



Published in final edited form as:

Neurobiol Aging. 2022 March ; 111: 95–106. doi:10.1016/j.neurobiolaging.2021.10.011.

Association between *WWOX/MAF* variants and dementia-related neuropathologic endophenotypes

Adam J. Dugan, Ph.D.^a, Peter T. Nelson, M.D., Ph.D.^{b,c}, Yuriko Katsumata, Ph.D.^{a,b}, Lincoln M. P. Shade, B.S.^a, Merilee A. Teylan, M.P.H.^d, Kevin L. Boehme, B.S.^e, Shubhabrata Mukherjee, Ph.D.^f, John S. K. Kauwe, Ph.D.^{e,f}, Timothy J. Hohman, Ph.D.^g, Julie A. Schneider, M.D., M.S.^h, Alzheimer's Disease Genetics Consortium, David W. Fardo, Ph.D.^{a,b,*}

^aDepartment of Biostatistics, College of Public Health, University of Kentucky, Lexington, KY, 40536, USA

^bSanders-Brown Center on Aging and Alzheimer's Disease Research Center, University of Kentucky, Lexington, KY, 40504, USA

^cPathology and Laboratory Medicine, University of Kentucky, Lexington, KY, USA

^dNational Alzheimer's Coordinating Center, Department of Epidemiology, University of Washington, Seattle, WA, 98105, USA

^eDepartment of Biology, Brigham Young University, Provo, UT 84602, USA

^fDepartment of Medicine, University of Washington, Seattle, WA 98104, USA

^gVanderbilt Memory & Alzheimer's Center, Department of Neurology, Vanderbilt University Medical Center, Nashville, TN, 37212, USA

^hDepartments of Neurology and Pathology, Rush University Medical Center, Chicago, IL, 60612, USA

Abstract

The genetic locus containing the *WWOX* and *MAF* genes was implicated as a clinical Alzheimer's disease (AD) risk locus in two recent large meta-analytic genome wide association studies (GWAS). In a prior GWAS, we identified a variant in *WWOX* as a suggestive risk

* **Corresponding Author:** David W. Fardo, Ph.D., 760 Press Avenue Healthy Kentucky Research Building 0679, Room 372, Lexington KY 40536-0679, david.fardo@uky.edu.

Adam J. Dugan, Ph.D., Conceptualization; Data curation; Formal analysis; Visualization; Writing - original draft; Writing - review & editing, Peter T. Nelson, M.D., Ph.D., Conceptualization; Investigation; Supervision; Writing - original draft; Writing - review & editing, Yuriko Katsumata, Ph.D., Data curation; Methodology; Software; Writing - review & editing, Lincoln M. P. Shade, B.S., Data curation; Software; Writing - review & editing, Merilee A. Teylan, M.P.H., Writing - review & editing, Kevin L. Boehme, B.S., Data curation; Writing - review & editing, Shubhabrata Mukherjee, Ph.D., Data curation; Writing - review & editing, John S. K. Kauwe, Ph.D., Data curation; Writing - review & editing, Timothy J. Hohman, Ph.D., Data curation; Writing - review & editing, Julie A. Schneider, M.D., M.S., Resources; Writing - review & editing, Alzheimer's Disease Genetics Consortium, Resources; Writing - review & editing, David W. Fardo, Ph.D., Conceptualization; Funding acquisition; Project administration; Resources; Supervision; Writing - original draft; Writing - review & editing

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

allele for hippocampal sclerosis (HS). We hypothesized that the *WWOX/MAF* locus may be preferentially associated with non-plaque- and non-tau-related neuropathological changes (NC). Data from research participants with GWAS and autopsy measures from the National Alzheimer's Coordinating Center (NACC) and the Religious Orders Study and the Rush Memory and Aging Project (ROSMAP) were meta-analyzed. Notably, no variants in the locus were significantly associated with ADNC. However, several *WWOX/MAF* variants had significant adjusted associations with limbic-predominant age-related TDP-43 encephalopathy NC (LATE-NC), HS, and brain arteriolosclerosis. These associations remained largely unchanged after adjustment for ADNC (operationalized with standard semiquantitative staging), suggesting that these associations are independent of ADNC. Thus, *WWOX* genetic variants associated with clinical AD-type dementia phenotype were associated pathologically with LATE-NC related brain changes, not ADNC.

Keywords

ADRD; Human; Hippocampus; ADNC; SNV; SNP

Introduction

The human WW domain-containing oxidoreductase (*WWOX*) and MAF bZIP transcription factor (*MAF*) genes are situated close to each other on chromosome 16q23. The normal functions of these genes are incompletely characterized. *WWOX* protein plays roles in transcription regulation, glucose metabolism, and central nervous system development (Kola et al. 2019), while the protein encoded by the *MAF* gene is a transcription factor that regulates cellular processes including T-cell susceptibility to apoptosis.

This locus has been implicated in human disease. *WWOX* has been hypothesized to play a role in neurodegenerative disease, particularly Alzheimer's disease (AD) (Chang et al. 2014; Dourlen et al. 2019; Sze et al. 2004). A large genome-wide association study (GWAS) of clinical AD suggested that *WWOX* confers AD risk in non-Hispanic White individuals (Kunkle et al. 2019) and a follow-up GWAS in African American individuals nominally replicated this association (Kunkle et al. 2020). More recently, the largest AD GWAS to date also found an association between AD and *MAF*, the gene just downstream of *WWOX* (Bellenguez et al. 2020). In addition to AD, *WWOX* has also shown suggestive linkage with autism and schizophrenia (Bacchelli et al. 2020; McClay et al. 2011) and *MAF* has been associated with thyroid-related diseases, such as Graves' disease and Hashimoto's disease (Campbell et al. 2016). Nonetheless, the neurochemistry of *WWOX* and *MAF* in the human brain, and in human disease, is still poorly understood.

It has become increasingly clear that AD and AD-related dementias (AD-ADRD) are highly complex at both the individual level (multiple pathologies per person) and in a population (many different combinations of mixed pathologies). Thus, multiple neuropathological changes are associated with the AD clinical syndrome and these neurodegenerative diseases often co-occur, especially in older age (Rahimi and Kovacs 2014; Nelson et al. 2019). A recent community-based cohort study looking at the prevalence of multiple proteinopathies

in older adults found that all individuals had the presence of at least one of tau, amyloid- β (A β), α -synuclein, or TAR-DNA binding protein 43 (TDP-43) pathologies at autopsy and only 6.4% of individuals presented with only a single proteinopathy (Karanth et al. 2020).

A previous GWAS found *WWOX* to be a gene suggestive for association with hippocampal sclerosis (HS) pathology (Nelson et al. 2014). The brain conditions which were previously referred to as “HS-Aging” and “HS dementia” are now subsumed under a broader disease category, and are characterized by the presence of comorbid TDP-43 proteinopathy, which is a more sensitive and specific feature. The condition was recently classified with the term limbic-predominant age-related TDP-43 encephalopathy (LATE) (Nelson et al. 2019). The presence of the neuropathological changes underlying LATE (LATE-NC) is associated with a dementia syndrome similar to AD (Nelson et al. 2019).

Given that we had found a suggestive link between *WWOX* variants and HS, and others found an association between the *WWOX/MAF* locus and clinical AD, we hypothesized that a more definitive conclusion could be reached via a pathology-based study of separate cohorts with both genetic and pathologic information (including TDP-43 proteinopathy) available. We investigated whether the *WWOX/MAF*AD association could be due to neuropathological changes other than AD-type pathological hallmarks, amyloid plaques and neurofibrillary tangles. GWAS data and autopsy-confirmed neuropathological endophenotypes were gathered from the National Alzheimer’s Coordinating Center (NACC) and from the Religious Orders Study and the Rush Memory and Aging Project (ROSMAP), to resolve novel associations between pathological findings and *WWOX/MAF* genetic variation.

Material and Methods

Study Participants

Phenotypic data from NACC (March 2021 data freeze) were linked with genotype data from the Alzheimer’s Disease Genetics Consortium (ADGC). Individuals who died at age 65 years or older were included in this study. Similar to other studies using NACC data (Katsumata et al. 2020; Dugan et al. 2021), individuals were excluded from the NACC cohort if at least one of 19 rare brain diseases were diagnosed (Supplemental Table 1) or if they were missing any adjustment variables or all of the endophenotypes under study.

The ROSMAP study has been described in detail elsewhere (Mahoney et al. 2019). Briefly, data were acquired from two well-characterized cohort studies of aging and dementia. The Religious Orders Study (ROS), begun in 1994, and the Rush Memory and Aging Project (MAP), begun in 1997, involve older adults who enrolled without dementia, agreed to annual clinical evaluations and organ donation at death, and signed an Anatomical Gift Act for brain donation. Written informed consent was obtained from participants, and research was carried out in accordance with Institutional Review Board (IRB)-approved protocols. ROSMAP data are available online at the Rush Alzheimer’s Disease Center Resource Sharing Hub (<https://www.radc.rush.edu/>), as well as on the Accelerating Medicines Partnership-Alzheimer’s Disease (AMP-AD) Knowledge Portal (syn3219045).

Neuropathological Endophenotype Definitions

In the NACC Neuropathology (NP) dataset, LATE-NC was defined as either present or absent using the “distribution of TDP-43 immunoreactive inclusions” variables indicating if TDP-43 proteinopathy was observed in either the hippocampus (NPTDPC NACC field), entorhinal/inferior temporal cortex (NPTDPD), or neocortex (NPTDPE) in a case lacking overall diagnosis of frontotemporal lobar degeneration (FTLD)-TDP. HS was defined as either present or absent based on the “hippocampal sclerosis of CA1 and/or subiculum” (NPHIPSCL) variable using the “unilateral,” “bilateral,” and “present but laterality not assessed” response categories. Arteriolosclerosis was defined similarly using the “arteriolosclerosis” (NACCARTE) variable and collapsing the “moderate” and “severe” response categories. Presence of neurofibrillary tangles was also defined dichotomously using the “Braak stage for neurofibrillary degeneration (B score)” (NACCBRAA) variable and collapsing the “stage V (B3)” and “stage VI (B3)” response categories. Presence of neuritic plaques was defined dichotomously using the “frequent neuritic plaques (C3)” response category of the “density of neocortical neuritic plaques (CERAD score) (C score)” (NACCNEUR) variable.

In ROSMAP, LATE-NC was defined dichotomously using the “TDP-43 stage” (tdp_st4) variable and collapsing the 2nd and 3rd stages in cases lacking FTLD-TDP. HS was defined dichotomously by the “hippocampal sclerosis was rated as definitely present with CA1 region affected” response category of the “definite presence of typical hippocampal sclerosis” (hspath_typ) variable. Arteriolosclerosis was defined dichotomously using the “arteriolosclerosis” (arteriol_scler) variable and collapsing the “moderate” and “severe” response categories. Presence of neurofibrillary tangles was defined dichotomously using the “semiquantitative measure of neurofibrillary tangles” (braaksc) variable and collapsing the “V” and “VI” response categories. Presence of neuritic plaques was defined dichotomously by the “definite” response category of the “semiquantitative measure of neuritic plaques” (ceradsc) variable.

Quality Control of Genotype Data

For NACC participants, genomic data from the ADGC imputed using the Haplotype Reference Consortium (ADGC-HRC) were used (McCarthy et al. 2016). The genetic data for ROSMAP was also imputed using the HRC and the methods have been described in detail elsewhere (Dumitrescu et al. 2020). Standard GWAS quality control (QC) procedures were performed separately on the ADGC and ROSMAP genotype data using PLINK1.9 (Marees et al. 2018; Purcell et al. 2007). Variants were excluded if they were missing in more than 5% of samples, if they had a minor allele frequency less than 1%, or if they had Hardy-Weinberg Equilibrium (HWE) p-values $< 1 \times 10^{-6}$ among AD controls. Individuals were excluded if they were missing more than 5% of genotypes. Two individuals were considered related if they had an identity by descent measure of at least 0.25, which indicates that they are second-degree relatives. For related pairs, the individual with the lowest call rate was excluded.

