



# Updated Analysis of the Surface Carbohydrate Gene Clusters in a Diverse Panel of *Acinetobacter baumannii* Isolates

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**ABSTRACT** To enhance the utility of the genetically diverse panel of *Acinetobacter baumannii* isolates reported recently by Galac and coworkers (M. R. Galac, E. Snesrud, F. Lebreton, J. Stam, et al., *Antimicrob Agents Chemother* 64:e00840-20, 2020, <https://doi.org/10.1128/AAC.00840-20>) and to identify the novel KL and OCL, all of the gene clusters that direct the biosynthesis of capsular polysaccharide and of the outer core of lipooligosaccharide, respectively, were reexamined. The nine KL and one OCL previously recorded as novel were identified, and nine further novel KL and two OCL were found.

**KEYWORDS** *Acinetobacter baumannii*, K locus, OC locus, diverse panel

**A** *Acinetobacter baumannii* is a leading cause of infections that are difficult to treat because of the high proportion of isolates that exhibit resistance to all or most of the therapeutically effective antibiotics. Consequently, a variety of potential new treatments are being explored. A recent study (1) compiled a genetically diverse panel of 100 non-redundant, sequenced *A. baumannii* clinical isolates that can be used to test and develop potential new therapies. The strain collection has been made publicly available and antibiotic resistance profiles, resistance gene families, and sequence types were recorded. As both vaccination and phage therapy approaches are affected by the specific composition and topology of surface polysaccharides (2, 3), information about the capsular polysaccharide (CPS or capsule) and the outer core (OC) of the lipooligosaccharide, the two surface structures known to exhibit considerable variation between isolates (4–7), is also important. Hence, the specific gene clusters responsible for synthesis of the variable portions of these carbohydrate structures, the K locus (KL) and the OC locus (OCL), found in each isolate were also identified using the recently developed Kaptive tool with *A. baumannii* reference databases (7) and recorded. Genome sequence data were also made available for all isolates in the panel (BioProject [PRJNA545079](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA545079)).

KL are assigned a new number when any difference in the gene content between *fkpA* and *lldP* is detected (5). The most recent published estimate of the diversity at the K locus reported 128 distinct KL (7), although this number continues to rise. However, currently only 92 reference KL sequences are available in the Kaptive database for *A. baumannii*. Consequently, a small proportion of KL and OCL in the diverse panel collection were recorded as novel (Fig. 3 in (1)). However, inaccurate assignments can arise in cases where a sequence differs by only a short segment from an available reference sequence and the closest available match is used.

Here, we have re-examined the KL and OCL in the 100 members of the diverse panel and manually re-examined the relevant DNA sequence in all cases where the Kaptive output was not classified as “perfect.” The novel types were also compared to a larger in-house collection of sequences for K and OC loci.

In the diverse panel, 9 isolates were recorded as carrying a novel KL type (1). Only one of these was found among the KL in the larger in-house database of 128 KL types, and 8 were confirmed as novel. Numbers were assigned to all novel KLs as in Table 1.

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**TABLE 1** Isolates with reassigned KL and OCL types

Isolate name	Assembly accession no.	Previously assigned <sup>a</sup>	Reassigned <sup>b</sup>	GenBank accession number <sup>c</sup>
K locus type				
MRSN 351524	GCA_006492685.1	novel	KL69	OK052595.1
MRSN 7153	GCA_006492295.1	novel	KL151	OK052589.1
MRSN 15574	GCA_006494565.1	novel	KL152	OK052588.1
MRSN 4943	GCA_006492305.1	novel	KL153	OK052587.1
MRSN 10372	GCA_006493855.1	novel	KL154	OK052586.1
MRSN 21660	GCA_006494015.1	novel	KL155	OK052585.1
MRSN 11816	GCA_006494285.1	novel	KL156	OK052584.1
MRSN 15093	GCA_006494575.1	novel	KL157	OK052583.1
MRSN 32076	GCA_006493005.1	novel	KL158	OK052582.1
MRSN 15075	GCA_006494055.1	KL46	KL121	OK052594.1
MRSN 32842	GCA_006492855.1	KL8	KL136	OK052593.1
MRSN 14193	GCA_006494215.1	KL19	KL146	OK052592.1
MRSN 489678	GCA_006492055.1	KL15	KL147	OK052591.1
MRSN 351162	GCA_006492905.1	KL50	KL150	OK052590.1
MRSN 14237	GCA_006494225.1	KL32	KL200-v <sup>*d</sup>	OK052581.1
MRSN 31196	GCA_006491855.1	KL1	KL1-v <sup>*</sup> + GI-1	
OC locus type				
MRSN 423159	GCA_006492625.1	novel	OCL3-v <sup>*</sup>	
MRSN 7113	GCA_006492505.1	OCL5	OCL13	OK052579.1
MRSN 14237	GCA_006494225.1	OCL7	OCL14	OK052580.1
MRSN 32304	GCA_006492945.1	OCL7	OCL14	

<sup>a</sup>Previously assigned in Galac et al. (1).

