



































OPEN

# Climatic and soil factors explain the two-dimensional spectrum of global plant trait variation

Julia S. Joswig <sup>1,2</sup> ✉, Christian Wirth <sup>1,3,4</sup>, Meredith C. Schuman <sup>2,5</sup>, Jens Kattge <sup>1,3</sup>, Björn Reu<sup>6</sup>, Ian J. Wright <sup>7</sup>, Sebastian D. Sippel <sup>8,9</sup>, Nadja Rüger <sup>3,10,11</sup>, Ronny Richter <sup>3,4,12</sup>, Michael E. Schaepman <sup>2</sup>, Peter M. van Bodegom <sup>13</sup>, J. H. C. Cornelissen<sup>14</sup>, Sandra Díaz <sup>15</sup>, Wesley N. Hattingh <sup>16</sup>, Koen Kramer <sup>17,18</sup>, Frederic Lens <sup>19,20</sup>, Ülo Niinemets <sup>21</sup>, Peter B. Reich <sup>22,23,24</sup>, Markus Reichstein <sup>1,3</sup>, Christine Römermann <sup>3,25</sup>, Franziska Schrodte <sup>26</sup>, Madhur Anand<sup>27</sup>, Michael Bahn <sup>28</sup>, Chaeho Byun <sup>29</sup>, Giandiego Campetella <sup>30</sup>, Bruno E. L. Cerabolini <sup>31</sup>, Joseph M. Craine <sup>32</sup>, Andres Gonzalez-Melo<sup>33</sup>, Alvaro G. Gutiérrez <sup>34</sup>, Tianhua He<sup>35,36</sup>, Pedro Higuchi <sup>37</sup>, Hervé Jactel<sup>38</sup>, Nathan J. B. Kraft<sup>39</sup>, Vanessa Minden<sup>40,41</sup>, Vladimir Onipchenko <sup>42</sup>, Josep Peñuelas <sup>43,44</sup>, Valério D. Pillar <sup>45</sup>, Ênio Sosinski <sup>46</sup>, Nadejda A. Soudzilovskaia<sup>47,48</sup>, Evan Weiher <sup>49</sup> and Miguel D. Mahecha<sup>3,50,51</sup>

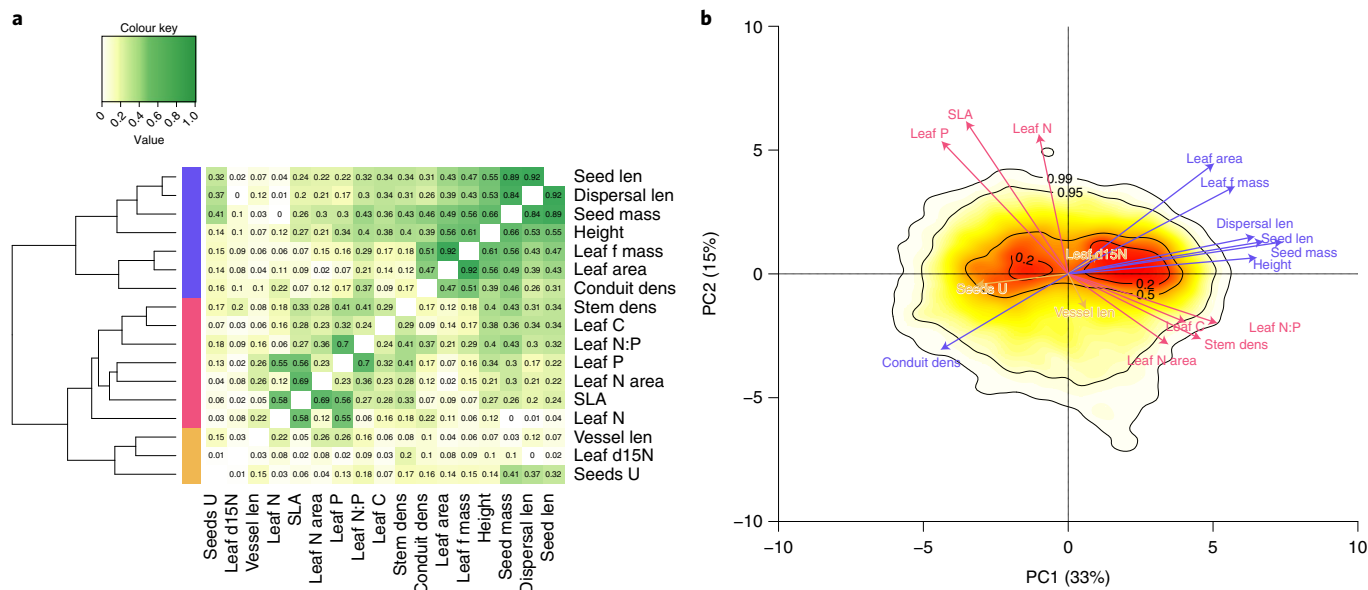
**Plant functional traits can predict community assembly and ecosystem functioning and are thus widely used in global models of vegetation dynamics and land-climate feedbacks. Still, we lack a global understanding of how land and climate affect plant traits. A previous global analysis of six traits observed two main axes of variation: (1) size variation at the organ and plant level and (2) leaf economics balancing leaf persistence against plant growth potential. The orthogonality of these two axes suggests they are differently influenced by environmental drivers. We find that these axes persist in a global dataset of 17 traits across more than 20,000 species. We find a dominant joint effect of climate and soil on trait variation. Additional independent climate effects are also observed across most traits, whereas independent soil effects are almost exclusively observed for economics traits. Variation in size traits correlates well with a latitudinal gradient related to water or energy limitation. In contrast, variation in economics traits is better explained by interactions of climate with soil fertility. These findings have the potential to improve our understanding of biodiversity patterns and our predictions of climate change impacts on biogeochemical cycles.**

Plant functional traits have proved useful in identifying life history strategies<sup>1,2</sup> for predicting plant community assembly<sup>3,4</sup> and for assessing the impact of vegetation composition and diversity on ecosystem functioning<sup>5,6</sup>. Consequently, vegetation models including coupled climate-vegetation models benefit from a better representation of plant trait variation to adequately analyse terrestrial biosphere dynamics under global change<sup>6,7</sup>. Today, in combination with advanced gap-filling techniques<sup>8</sup>, databases of plant traits have sufficient coverage to allow quantitative analyses of plant form and function at the global scale<sup>9</sup>. Analysing six fundamental traits, Díaz and colleagues<sup>10</sup> revealed that essential patterns of form and function across the plant kingdom can be captured by two main axes. The first reflects the size spectrum of whole plants and plant organs. The second axis corresponds to the ‘leaf economics spectrum’<sup>11</sup> emerging from the necessity for plants to balance leaf persistence against plant growth potential. The concept of a global spectrum of plant form and function has since been investigated from various perspectives<sup>12–14</sup>. It has been shown, for instance, that orthogonal axes of variation in size and economics traits emerge even in the extreme tundra biome<sup>13</sup> or at the scale of plant communities<sup>12</sup>. However, it remains unclear whether the two axes remain

dominant for extended sets of traits or when differentiating among growth forms. A particular knowledge gap is what environmental controls determine these two axes of plant form and function.

There is ample evidence that large-scale variation of individual plant traits is related to environmental gradients. Early plant biogeographers suggested that climate and soils together shape plant form and function<sup>15–17</sup> but could not propose a more precise theoretical framework describing these fundamental relationships. Over the last decades, examples have thus accumulated without an overall framework in which to place them<sup>13,18,19</sup>. For instance, tree height depends on water availability<sup>20,21</sup> while leaf economics traits depend on soil properties, especially soil nutrient supply, as well as on climatic conditions reflected in precipitation<sup>18,22,23</sup>. Leaf size, leaf dark respiration rate, specific leaf area (SLA), leaf N and P concentration, seed size and wood density, all show broad-scale correlations with climate or soil<sup>22,24–27</sup>. It has also been reported that many of these traits show latitudinal patterns<sup>24–27</sup>. Generalizing such insights is, however, not trivial, as soil properties partly mirror climate gradients, as a consequence of long-term soil formation through weathering, leaching and accumulation of organic matter—processes related to temperature and precipitation<sup>28</sup>; however,

A full list of affiliations appears at the end of the paper.



**Fig. 1 | Previously identified global axes of variation in size and economics traits hold for an extended trait set ( $n = 36,197$  species per ecoregion median).** The set of 17 investigated traits (Supplementary Table 5) can be primarily divided into size and economics traits, which load differently onto the two PC axes describing their global distribution. **a**, Heatmap of covariation. Trait correlations are indicated using absolute Pearson correlation coefficients, with green shades indicating high absolute correlation and yellow shades indicating low absolute correlation. On the left, the distance tree of traits derived from hierarchical clustering is illustrated. Three resulting groups are: (1) size-related traits (blue) consisting of conduit density (conduit dens), leaf area, leaf fresh mass (leaf f mass), plant height (height), seed mass, dispersal unit length (dispersal len) and seed length (seed len); (2) economics traits (red) comprising SLA, leaf N content per area (leaf N area), leaf N, P and C concentrations, leaf N/P ratio (leaf N:P) and stem specific density (stem dens); and (3) a third (yellow) consisting of the number of seeds per reproduction unit (seeds U), leaf  $\delta^{15}N$  (leaf d15N) and vessel element length (vessel len). **b**, The first two PCs of the PCA. Arrow tips refer to the loading of the traits (Supplementary Fig. 6). Contour lines delineate the colour scale that corresponds to the kernel density of species (dense, red to sparse, light yellow; 20%, 50%, 95% and 99% of all species). PC1 explains 33% of trait variation and PC2 15% (Supplementary Fig. 5).

climate-independent features reflecting geology and surface morphology also contribute to soil fertility<sup>28</sup>. Soil may furthermore buffer climate stresses; for example, by alleviating water deficit in periods of low precipitation<sup>29</sup>.

Combining the insights suggests that the global spectrum of plant traits reveals two internally correlated orthogonal groups and that many plant traits are individually linked to environmental gradients, we expect that both trait groups should closely follow gradients of climate and soil properties. Here, we investigate to what extent the major dimensions underpinning the global spectrum of plant form and function can be attributed to global gradients of climate and soil conditions; and to what extent these factors can jointly or independently explain the global spectrum of form and function.

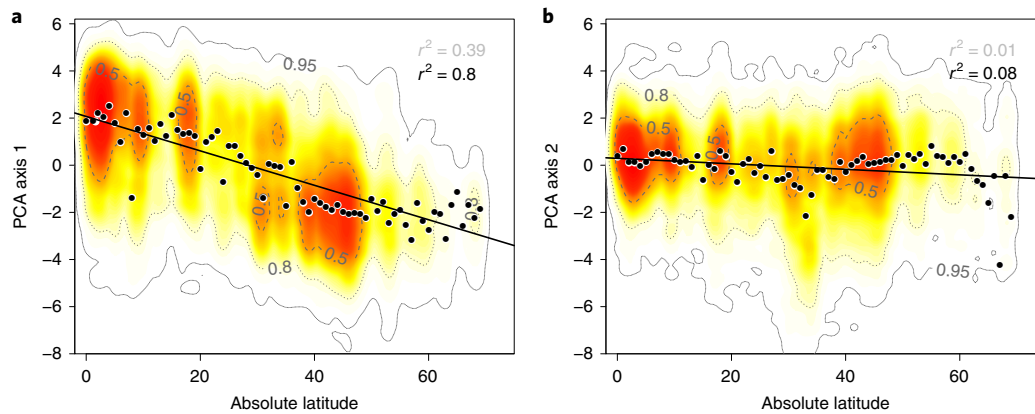
We compiled and analysed a dataset of 17 functional traits with a sufficient number of records in the TRY database<sup>9</sup> to characterize the main ecoregions of the world<sup>30</sup>, that is, environmentally homogeneous areas with distinct biota (Extended Data Fig. 1). The dataset is based on 225,206 georeferenced observations comprising records of 20,655 species. The trait data were complemented with 21 climate variables and 107 soil variables (Methods; Supplementary Tables 1 and 2). Trait–environment relationships were analysed for species medians aggregated to ecoregions using ridge regression<sup>31</sup>, a robust method (Supplementary Figs. 1–3) suitable to deal with high-dimensional, unbalanced and collinear predictors in combination with hierarchical partitioning<sup>32</sup> (Methods).

## Results

Our main analysis is based on median trait values of plant species per ecoregion. The rationale is that species presence indicates how the trait space can be realized in a given environment. Spatial

aggregation is a suitable means to increase the detectability of global trait patterns (Supplementary Fig. 3), as described in earlier studies, where traits have been binned by temperature classes<sup>33</sup> or for different altitudinal ranges<sup>22</sup>. Extreme outliers, for instance towering trees such as the Californian Sequoia (*Sequoiadendron sempervirens*), may still exist far away from the equator, where precipitation is sufficiently high<sup>20</sup> but their influence is outweighed in our approach by an increasing fraction of small-statured herbaceous species from tropical to temperate and boreal regions.

**Orthogonal axes and trait clusters.** To understand whether the axes of variation identified for the grouping of six traits<sup>10</sup> also hold for the extended set of 17 traits, we cluster their trait–trait correlations (Fig. 1a and Supplementary Fig. 4) and further represent these relations on the basis of their principal components (PCA; Methods). This analysis supports the clear distinction of size versus economics traits identified by Díaz and colleagues<sup>10</sup>. The group of size traits contains two subclusters. The first includes height and seed size traits: plant height (height), seed mass, seed length and dispersal unit length (dispersal length). The second subset contains traits that are linked through plant hydraulic scaling relationships<sup>34</sup> and contrasts high conduit density (that is, number of conduits per sapwood cross-sectional area) with high leaf area and leaf fresh mass (leaf f mass). Economics traits represent dry mass and nutrient investments in plant tissues, and the rate and duration of returns on those investments<sup>11</sup>. They are represented by leaf nitrogen content per leaf area (leaf N area), leaf nitrogen (leaf N), phosphorus (leaf P) and carbon (leaf C) content per dry mass, leaf N to P ratio (leaf N:P) and SLA. Stem specific density (stem density) takes an intermediate position (Fig. 1b) but more closely clusters



**Fig. 2 | Size traits, not economics traits vary with latitude: the PC1 of the PCA on 17 plant traits shows a clear latitudinal gradient while PC2 does not ( $n = 36,197$  species per ecoregion median).** Contour lines delineate the colour scale that corresponds to the kernel density of species (dense, red to sparse, light yellow; 5%, 95%, 99% quantiles). Mean estimates aggregated at  $1^\circ$  absolute latitude are indicated as black dots. The line refers to a linear model (ordinary least squares). **a**, PC1 representing mainly size traits (conduit density, leaf area, leaf fresh mass, plant height, seed mass, dispersal unit length, seed length) regressed against absolute latitude. Linear model:  $r^2 = 0.38$  without bins;  $r^2 = 0.84$  aggregated at  $1^\circ$  absolute latitude. **b**, PC2 representing mainly economics traits (leaf N, leaf N per area, leaf P, leaf N:P ratio, SLA, leaf C, stem density) regressed against absolute latitude. Linear model:  $r^2 = 0.01$  without bins;  $r^2 = 0.08$  aggregated at  $1^\circ$  absolute latitude.

with this set of economics traits (Fig. 1a), suggesting a syndrome of traits promoting slow to fast nutrient and carbon processing at the whole-plant level<sup>35–37</sup>. Furthermore, we identify a third group of traits that appear to be only weakly correlated with any other trait. This third group contains seed number per reproduction unit (seeds U), leaf  $\delta^{15}\text{N}$  (leaf  $\delta^{15}\text{N}$ ) and vessel element length (vessel length). The first two principal components (PC) of the PCA on the trait data represent 48% of the overall variation (Supplementary Fig. 5). PC1 is determined by size traits and accounts for 33% of the variance; PC2 is determined by economics traits and accounts for 15% of the variance (Fig. 1b and Supplementary Fig. 6). These two main axes remain clearly identifiable when the analysis is conducted separately for woody and non-woody species (Supplementary Figs. 7 and 8). The remaining PCs each account for less than 10% of variance (PC3 = 9.36%). In the following, we focus on the two groups of size and economics traits (Supplementary Fig. 5).

**Latitudinal trait variation.** As an investigation of broad-scale gradients among size and economics traits, we analyse latitudinal gradients of the first (PC1) and second (PC2) principal components. PC1—representing primarily size-related traits—shows a strong linear latitudinal signal (on the basis of species:  $r^2(\text{PC1}) = 0.37$ , at the ecoregion level  $r^2(\text{PC1}_{\text{aggregated}}) = 0.84$ ; Fig. 2a). By contrast, the axis representing primarily economic traits, PC2, shows little response to latitude (on the basis of species:  $r^2(\text{PC2}) = 0.01$ , at the ecoregion level  $r^2(\text{PC2}_{\text{aggregated}}) = 0.08$ ; Fig. 2b, for woody non-woody species Supplementary Fig. 9), except for a dip at  $35^\circ$  and declining sharply at  $60^\circ$  where the species density also drops (but see Supplementary Fig. 10 for comparison to an independent dataset from arctic latitudes which shows the same pattern). Latitudinal gradients are known to be strongly related to climate, due to the distribution of solar energy and general atmospheric circulation patterns. Therefore, we propose that those climate (and soil) aspects that co-vary with latitude consistently determine size traits, while they have little effect on economics traits, which are more strongly affected by latitude-independent soil (and climate) effects (Supplementary Fig. 11).

