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Frailty, Aging, and Periodontal Disease: Basic Biological Considerations

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Abstract

Aging is associated with the development of disease. Periodontal disease is one of the many diseases and conditions that increase in prevalence with age. In addition to the traditional focus on individual age-related conditions, there is now a greater recognition that multi-system conditions such as frailty play an important role in the health of older populations. Frailty is a clinical condition in older adults, which increases the risk of adverse health outcomes. Both frailty and periodontal disease are common chronic conditions in older populations and share several risk factors. There is likely a bi-directional relationship between periodontal disease and frailty. Co-morbid systemic diseases, poor physical functioning, and limited ability to self-care in frail older people have been implicated to be underlying the association between frailty and periodontal disease. In addition, both frailty and periodontal disease also have strong associations with inflammatory dysregulation and other age-related pathophysiological changes that may similarly underlie their development and progression. Investigating age-related changes in immune cells that regulate inflammation may lead to a better understanding of age-related disease.

Introduction

There has been a steady increase globally in aging populations over the last few decades. The increase in life expectancy, decrease in births, and improvement of health services have contributed to an increase in older populations. In the US, according to a recent report by the U.S. Department of Health and Human Services, the population aged 65 years and older increased by 34% between 2007 and 2017¹. Additionally, the population aged 85 years and above is projected to increase by 123% by 2040¹. Worldwide, it is projected that by 2050, 1 in 6 people will be aged 65 years and above². Areas showing the highest rate of aging are

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Eastern and South-Eastern Asia, Latin America and the Caribbean². Furthermore, there are gender differences in life expectancy globally, with women living nearly 5 years longer than men².

Aging is also strongly associated with the development of chronic diseases and age-related health conditions, which adversely affect health and quality of life in older people³. It has been reported that older individuals 65 years and above have at least one chronic condition¹. In the US, 38% had 0–1 chronic conditions, whereas 47% had 2–3 chronic conditions. Chronic conditions related to oral health also play a key role in the health of older populations. Periodontal disease is one of the most prevalent chronic diseases globally and becomes more prevalent with increasing age^{4,5}. Proper oral health care is essential in older populations to maintain their quality of life, functional performance, and overall systemic health.

In addition to the traditional focus on single (or related) chronic diseases, there is now a greater recognition that multi-system conditions such as frailty, play an important role in the health of older populations. Frailty is defined as "a clinically recognizable state of increased vulnerability, resulting from aging-associated decline in reserve and function across multiple physiologic systems such that the ability to cope with everyday or acute stressors is compromised"⁶. Frailty is a common clinical syndrome or condition in older adults, which increases the risk of adverse health outcomes, including falls, disability, hospitalization, mortality, and long-term care⁷. Similar to chronic diseases such as periodontal disease, the prevalence of frailty increases dramatically with increasing age.

There has been much research interest over the years in exploring associations of periodontal disease with chronic diseases, particularly cardiovascular disease, type 2 diabetes, and Alzheimer's Disease. More recently, there has also been increasing interest in understanding a similar association of periodontal disease with frailty. Rates of both frailty and periodontal disease are higher in older populations⁸. Co-morbidities, poor physical functioning and limited ability to self-care in frail older people have been implicated to be underlying the association between frailty and dental diseases such as periodontal disease. In addition, both frailty and periodontal disease also have strong associations with inflammation and other age-related pathophysiological changes that may similarly underlie their development and progression.

There are also suggestions that oral diseases, such as periodontal disease, could increase the risk of frailty. Unpacking these bi-directional associations between frailty and periodontal disease merits further research. A better understanding of these associations could lead to improved management and treatment of both frailty and of periodontal disease amongst frail older people. Moreover, understanding the mechanistic biological pathways underlying frailty and periodontal disease will enable richer insights with the potential to improve diagnostics and personalized treatment modalities for older individuals. This manuscript describes the epidemiology and risk factors of frailty and of periodontal disease, and the associations between the two chronic conditions. Additionally, the manuscript will address the molecular and cellular changes that occur with aging and how such pathophysiological

changes may contribute to both frailty and periodontal disease. The manuscript will finally draw together conclusions including implications for research and clinical care.

Epidemiology of frailty

Frailty has been described as "a medical syndrome with multiple causes and contributors that is characterized by diminished strength, endurance, and reduced physiologic function that increases an individual's vulnerability for developing increased dependency and/or death"⁹. A number of assessment tools for frailty have been developed in recent years, including the Fried Frailty Phenotype and the Frailty Index. The Fried Frailty Phenotype defines frailty as a clinical syndrome which is characterized by the following criteria: (1) unintentional weight loss, (2) self-reported exhaustion, (3) weakness (grip strength), (4) slow walking speed and (5) low physical activity¹⁰. Having three or more of these criteria are taken to identify an individual as being frail. Alternatively, the Frailty Index, measures frailty through the accumulation of deficits over time, and it mainly represents "a proxy measure of aging and mortality"¹¹. One important characteristic of the development of frailty is that there is no specific symptom that manifests first and therefore signaling the onset of frailty. However, it has been reported that the manifestation of exhaustion or weight loss can influence significantly the progression to frailty⁶. Furthermore, the manifestation of symptoms and progression of frailty differs between individuals⁶. The prevalence of frailty in older people can vary depending on the definition/instrument that was used¹². According to a large systematic review, the prevalence of frailty in community-dwelling individuals was 10.7%, while 41.6% were reported as pre-frail¹³.

Frailty becomes more common with increasing age^{13-16} . It has been reported that the prevalence of frailty was 4% in those 60-65 years old, 16% in individuals aged 80-84, and 26% in 85 year old community-dwelling populations¹³. Furthermore, women have higher rates of frailty compared to men¹³. Possible explanations are that women have lower muscle mass and strength than men¹⁰, but also women have a higher life expectancy in comparison with men¹³. Furthermore, there are several factors, which can affect the development and progression of frailty in older people. Previous studies have reported that psychological well-being, history of depression and use of antidepressants, diabetes, arthritis, smoking, body mass index (BMI), cognitive impairment, comorbidities and hospitalizations are predictors of incident frailty in older age^{16–20}. Additionally, level of mobility (walking speed), has been associated with development and progression of frailty¹⁵. Older individuals who reported good levels of mobility at baseline were more likely to maintain good physical function and observed no changes in their frailty status over time¹⁵. Also, increased levels of circulating inflammatory mediators are associated with development of frailty in older people (see Inflamm-aging section below). Frailty is also associated with a number of chronic diseases and adverse health outcomes in older people, such as cardiovascular disease, cognitive impairment, disability, falls, and mortality^{21–25}. In other words, frailty could be an indicator of worsening health status and therefore is an important condition to be considered in the management of older people's health.

Epidemiology of periodontal disease in older populations

Periodontal disease is recognized as one of the most common chronic diseases^{3,26}. In 2009–2010, the prevalence of periodontal disease in US adults over 30 years old was $47\%^{26}$. Interestingly, the prevalence differed substantially between young and older individuals, with a prevalence of 24% in 30–34 year old individuals and 70% in those 65 years old²⁶. Some differences in the prevalence of periodontal disease have also been reported by gender, whereby severe periodontal disease was more common in older men than women⁵. Smokers had higher levels of total (severe + mild/moderate) periodontal disease than non-smokers in individuals 65 years old⁵. Periodontal disease is of concern in older adults due to the associated root caries, tooth loss, and worsening masticatory ability, which can further effect, nutrition and speech and impact the patient's quality of life²⁷. The impact of periodontal disease is associated with increased systemic inflammation, cardiovascular disease, poor nutritional status, cognitive impairment, disability and decreased physical function and other comorbidities^{28–36}.

Periodontal disease and its association with frailty in older populations

Research has indicated that there is an association between oral health problems and frailty in older people^{8,37}. It is evident that both frailty and periodontal disease are common chronic conditions in older populations and share several risk factors (Figure 1). Both frailty and periodontal disease are associated with increased chronic systemic disease and both adversely influence quality of life by affecting food habits, physical activity, and functional independence^{7,24,30,34,35}. This section summarizes results from previous studies on the associations between periodontal disease and frailty.

The relationship between frailty and oral health is likely to be bi-directional. Frail older people, particularly those living in long-term care settings have very high levels of dental diseases, including periodontal disease⁸. It is possible that frailty influences and increases the risk of oral diseases in older people. Systemic conditions, such as diabetes mellitus and arthritis are more prevalent in frail older people and are also associated with periodontal disease. These comorbidities may increase the susceptibility of frail older people to periodontal disease⁸. The presence of co-morbidities is compounded by mobility limitations in frail individuals, which may contribute to inadequate oral hygiene and decreased access to professional dental care, which could further increase the risk of periodontal disease⁸.