NACC and ROSMAP genotype data were separately merged with 1000 Genomes data Phase 3. Principal components (PCs) were calculated for the merged data sets using the

“pca” procedure in PLINK1.9, and the first two PCs were plotted. The ADGC-HRC and ROSMAP individuals with first and second PCs that overlapped with those of the 1000 Genomes individuals of known European ancestry were identified and all other individuals were excluded from the analysis.

Variant-Level Associations

All statistical analyses were conducted in R programming language, version 4.1.1 (R Core Team 2021). The gene boundaries of *WWOX* and *MAF* were defined based on the canonical transcripts (*WWOX*: 78,133,309 – 79,246,564; *MAF*: 79,627,744 – 79,634,622) using the GRCh37/hg19 gene range list from PLINK (<https://www.cog-genomics.org/plink/1.9/resources>). Since both of the genome-wide significant *WWOX/MAF*AD variants (rs62039712 from Kunkle *et al.* and rs450674 from Bellenguez *et al.*) were in the intergenic region between the *WWOX* and *MAF* genes, we defined the *WWOX/MAF* locus to be from 78,133,309 to 79,634,622 \pm 250kb of flanking.

Associations between each endophenotype and each variant were tested separately in the NACC and ROSMAP datasets using binary logistic regression models assuming each of the three most commonly used modes of inheritance (MOI): additive, dominant, and recessive. Variants were excluded from the analyses if they were multiallelic, if there were fewer than 15 minor alleles present across all participants, or if the logistic regression analysis resulted in complete or quasi-complete separation. All regression models adjusted for age at death, sex, ADGC data selection round (for NACC data) or ROS/MAF study (for ROSMAP data), and the first three genetic PCs. Odds ratios (OR) were calculated for each variant by exponentiating the variant’s beta estimate. Since some endophenotypes were only available in a subset of participants, PCs were calculated separately for each endophenotype using the “pca” procedure in PLINK1.9. NACC and ROSMAP variant-level results were meta-analyzed using a fixed-effect, inverse-variance meta-analysis via the metagen function from the meta R package, version 4.18-0 (Balduzzi, Rücker, and Schwarzer 2019). Random-effect meta-analysis is appropriate, in general, for combining studies with varying studying designs but can result in problems concerning power and heterogeneity estimates, especially when few studies are combined (Jackson and Turner 2017; von Hippel 2015; Gavaghan, Moore, and McQuay 2000). We, thus, chose to employ fixed-effect meta-analysis. Plots of study-specific and meta-analyzed variant-level p-values were created using LocusZoom Standalone, version 1.4 (https://genome.sph.umich.edu/wiki/LocusZoom_Standalone) (Pruim *et al.* 2010). Linkage disequilibrium estimates were computed using LDlink assuming a CEU population (<https://ldlink.nci.nih.gov/>) (Machiela and Chanock 2015).

Variant Prioritization and Downstream Analyses

Statistically significant variants were identified using a Bonferroni-corrected threshold for significance that accounts for the effective number of independent tests in the *WWOX/MAF* \pm 250kb region. The effective number of independent tests in the region was calculated for each endophenotype subset using the method of Gao *et al.* (Gao, Starmer, and Martin 2008). Briefly, Pearson’s correlation coefficient was calculated for all pairs of variants and these coefficients were placed in a square matrix. The eigenvalues of the matrix were then

computed and ordered from largest to smallest and the effective number of independent tests was defined to be the smallest number of ordered eigenvalues that account for 99.5% of the sum of all eigenvalues. The Bonferroni-corrected threshold for identifying prioritized variants for an endophenotype was defined as a variant-level p-value less than 0.05 divided by the effective number of independent tests in the region for the endophenotype.

Prioritized variants were investigated for expression quantitative trait loci (eQTL) and splicing quantitative trait loci (sQTL) associations using the Genotype-Tissue Expression (GTEx) Project's V8 public data (Consortium 2013) and the BRAINEAC Brain eQTL Almanac (<http://braineac.org/>) (Ramasamy et al. 2014). Prioritized variants were also investigated for associations with other molecular mechanisms using the INFERRing the molecular mechanisms of Noncoding genetic variants (INFERNO) software assuming a threshold on r^2 of 0.5 and a threshold on linkage disequilibrium (LD) block size of 500 kb (<http://inferno.lisanwanglab.org/index.php>) (Amlie-Wolf et al. 2018). Prioritized variants were also investigated for association with clinical AD using two large data sources: the Phase 3 summary statistics from the clinical AD GWAS of Jansen *et al.* (Jansen et al. 2019) and the ADGC-HRC data.

Sensitivity Analyses

The dependency of the study's results on several analytic choices were investigated. In addition to including 250kb of flanking on both sides of the *WWOX/MAF* locus, all analyses were conducted assuming no flanking and 25kb of flanking. Since some neurodegenerative diseases are more pronounced at later ages and some variant effects may be age-dependent or only affect the age of onset, all analyses were also conducted on the subset of individuals with ages of death of 75 years or older. Finally, to determine if significant variant-level results were independent of ADNC, all analyses were also conducted while adjusting for the presence of neurofibrillary tangles and, separately, for the presence of neuritic plaques.

Results

In the NACC data set, n=3,749 individuals had available data for at least one of the endophenotypes along with GWAS data. In ROSMAP, a total of n=1,390 individuals had available data for at least one of the endophenotypes along with GWAS data. Table 1 shows a summary of individual characteristics and endophenotypes for both NACC and ROSMAP participants. NACC participants with neurofibrillary tangles (p<0.001), neuritic plaques (p<0.001), and brain arteriosclerosis (p<0.001) tended to be younger at death. Conversely, ROSMAP participants with an endophenotype present tended to be older at death and were less likely to be male (all p<0.05).

Variant-Level Associations

A total of 9,492 genetic variants in the *WWOX/MAF* locus passed QC in NACC and 8,953 variants passed QC in ROSMAP. A total of 8,256 variants were shared between NACC and ROSMAP and were included in the meta-analysis. Notably, rs62039712, the top *WWOX/MAF* variant from the Kunkle *et al.* clinical AD GWAS (Kunkle et al. 2019), did

not pass QC in either data set because it was missing in greater than 5% of individuals. No variants were in high enough linkage disequilibrium with rs62039712 in a CEU populations in LDlink to serve as proxies (no variants with $R^2 > 0.4$ within 500kb of rs62039712).

The *WWOX* variant previously found to be genome-wide suggestive for HS with a recessive MOI, rs55751884 (Nelson et al. 2014), had nominally significant adjusted associations with HS (OR=5.53, 95% CI: (1.51, 19.51), $p=0.01133$) in NACC and neuritic plaques in both NACC and ROSMAP (NACC: OR=2.38, 95% CI: (1.32, 4.57), $p=0.00330$; ROSMAP: OR=0.32, 95% CI: (0.07, 0.93), $p=0.03506$) assuming a recessive MOI (Table 2). However, while the NACC and ROSMAP odds ratios for rs55751884 on neuritic plaques were of similar magnitude, they did not point in the same direction despite having the same minor allele (NACC: OR=2.38; ROSMAP: OR=0.32) and the adjusted meta-analytic p-value did not reach nominal significance (OR=1.58, 95% CI: (0.91, 2.74), $p=0.10566$). Notably, the adjusted meta-analytic association between rs55751884 and HS remained nominally significant when restricted to participants not included in the 2014 HS GWAS (meta-analysis: OR=2.97, 95% CI: (1.13, 7.83), $p=0.02748$) representing a nominal replication of that HS association (Supplemental Table 2). When additive and dominant MOIs were assumed, additional adjusted associations reached nominal statistical significance including brain arteriolosclerosis in NACC (dominant MOI: OR=0.82, 95% CI: (0.69, 0.98), $p=0.03008$) (Supplemental Table 2).

The recently identified genome-wide significant clinical AD risk variant near the *MAF* gene, rs450674 which is between *WWOX* and *MAF* approximately 53kb away from *MAF*'s 3' end (Bellenguez et al. 2020), had a nominally significant adjusted association with neuritic plaques in NACC (OR=0.86, 95% CI: (0.78, 0.96), $p=0.00637$) and in the meta-analysis of NACC and ROSMAP (OR=0.90, 95% CI: (0.82, 0.99), $p=0.03227$) assuming an additive MOI (Table 2). No additional adjusted associations were found to be nominally significant when assuming a recessive or dominant MOI (Supplemental Table 2).

Variant Prioritization and Downstream Analyses

The largest estimate of the effective number of independent tests for the *WWOX/MAF* locus ± 250 kb was 1,364 in NACC and 804 in ROSMAP. The larger of these two estimates was used to compute the Bonferroni-corrected threshold for the *WWOX/MAF* locus ± 250 kb of 3.67×10^{-5} (0.05/1,364). Variants with p-values less than this threshold were prioritized for further investigation. Associations with ADNC endophenotypes were notably absent for the recently identified clinical AD loci of rs62039712 and rs450674 (Figure 1). Several loci in the *WWOX/MAF* region contained variants that approached the Bonferroni-corrected threshold for significance in NACC but none surpassed it (Supplemental Figure 1).

Eight thousand two hundred and fifty-six variants were shared between NACC and ROSMAP in the *WWOX/MAF* locus ± 250 kb and were meta-analyzed across NACC and ROSMAP. Five variants had meta-analytic p-values that met the Bonferroni-corrected significance threshold for the *WWOX/MAF* region ± 250 kb for at least one endophenotype and one MOI. Two of these variants, rs6564590 and rs7404901, were associated with LATE-NC assuming an additive MOI (OR=1.43, 95% CI: (1.22, 1.68), $p=1.07 \times 10^{-5}$ and OR=1.44, 95% CI: (1.22, 1.69), $p=1.56 \times 10^{-5}$, respectively) and are in high LD with one another

($R^2=0.735$ in LDlink). Two additional variants, rs9925100 and rs9930659, were associated with HS while assuming a recessive MOI (OR=2.44, 95% CI: (1.63, 3.61), $p=1.34\times 10^{-5}$ and OR=2.29, 95% CI: (1.57, 3.34), $p=1.82\times 10^{-5}$, respectively). These two variants are also in high LD with one another ($R^2 = 0.726$ in LDlink) and are located approximately 947kb downstream from the previously identified *WWOX* HS locus, rs55751884. The remaining variant, rs4435266, was associated with brain arteriolosclerosis while assuming a dominant MOI (OR=0.74, 95% CI: (0.64, 0.85), $p=2.02\times 10^{-5}$) (Table 3 and Figure 2). While the other endophenotypes were associated with variants in either the NACC-only or the ROSMAP-only analyses, they did not have any associations with meta-analytic p-values that met the Bonferroni-corrected significance threshold (Supplemental Figure 2).