**Climate and soil: joint and independent effects.** The differences in latitudinal relationships between the two PC axes support the hypothesis that different environmental factors should drive

variation within the separate groups of size versus economics traits. We assess the joint and independent effects of climate and soil on trait variability (ridge regression, RR; Table 1 and Fig. 3). Overall, size traits are better explained (RR;  $r^2 = 0.55$ ; maximum  $r^2 = 0.78$  for conduit density; Table 1) than are economics traits (RR;  $r^2 = 0.40$ ; maximum  $r^2 = 0.55$  for leaf N:P ratio; Table 1). We find a substantial joint effect of climate and soil variables—in every case larger than either unique effect—which reflects strong interactions between specific climate and soil predictors (RR with hierarchical partitioning (HP); Fig. 3b and Supplementary Fig. 12). However, we also observe independent effects of climate and soil (RR with HP; Fig. 3 and Table 1). The independent climate effects are observed across traits but size traits tend to be better explained by the independent climate effects than are economics traits. In contrast, independent soil predictors are relevant for all economics traits but not size traits (apart from a small contribution to leaf area). We interpret these results as evidence for the importance of both joint and independent effects of climate and soil variables for whole-plant strategies<sup>2,37,38</sup> which we show here at the global scale along with a dichotomous tendency of a stronger imprint of climate factors on size traits and of soil conditions on economics traits (Fig. 3, Supplementary Figs. 13–38 and Supplementary Table 3). We propose that the dominance of joint effects implies that interactions between soil and climate properties are of primary importance in plant trait ecology; as opposed to trait syndromes being defined by single environmental variables in isolation.

We next ask how the climate and soil datasets are interdependent and which predictors add the most relevant information. For this purpose, we related all traits to environmental variables in a redundancy analysis (RDA; Methods; Fig. 4). The RDA again identifies two main axes of size and economics traits (Fig. 4a), which are now shown together with the environmental variables that co-vary linearly with those traits (Fig. 4b and Supplementary Fig. 39). The first RDA axis corresponds to size traits (Fig. 4a) and represents an axis of water and energy (for example, precipitation, vapour pressure and temperature; Fig. 4b). Two attributes of soil texture important for water retention—the fraction of gravel and clay—also vary along this axis. The second RDA axis corresponds to economics traits (Fig. 4a) co-varying with an axis of soil variables generally associated with soil fertility (that is, soil texture (silt versus sand), water holding capacity, carbon concentration and stocks), as well as the climate variable mean solar radiation (Fig. 4b).

**Table 1 | Showing for each trait the variance explained ( $r^2$ ) by ridge regression models for 220 ecoregions and the independent effects for climate and soil listed from hierarchical partitioning that, respectively, add up with the joint effect to the variance explained by climate or soil**

Trait <sup>a</sup>	Group	Explained variance <sup>b</sup> by soil and climate ( $r^2$ )	Soil (independent effect <sup>b</sup> ) ( $r^2$ )	Climate (independent effect <sup>b</sup> ) ( $r^2$ )	Joint effect <sup>b</sup> ( $r^2$ )
		Ridge regression model	Hierarchical partitioning	Hierarchical partitioning	Hierarchical partitioning
Seed length	Size	0.4	-0.01	0.08	0.33
Dispersal length	Size	0.26	-0.01	0.03	0.24
Seed mass	Size	0.57	-0.01	0.09	0.49
Height	Size	0.52	0.01	0.1	0.41
Leaf f mass	Size	0.72	0	0.15	0.56
Leaf area	Size	0.63	0.03	0.13	0.47
Conduit density	Size	0.77	-0.01	0.22	0.56
Stem density	Economics	0.41	0.03	0.02	0.36
Leaf C	Economics	0.29	0.08	0.05	0.17
Leaf N:P	Economics	0.55	0.06	0.12	0.38
Leaf P	Economics	0.45	0.15	0.05	0.25
Leaf N area	Economics	0.39	0.03	0.02	0.33
SLA	Economics	0.41	0.09	0.13	0.19
Leaf N	Economics	0.26	0.06	0.16	0.05
Vessel length	Other	0.4	0	-0.03	0.42
Leaf d15N	Other	0.51	0.05	0.1	0.35
Seeds U	Other	0.1	-0.02	0.02	0.1

<sup>a</sup>For full versions of traits, see main text. Mean values; minimum and maximum values from different cross-validation runs in Supplementary Table 8. Negative values indicate a reduction of explained variance when respective variables are added to the ridge regression model.

## Discussion

This study shows that the proposed global spectrum of plant form and function fits well to a substantially extended trait space compared to the original study<sup>10</sup>, with seven traits that capture the whole-plant size spectrum and seven traits that capture the leaf economic spectrum and only three traits that do not fall along these dimensions (Fig. 1b). One explanation could be that the varying fraction of woody and non-woody species would drive these patterns. However, we showed that these two main trait groups remain clearly identifiable when the analysis is conducted separately, yet with fewer samples, for woody and non-woody species (Supplementary Fig. 8).

However, we cannot discard the possibility that additional traits may add relevant axes of trait variation. For example, our study does not include carbon fixation rates<sup>39</sup> or fire adaptation traits<sup>40</sup>, nor does it include any root traits—representing an essential gap to be filled at the global scale<sup>41</sup>. The respective data are too scarce to yet be integrated with global datasets. If such data were available they would have the potential to fundamentally change our perception of global plant form and function, and their relation to ecosystem functioning.

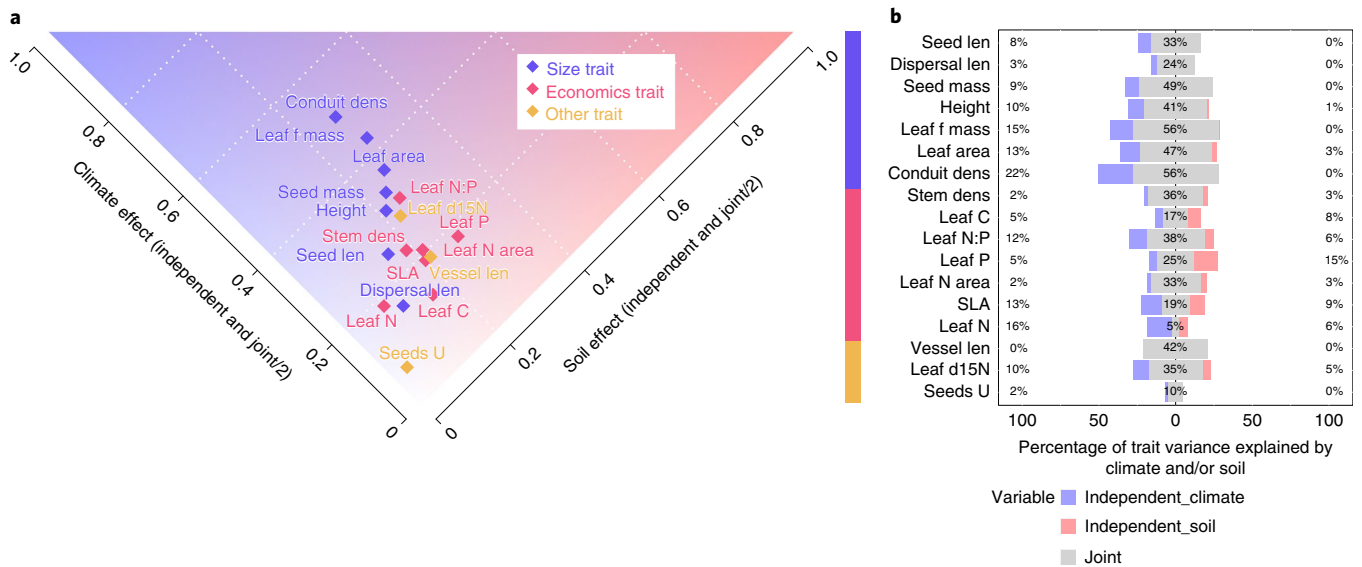
Variation in size traits, represented by PC1 in Fig. 1b, shows a clear latitudinal gradient (Fig. 2b). In contrast, variation in economics traits (represented by PC2) does not show a latitudinal trend. Only a dip is apparent at around 35° (absolute), in addition to a decrease at high latitudes above 60° (absolute) where available data become increasingly limited. However, comparison to a recent arctic dataset indicates that this decrease in variation at high latitudes reflects available observations (see Supplementary Fig. 10 for a comparison to independent data). These patterns might represent a response to nutrient limitation and drought<sup>42,43</sup> in water-scarce and nutrient-scarce deserts and Mediterranean regions (Supplementary

Fig. 40) or boreal and arctic areas characterized by short growing periods slowing down mineralization. The dip at ~35° indeed can be related to low water availability (Supplementary Fig. 41). At high latitudes, cold winters and short growing seasons constrain plant height<sup>13</sup> and require on average more conservative nutrient-use strategies (like evergreen leaves) and protection against frost damage than the global mean, despite the high functional diversity in economics traits observed at these latitudes<sup>13</sup>. Additional datasets may shed more light on specific conditions, for example see Bjorkman et al.<sup>19</sup>. Future studies should quantify how individual stressors, for example radiative stress or water stress, relate to global patterns of trait variation.

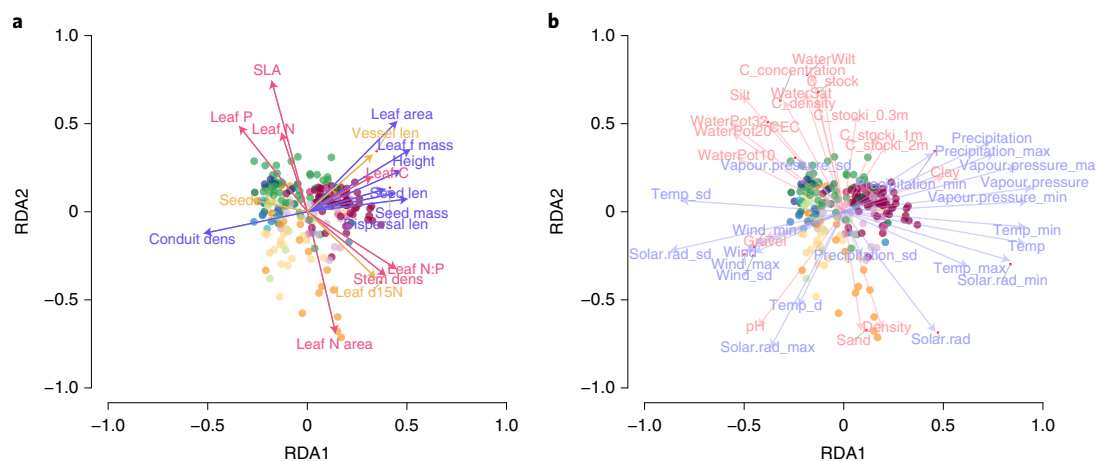
The climate and soil factors used in this analysis explain up to 77% of observed trait variation—a high fraction given that trait variation is widely known to be determined also by other factors such as biotic interactions (for example, soil biota) and anthropogenic effects or disturbances and local effects such as those of microclimate<sup>12,44–46</sup>. Recent findings on how different trait groups vary with the environment indicate that size and economics traits vary differently<sup>13</sup> and in particular respond differently to climate and soil<sup>19</sup>.

Our analyses reveal a dominant joint effect of climate and soil drivers on trait variation—as already suggested by a number of earlier studies<sup>18,19,22</sup> but not yet quantified globally.

The orthogonality of the two main dimensions of plant trait variation suggests that different aspects of climate and soil variables are relevant to explain plant trait patterns at the global scale (Supplementary Figs. 11–39). While latitude-related variables (mainly climate) explain size traits, variables that share less explanatory power with latitude (mainly soil) explain economics traits (Supplementary Table 4 and Supplementary Fig. 11). The RDA presented in Fig. 4 (Supplementary Fig. 39) provides some insight



**Fig. 3 | Climate and soil variables explain up to 77% of variance in size and economics traits.** Hierarchical partitioning<sup>32</sup> identifies the contribution of climate and soil variables to explain each trait ( $n=220$ , ecoregional median trait: blue, size; red, economics; yellow, other). The joint effect is the fraction explained by both climate and soil together, and is split equally among them. The independent effect is the fraction of  $r^2$  explained exclusively by either soil or climate variables. a, Tilted x-y plot of the soil versus climate variables to explain a trait. The axes show the sum of the respective joint and independent effect (hierarchical partitioning). The colours reflect the strength of: the independent effect of climate plus its share of the joint effect ( $r^2$ ; purple); and soils' independent effect plus its share of the joint effect (peach). The sum of both axes equals the total  $r^2$  explained by climate and soil; in cases where soil showed a negative independent effect only the climate-independent effect is shown (and vice versa but see Table 1). b, Percentage variation explained by climate (purple, percentages on the left), soil (peach, percentages on the right) and jointly (grey, percentages in the middle) for trait groups—size, economics and other. Total bar length = total  $r^2$  explained by climate and soil; in cases where soil showed a negative independent effect only the climate-independent effect is shown (and vice versa but see Table 1). For leaf area, climate and soil jointly explain 47%, the independent climate effect explains an additional 13% of the variance, while soil explains 3%, totalling 63% of variance explained. For trait abbreviations see Fig. 1.



**Fig. 4 | RDA of traits reveals the relationships of climate and soil factors associated with trait distributions ( $n=220$ , ecoregion median, only topsoil layer variables included; variance explained: RDA1 = 63%, RDA2 = 18%).** a, b, The output of the RDA is split into two plots: traits (a), where arrows are coloured according to trait groups (blue, size traits; red, economics traits; yellow, other traits; arrow length and point positions scaled to fit the plot); and environmental factors (b), where arrows are coloured according to predictor group (Climate, blue and soil, red variables; arrow length and point positions scaled to fit the plot). In a and b, points represent ecoregions and are coloured according to biome (red, tropics; green, temperate; yellow, desert; orange, Mediterranean; dark blue, tundra). Climate variable abbreviations are composed of the variable (average if not stated differently) and a suffix. Variables are: Solar.rad, solar radiation; Vapour.pressure, vapour pressure; Wind, average wind speed; Temp, temperature; Precipitation, precipitation. Suffixes are: no suffix, mean of respective variable; d, diurnal range; min, annual minimum of the respective variable; sd, seasonality of respective variable. Soil variable abbreviations (all topsoil) are: Density, soil density ( $\text{kg}/\text{m}^3$ ); pH, pH value; Sand, sand fraction (vol%); Silt, silt fraction (vol%); Clay, clay fraction (vol%); C\_concentration, organic carbon concentration; C\_density, organic carbon density; CEC, cation exchange capacity; C\_stock, soil organic carbon stock at depth 0.00 m; C\_stocki, organic carbon stock for depth intervals—0.3m (0–0.30 m), 1m (0–1 m), 2m (0–2 m); WaterWilt, available soil water capacity (volumetric fraction) until wilting point; WaterSat, saturated water content; WaterPot, available soil water capacity for moisture potentials—10 (–10 kPa;pF 2.0), 20 (–20 kPa;pF 2.3), 32 (–31.6 kPa;pF 2.5). For trait abbreviations see Fig. 1.

on the nature of these climate–soil interactions. The first RDA axis, which describes variation in size traits, resembles a latitudinal gradient. On one extreme end, ample water supply from high and frequent precipitation, abundant water vapour and constant rates of high solar radiation meet the fundamental requirements of plant physiology—water, sunlight and warm temperatures. Additionally, these conditions promote weathering of soil minerals but also microbial activity, contributing to fast turnover rates of organic matter supporting nutrient provisioning<sup>28,47</sup>; in brief, they represent conditions that allow plants to grow fast and tall in the race for light. Large vessels supporting large leaves promote high rates of water transport and thus growth, which is only possible because of the small risk of embolism under these benign water conditions<sup>43</sup>. The high carbon gains can be invested in large fruits and seeds (seed mass, seed length and dispersal unit length). Further along this gradient, the above-mentioned plant requirements become limited: water supply and temperatures are reduced and slow metabolic rates aboveground and belowground. In ecoregions of the boreal and desert biomes, conduit diameter is constrained by the risk of cavitation during freeze–thaw cycles<sup>43</sup> and water scarcity, amplified by little water holding capacity of gravel-rich soils. Our analysis thus indicates that size traits appear to be related to a latitudinal gradient of climatic favorability for plant growth determined by water and light availability.

Important correlates of water and nutrient availability are associated with the second RDA axis, describing variation in economics traits. Traits associated with an acquisitive strategy are related to indicators of soil fertility, most importantly silt and organic matter concentration as well as pH (refs. <sup>18,28</sup>). Soil pH is intermediate between the two axes, as might be expected given that pH reflects both broad-scale climate variation (especially aridity<sup>47</sup>) and a variety of processes related to nutrient availability and soil microbial communities<sup>18,48–50</sup>. Silt forms the substrate of our most fertile soils as its structure is able to retain water against gravitation (unlike sand) but renders it accessible to plants under drought conditions<sup>28,51</sup> (unlike clay). The high fertility is associated with a high concentration of organic matter, which has a high cation exchange capacity especially under high pH (ref. <sup>47</sup>). On the opposite end of the gradient, sandy soils require adaptations to both water and nutrient limitation. The trait configuration at the conservative end of the economics traits (low SLA, high tissue density and high organ longevity) represents an adaptation to both<sup>11,37</sup>. Various processes exist that lead to variation in the soil characteristics underlying the second RDA axis independent of latitude<sup>18</sup>—for example, sandstone as a geological substrate giving rise to sandy soils exists from the tropics to the arctic<sup>28,51</sup>. However, different climate variables related to solar radiation, temperature and precipitation, which influence long- and short-term soil development processes directly and indirectly via soil biology<sup>28,51,52</sup>, are related to this axis. Variation in economic traits is most probably the evolutionary response to exploiting this partly climate-independent edaphic niche axis.

Size traits are on average explained better than economics traits by the environmental variables considered in this study. The lower fraction of explained variance for economics traits could have several causes. Firstly, data on soil factors that are likely to be very important, such as soil nitrogen and phosphorus availability<sup>18,23</sup>, are not yet available at a global scale. Secondly, economics traits show relatively more within-site variation than across-site variation in comparison to size traits (Supplementary Fig. 42), probably because economics traits vary more than size traits within one plant; for example, leaf N per area and SLA vary with age and light availability<sup>53</sup>. Thirdly, soil heterogeneity within ecoregions—both abiotic and biotic—may weaken the relationship between economics traits and environmental variables<sup>12,54,55</sup>. Reasons for small-scale soil variation are, for example, topography, soil age and thus fertility<sup>56</sup> but also abundance of microbial communities and mycorrhiza

that interact with climate, pH, soil properties and also plant traits<sup>50</sup>. Trait–environment relationships due to smaller scale variation require well-resolved soil data. However, we note that soil physics and chemistry explain a large portion of variance along the trait PC axis three (which itself explains slightly less than 10% of variance in the PCA (9.36%); Supplementary Figs. 5, 6 and 38). We expect that with improved soil datasets and a higher resolution, the joint control of climate and soil on trait variation will probably appear even stronger and more evenly distributed between the two groups of driver variables.