Studies have also examined the influence of periodontal disease on the risk developing frailty. A number of studies have also focused on tooth loss, chewing difficulties, dental caries, oral pain, occlusion force, and their associations with frailty. However, fewer studies have focused on periodontal disease specifically, and they have reported mixed results. Differences in the assessment of frailty and periodontal disease may contribute to these inconsistent results. In one cross-sectional study of an aging population, individuals with remaining teeth reported better frailty status than those without any natural teeth³⁸ Other cross-sectional studies reported no differences in the prevalence of frailty when individuals with severe periodontal disease were compared to those with non-severe periodontal

disease^{37,39–41}. However, in a longitudinal study, it was reported that severe periodontal disease at baseline, was associated with incident (development of frailty) after 3 years of follow-up⁴². Interestingly, this association remained significant even after adjusting for an interaction term between number of teeth and diabetes⁴². Another longitudinal study with 3 years of follow-up did not report any associations between periodontal disease and incidence of frailty⁴⁰. A recent systematic review of five longitudinal studies demonstrated that oral health status was a predictor of frailty; however, the association of periodontal disease with frailty was inconsistent across the multiple studies⁴³.

Although frailty and periodontal disease in older populations appear to be linked, the direction of the association between the two conditions remains unclear. Nonetheless, there has been increasing awareness of the association of oral health and frailty and a growing appreciation of oral health as an important predictor of frailty⁴³. Studies have highlighted that these two age-related chronic conditions may be associated through similar age-related changes to inflammatory and other biological pathways. An improved understanding of these pathophysiological changes may provide better insight into the management of periodontal disease and frailty.

Biological Considerations underlying periodontal disease and frailty

The association of frailty, aging, and periodontal disease is demonstrated by the epidemiological data as presented in the previous sections. However, a basic biological understanding of the age-related changes that contribute to frailty and periodontal disease is not fully understood. In advancing our biological understanding of aging, much work has been directed at inflammation. Of the many pathophysiological changes that occur with aging, age-related changes to the inflammatory response may be the closest pathophysiological explanation underlying both frailty and periodontal disease.

Inflamm-aging

Inflamm-aging and Frailty

Inflamm-aging describes a chronically elevated and dysregulated inflammatory response that increases with age⁴⁴. It is unknown if inflamm-aging results from a chronic signal that maintains and prolongs the inflammatory response or from a defect in the resolution of the inflammatory response⁴⁵. This systemic inflammatory dysregulation is suspected to directly contribute to or result in a predisposition to the many age-related pathologies including periodontal disease and frailty⁴⁴. Inflamm-aging is generally characterized by elevated levels of circulating inflammatory markers. Circulating levels of interleukin-6 (IL-6), tumor necrosis factor a (TNFa), and C-reactive proteins (CRP) are elevated with increasing age, even in otherwise healthy individuals and in the absence of acute inflammatory stimulus^{46,47}. Increased circulating pro-inflammatory cytokines are also associated with frailty, including interleukin-1 beta (IL-1 β), interleukin-18 (IL-18), interleukin-8 (IL-8), and chemokine (C-X-C) motif ligand 10 (CXCL10)⁴⁸. Both frailty and inflamm-aging are associated with poor health outcomes and an association with similar age-related diseases such as Alzheimer's disease, Type II diabetes, atherosclerosis, and Parkinson's disease⁴⁹. The relationship between inflamm-aging and frailty appears strong. However, studies

have demonstrated conflicting results regarding the association of increased circulating proinflammatory cytokines and frailty. Longitudinal studies have demonstrated that circulating levels of CRP increase with age and higher levels were associated with adverse aging outcomes⁵⁰. Multiple studies have similarly reported that inflammation is a good predictor of frailty but also concluded that inflammation was still increased in those who did not have a diagnosis of frailty and were in otherwise good health^{51,52}. A meta-analysis was completed to better examine the association of frailty and systemic circulating proinflammatory cytokines. Analyzing 32 cross-sectional studies that included over 23,000 older participants, frailty was associated with significantly higher circulating levels of IL-6

and CRP compared to robust (non-frail) older adults⁴⁹. However, they found that when they analyzed the only three longitudinal studies on the topic, levels of IL-6 and CRP were not associated with development of frailty.

Inflamm-aging and Periodontal Disease

The pathogenic processes responsible for the age-related increase in periodontal disease is not fully understood. Periodontal disease arises from a dysregulated or excessive host inflammatory response to the subgingival microbial pathogens. Therefore, it is reasonable to suggest the dysregulated inflammatory response that is characteristic of inflamm-aging may contribute to the pathogenesis of periodontal disease.

Evidence is starting to demonstrate the effect of inflamm-aging within the periodontium. Examining old and young mice, older mice demonstrate increased alveolar bone loss, and the increased bone loss was associated with higher levels of pro-inflammatory cytokine expression within the gingiva compared to young mice⁵³. The increased cytokine expression included IL-1 β and TNFa which are known to promote osteoclastic activity and bone loss in periodontal disease⁵³. In humans, a differential inflammatory response to oral microbiota was demonstrated in an experimental gingivitis study comparing old and young subjects. All subjects were initially treated to achieve similar levels of oral health before abstaining from all oral hygiene practices for a given period of time. At the end of the study period old and young subjects had similar levels of biofilm accumulation; however, the older patients demonstrated more severe gingivitis, suggesting a more pathologic inflammatory response in the older patients⁵⁴.

The effect of inflamm-aging can be characterized locally within various tissues, which can provide insight into how the inflammatory dysregulation contributes to age-related disease. As bone loss is the clinical hallmark of periodontal disease, it would be of interest to better understand how inflamm-aging may directly affect bone. Osteoarthritis, osteoporosis, periodontal disease, and decreased regenerative capacity after injury are some of the age-related diseases and conditions that affect bone health. In osteoarthritis, the age-related increase in pro-inflammatory mediators are suspected to contribute to an increase in damage-associated molecular patterns within the synovial fluid that further drive the innate immune response⁵⁵. The prolonged activation of the innate immune response leads to the production of metalloproteinases leading to the local destruction of tissue and the progression of osteoarthritis⁵⁵. Osteoporosis demonstrates a multifactorial pathogenesis. However, osteoporosis is associated with systemic inflammation in postmenopausal women

as well as being associated with other systemic inflammatory conditions⁵⁶. In osteoporosis, inflamm-aging is suspected to disrupt the homeostatic bone resorption and deposition processes resulting in a shift towards the promotion of bone resorption⁵⁷. Fracture healing capacity is decreased with increasing age⁵⁸. Studies have suggested the decreased healing capacity to be a result of declining skeletal stem cell function and quantity⁵⁹, while others suggest the age-related inflammatory dysregulation perturbs the proceeding anabolic and remodeling processes of fracture healing⁶⁰. One study demonstrated how the age-related inflammatory dysregulation may be directly related to perturbed skeletal stem cell function. In a mouse model, the decreased skeletal stem cell function and quantity was attributed to the local and systemic pro-inflammatory environment, and inhibition of NF- κ B activation in old mice resulted in decreased inflammation and a restoration of skeletal stem cell function and quantity⁶¹.

Cellular and molecular changes of the aging immune response

While inflamm-aging appears to be an important driver of both frailty and periodontal disease, our understanding of what drives inflamm-aging is still limited. Investigating age-related changes in immune cells responsible for the regulation of inflammation may lead to a better understanding of the biology of aging and could lead to therapeutic targets to mitigate the effects of inflamm-aging in frailty and periodontal disease.

The following discusses the known age-related changes to the cells of the innate and adaptive immune system and other cellular and molecular changes that may contribute to the pathogenesis of frailty and periodontal disease. The following also highlights therapeutic strategies that have successfully targeted these age-related changes in human and animal studies (Table 1).

Neutrophil

Neutrophils function as the first line of defense for the innate immune system and mediate the initial host response to invading microorganisms⁶². In periodontal disease, neutrophils migrate to periodontal tissues and the gingival crevice in response to the chemotactic signals of the invading oral microorganisms⁶³. Neutrophils clear microorganisms via phagocytosis; however, the production of toxic reactive oxygen species by neutrophils used to kill microorganisms contributes to the host tissue damage⁶³. Therefore, proper regulation of neutrophil activity is important in the maintenance of periodontal health. Less is known about the effects of aged neutrophils specifically in the context of periodontal disease; however, age-related changes to neutrophils are well characterized in studies examining circulating neutrophils or their bone marrow progenitors.

