

Landscape and Clonal Dominance of Co-occurring Genomic Alterations in Non–Small-Cell Lung Cancer Harboring *MET* Exon 14 Skipping

Xiuning Le, MD, PhD¹; Lingzhi Hong, MD, PhD^{1,2}; Chuck Hensel, PhD³; Rongrong Chen, PhD⁴; Haley Kemp, MPAS¹; Niamh Coleman, MBBCh, PhD⁵; Christine A. Ciunci, MD⁶; Stephen V. Liu, MD⁷; Marcelo V. Negrao, MD¹; Jennifer Yen, PhD³; Xuefeng Xia, MD, PhD⁴; Juergen Scheuenpflug, PhD⁸; Christopher Stroh, PhD⁸; Dilafruz Juraeva, PhD⁸; Anne Tsao, MD¹; David Hong, MD, PhD⁵; Victoria Raymond, MS³; Paul Paik, MD⁹; Jianjun Zhang, MD, PhD¹; and John V. Heymach, MD, PhD¹

PURPOSE *MET* exon 14 skipping alterations (*MET*Ex14) comprise a diverse set of actionable oncogene drivers in non–small-cell lung cancer (NSCLC). Recent studies have established the efficacy of tyrosine kinase inhibitors for this patient population. The landscape of co-occurring genetic alterations in *MET*Ex14 NSCLC and their potential impact to therapeutic sensitivities has not yet been fully described.

MATERIALS AND METHODS *MET*Ex14 NSCLC cases were collected from three cohorts: the VISION trial, and data sets from Guardant360 and GenePlus. Clinicopathologic characteristics and *MET*Ex14 mutation sites were analyzed and compared across data sets. Co-occurring genetic alterations and the clonality relationships to *MET*Ex14 were evaluated.

RESULTS Of 40,824 NSCLCs, 692 *MET*Ex14 cases (1.7%) were identified, including 332 in Guardant360, 188 in VISION, and 172 in GenePlus. The demographics and mutation type and/or sites were similar in the Asian versus Western cohorts. *MET* amplification, which were found to be associated with sensitivity to *MET* kinase inhibitors, co-occurs in 7.6%-13.8% of cases, whereas kinase domain secondary mutation of *MET* co-occurs in 5%-6%. When co-occurring with *MET*Ex14, *EGFR* mutations were often identified as the dominant clone (78%, 7 of 9), whereas when co-occurring, *MET*Ex14 (39%, 7 of 18) and *KRAS* (44%, 8 of 18) had similar rates of clonal dominance. *PIK3CA* and *PTEN* mutations were almost always subclones (89%, 16 of 18) to *MET*Ex14. Moreover, *RET-CCDC6* fusion and *EGFR* mutation were detected following crizotinib treatment in two patients, suggesting novel mechanisms of resistance.

CONCLUSION *MET*Ex14 mutations frequently co-occur with other potential driver oncogenes with differing patterns of clonal dominance observed among the drivers. This cellular context can provide insights into whether *MET*Ex14 is acting as a primary oncogenic driver or resistance mechanism and help guide treatment choices.

JCO Precis Oncol 5:1802-1812. © 2021 by American Society of Clinical Oncology

Creative Commons Attribution Non-Commercial No Derivatives 4.0 License 

BACKGROUND

MET exon 14 skipping alterations (*MET*Ex14) have recently been established as an actionable oncogene driver in non–small cell lung cancer (NSCLC).¹ Small molecule *MET* tyrosine kinase inhibitors (TKIs) have shown efficacy in patients with *MET*Ex14 NSCLC, with objective response rate ranging from 25% to 68% and median progression-free survival at 7.6-13.8 months.²⁻⁷ From 2020 to 2021, capmatinib and tepotinib received US Food and Drug Administration approval for *MET*Ex14 NSCLC, representing a significant milestone in *MET* TKI development.

Obtaining a full understanding of co-occurring alterations with *MET*Ex14 could be crucial in providing novel insights

to increase our understanding of treatment sensitivity and resistance in *MET*Ex14 NSCLC, and thus, guide future therapeutic strategy development. Acquired *MET* kinase domain (KD) mutations in residues D1228 and Y1230 have been shown to cause *MET* TKI resistance.⁸ Recent studies also indicate that some co-occurring alterations are detected in TP53, RAS-MAPK, and PI3K pathways.⁹ Gene amplifications (eg, *EGFR*, *MDM2*, and *CDK4*) were also observed in 6%-35% of *MET*Ex14 NSCLC.¹⁰ Some of these genomic alterations have been confirmed as mechanisms of *MET* TKI resistance, especially *RAS-MAPK* and *PI3K* pathways.^{9,11-14} Furthermore, the efficacy of immunotherapy for patients with *MET*Ex14 NSCLC was low despite high programmed death-ligand 1 expression.¹⁵ However, comprehensive landscape

ASSOCIATED CONTENT

Data Supplement

Author affiliations and support information (if applicable) appear at the end of this article.

Accepted on October 15, 2021 and published at ascopubs.org/journal/po on December 13, 2021; DOI <https://doi.org/10.1200/P0.21.00135>

CONTEXT

Key Objective

To evaluate the mutational profile and co-occurring genetic alteration landscape of non–small-cell lung cancer harboring *MET* exon 14 skipping (*METex14*) and the clonality relationship between the *METex14* and co-occurring mutations inferred from the variant allele frequency and dissect the potential resistance mechanisms in cases with longitudinal biopsies.

Knowledge Generated

The demographics and mutation type and/or sites of *METex14* were similar in the Asian versus Western cohorts. *METex14* mutations frequently co-occur with other potential driver oncogenes with differing patterns of clonal dominance observed among the drivers, and usually nondominant subcloned when co-occurred with *EGFR* and *ERBB2*.

Relevance

METex14 can act as a primary oncogenic driver or resistance mechanism, suggesting that appropriate treatment choices can be potentially guided by co-occurring alterations.

description of co-occurring mutations with *METex14* in NSCLC is still missing.

Here, we leveraged three cohorts of *METex14* NSCLCs and aimed to evaluate the mutational profile and co-occurring genetic alteration landscape of *METex14* NSCLC across countries. We evaluated clonality relationship between the *METex14* and co-occurring mutations inferred from the variant allele frequency (VAF) and dissected the potential resistance mechanisms in cases with longitudinal biopsies.

MATERIALS AND METHODS

Study Population and Platform

Three data sets were queried for *METex14* NSCLC: Guardant360 (July 2019 to July 2020), GenePlus (both circulating tumor DNA [ctDNA] and tissue, February 2017 to April 2020), and VISION trial ctDNA cohort (NCT02864992) detected by Guardant360. The Data Supplement shows Guardant360 ctDNA 74 gene and VISION ctDNA 73 gene (without *CDK12*) panels. The Data Supplement also shows GenePlus 1,021 and 59 gene panels for ctDNA or tissue. The panels used in tissue or blood samples from GenePlus were summarized in the Data Supplement. There were 48 genes covered in all patients across the three data sets (Data Supplement).