Our analysis can serve as reference for model developments that increasingly consider plant functional traits as part of vegetation dynamics under climate change<sup>44</sup>.

Individual plants and their trait syndromes are considered to be viable only within specific environmental conditions<sup>2</sup>. Therefore trait–environment relationships should be scale-independent. However, different plant strategies can be successful under given environmental conditions, which in addition are often confounded by small-scale variation. In analyses to date, trait–environment relationships become more apparent for aggregations higher than the community scale<sup>12</sup>, where most of the small-scale variation is averaged out. In addition the difference between potential and actual vegetation is suggested to explain some of this gap<sup>13</sup>. Dynamic global vegetation models predict individual plant processes well but fail to produce reliable forecasts with a changing environment<sup>44</sup>. Deciphering at which spatial and temporal scale, or conditions, actual vegetation is representative of potential vegetation may advance our understanding of community assembly and necessary model complexity.

Trait–environment correlations identified in our study should not be confounded with causality. Yet, the ubiquitous importance of climate variables for explaining current differences in trait expression at ecoregion scale, suggests that trait shifts will occur with climate change. Trait shifts are constrained by available trait combinations in addition to other constraints such as species dispersal. For example, our results indicate that plant size increases with temperature so long as sufficient water is available (Fig. 4 and Supplementary Figs. 19, 20 and 21), in line with the finding that species become larger and large species are more prevalent at warmer and wetter sites in the tundra<sup>19</sup>. Global change is also reflected by soil degradation. Changes in soil parameters can be considered to also correspond with trait shifts, especially for economics traits. Human-induced soil degradation has many facets: often fertile topsoil is lost or toxic substances accumulate; rooting is impeded and altered by artificial fertilizers; while soil formation takes millenia<sup>57</sup>. The trait shifts may thus be similarly complex and depend on the extent and type of soil degradation. For example, in areas of wind and water erosion, species that tolerate lower nutrient availability may be more successful and this may be reflected in lower leaf nutrient contents (Fig. 4 and Supplementary Fig. 30). The fertilization of nutrient-poor grasslands, for example resulting from agricultural run-off, may shift these areas from more conservative to more competitive species with higher leaf nutrient contents.

Plants as a whole need to balance both size and economics traits. To sustain human livelihoods, it may be important to understand the local expression of trait shifts and their global consequences for biodiversity when viable trait combinations change.

In conclusion, the insights extracted here advance our understanding of broad-scale plant functional patterns. In particular, we highlight the combination of independent and particularly joint effects of climate and soil on trait variation, an interaction that has to date been neglected because few studies include both in a single analysis, at the global scale as we have done here. In doing so, we identify an important gap in knowledge: what is the nature of climate–soil interactions that drive whole-plant trait variation and what distinguishes the majority of climate and soil factors having

joint effects on plant traits from those with independent effects? These are the sorts of questions that require answers to increase our capacity to predict plant functional diversity in a changing environment. Such predictive power would contribute to a sound basis for assessing long-term feedbacks between global environmental change and the terrestrial biosphere, helping to constrain parameters of global coupled climate–vegetation models. Humans are currently modifying both climatic and edaphic conditions at the global scale. Climate envelope models used to predict vegetation shifts must be complemented by drivers related to large-scale anthropogenic alterations of soil conditions resulting, for example, from land-use change, atmospheric nitrogen deposition, fertilization, liming and salinization. Our global analysis provides an essential context for finer-scale studies to directly tackle questions of biological processes and mechanisms at landscape and community scales.

## Methods

We extracted data on 17 plant functional traits from a gap-filled version of TRY database<sup>9</sup> (Supplementary Table 5; [www.try-db.org](http://www.try-db.org), accession date July 2017, request no. 3282) which includes published literature<sup>11,58–101,101–310</sup>. Quality control was conducted according to the published protocol of TRY.<sup>9, 311</sup> Traits with  $z$ -score > 4 were excluded and those with  $z$ -score > 3 were checked for plausibility. Before this, missing data were imputed using a Bayesian hierarchical probabilistic matrix factorization (BHPMF) algorithm<sup>8,312</sup> for an extended dataset, derived from TRY (Supplementary Table 6). Imputation was done to be able to include the maximum number of species in our analyses. Then the 17 traits were selected among the traits with the largest total number of entries. The data were attributed to ecoregions<sup>30</sup> (Supplementary Table 7 and Extended Data Fig. 1) and aggregated to species median values. The imputed values were calculated using the whole dataset at the individual record level. BHPMF calculates the imputations from 1,000 Gibbs sampler (Markov chain Monte Carlo) imputations by taking the mean of every twentieth imputation of these 1,000 ‘versions’, after the first 200 are removed. Then the species median was calculated at the ecoregion level. We excluded observations that were not georeferenced because we could not attribute them to ecoregions. According to TRY regulations, data from experimental treatments (for example, fertilization) or from botanical gardens were also excluded. In total, we included 225,206 observations from 20,655 global unique species (36,197 unique species to ecoregion combinations). Throughout this study we used one of two aggregation levels: either species median per ecoregion (ER)<sup>30</sup> resulting in unique species values per ecoregion (termed A1,  $n = 36,197$  with  $n = 20,655$  globally unique species) or the aggregation to median ecoregions calculated from median species per ecoregion (termed A2,  $n = 220$ ). R was used for all analyses and figures<sup>313</sup>.

**Hierarchical probabilistic matrix factorization.** *Description.* BHPMF decomposes or factorizes probabilistically a matrix (probabilistic matrix factorization, PMF<sup>314</sup>) using information contained within different hierarchical levels (here, taxonomy) within a Bayesian framework<sup>8</sup>. The underlying premise of BHPMF is to gap-fill (or more accurately, to predict) traits of an individual plant using trait–trait correlations as well as intraspecific and interspecific trait variability.<sup>8</sup> Using a Gibbs sampler (a Markov Chain Monte Carlo algorithm), BHPMF also provides a prediction confidence in the form of standard deviations which is a per-value estimate of uncertainty in trait predictions<sup>8</sup>. BHPMF can fill gaps if there is at least one value per row (species) and column (trait).

**Implementation.** The largest possible dataset was retrieved at the time when study was conducted, including 172 traits of 652,957 individuals (Supplementary Table 6). For data preparation before BHPMF, all individual-level trait data were firstly log-transformed and secondly normalized via zlog transformation ( $z = \frac{x-\bar{x}}{s.d.}$ ). Log transformation was chosen to achieve a closer-to-normal distribution of values per trait<sup>311, 313</sup>. This transformation is considered necessary because a given difference for small trait values (absolute value) is likely to be physiologically more relevant than the same difference (absolute value) for large trait values.

BHPMF internally splits the datasets randomly into a training dataset (80%), a test dataset (10%) and a validation dataset (10%).

The training dataset is used during training of latent vectors, while the test data are tested against to improve the latent vectors, and finally the validation dataset serves as the basis for calculation of the root mean square error (RMSE) and stopping the optimization of latent vectors within BHPMF<sup>8</sup>. The validation dataset ensures ongoing amelioration of the model performance during the training process and stops the process after five consecutive iterations with stable RMSE. The test dataset is used only on the lowest taxonomic level (individuals  $\times$  traits). BHPMF was run with a maximum of 1,000 iterations, whereas the first 200 were discarded during the ‘burn-in’ phase, as predictions of these iterations are likely to be influenced by the initialization of BHPMF rather than being

part of the probability density distribution to be sampled by BHPMF. To avoid autocorrelation, only every twentieth iteration was used to calculate the resulting trait values. The mean of these predictions result in the final trait values used as the output. Compared to the original data, the imputed values are similar in terms of trait–trait correlation, according to the Procrustes test provided in ref. <sup>10</sup>.

**Trait clustering.** To define groups of correlated traits, we clustered species’ traits (species median per ecoregion, A1) on the basis of absolute pairwise Pearson correlation coefficients using a hierarchical clustering algorithm (‘complete linkage clustering’). Variables were transformed into distances previous to the clustering. Hierarchical clustering then attributes variables (here, traits) to groups of least distance and highest similarity. Traits were more like each other if they exhibited similar correlation patterns with all other traits. We set a distance between traits of 1 as the threshold for defining trait clusters. We used the R package ‘stats’ function ‘hclust’ included in R (ref. <sup>315</sup>).

**PCA.** Values for all 17 traits (unique species per ecoregion, A1) were natural log-transformed and then projected onto components (PCA). We used the R package FactoMineR<sup>316</sup> that scales data internally. After the PCA (A1), we extracted the variance explained (Fig. 1b and Supplementary Fig. 5) and respective loadings for the first five principle components (Fig. 1b and Supplementary Fig. 6), which are significant according to the number of axes to keep estimated using a sequential Bonferroni procedure (R package ade4 (refs. <sup>317–321</sup>), function testdim). For the analysis (ridge regression package ‘glmnet’<sup>322,323</sup>) for Fig. 3, all environmental variables (climate and soil) were first reduced with this package to 20 PCs.

**Environmental variables.** To represent climate conditions we used 21 variables derived from WorldClim at a resolution of 1 km for temperature, precipitation, vapour pressure, solar radiation and wind (Supplementary Table 1). To characterize soil conditions we used 107 variables derived from the ISRIC data product ‘SoilGrids’<sup>324–326</sup> (<https://soilgrids.org/> through ISRIC—WDC Soils). ‘SoilGrids’ provides global predictions of 17 fundamentally different soil characteristics (some for seven depths, that is 0, 5, 15, 30, 60, 100, 200 cm; Supplementary Table 2) at a resolution of 1 km. SoilGrids are publicly accessible environmental data (Creative Commons Attribution 4.0 International), with a collection of georeferenced soil profile data and are managed in World Soil Information Service<sup>324</sup>.

**Aggregation of traits and environmental variables to ecoregions.** To determine trait–environment relationships, we aggregated trait as well as environmental data to regions, here ecoregions<sup>30</sup> (ecoregion aggregation A2, see also above; Supplementary Table 7). Ecoregions are environmentally homogeneous areas, nested within biogeographic realms (defined by refs. <sup>327,328</sup>) and biomes (modified after refs. <sup>329,330</sup> but see ref. <sup>30</sup>). As a first estimate, ecoregions are distinct biotas<sup>328,331</sup> defined by the physiognomy of the prevailing climatic climax vegetation<sup>30</sup>. These areas of distinct biotas, areas of relatively uniform flora or fauna, are next subset into provinces with substantial differences of vegetation on the basis of a selection of plants and animals, maps and expert knowledge<sup>30,331</sup>. At global scale Olson et al.<sup>30</sup> defined 867 ecoregions. Ecoregions were chosen as the scale of aggregation for their high signal-to-noise ratio and the ability to correct for sampling bias. While the grid scale has higher spatial resolution, it lacks estimates of species richness (equivalent of Kier species richness<sup>174</sup>) and is not as well explained by the climate and soil (Supplementary Fig. 3) and distribution of grids is globally uneven (Extended Data Fig. 1) in comparison to ecoregions. The global sampling distribution is recognized to show a bias towards Europe<sup>8</sup>, which is even more pronounced in the lower level data (grid scale) than in the more aggregated one (Extended Data Fig. 1). Our method accounts for this oversampling and reproduces a stable pattern, even when species in oversampled ecoregions are deleted (Supplementary Fig. 2).

For each of the 867 ecoregions, we calculated the median ecoregion aggregate trait value from the median trait values of all species identified in each region. For further analyses, we only used regions with >20 species and a representation of >1% of the estimated species richness of the ecoregion<sup>30</sup>. Preliminary tests with different selection criteria (for example, number of species and inclusion or exclusion of 1% of species richness estimate by Kier et al.<sup>174</sup>) showed that lower numbers of species per ecoregion result in weaker explained variance, while stricter rules reduced the number of ecoregions. These selection criteria serve as a quality control because ecoregions with poor representation of species richness are excluded, as we can expect the regression to the mean to be stronger with more species data. A total of 220 ecoregions met these criteria and were included in the analysis. On average, these ecoregion-level trait values were based on 164 species-level trait medians (with a maximum of 1,245 species in Tapajós-Xingu moist forests; Supplementary Table 7). In total, we aggregated 36,197 median species trait values to ecoregions. These ecoregions cover the global latitudinal gradient (Fig. 2) as well as a substantial fraction of the geographic space (Extended Data Fig. 1). To aggregate environmental variables to ecoregions, we associated each trait observation with its corresponding values of climate and soil variables. Then, we averaged over all values within one ecoregion. Thus, the selected environmental variables represent averages that are weighted by the number and locations of trait observations within ecoregions.

**Model building.** Ecoregion trait values (natural log-transformed, A2) were related to all environmental variables using ridge regression<sup>31</sup>, which is a well-established linear regression method that is suitable to deal with a large number of collinear predictors and uneven numbers of predictors for climate and soil. We used the R package 'glmnet'<sup>32,33</sup>. From aggregating trait values to ecoregion medians we obtain 220 samples for each trait. The environmental predictors of climate and soil were reduced to 20 each by means of a PCA. In addition, the environmental predictors show relatively high collinearity, thus duplicated information. Ridge regression addresses collinearity among predictors by shrinking (regularizing) regression coefficients according to a penalty on the L2 norm of the vector of regression coefficients. The regularization parameter lambda was obtained via tenfold cross validation. The variance explained was derived from an iterative holdout set (tenfold cross validation), that is, prediction of 90% of randomly sampled ecoregions for inclusion in model building and then predicting the remaining 10% of the data to evaluate the quality of the models. The final model predicts the remaining 10% of unused ecoregions. This prediction-loop was repeated until all ER trait values are predicted, that is, resulting in different linear models. Repeated  $r^2$ : the  $r^2$  is the squared correlation of predicted versus original ER trait values. This procedure was repeated 50 times and the explained variances ( $r^2$ ) mean, minimum and maximum were calculated. For the purpose of defining how much of the explained variance is due to independent and joint information in the data streams, we used hierarchical partitioning<sup>32</sup>. Model outputs ( $r^2$ ) of all repetitions ( $n=50$ , if not indicated differently) were used as input (ridge regression, partial least squares (PLS) with and without PCA, random forest).

**Redundancy analysis.** To relate trait–trait covariation to trait–environment covariation, we performed a redundancy analysis (R package 'vegan'). Ecoregion-aggregated traits (A2) were normalized and natural log-transformed. Scaled climate and soil variables were used as predictors. To decrease the factor that quantifies collinearity (variance inflation factor, vif), only the topsoil layer was selected (Fig. 4). For Supplementary Fig. 39, additional model tuning based on vif, with the exclusion of two variables with vif > 20, led to a model with vif < 10, which can be considered low cocorrelation.

**Reporting Summary.** Further information on research design is available in the Nature Research Reporting Summary linked to this article.

## Data availability

Plant trait data were accessed from the TRY database (<https://try-db.org>, request no. 3282, date accessed July 2017, see also Extended Data Fig. 1). All TRY data required to reproduce this analysis, and the corresponding R scripts, are provided in an open TRY File Archive (<https://www.try-db.org/TryWeb/Data.php>). Climate data WorldClim are publicly available via <https://www.worldclim.org/> (accessed May 2018). Soil data, namely SoilGrids (<https://soilgrids.org/>, accessed June 2018) are publicly available. Ecoregion information<sup>30</sup> shapefiles are publicly available (accessed January 2014, [Sciencebase.gov](http://Sciencebase.gov)). The estimate of species richness per ecoregion<sup>174</sup> is publicly available (accessed January 2014, [databasin.org](http://databasin.org)). Data for this study can be accessed on Github ([https://github.com/julijawig/Repo\\_ClimateSoil\\_TraitSpectrum](https://github.com/julijawig/Repo_ClimateSoil_TraitSpectrum)). For Extended Data Fig. 1 and Supplementary Fig. 7, the Geodata product of the Missions Database 'ArcWorld Supplement' (GMI) was used.

## Code availability

The code is available on Github ([https://github.com/julijawig/Repo\\_ClimateSoil\\_TraitSpectrum](https://github.com/julijawig/Repo_ClimateSoil_TraitSpectrum)).