The total number of neutrophils in the circulation or the neutrophil progenitor cells within the bone marrow are unchanged as a function of age⁶⁴. However, it is not clear if age affects the neutrophil chemotaxis or the migratory response. *In vitro* studies have demonstrated the chemotactic response and the ability of neutrophils to effectively migrate to the site of infection were largely unchanged as a function of age^{65,66} or slightly decreased in old age compared to young control samples^{67,68}. In neutrophils from healthy older human subjects, the chemotactic response to granulocyte-colony stimulating factor (G-CSF) and

N-formyl-Met-Leu-Phe (FMLP) peptide was reduced, but the response to granulocyte macrophage-colony stimulating factor (GM-CSF) and IL-3 was largely unchanged as a function of age^{64,69}. Similarly, molecules involved in the extravasation of neutrophils out of the vessel and into the tissue, such as CD15, CD11a and CD11b have been reported to be unaffected by age or slightly increased^{65,70,71}.

While it appears that age has less of an effect on the number of circulating neutrophils or the ability of the neutrophils to migrate to the site of infection, the antimicrobial activity of neutrophils appears to be affected by age-related changes. Bacterial phagocytosis by neutrophils has been consistently demonstrated to be decreased in samples from healthy old subjects compared to young^{72,73}. Such decreases in phagocytosis have been implicated in the increased susceptibility to bacterial infections in elderly populations. Some studies have suggested that the decreased phagocytosis is a result of decreased recognition of antibodies or complement by neutrophils or from an impairment of FC-mediated phagocytosis^{65,70}. Additionally, the expression of CD16 on neutrophils allows for the recognition of bacteria and induce phagocytosis, and reduced expression of CD16 in response to bacterial stimuli was demonstrated in neutrophils from elderly subjects compared to young⁷⁴. Once phagocytized, it appears age-related changes may also result in a decreased microbial killing ability. Neutrophil microbicidal activity via production of reactive oxygen species and cytotoxic proteases has been reported to be reduced in old neutrophils compared to $young^{68,70,75}$. However, the age-related decrease in superoxide production may be stimulus specific, as one study demonstrated decreased superoxide production in response to Staphylococcus aureus but normal production in response to Escherichia coli by old neutrophils compared to young⁷².

The antimicrobial activities of aged neutrophils may be further compromised by the demonstrated age-related decrease in formation of neutrophil extracellular traps (NETs). The formation of NETs is a form of cell death in neutrophils, known as NETosis, where the nuclear and plasma membrane disintegrate and a decondensed DNA structure is released extracellularly⁷⁶. The NET is able to physically entrap microorganisms in the extracellular space and contains antimicrobial agents, including neutrophil elastase, cathepsin G, and myeloperoxidase, to kill the microorganisms as well as eliminate their virulence factors⁷⁷. As NET formation appears to be an important antimicrobial property of neutrophils, age-related changes affecting NET formation may results in an increase susceptibility to infection. In vitro studies have demonstrated weaker NET formation by neutrophils from old subjects compared to young⁷⁸⁻⁸⁰. In an aging mouse study, the decreased NET formation in old mice was associated with a higher dissemination of Staphylococcus aureus and a more severe infection⁸⁰. The mechanistic defect in age-related decreased NET formation is unclear. One study has suggested that NET formation can be stimulated through activation of Toll-like receptor 2 (TLR2) and that aged neutrophils were less responsive to TLR2 ligands, resulting in a preference for apoptosis over NETosis⁷⁹.

The decreased phagocytic and microbicidal activity of aged neutrophils may be further compounded by their decreased lifespan at the site of infection. Neutrophils have a typical short life-span once recruited to the site of infection but the lifespan can be extended via survival signals that continue to propagate the inflammatory response such as GM-CSF,

GCSF, and LPS⁸¹. However, neutrophils from elderly subjects demonstrated decreased response to such survival signals and thus higher rates of early apoptosis⁸².

In summary, age-related changes to neutrophils may contribute to the pathogenesis of periodontal disease in multiple ways. The antimicrobial properties of the neutrophil are diminished with increasing age. Decreased phagocytosis and decreased NET formation may result in increased susceptibility to the pathogenic microbiota present within the periodontium. Inadequate clearance and the continued presence of the pathogenic microbiota would result in the prolonged propagation of the inflammatory response within the periodontium. In addition, the increased apoptosis of aged neutrophils may further limit the effectiveness of microbial clearance from the periodontium. The resolution of inflammation requires adequate clearance of apoptotic neutrophils from the tissue by other phagocytes as their sustained presence will continue to propagate the inflammatory response. Thus, higher levels of apoptotic neutrophils within the periodontium may result in further inflammatory propagation and a resulting increase in collateral damage of the host tissue.

Macrophage

The macrophage is a key modulator of the innate immune system during infection, disease, or injury. Generally, macrophage phenotypes have been defined along a spectrum of proinflammatory (M1) and anti-inflammatory (M2) phenotypes that participate in distinct processes of inflammatory propagation and resolution^{83,84}. Our understanding of the phenotypic diversity of macrophages is continually growing and new distinct phenotypes are regularly being characterized. Therefore, here we refer to M1 macrophages as generally a pro-inflammatory phenotype and M2 macrophage as generally an anti-inflammatory phenotype without reference to the multitude of diverse phenotypes previously identified.

During periodontal disease, M1 macrophages are presents early during the inflammatory response within the periodontium in response to the presence of subgingival bacterial pathogens. M1 macrophages mediate the host defense against bacteria and propagate the inflammatory response through production of pro-inflammatory cytokines (iNOS, TNF α , IL-1 β , IL-6)^{84,85}. After adequate removal of the bacterial pathogens and their associated pro-inflammatory signaling, a timely phenotypic switch away from M1 is required to prevent collateral tissue damage. The M2 macrophage phenotype acts to resolve inflammation through the production of anti-inflammatory cytokines (IL-10, Arg) and to promote healing of damaged tissues through the production of growth factors (TGF^β, VEGF)⁸⁶. The presence of M1 and M2 type macrophages was shown to be related to increased and decreased severity of periodontal disease respectively^{87,88}. A higher prevalence of M1 macrophages was associated with increased inflammatory cytokines within the gingiva of humans with periodontal disease compared to healthy controls⁸⁹. These findings suggest that the presence of periodontal disease or the severity of disease is a function of the macrophage phenotype, and dysfunctional macrophage activity may contribute to the pathogenesis of periodontal disease.

Age-related changes to the macrophage may perturb the phenotypic presentation and contribute to inflammatory pathologies⁹⁰. An increase in M1-related gene expression was demonstrated in the gingiva of healthy aged non-human primate compared to young control

groups⁹¹. Proinflammatory cytokines secreted by M1 macrophages, such as IL-1β and TNF- α , were also found to be elevated in the gingiva of old mice compared to young⁵³. However, studies evaluating cell autonomous age-related changes to macrophage function and activity have reported conflicting results. Production of pro-inflammatory cytokines by old macrophages compared to young has been reported to be increased⁹², decreased⁹³, or not affected by age⁹⁴. Similarly, phagocytic activity in old macrophages compared to young has been reported to be increased⁹⁵, decreased⁹⁶, or not affected by age⁹⁷. The heterogenous literature on macrophage aging may be a function of a lack of replicability between in vitro and in vivo studies. It is important to consider that the in vivo microenvironment is also affected by age and certainly different than the in vitro cell culture environment utilized in many experiments⁹⁸. Macrophages are of hematopoietic origin with progenitor cells that reside in the bone marrow. Age-related changes to the bone marrow microenvironment have been well characterized and include increased marrow cavity, increased vascular volume, and a shift in endothelial and mesenchymal cell population that support the hematopoietic stem cell niche^{99,100}. Therefore, the aged microenvironment in which macrophages and their progenitor cells occupy may have a strong effect on cell behavior. Similarly, the *in vitro* cell culture environment may also have a strong effect on cell behavior and obscure the intrinsic age-related differences.