METex14 Detection

For Guardant360 (also VISION), single-nucleotide variant or indel variant that overlaps any of the two splice regions of *MET* exon 14 (chromosome 7:116411902-116412043; human genome [hg19]) defined as eight bp into the intron or three bp into the exon was identified with the Guardant360 assay. Detection of indels larger than 50 bp is described in previous publication.¹⁶ For GenePlus, the regions defined as *METex14* were the same to Guardant360. Additionally, variants that affect bases as far as 26 bp into the intron were also identified as *METex14*.

Actionable Mutation Determination

The actionability of each mutation was determined when it was considered as pathogenic by Catalogue Of Somatic Mutations In Cancer (COSMIC) Score.¹⁷

Estimation of Mutation Clonality

Variant clonality was determined by normalizing VAF to the maximum somatic VAF in a sample. Variants were classified as clonal if the normalized value was ≥ 0.5 , subclonal for values < 0.5 but ≥ 0.05 , and subclonal minor if < 0.05 .

Statistical Analysis

Group comparisons were performed using a 2-tailed chi-square test, with significance threshold of *P* value $< .05$. Analyses were performed using GraphPad Prism 8.0.

RESULTS

Clinicopathologic Characteristics of ctDNA Detected *METex14* NSCLC

A total of 692 patients with NSCLC with *METex14* were identified from three independent data sets of a combined total of 40,824 patients with NSCLC with an overall incidence of 1.7%, including Guardant360 (332 of 20,987, 1.6%), GenePlus (172 of 14,657, 1.2%), and VISION trial (188 of 5,180, 3.6%). Patient demographics and tumor characteristics were summarized in the Data Supplement. In all three data sets, *METex14* occurred with higher frequency in adenocarcinoma, with increasing age and equal sex distribution.

When the GenePlus ctDNA cohort ($n = 37$) was compared with the Western data sets (Guardant360 plus VISION, $n = 520$), there were no differences in age (median 70.5 v 73 years) and sex distribution (female 54% v 57%), and similar patient demographic characteristics were noted in both Asian and Western data sets. Prevalence of *METex14* was higher in the VISION trial (3.6%) than the other two real-world cohorts (Guardant360 [1.6%] and GenePlus [1.2%]), likely because VISION trial excluded *EGFR* and *ALK*-positive patients at initial screening.

Mutation Characteristics of *METex14* in NSCLC

Next, we characterized the mutational landscape of *METex14* NSCLC from the three data sets. The positions of *MET* mutations and the prevalence by functional alterations

are shown in [Figure 1](#) and [Table 1](#). The functional sites of *MET* mutations were similar across the three data sets, allowing the differences among platforms and *MET*ex14 detection methods. In Guardant360, the prevalence by functional alteration sites were as follows: donor (44.3%), acceptor (30.4%), D1010 (20.5%), and Y1003 (3.9%). The most frequent mutation type was base substitution (55.7%), followed by indel (43.4%). Both the VISION and the GenePlus data sets revealed a remarkably similar pattern in terms of prevalence by functional alteration sites and most frequent mutation type ([Fig 1](#), [Table 1](#)). Regarding acceptor versus donor sites and SNVs versus indels, there was no significant difference between the Asian and Western data sets.

Co-occurring *MET* Amplification (*MET*amp) With *MET*ex14 in NSCLC

The frequency of *MET*amp co-occurred with *MET*ex14 was 8.4% (Guardant360), 13.8% (VISION), and 7.6% (GenePlus), respectively (Data Supplement). The mean VAF of *MET*ex14 in cases concomitant with *MET*amp was significantly higher than those without *MET*amp across three data sets ($P < .001$, Data Supplement). The distribution of gene copy number for Guardant360 and GenePlus is displayed in the Data Supplement. Most of the patients had an *MET* gene copy number between 2 and 4 (24 of 28 in Guardant360; 9 of 13 in GenePlus).

In VISION trial, four in five co-occurring *MET*ex14 and *MET*amp cases (80%) had a partial response and one had target lesion tumor reduction, but progressive disease because of a new lesion. The response rate was numerically higher than in patients whose tumor did not have co-occurring *MET*amp (4 of 5, 80% v 28 of 61, 46%), suggesting co-occurring *MET*amp might be associated with responsiveness to targeted therapy,⁴ although the number in the *MET*amp group was too small for a statistical comparison.

Secondary Mutations Within *MET* in *MET*ex14 NSCLC

Secondary mutations located in *MET* KDs, such as D1228 and Y1230, have been reported to be associated with resistance to MET TKI.^{8,11} In Guardant360, 46 (13.9%) had *MET* secondary mutations including 17 (5%) having at least one secondary mutation in the KDs, including H1094C/Y, D1228H/N, and Y1230C/H ([Fig 2B](#)). In GenePlus, secondary mutations were detected in 29 (16.9%) patients, including 11 (6%) in the KDs ([Fig 2B](#)). Most of the KD mutations were deemed pathogenic on the basis of COSMIC prediction score > 0.95 . The VISION data set only included TKI-naïve (part of trial eligibility criteria) patients and had no secondary mutations.

Co-occurring Genetic Alterations in Bypass Pathways in *MET*ex14 NSCLC

Next, we evaluated other co-occurring genetic alterations with *MET*ex14 in NSCLC. For a fair comparison, we focused

only on the 48 cancer genes tested in all patients across the three data sets. First, we compared whether any comutations were enriched in *MET*ex14 NSCLC compared with NSCLC without *MET*ex14. In the Guardant360 and GenePlus cohorts, no gene alterations were enriched for co-occurring with *MET*ex14. *EGFR*, *KRAS*, and *TP53* were significantly enriched in the non-*MET*ex14 tumors in both cohorts ([Figs 3A](#) and [3B](#)), consistent with the notion that *MET*ex14 is a de novo oncogenic driver.

We then focused on comutations in key bypass pathways, including RAS-MAPK, EGFR and ERBB2, PI3K and AKT, and cell cycle (*CDK4/6*) pathways (Data Supplement). In Guardant360, 136 (41%) had at least one co-occurring alteration in those pathways, including *EGFR* (24 of 332), *ERBB2* (13 of 332); *KRAS* (20 of 332), *BRAF* (12 of 332), *NRAS* (7 of 332), *NF1* (3 of 332), and *PIK3CA* (33 of 332), *PTEN* (7 of 332), and *AKT1* (1 of 332) (Data Supplement). In the VISION trial, *EGFR* driver mutations were specifically excluded. Fifty-one (27%) patients had at least one co-occurring alteration; *ALK* and *RET* fusions, each occurred once (Data Supplement). In GenePlus data set, 49 of 172 (28%) *MET*ex14 cases had at least one co-occurring alteration (Data Supplement), with similar distribution to Guardant360 data set.

