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Pulse oximetry plethysmography: a new approach for physiology-directed CPR?

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The overarching aim of cardiopulmonary resuscitation (CPR) is to generate sufficient myocardial and cerebral blood flow to allow for survival with favorable neurologic outcome.¹ The adequacy of myocardial and cerebral blood flow during CPR depends, in part, on force of chest compressions, rate of compressions, chest compression fraction, and allowing full chest recoil for sufficient venous return.^{2–6} Therefore, the core tenets of CPR are to push hard and push fast, minimize interruptions, and allow full chest recoil.⁷ But how hard and fast should we push?

One approach to determine CPR quality is to measure the depth and rate of compressions (i.e., CPR mechanics) in hopes that achieving mechanical targets will provide adequate myocardial and cerebral perfusion. However, variability in chest wall size and compliance and heterogeneity in cardiac arrest etiology and underlying physiology suggest that a “one-size-fits-all” approach may not be optimal.⁸ Not surprisingly, translational animal studies show that physiology-directed CPR titrated to coronary perfusion pressure (CoPP), blood pressure, or end-tidal carbon dioxide (ETCO₂) can improve myocardial and cerebral blood flow and result in superior survival rates and neurologic outcomes.^{9–13} Clinical studies confirm that higher CoPP, arterial diastolic blood pressures (DBP), and ETCO₂ during CPR are associated with survival outcomes, thus supporting the premise of physiology-directed CPR.^{1, 14–16} However, measurement of CoPP or DBP requires invasive hemodynamic monitoring, and ETCO₂ measurement is most reliable in patients who are tracheally intubated. These limitations of established physiologic indicators of CPR quality provide the impetus for evaluating additional, non-invasive tools that can be used when CoPP, DBP, or ETCO₂ are unavailable.

Pulse oximeters are widely used clinical monitors that are commonly applied to patients both in and out of the hospital to measure the oxygenation of peripheral blood. Independent of pulse oximetry's utility for determining oxygen saturation, the pulse oximetry plethysmography (POP) waveform generated by measuring pulsatile blood flow

has clinical utility. For example, the POP waveform may be useful to assess vasomotor tone,¹⁷ volume status^{18, 19} and blood pressure.²⁰ Case reports and small series suggest that POP during CPR may provide clinically relevant information.^{21–23} Promising animal studies indicate that POP can be used both for monitoring CPR quality²⁴ and for identification of return of spontaneous circulation (ROSC) during active CPR.^{25, 26} In a porcine model, Xu et al. demonstrated correlations of POP amplitude (Amp) and POP area under the curve (AUCp) with higher-quality CPR, CoPP, and ETCO₂.²⁷

In this issue of *Resuscitation*, Xu and colleagues build upon their prior translational laboratory work with a large clinical study in which POP was evaluated during CPR to assess its utility in discriminating between patients with and without ROSC.²⁸ They measured POP Amp, AUCp, and ETCO₂ in 150 out-of-hospital cardiac arrest (OHCA) patients and 291 in-hospital cardiac arrest (IHCA) patients across 14 teaching hospitals in China. In the 299 patients with evaluable POP and ETCO₂ data, they found that those with ROSC had higher Amp, AUCp, and ETCO₂ values during CPR than those without ROSC. During the early stage of CPR, POP and ETCO₂ had similar abilities in discriminating between patients with and without ROSC. However, ETCO₂ performed better than POP in the final two minutes of CPR and over the course of the entire resuscitation event. Additionally, the authors proposed cutoff values for POP Amp and AUCp to predict ROSC.

There are important limitations to the study that must be considered in interpreting and applying its results. Most notably, the investigators excluded 585 patients with conditions anticipated to be associated with extremely poor perfusion or low hemoglobin concentrations. This included patients with rib fractures, hemorrhagic shock, pulmonary embolism, pericardial tamponade, severe anemia, and tension pneumothorax without drainage. It was reasonable to exclude these patients in an initial clinical study to enrich the study cohort into one in which POP's potential benefit could be detected. However, the sum of these conditions was common in the patient population. Importantly, these conditions are not always evident to the rescuers providing CPR, and their association with POP efficacy is unclear. Thus, these exclusion criteria limit the generalizability of the data at present. Future prospective studies should attempt to assess POP's utility in broader OHCA and IHCA populations. Additionally, calculating POP Amp and AUCp requires post-event processing algorithms and analyses that are not immediately available at the bedside, thus limiting its current clinical applicability.