Received: 21 January 2021; Accepted: 10 November 2021;  
Published online: 23 December 2021

## References

- Westoby, M. A leaf–height–seed (LHS) plant ecology strategy scheme. *Plant Soil* **199**, 213–227 (1998).
- Kraft, N. J. B. et al. Community assembly, coexistence and the environmental filtering metaphor. *Funct. Ecol.* **29**, 592–599 (2015).
- McGill, B. J., Enquist, B. J., Weiher, E. & Westoby, M. Rebuilding community ecology from functional traits. *Trends Ecol. Evol.* **21**, 178–185 (2006).
- Lavorel, S. & Garnier, E. Predicting changes in community composition and ecosystem functioning from plant traits: revisiting the Holy Grail. *Funct. Ecol.* **16**, 545–556 (2002).
- Musavi, T. et al. Potential and limitations of inferring ecosystem photosynthetic capacity from leaf functional traits. *Ecol. Evol.* **6**, 7352–7366 (2016).
- Scheiter, S., Langan, L. & Higgins, S. I. Next-generation dynamic global vegetation models: learning from community ecology. *New Phytol.* **198**, 957–969 (2013).
- Van Bodegom, P. M., Douma, J. C. & Verheijen, L. M. A fully traits-based approach to modeling global vegetation distribution. *Proc. Natl Acad. Sci. USA* **111**, 13733–13738 (2014).
- Schrod, F. et al. BHPMF—a hierarchical Bayesian approach to gap-filling and trait prediction for macroecology and functional biogeography. *Glob. Ecol. Biogeogr.* **24**, 1510–1521 (2015).
- Kattge, J. et al. TRY plant trait database—enhanced coverage and open access. *Glob. Change Biol.* **26**, 119–188 (2020).
- Diaz, S. et al. The global spectrum of plant form and function. *Nature* **529**, 167–171 (2015).
- Wright, I. J. et al. The worldwide leaf economics spectrum. *Nature* **428**, 821–827 (2004).
- Bruelheide, H. et al. Global trait–environment relationships of plant communities. *Nat. Ecol. Evol.* **2**, 1906–1917 (2018).
- Thomas, H. J. et al. Global plant trait relationships extend to the climatic extremes of the tundra biome. *Nat. Commun.* **11**, 1351 (2020).
- Kong, D. et al. Nonlinearity of root trait relationships and the root economics spectrum. *Nat. Commun.* **10**, 2203 (2019).
- Schimper, A. *Plant-Geography Upon A Physiological Basis* (Clarendon Press, 1903).
- Warming, E. *Oecology Of Plants* (Oxford, 1909).
- Raunkjær, C. in *Life Forms of Plants and Statistical Plant Geography*, 4–16 (Clarendon Press, 1934).
- Maire, V. et al. Global effects of soil and climate on leaf photosynthetic traits and rates. *Glob. Ecol. Biogeogr.* **24**, 706–717 (2015).
- Bjorkman, A. D. et al. Plant functional trait change across a warming tundra biome. *Nature* **562**, 57–62 (2018).
- Olson, M. E. et al. Plant height and hydraulic vulnerability to drought and cold. *Proc. Natl Acad. Sci. USA* **115**, 7551–7556 (2018).
- Moles, A. T. et al. Global patterns in plant height. *J. Ecol.* **97**, 923–932 (2009).
- Ordoñez, J. C. et al. A global study of relationships between leaf traits, climate and soil measures of nutrient fertility. *Glob. Ecol. Biogeogr.* **18**, 137–149 (2009).
- Simpson, A. H., Richardson, S. J. & Laughlin, D. C. Soil–climate interactions explain variation in foliar, stem, root and reproductive traits across temperate forests. *Glob. Ecol. Biogeogr.* **25**, 964–978 (2016).
- Wright, I. J. et al. Global climatic drivers of leaf size. *Science* **357**, 917–921 (2017).
- Atkin, O. K. et al. Global variability in leaf respiration in relation to climate, plant functional types and leaf traits. *New Phytol.* **206**, 614–636 (2015).
- Asner, G. P., Knapp, D. E., Anderson, C. B., Martin, R. E. & Vaughn, N. Large-scale climatic and geophysical controls on the leaf economics spectrum. *Proc. Natl Acad. Sci. USA* **113**, E4043–E4051 (2016).
- Moles, A. T. et al. Global patterns in seed size. *Glob. Ecol. Biogeogr.* **16**, 109–116 (2007).
- Blume, H.-P. et al. *Soil Science* 1st edn. (Springer, Berlin-Heidelberg, 2016).
- Seneviratne, S. I. et al. Investigating soil moisture–climate interactions in a changing climate: a review. *Earth-Sci. Rev.* **99**, 125–161 (2010).
- Olson, D. M. et al. Terrestrial ecoregions of the world: a new map of life on Earth. *BioScience* **51**, 933–938 (2001).
- Hastie, T., Tibshirani, R. & Friedman, J. *The Elements of Statistical Learning* (Springer, 2008).
- Chevan, A. & Sutherland, M. Hierarchical partitioning. *Am. Stat.* **45**, 90–96 (1991).
- Reich, P. B. & Oleksyn, J. Global patterns of plant leaf N and P in relation to temperature and latitude. *Proc. Natl Acad. Sci. USA* **101**, 11001–11006 (2004).
- Corner, E. J. H. The Durian theory or the origin of the modern tree. *Ann. Bot.* **XIII**, 367–414 (1949).
- Pietsch, K. A. et al. Global relationship of wood and leaf litter decomposability: the role of functional traits within and across plant organs. *Glob. Ecol. Biogeogr.* **23**, 1046–1057 (2014).
- FloresàMoreno, H. et al. Robustness of trait connections across environmental gradients and growth forms. *Glob. Ecol. Biogeogr.* **28**, 1806–1826 (2019).
- Chapin, F. S. The mineral nutrition of wild plants. *Annu. Rev. Ecol. Syst.* **11**, 233–260 (1980).
- Vitousek, P. *Nutrient Cycling and Limitation: Hawai'i as a Model System* (Princeton Univ. Press, 2004).
- Shipley, B., Vile, D., Garnier, E., Wright, I. J. & Poorter, H. Functional linkages between leaf traits and net photosynthetic rate: reconciling empirical and mechanistic models. *Funct. Ecol.* **19**, 602–615 (2005).
- He, T., Belcher, C. M., Lamont, B. B. & Lim, S. L. A 350-million-year legacy of fire adaptation among conifers. *J. Ecol.* **104**, 352–363 (2016).
- Bergmann, J., Ryo, M., Prati, D., Hempel, S. & Rillig, M. C. Root traits are more than analogues of leaf traits: the case for diaspore mass. *New Phytol.* **216**, 1130–1139 (2017).
- Aerts, R. The advantages of being evergreen. *Trends Ecol. Evol.* **10**, 402–407 (1995).
- Zanne, A. E. et al. Functional biogeography of angiosperms: life at the extremes. *New Phytol.* **218**, 1697–1709 (2018).
- Franklin, O. et al. Organizing principles for vegetation dynamics. *Nat. Plants* **6**, 444–453 (2020).



45. Legay, N. et al. Contribution of above- and below-ground plant traits to the structure and function of grassland soil microbial communities. *Ann. Bot.* **114**, 1011–1021 (2014).
46. Grime, J. P. Vegetation classification by reference to strategies. *Nature* **250**, 26–31 (1974).
47. Slessarev, E. W. et al. Water balance creates a threshold in soil pH at the global scale. *Nature* **540**, 567–569 (2016).
48. Fierer, N. & Jackson, R. B. The diversity and biogeography of soil bacterial communities. *Proc. Natl Acad. Sci. USA* **103**, 626–631 (2006).
49. Sinsabaugh, R. L. & Follstad Shah, J. J. Ecoenzymatic stoichiometry and ecological theory. *Annu. Rev. Ecol. Evol. Syst.* **43**, 313–343 (2012).
50. de Vries, F. T. et al. Abiotic drivers and plant traits explain landscape-scale patterns in soil microbial communities. *Ecol. Lett.* **15**, 1230–1239 (2012).
51. Zech, W., Schad, P. & Hintermaier-Erhard, G. *Böden der Welt—Ein Bildatlas* (Springer Spectrum, 2014).
52. Rosenberg, E. et al. (eds) *The Prokaryotes: Prokaryotic Communities and Ecophysiology* 4th edn. (Springer-Verlag, 2013).
53. Niinemets, Ä. Leaf age dependent changes in within-canopy variation in leaf functional traits: a meta-analysis. *J. Plant Res.* **129**, 313–338 (2016).
54. Butler, E. E. et al. Mapping local and global variability in plant trait distributions. *Proc. Natl Acad. Sci. USA* **114**, E10937–E10946 (2017).
55. Freschet, G. T. et al. Global to community scale differences in the prevalence of convergent over divergent leaf trait distributions in plant assemblages. *Global Ecol. Biogeogr.* **20**, 755–765 (2011).
56. Yemefack, M., Rossiter, D. G. & Njomgang, R. Multi-scale characterization of soil variability within an agricultural landscape mosaic system in southern Cameroon. *Geoderma* **125**, 117–143 (2005).
57. Oldeman, L., Hakkeling, R. & Sombroek, W. *Global Assessment of Soil Degradation (GLASOD): World Map of the Status of Human-induced Soil Degradation* (United Nations Environment Programme, 1991).
58. Ackerly, D. D. & Cornwell, W. K. A trait-based approach to community assembly: partitioning of species trait values into within- and among-community components. *Ecol. Lett.* **10**, 135–145 (2007).
59. Adler, P. B. *A Comparison of Livestock Grazing Effects on Sagebrush Steppe, USA, and Patagonian Steppe, Argentina*. PhD thesis (Colorado State University, 2003).
60. Adler, P. B., Milchunas, D. G., Lauenroth, W. K., Sala, O. E. & Burke, I. C. Functional traits of graminoids in semi-arid steppes: a test of grazing histories. *J. Appl. Ecol.* **41**, 653–663 (2004).
61. Adriaenssens, S. Dry deposition and canopy exchange for temperate tree species under high nitrogen deposition. PhD thesis, Ghent Univ. (2012).
62. Atkin, O. K., Schortemeyer, M., McFarlane, N. & Evans, J. R. The response of fast- and slow-growing Acacia species to elevated atmospheric CO<sub>2</sub>: an analysis of the underlying components of relative growth rate. *Oecologia* **120**, 544–554 (1999).
63. Atkin, O. K., Westbeek, M., Cambridge, M. L., Lambers, H. & Pons, T. L. Leaf respiration in light and darkness (a comparison of slow- and fast-growing *Poa* species). *Plant Physiol.* **113**, 961–965 (1997).
64. Auger, S. *L'Importance de la Variabilité Interspécifique des Traits Fonctionnels par Rapport à la Variabilité Intraspécifique Chez les Jeunes Arbres en Forêt Mature*. MSc thesis (Université de Sherbrooke, 2012).
65. Bahn, M. et al. in *Land-Use Changes in European Mountain Ecosystems. ECOMONT—Concept and Results* (eds Cernusca, A. et al.) 247–255 (Blackwell Wissenschaft, 1999).
66. Baker, T. R. et al. Do species traits determine patterns of wood production in Amazonian forests? *Biogeosciences* **6**, 297–307 (2009).
67. Bakker, C., Van Bodegom, P. M., Nelissen, H. J. M., Ernst, W. H. O. & Aerts, R. Plant responses to rising water table and nutrient management in calcareous dune slacks. *Plant Ecol.* **185**, 19–28 (2006).
68. Bakker, C., Rodenburg, J. & van Bodegom, P. M. Effects of Ca- and Fe-rich seepage on P availability and plant performance in calcareous dune soils. *Plant Soil* **275**, 111–122 (2005).
69. Baraloto, C. et al. Decoupled leaf and stem economics in rainforest trees. *Ecol. Lett.* **13**, 1338–1347 (2010).
70. Baraloto, C. et al. Functional trait variation and sampling strategies in species-rich plant communities. *Funct. Ecol.* **24**, 208–216 (2010).
71. Beckmann, M., Hock, M., Bruehlheide, H. & Erfmeier, A. The role of UV-B radiation in the invasion of *Hieracium pilosella*—a comparison of German and New Zealand plants. *Environ. Exp. Bot.* **75**, 173–180 (2012).
72. Blanco, C. C., Sosinski, E. E., dos Santos, B. R. C., da Silva, M. A. & Pillar, V. D. On the overlap between effect and response plant functional types linked to grazing. *Community Ecol.* **8**, 57–65 (2007).
73. Blonder, B. et al. The shrinkage effect biases estimates of paleoclimate. *Am. J. Bot.* **99**, 1756–1763 (2012).
74. Blonder, B., Violle, C. & Enquist, B. J. Assessing the causes and scales of the leaf economics spectrum using venation networks in *Populus tremuloides*. *J. Ecol.* **101**, 981–989 (2013).
75. Blonder, B. et al. Testing models for the leaf economics spectrum with leaf and whole-plant traits in *Arabidopsis thaliana*. *AoB Plants* **7**, plv049 (2015).
76. Blonder, B., Violle, C., Bentley, L. P. & Enquist, B. J. Venation networks and the origin of the leaf economics spectrum. *Ecol. Lett.* **14**, 91–100 (2011).
77. Bocanegra-González K.T., Fernández-Méndez, F. & David Galvis-Jiménez, J. Funtional groups of tres in secondary forests of the bajo calima region (Buenaventura, Colombia) *Boletín Científico Centro de Museos Museo de Historia natura* **19**, (2015).
78. Bodegom, P. M. V., Kanter, M. D. & Aerts, C. B. R. Radial oxygen loss, a plastic property of dune slack plant species. *Plant Soil* **271**, 351–364 (2005).
79. Bond-Lamberty, C. W. B. & Gower, S. T. Above- and belowground biomass and sapwood area allometric equations for six boreal tree species of northern Manitoba. *Can. J. For. Res.* **32**, 1441–1450 (2002).
80. Bond-Lamberty, C. W. B. & Gower, S. T. Leaf area dynamics of a boreal black spruce fire chronosequence. *Tree Physiol.* **22**, 993–1001 (2002).
81. Bond-Lamberty, C. W. B. & Gower, S. T. The use of multiple measurement techniques to refine estimates of conifer needle geometry. *Can. J. For. Res.* **33**, 101–105 (2003).
82. Bond-Lamberty, C. W. B. & Gower, S. Net primary production and net ecosystem production of a boreal black spruce fire chronosequence. *Glob. Change Biol.* **10**, 473–487 (2004).
83. Bragazza, L. Conservation priority of Italian alpine habitats: a floristic approach based on potential distribution of vascular plant species. *Biodivers. Conserv.* **18**, 2823–2835 (2009).
84. Choat, B. et al. Global convergence in the vulnerability of forests to drought. *Nature* **491**, 752–755 (2012).
85. Briemle, G., Nitsche, S. & Nitsche, L. in *BIOLFLOR—Eine Datenbank mit Biologisch-ökologischen Merkmalen zur Flora von Deutschland* (eds Klotz, S. et al.) 203–225 (Bundesamt für Naturschutz, 2002).
86. Brown, K. et al. Assessing natural resource use by forest-reliant communities in Madagascar using functional diversity and functional redundancy metrics. *PLoS ONE* <https://doi.org/10.1371/journal.pone.0024107> (2011).
87. Burrascano, S. et al. Wild boar rooting intensity determines shifts in understory composition and functional traits. *Community Ecol.* **16**, 244–253 (2015).
88. Butterfield, B. J. & Briggs, J. M. Regeneration niche differentiates functional strategies of desert woody plant species. *Oecologia* **165**, 477–487 (2011).
89. Byun, C., de Blois, S. & Brisson, J. Plant functional group identity and diversity determine biotic resistance to invasion by an exotic grass. *J. Ecol.* **101**, 128–139 (2013).
90. Campbell, C. et al. Acclimation of photosynthesis and respiration is asynchronous in response to changes in temperature regardless of plant functional group. *New Phytol.* **176**, 375–389 (2007).
91. Campetella, G. et al. Patterns of plant trait–environment relationships along a forest succession chronosequence. *Agric. Ecosyst. Environ.* **145**, 38–48 (2011).
92. Carswell, F. E. et al. Photosynthetic capacity in a central Amazonian rain forest. *Tree Physiol.* **20**, 179–186 (2000).
93. Castro-Diez, P., Puyravaud, J. P., Cornelissen, J. H. C. & Villar-Salvador, P. Stem anatomy and relative growth rate in seedlings of a wide range of woody plant species and types. *Oecologia* **116**, 57–66 (1998).
94. Castro-Diez, P., Puyravaud, J. P. & Cornelissen, J. H. C. Leaf structure and anatomy as related to leaf mass per area variation in seedlings of a wide range of woody plant species and types. *Oecologia* **124**, 476–486 (2000).
95. Cavender-Bares, A. K. J. & Miles, B. Phylogenetic structure of Floridian plant communities depends on taxonomic and spatial scale. *Ecology* **87**, 109–122 (2006).
96. Cavender-Bares, L. S. J. & Savage, J. Atmospheric and soil drought reduce nocturnal conductance in live oaks. *Tree Physiol.* **27**, 522–620 (2007).
97. Cerabolini, B. E. L. et al. Can CSR classification be generally applied outside Britain? *Plant Ecol.* **210**, 253–261 (2010).
98. Chave, J. et al. Towards a worldwide wood economics spectrum. *Ecol. Lett.* **12**, 351–366 (2009).
99. Chen, Y., Han, W., Tang, L., Tang, Z. & Fang, J. Leaf nitrogen and phosphorus concentrations of woody plants differ in responses to climate, soil and plant growth form. *Ecography* **36**, 178–184 (2011).
100. Choat, B. et al. Global convergence in the vulnerability of forests to drought. *Nature* **491**, 752–755 (2012).
101. Choat, B., Sack, L. & Holbrook, N. M. Diversity of hydraulic traits in nine *Cordia* species growing in tropical forests with contrasting precipitation. *New Phytol.* **175**, 686–698 (2007).
102. Coomes, D. A., Heathcote, S., Godfrey, E. R. & Shepherd, J. J. Scaling of xylem vessels and veins within the leaves of oak species. *Biol. Lett.* **4**, 302–306 (2008).
103. Cornelissen, J., Aerts, R., Cerabolini, B., Werger, M. & van der Heijden, M. Carbon cycling traits of plant species are linked with mycorrhizal strategy. *Oecologia* **129**, 611–619 (2001).
104. Cornelissen, J. H. C. An experimental comparison of leaf decomposition rates in a wide range of temperate plant species and types. *J. Ecol.* **84**, 573–582 (1996).
105. Cornelissen, J. H. C., Diez, P. C. & Hunt, R. Seedling growth, allocation and leaf attributes in a wide range of woody plant species and types. *J. Ecol.* **84**, 755–765 (1996).