Clonality Relationship Between *MET*ex14 and Other Driver Oncogenes in NSCLC

Oncogene variant clonality can be deduced from VAF to infer dominant versus nondominant clonal relationship. We annotated comutations using COSMIC score to identify the potential activating mutations and found that 76 unique cases (23%) had at least one activating comutation in Guardant360 data set ([Fig 3C](#)). We then evaluated the relative clonality inferred by VAF for *MET*ex14 and the second activating mutations to dissect potential resistance mechanisms. In the *EGFR* cases, most cases (7 of 9, 78%) had *EGFR* as the dominant clone, as was the one case with *ERBB2* mutation (A775_G776insYVMA, VAF 31.4%) being dominant compared with 0.56% in *MET*ex14 (GH#128, [Fig 3C](#) and Data Supplement). *ALK* fusions also had high VAF in both of two co-occurring cases (GH#201 and GH#213, [Fig 3C](#)). For *PIK3CA* and *PTEN* mutations, the clonality of *MET*ex14 was higher in 16 of 18 (89%) cases. Interestingly, for *KRAS* and *MET*ex14 co-occurring cases, 44% (8 of 18) had *KRAS* as the dominant clone, and 39% of cases were *MET*ex14-dominant. *KRAS* G12C and *BRAF* V600E were both detected in one single patient (GH#89, [Fig 3C](#)), and *MET*ex14 was the dominant mutation.

Eighteen out of 188 (9.6%) patients in the VISION cohort had co-occurring alterations and the dominant clonality of *MET*ex14 was observed in 2 of 5 (40%) patients with co-occurring in *EGFR* and *ERBB2*, 0 of 1 in *ALK* (*EML4-ALK* fusion), 3 of 4 (75%) in *K/NRAS*, and 7 of 8 (87.5%) in *PIK3CA* and *PTEN* (Data Supplement). In GenePlus, similar pattern was seen (Data Supplement). In summary, when co-

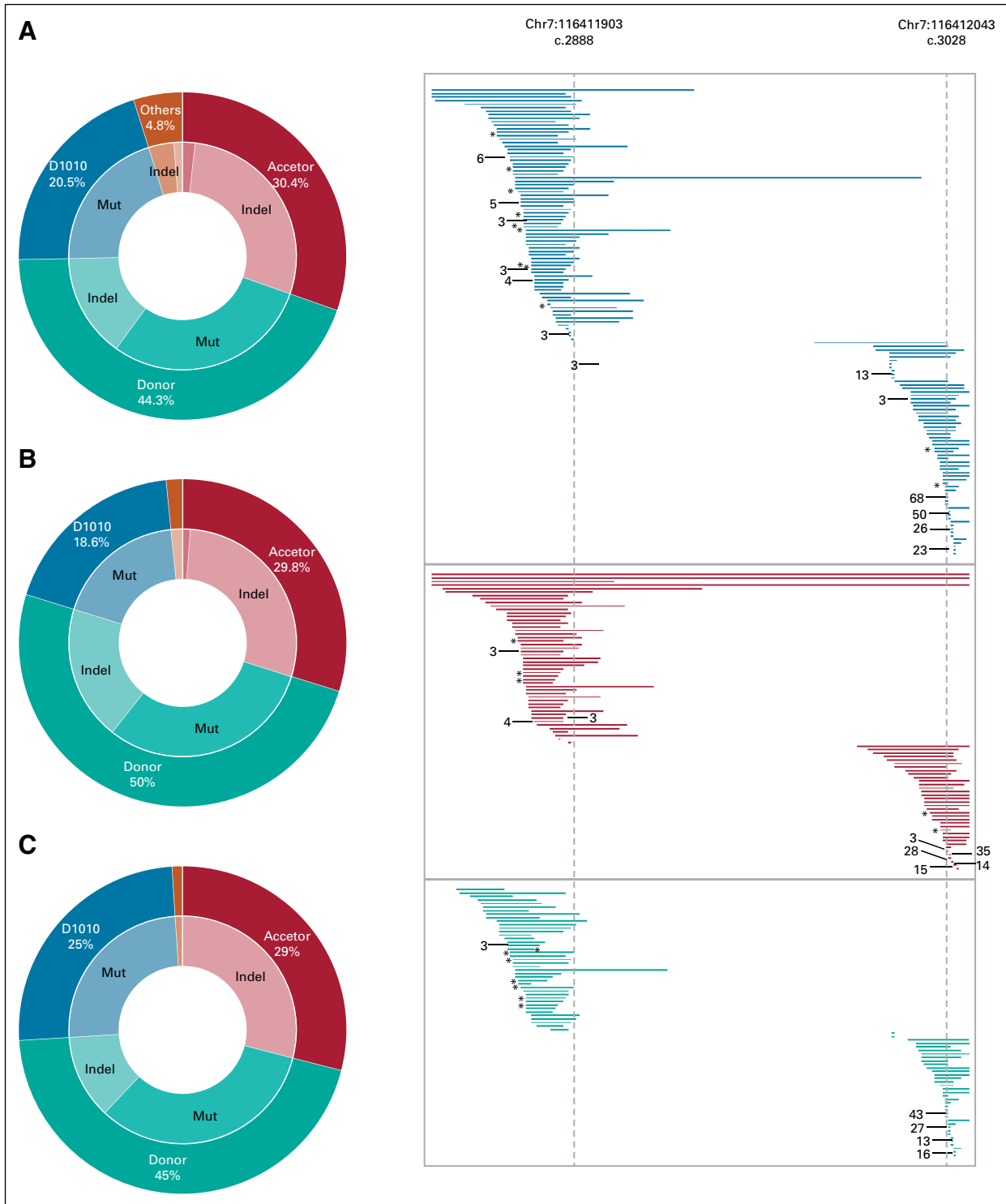


FIG 1. *MET* exon 14 skipping alterations mutation distribution in the three data sets: (A) Guardant360, (B) VISION, and (C) GenePlus. Genomic positions with alterations occurring in more than one case are indicated with an asterisk (*) for two and the number of cases is greater than two. Mut, mutation.

occurring with *EGFR* and *ERBB2* mutations, *MET*ex14 alterations were more frequently observed as a subclone, whereas in the *KRAS* and *BRAF* co-occurring cases, *KRAS* and *MET*ex14 had similar frequency of being clonal.

We next explored the potential impact of clonality of *MET*ex14 and coalterations on response to *MET* inhibitors, leveraging the VISION cohort. In the 62 patients with outcome data, 52 had tumors with clonal *MET*ex14 and 10

TABLE 1. Patient Characteristics and METex14 Functional Site by Data Set

Characteristics	Guardant360	Vision	GenePlus
No. of cases tested = 40,824	20,987	5,180	14,657
No. of METex14 = 692	332	188	172
Frequency, %	1.6	3.6	1.2
Age at tested, years			
Median (range)	73 (53-81)	72 (49-89)	69 (36-95)
Sex, No. (%)			
Female	197 (59)	101 (54)	83 (48)
Male	135 (41)	87 (46)	89 (52)
Race, No. (%)			
White	—	136 (72)	0
Asian	—	32 (17)	172 (100)
NA	332	20	—
Smoking history, No. (%)			
No	—	87 (46)	73 (42)
Yes	—	83 (44)	36 (21)
NA	332	18 (9)	63 (37)
Histologic subtype, No. (%)			
Adenocarcinoma	268 (81)	121 (64)	105 (61)
Squamous	34 (10)	16 (9)	9 (5)
Others	30 (9)	51 (27)	58 (33)
Functional site, No. (%)			
Acceptor site	101 (30.4)	56 (29.8)	50 (29)
Indel	95 (28.6)	54 (28.7)	50 (29)
Base substitution	6 (1.8)	2 (1.1)	0
Donor site	147 (44.3)	94 (50)	77 (45)
Indel	49 (14.7)	36 (19.1)	20 (12)
Base substitution	98 (29.5)	58 (30.8)	57 (33)
Y1003	13 (3.9)	—	2 (1)
D1010	68 (20.5)	35 (18.6)	43 (25)
Whole exon 14 deletion	0	3 (1.6)	0
Others	3 (0.9)	0	0

Abbreviation: METex14, MET exon 14 skipping alterations.

had subclonal. The response rate for clonal group was 46.1% and subclonal group was 50% (Data Supplement), suggesting similar responses to tepotinib regardless of clonality; however, this analysis was underpowered because of the low number in the subclonal cohort.