None of the prioritized meta-analytic variants were found to be associated with eQTLs or sQTLs for *WWOX*, *MAF*, or any other proximal genes in GTEx. However, all five variants were found to have notable associations in BRAINEAC. The two HS variants, rs9925100 and rs9930659, had nominally significant eQTL associations for *WWOX* (both brain tissue-wide p-values $< 3.9\times 10^{-3}$) with the hippocampus and putamen regions having the strongest single-tissue associations. Both of the LATE-NC variants, rs6564590 and rs7404901, had nominally significant eQTL associations with *MAF* (brain tissue-wide $p=0.040$ and $p=0.012$, respectively), with the thalamus region having the strongest single-tissue association for rs6564590 ($p=0.0096$) and the frontal cortex region having the strongest single-tissue association for rs7404901 ($p=0.0036$). The brain arteriolosclerosis variant, rs4435266, also had a nominally significant eQTL association with *MAF* (brain tissue-wide $p=0.019$) with the cerebellum region having the strongest single-tissue association ($p=0.0088$). Additionally, in INFERNO the LATE-NC variants were found to be eQTLs for Roadmap enhancers in the blood and immune organ tissues, the brain arteriolosclerosis variant was found to be an eQTL for Roadmap enhancers in blood and skeletal muscle tissues, and the HS variants were found to be eQTLs for both Roadmap and FANTOM5 enhancers in the blood and Roadmap enhancers in immune organ and skeletal muscle tissues. Of the five prioritized variants, only rs7404901 was nominally significant in the Jansen *et al.* GWAS of clinical AD (Jansen *et al.* 2019), though the effect was weak ($n=426,823$, OR=1.005, 95% CI: (1.000, 1.009), $p=0.03818$). Three of the prioritized variants – rs6564590, rs9925100, and rs9930659 – had nominally significant marginal associations with clinical AD status in ADGC-HRC and the corresponding effect estimates were also small ($n=21,439$, OR=1.05, 95% CI: (1.01, 1.09), $p=0.01115$; $n=21,507$, OR=0.96, 95% CI: (0.92, 1.00), $p=0.04813$; and $n=20,865$, OR=0.94, 95% CI: (0.91, 0.98), $p=0.00521$, respectively).

Sensitivity Analyses

Varying *WWOX/MAF* Flanking.—The Bonferroni-corrected threshold for the *WWOX/MAF* locus was estimated to be 3.12×10^{-5} (0.05/1,214) with 25kb of flanking and 4.19×10^{-5} (0.05/1,194) with 0kb of flanking. Not surprisingly, all five of the prioritized meta-analytic variants along with the majority of the NACC-only and ROSMAP-only variants identified in the primary analysis with 250kb of flanking were also identified when the flanking was reduced. The exception to this were several variants associated with neuritic plaques in the NACC-only analysis which were located just upstream of *WWOX* near the *CLEC3A* gene. Of the 12 neuritic plaque variants that surpassed the

Bonferroni-corrected threshold in the NACC-only analysis when the *WWOX/MAF* locus was flanked by 250kb, only one variant (rs79416778) remained with no flanking. This result highlights the specificity of the neuritic plaques association, not only with NACC, but with the *CLEC3A* gene specifically. See Supplemental Table 4 for the complete results.

Age of Death 75+: The odds ratio estimates for all five of the prioritized meta-analytic variants remained largely unchanged when the analyses were restricted to only those individuals with an age of death of 75 years of age or older. Additionally, the meta-analytic p-values for the associations between rs6564590 and LATE-NC (OR=1.42, 95% CI: (1.20, 1.66), $p=2.85\times 10^{-5}$) and rs4435266 and brain arteriolosclerosis (OR=0.73, 95% CI: (0.63, 0.85), $p=3.07\times 10^{-5}$) and remained below the Bonferroni-corrected threshold. The remaining associations were all nominally significant, but did not quite meet the Bonferroni-corrected threshold (all remaining p-values $\leq 6.12\times 10^{-5}$). These findings suggest that age of death does not meaningfully impact the associations between the prioritized variants and the endophenotypes. See Supplemental Table 5 for the complete results.

Adjusting for ADNC.—The odds ratio estimates for all five of the prioritized meta-analytic variants remained largely unchanged when the analyses were adjusted for neurofibrillary tangles and, separately, neuritic plaques. Additionally, all of the meta-analytic p-values remained nominally significant after adjustment for neurofibrillary tangles and, separately, neuritic plaques. These findings suggest that the associations of the prioritized variants are independent of ADNC. See Supplemental Table 6 for the complete results.

Discussion

Using autopsy-confirmed neuropathologic endophenotypes, we evaluated the genetic associations between the *WWOX/MAF* locus and several neurodegenerative diseases using neuropathological changes to operationalize the presence and severity of the diseases. We found significant adjusted meta-analytic associations between *WWOX* variants and LATE-NC, HS, and brain arteriolosclerosis. While previous GWASs linked variants in the *WWOX/MAF* locus with HS and clinical AD, the associations with LATE-NC and brain arteriolosclerosis have never been reported. Furthermore, since these associations remained nominally significant after adjustment for AD-related neuropathological changes and none of the variants were robustly associated with clinical AD, it suggests that the LATE, HS, and brain arteriolosclerosis neuropathological changes associated with *WWOX/MAF* are independent of ADNC.

The novel neuritic plaque signal found in the NACC-only analyses near the *CLEC3A* gene is intriguing since other *CLEC* family genes have been linked to AD and inflammation (Wang, Liu, et al. 2020; Porcellini et al. 2010; Meng et al. 2020). Additionally, a recent genome-wide interaction analysis found evidence of variant-by-variant interactions for neurofibrillary tangles involving variants near *CLEC3A* and *WWOX* (Wang, Yang, et al. 2020). Further investigations into the influence of *CLEC3A* on neurodegenerative disease are warranted.

The previously identified AD-associated *WWOX/MAF* variant, rs62039712, did not pass QC in either dataset and did not have any proxy variants, and was only available in two of

the 12 Stage 1 GWAS cohorts from Kunkle *et al.* These factors indicate that rs62039712 was difficult to impute. The recently identified AD-associated *MAF* variant, rs450674 (Bellenguez et al. 2020), which is located approximately 362kb downstream of *WFOX* and is not in linkage disequilibrium with rs62039712, was found to have a nominally significant adjusted association with neuritic plaques in NACC (OR=0.86, 95% CI: (0.78, 0.96), p=0.00637) and in the meta-analysis of NACC and ROSMAP (OR=0.90, 95% CI: (0.82, 0.99), p=0.03227). However, since Kunkle *et al.*, Bellenguez *et al.*, and the present study have all utilized data from the ADGC – albeit using different, more specific phenotypes in the case of the current study – this region of the genome should be investigated further in other datasets to better understand its influence on AD risk.

We examined the associations of the genome-wide suggestive HS *WFOX* variant, rs55751884, with neuropathological endophenotypes. The rs55751884 variant was nominally significant in adjusted association tests for neuritic plaques in both NACC and ROSMAP (NACC: OR=2.38, 95% CI: (1.32, 4.57), p=0.00330; ROSMAP: OR=0.32, 95% CI: (0.07, 0.93), p=0.03506), HS in NACC (OR=5.53, 95% CI: (1.51, 19.51), p=0.01133), and borderline significant for neurofibrillary tangles in NACC (OR=1.66, 95% CI: (0.94, 3.07), p=0.08202). Even though the meta-analytic association between HS and rs55751884 did not reach the Bonferroni-corrected threshold for significance in our current study, that same region of *WFOX* had the strongest association with arteriolosclerosis in ROSMAP, which also merits additional investigation.

Given the abundant evidence that mixed pathologies are highly prevalent in elderly populations, the hypothesis that *WFOX* is associated with several neuropathological endophenotypes fits in with recent studies looking at genetic pleiotropy in neurological conditions (Choronenky, Fardo, and Nelson 2019). Pleiotropic effects have been found between AD and Parkinson's disease (Ibanez et al. 2018), AD and amyotrophic lateral sclerosis (Montibeller and de Belleruche 2018), early-onset AD and frontotemporal dementia (Cochran et al. 2020), AD-related psychosis and schizophrenia (Creese et al. 2019), LATE-NC and FTLT-TDP (Nelson et al. 2019), and, more recently, LATE-NC and HS (Dugan et al. 2021). A specific example is the *MAPT* gene which is a risk allele for many tauopathies, and also for Parkinson's disease (not a condition linked to tau pathology) (Lin and Farrer 2014). Pleiotropic effects have also been found between AD-related neuropathological changes like neuritic plaques, neurofibrillary tangles, and cerebral amyloid angiopathy (Chung et al. 2018). Further, it has been shown that brain arteriolosclerosis is linked to HS and LATE-NC (Blevins et al. 2020; Neltner et al. 2014). Our data indicate that the associations between the *WFOX/MAF* locus and LATE-NC, HS, and brain arteriolosclerosis were independent of ADNC. Thus, *WFOX* is apparently associated with more than one clinico-pathologic entity. Since *WFOX* is also known to play a role in molecular functions (Teng et al. 2013), autism spectrum disorder (Bacchelli et al. 2020), multiple sclerosis (Beecham et al. 2013), schizophrenia (McClay et al. 2011), and brain volume (Xia et al. 2017), it is a good target for additional follow-up studies.

There are limitations to our study. While the NACC and ROSMAP cohorts are quite different due to varying recruitment models, our primary results showed agreement between the two cohorts. For the five newly identified variants, the allele frequencies and effect

estimates were broadly consistent across NACC and ROSMAP, suggesting that these variants are similarly prevalent and associative in both cohorts (Table 3). The two studies did show some disagreement. When the *WWOX/MAF* variants previously reported were investigated in the present study, rs55751884 showed diverging results between NACC and ROSMAP for neurofibrillary tangles and neuritic plaques and differing effect sizes for HS and LATE-NC (Table 2). Additionally, the association between *CLEC3A* variants and neuritic plaques was seen only in the NACC cohort.

Because data come from studies employing variable study designs and are highly homogeneous, the degree to which findings are generalizable is unknown, especially concerning individuals of non-Caucasian ancestries. Additionally, while a random effects meta-analysis can be optimal for combining heterogeneous studies, we employed a fixed effects meta-analysis as we were only combining two studies and a random effects approach can be underpowered in such situations (Jackson and Turner 2017). These suggestive findings extend prior research in the field that linked the *WWOX/MAF* locus with neurodegenerative phenotypes. Yet these findings need corroborative evaluations in additional data sets to evaluate the relationships between genetics and neuropathologic data.

Conclusions

In conclusion, we showed using large genetic datasets and autopsy-derived endophenotypes that neuropathological endophenotypes related to LATE, HS, and brain arteriosclerosis were associated with *WWOX/MAF* gene variants. While clinical diagnoses of AD may be helpful for discovering dementia-related genetic variation, our study adds to the growing body of literature highlighting the complexity of dementia phenotypes, and the benefit of leveraging autopsy-derived data for studies of aging-related brain disease.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgements

We are grateful to the research volunteers, clinicians and colleagues that enabled us to perform these studies. The following NIH grants supported this work: R56-AG057191, R01-AG057187, P30-AG028383, R01-AG059716, and K01-AG049164.

The NACC database is funded by NIA/NIH Grant U01 AG016976. NACC data are contributed by the NIA-funded ADCs: P30 AG019610 (PI Eric Reiman, MD), P30 AG013846 (PI Neil Kowall, MD), P30 AG062428-01 (PI James Leverenz, MD) P50 AG008702 (PI Scott Small, MD), P50 AG025688 (PI Allan Levey, MD, PhD), P50 AG047266 (PI Todd Golde, MD, PhD), P30 AG010133 (PI Andrew Saykin, PsyD), P50 AG005146 (PI Marilyn Albert, PhD), P30 AG062421-01 (PI Bradley Hyman, MD, PhD), P30 AG062422-01 (PI Ronald Petersen, MD, PhD), P50 AG005138 (PI Mary Sano, PhD), P30 AG008051 (PI Thomas Wisniewski, MD), P30 AG013854 (PI Robert Vassar, PhD), P30 AG008017 (PI Jeffrey Kaye, MD), P30 AG010161 (PI David Bennett, MD), P50 AG047366 (PI Victor Henderson, MD, MS), P30 AG010129 (PI Charles DeCarli, MD), P50 AG016573 (PI Frank LaFerla, PhD), P30 AG062429-01 (PI James Brewer, MD, PhD), P50 AG023501 (PI Bruce Miller, MD), P30 AG035982 (PI Russell Swerdlow, MD), P30 AG028383 (PI Linda Van Eldik, PhD), P30 AG053760 (PI Henry Paulson, MD, PhD), P30 AG010124 (PI John Trojanowski, MD, PhD), P50 AG005133 (PI Oscar Lopez, MD), P50 AG005142 (PI Helena Chui, MD), P30 AG012300 (PI Roger Rosenberg, MD), P30 AG049638 (PI Suzanne Craft, PhD), P50 AG005136 (PI Thomas Grabowski, MD), P30 AG062715-01 (PI Sanjay Asthana, MD, FRCP), P50 AG005681 (PI John Morris, MD), P50 AG047270 (PI Stephen Strittmatter, MD, PhD)

The Genotype-Tissue Expression (GTEx) Project was supported by the Common Fund of the Office of the Director of the National Institutes of Health, and by NCI, NHGRI, NHLBI, NIDA, NIMH, and NINDS. The data used for the analyses described in this manuscript were obtained from the GTEx Portal (<https://www.gtexportal.org/home/datasets>) on 9/17/2020.