106. Cornelissen, J. H. C., Werger, M. J. A., Castro-Diez, P., van Rheenen, J. W. A., & Rowland, A. P. Foliar nutrients in relation to growth, allocation and leaf traits in seedlings of a wide range of woody plant species and types. *Oecologia* **111**, 460–469 (1997).
107. Cornelissen, J. H. C. et al. Leaf structure and defence control litter decomposition rate across species and life forms in regional floras on two continents. *New Phytol.* **143**, 191–200 (1999).
108. Cornelissen, J. H. C. A triangular relationship between leaf size and seed size among woody species: allometry, ontogeny, ecology and taxonomy. *Oecologia* **118**, 248–255 (1999).
109. Cornelissen, J. H. C., Aerts, R., Cerabolini, B., Werger, M. J. A. & van der Heijden, M. G. A. Carbon cycling traits of plant species are linked with mycorrhizal strategy. *Oecologia* **129**, 611–619 (2001).
110. Cornelissen, J. H. C. et al. Leaf digestibility and litter decomposability are related in a wide range of subarctic plant species and types. *Funct. Ecol.* **18**, 779–786 (2004).
111. Cornelissen, J. H. C. et al. Functional traits of woody plants: correspondence of species rankings between field adults and laboratory-grown seedlings? *J. Veg. Sci.* **14**, 311–322 (2003).
112. Cornelissen, J. H. C., Diez, P. C. & Hunt, R. Seedling growth, allocation and leaf attributes in a wide range of woody plant species and types. *J. Ecol.* **84**, 755 (1996).
113. Cornelissen, J. H. C. et al. Leaf structure and defence control litter decomposition rate across species and life forms in regional floras on two continents. *New Phytol.* **143**, 191–200 (1999).
114. Schwilk, D. W., Cornwell, W. K. & Ackerly, D. D. A trait-based test for habitat filtering: convex hull volume. *Ecology* **87**, 1465–1471 (2006).
115. Cornwell, W. K. & Ackerly, D. D. Community assembly and shifts in plant trait distributions across an environmental gradient in coastal California. *Ecol. Monogr.* **79**, 109–126 (2009).
116. Cornwell, W. K., Bhaskar, R., Sack, L., Cordell, S. & Lurch, C. K. Adjustment of structure and function of Hawaiian *Metrosideros polymorpha* at high vs. low precipitation. *Funct. Ecol.* **21**, 1063–1071 (2007).
117. Cornwell, W. K. et al. Plant species traits are the predominant control on litter decomposition rates within biomes worldwide. *Ecol. Lett.* **11**, 1065–1071 (2008).
118. Craine, J. M. et al. Global patterns of foliar nitrogen isotopes and their relationships with climate, mycorrhizal fungi, foliar nutrient concentrations, and nitrogen availability. *New Phytol.* **183**, 980–992 (2009).
119. Craine, J. M., Lee, W. G., Bond, W. J., Williams, R. J. & Johnson, L. C. Environmental constraints on a global relationship among leaf and root traits of grasses. *Ecology* **86**, 12–19 (2005).
120. Craine, J. M. et al. Functional consequences of climate change-induced plant species loss in a tallgrass prairie. *Oecologia* **165**, 1109–1117 (2011).
121. Craine, J. M. et al. Global diversity of drought tolerance and grassland climate-change resilience. *Nat. Clim. Change* **3**, 63–67 (2012).
122. Craven, D. et al. Between and within-site comparisons of structural and physiological characteristics and foliar nutrient content of 14 tree species at a wet, fertile site and a dry, infertile site in Panama. *For. Ecol. Manag.* **238**, 335–346 (2007).
123. Craven, D. et al. Seasonal variability of photosynthetic characteristics influences growth of eight tropical tree species at two sites with contrasting precipitation in Panama. *For. Ecol. Manag.* **261**, 1643–1653 (2011).
124. Dainese, M. & Bragazza, L. Plant traits across different habitats of the Italian alps: a comparative analysis between native and alien species. *Alpine Bot.* **122**, 11–21 (2012).
125. de Araujo, A. et al. *LBA-ECO CD-02 C and N Isotopes in Leaves and Atmospheric CO<sub>2</sub>, Amazonas, Brazil* (ORNL DAAC, 2012); <http://daac.ornl.gov>
126. de Vries, F. T. & Bardgett, R. D. Plant community controls on short-term ecosystem nitrogen retention. *New Phytol.* **210**, 861–874 (2016).
127. Demey, A. et al. Nutrient input from hemiparasitic litter favors plant species with a fast-growth strategy. *Plant Soil* **371**, 53–66 (2013).
128. Diaz, S. et al. The plant traits that drive ecosystems: evidence from three continents. *J. Veg. Sci.* **15**, 295–304 (2004).
129. Domingues, T. F., Berry, J. A., Martinelli, L. A., Ometto, J. P. H. B. & Ehleringer, J. R. Parameterization of canopy structure and leaf-level gas exchange for an eastern Amazonian tropical rain forest (Tapajós National Forest, Pará, Brazil). *Earth Interact.* <https://doi.org/10.1175/EI149.1> (2005).
130. Domingues, T. F., Martinelli, L. A. & Ehleringer, J. R. Ecophysiological traits of plant functional groups in forest and pasture ecosystems from eastern Amazonia, Brazil. *Plant Ecol.* **193**, 101–112 (2007).
131. Domingues, T. F. et al. Co-limitation of photosynthetic capacity by nitrogen and phosphorus in West Africa woodlands. *Plant Cell Environ.* **33**, 959–980 (2010).
132. Duarte, Ld. S., Carlucci, M. B., Hartz, S. M. & Pillar, V. D. Plant dispersal strategies and the colonization of Araucaria forest patches in a grassland-forest mosaic. *J. Veg. Sci.* **18**, 847–858 (2007).
133. DunbarCo, S., Sporck, M. J. & Sack, L. Leaf trait diversification and design in seven rare taxa of the Hawaiian *Plantago* radiation. *Int. J. Plant Sci.* **170**, 61–75 (2009).
134. Durka, W. In *BIOLFLOR—Eine Datenbank mit Biologisch-ökologischen Merkmalen zur Flora von Deutschland* (eds Klotz, S. et al.) 75–91 (Bundesamt für Naturschutz, 2002).
135. Durka, W. In *BIOLFLOR—Eine Datenbank mit Biologisch-ökologischen Merkmalen zur Flora von Deutschland* (eds Klotz, S. et al.) 57–74 (Bundesamt für Naturschutz, 2002).
136. Durka, W. In *BIOLFLOR—Eine Datenbank mit Biologisch-ökologischen Merkmalen zur Flora von Deutschland* (eds Klotz, S. et al.) 133–175 (Bundesamt für Naturschutz, 2002).
137. Medlyn, B. E. & Jarvis, P. G. Design and use of a database of model parameters from elevated [CO<sub>2</sub>] experiments. *Ecol. Model.* **124**, 69–83 (1999).
138. Everwand, G., Fry, E. L., Eggers, T. & Manning, P. Seasonal variation in the capacity for plant trait measures to predict grassland carbon and water fluxes. *Ecosystems* **17**, 1095–1108 (2014).
139. Fazaeyli, F., Banerjee, A., Kattge, J., Schrodt, F. & Reich, P. B. Uncertainty quantified matrix completion using Bayesian Hierarchical Matrix factorization. In *Proc. 13th International Conference on Machine Learning and Applications* (eds Ferri, C. et al.) 312–317 (International Conference on Machine Learning and Applications (ICMLA), 2014).
140. Fagüндеz, J. & Izco, J. Seed morphology of the European species of *Erica* L. sect. *Arsace* Salisb. ex Benth. (Ericaceae). *Acta Bot. Gall.* **157**, 45–54 (2010).
141. Fonseca, C. R., Overton, J. M., Collins, B. & Westoby, M. Shifts in trait-combinations along rainfall and phosphorus gradients. *J. Ecol.* **88**, 964–977 (2000).
142. Fortunel, C. et al. Leaf traits capture the effects of land use changes and climate on litter decomposability of grasslands across Europe. *Ecology* **90**, 598–611 (2009).
143. Frainer, A. & McKie, B. G. Shifts in the diversity and composition of consumer traits constrain the effects of land use on stream ecosystem functioning. *Adv. Ecol. Res.* **52**, 169–200 (2015).
144. Frenette-Dussault, C., Shipley, B., Léger, J.-F., Meziane, D. & Hingrat, Y. Functional structure of an arid steppe plant community reveals similarities with Grime's C-S-R theory. *J. Veg. Sci.* **23**, 208–222 (2011).
145. Freschet, G. T., Cornelissen, J. H. C., van Logtestijn, R. S. P. & Aerts, R. Evidence of the plant economics spectrum in a subarctic flora. *J. Ecol.* **98**, 362–373 (2010).
146. Freschet, G. T., Cornelissen, J. H. C., van Logtestijn, R. S. P. & Aerts, R. Substantial nutrient resorption from leaves, stems and roots in a subarctic flora: what is the link with other resource economics traits? *New Phytol.* **186**, 879–889 (2010).
147. Fry, E. L., Power, S. A. & Manning, P. Trait-based classification and manipulation of plant functional groups for biodiversity–ecosystem function experiments. *J. Veg. Sci.* **25**, 248–261 (2013).
148. Fyllas, N. M. et al. Basin-wide variations in foliar properties of Amazonian forest: phylogeny, soils and climate. *Biogeosciences* **6**, 2677–2708 (2009).
149. Gachet, S., Vêla, E. & Taton, T. BASECO: a floristic and ecological database of Mediterranean French flora. *Biodivers. Conserv.* **14**, 1023–1034 (2005).
150. Gallagher, R. V. & Leishman, M. R. A global analysis of trait variation and evolution in climbing plants. *J. Biogeogr.* **39**, 1757–1771 (2012).
151. Garnier, E. et al. Assessing the effects of land-use change on plant traits, communities and ecosystem functioning in grasslands: a standardized methodology and lessons from an application to 11 European sites. *Ann. Bot.* **99**, 967–985 (2007).
152. Givnish, T. J., Montgomery, R. A. & Goldstein, G. Adaptive radiation of photosynthetic physiology in the Hawaiian lobeliads: light regimes, static light responses, and whole-plant compensation points. *Am. J. Bot.* **91**, 228–246 (2004).
153. Guerin, G. R., Wen, H. & Lowe, A. J. Leaf morphology shift linked to climate change. *Biol. Lett.* **8**, 882–886 (2012).
154. Gutiérrez, A. G. & Huth, A. Successional stages of primary temperate rainforests of Chiloé Island, Chile. *Perspect. Plant Ecol. Evol. Syst.* **14**, 243–256 (2012).
155. Guy, A. L., Mischkolz, J. M. & Lamb, E. G. Limited effects of simulated acidic deposition on seedling survivorship and root morphology of endemic plant taxa of the Athabasca sand dunes in well-watered greenhouse trials. *Botany* **91**, 176–181 (2013).
156. Han, W. et al. Floral, climatic and soil pH controls on leaf ash content in China's terrestrial plants. *Glob. Ecol. Biogeogr.* **21**, 376–382 (2011).
157. Han, W., Fang, J., Guo, D. & Zhang, Y. Leaf nitrogen and phosphorus stoichiometry across 753 terrestrial plant species in China. *New Phytol.* **168**, 377–385 (2005).
158. Hao, G.-Y., Sack, L., Wang, A.-Y., Cao, K.-F. & Goldstein, G. Differentiation of leaf water flux and drought tolerance traits in hemiepiphytic and non-hemiepiphytic *Ficus* tree species. *Funct. Ecol.* **24**, 731–740 (2010).
159. He, J.-S. et al. A test of the generality of leaf trait relationships on the Tibetan plateau. *New Phytol.* **170**, 835–848 (2006).

160. Hickler, T. *Plant Functional Types and Community Characteristics along Environmental Gradients on Öland's Great Alvar (Sweden)*. Masters thesis (University of Lund, 1999).
161. Hoof, J., Sack, L., Webb, D. T. & Nilsen, E. T. Contrasting structure and function of pubescent and glabrous varieties of Hawaiian *Metrosideros polymorpha* (Myrtaceae) at high elevation. *Biotropica* **40**, 113–118 (2008).
162. Husson, A. F., Josse, J., Le, S., Mazet, J. & Husson, M. F. Package 'FactoMineR' (CRAN, 2017).
163. Jacobs, B. et al. Unraveling the Phylogeny of Heptacodium and Zabelia (Caprifoliaceae): An Interdisciplinary Approach. *Syst. Bot.* **36**, 231–252 (2011).
164. Jansen, S., Decraene, L. P. R. & Smets, E. On the wood and stem anatomy of *Monococcus echinophorus* (Phytolaccaceae s.l.). *Syst. Geogr. Plants* **70**, 171 (2000).
165. Jansen, S. et al. Contributions to the wood anatomy of the Rubioideae (Rubiaceae). *J. Plant Res.* **114**, 269–289 (2001).
166. Jansen, S., Piesschaert, F. & Smets, E. Wood anatomy of Elaeagnaceae, with comments on vested pits, helical thickenings, and systematic relationships. *Am. J. Bot.* **87**, 20 (2000).
167. Jansen, S., Robbrecht, E., Beeckman, H. & Smets, E. *Gaertnera* and *Pagamea*: genera within the Psychotriaceae or constituting the tribe Gaertneraceae? A wood anatomical and palynological approach. *Bot. Acta* **109**, 466–476 (1996).
168. S., J., E., R., H., B. & Smets, E. Comparative wood anatomy of African Coffeae (Rubiaceae-Rubioideae). *Belg. J. Bot.* **130**, 47–58 (1997).
169. Kattge, J., Knorr, W., Raddatz, T. & Wirth, C. Quantifying photosynthetic capacity and its relationship to leaf nitrogen content for global-scale terrestrial biosphere models. *Glob. Change Biol.* **15**, 976–991 (2009).
170. Kazakou, E., Vile, D., Shipley, B., Gallet, C. & Garnier, E. Co-variations in litter decomposition, leaf traits and plant growth in species from a Mediterranean old-field succession. *Funct. Ecol.* **20**, 21–30 (2006).
171. Kerkhoff, A. J., Fagan, W. F., Elser, J. J. & Enquist, B. J. Phylogenetic and growth form variation in the scaling of nitrogen and phosphorus in the seed plants. *Am. Nat.* **168**, E103–E122 (2006).
172. Kew, R. B. G. *Seed Information Database—SID* (Kew, 2008); <http://data.kew.org/sid/>
173. Kichenin, E., Wardle, D. A., Peltzer, D. A., Morse, C. W. & Freschet, G. T. Contrasting effects of plant inter- and intraspecific variation on community-level trait measures along an environmental gradient. *Funct. Ecol.* **27**, 1254–1261 (2013).
174. Kier, G. et al. Global patterns of plant diversity and floristic knowledge. *J. Biogeogr.* **32**, 1107–1116 (2005).
175. Kirkup, D., Malcolm, P., Christian, G. & Paton, A. Towards a digital African flora. *Taxon* **54**, 457 (2005).
176. Kleyer, M. et al. The LEDA traitbase: a database of life-history traits of the northwest European flora. *J. Ecol.* **96**, 1266–1274 (2008).
177. Klotz, S. & Kühn, I. in *BIOLFLOR—Eine Datenbank mit Biologisch-ökologischen Merkmalen zur Flora von Deutschland* (eds Klotz, S. et al.) 119–126 (Bundesamt für Naturschutz, 2002).
178. Klotz, S. & Kühn, I. in *BIOLFLOR—Eine Datenbank mit Biologisch-ökologischen Merkmalen zur Flora von Deutschland* (eds Klotz, S. et al.) 241–246 (Bundesamt für Naturschutz, 2002).
179. Klotz, S. & Kühn, I. in *BIOLFLOR—Eine Datenbank mit Biologisch-ökologischen Merkmalen zur Flora von Deutschland* (eds Klotz, S. et al.) 273–281 (Bundesamt für Naturschutz, 2002).
180. Klotz, S. & Kühn, I. in *BIOLFLOR—Eine Datenbank mit Biologisch-ökologischen Merkmalen zur Flora von Deutschland* (eds Klotz, S. et al.) 197–201 (Bundesamt für Naturschutz, 2002).
181. Koike, F. Plant traits as predictors of woody species dominance in climax forest communities. *J. Veg. Sci.* **12**, 327–336 (2001).
182. Kraft, N. J. B. & Ackerly, D. D. Functional trait and phylogenetic tests of community assembly across spatial scales in an Amazonian forest. *Ecol. Monogr.* **80**, 401–422 (2010).
183. Kraft, N. J. B., Valencia, R. & Ackerly, D. D. Functional traits and niche-based tree community assembly in an Amazonian forest. *Science* **322**, 580–582 (2008).
184. Krumbiegel, A. in *BIOLFLOR—Eine Datenbank mit Biologisch-ökologischen Merkmalen zur Flora von Deutschland* (eds Klotz, S. et al.) 93–118 (Bundesamt für Naturschutz, 2002).
185. Kühn, I. in *BIOLFLOR—Eine Datenbank mit Biologisch-ökologischen Merkmalen zur Flora von Deutschland* (eds Klotz, S. et al.) 47–56 (Bundesamt für Naturschutz, 2002).
186. Kuhn, I., Durka, W. & Klotz, S. Biolflor—a new plant-trait database as a tool for plant invasion ecology. *Divers. Distrib.* **10**, 363–365 (2004).
187. Kühn, I. & Klotz, S. in *BIOLFLOR—Eine Datenbank mit Biologisch-ökologischen Merkmalen zur Flora von Deutschland* (eds Klotz, S. et al.) 227–239 (Bundesamt für Naturschutz, 2002).
188. Kurokawa, H. & Nakashizuka, T. Leaf herbivory and decomposability in a Malaysian tropical rain forest. *Ecology* **89**, 2645–2656 (2008).
189. Laughlin, D. C., Fulé, P. Z., Huffman, D. W., Crouse, J. & Laliberté, E. Climatic constraints on trait-based forest assembly. *J. Ecol.* **99**, 1489–1499 (2011).
190. Laughlin, D. C., Leppert, J. J., Moore, M. M. & Sieg, C. H. A multi-trait test of the leaf-height-seed plant strategy scheme with 133 species from a pine forest flora. *Funct. Ecol.* **24**, 493–501 (2009).
191. Lens, F. Comparative wood anatomy of Epacrids (Styphelioideae, Ericaceae s.l.). *Ann. Bot.* **91**, 835–856 (2003).
192. Lens, F., Baas, P., Jansen, S. & Smets, E. A search for phylogenetically informative wood characters within Lecythidaceae s.l. *Am. J. Bot.* **94**, 483–502 (2007).
193. Lens, F., Dressler, S., Jansen, S., van Evelghem, L. & Smets, E. Relationships within balsaminoid Ericales: a wood anatomical approach. *Am. J. Bot.* **92**, 941–953 (2005).
194. Lens, F., Eeckhout, S., Zwartjes, R., Smets, E. & Janssens, S. B. The multiple fuzzy origins of woodiness within Balsaminaceae using an integrated approach: where do we draw the line? *Ann. Bot.* **109**, 783–799 (2011).
195. Lens, F., Endress, M. E., Baas, P., Jansen, S. & Smets, E. Vessel grouping patterns in subfamilies Apocynoideae and Periplocoideae confirm phylogenetic value of wood structure within Apocynaceae. *Am. J. Bot.* **96**, 2168–2183 (2009).
196. Lens, F., Groeninckx, I., Smets, E. & Dessein, S. Woodiness within the Spermaceae–Knoxieae alliance (Rubiaceae): retention of the basal woody condition in Rubiaceae or recent innovation? *Ann. Bot.* **103**, 1049–1064 (2009).
197. Lens, F., Jansen, S., Caris, P., Serlet, L. & Smets, E. Comparative wood anatomy of the primuloid clade (Ericales s.l.). *Syst. Bot.* **30**, 163–183 (2005).
198. Lens, F., Jansen, S., Robbrecht, E. & Smets, E. Wood anatomy of the Vangueriae (Ixoroideae-Rubiacaceae), with special emphasis on some geofrutices. *IAWA J.* **21**, 443–455 (2000).
199. Lens, F. et al. The wood anatomy of the polyphyletic Icacinae s.l., and their relationships within asterids. *Taxon* **57**, 525–552 (2008).
200. Lens, F., Kron, K. A., Luteyn, J. L., Smets, E. & Jansen, S. Comparative wood anatomy of the blueberry tribe (Vaccinieae, Ericaceae s.l.). *Ann. Missouri Bot. Gard.* **91**, 566–592 (2004).
201. Lens, F., Smets, E. & Jansen, S. Comparative wood anatomy of Andromedeae s.s., Gaultherieae, Lyonieae and Oxydendreae (Vaccinioideae, Ericaceae s.l.). *Bot. J. Linn. Soc.* **144**, 161–179 (2004).
202. Lens, F., Smets, E. & Melzer, S. Stem anatomy supports *Arabidopsis thaliana* as a model for insular woodiness. *New Phytol.* **193**, 12–17 (2011).
203. Lens, F. et al. Testing hypotheses that link wood anatomy to cavitation resistance and hydraulic conductivity in the genus *Acer*. *New Phytol.* **190**, 709–723 (2010).
204. Li, H., Liang, Y., Xu, Q. & Cao, D. Key wavelengths screening using competitive adaptive reweighted sampling method for multivariate calibration. *Anal. Chim. Acta* **648**, 77–84 (2009).
205. Louault, F., Pillar, V. D., Aufrère, J., Garnier, E. & Soussana, J. F. Plant traits and functional types in response to reduced disturbance in a semi-natural grassland. *J. Veg. Sci.* **16**, 151–160 (2005).
206. Loveys, B. R. et al. Thermal acclimation of leaf and root respiration: an investigation comparing inherently fast- and slow-growing plant species. *Glob. Change Biol.* **9**, 895–910 (2003).
207. Malhado, A. C. M. et al. Drip-tips are associated with intensity of precipitation in the Amazon rain forest. *Biotropica* **44**, 728–737 (2012).
208. Malhado, A. C. M. et al. Spatial trends in leaf size of Amazonian rainforest trees. *Biogeosciences* **6**, 1563–1576 (2009).
209. Malhado, A. C. M. et al. Spatial distribution and functional significance of leaf lamina shape in Amazonian forest trees. *Biogeosciences* **6**, 1577–1590 (2009).
210. Malhado, A. C. M. et al. Are compound leaves an adaptation to seasonal drought or to rapid growth? Evidence from the Amazon rain forest. *Glob. Ecol. Biogeogr.* **19**, 852–862 (2010).
211. Manning, P., Houston, K. & Evans, T. Shifts in seed size across experimental nitrogen enrichment and plant density gradients. *Basic Appl. Ecol.* **10**, 300–308 (2009).
212. Markesteijn, L., Poorter, L., Paz, H., Sack, L. & Bongers, F. Ecological differentiation in xylem cavitation resistance is associated with stem and leaf structural traits. *Plant Cell Environ.* **34**, 137–148 (2011).
213. Martin, R. E., Asner, G. P. & Sack, L. Genetic variation in leaf pigment, optical and photosynthetic function among diverse phenotypes of *Metrosideros polymorpha* grown in a common garden. *Oecologia* **151**, 387–400 (2007).
214. McDonald, P. G., Fonseca, C. R., Overton, J. M. & Westoby, M. Leaf-size divergence along rainfall and soil-nutrient gradients: is the method of size reduction common among clades? *Funct. Ecol.* **17**, 50–57 (2003).
215. McKenna, M. F. & Shipley, B. Interacting determinants of interspecific relative growth: empirical patterns and a theoretical explanation. *Écoscience* **6**, 286–296 (1999).
216. Medlyn, B. E. et al. Effects of elevated [CO<sub>2</sub>] on photosynthesis in European forest species: a meta-analysis of model parameters. *Plant Cell Environ.* **22**, 1475–1495 (1999).