Of the 18 patients with concomitant alterations, seven had clinical outcome information with tepotinib (Data Supplement). One case with both METex14 and ERBB2 (L796_V797del) had a reduction in tumor size. The METex14 was not the clonal mutation for this patient. No response was seen in the remaining six patients with point mutations in PI3KCA (n = 2), K/NRAS (n = 2), and PTEN (n = 2) at baseline, regardless of the clonality of METex14.

Longitudinal ctDNA Analysis for METex14 NSCLC

In Guardant360, 23 cases had more than one blood sample collected at different time points. Two cases had acquired secondary mutations in the MET KDs (Fig 3D), Y1230C (GH#070) and D1228N (GH#146). In GH#146 (Data Supplement) with METex14 (VAF 9.9%) and an acquired MET D1228N (VAF 1.5%), an EGFR exon 20 insertion (A767_V769dup, VAF 3.9%) and EGFR amplification were also observed, all following therapy with crizotinib, suggesting that alterations in EGFR can mediate polyclonal resistance to MET TKI therapy. Conversely, EGFR driver alterations (defined with high VAF) were detected in 3 cases (GH#127, #134, and #162, Fig 3D), suggesting METex14 could be a resistance mechanism to EGFR TKI, consistent with previous publications.^{18,19} In GH#162, nine different ctDNA samples was taken over three years, and METex14 was acquired at the fifth ctDNA. EML4-ALK and BRAF V600E were also observed over the course of treatment, suggesting a high heterogeneity of resistance mechanisms. In GH#107, RET-CCDC6 fusion was acquired, following crizotinib and chemoimmunotherapy, with VAF of 1.53% relative to METex14 VAF 16.5%. The emergence of RET-CCDC6 was also confirmed in tumor tissue by FISH assay, appearing only after crizotinib, indicating RET fusion as an acquired potential resistant mechanism to MET TKI.

In the GenePlus data set, six patients had samples collected at different time points. Five were collected before and after crizotinib treatment, and one was taken before and after afatinib. MET D1228N was identified in two post-crizotinib cases (GP#52 and #53, Fig 3D and Data Supplement). In GP# 46, EGFR exon 20 insertion (S768_D770dup, dominant clone), METex14, and EGFR amplification were present before treatment. Following afatinib treatment, EGFR exon 20 and METex14 remained detectable in ctDNA, with loss of EGFR amplification. Taken together, we confirm secondary MET mutations as a consistent resistance mechanism to MET TKI therapy, and provide evidence of novel resistance mechanisms, such as acquired RET fusion or EGFR mutation.

Tissue and Liquid Biopsy Concordance for METex14 NSCLC

Ten cases from GenePlus cohort had both tissue and ctDNA profiling at the same time from the same patients (Data Supplement). METex14 were identified both in tissue and ctDNA at identical functional sites for all, demonstrating perfect concordance. Among them, four had identical comutation results. The remaining six had different co-occurring genomic alterations in TP53 (ctDNA only), MDM2 amplification (ctDNA only), PTEN mutation (tissue only), and four genomic mutations (EGFR, NF1, TP53, and RBI, tissue only, Data Supplement).

DISCUSSION

In this retrospective multicohort study, we identified 692 cases of METex14 NSCLC, including 557 ctDNA cases,