The National Institutes of Health, National Institute on Aging (NIH-NIA) supported this work through the following grants: ADGC, U01 AG032984, RC2 AG036528; Samples from the National Cell Repository for Alzheimer's Disease (NCRAD), which receives government support under a cooperative agreement grant (U24 AG21886) awarded by the National Institute on Aging (NIA), were used in this study.

We thank contributors who collected samples used in this study, as well as patients and their families, whose help and participation made this work possible; Data for this study were prepared, archived, and distributed by the National Institute on Aging Alzheimer's Disease Data Storage Site (NIAGADS) at the University of Pennsylvania (U24-AG041689-01); NACC, U01 AG016976; NIA LOAD, U24 AG026395, R01AG041797; Banner Sun Health Research Institute P30 AG019610; Boston University, P30 AG013846, U01 AG10483, R01 CA129769, R01 MH080295, R01 AG017173, R01 AG025259, R01AG33193; Columbia University, P50 AG008702, R37 AG015473; Duke University, P30 AG028377, AG05128; Emory University, AG025688; Group Health Research Institute, U01 AG006781, U01 HG004610, U01 HG006375; Indiana University, P30 AG101333; Johns Hopkins University, P50 AG005146, R01 AG020688; Massachusetts General Hospital, P50 AG005134; Mayo Clinic, P50 AG016574; Mount Sinai School of Medicine, P50 AG005138, P01 AG002219; New York University, P30 AG08051, UL1 RR029893, 5R01AG012101, 5R01AG022374, 5R01AG013616, 1RC2AG036502, 1R01AG035137; Northwestern University, P30 AG013854; Oregon Health & Science University, P30 AG008017, R01 AG026916; Rush University, P30 AG010161, R01 AG019085, R01 AG15819, R01 AG17917, R01 AG30146; TGen, R01 NS059873; University of Alabama at Birmingham, P50 AG016582; University of Arizona, R01 AG031581; University of California, Davis, P30 AG010129; University of California, Irvine, P50 AG016573; University of California, Los Angeles, P50 AG016570; University of California, San Diego, P50 AG005131; University of California, San Francisco, P50 AG023501, P01 AG019724; University of Kentucky, P30 AG028383, AG05144; University of Michigan, P50 AG008671; University of Pennsylvania, P30 AG010124; University of Pittsburgh, P50 AG005133, AG030653, AG041718, AG07562, AG02365; University of Southern California, P50 AG005142; University of Texas Southwestern, P30 AG012300; University of Miami, R01 AG027944, AG010491, AG027944, AG021547, AG019757; University of Washington, P50 AG005136; University of Wisconsin, P50 AG033514; Vanderbilt University, R01 AG019085; and Washington University, P50 AG005681, P01 AG03991.

The Kathleen Price Bryan Brain Bank at Duke University Medical Center is funded by NINDS grant # NS39764, NIMH MH60451 and by Glaxo Smith Kline. Genotyping of the TGEN2 cohort was supported by Kronos Science. The TGen series was also funded by NIA grant AG041232 to AJM and MJH, The Banner Alzheimer's Foundation, The Johnnie B. Byrd Sr.

Alzheimer's Institute, the Medical Research Council, and the state of Arizona and also includes samples from the following sites: Newcastle Brain Tissue Resource (funding via the Medical Research Council, local NHS trusts and Newcastle University), MRC London Brain Bank for Neurodegenerative Diseases (funding via the Medical Research Council), South West Dementia Brain Bank (funding via numerous sources including the Higher Education Funding Council for England (HEFCE), Alzheimer's Research Trust (ART), BRACE as well as North Bristol NHS Trust Research and Innovation Department and DeNDroN), The Netherlands Brain Bank (funding via numerous sources including Stichting MS Research, Brain Net Europe, Hersenstichting Nederland Breinbrekend Werk, International Parkinson Fonds, Internationale Stichting Alzheimer Onderzoek), Institut de Neuropatologia, Servei Anatomia Patologica, Universitat de Barcelona.

ADNI data collection and sharing was funded by the National Institutes of Health Grant U01 AG024904 and Department of Defense award number W81XWH-12-2-0012. ADNI is funded by the National Institute on Aging, the National Institute of Biomedical Imaging and Bioengineering, and through generous contributions from the following: AbbVie, Alzheimer's Association; Alzheimer's Drug Discovery Foundation; Araclon Biotech; BioClinica, Inc.; Biogen; Bristol-Myers Squibb Company; CereSpir, Inc.; Eisai Inc.; Elan Pharmaceuticals, Inc.; Eli Lilly and Company; EuroImmun; F. Hoffmann-La Roche Ltd and its affiliated company Genentech, Inc.; Fujirebio; GE Healthcare; IXICO Ltd.; Janssen Alzheimer Immunotherapy Research & Development, LLC.; Johnson & Johnson Pharmaceutical Research & Development LLC.; Lumosity; Lundbeck; Merck & Co., Inc.; Meso Scale Diagnostics, LLC.; NeuroRx Research; Neurotrack Technologies; Novartis Pharmaceuticals Corporation; Pfizer Inc.; Piramal Imaging; Servier; Takeda Pharmaceutical Company; and Transition Therapeutics.

The Canadian Institutes of Health Research is providing funds to support ADNI clinical sites in Canada. Private sector contributions are facilitated by the Foundation for the National Institutes of Health (www.fnih.org).

The grantee organization is the Northern California Institute for Research and Education, and the study is coordinated by the Alzheimer's Disease Cooperative Study at the University of California, San Diego. ADNI data are disseminated by the Laboratory for Neuro Imaging at the University of Southern California. We thank Drs. D. Stephen Snyder and Marilyn Miller from NIA who are ex-officio ADGC members.

Support was also from the Alzheimer's Association (LAF, IIRG-08-89720; MP-V, IIRG-05-14147) and the US Department of Veterans Affairs Administration, Office of Research and Development, Biomedical Laboratory Research Program. P.S.G.-H. is supported by Wellcome Trust, Howard Hughes Medical Institute, and the Canadian Institute of Health Research.

Abbreviations

AD	Alzheimer's disease
ADGC	Alzheimer's disease genetics consortium
ADNC	Alzheimer's disease neuropathological changes
ADRD	Alzheimer's disease-related dementias
AD-ADRD	Alzheimer's disease and Alzheimer's disease-related dementias
eQTL	expression quantitative trait loci
FTLD	frontotemporal lobar degeneration
GTE_x	Genotype-Tissue Expression
GWAS	genome wide association study
HS	hippocampal sclerosis
HWE	Hardy-Weinberg equilibrium
LATE	limbic-predominant age-related TDP-43 encephalopathy
LATE-NC	limbic-predominant age-related TDP-43 encephalopathy neuropathological changes
LD	linkage disequilibrium
MOI	mode of inheritance
NACC	National Alzheimer's Coordinating Center
NP	neuropathology
OR	odds ratio
PC	principal component
QC	quality control
ROSMAP	Religious Orders Study and the Rush Memory and Aging Project
sQTL	splicing quantitative trait loci
TDP-43	TAR-DNA binding protein 43
WWOX	WW domain-containing oxidoreductase

References

- Amlie-Wolf Alexandre, Tang Mitchell, Mlynarski Elisabeth E, Kuksa Pavel P, Valladares Otto, Katanic Zivadin, Tsuang Debby, Brown Christopher D, Schellenberg Gerard D, and Wang Li-San. 2018. 'INFERNO: inferring the molecular mechanisms of noncoding genetic variants', *Nucleic Acids Research*, 46: 8740–53. [PubMed: 30113658]
- Bacchelli E, Cameli C, Viggiano M, Iglizzo R, Mancini A, Tancredi R, Battaglia A, and Maestrini E. 2020. 'An integrated analysis of rare CNV and exome variation in Autism Spectrum Disorder using the Infinium PsychArray', *Sci Rep*, 10: 3198. [PubMed: 32081867]
- Balduzzi Sara, Rücker Gerta, and Schwarzer Guido. 2019. 'How to perform a meta-analysis with R: a practical tutorial', *Evidence Based Mental Health*, 22: 153–60. [PubMed: 31563865]
- Beecham AH, Patsopoulos NA, Xifara DK, Davis MF, Kempainen A, Cotsapas C, Shah TS, Spencer C, Booth D, Goris A, Oturai A, Saarela J, Fontaine B, Hemmer B, Martin C, Zipp F, D'Alfonso S, Martinelli-Boneschi F, Taylor B, Harbo HF, Kockum I, Hillert J, Olsson T, Ban M, Oksenberg JR, Hintzen R, Barcellos LF, Agliardi C, Alfredsson L, Alizadeh M, Anderson C, Andrews R, Søndergaard HB, Baker A, Band G, Baranzini SE, Barizzone N, Barrett J, Bellenguez C, Bergamaschi L, Bernardinelli L, Berthele A, Biberacher V, Binder TM, Blackburn H, Bomfim IL, Brambilla P, Broadley S, Brochet B, Brundin L, Buck D, Butzkueven H, Caillier SJ, Camu W, Carpentier W, Cavalla P, Celiuș EG, Coman I, Comi G, Corrado L, Cosemans L, Courno-Rebeix I, Cree BA, Cusi D, Damotte V, Defer G, Delgado SR, Deloukas P, di Sapio A, Dillthey AT, Donnelly P, Dubois B, Duddy M, Edkins S, Elovaara I, Esposito F, Evangelou N, Fiddes B, Field J, Franke A, Freeman C, Frohlich IY, Galimberti D, Gieger C, Gourraud PA, Graetz C, Graham A, Grummel V, Guaschino C, Hadjixenofontos A, Hakonarson H, Halfpenny C, Hall G, Hall P, Hamsten A, Harley J, Harrower T, Hawkins C, Hellenthal G, Hillier C, Hobart J, Hoshi M, Hunt SE, Jagodic M, Jel i I, Jochim A, Kendall B, Kermode A, Kilpatrick T, Koivisto K, Konidari I, Korn T, Kronsbein H, Langford C, Larsson M, Lathrop M, Lebrun-Frenay C, Lechner-Scott J, Lee MH, Leone MA, Leppä V, Liberatore G, Lie BA, Lill CM, Lindén M, Link J, Luessi F, Lycke J, Macchiardi F, Männistö S, Manrique CP, Martin R, Martinelli V, Mason D, Mazibrada G, McCabe C, Mero IL, Mescheriakova J, Moutsianas L, Myhr KM, Nagels G, Nicholas R, Nilsson P, Piehl F, Pirinen M, Price SE, Quach H, Reunanen M, Robberecht W, Robertson NP, Rodegher M, Rog D, Salvetti M, Schnetz-Boutaud NC, Sellebjerg F, Selter RC, Schaefer C, Shaunak S, Shen L, Shields S, Siffrin V, Slee M, Sorensen PS, Sorosina M, Sospedra M, Spurkland A, Strange A, Sundqvist E, Thijs V, Thorpe J, Ticca A, Tienari P, van Duijn C, Visser EM, Vucic S, Westerlind H, Wiley JS, Wilkins A, Wilson JF, Winkelmann J, Zajicek J, Zindler E, Haines JL, Pericak-Vance MA, Ivinson AJ, Stewart G, Hafler D, Hauser SL, Compston A, McVean G, De Jager P, Sawcer SJ, and McCauley JL. 2013. 'Analysis of immune-related loci identifies 48 new susceptibility variants for multiple sclerosis', *Nat Genet*, 45: 1353–60. [PubMed: 24076602]
- Bellenguez Céline, Küçükali Fahri, Jansen Iris, Andrade Victor, Morenau-Grau Sonia, Amin Najaf, Grenier-Boley Benjamin, Boland Anne, Kleineidam Luca, Holmans Peter, Garcia Pablo, Martin Rafael Campos, Naj Adam, Qiong Yang, Bis Joshua C., Damotte Vincent, Van der Lee Sven, Costa Marcos, Chapuis Julien, Giedraitis Vilmentas, Bullido María Jesús, de Munáin Adolfo López, Pérez-Tur Jordi, Sánchez-Juan Pascual, Sánchez-Valle Raquel, Álvarez Victoria, Pastor Pau, Medina Miguel, Van Dongen Jasper, Van Broeckhoven Christine, Vandenberghe Rik, Engelborghs Sebastiaan, Nicolas Gael, Pasquier Florence, Hanon Olivier, Dufouil Carole, Berr Claudine, Debette Stéphanie, Dartigues Jean-François, Spalletta Gianfranco, Nacmias Benedetta, Solfrezi Vincenzo, Borroni Barbara, Tremolizzo Lucio, Seripa Davide, Caffarra Paolo, Daniele Antonio, Galimberti Daniela, Rainero Innocenzo, Benussi Luisa, Squassina Alesio, Mecoci Patrizia, Parnetti Lucilla, Masullo Carlo, Arosio Beatrice, Hardy John, Mead Simon, Morgan Kevin, Holmes Clive, Kehoe Patrick, Woods Bob, Sha Jin, Zhao Yi, Lee Chien-Yueh, Kuksa Pavel P., Hamilton-Nelson Kara L., Kunkle Brian W., Bush William S., Martin Eden R., Wang Li-San, Mayeux Richard, Farrer Lindsay A., Haines Jonathan L., Pericak-Vance Margaret A., Wang Ruiqi, Satizabal Claudia, Psaty Bruce, Lopez Oscar, Sanchez-Garcia Florentino, Nordestgaard Børge G., Tybjærg-Hansen Anne, Thomassen Jesper Qvist, Graff Caroline, Papenberg Goran, Soininen Hilikka, Kivipelto Miia, Haapasalo Annakaisa, Ngandu Tiia, Koivisto Anne, kuulasmaa teemu, Porcel Laura Molina, Kornhuber Johannes, Peters Oliver, schneider Anja, Scarmeas Nikolaos, Dichgans Martin, Froelich Lutz, Rujescu Dan, Diehl-Schmid Janine, Grimmer Timo, Schmid Matthias, Möthen Markus M,