217. Medlyn, B. E. et al. Stomatal conductance of forest species after long-term exposure to elevated CO<sub>2</sub> concentration: a synthesis. *New Phytol.* **149**, 247–264 (2001).
218. Meir, P. et al. Acclimation of photosynthetic capacity to irradiance in tree canopies in relation to leaf nitrogen concentration and leaf mass per unit area. *Plant Cell Environ.* **25**, 343–357 (2002).
219. Meir, P., Levy, P. E., Grace, J. & Jarvis, P. G. Photosynthetic parameters from two contrasting woody vegetation types in West Africa. *Plant Ecol.* **192**, 277–287 (2007).
220. Mencuccini, M. The ecological significance of long-distance water transport: short-term regulation, long-term acclimation and the hydraulic costs of stature across plant life forms. *Plant Cell Environ.* **26**, 163–182 (2003).
221. Meng, T.-T. et al. Responses of leaf traits to climatic gradients: Adaptive variation versus compositional shifts. *Biogeosciences* **12**, 5339–5352 (2015).
222. Messier, J., McGill, B. J., Enquist, B. J. & Lechowicz, M. J. Trait variation and integration across scales: is the leaf economic spectrum present at local scales? *Ecography* **40**, 685–697 (2016).
223. Messier, J., McGill, B. J. & Lechowicz, M. J. How do traits vary across ecological scales? A case for trait-based ecology. *Ecol. Lett.* **13**, 838–848 (2010).
224. Meziane, D. & Shipley, B. Interacting components of interspecific relative growth rate: constancy and change under differing conditions of light and nutrient supply. *Funct. Ecol.* **13**, 611–622 (1999).
225. Milla, R. & Reich, P. B. Multi-trait interactions, not phylogeny, fine-tune leaf size reduction with increasing altitude. *Ann. Bot.* **107**, 455–465 (2011).
226. Minden, V., Andrantschke, S., Spalke, J., Timmermann, H. & Kleyer, M. Plant trait–environment relationships in salt marshes: deviations from predictions by ecological concepts. *Perspect. Plant Ecol. Evol. Syst.* **14**, 183–192 (2012).
227. Minden, V. & Kleyer, M. Testing the effect–response framework: key response and effect traits determining above-ground biomass of salt marshes. *J. Veg. Sci.* **22**, 387–401 (2011).
228. Mischkolz, J. M. *Selecting and Evaluating Native Forage Mixtures for the Mixed Grass Prairie*. Msc thesis (University of Saskatchewan, 2013).
229. Moretti, M. & Legg, C. Combining plant and animal traits to assess community functional responses to disturbance. *Ecography* **32**, 299–309 (2009).
230. Müller, S. C., Overbeck, G. E., Pfadenhauer, J. & Pillar, V. D. Plant functional types of woody species related to fire disturbance in forest–grassland ecotones. *Plant Ecol.* **189**, 1–14 (2006).
231. Nakahashi, C. D., Frole, K. & Sack, L. Bacterial leaf nodule symbiosis in *Ardisia* (Myrsinaceae): does it contribute to seedling growth capacity? *Plant Biol.* **7**, 495–500 (2005).
232. Niinemets, U. Components of leaf dry mass per area—thickness and density—alter leaf photosynthetic capacity in reverse directions in woody plants. *New Phytol.* **144**, 35–47 (1999).
233. Niinemets, U. Global-scale climatic controls of leaf dry mass per area, density, and thickness in trees and shrubs. *Ecology* **82**, 453–469 (2001).
234. Ogaya, R. & Peñuelas, J. Comparative field study of *Quercus ilex* and *Phillyrea latifolia*: photosynthetic response to experimental drought conditions. *Environ. Exp. Bot.* **50**, 137–148 (2003).
235. Ogaya, R. & Peñuelas, J. Contrasting foliar responses to drought in *Quercus ilex* and *Phillyrea latifolia*. *Biol. Plant.* **50**, 373–382 (2006).
236. Ogaya, R. & Peñuelas, J. Tree growth, mortality, and above-ground biomass accumulation in a holm oak forest under a five-year experimental field drought. *Plant Ecol.* **189**, 291–299 (2006).
237. Ogaya, R. & Peñuelas, J. Changes in leaf δ<sup>13</sup>C and δ<sup>15</sup>N for three Mediterranean tree species in relation to soil water availability. *Acta Oecol.* **34**, 331–338 (2008).
238. Onoda, Y. et al. Global patterns of leaf mechanical properties. *Ecol. Lett.* **14**, 301–312 (2011).
239. Ordoñez, J. C. et al. Leaf habit and woodiness regulate different leaf economy traits at a given nutrient supply. *Ecology* **91**, 3218–3228 (2010).
240. Otto, B. in *BIOLFLORE—Eine Datenbank mit Biologisch-ökologischen Merkmalen zur Flora von Deutschland* (eds Klotz, S. et al.) 177–196 (Bundesamt für Naturschutz, 2002).
241. Overbeck, G. E., Müller, S. C., Pillar, V. D. & Pfadenhauer, J. Fine-scale post-fire dynamics in southern Brazilian subtropical grassland. *J. Veg. Sci.* **16**, 655–664 (2005).
242. Overbeck, G. E. & Pfadenhauer, J. Adaptive strategies in burned subtropical grassland in southern Brazil. *Flora* **202**, 27–49 (2007).
243. Baas, P., Smets, E. & Jansen, S. Vegetative anatomy and affinities of *Dirachma socotrana* (Dirachmaceae). *Syst. Bot.* **26**, 231–241 (2001).
244. Pakeman, R. J. et al. Impact of abundance weighting on the response of seed traits to climate and land use. *J. Ecol.* **96**, 355–366 (2008).
245. Pakeman, R. J., Lep, J., Kleyer, M., Lavorel, S. & Garnie, E. Relative climatic, edaphic and management controls of plant functional trait signatures. *J. Veg. Sci.* **20**, 148–159 (2009).
246. Papanastasis, M. et al. Leaf traits capture the effects of land use changes and climate on litter decomposability of grasslands across Europe. *Ecology* **90**, 598–611 (2009).
247. Patiño, S. et al. Branch xylem density variations across the Amazon basin. *Biogeosciences* **6**, 545–568 (2009).
248. Paula, S. et al. Fire-related traits for plant species of the Mediterranean basin. *Ecology* **90**, 1420–1420 (2009).
249. Paula, S. & Pausas, J. G. Burning seeds: germinative response to heat treatments in relation to resprouting ability. *J. Ecol.* **96**, 543–552 (2008).
250. Peco, B., de Pablos, I., Traba, J. & Levasor, C. The effect of grazing abandonment on species composition and functional traits: the case of Dehesa grasslands. *Basic Appl. Ecol.* **6**, 175–183 (2005).
251. Peñuelas, J. et al. Faster returns on ‘leaf economics’ and different biogeochemical niche in invasive compared with native plant species. *Glob. Change Biol.* **16**, 2171–2185 (2009).
252. Peñuelas, J. et al. Higher allocation to low cost chemical defenses in invasive species of Hawaii. *J. Chem. Ecol.* **36**, 1255–1270 (2010).
253. Petter, G. et al. Functional leaf traits of vascular epiphytes: vertical trends within the forest, intra- and interspecific trait variability, and taxonomic signals. *Funct. Ecol.* **30**, 188–198 (2015).
254. Pierce, S., Brusa, G., Sartori, M. & Cerabolini, B. E. L. Combined use of leaf size and economics traits allows direct comparison of hydrophyte and terrestrial herbaceous adaptive strategies. *Ann. Bot.* **109**, 1047–1053 (2012).
255. Pierce, S., Brusa, G., Vagge, I. & Cerabolini, B. E. L. Allocating CSR plant functional types: the use of leaf economics and size traits to classify woody and herbaceous vascular plants. *Funct. Ecol.* **27**, 1002–1010 (2013).
256. Pierce, S., Ceriani, R. M., De Andreis, R., Luzzaro, A. & Cerabolini, B. The leaf economics spectrum of Poaceae reflects variation in survival strategies. *Plant Biosyst.* **141**, 337–343 (2007).
257. Pierce, S., Luzzaro, A., Caccianiga, M., Ceriani, R. M. & Cerabolini, B. Disturbance is the principal  $\alpha$ -scale filter determining niche differentiation, coexistence and biodiversity in an alpine community. *J. Ecol.* **95**, 698–706 (2007).
258. Pillar, V. D. & Sosinski, E. E. An improved method for searching plant functional types by numerical analysis. *J. Veg. Sci.* **14**, 323–332 (2003).
259. Powers, J. S. & Tiffin, P. Plant functional type classifications in tropical dry forests in Costa Rica: leaf habit versus taxonomic approaches. *Funct. Ecol.* **24**, 927–936 (2010).
260. Prentice, I. C. et al. Evidence of a universal scaling relationship for leaf CO<sub>2</sub> drawdown along an aridity gradient. *New Phytol.* **190**, 169–180 (2010).
261. Preston, K. A., Cornwell, W. K. & DeNoyer, J. L. Wood density and vessel traits as distinct correlates of ecological strategy in 51 California coast range angiosperms. *New Phytol.* **170**, 807–818 (2006).
262. Price, C. A. & Enquist, B. J. Scaling mass and morphology in leaves: an extension of the WBE model. *Ecology* **88**, 1132–1141 (2007).
263. Price, C. A., Enquist, B. J. & Savage, V. M. A general model for allometric covariation in botanical form and function. *Proc. Natl Acad. Sci. USA* **104**, 13204–13209 (2007).
264. Pyankov, V. I., Kondratyuk, A. V. & Shipley, B. Leaf structure and specific leaf mass: the alpine desert plants of the Eastern Pamirs, Tadjikistan. *New Phytol.* **143**, 131–142 (1999).
265. Quero, J. L. et al. Relating leaf photosynthetic rate to whole-plant growth: drought and shade effects on seedlings of four *Quercus* species. *Funct. Plant Biol.* **35**, 725 (2008).
266. Quested, H. M. et al. Decomposition of sub-arctic plants with differing nitrogen economies: a functional role for hemiparasites. *Ecology* **84**, 3209–3221 (2003).
267. Reich, P. B., Oleksyn, J. & Wright, I. J. Leaf phosphorus influences the photosynthesis–nitrogen relation: a cross-biome analysis of 314 species. *Oecologia* **160**, 207–212 (2009).
268. Reich, P. B. et al. Scaling of respiration to nitrogen in leaves, stems and roots of higher land plants. *Ecol. Lett.* **11**, 793–801 (2008).
269. Auger, S. & Shipley, B. Inter-specific and intra-specific trait variation along short environmental gradients in an old-growth temperate forest. *J. Veg. Sci.* **24**, 419–428 (2012).
270. Sack, L., Cowan, P. D., Jaikumar, N. & Holbrook, N. M. The ‘hydrology’ of leaves: co-ordination of structure and function in temperate woody species. *Plant Cell Environ.* **26**, 1343–1356 (2003).
271. Sack, L. & Frole, K. Leaf structural diversity is related to hydraulic capacity in tropical rain forest trees. *Ecology* **87**, 483–491 (2006).
272. Sack, L., Melcher, P. J., Liu, W. H., Middleton, E. & Pardee, T. How strong is intracanalopy leaf plasticity in temperate deciduous trees? *Am. J. Bot.* **93**, 829–839 (2006).
273. Sack, L., Tyree, M. T. & Holbrook, N. M. Leaf hydraulic architecture correlates with regeneration irradiance in tropical rainforest trees. *New Phytol.* **167**, 403–413 (2005).
274. Sanda V., Bită-Nicolae, C. D. & Barabas, N. *The Flora of Spontaneous and Cultivated Cormophytes from Romania* (in Romanian) (Editura Ion Bacău, 2003).