Regardless of these limitations, we commend the authors for this exciting exploratory contribution to the literature. The authors first established the physiologic premise of POP use in animal studies, and they now provide initial clinical data to support its role in assessing CPR quality and predicting outcome. Notably, this technology is ubiquitous, cheap, non-invasive, and easily applied, and therefore has the potential to impact nearly every cardiac arrest victim, including when invasive monitoring data is not available. Use of POP in conjunction with other physiologic indicators of CPR quality may better inform providers about multiple intertwined components of cardiac arrest physiology. Although further study is needed to understand how to interpret and apply this technology in real-time, the use of the pulse oximeter to inform physiology-directed CPR carries great promise.

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References:

1. Paradis NA, Martin GB, Rivers EP, et al. Coronary perfusion pressure and the return of spontaneous circulation in human cardiopulmonary resuscitation. *JAMA*. Feb 23 1990;263(8):1106–13. [PubMed: 2386557]
2. Idris AH, Guffey D, Pepe PE, et al. Chest compression rates and survival following out-of-hospital cardiac arrest. *Crit Care Med*. Apr 2015;43(4):840–8. doi:10.1097/CCM.0000000000000824 [PubMed: 25565457]
3. Vadeboncoeur T, Stolz U, Panchal A, et al. Chest compression depth and survival in out-of-hospital cardiac arrest. *Resuscitation*. Feb 2014;85(2):182–8. doi:10.1016/j.resuscitation.2013.10.002 [PubMed: 24125742]
4. Talikowska M, Tohira H, Finn J. Cardiopulmonary resuscitation quality and patient survival outcome in cardiac arrest: A systematic review and meta-analysis. *Resuscitation*. Nov 2015;96:66–77. doi:10.1016/j.resuscitation.2015.07.036 [PubMed: 26247143]
5. Christenson J, Andrusiek D, Everson-Stewart S, et al. Chest compression fraction determines survival in patients with out-of-hospital ventricular fibrillation. *Circulation*. Sep 29 2009;120(13):1241–7. doi:10.1161/CIRCULATIONAHA.109.852202 [PubMed: 19752324]
6. Yannopoulos D, McKnite S, Aufderheide TP, et al. Effects of incomplete chest wall decompression during cardiopulmonary resuscitation on coronary and cerebral perfusion pressures in a porcine model of cardiac arrest. *Resuscitation*. Mar 2005;64(3):363–72. doi:10.1016/j.resuscitation.2004.10.009 [PubMed: 15733767]
7. Meaney PA, Bobrow BJ, Mancini ME, et al. Cardiopulmonary resuscitation quality: [corrected] improving cardiac resuscitation outcomes both inside and outside the hospital: a consensus statement from the American Heart Association. *Circulation*. Jul 23 2013;128(4):417–35. doi:10.1161/CIR.0b013e31829d8654 [PubMed: 23801105]
8. Sainio M, Hoppu S, Huhtala H, Eilevstjonn J, Olkkola KT, Tenhunen J. Simultaneous beat-to-beat assessment of arterial blood pressure and quality of cardiopulmonary resuscitation in out-of-hospital and in-hospital settings. *Resuscitation*. Nov 2015;96:163–9. doi:10.1016/j.resuscitation.2015.08.004 [PubMed: 26310837]
9. Morgan RW, Kilbaugh TJ, Shoap W, et al. A hemodynamic-directed approach to pediatric cardiopulmonary resuscitation (HD-CPR) improves survival. *Resuscitation*. Feb 2017;111:41–47. doi:10.1016/j.resuscitation.2016.11.018 [PubMed: 27923692]
10. Naim MY, Sutton RM, Friess SH, et al. Blood Pressure- and Coronary Perfusion Pressure-Targeted Cardiopulmonary Resuscitation Improves 24-Hour Survival From Ventricular Fibrillation Cardiac Arrest. *Crit Care Med*. Nov 2016;44(11):e1111–e1117. doi:10.1097/CCM.0000000000001859 [PubMed: 27414479]
11. Sutton RM, Friess SH, Naim MY, et al. Patient-centric blood pressure-targeted cardiopulmonary resuscitation improves survival from cardiac arrest. *Am J Respir Crit Care Med*. Dec 1 2014;190(11):1255–62. doi:10.1164/rccm.201407-1343OC [PubMed: 25321490]
12. Hamrick JL, Hamrick JT, Lee JK, Lee BH, Koehler RC, Shaffner DH. Efficacy of chest compressions directed by end-tidal CO2 feedback in a pediatric resuscitation model of basic life support. *J Am Heart Assoc*. Apr 14 2014;3(2):e000450. doi:10.1161/JAHA.113.000450 [PubMed: 24732917]
13. Lautz AJ, Morgan RW, Karlsson M, et al. Hemodynamic-Directed Cardiopulmonary Resuscitation Improves Neurologic Outcomes and Mitochondrial Function in the Heart and Brain. *Crit Care Med*. Mar 2019;47(3):e241–e249. doi:10.1097/CCM.0000000000003620 [PubMed: 30779720]
14. Sheak KR, Wiebe DJ, Leary M, et al. Quantitative relationship between end-tidal carbon dioxide and CPR quality during both in-hospital and out-of-hospital cardiac arrest. *Resuscitation*. Apr 2015;89:149–54. doi:10.1016/j.resuscitation.2015.01.026 [PubMed: 25643651]