Grünblatt Edna, Popp Julius, Scherbaum Norbert, Mehrabian Shima, Deckert Jürgen, Aarsland Dag, Selbæk Geir, Saltvedt Ingvild, Djurovic Srdjan, Holstege Henne, Pijnenburg Yolande A.L., Van Swieten John, Ramakers Inez, Van der Lugt Aad, Claassen Jurgen A.H.R, Jan Biessels Geert, Scheltens Philip, Antúnez Carmen, Mir Pablo, Real Luis Miguel, García-Alberca Jose María, Piñol-Ripoll Gerard, Garcia-Ribas Guillermo, Serrano-Ríos Manuel, Williams Julie, Sachdev Perminder, Amouyel Philippe, Mather Karen, Jessen Frank, de Mendonça Alexandre, Hort Jakub, Tsolaki Magda, Frikke-Schmidt Ruth, Clarimon Jordi, Deleuze Jean-François, Seshadri Sudha, Schellenberg Gerald, Rossi Giacomina, Andreassen Ole, Ingelsson Martin, Hiltunen Mikko, Sleegers Kristel, Van Duijn Cornelia, Sims Rebecca, Van der Flier Wiesje M., Ruiz Agustin, Ramirez Alfredo, and Lambert Jean-Charles. 2020. 'Large meta-analysis of genome-wide association studies expands knowledge of the genetic etiology of Alzheimer's disease and highlights potential translational opportunities', medRxiv: 2020.10.01.20200659.

Blevins Brittney L., Vinters Harry V., Love Seth, Wilcock Donna M., Grinberg Lea T., Schneider Julie A., Kalaria Rajesh N., Katsumata Yuriko, Gold Brian T., Wang Danny J. J., Ma Samantha J., Shade Lincoln M. P., Fardo David W., Hartz Anika M. S., Jicha Gregory A., Nelson Karin B., Magaki Shino D., Schmitt Frederick A., Teylan Merilee A., Ighodaro Eseosa T., Phe Panhavuth, Abner Erin L., Cykowski Matthew D., Van Eldik Linda J., and Nelson Peter T.. 2020. 'Brain arteriolosclerosis', *Acta neuropathologica*.

Campbell Purdey, Brix Thomas H., Wilson Scott G., Ward Lynley C., Hui Jennie, Beilby John P., Hegedüs Laszlo, and Walsh John P. 2016. 'Common genetic variants associated with thyroid function may be risk alleles for Hashimoto's disease and Graves' disease', *Clinical Endocrinology*, 84: 278–83. [PubMed: 25683181]

Chang Hsin-Tzu, Liu Chan-Chuan, Chen Shur-Tzu, Yap Ye Vone, Chang Nan-Shang, and Sze Chun I.. 2014. 'WW domain-containing oxidoreductase in neuronal injury and neurological diseases', *Oncotarget*, 5: 11792–99. [PubMed: 25537520]

Chornenkyy Y, Fardo DW, and Nelson PT. 2019. 'Tau and TDP-43 proteinopathies: kindred pathologic cascades and genetic pleiotropy', *Lab Invest*, 99: 993–1007. [PubMed: 30742063]

Chung Jaeyoon, Zhang Xiaoling, Allen Mariet, Wang Xue, Ma Yiyi, Beecham Gary, Montine Thomas J., Younkin Steven G., Dickson Dennis W., Golde Todd E., Price Nathan D., Ertekin-Taner Nilüfer, Lunetta Kathryn L., Mez Jesse, Mayeux Richard, Haines Jonathan L., Pericak-Vance Margaret A., Schellenberg Gerard, Jun Gyungah R., Farrer Lindsay A., and Consortium Alzheimer's Disease Genetics. 2018. 'Genome-wide pleiotropy analysis of neuropathological traits related to Alzheimer's disease', *Alzheimer's research & therapy*, 10: 22.

Cochran J. Nicholas, Geier Ethan G., Bonham Luke W., Newberry J. Scott, Amaral Michelle D., Thompson Michelle L., Lasseigne Brittany N., Karydas Anna M., Roberson Erik D., Cooper Gregory M., Rabinovici Gil D., Miller Bruce L., Myers Richard M., and Yokoyama Jennifer S.. 2020. 'Non-coding and Loss-of-Function Coding Variants in TET2 are Associated with Multiple Neurodegenerative Diseases', *The American Journal of Human Genetics*, 106: 632–45. [PubMed: 32330418]

Consortium, G. TEx. 2013. 'The Genotype-Tissue Expression (GTEx) project', *Nature Genetics*, 45: 580–85. [PubMed: 23715323]

Creese Byron, Vassos Evangelos, Bergh Sverre, Athanasiu Lavinia, Johar Iskandar, Rongve Arvid, Ingrid Tøndel Medbøen Miguel Vasconcelos Da Silva, Aakhus Eivind, Andersen Fred, Bettella Francesco, Braekhus Anne, Djurovic Srdjan, Paroni Giulia, Proitsi Petroula, Saltvedt Ingvild, Seripa Davide, Stordal Eystein, Fladby Tormod, Aarsland Dag, Andreassen Ole A., Ballard Clive, Selbaek Geir, consortium on behalf of the AddNeuroMed, and Initiative the Alzheimer's Disease Neuroimaging. 2019. 'Examining the association between genetic liability for schizophrenia and psychotic symptoms in Alzheimer's disease', *Translational Psychiatry*, 9: 273. [PubMed: 31641104]

Dourlen P, Kilinc D, Malmanche N, Chapuis J, and Lambert JC. 2019. 'The new genetic landscape of Alzheimer's disease: from amyloid cascade to genetically driven synaptic failure hypothesis?', *Acta neuropathologica*, 138: 221–36. [PubMed: 30982098]

Dugan AJ, Nelson PT, Katsumata Y, Shade LMP, Boehme KL, Teylan MA, Cykowski MD, Mukherjee S, Kauwe JSK, Hohman TJ, Schneider JA, and Fardo DW. 2021. 'Analysis of genes (TMEM106B, GRN, ABCC9, KCNMB2, and APOE) implicated in risk for LATE-NC and hippocampal sclerosis

provides pathogenetic insights: a retrospective genetic association study', *Acta Neuropathol Commun*, 9: 152. [PubMed: 34526147]

- Dumitrescu Logan, Mahoney Emily R, Mukherjee Shubhabrata, Lee Michael L, Bush William S, Engelman Corinne D, Lu Qiongshi, Fardo David W, Trittschuh Emily H, Mez Jesse, Kaczorowski Catherine, Saucedo Hector Hernandez, Widaman Keith F, Buckley Rachel, Properzi Michael, Mormino Elizabeth, Yang Hyun-Sik, Harrison Tessa, Hedden Trey, Nho Kwangsik, Andrews Shea J, Tommet Doug, Hadad Niran, Sanders R Elizabeth, Ruderfer Douglas M, Gifford Katherine A, Moore Annah M, Cambronero Francis, Zhong Xiaoyuan, Raghavan Neha S, Vardarajan Badri, The Alzheimer's Disease Neuroimaging Initiative, A4 Study Team Alzheimer's Disease Genetics Consortium, Pericak-Vance Margaret A, Farrer Lindsay A, Wang Li-San, Cruchaga Carlos, Schellenberg Gerard, Cox Nancy J, Haines Jonathan L, Keene C Dirk, Saykin Andrew J, Larson Eric B, Sperling Reisa A, Mayeux Richard, Bennett David A, Schneider Julie A, Crane Paul K, Jefferson Angela L, and Hohman Timothy J. 2020. 'Genetic variants and functional pathways associated with resilience to Alzheimer's disease', *Brain*, 143: 2561–75. [PubMed: 32844198]
- Gao Xiaoyi, Starmer Joshua, and Martin Eden R.. 2008. 'A multiple testing correction method for genetic association studies using correlated single nucleotide polymorphisms', *Genetic Epidemiology*, 32: 361–69. [PubMed: 18271029]
- Gavaghan DJ, Moore AR, and McQuay HJ. 2000. 'An evaluation of homogeneity tests in meta-analyses in pain using simulations of individual patient data', *Pain*, 85: 415–24. [PubMed: 10781914]
- Ibanez Laura, Dube Umer, Davis Albert A., Fernandez Maria V., Budde John, Cooper Breanna, Monica Diez-Fairen Sara Ortega-Cubero, Pastor Pau, Perlmutter Joel S., Cruchaga Carlos, and Benitez Bruno A.. 2018. 'Pleiotropic Effects of Variants in Dementia Genes in Parkinson Disease', *Frontiers in Neuroscience*, 12.
- Jackson D, and Turner R. 2017. 'Power analysis for random-effects meta-analysis', *Res Synth Methods*, 8: 290–302. [PubMed: 28378395]
- Jansen Iris E., Savage Jeanne E., Watanabe Kyoko, Bryois Julien, Williams Dylan M., Steinberg Stacy, Sealock Julia, Karlsson Ida K., Hägg Sara, Athanasiu Lavinia, Voyle Nicola, Proitsi Petroula, Witoelar Aree, Stringer Sven, Aarsland Dag, Almdahl Ina S., Andersen Fred, Bergh Sverre, Bettella Francesco, Bjornsson Sigurbjorn, Brækhus Anne, Bråthen Geir, de Leeuw Christiaan, Desikan Rahul S., Djurovic Srdjan, Dumitrescu Logan, Fladby Tormod, Hohman Timothy J., Jonsson Palmi V., Kiddle Steven J., Rongve Arvid, Saltvedt Ingvild, Sando Sigrid B., Selbæk Geir, Shoai Maryam, Skene Nathan G., Snaedal Jon, Stordal Eystein, Ulstein Ingun D., Wang Yunpeng, White Linda R., Hardy John, Hjerling-Leffler Jens, Sullivan Patrick F., van der Flier Wiesje M., Dobson Richard, Davis Lea K., Stefansson Hreinn, Stefansson Kari, Pedersen Nancy L., Ripke Stephan, Andreassen Ole A., and Posthuma Danielle. 2019. 'Genome-wide meta-analysis identifies new loci and functional pathways influencing Alzheimer's disease risk', *Nature Genetics*, 51: 404–13. [PubMed: 30617256]
- Karant Shama, Nelson Peter T., Katsumata Yuriko, Kryscio Richard J., Schmitt Frederick A., Fardo David W., Cykowski Matthew D., Jicha Gregory A., Van Eldik Linda J., and Abner Erin L.. 2020. 'Prevalence and Clinical Phenotype of Quadruple Misfolded Proteins in Older Adults', *JAMA Neurology*.
- Katsumata Yuriko, Abner Erin L., Karant Shama, Teylan Merilee A., Mock Charles N., Cykowski Matthew D., Lee Edward B., Boehme Kevin L., Mukherjee Shubhabrata, Kauwe John S. K., Kryscio Richard J., Schmitt Frederick A., Fardo David W., and Nelson Peter T.. 2020. 'Distinct clinicopathologic clusters of persons with TDP-43 proteinopathy', *Acta neuropathologica*.
- Ko la Katarzyna, El bieta Płuciennik Ewa Stycze -Binkowska, Nowakowska Magdalena, Orzechowska Magdalena, and Bednarek Andrzej K.. 2019. 'The WWOX Gene Influences Cellular Pathways in the Neuronal Differentiation of Human Neural Progenitor Cells', *Frontiers in Cellular Neuroscience*, 13.
- Kunkle BW, Schmidt M, Klein HU, Naj AC, Hamilton-Nelson KL, Larson EB, Evans DA, De Jager PL, Crane PK, Buxbaum JD, Ertekin-Taner N, Barnes LL, Fallin MD, Manly JJ, Go RCP, Obisesan TO, Kamboh MI, Bennett DA, Hall KS, Goate AM, Foroud TM, Martin ER, Wang LS, Byrd GS, Farrer LA, Haines JL, Schellenberg GD, Mayeux R, Pericak-Vance MA, Reitz C, Graff-Radford NR, Martinez I, Ayodele T, Logue MW, Cantwell LB, Jean-Francois M, Kuzma AB, Adams LD, Vance JM, Cuccaro ML, Chung J, Mez J, Lunetta KL, Jun GR, Lopez OL,

