

Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.



See related article on page 454

COMMENTARY

The American Journal of **PATHOLOGY** aip.amipathol.org

Check for updates

Lung Injury and Repair in Coronavirus Disease 2019—Related Acute Lung Injury

Thomas R. Martin

From the Division of Pulmonary, Critical Care and Sleep Medicine, Department of Medicine, University of Washington School of Medicine, Seattle, Washington

The alveolar epithelium is the largest surface area of the body in continuous contact with the outside environment and is critical for effective gas exchange to support metabolism. The alveolar walls consist of three distinct layers: the alveolar epithelium, comprising type I and type II alveolar epithelial cells; a matrix scaffold; and the microvascular endothelium. Most of the alveolar surface is covered by thin type I epithelial cells that cover the matrix framework and are closely opposed to microvascular endothelial cells. Type II epithelial cells are found in each alveolar unit and produce surfactant lipids and proteins that reduce surface tension at low lung volumes and function in antimicrobial defense. The alveolar epithelium is a tight barrier that prevents fluid movement into the airspaces. Alveolar epithelial cells also reabsorb fluid from the airspaces by active transport through specialized ion channels. By contrast, the microvascular endothelial barrier changes permeability more readily in response to intravascular stimuli, resulting in interstitial edema that is cleared by lymphatic vessels. Alveolar edema forms in response to increased microvascular hydrostatic pressure, as in left heart failure, or an increase in endothelial or epithelial permeability when endothelial or epithelial cell damage occurs. Rapid repair of increased epithelial or endothelial permeability is essential to support continuing gas exchange. In some cases, such as pneumococcal pneumonia, the repair process reestablishes normal alveolar architecture without significant fibrosis, whereas in others repair leads to fibrosis, which can distort alveolar units and lead to long-term abnormalities in gas exchange and pulmonary function. Accordingly, a great deal of research has focused on identifying the mechanisms that govern normal alveolar repair versus fibrosis after various forms of acute lung injury.¹

The coronavirus disease 2019 (COVID-19) pandemic has focused attention on severe lung injury, termed adult respiratory distress syndrome (ARDS). New work by Ting et al² uses COVID-19 cases to generate a new perspective on lung injury and repair in ARDS. ARDS is a form of severe lung injury caused by a variety of different stimuli that usually results in diffuse alveolar damage, characterized by increased permeability, alveolar edema, leukocyte infiltration, type II pneumocyte proliferation, hyaline membrane formation, surfactant dysfunction, and alveolar collapse, all of which result in life-threatening hypoxemia that requires high-level respiratory support.^{3,4} In ARDS from a variety of causes, evidence of active repair processes is present in the first days after onset of the illness, reflected by increased concentrations of procollagen III. transforming growth factor- α , and other growth factors in bronchoalveolar lavage fluid.^{5–7} Although lung function improves in most survivors of ARDS, some experience a persistent fibroproliferative response that impairs pulmonary function and causes lasting disability.^{8,9} The lungs of patients with COVID-19-related ARDS who have undergone autopsy have shown diffuse alveolar damage that is difficult to distinguish from other causes of ARDS.^{10,11} Because COVID-19-related ARDS occurs after a well-defined viral stimulus [severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2)], COVID-19 provides a relevant clinical condition in which to study factors that cause lung injury and repair, to reduce the heterogeneity inherent in studying ARDS from diverse causes.

Alveolar type II cells are the principal cells that give rise to the type I pneumocytes that make up most of the alveolar surface.¹ In normal homeostasis, type II cells are maintained by a subpopulation of rare progenitor cells that are

Accepted for publication January 5, 2022.

Disclosures: None declared.

Address correspondence to Thomas R. Martin, M.D., Department of Medicine, University of Washington School of Medicine, Seattle, WA 98195-6522. E-mail: trmartin@uw.edu.

long-lived and self-renewing. Type II progenitor cells can be isolated by unique surface markers and have distinct transcriptional and functional phenotypes.^{12,13} They are usually found near fibroblasts that express Wnt genes, generating a niche for type II progenitor cells.¹² Following injury, remaining type II cells and the subpopulation of type II progenitor cells proliferate, then enter a non-proliferative transitional state with cuboidal or partially spread morphology, then flatten and acquire characteristics of mature type I cells.^{12–14} Lineage tracing studies in mice with bleomycin-induced lung injury identified the transitional cells expressing the cytokeratin 8 marker (KRT 8^+) as key cells in the progression from type II cells to mature type I cells.¹⁵ Studies of lung tissue from patients with idiopathic pulmonary fibrosis (IPF), a form of chronic fibrotic lung disease, have shown persistent transitional cells and a relative paucity of mature type I cells, leading to the hypothesis that a delay or block in progression from the transitional state to mature type I epithelial cells may be a key part of ineffective repair leading to fibrosis.^{15–18} The role of transitional epithelial cells in ineffective repair and whether ineffective repair occurs relatively early following acute lung injury are key questions to be resolved.