275. Sandel, B., Corbin, J. D. & Krupa, M. Using plant functional traits to guide restoration: a case study in California coastal grassland. *Ecosphere* **2**, art23 (2011).
276. Sardans, J., Penuelas, J. & Ogaya, R. Drought-induced changes in C and N stoichiometry in a *Quercus ilex* Mediterranean forest. *For. Sci.* **54**, 513–522 (2008).
277. Sardans, J., Penuelas, J., Prieto, P. & Estiarte, M. Changes in Ca, Fe, Mg, Mo, Na, and S content in a Mediterranean shrubland under warming and drought. *J. Geophys. Res.* <https://doi.org/10.1029/2008jg000795> (2008).
278. Scherer-Lorenzen, M., Schulze, E., Don, A., Schumacher, J. & Weller, E. Exploring the functional significance of forest diversity: a new long-term experiment with temperate tree species (biotree). *Perspect. Plant Ecol. Evol. Syst.* **9**, 53–70 (2007).
279. Schurr, F. M. et al. Colonization and persistence ability explain the extent to which plant species fill their potential range. *Global Ecol. Biogeogr.* **16**, 449–459 (2007).
280. Schwallier, R. et al. Evolution of wood anatomical characters in Nepenthes and close relatives of Caryophyllales. *Ann. Bot.* **119**, 1179–1193 (2017).
281. Schweingruber, F. H., & Poschold, P. Growth rings in herbs and shrubs: life span, age determination and stem anatomy. *Forest Snow Landsc. Res.* **79**, 195–415 (2005).
282. Scoffoni, C., Pou, A., Aasamaa, K. & Sack, L. The rapid light response of leaf hydraulic conductance: new evidence from two experimental methods. *Plant Cell Environ.* **31**, 1803–1812 (2008).
283. Shiodera, S., Rahajoe, J. S. & Kohyama, T. Variation in longevity and traits of leaves among co-occurring understorey plants in a tropical montane forest. *J. Trop. Ecol.* **24**, 121–133 (2008).
284. Shipley, B. The use of above-ground maximum relative growth rate as an accurate predictor of whole-plant maximum relative growth rate. *Funct. Ecol.* **3**, 771 (1989).
285. Shipley, B. Trade-offs between net assimilation rate and specific leaf area in determining relative growth rate: relationship with daily irradiance. *Funct. Ecol.* **16**, 682–689 (2002).
286. Shipley, B. & Lechowicz, M. J. The functional co-ordination of leaf morphology, nitrogen concentration, and gas exchange in 40 wetland species. *Écoscience* **7**, 183–194 (2000).
287. Shipley, B. & Parent, M. Germination responses of 64 wetland species in relation to seed size, minimum time to reproduction and seedling relative growth rate. *Funct. Ecol.* **5**, 111 (1991).
288. Shipley, B. & Vu, T.-T. Dry matter content as a measure of dry matter concentration in plants and their parts. *New Phytol.* **153**, 359–364 (2002).
289. Spasojevic, M. J. & Suding, K. N. Inferring community assembly mechanisms from functional diversity patterns: the importance of multiple assembly processes. *J. Ecol.* **100**, 652–661 (2012).
290. Swaine, E. K. *Ecological and Evolutionary Drivers of Plant Community Assembly in a Bornean Rain Forest*. PhD Thesis (University of Aberdeen, 2007).
291. Trefflich, A., Klotz, S. & Kuhn, I. in *BIOLFLOR—Eine Datenbank mit Biologisch-ökologischen Merkmalen zur Flora von Deutschland* (eds Klotz, S. et al.) 127–131 (Bundesamt für Naturschutz, 2002).
292. Tucker, S. S., Craine, J. M. & Nippert, J. B. Physiological drought tolerance and the structuring of tallgrass prairie assemblages. *Ecosphere* **2**, art48 (2011).
293. Ciocarlan, V. *The Illustrated Flora of Romania. Pteridophyta et Spermatopyta* (in Romanian) (Editura Ceres, 2009).
294. van Bodegom, P. M., Sorrell, B. K., Oosthoek, A., Bakker, C. & Aerts, R. Separating the effects of partial submergence and soil oxygen demand on plant physiology. *Ecology* **89**, 193–204 (2008).
295. Vergutz, L. et al. *A Global Database of Carbon and Nutrient Concentrations of Green and Senesced Leaves* (ORNL DAAC, 2012); <https://doi.org/10.3334/ORNLDAAC/1106>
296. Vergutz, L., Manzoni, S., Porporato, A., Novais, R. F. & Jackson, R. B. Global resorption efficiencies and concentrations of carbon and nutrients in leaves of terrestrial plants. *Ecol. Monogr.* **82**, 205–220 (2012).
297. Vile, D. *Significations Fonctionnelle et Ecologique des Traits des Espèces Vegetales: Exemple dans une Succession Post-culturelle Méditerranéenne et Généralisations*. PhD thesis (University of Montpellier II, 2005).
298. Von Holle, B. & Simberloff, D. Testing Fox's assembly rule: does plant invasion depend on recipient community structure? *Oikos* **105**, 551–563 (2004).
299. Williams, M., Shimabukuro, Y. E. & Rastetter, E. B. *LBA-ECO CD-09 Soil and Vegetation Characteristics, Tapajos National Forest, Brazil* (ORNL DAAC, 2012); <https://doi.org/10.3334/ORNLDAAC/1104>
300. Willis, C. G. et al. Phylogenetic community structure in Minnesota oak savanna is influenced by spatial extent and environmental variation. *Ecography* **33**, 565–577 (2010).
301. Wilson, K. B., Baldocchi, D. D. & Hanson, P. J. Spatial and seasonal variability of photosynthetic parameters and their relationship to leaf nitrogen in a deciduous forest. *Tree Physiol.* **20**, 565–578 (2000).
302. Wirth, C. & Lichstein, J. W. in *Old-Growth Forests: Function, Fate and Value* (eds Wirth, C. et al.) 81–113 (Springer, 2009).
303. Wohlfahrt, G. et al. Inter-specific variation of the biochemical limitation to photosynthesis and related leaf traits of 30 species from mountain grassland ecosystems under different land use. *Plant Cell Environ.* **22**, 1281–1296 (1999).
304. Wright, I. J. et al. Relationships among ecologically important dimensions of plant trait variation in seven neotropical forests. *Ann. Bot.* **99**, 1003–1015 (2007).
305. Wright, J. P. & Sutton-Grier, A. Does the leaf economic spectrum hold within local species pools across varying environmental conditions? *Funct. Ecol.* **26**, 1390–1398 (2012).
306. Wright, S. J. et al. Functional traits and the growth–mortality trade-off in tropical trees. *Ecology* **91**, 3664–3674 (2010).
307. Xu, L. & Baldocchi, D. D. Seasonal trends in photosynthetic parameters and stomatal conductance of blue oak (*Quercus douglasii*) under prolonged summer drought and high temperature. *Tree Physiol.* **23**, 865–877 (2003).
308. Yguel, B. et al. Phytophagy on phylogenetically isolated trees: why hosts should escape their relatives. *Ecol. Lett.* **14**, 1117–1124 (2011).
309. Zanne, A. E. et al. *Global Wood Density Database* (EOL, 2009); <https://opendata.eol.org/dataset/dde86ffb-7741-44a1-acf2-808b3dd6bc97/resource/d1e2b018-a7ce-444b-ac8a-ac43b2355cc9/download/archive>
310. Zanne, A. E. et al. Angiosperm wood structure: global patterns in vessel anatomy and their relation to wood density and potential conductivity. *Am. J. Bot.* **97**, 207–215 (2010).
311. Kattge, V. et al. TRY - a global database of plant traits. *Global Change Biol* **9**, 2905–2935 (2011).
312. Shan, H. et al. *Gap Filling in the Plant Kingdom—Trait Prediction Using Hierarchical Probabilistic Matrix Factorization* (ICML, 2012); <http://arxiv.org/abs/1206.6439>
313. R Core Team. *R: A Language and Environment for Statistical Computing* (R Foundation for Statistical Computing, 2021).
314. Salakhutdinov, R. & Mnih, A. Probabilistic matrix factorization. In *Proc. 20th International Conference on Neural Information Processing Systems* (eds Platt, J. C. et al.) 1257–1264 (Curran Associates Inc., 2007).
315. R Core Team. *R: A Language and Environment for Statistical Computing* (R Foundation for Statistical Computing, 2013).
316. Lê, S., Josse, J. & Husson, F. FactoMineR: a package for multivariate analysis. *J. Stat. Softw.* **25**, 1–18 (2008).
317. Dray, S. & Dufour, A.-B. The ade4 package: implementing the duality diagram for ecologists. *J. Stat. Softw.* **22**, 1–20 (2007).
318. Bougeard, S. & Dray, S. Supervised multiblock analysis in R with the ade4 package. *J. Stat. Softw.* **86**, 1–17 (2018).
319. Chessel, D., Dufour, A.-B. & Thioulouse, J. The ade4 package—I: one-table methods. *R News* **4**, 5–10 (2004).
320. Dray, S., Dufour, A.-B. & Chessel, D. The ade4 package—II: two-table and K-table methods. *R News* **7**, 47–52 (2007).
321. Thioulouse, J. et al. *Multivariate Analysis of Ecological Data with ade4* (Springer, 2018).
322. Friedman, J., Hastie, T. & Tibshirani, R. Regularization paths for generalized linear models via coordinate descent. *J. Stat. Softw.* **33**, 1–22 (2010).
323. Simon, N., Friedman, J., Hastie, T. & Tibshirani, R. Regularization paths for Cox's proportional hazards model via coordinate descent. *J. Stat. Softw.* **39**, 1–13 (2011).
324. Batjes, N. H., Ribeiro, E. & van Oostrum, A. Standardised soil profile data to support global mapping and modelling (WoSIS snapshot 2019). *Earth Syst. Sci. Data* **12**, 299–320 (2020).
325. Hengl, T. et al. SoilGrids1km—global soil information based on automated mapping. *PLoS ONE* **9**, e105992 (2014).
326. Arrouays, D. et al. Soil legacy data rescue via GlobalSoilMap and other international and national initiatives. *GeoRes* **14**, 1–19 (2017).
327. Richard, P. & Pielou, E. C. *Biogeography* (John Wiley & Sons, 1979).
328. Udvardy, M. D. F. *A Classification of the Biogeographical Provinces of the World* (International Union for Conservation of Nature and Natural Resources, 1975).
329. Dinerstein, E. et al. *A Conservation Assessment of the Terrestrial Ecoregions of Latin America and the Caribbean* (The World Bank, 1995).
330. Ricketts, T. H. et al. *Terrestrial Ecoregions of North America: A Conservation Assessment* (Island Press, 1999).
331. Dasmann, R. F. *A System for Defining and Classifying Natural Regions for Purposes of Conservation: A Progress Report* (IUCN, 1973).

## Acknowledgements

The study was supported by the TRY initiative on plant traits (<http://www.try-db.org>). The TRY database is hosted at the Max Planck Institute for Biogeochemistry (MPI BGC, Germany) and supported by Future Earth and the German Centre for Integrative Biodiversity Research (iDiv) Halle-Jena-Leipzig. We would like to thank all PIs contributing to the TRY database, whose efforts allowed this analysis. In detail, we thank: J.H.C. Cornelissen, R. Milla, W. Cornwell, K. Kramer, S. Gachet, Ingolf Kühn, P. Poschold, M. Scherer, J. Pausas, B. Sandal, K. Verheyen, J. Penuelas, N. Soudzilovskaia,

P. Reich, J. Fang, S. Harrison, R. Gallagher, B. Hawkins, B. Finegan, J. Powers, F. Lenti, S. Higgins, B. Medlyn, H. Ford, V. Pillar, M. Bahn, E. Sosinski, T. He, B. Cerabolini, J. Cavender-Bares, I. J. Wright, F. Louault, B. Amiaud, G. Gonzalez-Melo, P. Adler, F. Schurr, J. Craine, Y. Niinemets, A. Zanne, H. Jactel, M. Harze, R. Montgomery, C. Römermann, T. Hickler, A. Pahl, M. Dainese, D. Kirkup, J. Dickie, W. Hattingh, P. Higuchi, T. Domingues, A. Araujo, M. Williams, C. Price, B. Shipley, L. Sack, B. Schamp, W. Han, Y. Onoda, K. Fleischer, J.P. Wright, G. Guerin, F. de Vries, D.D. Baldocchi, J. Kattge, B. Blonder, K. Brown, D. Campetella, G. Frechet, Q. Read, N. G. Swenson, V. Lanta, E. Weiher, M. Leishman, A. Siefert, M. Spasojevic, R. Jackson, J. Messier, S. J. Wright, D. Craven, J. Molofsky, P. Meir, E. Forey, A. Totte, C. Frenette Dussault, O. Atkin, F. Koike, D. Laughlin, S. Burrascano, K. Ollerer, N. Gross, A. Madhur, P. Begonna, B. Bond-Lamberty, B. von Holle, W. Green, B. Yguel, A. C. Malhado, P. Manning, G. Zott, E. Lamb, J. Fagundes, Z. Wang, S. Diaz, C. Byun, W. Bond, B. Enquist, C. Baraloto, P. Manning, M. Kleyer, W. Ozinga, J. Ordóñez, J. Lloyd, H. Poorter, E. Garnier, F. Valladares, C. Pladevall, G. Freschet, M. Moretti, H. Kurokawa, V. Minden, A. Demey, F. Fernandez-Méndez, J. Butterfield, T. Domingu, E. Swaine, L. Poorter, S. Shiodera, T. Chapin, M. Beckmann, J.A. Gutierrez, M. Mencuccini, S. Jansen, and N. J. B. Kraft. We appreciate the discussions at the MPI BGC. We thank F. Fazayeli for preparing the gap-filled trait data. We thank F. Gans and U. Weber for preparing ancillary data and B. Ahrens for pointing out some soil data availability. We acknowledge Environmental Systems Research Institute (ESRI) and its licensor(s) for the Geodata product of the Missions Database 'ArcWorld Supplement' (GMI), published by Global Mapping International and originated from Global Mapping International for producing Extended Data Fig. 1 and Supplementary Fig. 7 and available in ArcGIS software by ESRI. ArcGIS and ArcMap are the intellectual property of ESRI and are used herein under license. For more information about ESRI software, please visit [www.esri.com](http://www.esri.com). The authors affiliated with the MPI BGC acknowledge funding by the European Union's Horizon 2020 project BACI under grant agreement no. 640176. We are thankful to the data providers for the SoilGrids, hosted by ISRIC. J.S.J. acknowledges the International Max Planck Research School for global biogeochemical cycles, J.S.J., M.E.S. and M.C.S. acknowledge support from the University of Zurich University Research Priority Program on Global Change and Biodiversity. P.B.R., M.E.S. and M.C.S. acknowledge membership in the US NSF 20-508 BII-Implementation project, 'The causes and consequences of plant biodiversity across scales in a rapidly changing world'. M.E.S. acknowledges the NOMIS grant of Remotely Sensing Ecological Genomics that funds J.S.J. and M.C.S. C.W. acknowledges the support of the Max Planck Society via its fellowship programme. N.R. was funded by a research grant from Deutsche Forschungsgemeinschaft DFG (RU 1536/3-1). K.K. was supported by the project Resilient Forests (KB-29-009-003) of the Dutch Ministry of Economic Affairs. The trait data supplied were co-funded by the EU-FP7-KBBE project: BACCARA—Biodiversity and climate change, a risk analysis (project ID 226299). I.W. acknowledges support from the Australian Research Council (DP170103410). J.P. acknowledges financial support from the European Research Council Synergy grant ERC-SyG-2013-610028 IMBALANCE-P. N.A.S. is financed by a VIDI grant (016.161.318) issued by the Netherlands Organization of Scientific Research. The data V.M. provided were funded by II. Oldenburgischer Deichband and the Wasserverbandstag e.V. (NWS 10/05). We thank M. Kleyer for his critical input. P.H. and V.D.P. have been supported by CNPq (grant nos 369617/2017-2 and 307689/2014-0, respectively). C.B. was supported by the National Research Foundation of Korea (NRF) grant funded by the Korea government (MSIT) (2018R1C1B6005351). A.G.G. was

funded by FONDECYT grant nos 11150835 and 1200468. V.O. thanks Russian science foundation (RSF, 19-14-00038) for financial support.

## Author contributions

B.R., C.W., J.K., J.S.J. and M.D.M. were responsible for conceptualization. B.R., C.W., J.S.J. and M.D.M. developed the methodology. J.S.J. undertook the formal analysis, validation, visualization, investigation and project administration. B.R., C.R., C.W., F.L., F.S., I.W., J.H.C.C., J.K., J.S.J., K.K., M.D.M., M.R., N.R., P.B.R., P.v.B., R.R., S.D., S.D.S., Ü.N. and W.N.H. wrote the original draft. J.K. and J.S.J. carried out data curation. A.G.M., A.G.G., B.E.L.C., C.B., C.R., C.W., E.S., E.R.W., F.L., G.C., H.J., I.W., J.M.C., J.H.C.C., J.P., K.K., M.A., M.B., N.K., N.A.S., P.H., P.B.R., P.v.B., S.D., T.H., Ü.N., V.M., V.O., V.D.P. and W.N.H. obtained resources. A.G.M., A.G.G., B.E.L.C., B.R., C.B., C.R., C.W., E.S., E.R.W., F.L., F.S., G.C., H.J., I.W., J.M.C., J.S.J., J.H.C.C., J.P., K.K., M.A., M.B., M.C.S., M.D.M., M.R., M.E.S., N.K., N.R., N.A.S., P.H., P.B.R., P.v.B., S.D., S.D.S., T.H., Ü.N., V.M., V.O., V.D.P. and W.N.H. wrote, reviewed and edited the manuscript. B.R., C.W., J.K., M.C.S., M.D.M., M.E.S. and N.R. undertook supervision. C.W., J.K., M.R. and M.E.S. were involved in project administration and funding. B.R., J.S.J., S.D.S. and R.R. ran the software.

## Competing interests

The authors declare no competing interests.

## Additional information

**Extended data** is available for this paper at <https://doi.org/10.1038/s41559-021-01616-8>.

**Supplementary information** The online version contains supplementary material available at <https://doi.org/10.1038/s41559-021-01616-8>.

**Correspondence and requests for materials** should be addressed to Julia S. Joswig.

**Peer review information** *Nature Ecology & Evolution* thanks Haydn J.D. Thomas, Vanessa Buzzard and the other, anonymous, reviewer(s) for their contribution to the peer review of this work. Peer reviewer reports are available.