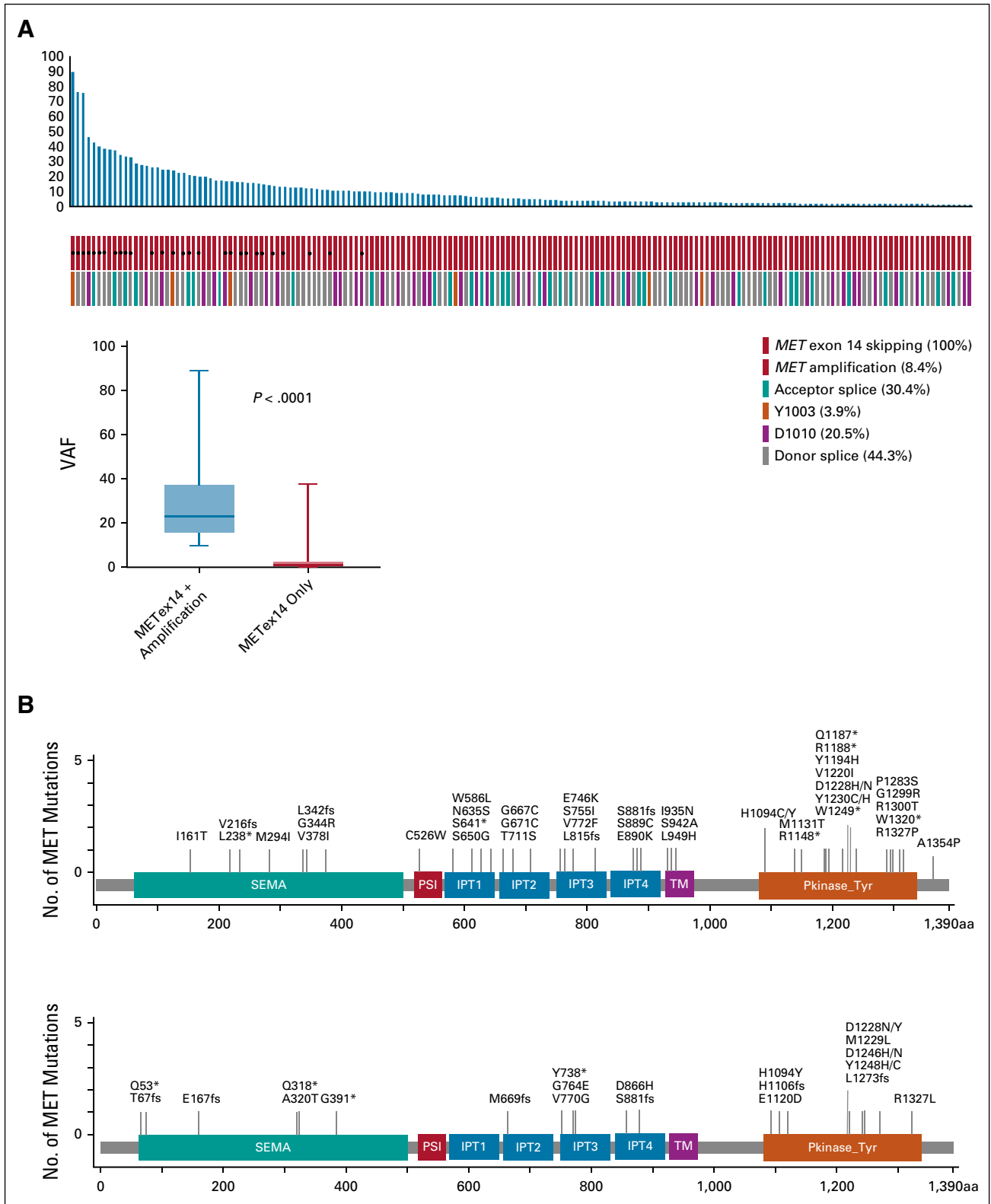


FIG 2. Co-occurring *MET* amplification and secondary mutations. (A) VAF of *MET*ex14 concurrent with *MET* amplification versus *MET*ex14 only in Guardant360. (B) Second site *MET* mutations in patients with *MET*ex14 non-small-cell lung cancer in Guardant360 and GenePlus data sets. IPT, immunoglobulin plexins transcription domains; *MET*ex14, *MET* exon 14 skipping alterations; PSI, plexins-semaphorin-integrin domain; TM, transmembrane domain; VAF, variant allele frequency.

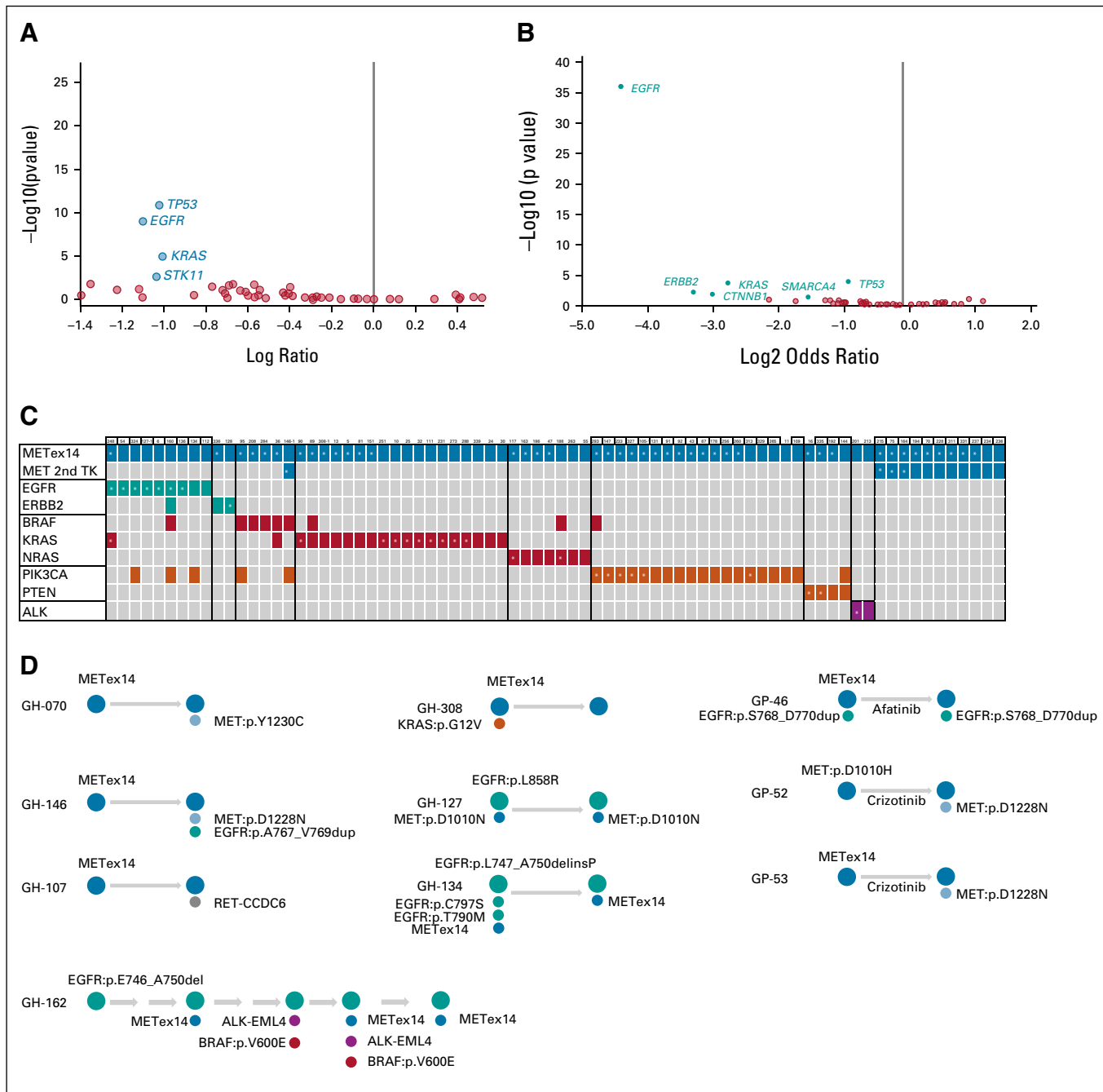


FIG 3. Co-occurring mutations with *METex14* and longitudinal ctDNA cases. (A and B) Volcano plots showing the difference of co-occurring alterations between *METex14* NSCLC and non-*METex14* NSCLC in (A) Guardant360 and (B) GenePlus data sets. (C) Cases with co-occurring alterations with *METex14* were shown, and the gene with dominant clone was labeled by star. (D) Genomic alterations of longitudinal cases at different time for patients with *METex14* NSCLC in Guardant360 (GH) and GenePlus (GP) data sets. *METex14*, *MET* exon 14 skipping alterations; NSCLC, non-small-cell lung cancer; TK, tyrosine kinase domain.

which is, to our knowledge, the largest ctDNA *METex14* NSCLC cohort reported to date. Our cohort confirms that patients with *METex14* NSCLC are older with equal sex distribution,²⁰⁻²² with a relatively high incidence in non-adenocarcinoma pathology. We separately confirmed these clinicopathologic features in both the Asian cohort and the predominantly Western data sets. The incidence of

METex14 in GenePlus data set was 1.17% in 14,657 cases, similar to prior reports of 0.9% in 1,296 Chinese lung cancer cases,^{23,24} also similar to the Guardant360 real-world data set with an incidence of 1.6%. Furthermore, the types of *METex14* alterations and the location were similar (Table 1), supporting that there are no significant differences between Asian and Western patients with *METex14* NSCLC.