To study characteristics of alveolar injury and epithelial repair in COVID-19-related lung injury, Ting et al² used autopsy material from the lungs of three patients who died within 14 days of onset of COVID-19-related ARDS. For comparison, they used archived lung tissue from three patients with non-COVID-19 ARDS, lung tissue from patients with IPF who were undergoing lung transplantation, and normal tissue from lungs rejected for transplantation. As previously reported for COVID-19 and non-COVID-19 ARDS, the lungs of all of the patients showed extensive diffuse alveolar damage, consistent with severe epithelial injury. Labeling of COVID-19 lung tissue for type II and type I epithelial cell markers [pro-surfactant protein C (pro-SPC) and human type I antigen 56 (HTI-56), respectively] showed extensive loss of mature type II cells and widespread damage to type I cells. In COVID-19 lungs, hyperplastic cuboidal epithelial cells were identified, which is a common finding in diffuse alveolar damage, but for the most part these cells did not express the pro-SPC marker, suggesting that they were not mature type II cells. Labeling of lung tissue with the KRT8 marker showed prominent labeling throughout alveolar regions, suggesting that KRT8⁺ transitional cells were abundant in COVID-19-related as well as non-COVID-19-related injured lungs. KRT8 is an intracellular cytokeratin marker that is expressed at high levels in transitional alveolar epithelial cells of mice with bleomycin-induced lung injury.¹⁵ KRT8⁺ transitional cells have a flattened morphology and show evidence of p53 and NF-kB activation and have been identified in different animal models of lung injury, as well as lungs of humans with IPF. KRT8⁺ epithelial cells have been proposed as key transitional cells in the progression from type II to type I cells, and abnormal persistence of KRT8⁺ cells is thought to be a key feature in the progression from acute lung injury to chronic fibrosis.^{15,16}

The findings of Ting et al^2 show that type II epithelial cells are lost and KRT8⁺ transitional epithelial cells appear early after the onset of acute lung injury in humans, but raise the question of whether the appearance of transitional cells is a marker of impending fibrosis. Despite the widespread loss of type II cells and the abundance of transitional cells, the lungs from COVID-19 and non-COVID-19 ARDS decedents did not show evidence of collagen deposition, myofibroblast accumulation, or distortion of the underlying alveolar matrix structure, all of which are common findings in IPF and fibroproliferative ARDS.⁸ Although most KRT8⁺ transitional cells did not express the HTI-56 type I cell marker, HTI-56 was detected on rare KRT8⁺ cells in the COVID-19 lungs, suggesting incomplete progression of transitional cells to the mature type I epithelial phenotype. Further labeling of lung sections with markers of cell cycle arrest and senescence showed that although epithelial markers of cell cycle arrest (CDKN1A, CDKN2B, TP53, and CCND1) were detectable in lung tissue of COVID-19 and non-COVID-19 ARDS patients, only the lung tissue from patients with IPF expressed a relatively specific marker of senescence, p26 (CDKN2A). Taken together, these findings led the authors to hypothesize that in patients with early COVID-19 and non-COVID-19 ARDS, proliferating type II epithelial cells or related progenitor cells become transitional cells in a state of cell cycle arrest and then differentiate into mature type I cells to restore normal alveolar architecture, whereas in fibroproliferative ARDS or in IPF, the transitional cells become senescent and lose the capacity for type I differentiation, leading to the development of fibrosis. These findings are important, because they show that transitional cells arise relatively early in the course of severe lung injury due to COVID-19 as well as non-COVID-19 etiologies, before evidence of fibrosis can be detected. Whether fibroproliferative changes might have occurred in the lungs of these patients over a longer period cannot be determined from these autopsy samples.

The findings of Ting et al^2 are important because they provide human evidence in support of translational studies of alveolar regeneration in mice.¹⁵ They focus attention on transitional cells in lung epithelial repair and the factors that govern the progression from type II cells to transitional cells and then normal type I epithelial cells, versus the development of senescence in transitional epithelial cells with associated fibrosis. The inflammatory milieu of the injured lung is a complex environment, containing a variety of growth factors and cytokines that influence the balance between normal epithelial regeneration and fibrosis, including tumor necrosis factor- α , IL-1 β , transforming growth factor- α , keratinocyte growth factor, granulocyte-macrophage colony-stimulating factor, and others, and persistent inflammation has been proposed as an important factor promoting fibrosis.^{5,19-22} Sorting out the key factors that promote progression from KRT8⁺ transitional cells to mature type I pneumocytes

versus the alternative mechanisms that activate fibrosis is an important goal of future research.

