	S7A	C, NC, S, T
	30	D30E	C, NC, S,
	32	E32K	C, NC, S
	41	Q41H	C, NC, S
	44	L44R	C, NC, S
	48	V48I	C, NC, S, T
	52	C52R	C, NC, S
	54	T54I	C, S
	57	H57Q	C, NC, S
	61	S61T	C, NC, S, T
	65	A65P	C, NC, S, T
	71	I71V	C, NC, S, T
	72	T72I	C, NC, S, T
	79	D79N	C, NC, S
	80	Q80E	C, NC, S
	86	Q86P	C, NC, S, T
	87	A87G	C, NC, S, T
	103	D103N	C, NC, S, T
	107	V107A	C, NC, S
112	D112N	C, NC, S	
117	R117P	C, NC, S	
121	D121N	C, NC, S	
122	S122G	C, NC, S, T	
130	R130K	C, NC, S	
132	V132I	C, NC, S, T	
168	D168E	NC, S	
170	V170I	C, NC, S, T	
NS5A	31	L31M	T
	34	V34I	C, NC, S, T
	37	F37L	C, NC, S, T
	54	Q54H	C, NC, S, T
	75	V75A	C, NC, S, T
	78	K78R	C, NC, S, T
	79	T79A	C, NC, S
	93	Y93H	C, NC, S, T
	101	S101C	NC, S
	120	E120K	C, NC, S,
126	D126N	C, S	
CORE	70	Q70R	C, NC, S, T
	75	A75T	C, NC, S, T
	91	M91L	C, NC, S, T
	110	T110N	C, NC, S, T

Notes: C, cancer; NC, non-cancer; S, serum; T, hepatic tissue.

Data Sharing Statement

All the data on the study may be requested to the authors, Mariantonietta Pisaturo and Nicola Coppola.

Consent to Participate

All patients signed their informed consent for liver biopsy, the collection and storage of biological samples and for the anonymous use of their data for research purposes.

Consent to Publish

The manuscript has been read and approved by all authors.

Author Contributions

All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis and interpretation, or in all these areas; took part in drafting, revising or critically reviewing the article; gave final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

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Disclosure

The authors declare that we have no conflicts of interest related to this work.

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