15. Paiva EF, Paxton JH, O'Neil BJ. The use of end-tidal carbon dioxide (ETCO₂) measurement to guide management of cardiac arrest: A systematic review. *Resuscitation*. Feb 2018;123:1–7. doi:10.1016/j.resuscitation.2017.12.003 [PubMed: 29217394]
16. Berg RA, Sutton RM, Reeder RW, et al. Association Between Diastolic Blood Pressure During Pediatric In-Hospital Cardiopulmonary Resuscitation and Survival. *Circulation*. Apr 24 2018;137(17):1784–1795. doi:10.1161/CIRCULATIONAHA.117.032270 [PubMed: 29279413]
17. Shelley KH, Murray WB, Chang D. Arterial-pulse oximetry loops: a new method of monitoring vascular tone. *J Clin Monit*. Jul 1997;13(4):223–8. doi:10.1023/a:1007361020825 [PubMed: 9269615]
18. Stewart CL, Mulligan J, Grudic GZ, Talley ME, Jurkovich GJ, Moulton SL. The Compensatory Reserve Index Following Injury: Results of a Prospective Clinical Trial. *Shock*. Sep 2016;46(3 Suppl 1):61–7. doi:10.1097/SHK.0000000000000647 [PubMed: 27172155]
19. Cannesson M, Attof Y, Rosamel P, et al. Respiratory variations in pulse oximetry plethysmographic waveform amplitude to predict fluid responsiveness in the operating room. *Anesthesiology*. Jun 2007;106(6):1105–11. doi:10.1097/01.anes.0000267593.72744.20 [PubMed: 17525584]
20. Talke P, Nichols RJ Jr., Traber DL. Does measurement of systolic blood pressure with a pulse oximeter correlate with conventional methods? *J Clin Monit*. Jan 1990;6(1):5–9. doi:10.1007/BF02832176 [PubMed: 2295896]
21. Griffin M, Cooney C. Pulse oximetry during cardiopulmonary resuscitation. *Anaesthesia*. Nov 1995;50(11):1008. doi:10.1111/j.1365-2044.1995.tb05907.x
22. Narang VP. Utility of the pulse oximeter during cardiopulmonary resuscitation. *Anesthesiology*. Aug 1986;65(2):239–40. doi:10.1097/00000542-198608000-00039
23. Spittal MJ. Evaluation of pulse oximetry during cardiopulmonary resuscitation. *Anaesthesia*. Aug 1993;48(8):701–3. doi:10.1111/j.1365-2044.1993.tb07185.x [PubMed: 8214462]
24. Fu Y, Yin L, Seery S, et al. Pulse rate as an alternative, real-time feedback indicator for chest compression rate: a porcine model of cardiac arrest. *J Clin Monit Comput*. Oct 2021;35(5):1159–1167. doi:10.1007/s10877-020-00576-x [PubMed: 32780354]
25. Li C, Xu J, Han F, et al. Identification of return of spontaneous circulation during cardiopulmonary resuscitation via pulse oximetry in a porcine animal cardiac arrest model. *J Clin Monit Comput*. Oct 2019;33(5):843–851. doi:10.1007/s10877-018-0230-4 [PubMed: 30498975]
26. Wijshoff RW, van der Sar T, Peeters WH, et al. Detection of a spontaneous pulse in photoplethysmograms during automated cardiopulmonary resuscitation in a porcine model. *Resuscitation*. Nov 2013;84(11):1625–32. doi:10.1016/j.resuscitation.2013.07.019 [PubMed: 23907100]
27. Xu J, Li C, Zheng L, et al. Pulse Oximetry: A Non-Invasive, Novel Marker for the Quality of Chest Compressions in Porcine Models of Cardiac Arrest. *PLoS One*. 2015;10(10):e0139707. doi:10.1371/journal.pone.0139707 [PubMed: 26485651]
28. Xu J, Li C, Tang H, et al. Pulse oximetry waveform: a non-invasive physiological predictor for the return of spontaneous circulation in cardiac arrest patients ---- A multicenter, prospective observational study. *Resuscitation*. Oct 5 2021;doi:10.1016/j.resuscitation.2021.09.032