Hendrie HC, Reiman EM, Kowall NW, Leverenz JB, Small SA, Levey AI, Golde TE, Saykin AJ, Starks TD, Albert MS, Hyman BT, Petersen RC, Sano M, Wisniewski T, Vassar R, Kaye JA, Henderson VW, DeCarli C, LaFerla FM, Brewer JB, Miller BL, Swerdlow RH, Van Eldik LJ, Paulson HL, Trojanowski JQ, Chui HC, Rosenberg RN, Craft S, Grabowski TJ, Asthana S, Morris JC, Strittmatter SM, and Kukull WA. 2020. 'Novel Alzheimer Disease Risk Loci and Pathways in African American Individuals Using the African Genome Resources Panel: A Meta-analysis', *JAMA Neurol*.

Kunkle Brian W., Benjamin Grenier-Boley Rebecca Sims, Bis Joshua C., Damotte Vincent, Naj Adam C., Boland Anne, Vronskaya Maria, van der Lee Sven J., Amlie-Wolf Alexandre, Bellenguez Céline, Frizatti Aura, Chouraki Vincent, Martin Eden R., Slegers Kristel, Badarinarayan Nandini, Jakobsdottir Johanna, Hamilton-Nelson Kara L., Moreno-Grau Sonia, Olaso Robert, Raybould Rachel, Chen Yuning, Kuzma Amanda B., Hiltunen Mikko, Morgan Taniesha, Ahmad Shahzad, Vardarajan Badri N., Epelbaum Jacques, Hoffmann Per, Boada Merce, Beecham Gary W., Garnier Jean-Guillaume, Harold Denise, Fitzpatrick Annette L., Valladares Otto, Moutet Marie-Laure, Gerrish Amy, Smith Albert V., Qu Liming, Bacq Delphine, Denning Nicola, Jian Xueqiu, Zhao Yi, Maria Del Zompo Nick C. Fox, Choi Seung-Hoan, Mateo Ignacio, Hughes Joseph T., Adams Hieab H., Malamon John, Florentino Sanchez-Garcia Yogen Patel, Brody Jennifer A., Dombroski Beth A., Naranjo Maria Candida Deniz, Daniilidou Makrina, Eiriksdottir Gudny, Mukherjee Shubhabrata, Wallon David, Uphill James, Aspelund Thor, Cantwell Laura B., Garzia Fabienne, Galimberti Daniela, Hofer Edith, Butkiewicz Mariusz, Fin Bertrand, Scarpini Elio, Sarnowski Chloe, Bush Will S., Meslage Stéphane, Kornhuber Johannes, White Charles C., Song Yuenjoo, Barber Robert C., Engelborghs Sebastiaan, Sordon Sabrina, Vojnovic Dina, Adams Perrie M., Vandenbergh Rik, Mayhaus Manuel, Cupples L. Adrienne, Albert Marilyn S., De Deyn Peter P., Gu Wei, Himali Jayanadra J., Beekly Duane, Squassina Alessio, Hartmann Annette M., Orellana Adelina, Blacker Deborah, Rodriguez-Rodriguez Eloy, Lovestone Simon, Garcia Melissa E., Doody Rachelle S., Munoz-Fernandez Carmen, Sussams Rebecca, Lin Honghuang, Fairchild Thomas J., Benito Yolanda A., Holmes Clive, Karamuji - o mi Hata, Frosch Matthew P., Thonberg Hakan, Maier Wolfgang, Roshchupkin Gennady, Ghetti Bernardino, Giedraitis Vilmantas, Kawalia Amit, Li Shuo, Huebinger Ryan M., Kilander Lena, Moebus Susanne, Hernández Isabel, Kamboh M. Ilyas, Brundin RoseMarie, Turton James, Yang Qiong, Katz Mindy J., Concari Letizia, Lord Jenny, Beiser Alexa S., Keene C. Dirk, Helisalmi Seppo, Kloszewska Iwona, Kukull Walter A., Koivisto Anne Maria, Lynch Aoibhinn, Tarraga Lluís, Larson Eric B., Haapasalo Annakaisa, Lawlor Brian, Mosley Thomas H., Lipton Richard B., Solfrizzi Vincenzo, Gill Michael, Longstreth WT, Montine Thomas J., Frisardi Vincenzo, Monica Diez-Fairen Fernando Rivadeneira, Petersen Ronald C., Deramecourt Vincent, Alvarez Ignacio, Salani Francesca, Ciaramella Antonio, Boerwinkle Eric, Reiman Eric M., Fievet Nathalie, Rotter Jerome I., Reisch Joan S., Hanon Olivier, Cupidi Chiara, Uitterlinden A. G. Andre, Royall Donald R., Dufouil Carole, Maletta Raffaele Giovanni, de Rojas Itziar, Sano Mary, Brice Alexis, Cecchetti Roberta, St George-Hyslop Peter, Ritchie Karen, Tsolaki Magda, Tsuang Debby W., Dubois Bruno, Craig David, Wu Chuang-Kuo, Soininen Hilikka, Avramidou Despoina, Albin Roger L., Fratiglioni Laura, Germanou Antonia, Apostolova Liana G., Keller Lina, Koutroumani Maria, Arnold Steven E., Panza Francesco, Gkatzima Olympia, Asthana Sanjay, Hannequin Didier, Whitehead Patrice, Atwood Craig S., Caffarra Paolo, Hampel Harald, Quintela Inés, Carracedo Ángel, Lannfelt Lars, Rubinsztein David C., Barnes Lisa L., Pasquier Florence, Frölich Lutz, Barral Sandra, McGuinness Bernadette, Beach Thomas G., Johnston Janet A., Becker James T., Passmore Peter, Bigio Eileen H., Schott Jonathan M., Bird Thomas D., Warren Jason D., Boeve Bradley F., Lupton Michelle K., Bowen James D., Proitsi Petra, Boxer Adam, Powell John F., Burke James R., Kauwe John S. K., Burns Jeffrey M., Mancuso Michelangelo, Buxbaum Joseph D., Bonuccelli Ubaldo, Cairns Nigel J., McQuillin Andrew, Cao Chuanhai, Livingston Gill, Carlson Chris S., Bass Nicholas J., Carlsson Cynthia M., Hardy John, Carney Regina M., Bras Jose, Carrasquillo Minerva M., Guerreiro Rita, Allen Mariet, Chui Helena C., Fisher Elizabeth, Masullo Carlo, Crocco Elizabeth A., DeCarli Charles, Bisceglia Gina, Dick Malcolm, Ma Li, Duara Ranjan, Graff-Radford Neill R., Evans Denis A., Hodges Angela, Faber Kelley M., Scherer Martin, Fallon Kenneth B., Riemenschneider Matthias, Fardo David W., Heun Reinhard, Farlow Martin R., Heike Kölsch Steven Ferris, Leber Markus, Foroud Tatiana M., Heuser Isabella, Galasko Douglas R., Giegling Ina, Gearing Marla, Michael Hüll Daniel H. Geschwind, Gilbert John R., Morris John, Green Robert C., Mayo Kevin, Growdon John H., Feulner Thomas,

Hamilton Ronald L., Harrell Lindy E., Drichel Dmitriy, Honig Lawrence S., Cushion Thomas D., Huentelman Matthew J., Hollingworth Paul, Hulette Christine M., Hyman Bradley T., Marshall Rachel, Jarvik Gail P., Meggy Alun, Abner Erin, Menzies Georgina E., Jin Lee-Way, Leonenko Ganna, Real Luis M., Jun Gyungah R., Baldwin Clinton T., Grozeva Detelina, Karydas Anna, Russo Giancarlo, Kaye Jeffrey A., Kim Ronald, Jessen Frank, Kowall Neil W., Vellas Bruno, Kramer Joel H., Vardy Emma, LaFerla Frank M., Jöckel Karl-Heinz, Lah James J., Dichgans Martin, Leverenz James B., Mann David, Levey Allan I., Pickering-Brown Stuart, and Lieberman Andrew P. 2019. 'Genetic meta-analysis of diagnosed Alzheimer's disease identifies new risk loci and implicates A β , tau, immunity and lipid processing', *Nature Genetics*, 51: 414–30. [PubMed: 30820047]

Lin MK, and Farrer MJ. 2014. 'Genetics and genomics of Parkinson's disease', *Genome Med*, 6: 48. [PubMed: 25061481]

Machiela MJ, and Chanock SJ. 2015. 'LDlink: a web-based application for exploring population-specific haplotype structure and linking correlated alleles of possible functional variants', *Bioinformatics (Oxford, England)*, 31: 3555–7.

Mahoney Emily R., Dumitrescu Logan, Moore Annah M., Cambronero Francis E., De Jager Philip L., Koran Mary Ellen I., Petyuk Vladislav A., Robinson Renā A. S., Goyal Sandeep, Schneider Julie A., Bennett David A., Jefferson Angela L., and Hohman Timothy J.. 2019. 'Brain expression of the vascular endothelial growth factor gene family in cognitive aging and alzheimer's disease', *Molecular Psychiatry*.