Intrinsic age-related changes to the macrophage are becoming more evident using unbiased, next generation sequencing methodologies that don't require exposure of isolated cells to cell culture. Macrophages isolated from a fracture callus in old mice were transcriptionally distinct from the macrophages isolated from a callus of a young mouse as measured by RNAseq⁶⁰. The significant differentially expressed genes in old mice demonstrated increased pro-inflammatory cytokine expression and M1 markers compared to young, and such differences were associated with poor fracture healing outcomes in old mice compared to young⁶⁰. Similar findings were demonstrated in alveolar lung macrophages from young and old mice using single cell RNAseq. Macrophages from the old mice demonstrated increased population heterogeneity with increased pro-inflammatory gene expression¹⁰¹. Other single cell RNAseq studies of the brain have demonstrated that age-related transcriptional changes are not a universal program across all cell types, and that specific cell types, including macrophages demonstrate unique aging profiles distinct from other cells within the same tissue¹⁰².

These age-related changes appear deleterious to the normal macrophage function as evidenced by studies that have directed treatment at aged macrophages. Improved fracture healing was demonstrated in old mice by blocking the macrophage infiltration into the fracture callus⁶⁰. In the same study, there was no effect on fracture healing in young mice with macrophage blockade. In other studies, delayed fracture or cutaneous healing was demonstrated with increasing age, but the age-related healing delay could be rescued with the transplantation of young macrophages or young macrophage progenitors into the old mice^{103,104}.

In summary, a full understanding of the age-related changes to macrophages and its effect on periodontal disease is lacking. However, our current understanding demonstrates age-related changes affecting cytokine secretion, phagocytosis and wound healing, all of which are

essential macrophage processes that could contribute to the pathogenesis of periodontal disease. Treatments directed at aged macrophages have demonstrated some initial support in wound healing and may justify adapting a similar strategy for the treatment of periodontal disease in elderly patients.

T Cells and the adaptive immune response

Activation of the adaptive immune response is generally characterized by the activity of T cells and B cells and their associated production of antibodies and cellular immunity¹⁰⁵. Naïve lymphocytes can recognize antigens and mount a response during infection and memory lymphocytes are primed to allow for a more rapid response to previously experienced antigens. In the periodontium, low levels of T and B cells are evident during health demonstrating a diverse immunoglobulin repertoire within the crevicular fluid in response to the oral microbiota¹⁰⁶. During periodontal disease, a stronger adaptive immune response is mounted and the immunoglobulin repertoire shifts towards a more homogenous response towards the pathogenic microorganisms¹⁰⁶.

Age-related changes to the adaptive immune response have been generally described as immune senescence and has been attributed to the increased susceptibility to infectious disease amongst the elderly. Age-related changes to T and B cells affect their activation and proliferation centrally as well as their recruitment and function locally at the site of inflammation¹⁰⁷. With increased age there is an involution of the thymus, the site of naïve T cell production, resulting in an overall decline in naïve T cell numbers¹⁰⁸. Similarly, decreased generation of progenitor B cells from the hematopoietic environment of the bone marrow is observed with increased age¹⁰⁷. Overall, these age-related declines in naïve lymphocytes result in a decline of antigen-specific immunity and a resulting increased susceptibility to infection in the elderly¹⁰⁵. The immunodeficient status associated with increased age could be expected to result in a susceptibility to subgingival microbial infection during periodontal disease.

As T cells interact with antigen presenting cells, they further differentiate into subpopulations with distinct functions. CD4+ T helper cells are considered a memory T cell and are generally characterized into two subsets based on functional characteristics. Th1 subsets produce pro-inflammatory cytokines that are cytotoxic and propagate the inflammatory response, while the Th2 subset produce largely anti-inflammatory cytokines that are involved in B cell activation and humoral immune response^{109,110}. With increased age, CD4+ T helper cells appear less responsive to receptor stimulation with a resulting decrease in Th2 cell differentiation and a preference towards Th1 differentiation with the associated pro-inflammatory cytokine profile^{111,112}. An increased ratio of Th1 over Th2 cells is present in the periodontium during chronic periodontitis and increased Th1 cells are associated with increased bone resorption¹¹³.

A further subset of Th1 cells that has been implicated in periodontal disease is Th17 cells, named after their characteristic production of the pro-inflammatory cytokine IL-17¹¹⁴. Th17 cell expansion and IL-17 expression are pathogenic mediators in periodontal disease^{115,116}. There are multiple cellular sources of IL-17 within the gingiva; however, during periodontal disease Th17 cells represent 80% of the IL-17+ cells¹¹⁵. Inhibition

of Th17 cell differentiation was shown to decrease periodontal disease severity¹¹⁵. Interestingly, Th17 cells demonstrate increased homeostatic expansion within the gingiva of old mice and humans compared to young controls¹¹⁷. Similar increased Th17 expansion was demonstrated in the circulating blood samples of healthy old human subjects compared to young controls¹¹⁸. Increased Th17 cells expansion was also noted in other age related disease, including corneal epithelial disease where treatment directed at reducing Th17 cell numbers decreased disease severity in aged mouse models¹¹⁹. However, it is unknown what drives this age-related expansion of Th17 cells. Th17 cell expansion is stimulated by IL-6 and IL-23 signaling¹¹⁴. There are multiple cellular sources of IL-6 and IL-23 and macrophages demonstrating the M1-like phenotype secrete both cytokines. The previous section described a shift towards an M1 macrophage phenotype with increasing age. Such age-related changes in the macrophage may drive downstream cellular effectors of the immune response, such as Th17 cells, and further contribute to age-associated inflammatory dysregulation.

In summary, aging affects the adaptive immune response, in part, by reducing the T and B cell naïve repertoire during microbial infection. In addition, aging is characterized by a shift in the T cell phenotypes towards the pro-inflammatory TH1 and TH17 subpopulations, which demonstrate a cytokine profile that promotes bone resorption and contribute to the pathogenesis of periodontal disease.

Cellular Senescence

Cellular senescence defines an exit from the normal cell cycle where the cell is no longer able to replicate but remains resistant to apoptosis¹²⁰. Cellular senescence can be initiated when a cell reaches the end of its replicative capacity or upon exposure to various stressors^{120,121}. The process of cellular senescence appears to be a normal protective measure to prevent malignancies by stopping the proliferation of damaged or premalignant cells¹²². However, senescent cells accumulate within tissue with increasing age and such accumulation is suspected to drive age-related pathologies^{123,124}. It is not clear if the age-related increased accumulation of senescent cells is a result of more cells becoming senescent with increasing age or dysfunctional clearance of senescent cells with increasing age, or both¹²⁵.

The detrimental impact of senescent cells within tissue appears to be a result of its characteristic pro-inflammatory phenotype known as the senescent-associated secretory phenotype (SASP)¹²⁶. The SASP is characterized by the secretion of numerous pro-inflammatory cytokines, chemokines, and other proteins and molecules. The role of the SASP appears to be to maintain the cell cycle arrest in an autocrine manner as well as to induce senescence in neighboring cells in a further effort to prevent malignancy¹²⁷. However, the accumulation and continued production of SASP compounds is detrimental to the surrounding tissues and cells resulting in impaired tissue function and the development of a chronic inflammatory environment¹²⁸. For this reason, cellular senescence is suspected to be a primary diver of inflamm-aging¹²⁹. As inflamm-aging is associated with age-related diseases, senescent cell accumulation has also been associated with a wide variety of chronic inflammatory conditions that increase in prevalence with age. In human and animal studies,

accumulation of senescent cells has been associated with macular degeneration¹³⁰, chronic obstruction pulmonary disease¹³¹, dementia¹³², Parkinson disease¹³³, osteoarthritis¹³⁴, and atherosclerosis¹³⁵.

Less is understood about the contribution of cellular senescence to the pathogenesis of periodontal disease. The SASP is a source of pro-inflammatory cytokines within the tissue and includes pro-inflammatory cytokines (IL-6, IL-1β, TNFa) that are also implicated in periodontal disease^{126,136}. Senescent cells have also been identified within the periodontium. Cellular senescent markers were demonstrated in osteocytes isolated from mouse alveolar bone¹³⁷. In addition, LPS was shown to to induce senescence of alveolar osteocytes in vitro, suggesting a role of bacteria in contributing to senescent cell accumulation within the periodontium¹³⁷. Periodontal derived ligament stem cells (PDLSCs) isolated from older adults were shown to have higher levels of senescent cell markers compared to cells isolated from younger subjects¹³⁸. The older PDLSCs were also shown to be less osteoinductive¹³⁸. Similarly, another study demonstrated that induction of senescence in human periodontal ligament fibroblasts results in decreased osteoblastic differentiation potential *in vitro*¹³⁹. Bone outside of the oral cavity has also been shown to be affected by cellular senescence. Osteoprogenitor cells, osteoblasts, and osteocytes from the bone marrow of old mice had higher levels of senescent markers compared to cells in young mice¹⁴⁰. These findings suggest that senescent cell could accumulate within the periodontium with increasing age and contribute to the pathogenesis of periodontal disease by serving as a source of pro-inflammatory cytokines and limiting the regenerative capacity of the soft and hard tissues of the periodontium.