Co-occurring genetic alterations with an oncogene driver can associate with clinical response or resistance. We found *MET*amp as the most frequent co-occurring alterations in *MET*ex14 NSCLC, at approximately 8%. A number of recent studies have reported on the outcomes of patients with *MET*ex14 and *MET*amp NSCLC to MET TKI, including four of the five patients achieving partial response in the VISION study⁴ and 75%-80% partial response in cohort 4 and 5b in GEOMETRY mono-1 study.³ With the small sample sizes acknowledged, these data indicate that *MET*ex14 concurrent with *MET*amp may have higher sensitivity to MET TKI. In our analysis, we found significantly higher VAF for *MET*ex14 when *MET*amp is detected. Although it is likely that the high VAF of *MET*ex14 with *MET*amp was related to increased copy number, as previously demonstrated in *EGFR*-mutant NSCLC,²⁵ *MET*ex14 and *MET*amp tumors represent a subgroup that are deeply addicted to aberrant *MET* signaling for tumorigenesis, and thus, a subgroup where the benefit of MET TKI is likely to be most pronounced. By contrast, secondary mutations in the *MET*KD are resistant mechanisms to MET TKI. In our study, a number of KD mutations, including D1228N in both Guardant360 and GenePlus, were detected, at a rate of 5%-6%, representing potential resistance mechanisms.

Using relative VAF to infer clonality, we analyzed other functional oncogenic alterations co-occurring with *MET*ex14. In the Guardant360 data set, most of the *EGFR* (9) and *ERBB2* (1) mutations had higher clonality than *MET*ex14, indicating that *EGFR* and *ERBB2* mutations were the dominant oncogene drivers, and suggesting that *MET*ex14 alterations were the potential drivers of resistance. This is consistent with the established notion that *MET*ex14 and *MET*amp are resistance mechanisms to EGFR TKI for *EGFR*-mutant NSCLC.^{18,19,26}

KRAS-activating mutations were found in only three cases in GenePlus cohort; however, 18 cases were identified in the Guardant data set,^{10,12,24} consistent with prior reports showing RAS alterations are more common in Western lung cancer populations.²⁷ *KRAS* mutations and *KRAS* and *BRAF* amplification constitute a cause of resistance to MET TKI on the basis of previous clinical and preclinical studies.^{9,12,28} We found that *KRAS* mutation and *MET*ex14 demonstrate similar tendency to be the dominant clone when co-occurring, as opposed to *EGFR* and *ERBB2*, which are usually the dominant driver. Now that both *KRAS* G12C²⁹ and *MET*ex14 have available targeted therapy

options, it is of great importance to recognize the dominant clone to prioritize treatment: ultimately, it is likely that dual inhibition of *KRAS* and *MET* pathways may be needed for these cases.

Through longitudinal ctDNA analysis, we identified, to our knowledge, the first reported case of acquired *RET-CCDC6* fusion co-occurring with *MET*ex14 (GH #107; Fig 3C). Acquired *RET* fusion has been reported in osimertinib-resistant *EGFR*-mutant NSCLC, where acquired resistance was overcome by *EGFR* plus *RET* inhibition.³⁰ Similar to *ALK* fusion concurrent with *MET*ex14, either combination therapy or a multikinase inhibitor would merit investigation as a therapeutic option.

Our study analyzed data sets from various sequencing platforms used in real-world and a clinical trial, which brings strengths as well as weaknesses. The addition of GenePlus cohort allowed a general comparison between Asian and Western populations, but the conclusion is limited by the heterogeneity of laboratory assays and sample source. Furthermore, the comutation and clonality analysis were only comparable across cohorts in the mutually tested 48 genes in all panels. Although they covered key oncogenic pathways in cancer, other genes of potential interest, such as *SMAD4* and *EZH2*, cannot be compared because they were only tested in Guardant360 and GenePlus 1,021 platforms. One important inherited limitation of real-world study is the incomplete data. In our study, for example, we did not have clinical outcome data to different therapies in the Guardant360 and GenePlus cohorts. As such, future studies, both experimental and clinical, are warranted to validate these provocative genomics findings and their clinical implications.

In conclusion, we describe a large cohort of *MET*ex14 NSCLC, mostly identified through ctDNA detection. We demonstrate that co-occurring *MET*amp is associated with high *MET*ex14 VAF and potential targeted therapy benefit, whereas *MET* KD secondary mutations are associated with targeted therapy resistance. When *MET*ex14 co-occurs with *EGFR* and *ERBB2* mutations, *MET*ex14 most commonly serves as a nondominant subclone and is a potential mediator of EGFR TKI resistance. Finally, we reveal emerging novel resistance mechanisms to MET TKI, such as *RET* fusion, which warrant future translational and therapeutic studies to overcome resistance.

AFFILIATIONS

¹Department of Thoracic/Head and Neck Medical Oncology, The University of Texas MD Anderson Cancer Center, Houston, TX

²Department of Oncology, Nanjing First Hospital, Nanjing Medical University, Nanjing, China

³Guardant Health Inc, Redwood City, CA

⁴Geneplus-Beijing Institute, Beijing, China

⁵Department of Investigational Cancer Therapeutics, The University of Texas MD Anderson Cancer Center, Houston, TX

⁶Division of Hematology/Oncology, Department of Medicine, Abramson Cancer Center, University of Pennsylvania, Philadelphia, PA

⁷Division of Oncology, Department of Medicine, Lombardi Comprehensive Cancer Center of Georgetown University, Washington, DC

⁸Merck KGaA (EMD Serono), Darmstadt, Germany

⁹Thoracic Oncology, Division of Solid Tumor Oncology, Department of Medicine, Memorial Sloan Kettering Cancer Center, New York, NY

CORRESPONDING AUTHOR

John V. Heymach, MD, PhD, Department of Thoracic/Head and Neck Medical Oncology, The University of Texas MD Anderson Cancer Center, Houston, TX 77030; e-mail: jheykach@mdanderson.org.

EQUAL CONTRIBUTION

X.L. and L.H. contributed equally to this work. J.Z. and J.V.H. shared the senior authorship.

SUPPORT

This work was supported by the generous philanthropic contributions to The University of Texas MD Anderson Lung Moon Shot Program and the MD Anderson Cancer Center Support Grant P30 CA016672.

AUTHOR CONTRIBUTIONS

Conception and design: Xiuning Le, Lingzhi Hong, Chuck Hensel, Rongrong Chen, Christopher Stroh, Anne Tsao, Paul Paik, Jianjun Zhang, John Heymach

Provision of study materials or patients: Christine A. Ciunci, Xuefeng Xia, David Hong, Victoria Raymond, Paul Paik

Collection and assembly of data: Xiuning Le, Lingzhi Hong, Chuck Hensel, Rongrong Chen, Haley Kemp, Niamh Coleman, Stephen V. Liu, Christopher Stroh, David Hong, Victoria Raymond, Paul Paik, Jianjun Zhang, John Heymach

Data analysis and interpretation: Xiuning Le, Lingzhi Hong, Chuck Hensel, Rongrong Chen, Niamh Coleman, Christine A. Ciunci, Stephen V. Liu, Marcelo V. Negrao, Jennifer Yen, Xuefeng Xia, Juergen Scheuenpflug, Christopher Stroh, Dilafuz Juraeva, Anne Tsao, David Hong, Victoria Raymond, Paul Paik, Jianjun Zhang, John Heymach

Manuscript writing: All authors

Final approval of manuscript: All authors

Accountable for all aspects of the work: All authors

AUTHORS' DISCLOSURES OF POTENTIAL CONFLICTS OF INTEREST

The following represents disclosure information provided by authors of this manuscript. All relationships are considered compensated unless otherwise noted. Relationships are self-held unless noted. I = Immediate Family Member, Inst = My Institution. Relationships may not relate to the subject matter of this manuscript. For more information about ASCO's conflict of interest policy, please refer to www.asco.org/rwc or ascopubs.org/po/author-center.