The study by Ting et al^2 has some limitations that are inherent in studies of human autopsy material. First, the COVID-19 and non-COVID-19 ARDS lung samples were obtained at single times within 2 weeks of onset of acute lung injury, so the data do not provide insight into the progression of the findings over longer times. Second, although the investigators performed comprehensive analyses on the lung tissue samples, they studied tissue from a limited number of patients, raising the question of how uniform the findings might be in a larger population. Third, although the index cases of COVID-19 ARDS all had a common primary viral injury, all of them also were mechanically ventilated, adding a second and different injurious stimulus that must be considered. Although the authors did not find evidence of persistent viral infection in the lungs, how initial viral damage to type II pneumocytes might interact with ongoing injury from mechanical forces in the lungs remains an important question. The study was not designed to consider how other levels of complexity, including damage to the alveolar wall matrix and epithelial mesenchymal cross talk, might contribute to aberrant repair.²³⁻²⁵ Last, although the idea that development of senescence in KRT8⁺ transitional cells due to repetitive injury may be linked with the onset of fibrosis is intriguing, the IPF tissue samples were collected much later in the course of that illness, so the direct relevance of the findings for the evolution from acute to chronic fibroproliferative lung injury remains to be proven.

Despite these limitations, the study by Ting et al² provides important directions for future research on lung repair. The factors that favor the maturation of KRT8⁺ transitional cells into normal type I epithelial cells versus factors that lead to a senescent state need to be clarified and studied in a larger number of patients as well as relevant animal models. The results could provide clues about new therapeutic approaches to promote normal repair and/or prevent fibrosis. The results may also be useful in evaluating whether currently available therapies for chronic fibrosis might be helpful early after the onset of acute lung injury.

References

- Zemans RL, Henson PM, Henson JE, Janssen WJ: Conceptual approaches to lung injury and repair. Ann Am Thorac Soc 2015, 12(Suppl 1):S9–S15
- Ting C, Aspal M, Vaishampayan N, Huang SK, Riemondy KA, Wang F, Farver C, Zemans RL: Fatal COVID-19 and non-COVID-19 acute respiratory distress syndrome is associated with incomplete alveolar type 1 epithelial cell differentiation from the transitional state without fibrosis. Am J Pathol 2022, 192:454–467
- Matthay MA, Zemans RL, Zimmerman GA, Arabi YM, Beitler JR, Mercat A, Herridge M, Randolph AG, Calfee CS: Acute respiratory distress syndrome. Nat Rev Dis Primers 2019, 5:18
- Bachofen M, Weibel ER: Alterations of the gas exchange apparatus in adult respiratory insufficiency associated with septicemia. Am Rev Respir Dis 1977, 116:589–615