Senescent cell accumulation within young subjects has been associated with chronic disease. Chronic inflammatory conditions such as obesity¹⁴¹, atherosclerosis¹⁴², and chronic kidney disease¹⁴³ have all demonstrated increased senescent cell accumulation within the affected tissues and organs regardless of age. It is not known if senescent cells accumulate within the periodontium during periodontal disease. If so, periodontal disease at any age could result in the accumulation of senescent cells within the periodontium, which would further increase the susceptibility to periodontal disease in the future. In this manner, treatment of periodontal disease may be best directed at removing the bacterial etiologic agents as well as removing the senescent cells that have accumulated within the tissue.

The role of senescent cells in aging has further been supported by studies demonstrating that the clearance of senescent cells can ameliorate age-related pathologies. The initial studies utilized genetically engineered mouse models that allowed for inducible elimination of senescent cells and demonstrated that removal of senescent cells throughout the lifetime of the animal resulted in the delay of age-related disease and extension of lifespan¹⁴⁴. More recent efforts have been focused on the development of senolytic drugs that attempt to induce apoptosis in senescent cells as a means of limiting their accumulation within tissue. Many studies have further demonstrated the benefit of the removal of senescent cells in multiple disease and tissue-specific contexts. In animal models, the removal of senescent cells was show to prevent age-related bone loss¹⁴⁵, prevent hepatic steatosis¹⁴⁶, and improve cognitive declines in an Alzheimer's model¹⁴⁷. In human trials, Dasatinib and Quercetin have been used in combination as a senolytic treatment. In the first human

trial, patients with idiopathic pulmonary fibrosis, a condition characterized by high levels of senescence cells within lung tissue, received treatment with the senolytic combination and demonstrated improvement in their physical function¹⁴⁸. In another human trial, a cohort of diabetic kidney disease patients also received the Dasatinib and Quercetin senolytic treatment and demonstrated a decreased quantity of senescent cells in multiple different tissues¹⁴⁹. Other senolytic agents have been characterized and tested and there is much interest in understanding the potential benefits of senolytic therapy to treat the myriad of age-related disease. Questions remain about proper administration of senolytic agents as to whether they should be administered throughout the lifetime to prevent senescent cell accumulation, or if administration is best focused as a treatment of chronic inflammatory conditions.

In summary, senescent cells have been shown to accumulate within the periodontium. The associated SASP produces cytokines known to contribute to the pathogenesis of periodontal disease, and senolytic drugs have demonstrated success as a treatment for age-related inflammatory pathologies. Therefore, treatment directed at the removal of senescent cells within the periodontium may be a justified therapeutic approach to managing periodontal disease.

mTOR and aging

To better understand aging, much effort has been placed in trying to identify a more singular process or mediator of aging that similarly changes throughout the organism. For example, epigenetic changes, cellular senescence, telomere shortening, and mitochondrial dysfunction have all been show to increase with age across multiple cell types and organisms and contribute to age-related pathologies¹⁵⁰. Additionally, mammalian target of rapamycin (mTOR) is a protein kinase that has received much attention for its potential role in modulating aging. mTOR consists of an extensive and wide-ranging signaling network involved in protein production, autophagy, and metabolism across all eukaryotic cells¹⁵¹.

It appears the activation of the mTOR pathway stimulates multiple cellular and molecular activities that contribute to aging. For one, activation of mTOR promotes translation and protein production, and an accumulation of misfolded and altered proteins are characteristic of aging¹⁵². Decreased mTOR activity is suspected to decrease the cellular burden of the altered proteins and their harmful metabolic by-products and result in improved cellular function¹⁵². Additionally, mTOR activity promotes the production of transcription factors that regulate pro-inflammatory cytokine synthesis¹⁵². Increased pro-inflammatory cytokine expression is a hallmark of aging as described in the previous sections. An additional hallmark of aging is the accumulation of senescent cells within tissue. Senescent cells demonstrate hyperactivity of the mTOR pathway, which likely contributes to the pathologic increase of pro-inflammatory cytokine production¹⁵³. Finally, autophagy is inversely regulated by mTOR activity, where increased mTOR activity resulted in decreased autophagy. Autophagy is an essential cellular process to recycle and remove degenerated or damaged organelles, and the accumulation of these organelles within the cell can result in mitochondrial damage and cellular dysfunction. Increased mTOR activity results

in decreased autophagy and likely contributes the cellular dysfunction associated with aging¹⁵⁰.

The strongest implication of mTOR activity in the aging process is the broad base of evidence that demonstrates that the inhibition of mTOR activity can increase both lifespan and health span¹⁵⁴. Genetic depletion of mTOR was shown to extend lifespan in flies and mammals^{155,156}. Additionally, treatment with rapamycin, a pharmacologic that inhibits mTOR activity, demonstrates similar improvements in lifespan and health span^{157,158}. Interestingly, there are a few studies that have examined the effect of rapamycin treatment on periodontal disease in aged animal models. It had previously been shown that old mice demonstrate increased alveolar bone loss and an associated increased local pro-inflammatory cytokine expression compared to young mice⁵³. In one study, mice were treated with rapamycin for their entire adult life and periodontal disease was then evaluated when the mice reached old age (35 months). Mice treated with rapamycin demonstrated significantly increased alveolar bone compared to the age-matched controls¹⁵⁹. In another study, old mice (20 month) were treated with a course of rapamycin for only 8 weeks. The short course of treatment resulted in significantly increased alveolar bone compared to age-matched controls, which was also associated with a local decrease of NF-kB pro-inflammatory signaling within the periodontium¹⁶⁰. Interestingly, the same study showed that the short course of rapamycin treatment resulted in new bone formation, as the treated groups had significantly more alveolar bone compared to pre-treatment levels for the same animal (as measured by *in vivo* micro-CT)¹⁶⁰. In summary, these results suggest that targeting mTOR may be a future treatment strategy for periodontal disease in older patient populations. In addition, the findings further implicate periodontal disease as an age-related disease by showing that targeting a dysregulated pathway central to the aging process (mTOR) improves periodontal health.

Conclusions

The association of aging, frailty, and periodontal disease is supported by the epidemiological and basic cellular and molecular research presented here (Figure 1). With a rapidly aging population globally, there is a pressing need to address the challenges of periodontal disease and frailty in older populations. Evidence suggests a bi-directional relationship between frailty and periodontal disease, which remains to be fully understood. Nonetheless, there is a need for improved prevention and management of periodontal disease, particularly in frail older people. Our current understanding provides strong support that the pathophysiological changes that occur with aging contribute to periodontal disease and could explain, in part, the increased prevalence of periodontal disease in frail older populations. An appreciation for the age-related changes that contribute to periodontal disease will lead to expanded translational work developing novel approaches to diagnosis and treatment of periodontal disease. Multiple cellular and molecular targets have been implicated as pathophysiological changes that occur with aging and periodontal disease (Table 1). Animal models have demonstrated proof of principle that treatment directed at the altered physiology of aging can be therapeutically beneficial, and recent clinical trials have shown this approach to be successful in humans. Directing treatment at the specific physiological changes that occur with age could be an important step towards delivering personalized healthcare.