Marees Andries T., de Kluiver Hilde, Stringer Sven, Vorspan Florence, Curis Emmanuel, Marie-Claire Cynthia, and Derks Eske M.. 2018. 'A tutorial on conducting genome-wide association studies: Quality control and statistical analysis', *International journal of methods in psychiatric research*, 27: e1608–e08. [PubMed: 29484742]

McCarthy S, Das S, Kretschmar W, Delaneau O, Wood AR, Teumer A, Kang HM, Fuchsberger C, Danecek P, Sharp K, Luo Y, Sidore C, Kwong A, Timpson N, Koskinen S, Vrieze S, Scott LJ, Zhang H, Mahajan A, Veldink J, Peters U, Pato C, van Duijn CM, Gillies CE, Gandin I, Mezzavilla M, Gilly A, Cocca M, Traglia M, Angius A, Barrett JC, Boomsma D, Branham K, Breen G, Brummett CM, Busonero F, Campbell H, Chan A, Chen S, Chew E, Collins FS, Corbin LJ, Smith GD, Dedoussis G, Dorr M, Farmaki AE, Ferrucci L, Forer L, Fraser RM, Gabriel S, Levy S, Groop L, Harrison T, Hattersley A, Holmen OL, Hveem K, Kretzler M, Lee JC, McGue M, Meitinger T, Melzer D, Min JL, Mohlke KL, Vincent JB, Nauck M, Nickerson D, Palotie A, Pato M, Pirastu N, McInnis M, Richards JB, Sala C, Salomaa V, Schlessinger D, Schoenherr S, Slagboom PE, Small K, Spector T, Stambolian D, Tuke M, Tuomilehto J, Van den Berg LH, Van Rheenen W, Volker U, Wijmenga C, Toniolo D, Zeggini E, Gasparini P, Sampson MG, Wilson JF, Frayling T, de Bakker PI, Swertz MA, McCarroll S, Kooperberg C, Dekker A, Altshuler D, Willer C, Iacono W, Ripatti S, Soranzo N, Walter K, Swaroop A, Cucca F, Anderson CA, Myers RM, Boehnke M, McCarthy MI, and Durbin R. 2016. 'A reference panel of 64,976 haplotypes for genotype imputation', *Nat Genet*, 48: 1279–83. [PubMed: 27548312]

McClay JL, Adkins DE, Aberg K, Bukszár J, Khachane AN, Keefe RS, Perkins DO, McEvoy JP, Stroup TS, Vann RE, Beardsley PM, Lieberman JA, Sullivan PF, and van den Oord EJ. 2011. 'Genome-wide pharmacogenomic study of neurocognition as an indicator of antipsychotic treatment response in schizophrenia', *Neuropsychopharmacology*, 36: 616–26. [PubMed: 21107309]

Meng Danyang, Ma Xiaohua, Li Hui, Wu Xuechun, Cao Yongjun, Miao Zhigang, and Zhang Xia. 2020. 'A Role of the Podoplanin-CLEC-2 Axis in Promoting Inflammatory Response After Ischemic Stroke in Mice', *Neurotoxicity Research*.

Montibeller L, and de Belleruche J. 2018. 'Amyotrophic lateral sclerosis (ALS) and Alzheimer's disease (AD) are characterised by differential activation of ER stress pathways: focus on UPR target genes', *Cell stress & chaperones*, 23: 897–912. [PubMed: 29725981]

Nelson Peter T, Dickson Dennis W, Trojanowski John Q, Jack Clifford R, Boyle Patricia A, Arfanakis Konstantinos, Rademakers Rosa, Alafuzoff Irina, Attems Johannes, Brayne Carol, Coyle-Gilchrist Ian T S, Chui Helena C, Fardo David W, Flanagan Margaret E, Halliday Glenda, Hokkanen Suvi R K, Hunter Sally, Jicha Gregory A, Katsumata Yuriko, Kawas Claudia H, Keene C Dirk, Kovacs Gabor G, Kukull Walter A, Levey Allan I, Makkinejad Nazanin, Montine Thomas J, Murayama Shigeo, Murray Melissa E, Nag Sukriti, Rissman Robert A, Seeley William W,

- Sperling Reisa A, White Charles L III, Lei Yu, and Schneider Julie A. 2019. 'Limbic-predominant age-related TDP-43 encephalopathy (LATE): consensus working group report', *Brain*, 142: 1503–27. [PubMed: 31039256]
- Nelson Peter T., Estus Steven, Abner Erin L., Parikh Ishita, Malik Manasi, Neltner Janna H., Ighodaro Eseosa, Wang Wang-Xia, Wilfred Bernard R., Wang Li-San, Kukull Walter A., Nandakumar Kannabiran, Farman Mark L., Poon Wayne W., Corrada Maria M., Kawas Claudia H., Cribbs David H., Bennett David A., Schneider Julie A., Larson Eric B., Crane Paul K., Valladares Otto, Schmitt Frederick A., Kryscio Richard J., Jicha Gregory A., Smith Charles D., Scheff Stephen W., Sonnen Joshua A., Haines Jonathan L., Pericak-Vance Margaret A., Mayeux Richard, Farrer Lindsay A., Van Eldik Linda J., Horbinski Craig, Green Robert C., Gearing Marla, Poon Leonard W., Kramer Patricia L., Woltjer Randall L., Montine Thomas J., Partch Amanda B., Rajic Alexander J., Richmire KatieRose, Monsell Sarah E., Consortium Alzheimer' Disease Genetic, Schellenberg Gerard D., and Fardo David W.. 2014. 'ABCC9 gene polymorphism is associated with hippocampal sclerosis of aging pathology', *Acta neuropathologica*, 127: 825–43. [PubMed: 24770881]
- Neltner JH, Abner EL, Baker S, Schmitt FA, Kryscio RJ, Jicha GA, Smith CD, Hammack E, Kukull WA, Brenowitz WD, Van Eldik LJ, and Nelson PT. 2014. 'Arteriolosclerosis that affects multiple brain regions is linked to hippocampal sclerosis of ageing', *Brain*, 137: 255–67. [PubMed: 24271328]
- Porcellini Elisa, Carbone Ilaria, Ianni Manuela, and Licastro Federico. 2010. 'Alzheimer's disease gene signature says: beware of brain viral infections', *Immunity & Ageing*, 7: 16. [PubMed: 21156047]
- Pruim Randall J., Welch Ryan P., Sanna Serena, Teslovich Tanya M., Chines Peter S., Gliedt Terry P., Boehnke Michael, Abecasis Gonçalo R., and Willer Cristen J.. 2010. 'LocusZoom: regional visualization of genome-wide association scan results', *Bioinformatics (Oxford, England)*, 26: 2336–37.
- Purcell Shaun, Neale Benjamin, Kathe Todd-Brown Lori Thomas, Ferreira Manuel A. R., Bender David, Maller Julian, Sklar Pamela, de Bakker Paul I. W., Daly Mark J., and Sham Pak C.. 2007. 'PLINK: a tool set for whole-genome association and population-based linkage analyses', *American journal of human genetics*, 81: 559–75. [PubMed: 17701901]
- R Core Team. 2021. "R: A Language and Environment for Statistical Computing." In. Vienna, Austria: R Foundation for Statistical Computing.
- Rahimi Jasmin, and Kovacs Gabor G.. 2014. 'Prevalence of mixed pathologies in the aging brain', *Alzheimer's research & therapy*, 6: 82–82.
- Ramasamy A, Trabzuni D, Guelfi S, Varghese V, Smith C, Walker R, De T, Coin L, de Silva R, Cookson MR, Singleton AB, Hardy J, Ryten M, and Weale ME. 2014. 'Genetic variability in the regulation of gene expression in ten regions of the human brain', *Nat Neurosci*, 17: 1418–28. [PubMed: 25174004]
- Sze CI, Su M, Pugazhenti S, Jambal P, Hsu LJ, Heath J, Schultz L, and Chang NS. 2004. 'Down-regulation of WW domain-containing oxidoreductase induces Tau phosphorylation in vitro. A potential role in Alzheimer's disease', *J Biol Chem*, 279: 30498–506. [PubMed: 15126504]
- Teng CC, Yang YT, Chen YC, Kuo YM, and Sze CI. 2013. 'Role of WWOX/WOX1 in Alzheimer's disease pathology and in cell death signaling', *Front Biosci (Schol Ed)*, 5: 72–85. [PubMed: 23277037]
- von Hippel PT 2015. 'The heterogeneity statistic I(2) can be biased in small meta-analyses', *BMC Med Res Methodol*, 15: 35. [PubMed: 25880989]
- Wang H, Yang J, Schneider JA, De Jager PL, Bennett DA, and Zhang HY. 2020. 'Genome-wide interaction analysis of pathological hallmarks in Alzheimer's disease', *Neurobiology of aging*, 93: 61–68. [PubMed: 32450446]
- Wang Xin, Liu Guo-Jun, Gao Qiang, Li Na, and Wang Rui-tao. 2020. 'C-type lectin-like receptor 2 and zonulin are associated with mild cognitive impairment and Alzheimer's disease', *Acta Neurologica Scandinavica*, 141: 250–55. [PubMed: 31715011]
- Xia K, Zhang J, Ahn M, Jha S, Crowley JJ, Szatkiewicz J, Li T, Zou F, Zhu H, Hibar D, Thompson P, Sullivan PF, Styner M, Gilmore JH, and Knickmeyer RC. 2017. 'Genome-wide association

analysis identifies common variants influencing infant brain volumes', *Transl Psychiatry*, 7: e1188. [PubMed: 28763065]

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript

Article Highlights:

- The *WWOX/MAF* locus has been identified as a potentially harboring AD risk variants
- The present study failed to find associations with AD-related endophenotypes
- However, several other non-AD endophenotypes were associated with *WWOX/MAF* variants
- The novel associations were unchanged by adjustment for AD-related endophenotypes

Statement of Verification

The work under consideration here has not been published previously, is not under consideration for publication elsewhere, and its publication is approved by all authors and tacitly by the responsible authorities where the work was carried out. If accepted for publication, it will not be published elsewhere in the same form, in English or in any other language, including electronically without the written consent of the copyright-holder.

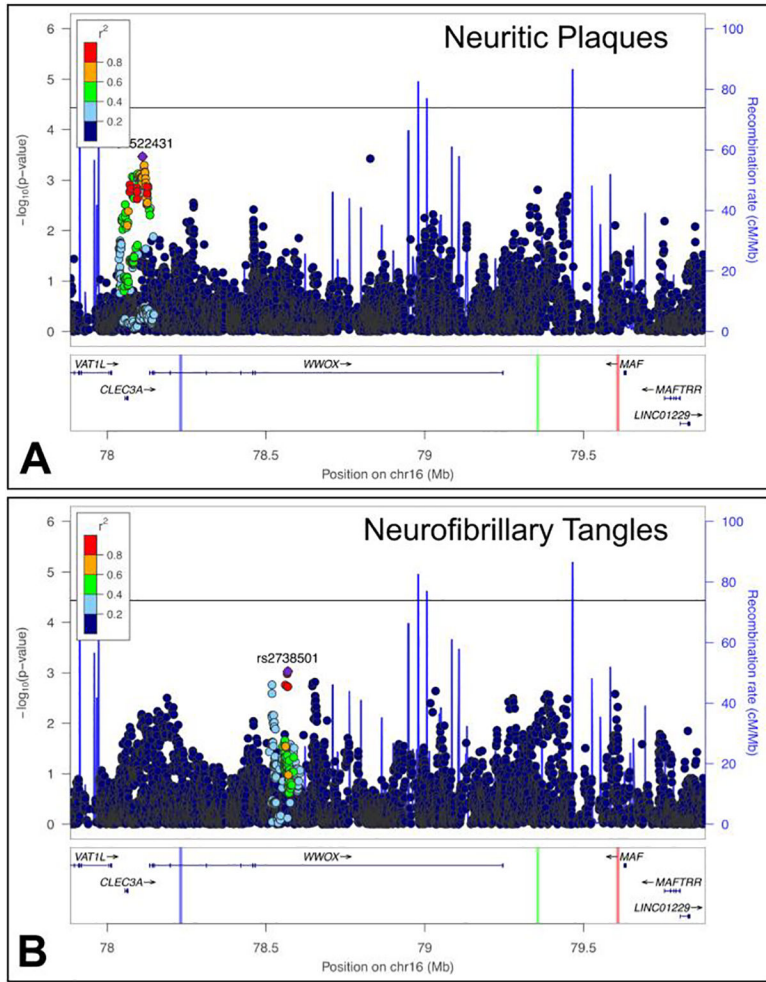


Figure 1: LocusZoom plots of the *WWOX/MAF* region \pm 250kb for **A** neuritic plaques and **B** neurofibrillary tangles, both assuming an additive MOI. Meta-analytic variant-level p-values were adjusted for age at death, sex, Alzheimer’s Disease Genetics Consortium (ADGC) cohort or Religious Orders Study and Memory and Aging Project (ROSMAP) study, and first three genetic principal components and meta-analyzed across the NACC and ROSMAP cohorts. The horizontal line at 4.44 represents the Bonferroni-corrected threshold for significance for the *WWOX/MAF* locus \pm 250kb. The blue region on the gene window highlights the location of rs55751884, the variant previously found to be genome-wide suggestive for HS; the green region on the gene window highlights the location of rs62039712, the variant previously found to be genome-wide significant for clinical AD; and the red region on the gene window highlights the location of rs450674, an *MAF* variant recently found to be associated with clinical AD.