<sup>b</sup>-v<sup>\*</sup> indicates that locus is found across two or more contigs, suggesting a variant type.

<sup>c</sup>Fully annotated sequence of novel KL or OCL gene cluster.

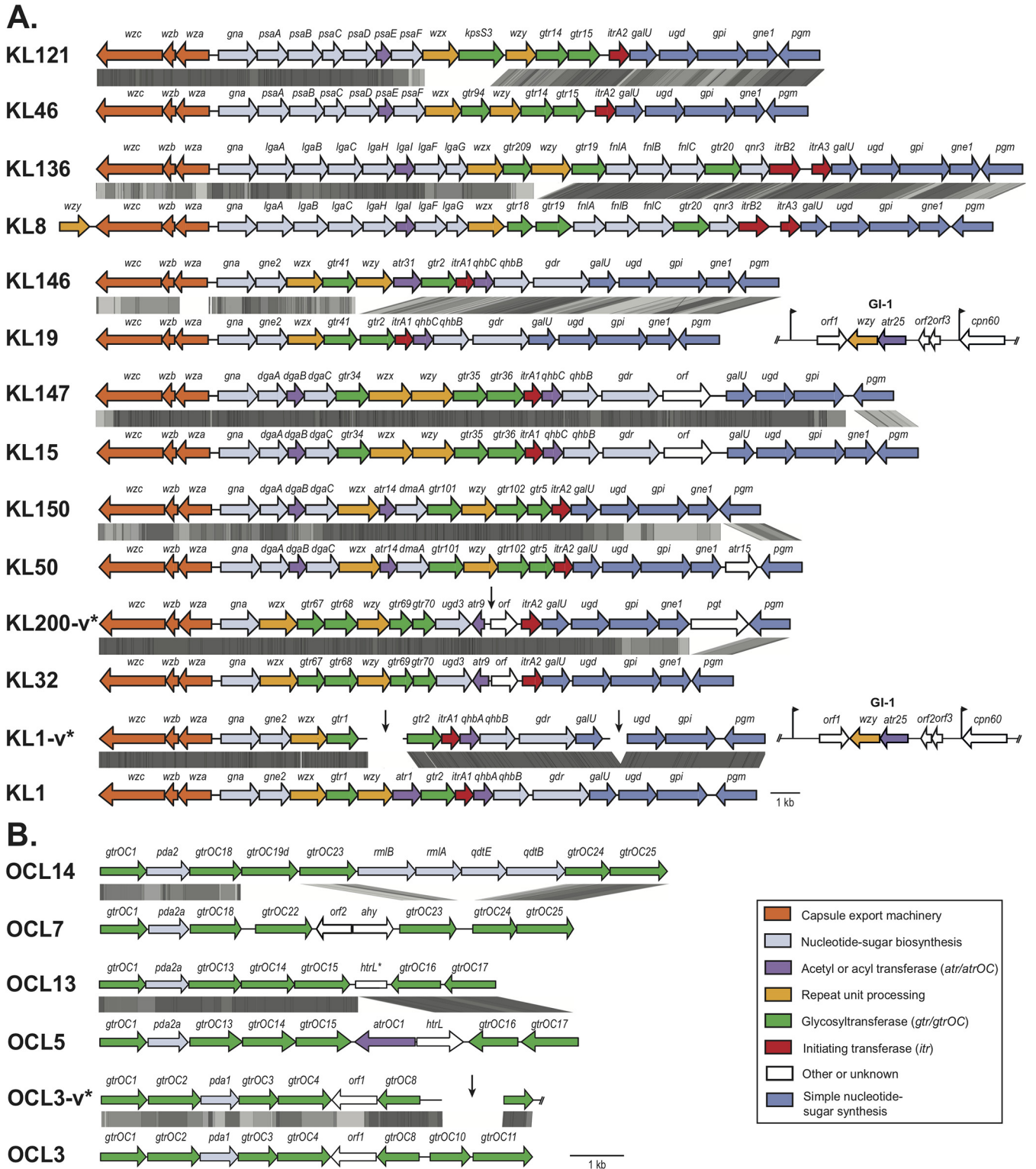
<sup>d</sup>KL200 was found in two contigs that could be directly abutted (see Fig. 1A for position of contig break). A complete sequence with the joined contigs was annotated and submitted to GenBank under accession number [OK052581.1](#).