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References

- 1. Administration for Community Living A on A. 2018 Profile of Older Americans. Services USD of H and H, ed. 2018.
- United Nations D of E and SA. World Population Ageing 2019. Highlights. 2019. https://www.un.org/en/development/desa/population/publications/pdf/ageing/ WorldPopulationAgeing2019-Highlights.pdf.
- 3. Kandelman D, Petersen PE, Ueda H. Oral health, general health, and quality of life in older people. Spec Care Dent. 2008;28(6):224–236. doi:10.1111/j.17544505.2008.00045.x
- Marcenes W, Kassebaum NJ, Bernabe E, et al. Global burden of oral conditions in 1990–2010: a systematic analysis. J Dent Res. 2013;92(7):592–597. doi:10.1177/0022034513490168 [PubMed: 23720570]
- 5. Eke PI, Wei L, Borgnakke WS, et al. Periodontitis prevalence in adults 65 years of age, in the USA. Periodontol 2000. 2016;72(1):76–95. doi:10.1111/prd.12145 [PubMed: 27501492]
- 6. Xue Q-L. The Frailty Syndrome: Definition and Natural History. Clin Geriatr Med. 2011;27(1):1– 15. doi:10.1016/j.cger.2010.08.009 [PubMed: 21093718]
- 7. Clegg A, Young J, Iliffe S, Rikkert MO, Rockwood K. Frailty in elderly people. Lancet. 2013;381:752–762. http://www.sciencedirect.com/science/article/pii/S0140673612621679 http:// ac.els-cdn.com/S0140673612621679/1-s2.0-S0140673612621679-main.pdf? _tid=db79f8f6-62f6-11e7-8a46-00000aacb361&acdnat=1499419984_7b6945e59763738607e6aab4 d183b5e7. [PubMed: 23395245]
- 8. van der Putten GJ, De Visschere L, van der Maarel-Wierink C, Vanobbergen J, Schols J. The importance of oral health in (frail) elderly people a review. Eur Geriatr Med. 2013;4(5):339–344. doi:10.1016/j.eurger.2013.07.007
- 9. Morley JE, Vellas B, Abellan van Kan G, et al. Frailty Consensus: A Call to Action. J Am Med Dir Assoc. 2013;14(6):392–397. doi:10.1016/j.jamda.2013.03.022 [PubMed: 23764209]
- Fried LP, Tangen CM, Walston J, et al. Frailty in older adults: Evidence for a phenotype. Journals Gerontol - Ser A Biol Sci Med Sci. 2001. doi:10.1093/gerona/56.3.m146
- Mitnitski AB, Mogilner AJ, Rockwood K. Accumulation of deficits as a proxy measure of aging. ScientificWorldJournal. 2001;1:323–336. doi:10.1100/tsw.2001.58 [PubMed: 12806071]
- Rohrmann S Epidemiology of Frailty in Older People. In: Veronese N, ed. Frailty and Cardiovascular Diseases : Research into an Elderly Population. Cham: Springer International Publishing; 2020:21–27. doi:10.1007/978-3-030-33330-0_3LB-Rohrmann2020
- Collard RM, Boter H, Schoevers RA, Oude Voshaar RC. Prevalence of Frailty in Community-Dwelling Older Persons: A Systematic Review. J Am Geriatr Soc. 2012;60(8):1487–1492. doi:10.1111/j.1532-5415.2012.04054.x [PubMed: 22881367]
- Rockwood K, Mitnitski A. Frailty in Relation to the Accumulation of Deficits. Journals Gerontol Ser A. 2007;62(7):722–727. doi:10.1093/gerona/62.7.722
- 15. Fallah N, Mitnitski A, Searle SD, Gahbauer EA, Gill TM, Rockwood K. Transitions in Frailty Status in Older Adults in Relation to Mobility: A Multistate Modeling Approach Employing a Deficit Count. J Am Geriatr Soc. 2011;59(3):524–529. doi:10.1111/j.1532-5415.2011.03300.x [PubMed: 21391943]
- Hajek A, Brettschneider C, Posselt T, et al. Predictors of frailty in old age–results of a longitudinal study. J Nutr Health Aging. 2016;20(9):952–957. doi:10.1007/s12603-015-0634-5 [PubMed: 27791226]
- Lakey SL, LaCroix AZ, Gray SL, et al. Antidepressant Use, Depressive Symptoms, and Incident Frailty in Women Aged 65 and Older from the Women's Health Initiative Observational Study. J Am Geriatr Soc. 2012;60(5):854–861. doi:10.1111/j.1532-5415.2012.03940.x [PubMed: 22568404]

- Ottenbacher KJ, Graham JE, Al Snih S, et al. Mexican Americans and frailty: findings from the Hispanic established populations epidemiologic studies of the elderly. Am J Public Heal. 2009;99(4):673–679. doi:10.2105/ajph.2008.143958
- Gale CR, Cooper C, Deary IJ, Aihie Sayer A. Psychological well-being and incident frailty in men and women: the English Longitudinal Study of Ageing. Psychol Med. 2014;44(4):697–706. doi:10.1017/S0033291713001384 [PubMed: 23822897]
- Lee JSW, Auyeung T-W, Leung J, Kwok T, Woo J. Transitions in Frailty States Among Community-Living Older Adults and Their Associated Factors. J Am Med Dir Assoc. 2014;15(4):281–286. doi:10.1016/j.jamda.2013.12.002 [PubMed: 24534517]
- Papachristou E, Wannamethee SG, Lennon LT, et al. Ability of Self-Reported Frailty Components to Predict Incident Disability, Falls, and All-Cause Mortality: Results From a Population-Based Study of Older British Men. J Am Med Dir Assoc. 2017;18(2):152–157. doi:10.1016/ j.jamda.2016.08.020 [PubMed: 27742583]
- 22. Soysal P, Arik F, Smith L, Jackson SE, Isik AT. Inflammation, Frailty and Cardiovascular Disease. In: Veronese N, ed. Frailty and Cardiovascular Diseases : Research into an Elderly Population. Cham: Springer International Publishing; 2020:55–64. doi:10.1007/978-3-030-33330-0_7LB-Soysal2020
- Robertson DA, Savva GM, Kenny RA. Frailty and cognitive impairment—A review of the evidence and causal mechanisms. Ageing Res Rev. 2013;12(4):840–851. doi:10.1016/ j.arr.2013.06.004 [PubMed: 23831959]
- Jayanama K, Theou O, Blodgett JM, Cahill L, Rockwood K. Frailty, nutrition-related parameters, and mortality across the adult age spectrum. BMC Med. 2018;16(1):188. doi:10.1186/ s12916-018-1176-6 [PubMed: 30360759]
- 25. Kojima G, Iliffe S, Walters K. Frailty index as a predictor of mortality: a systematic review and meta-analysis. Age Ageing. 2017;47(2):193–200. doi:10.1093/ageing/afx162
- 26. Eke PI, Dye BA, Wei L, Thornton-Evans GO, Genco RJ. Prevalence of Periodontitis in Adults in the United States: 2009 and 2010. J Dent Res. 2012;91(10):914–920. doi:10.1177/0022034512457373 [PubMed: 22935673]
- Boehm TK, Scannapieco FA. The epidemiology, consequences and management of periodontal disease in older adults. J Am Dent Assoc. 2007;138 Suppl:26s–33s. doi:10.14219/ jada.archive.2007.0360 [PubMed: 17761843]
- 28. Kotronia E, Wannamethee SG, Papacosta AO, et al. Poor Oral Health and Inflammatory, Hemostatic, and Cardiac Biomarkers in Older Age: Results From Two Studies in the UK and USA. Journals Gerontol Ser A. 2020. doi:10.1093/gerona/glaa096
- Beck JD, Offenbacher S. Systemic Effects of Periodontitis: Epidemiology of Periodontal Disease and Cardiovascular Disease. J Periodontol. 2005;76(11S):2089–2100. doi:10.1902/jop.2005.76.11-S.2089
- Woelber JP, Bremer K, Vach K, et al. An oral health optimized diet can reduce gingival and periodontal inflammation in humans - a randomized controlled pilot study. BMC Oral Health. 2016;17(1 LB-Woelber2016):28. doi:10.1186/s12903-016-0257-1 [PubMed: 27460471]
- Hamasaki T, Kitamura M, Kawashita Y, Ando Y, Saito T. Periodontal disease and percentage of calories from fat using national data. J Periodontal Res. 2017;52(1):114–121. doi:10.1111/ jre.12375 [PubMed: 27028150]
- 32. Singhrao SK, Harding A, Poole S, Kesavalu L, Crean S. *Porphyromonas gingivalis* Periodontal Infection and Its Putative Links with Alzheimer's Disease. Mediators Inflamm. 2015;2015:137357. doi:10.1155/2015/137357 [PubMed: 26063967]
- Yu Y, Lai Y, Cheung Wai S, Kuo H. Oral Health Status and Self-Reported Functional Dependence in Community-Dwelling Older Adults. J Am Geriatr Soc. 2011;59(3):519–523. doi:doi:10.1111/ j.1532-5415.2010.03311.x [PubMed: 21391942]
- 34. Kotronia E, Wannamethee SG, Papacosta AO, et al. Oral Health, Disability and Physical Function: Results From Studies of Older People in the United Kingdom and United States of America. J Am Med Dir Assoc. 2019. doi:10.1016/j.jamda.2019.06.010