Open Payments is a public database containing information reported by companies about payments made to US-licensed physicians ([Open Payments](http://OpenPayments)).

Xiuning Le

Consulting or Advisory Role: AstraZeneca, Lilly, EMD Serono, Spectrum Pharmaceuticals, Daiichi Sankyo/Lilly

Research Funding: Lilly (Inst), Boehringer Ingelheim (Inst)

Chuck Hensel

Employment: Guardant Health

Stock and Other Ownership Interests: Guardant Health

Travel, Accommodations, Expenses: Guardant Health

Christine A. Ciunci

Honoraria: Imedex

Research Funding: Celgene (Inst), Merck (Inst), Bristol Myers Squibb (Inst), MacroGenics (Inst)

Stephen V. Liu

Consulting or Advisory Role: Genentech, Pfizer, Lilly, Bristol Myers Squibb, AstraZeneca, Takeda, Regeneron, G1 Therapeutics, Guardant Health, Janssen Oncology, MSD Oncology, Jazz Pharmaceuticals, Blueprint Medicines, Inivata, PharmaMar, Daiichi Sankyo/UCB Japan, BeiGene, Amgen, Turning Point Therapeutics, Elevation Oncology

Research Funding: Genentech/Roche (Inst), Pfizer (Inst), Bayer, Merck (Inst), AstraZeneca (Inst), Blueprint Medicines (Inst), Lilly (Inst), Rain Therapeutics (Inst), Alkermes (Inst), Bristol Myers Squibb (Inst), Turning Point Therapeutics (Inst), RAPT Therapeutics (Inst), Merus (Inst), Elevation Oncology (Inst), Erasca Inc (Inst)

Travel, Accommodations, Expenses: AstraZeneca, Roche/Genentech, MSD Oncology

Marcelo V. Negrao

Consulting or Advisory Role: Mirati Therapeutics

Research Funding: Mirati Therapeutics, AstraZeneca, Novartis, Pfizer, ZIOPHARM Oncology, Checkmate Pharmaceuticals

Jennifer Yen

Employment: Guardant Health

Stock and Other Ownership Interests: Guardant Health

Xuefeng Xia

Employment: Geneplus

Leadership: Geneplus

Juergen Scheuenpflug

Employment: Merck KGaA

Leadership: Merck KGaA

Stock and Other Ownership Interests: Merck KGaA

Christopher Stroh

Employment: Merck KGaA

Stock and Other Ownership Interests: Merck KGaA

Patents, Royalties, Other Intellectual Property: Predictive biomarker for anti-EGFR therapy

Dilafuz Juraeva

Employment: Merck

Stock and Other Ownership Interests: Merck KGaA

Anne Tsao

Consulting or Advisory Role: Novartis, Boehringer Ingelheim, Genentech/Roche, Lilly, Bristol Myers Squibb, Epizyme, AstraZeneca/MedImmune, ARIAD, EMD Serono, Takeda, HERON

Research Funding: Merck, Genentech/Roche, Seattle Genetics, Millennium, Bristol Myers Squibb, Boehringer Ingelheim, Polaris, EMD Serono, Takeda

Patents, Royalties, Other Intellectual Property: UptoDate

David Hong

Stock and Other Ownership Interests: OncoResponse, Telperian

Consulting or Advisory Role: Bayer, Guidepoint Global, GLG, Alphasights, Axiom Biotechnologies, Medscape, Numab, Pfizer, Seattle Genetics, Takeda, Trieza Therapeutics, WebMD, Infinity Pharmaceuticals, Amgen, Adaptimmune, Boxer Capital, EcoR1 Capital, Tavistock Life Sciences, Baxter, COG, Genentech, GroupH, Janssen, Acuta, HCW Precision, Prime Oncology, ST Cube, Alkermes, Aumbiosciences, Antheneum, Barclays, Bridgebio, CDR-Life, Cor2Ed, Gilead Sciences, Immunogen, Liberium, Oncologia Brasil, Pharma Intelligence, POET Congress, Turning Point Therapeutics, ZIOPHARM Oncology

Research Funding: Genentech (Inst), Amgen (Inst), Daiichi Sankyo (Inst), Adaptimmune (Inst), AbbVie (Inst), Bayer (Inst), Infinity Pharmaceuticals (Inst), Kite, a Gilead Company (Inst), MedImmune (Inst), National Cancer Institute (Inst), Fate Therapeutics (Inst), Pfizer (Inst), Novartis (Inst), Numab (Inst), Turning Point Therapeutics (Inst), Verastem (Inst), Kyowa (Inst), Loxo (Inst), Merck (Inst), Eisai (Inst), Genmab (Inst), Mirati Therapeutics (Inst), Mologen (Inst), Takeda (Inst), AstraZeneca (Inst),

Navire (Inst), VM Pharma (Inst), Erasca Inc (Inst), Bristol Myers Squibb (Inst), Adlai Nortye (Inst), Seagen (Inst), Deciphera (Inst), Pyramid Biosciences (Inst), Lilly (Inst)

Travel, Accommodations, Expenses: Genmab, Society for Immunotherapy of Cancer, Bayer Schering Pharma, ASCO, AACR, Telperian

Victoria Raymond

Employment: Guardant Health

Stock and Other Ownership Interests: Guardant Health, Trovagene

Paul Paik

Honoraria: Takeda, Calithera Biosciences, Boehringer Ingelheim, EMD Serono, AstraZeneca, Xencor, Bicara Therapeutics, GlaxoSmithKline

Consulting or Advisory Role: EMD Serono, Takeda, Calithera Biosciences, AstraZeneca, Boehringer Ingelheim, GlaxoSmithKline, Xencor, Bicara Therapeutics

Research Funding: EMD Serono, Boehringer Ingelheim, Bicara Therapeutics

Jianjun Zhang

Honoraria: Roche, Sino-USA Biomedical Platform, Geneplus, Origimed, Innovent Biologics, CancerNet, Zhejiang Cancer Hospital

Consulting or Advisory Role: AstraZeneca, Geneplus, Capital Medical University, AstraZeneca, Johnson & Johnson/Janssen, Novartis

Research Funding: Merck, Novartis

Travel, Accommodations, Expenses: Innovent Biologics, Zhejiang Cancer Hospital

John V. Heymach

Stock and Other Ownership Interests: Cardinal Spine, Bio-Tree

Consulting or Advisory Role: AstraZeneca, Bristol Myers Squibb, Spectrum Pharmaceuticals, Guardant Health, Hengrui Pharmaceutical, GlaxoSmithKline, EMD Serono, Lilly, Takeda, Sanofi/Aventis, Genentech/Roche, Boehringer Ingelheim, Catalyst Biotech, Foundation Medicine, Novartis, Mirati Therapeutics, BrightPath Biotherapeutics, Janssen, Nexus Health Systems, Pneuma Respiratory, Kairos Ventures, Roche, Leads Biolabs

Research Funding: AstraZeneca (Inst), Spectrum Pharmaceuticals, GlaxoSmithKline

Patents, Royalties, Other Intellectual Property: Licensing agreement between Spectrum and MD Anderson (including myself) regarding intellectual property for treatment of EGFR and HER2 exon 20 mutations

No other potential conflicts of interest were reported.