**Reprints and permissions information** is available at [www.nature.com/reprints](http://www.nature.com/reprints).

**Publisher's note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

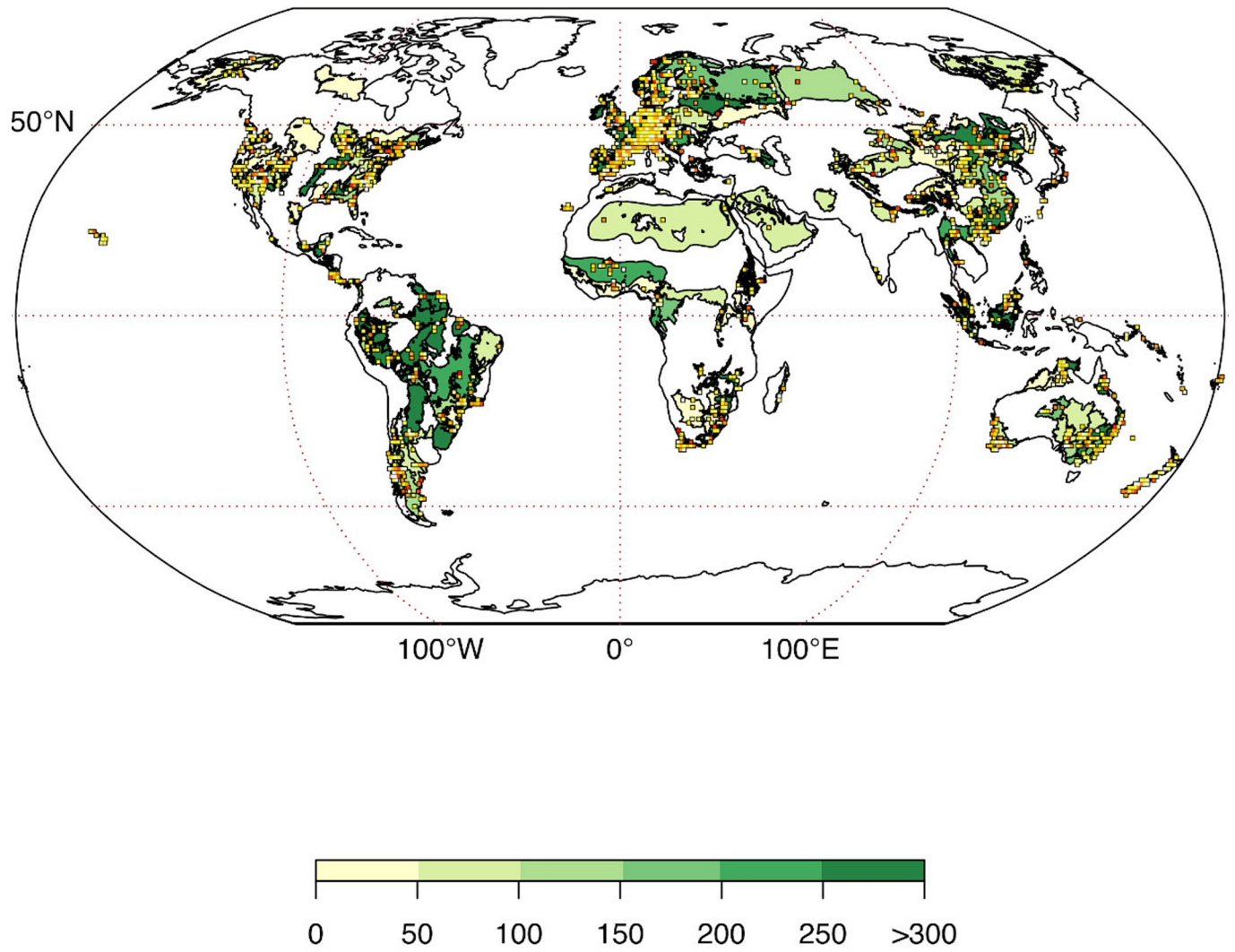


**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2021

<sup>1</sup>Max-Planck-Institute for Biogeochemistry, Jena, Germany. <sup>2</sup>Remote Sensing Laboratories, Department of Geography, University of Zurich, Zurich, Switzerland. <sup>3</sup>German Centre for Integrative Biodiversity Research (iDiv), Leipzig, Germany. <sup>4</sup>Institute of Systematic Botany and Functional Biodiversity, University of Leipzig, Leipzig, Germany. <sup>5</sup>Department of Chemistry, University of Zurich, Zurich, Switzerland. <sup>6</sup>Escuela de Biología, Universidad Industrial de Santander, Bucaramanga, Colombia. <sup>7</sup>Department of Biological Sciences, Macquarie University, Sydney, New South Wales, Australia. <sup>8</sup>Institute for Atmospheric and Climate Science, ETH Zurich, Zurich, Switzerland. <sup>9</sup>Norwegian Institute of Bioeconomy Research, Oslo, Norway. <sup>10</sup>Department of Economics, University of Leipzig, Leipzig, Germany. <sup>11</sup>Smithsonian Tropical Research Institute, Panamá, Panamá. <sup>12</sup>Geoinformatics and Remote Sensing, Institute for Geography, University of Leipzig, Leipzig, Germany. <sup>13</sup>Environmental Biology Department, Institute of Environmental Sciences, CML, Leiden University, Leiden, the Netherlands. <sup>14</sup>Systems Ecology, Department of Ecological Science, Faculty of Science, Vrije Universiteit Amsterdam, Amsterdam, the Netherlands. <sup>15</sup>Instituto Multidisciplinario de Biología Vegetal (IMBIV), CONICET and FCEfyN, Universidad Nacional de Córdoba, Córdoba, Argentina. <sup>16</sup>Global Systems and Analytics, Nova Pioneer, Johannesburg, South Africa. <sup>17</sup>Chairgroup Forest Ecology and Forest Management, Wageningen University, Wageningen, the Netherlands. <sup>18</sup>Land Life Company, Amsterdam, the Netherlands. <sup>19</sup>Research Group Functional Traits, Naturalis Biodiversity Center, Leiden, the Netherlands. <sup>20</sup>Plant Sciences, Institute of Biology Leiden, Leiden University, Leiden, the Netherlands. <sup>21</sup>Estonian University of Life Sciences, Tartu, Estonia. <sup>22</sup>Department of Forest Resources, University of Minnesota, St Paul, MN, USA. <sup>23</sup>Hawkesbury Institute for the Environment, Western Sydney University, Penrith, New South Wales, Australia. <sup>24</sup>Institute for Global Change Biology and School for Environment and Sustainability, University of Michigan, Ann Arbor, MI, USA. <sup>25</sup>Department of Plant Biodiversity, Institute of Ecology and Evolution, Friedrich-Schiller University, Jena, Germany. <sup>26</sup>School of Geography, University of Nottingham, Nottingham, UK. <sup>27</sup>School of Environmental Sciences, University of Guelph, Guelph, Canada. <sup>28</sup>Department of Ecology, University of Innsbruck, Innsbruck, Austria. <sup>29</sup>Department of Biological Sciences and Biotechnology, Andong National University, Andong, Korea. <sup>30</sup>Plant Diversity and Ecosystems Management Unit, School of Biosciences and Veterinary Medicine, University of Camerino, Camerino, Italy. <sup>31</sup>Department of Biotechnologies and Life Sciences (DBSV), University of Insubria, Varese, Italy. <sup>32</sup>Jonah Ventures LLC, Boulder, CO, USA. <sup>33</sup>Facultad de Ciencias Naturales y Matemáticas, Universidad del Rosario, Bogotá, Colombia. <sup>34</sup>Departamento de Ciencias Ambientales y Recursos Naturales Renovables, Facultad de Ciencias Agronómicas, Universidad de Chile, Santiago, Chile. <sup>35</sup>School of Molecular and Life Sciences, Curtin University, Perth, Western Australia, Australia. <sup>36</sup>College of Science, Health, Engineering and Education, Murdoch University, Murdoch, Western Australia, Australia. <sup>37</sup>Department of Forestry, Universidade do Estado de Santa Catarina, Lages, Brazil. <sup>38</sup>INRAE University Bordeaux, BIOGECO, Cestas, France. <sup>39</sup>Department of Ecology and Evolutionary Biology, University of California, Los Angeles, CA, USA. <sup>40</sup>Department of Biology, Vrije Universiteit Brussel, Brussels, Belgium. <sup>41</sup>Landscape

Ecology Group, Institute of Biology and Environmental Sciences, University of Oldenburg, Oldenburg, Germany. <sup>42</sup>Department of Ecology and Plant Geography, Moscow State Lomonosov University, Moscow, Russia. <sup>43</sup>CSIC, Global Ecology Unit CREAL-CSIC-UAB, Bellaterra, Spain. <sup>44</sup>CREAF, Cerdanyola del Vallés, Spain. <sup>45</sup>Department of Ecology, Universidade Federal do Rio Grande do Sul, Porto Alegre, Brazil. <sup>46</sup>Embrapa Recursos Genéticos e Biotecnologia, Brasília, Brazil. <sup>47</sup>Centre for Environmental Sciences, Hasselt University, Diepenbeek, Belgium. <sup>48</sup>Institute of Environmental Sciences, Leiden University, Leiden, the Netherlands. <sup>49</sup>Department of Biology, University of Wisconsin, Eau Claire, WI, USA. <sup>50</sup>Remote Sensing Centre for Earth System Research, University of Leipzig, Leipzig, Germany. <sup>51</sup>Helmholtz Centre for Environmental Research, Leipzig, Germany. ✉e-mail: [juliajoswigjj@gmail.com](mailto:juliajoswigjj@gmail.com)



**Extended Data Fig. 1 | Map of Ecoregion data.** Map of ecoregions (30) included in this study (n=220).



## Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our [Editorial Policies](#) and the [Editorial Policy Checklist](#).

### Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a Confirmed

- The exact sample size ( $n$ ) for each experimental group/condition, given as a discrete number and unit of measurement
- A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
- The statistical test(s) used AND whether they are one- or two-sided  
*Only common tests should be described solely by name; describe more complex techniques in the Methods section.*
- A description of all covariates tested
- A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
- A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
- For null hypothesis testing, the test statistic (e.g.  $F$ ,  $t$ ,  $r$ ) with confidence intervals, effect sizes, degrees of freedom and  $P$  value noted  
*Give  $P$  values as exact values whenever suitable.*
- For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
- For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
- Estimates of effect sizes (e.g. Cohen's  $d$ , Pearson's  $r$ ), indicating how they were calculated

*Our web collection on [statistics for biologists](#) contains articles on many of the points above.*

### Software and code

Policy information about [availability of computer code](#)

Data collection

Trait data: TRY ([www.try-db.org](http://www.try-db.org), accession date July 2017, request nb.3282)  
 Climate data: WorldClim is publicly available via <https://www.worldclim.org/> (accession date: May 2018),  
 Soil data: SoilGrids ([soilgrids.org](http://soilgrids.org), accession date: June 2018) is publicly available  
 Ecoregions: of Olson et al. 2001 Shapefiles are publicly available (accession date January 2014, <https://www.sciencebase.gov/catalog/item/508fece8e4b0a1b43c29ca22>)  
 Estimate of species richness per ecoregion: Kier species richness (accession date January 2014, <https://databasin.org/datasets/43478f840ac84173979b22631c2ed672/>)  
 Spatial data (licenced, see third party rights form)

Data analysis

R, R-Studio, Microsoft Excel

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio [guidelines for submitting code & software](#) for further information.

## Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our [policy](#)

Plant trait data were accessed from the TRY data base ([try-db.org](http://try-db.org), request number: 3282, accession date: July 2017, see also Figure \ref{fig\_extDat}). All TRY data required to reproduce this analysis, and the corresponding R-scripts, are provided in an open TRY File Archive (<https://www.try-db.org/TryWeb/Data.php>). Climate data WorldClim is publicly available via <https://www.worldclim.org/> (accession date: May 2018), Soil data, namely SoilGrids ([soilgrids.org](http://soilgrids.org), accession date: June 2018) is publicly available. Ecoregion information \cite{Olson2001} shapefiles are publicly available (accession date January 2014, <https://www.sciencebase.gov/catalog/item/508fece8e4b0a1b43c29ca22>), The estimate of species richness per ecoregion (Kier et al. 2005) is publicly available (accession date January 2014, <https://databasin.org/datasets/43478f840ac84173979b22631c2ed672/>). Data for this study can be accessed on Github (<https://github.com/juliajoswig/RepoClimateSoilTraitSpectrum>).

For the extended data figure 1, Supplementary Fig.7, the Geodata product of the Missions Database “ArcWorld Supplement” (GMI) was used, published by Global Mapping International and originated from Global Mapping International for producing and available in ArcGIS software by Esri. ArcGIS and ArcMapTM are the intellectual property of Esri and are used herein under license. Copyright Esri. All rights reserved. For more information about Esri software, please visit [www.esri.com](http://www.esri.com).

## Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

- Life sciences       Behavioural & social sciences       Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see [nature.com/documents/nr-reporting-summary-flat.pdf](https://nature.com/documents/nr-reporting-summary-flat.pdf)

## Ecological, evolutionary & environmental sciences study design

All studies must disclose on these points even when the disclosure is negative.

### Study description

The overarching objective of this study is to test whether the major dimensions underpinning the global spectrum of plant variation, such as the size and leaf economics spectra, are the result of joint and/or independent variation of climate and soil. For this study, we compiled and analyze a dataset of 17 functional traits for 225,206 geo-referenced observations comprising records of 20,655 global unique species (36,197 unique species to ecoregion combinations) covering the main worlds’ ecoregions – environmentally homogeneous areas characterized by distinct plant assemblages (Methods, Extended Data Figure 1). The trait data were complemented with 21 climate variables (Supplementary Tab. 1) as well as 107 soil variables (Supplementary Tab. 2). Trait-environment relationships were analyzed on the basis of ecoregions using a combination of a regression technique and hierarchical partitioning (for details see Methods).

### Research sample

We extracted data of 17 plant functional traits from the TRY data base1 (Supplementary Tab. 5, Supplementary Tab. 6, [www.try-db.org](http://www.try-db.org), accession date July 2017, request nb.3282) including published literature (see manuscript). To be able to include the maximum number of species in our analyses, we used the gap-filled version of TRY, with missing data were imputed using a Bayesian Hierarchical Probabilistic Matrix Factorization (BHPMF) algorithm, but observed values were kept. We excluded observations that were not geo-referenced. In total, we included 225,206 observations from 20,655 global unique species (36,197 unique species to ecoregion combinations). The data were stratified by ecoregions (see below), aggregated to species median values per ecoregion and log-transformed.

To represent climate conditions we used 21 variables derived from the WorldClim reanalysis product at a resolution of 1 km (accession date May 2018). We extracted values for temperature (annual average, diurnal range, max of warmest month, min of coldest month, sd), precipitation (annual average, min, max, sd), vapour pressure (annual average, min, max, sd), solar radiation (annual average, min, max, sd) and wind (annual average, min, max, sd; see Supplementary Tab. 1).

To characterise soil conditions we used 107 variables derived from the ISRIC data product ‘SoilGrids’ ([soilgrids.org](http://soilgrids.org) through ISRIC - WDC Soils, Supplementary Tab. 2). ‘SoilGrids’ provides global predictions of soil characteristics for seven depths, i.e. 0, 5, 15, 30, 60, 100, 200 cm at a resolution of 1 km.

In total, we included 128 environmental variables. Before performing the ridge regression, climate and soil variables were reduced to 20 variables each by means of a principle component analysis (PCA).

### Sampling strategy

The data were stratified by ecoregions, aggregated to species median values per ecoregion and log-transformed. To determine trait-environment relationships, we aggregated trait as well as environmental data to ecoregions (Supplementary Tab.7) Ecoregions are environmentally homogeneous areas characterized by distinct plant assemblages. At global scale Olson et al. defined 867 ecoregions. For each of the 867 ecoregions, we calculated the median of all species median trait values. For further analyses we only included regions with >20 species and a representation of >1% of the estimated species richness of the ecoregion.

To aggregate environmental variables to ecoregion level, we associated each geo-referenced trait observation with its corresponding values of climate and soil variables. Then, we averaged over all values within one ecoregion. Thus, the selected environmental

variables represent averages that are weighted by the number and location of trait observations within ecoregions.

**Data collection** Trait data was collected in the framework of TRY (Kattge et al. 2011).

**Timing and spatial scale** Trait-data: geo-referenced global point data  
Climate- data: gridded data (1km resolution)  
Soil: gridded data of up to 7 different depths (1km resolution)

**Data exclusions** For trait observations, we only included geo-referenced data.  
For environmental data, we only included the grid-level information if a trait data point was located there.  
For ecoregions we only included regions with >20 species and a representation of >1% of the estimated species richness of the ecoregion.

**Reproducibility** All attempts to repeat the results were successful.  
Per default, the ridge regression was repeated 50 times for each trait. The mean, minimum and maximum of r2 and independent effect give an estimate of model spreads (Table 1, Supplementary Tab. 9).  
All TRY data required to reproduce this analysis, and the corresponding R-scripts, are provided in an open TRY File Archive (<https://www.try-db.org/TryWeb/Data.php>).

**Randomization** To estimate how much of the ecoregion trait variation (r2) is explained by random environmental variables, we performed the ridge regression followed by hierarchical partitioning (see methods „ridge regression“ and “hierarchical partitioning”) and paired climate or soil with noise. The noise data set comprised randomly sampled values for a variable set, as large as the soil variable set (n=107). We performed ridge regression analysis with noise data, together with climate or soil. Then we calculated the independent effect of noise from climate or soil data.  
The independent effect of randomized data (noise) is never above 0 (negative values due to large difference between r2\_total and r2\_noise and model variability; Supplement Fig 2).  
Moreover, we sampled for each ecoregion the minimum requirement for selection (Kier species richness of 1% and 20 species) and erased randomly (n=3) the rest of the species observations. See Supplement Fig. 2.

**Blinding** *Describe the extent of blinding used during data acquisition and analysis. If blinding was not possible, describe why OR explain why blinding was not relevant to your study.*

Did the study involve field work?  Yes  No

## Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

### Materials & experimental systems

### Methods

- | n/a                                 | Involved in the study                                  |
|-------------------------------------|--|
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Antibodies                    |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Eukaryotic cell lines         |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Palaeontology and archaeology |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Animals and other organisms   |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Human research participants   |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Clinical data                 |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Dual use research of concern  |

- | n/a                                 | Involved in the study                           |
|-------------------------------------|---|
| <input checked="" type="checkbox"/> | <input type="checkbox"/> ChIP-seq               |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Flow cytometry         |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> MRI-based neuroimaging |