- 35. Hämäläinen P, Rantanen T, Keskinen M, Meurman JH. Oral health status and change in handgrip strength over a 5-year period in 80-year-old people. Gerodontology. 2004;21(3):155–160. doi:10.1111/j.1741-2358.2004.00022.x [PubMed: 15369018]
- 36. Borgnakke WS, Glick M, Genco RJ. Periodontitis: The canary in the coal mine. J Am Dent Assoc. 2013;144(7):764–766. doi:10.14219/jada.archive.2013.0180 [PubMed: 23813251]
- 37. de Andrade FB, Lebrão ML, Santos JLF, de Oliveira Duarte YA. Relationship Between Oral Health and Frailty in Community-Dwelling Elderly Individuals in Brazil. J Am Geriatr Soc. 2013;61(5):809–814. doi:10.1111/jgs.12221 [PubMed: 23647172]
- Hoeksema AR, Peters LL, Raghoebar GM, Meijer HJA, Vissink A, Visser A. Health and quality of life differ between community living older people with and without remaining teeth who recently received formal home care: a cross sectional study. Clin Oral Investig. 2018;22(7):2615–2622. doi:10.1007/s00784-018-2360-y
- Castrejón-Pérez RC, Borges-Yáñez SA, Gutiérrez-Robledo LM, Ávila-Funes JA. Oral health conditions and frailty in Mexican community-dwelling elderly: a cross sectional analysis. BMC Public Health. 2012;12(1):773. doi:10.1186/1471-2458-12-773 [PubMed: 22971075]
- Ramsay SE, Papachristou E, Watt RG, et al. Influence of Poor Oral Health on Physical Frailty: A Population-Based Cohort Study of Older British Men. J Am Geriatr Soc. 2018;66(3):473–479. doi:10.1111/jgs.15175 [PubMed: 29266166]
- Satake A, Kobayashi W, Tamura Y, et al. Effects of oral environment on frailty: particular relevance of tongue pressure. Clin Interv Aging. 2019;14:1643–1648. doi:10.2147/CIA.S212980 [PubMed: 31564844]
- Castrejón-Pérez RC, Jiménez-Corona A, Bernabé E, et al. Oral Disease and 3-Year Incidence of Frailty in Mexican Older Adults. Journals Gerontol Ser A. 2016;72(7):951–957. doi:10.1093/ gerona/glw201
- 43. Hakeem FF, Bernabé E, Sabbah W. Association between oral health and frailty: A systematic review of longitudinal studies. Gerodontology. 2019. doi:10.1111/ger.12406
- 44. FRANCESCHI C, BONAFÈ M, VALENSIN S, et al. Inflamm-aging: An Evolutionary Perspective on Immunosenescence. Ann N Y Acad Sci. 2006. doi:10.1111/j.1749-6632.2000.tb06651.x
- 45. Xia S, Zhang X, Zheng S, et al. An Update on Inflamm-Aging: Mechanisms, Prevention, and Treatment. J Immunol Res. 2016. doi:10.1155/2016/8426874
- 46. Michaud M, Balardy L, Moulis G, et al. Proinflammatory cytokines, aging, and age-related diseases. J Am Med Dir Assoc. 2013. doi:10.1016/j.jamda.2013.05.009
- Puzianowska-Ku nicka M, Owczarz M, Wieczorowska-Tobis K, et al. Interleukin-6 and C-reactive protein, successful aging, and mortality: The PolSenior study. Immun Ageing. 2016. doi:10.1186/ s12979-016-0076-x
- Kane AE, Sinclair DA. Frailty biomarkers in humans and rodents: Current approaches and future advances. Mech Ageing Dev. 2019. doi:10.1016/j.mad.2019.03.007
- 49. Soysal P, Stubbs B, Lucato P, et al. Inflammation and frailty in the elderly: A systematic review and meta-analysis. Ageing Res Rev. 2016. doi:10.1016/j.arr.2016.08.006
- 50. Lassale C, Batty GD, Steptoe A, et al. Association of 10-Year C-Reactive Protein Trajectories with Markers of Healthy Aging: Findings from the English Longitudinal Study of Aging. Journals Gerontol - Ser A Biol Sci Med Sci. 2019. doi:10.1093/gerona/gly028
- Bruunsgaard H, Pedersen M, Pedersen BK. Aging and proinflammatory cytokines. Curr Opin Hematol. 2001. doi:10.1097/00062752-200105000-00001
- 52. Ballou SP, Lozanski GB, Hodder S, et al. Quantitative and qualitative alterations of acute-phase proteins in healthy elderly persons. Age Ageing. 1996. doi:10.1093/ageing/25.3.224
- 53. Liang S, Hosur KB, Domon H, Hajishengallis G. Periodontal inflammation and bone loss in aged mice. J Periodontal Res. 2010. doi:10.1111/j.1600-0765.2009.01245.x
- 54. Fransson C The effect of age on the development of gingivitis Clinical, microbiological and histological findings. J Clin Periodontol. 1996. doi:10.1111/j.1600-051X.1996.tb00561.x
- 55. Millerand M, Berenbaum F, Jacques C. Danger signals and inflammaging in osteoarthritis. Clin Exp Rheumatol. 2019.
- 56. Weitzmann MN, Ofotokun I. Physiological and pathophysiological bone turnover-role of the immune system. Nat Rev Endocrinol. 2016. doi:10.1038/nrendo.2016.91

- 57. Ginaldi L, Mengoli LP, Sirufo MM, De Martinis M. Osteoporosis, Inflammation, and Aging. In: Fulop T, Franceschi C, Hirokawa K, Pawelec G, eds. Handbook of Immunosenescence: Basic Understanding and Clinical Implications. Cham: Springer International Publishing; 2019:2437– 2467. doi:10.1007/978-3-319-99375-1_64
- Clark D, Nakamura M, Miclau T, Marcucio R. Effects of Aging on Fracture Healing. Curr Osteoporos Rep. 2017. doi:10.1007/s11914-017-0413-9
- Oh J, Lee YD, Wagers AJ. Stem cell aging: Mechanisms, regulators and therapeutic opportunities. Nat Med. 2014. doi:10.1038/nm.3651
- 60. Clark D, Brazina S, Yang F, et al. Age-related changes to macrophages are detrimental to fracture healing in mice. Aging Cell. 2020. doi:10.1111/acel.13112
- 61. Josephson AM, Bradaschia-Correa V, Lee S, et al. Age-related inflammation triggers skeletal stem/progenitor cell dysfunction. Proc Natl Acad Sci U S A. 2019. doi:10.1073/pnas.1810692116
- 62. Hasturk H, Kantarci A, Van Dyke TE. Oral inflammatory diseases and systemic inflammation: Role of the macrophage. Front Immunol. 2012. doi:10.3389/fimmu.2012.00118
- Van Dyke TE, Serhan CN. Resolution of inflammation: a new paradigm for the pathogenesis of periodontal diseases. J Dent Res. 2003;82(2):82–90. doi:10.1177/154405910308200202 [PubMed: 12562878]
- 64. Chatta GS, Andrews RG, Rodger E, Schrag M, Hammond WP, Dale DC. Hematopoietic progenitors and aging: alterations in granulocytic precursors and responsiveness to recombinant human G-CSF, GM-CSF, and IL-3. J Gerontol. 1993;48(5):M207–12. doi:10.1093/geronj/ 48.5.m207 [PubMed: 7690056]
- MacGregor RR, Shalit M. Neutrophil function in healthy elderly subjects. Journals Gerontol. 1990. doi:10.1093/geronj/45.2.M55
- 66. Phair JP, Kauffman CA, Bjornson A, Gallagher J, Adams L, Hess EV. Host defenses in the aged: Evaluation of components of the inflammatory and immune responses. J Infect Dis. 1978. doi:10.1093/infdis/138.1.67
- 67. McLaughlin B, O'Malley K, Cotter TG. Age-related differences in granulocyte chemotaxis and degranulation. Clin Sci. 1986. doi:10.1042/cs0700059
- Corberand J, Ngyen F, Laharrague P, et al. Polymorphonuclear functions and aging in humans. J Am Geriatr Soc. 1981;29(9):391–397. doi:10.1111/j.1532-5415.1981.tb02376.x [PubMed: 7264130]
- 69. Hajishengallis G Too old to fight? Aging and its toll on innate immunity. Mol Oral Microbiol. 2010. doi:10.1111/j.2041-1014.2009.00562.x
- 70. Esparza B, Sanchez H, Ruiz M, Barranquero M, Sabino E, Merino F. Neutrophil function in elderly persons assessed by flow cytometry. Immunol Invest. 1996;25(3):185–190. doi:10.3109/08820139609059301 [PubMed: 9157053]
- Rao KMK. Age-related decline in ligand-induced actin polymerization in human leukocytes and platelets. Journals Gerontol. 1986. doi:10.1093/geronj/41.5.561
- 72. Wenisch C, Patruta S, Daxböck F, Krause R, Hörl W. Effect of age on human neutrophil function. J Leukoc Biol. 2000;67(1):40–45. doi:10.1002/jlb.67.1.40 [PubMed: 10647996]
- Mege JL, Capo C, Michel B, Gastaut JL, Bongrand P. Phagocytic cell function in aged subjects. Neurobiol Aging. 1988;9(2):217–220. doi:10.1016/s0197-4580(88)80054-x [PubMed: 2836744]
- 74. Butcher SK, Chahal H, Nayak L, et al. Senescence in innate immune responses: reduced neutrophil phagocytic capacity and CD16 expression in elderly humans. J Leukoc Biol. 2001;70(6):881–886. [PubMed: 11739550]
- 75. Fülöp T, Fóris G, Wórum I, Leövey A. Age-dependent alterations of Fc gamma receptor-mediated effector functions of human polymorphonuclear leucocytes. Clin Exp Immunol. 1985.
- 76. Remijsen Q, Kuijpers TW, Wirawan E, Lippens S, Vandenabeele P, Vanden Berghe T. Dying for a cause: NETosis, mechanisms behind an antimicrobial cell death modality. Cell Death Differ. 2011. doi:10.1038/cdd.2011.1
- 77. Brinkmann V, Reichard U, Goosmann C, et al. Neutrophil Extracellular Traps Kill Bacteria Brinkmann Science 2004.pdf. Science. 2004. doi:10.1126/science.1092385
- Hazeldine J, Harris P, Chapple IL, et al. Impaired neutrophil extracellular trap formation: A novel defect in the innate immune system of aged individuals. Aging Cell. 2014. doi:10.1111/acel.12222