- Madtes DK, Rubenfeld G, Klima LD, Milberg JA, Steinberg KP, Martin TR, Raghu G, Hudson LD, Clark JG: Elevated transforming growth factor-alpha levels in bronchoalveolar lavage fluid of patients with acute respiratory distress syndrome. Am J Respir Crit Care Med 1998, 158:424–430
- Clark JG, Milberg JA, Steinberg KP, Hudson LD: Type III procollagen peptide in the adult respiratory distress syndrome: association of increased peptide levels in bronchoalveolar lavage fluid with increased risk for death. Ann Intern Med 1995, 122:17–23
- Chesnutt AN, Matthay MA, Tibayan FA, Clark JG: Early detection of type III procollagen peptide in acute lung injury: pathogenetic and prognostic significance. Am J Respir Crit Care Med 1997, 156: 840–845
- Burnham EL, Janssen WJ, Riches DW, Moss M, Downey GP: The fibroproliferative response in acute respiratory distress syndrome: mechanisms and clinical significance. Eur Respir J 2014, 43: 276–285
- 9. Herridge MS, Cheung AM, Tansey CM, Matte-Martyn A, Diaz-Granados N, Al-Saidi F, Cooper AB, Guest CB, Mazer CD, Mehta S, Stewart TE, Barr A, Cook D, Slutsky AS, Canadian Critical Care Trials Group: One-year outcomes in survivors of the acute respiratory distress syndrome. N Engl J Med 2003, 348:683–693
- Peiris S, Mesa H, Aysola A, Manivel J, Toledo J, Borges-Sa M, Aldighieri S, Reveiz L: Pathological findings in organs and tissues of patients with COVID-19: a systematic review. PLoS One 2021, 16: e0250708
- 11. Konopka KE, Nguyen T, Jentzen JM, Rayes O, Schmidt CJ, Wilson AM, Farver CF, Myers JL: Diffuse alveolar damage (DAD) resulting from coronavirus disease 2019 infection is morphologically indistinguishable from other causes of DAD. Histopathology 2020, 77: 570–578
- Nabhan AN, Brownfield DG, Harbury PB, Krasnow MA, Desai TJ: Single-cell Wnt signaling niches maintain stemness of alveolar type 2 cells. Science 2018, 359:1118–1123
- Zacharias WJ, Frank DB, Zepp JA, Morley MP, Alkhaleel FA, Kong J, Zhou S, Cantu E, Morrisey EE: Regeneration of the lung alveolus by an evolutionarily conserved epithelial progenitor. Nature 2018, 555: 251–255
- Desai TJ, Brownfield DG, Krasnow MA: Alveolar progenitor and stem cells in lung development, renewal and cancer. Nature 2014, 507: 190–194
- 15. Strunz M, Simon LM, Ansari M, Kathiriya JJ, Angelidis I, Mayr CH, Tsidiridis G, Lange M, Mattner LF, Yee M, Ogar P, Sengupta A, Kukhtevich I, Schneider R, Zhao Z, Voss C, Stoeger T, Neumann JHL, Hilgendorff A, Behr J, O'Reilly M, Lehmann M, Burgstaller G, Konigshoff M, Chapman HA, Theis FJ, Schiller HB: Alveolar regeneration through a Krt8+ transitional stem cell state that persists in human lung fibrosis. Nat Commun 2020, 11:3559
- 16. Kobayashi Y, Tata A, Konkimalla A, Katsura H, Lee RF, Ou J, Banovich NE, Kropski JA, Tata PR: Persistence of a regenerationassociated, transitional alveolar epithelial cell state in pulmonary fibrosis. Nat Cell Biol 2020, 22:934–946
- 17. Jiang P, Gil de Rubio R, Hrycaj SM, Gurczynski SJ, Riemondy KA, Moore BB, Omary MB, Ridge KM, Zemans RL: Ineffectual type 2-totype 1 alveolar epithelial cell differentiation in idiopathic pulmonary fibrosis: persistence of the KRT8(hi) transitional state. Am J Respir Crit Care Med 2020, 201:1443–1447
- Auyeung VC, Sheppard D: Stuck in a moment: does abnormal persistence of epithelial progenitors drive pulmonary fibrosis? Am J Respir Crit Care Med 2021, 203:667–669
- 19. Park WY, Goodman RB, Steinberg KP, Ruzinski JT, Radella F 2nd, Park DR, Pugin J, Skerrett SJ, Hudson LD, Martin TR: Cytokine balance in the lungs of patients with acute respiratory distress syndrome. Am J Respir Crit Care Med 2001, 164:1896–1903
- 20. Verghese GM, McCormick-Shannon K, Mason RJ, Matthay MA: Hepatocyte growth factor and keratinocyte growth factor in the pulmonary edema fluid of patients with acute lung injury; biologic

and clinical significance. Am J Respir Crit Care Med 1998, 158: 386–394

- 21. Matute-Bello G, Liles WC, Radella F 2nd, Steinberg KP, Ruzinski JT, Hudson LD, Martin TR: Modulation of neutrophil apoptosis by granulocyte colony-stimulating factor and granulocyte/macrophage colony-stimulating factor during the course of acute respiratory distress syndrome. Crit Care Med 2000, 28:1–7
- 22. Choi J, Park JE, Tsagkogeorga G, Yanagita M, Koo BK, Han N, Lee JH: Inflammatory signals induce AT2 cell-derived damage-associated transient progenitors that mediate alveolar regeneration. Cell Stem Cell 2020, 27:366–382.e7
- Yao L, Zhou Y, Li J, Wickens L, Conforti F, Rattu A, Ibrahim FM, Alzetani A, Marshall BG, Fletcher SV, Hancock D, Wallis T,

Downward J, Ewing RM, Richeldi L, Skipp P, Davies DE, Jones MG, Wang Y: Bidirectional epithelial-mesenchymal crosstalk provides selfsustaining profibrotic signals in pulmonary fibrosis. J Biol Chem 2021, 297:101096

- 24. Schmidt EP, Yang Y, Janssen WJ, Gandjeva A, Perez MJ, Barthel L, Zemans RL, Bowman JC, Koyanagi DE, Yunt ZX, Smith LP, Cheng SS, Overdier KH, Thompson KR, Geraci MW, Douglas IS, Pearse DB, Tuder RM: The pulmonary endothelial glycocalyx regulates neutrophil adhesion and lung injury during experimental sepsis. Nat Med 2012, 18:1217–1223
- Deng Z, Fear MW, Suk Choi Y, Wood FM, Allahham A, Mutsaers SE, Prele CM: The extracellular matrix and mechanotransduction in pulmonary fibrosis. Int J Biochem Cell Biol 2020, 126:105802