The sequences of the novel KL were annotated according to the standard nomenclature scheme (5, 7), and GenBank accession numbers for the annotated sequences are listed in Table 1. The organization and gene content of these KL is shown in Fig. S1.

We found that 75 of the 91 reported loci had been correctly identified by Kaptive and a further 10 were potentially correctly identified but interrupted or potentially interrupted by an insertion sequence as the locus was found in two or more contigs (see Table S1). For three of four KL19 and the two KL24 assignments, which rely on a genomic island (GI) to supply the K unit polymerase Wzy (8, 9), the presence of the appropriate GI (GI-1 for KL19, GenBank accession number [KU165787.1](#); and GI-2 for KL24, GenBank accession number [KX756650.1](#)) in the genome was confirmed. The gene cluster in the fourth KL19 assignment was closely related to KL19 but includes a *wzy* gene (Fig. 1A), and the GI was not present in that isolate. In addition, for one isolate assigned as KL1, a segment normally present in KL1 that includes the *wzy* and *atr1* gene was not found (Fig. 1A) but GI-1 was present in the genome of this isolate.

Six further isolates included KL that were closely related to the assigned KL, but closer matches for one was found in our in-house database and five were novel (Table 1). In all six cases, the new KL differed from the assigned KL only in a short region. A comparison of the gene content and organization of the KL originally assigned and the new KL for each of these pairs of KL is shown in Fig. 1A. KL200 was found in two contigs and may be interrupted by an IS. Two of these KL pairs differ only in the presence/absence of genes that are not known to directly affect the capsule structure (shown in white in Fig. 1), and these KL are predicted to produce the same capsule structure. Two of the remaining include different *wzy* genes, and the linkage between the oligomeric units that is formed by the encoded Wzy polymerases is likely to differ, changing the topology of the CPS. These differences are likely to be important as a recent study showed that most phage depolymerases examined target the Wzy linkage (2).

Diversity among the OC loci is more limited than for the K loci, with only 16 types identified in the species to date (6, 7), 12 of which are in the current Kaptive OCL



**FIG 1** Organization of reassigned KL and OCL. (A) The KL gene clusters. (B) The OCL gene clusters. Each reassigned KL or OCL type is compared with the KL or OCL type it was originally assigned to. Figure is drawn to scale with scale bar for KL shown below part A and scale bar for OCL shown below part B. Color scheme (legend bottom right) indicates the functional categories of the gene products. -v\* indicates loci that were found across more than one contig, with downward facing black arrows showing the positions of the breaks. Gray shading indicates the level of protein sequence identity 80 to 100% generated by tblastx comparisons with Easyfig (10).

reference database (7). One OCL found in isolates in the diverse panel was recorded as novel (1), and our re-analysis found that it was a variant of OCL3 that lacked a 1.2 kb internal segment that includes two glycosyltransferase genes (Table 1, Fig. 1A). Here, we found that the OCL in 89 of the remaining 99 isolates were correctly assigned by Kaptive, and a further 7 were potentially correctly identified but found in two or more contigs, suggesting interruption by an insertion sequence (see Table S1). The remaining three OCL cases, one recorded as OCL5 and two assigned as OCL7, were re-assigned to OCL13 or OCL14, respectively (Table 1), which could not have been detected with the current Kaptive database. Fully annotated OCL13 and OCL14 sequences have been deposited in GenBank and their accession numbers are listed in Table 1, and alignments of the two pairs are shown in Fig. 1B. OCL5, OCL7, OCL13, and OCL14 are all members of the OCL B group (6).

The re-analysis of the collection established by Galac et al. (1) has confirmed the utility of the Kaptive tool with current KL and OCL *A. baumannii* reference databases and highlighted some difficulties encountered when the KL or OCL gene cluster is interrupted by an IS or present in more than one contig. The current diverse panel does not include representatives of all KL types, with only 57 KL types (Fig. S1) and 12 OCL types (Fig. S2) represented. However, most of the KL and OCL found in the globally distributed GC1 and GC2 clones (7) that include most of the multiply, extensively, and pan resistant isolates recovered in hospitals were represented. Nonetheless, compilation of a panel that includes greater KL diversity may be warranted. An update of the Kaptive database that includes these newly identified KL and OCL as well as all KL and OCL found in all genomes of *A. baumannii* isolates that are publicly available is pending.

**Data availability.** Fully annotated sequences of novel KL and OCL were deposited into NCBI GenBank under accession numbers listed in Table 1. Genome sequence data for the diverse panel published by Galac et al. (1) are available in NCBI in BioProject [PRJNA545079](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA545079), and individual accession numbers are listed in Table S1.

## SUPPLEMENTAL MATERIAL

Supplemental material is available online only.

**SUPPLEMENTAL FILE 1**, PDF file, 4.2 MB.

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