REFERENCES

- Awad MM, Oxnard GR, Jackman DM, et al: MET exon 14 mutations in non-small-cell lung cancer are associated with advanced age and stage-dependent MET genomic amplification and c-met overexpression. *J Clin Oncol* 34:721-730, 2016
- Drilon A, Clark JW, Weiss J, et al: Antitumor activity of crizotinib in lung cancers harboring a MET exon 14 alteration. *Nat Med* 26:47-51, 2020
- Wolf J, Seto T, Han JY, et al: Capmatinib in MET exon 14-mutated or MET-amplified non-small-cell lung cancer. *N Engl J Med* 383:944-957, 2020
- Paik PK, Felip E, Veillon R, et al: Tepotinib in non-small-cell lung cancer with MET exon 14 skipping mutations. *N Engl J Med* 383:931-943, 2020
- Lu S, Fang J, Li X, et al: Phase II study of savolitinib in patients (pts) with pulmonary sarcomatoid carcinoma (PSC) and other types of non-small cell lung cancer (NSCLC) harboring MET exon 14 skipping mutations (METex14+). *J Clin Oncol* 38, 2020 (suppl; abstr 9519)
- Awad MM, Leonardi GC, Kravets S, et al: Impact of MET inhibitors on survival among patients with non-small cell lung cancer harboring MET exon 14 mutations: A retrospective analysis. *Lung Cancer* 133:96-102, 2019
- Offin M, Luo J, Guo R, et al: CNS metastases in patients with MET exon 14-altered lung cancers and outcomes with crizotinib. *JCO Precis Oncol* 4, 2020. doi:10.1200/PO.20.00098
- Fujino T, Kobayashi Y, Suda K, et al: Sensitivity and resistance of MET exon 14 mutations in lung cancer to eight MET tyrosine kinase inhibitors in vitro. *J Thorac Oncol* 14:1753-1765, 2019
- Rotow JK, Gui P, Wu W, et al: Co-occurring alterations in the RAS-MAPK pathway limit response to MET inhibitor treatment in MET exon 14 skipping mutation-positive lung cancer. *Clin Cancer Res* 26:439-449, 2020
- Schrock AB, Frampton GM, Suh J, et al: Characterization of 298 patients with lung cancer harboring MET exon 14 skipping alterations. *J Thorac Oncol* 11:1493-1502, 2016
- Recondo G, Bahcall M, Spurr LF, et al: Molecular mechanisms of acquired resistance to MET tyrosine kinase inhibitors in patients with MET exon 14-mutant NSCLC. *Clin Cancer Res* 26:2615-2625, 2020
- Suzawa K, Offin M, Lu D, et al: Activation of KRAS mediates resistance to targeted therapy in MET exon 14-mutant non-small cell lung cancer. *Clin Cancer Res* 25:1248-1260, 2019
- Jamme P, Fernandes M, Copin MC, et al: Alterations in the PI3K pathway drive resistance to MET inhibitors in NSCLC harboring MET exon 14 skipping mutations. *J Thorac Oncol* 15:741-751, 2020
- Jorge SE, Schulman S, Freed JA, et al: Responses to the multitargeted MET/ALK/ROS1 inhibitor crizotinib and co-occurring mutations in lung adenocarcinomas with MET amplification or MET exon 14 skipping mutation. *Lung Cancer* 90:369-374, 2015
- Sabari JK, Leonardi GC, Shu CA, et al: PD-L1 expression, tumor mutational burden, and response to immunotherapy in patients with MET exon 14 altered lung cancers. *Ann Oncol* 29:2085-2091, 2018
- Odegaard JI, Vincent JJ, Mortimer S, et al: Validation of a plasma-based comprehensive cancer genotyping assay utilizing orthogonal tissue- and plasma-based methodologies. *Clin Cancer Res* 24:3539-3549, 2018
- Tate JG, Bamford S, Jubb HC, et al: COSMIC: the catalogue of somatic mutations in cancer. *Nucleic Acids Res* 47 (D1): D941-D947, 2019
- Suzawa K, Offin M, Schoenfeld AJ, et al: Acquired MET exon 14 alteration drives secondary resistance to epidermal growth factor receptor tyrosine kinase inhibitor in EGFR-mutated lung cancer. *JCO Precis Oncol* 3, 2019. doi:10.1200/PO.19.00011
- Schoenfeld AJ, Chan JM, Kubota D, et al: Tumor analyses reveal squamous transformation and off-target alterations as early resistance mechanisms to first-line osimertinib in EGFR-mutant lung cancer. *Clin Cancer Res* 26:2654-2663, 2020
- Zhang YL, Yuan JQ, Wang KF, et al: The prevalence of EGFR mutation in patients with non-small cell lung cancer: A systematic review and meta-analysis. *Oncotarget* 7:78985-78993, 2016
- Bergthson K, Shaw AT, Ou SH, et al: ROS1 rearrangements define a unique molecular class of lung cancers. *J Clin Oncol* 30:863-870, 2012
- Shaw AT, Yeap BY, Mino-Kenudson M, et al: Clinical features and outcome of patients with non-small-cell lung cancer who harbor EML4-ALK. *J Clin Oncol* 27:4247-4253, 2009
- Liu SY, Gou LY, Li AN, et al: The unique characteristics of MET exon 14 mutation in Chinese patients with NSCLC. *J Thorac Oncol* 11:1503-1510, 2016

24. Yang H, Zhou Z, Lin L, et al: Characterization of MET exon 14 alteration and association with clinical outcomes of crizotinib in Chinese lung cancers. *Lung Cancer* 148:113-121, 2020
25. Lam VK, Zhang J, Wu CC, et al: Genotype-specific differences in circulating tumor DNA levels in advanced NSCLC. *J Thorac Oncol* 16:601-609, 2020
26. Le X, Puri S, Negrao MV, et al: Landscape of EGFR-dependent and -independent resistance mechanisms to osimertinib and continuation therapy beyond progression in EGFR-mutant NSCLC. *Clin Cancer Res* 24:6195-6203, 2018
27. Carrot-Zhang J, Chambwe N, Damrauer JS, et al: Comprehensive analysis of genetic ancestry and its molecular correlates in cancer. *Cancer Cell* 37:639-654.e6, 2020
28. Bahcall M, Awad MM, Sholl LM, et al: Amplification of wild-type KRAS imparts resistance to crizotinib in MET exon 14 mutant non-small cell lung cancer. *Clin Cancer Res* 24:5963-5976, 2018
29. Hong DS, Fakih MG, Strickler JH, et al: KRAS(G12C) inhibition with sotorasib in advanced solid tumors. *N Engl J Med* 383:1207-1217, 2020
30. Piotrowska Z, Isozaki H, Lennerz JK, et al: Landscape of acquired resistance to osimertinib in EGFR-mutant NSCLC and clinical validation of combined EGFR and RET inhibition with osimertinib and BLU-667 for acquired RET fusion. *Cancer Discov* 8:1529-1539, 2018