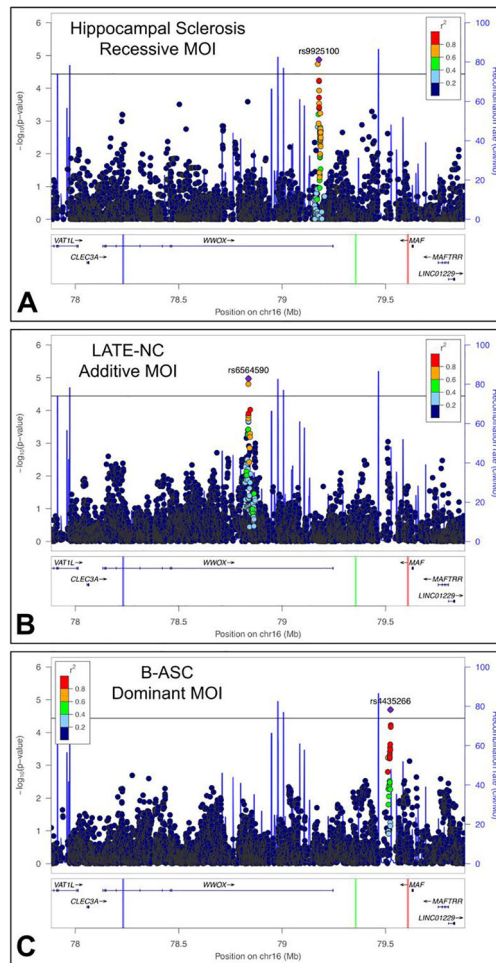


Figure 2: LocusZoom plots of the *WWOX/MAF* region \pm 250kb for **A** hippocampal sclerosis (HS) assuming a recessive mode of inheritance (MOI), **B** limbic-predominant age-related TDP-43 encephalopathy neuropathological changes (LATE-NC) assuming an additive MOI, and **C** brain arteriolosclerosis assuming a dominant MOI. Variant-level p-values were adjusted for age at death, sex, Alzheimer’s Disease Genetics Consortium (ADGC) cohort or Religious Orders Study and Memory and Aging Project (ROSMAP) study, and first three genetic principal components. The horizontal line at 4.44 represents the Bonferroni-corrected threshold for significance for the *WWOX/MAF* locus \pm 250kb. The blue region on the gene window highlights the location of rs55751884, the variant previously found to be genome-wide suggestive for HS; the green region on the gene window highlights the location of rs62039712, the variant previously found to be genome-wide significant for clinical AD; and the red region on the gene window highlights the location of rs450674, an *MAF* variant recently found to be associated with clinical AD.

Table 1:

Individual characteristics stratified by endophenotype status for National Alzheimer's Coordinating Center (NACC) and Religious Orders Study and Rush Memory and Aging Project (ROSMAP) participants.

Endophenotype Status	NACC			ROSMAP		
	Number of Participants (%)	Age at Death, Mean (SD)	Female, N (%)	Number of Participants (%)	Age at Death, Mean (SD)	Female, N (%)
Hippocampal Sclerosis	N=631	85.9 (8.3)	319 (50.6)	N=1200	89.6 (6.5)	812 (67.7)
Absent	542 (85.9)	85.9 (8.4)	270 (49.8)	1091 (90.9)	89.3 (6.5)	729 (66.8)
Present	89 (14.1)	86.0 (7.5)	49 (55.1)	109 (9.1)	92.4 (6.0)	83 (76.1)
LATE-NC	N=412	85.1 (7.9)	207 (50.2)	N=1130	89.8 (6.4)	775 (68.6)
Absent	291 (70.6)	84.9 (8.1)	138 (47.4)	733 (64.9)	88.8 (6.6)	471 (64.3)
Present	121 (29.4)	85.4 (7.3)	69 (57.0)	397 (35.1)	91.8 (5.6)	304 (76.6)
Neurofibrillary Tangles	N=3760	82.5 (8.2)	1939 (51.6)	N=1390	89.4 (6.5)	944 (67.9)
Braak Stage 0 to IV	1236 (32.9)	85.0 (8.4)	639 (51.7)	1046 (75.3)	88.9 (6.7)	679 (64.9)
Braak Stage V or VI	2524 (67.1)	81.2 (7.9)	1300 (51.5)	344 (24.7)	91.0 (5.6)	265 (77.0)
Neuritic Plaques	N=3764	82.5 (8.2)	1940 (51.5)	N=1222	89.5 (6.5)	825 (67.5)
None/Sparse/Moderate	1269 (33.7)	85.9 (8.3)	612 (48.2)	810 (66.3)	89.2 (6.8)	510 (63.0)
Frequent	2495 (66.3)	80.7 (7.6)	1328 (53.2)	412 (33.7)	90.2 (5.8)	315 (76.5)
Brain Arteriolosclerosis	N=2999	82.9 (8.3)	1514 (50.5)	N=1390	89.4 (6.5)	944 (67.9)
None/Mild	1720 (57.4)	81.9 (8.4)	832 (48.4)	1013 (72.9)	89.0 (6.4)	666 (65.7)
Moderate/Severe	1279 (42.6)	84.3 (8.0)	682 (53.3)	377 (27.1)	84.3 (8.0)	278 (73.7)

NACC = National Alzheimer's Coordinating Center; ROSMAP = Religious Orders Study and Rush Memory and Aging Project; SD = standard deviation; LATE-NC = limbic-predominant age-related TDP-43 encephalopathy neuropathological changes.

Table 2:

Adjusted results for previously published HS and clinical AD variants in the *WWOX/MAF* locus. All analyses adjusted for age at death, sex, Alzheimer's Disease Genetics Consortium (ADGC) cohort or Religious Orders Study and Memory and Aging Project (ROSMAP) study, and the first three genetic principal components. The rs55751884 results are reported assuming a recessive mode of inheritance (MOI) as that was the MOI with the strongest association in Nelson *et al.*, 2014. The rs450674 results are reported assuming an additive MOI as that was the MOI reported in Bellenguez *et al.*, 2020.

Variant	Effect Allele	MOI	Endophenotype	NACC			ROSMAP			Meta-Analysis		
				OR	95% CI	P-value	OR	95% CI	P-value	OR	95% CI	P-value
rs55751884	C	Rec.	Hippocampal Sclerosis	5.53	1.51–19.51	0.01133	1.88	0.53–5.21	0.29671	3.04	1.32–6.97	0.00878
			LATE-NC	3.69	0.75–20.26	0.10571	1.21	0.48–2.92	0.67826	1.58	0.73–3.44	0.24823
			Neurofibrillary Tangles	1.66	0.94–3.07	0.08202	0.57	0.21–1.38	0.22405	1.23	0.75–2.02	0.41942
			Neuritic Plaques	2.38	1.32–4.57	0.00330	0.32	0.07–0.93	0.03506	1.58	0.91–2.74	0.10566
			Brain Arteriolosclerosis	1.24	0.73–2.08	0.42069	1.48	0.63–3.32	0.35418	1.30	0.84–2.02	0.23621
rs450674	C	Add.	Hippocampal Sclerosis	0.77	0.48–1.23	0.26429	1.29	0.85–2.00	0.23481	1.02	0.74–1.39	0.91268
			LATE-NC	1.05	0.76–1.46	0.75143	0.97	0.81–1.17	0.75133	0.99	0.84–1.16	0.90518
			Neurofibrillary Tangles	0.93	0.83–1.04	0.20840	0.95	0.79–1.13	0.56609	0.94	0.85–1.03	0.16999
			Neuritic Plaques	0.86	0.78–0.96	0.00637	1.05	0.87–1.27	0.62292	0.90	0.82–0.99	0.03227
			Brain Arteriolosclerosis	0.93	0.84–1.04	0.22636	1.04	0.87–1.25	0.66185	0.96	0.87–1.06	0.41977

NACC = National Alzheimer's Coordinating Center; ROSMAP = Religious Orders Study and Rush Memory and Aging Project; LATE-NC = limbic-predominant age-related TDP-43 encephalopathy neuropathological changes; MOI = mode of inheritance; Rec. = recessive MOI; Add. = additive MOI; OR = odds ratio; and CI = confidence interval.

Table 3:

Variant-level results for variants with uncorrected meta-analytic p-values that met the Bonferroni-corrected threshold for significance for the *WWOX/MAF* locus \pm 250kb. All analyses adjusted for age at death, sex, Alzheimer's Disease Genetics Consortium (ADGC) cohort or Religious Orders Study and Memory and Aging Project (ROSMAP) study, and the first three genetic principal components.

Endophenotype	Variant	Effect Allele	MOI	NACC				ROSMAP				Meta-Analysis		
				Allele Freq.	OR	95% CI	P-value	Allele Freq.	OR	95% CI	P-value	OR	95% CI	P-value
LATE-NC	rs6564590	G	Add.	0.428	2.09	1.50–2.94	8.65×10^{-6}	0.421	1.28	1.07–1.54	0.00783	1.43	1.22–1.68	1.07×10^{-5}
	rs7404901	C	Add.	0.349	1.94	1.38–2.74	0.00011	0.354	1.31	1.09–1.58	0.00435	1.44	1.22–1.69	1.56×10^{-5}
HS	rs9925100	C	Rec.	0.316	2.32	1.19–4.35	0.01481	0.328	2.49	1.48–4.07	0.00087	2.44	1.63–3.61	1.34×10^{-5}
	rs9930659	C	Rec.	0.350	2.87	1.54–5.20	0.00112	0.364	1.98	1.20–3.17	0.00829	2.29	1.57–3.34	1.82×10^{-5}
B-ASC	rs4435266	A	Dom.	0.182	0.75	0.64–0.88	0.00041	0.184	0.71	0.54–0.94	0.01447	0.74	0.64–0.85	2.02×10^{-5}

NACC = National Alzheimer's Coordinating Center; ROSMAP = Religious Orders Study and Rush Memory and Aging Project; HS = hippocampal sclerosis; LATE-NC = limbic-predominant age-related TDP-43 encephalopathy neuropathological changes; B-ASC = brain arteriolosclerosis; MOI = mode of inheritance; Rec. = recessive MOI; Add. = additive MOI; Dom. = dominant MOI; OR = odds ratio; and CI = confidence interval.