- 79. Xu F, Zhang C, Zou Z, et al. Aging-related Atg5 defect impairs neutrophil extracellular traps formation. Immunology. 2017. doi:10.1111/imm.12740
- 80. Tseng CW, Kyme PA, Arruda A, Ramanujan VK, Tawackoli W, Liu GY. Innate immune dysfunctions in aged mice facilitate the systemic dissemination of methicillin-resistant S. aureus. PLoS One. 2012;7(7):e41454. doi:10.1371/journal.pone.0041454 [PubMed: 22844481]
- Tortorella C, Piazzolla G, Spaccavento F, Pece S, Jirillo E, Antonaci S. Spontaneous and fas-induced apoptotic cell death in aged neutrophils. J Clin Immunol. 1998. doi:10.1023/ A:1023286831246
- 82. F T, F C, A P, et al. Changes in apoptosis of human polymorphonuclear granulocytes with aging. In: Mechanisms of Ageing and Development.; 1997.
- Wynn TA, Chawla A, Pollard JW. Macrophage biology in development, homeostasis and disease. Nature. 2013. doi:10.1038/nature12034
- Yu T, Zhao L, Huang X, et al. Enhanced Activity of the Macrophage M1/M2 Phenotypes and Phenotypic Switch to M1 in Periodontal Infection. J Periodontol. 2016. doi:10.1902/ jop.2016.160081
- Sima C, Glogauer M. Macrophage subsets and osteoimmunology: Tuning of the immunological recognition and effector systems that maintain alveolar bone. Periodontol 2000. 2013. doi:10.1111/ prd.12032
- Biswas SK, Chittezhath M, Shalova IN, Lim JY. Macrophage polarization and plasticity in health and disease. Immunol Res. 2012. doi:10.1007/s12026-012-8291-9
- 87. Garlet GP. Critical reviews in oral biology & medicine: Destructive and protective roles of cytokines in periodontitis: A re-appraisal from host defense and tissue destruction viewpoints. J Dent Res. 2010. doi:10.1177/0022034510376402
- 88. Lappin DF, Macleod CP, Kerr A, Mitchell T, Kinane DF. Anti-inflammatory cytokine IL-10 and T cell cytokine profile in periodontitis granulation tissue. Clin Exp Immunol. 2001. doi:10.1046/ j.1365-2249.2001.01448.x
- 89. Zhou LN, Bi CS, Gao LN, An Y, Chen F, Chen FM. Macrophage polarization in human gingival tissue in response to periodontal disease. Oral Dis. 2019. doi:10.1111/odi.12983
- 90. van Beek AA, Van den Bossche J, Mastroberardino PG, de Winther MPJ, Leenen PJM. Metabolic Alterations in Aging Macrophages: Ingredients for Inflammaging? Trends Immunol. 2019. doi:10.1016/j.it.2018.12.007
- Gonzalez OA, Novak MJ, Kirakodu S, et al. Differential Gene Expression Profiles Reflecting Macrophage Polarization in Aging and Periodontitis Gingival Tissues. Immunol Invest. 2015. doi:10.3109/08820139.2015.1070269
- 92. Mariani E, Pulsatelli L, Neri S, et al. RANTES and MIP-1α production by T lymphocytes, monocytes and NK cells from nonagenarian subjects. Exp Gerontol. 2002. doi:10.1016/ S0531-5565(01)00187-5
- Nyugen J, Agrawal S, Gollapudi S, Gupta S. Impaired functions of peripheral blood monocyte subpopulations in aged humans. J Clin Immunol. 2010. doi:10.1007/s10875-010-9448-8
- 94. Seidler S, Zimmermann HW, Bartneck M, Trautwein C, Tacke F. Age-dependent alterations of monocyte subsets and monocyte-related chemokine pathways in healthy adults. BMC Immunol. 2010. doi:10.1186/1471-2172-11-30
- 95. Lynch AM, Murphy KJ, Deighan BF, et al. The impact of glial activation in the aging brain. Aging Dis. 2010.
- 96. Aprahamian T, Takemura Y, Goukassian D, Walsh K. Ageing is associated with diminished apoptotic cell clearance in vivo. Clin Exp Immunol. 2008. doi:10.1111/j.1365-2249.2008.03658.x
- 97. Fietta A, Merlini C, Dos Santos C, Rovida S, Grassi C. Influence of aging on some specific and nonspecific mechanisms of the host defense system in 146 healthy subjects. Gerontology. 1994. doi:10.1159/000213591
- Linehan E, Dombrowski Y, Snoddy R, Fallon PG, Kissenpfennig A, Fitzgerald DC. Aging impairs peritoneal but not bone marrow-derived macrophage phagocytosis. Aging Cell. 2014. doi:10.1111/ acel.12223
- Pinho S, Frenette PS. Haematopoietic stem cell activity and interactions with the niche. Nat Rev Mol Cell Biol. 2019. doi:10.1038/s41580-019-0103-9

- 100. Frisch BJ, Hoffman CM, Latchney SE, et al. Aged marrow macrophages expand platelet-biased hematopoietic stem cells via interleukin-1B. JCI Insight. 2019. doi:10.1172/jci.insight.124213
- 101. Lafuse WP, Rajaram MVS, Wu Q, et al. Identification of an Increased Alveolar Macrophage Subpopulation in Old Mice That Displays Unique Inflammatory Characteristics and Is Permissive to Mycobacterium tuberculosis Infection. J Immunol. 2019. doi:10.4049/ jimmunol.1900495
- 102. Ximerakis M, Lipnick SL, Innes BT, et al. Single-cell transcriptomic profiling of the aging mouse brain. Nat Neurosci. 2019;22(10):1696–1708. doi:10.1038/s41593-019-0491-3 [PubMed: 31551601]
- 103. Danon D, Kowatch MA, Roth GS. Promotion of wound repair in old mice by local injection of macrophages. Proc Natl Acad Sci U S A. 1989. doi:10.1073/pnas.86.6.2018
- 104. Xing Z, Lu C, Hu D, Miclau T, Marcucio RS. Rejuvenation of the inflammatory system stimulates fracture repair in aged mice. J Orthop Res. 2010. doi:10.1002/jor.21087
- 105. ping Weng N. Aging of the Immune System: How Much Can the Adaptive Immune System Adapt? Immunity. 2006. doi:10.1016/j.immuni.2006.05.001
- 106. Ebersole JL, Dawson DR, Morford LA, Peyyala R, Miller CS, Gonzaléz OA. Periodontal disease immunology: "Double indemnity" in protecting the host. Periodontol 2000. 2013. doi:10.1111/ prd.12005
- 107. Linton PJ, Dorshkind K. Age-related changes in lymphocyte development and function. Nat Immunol. 2004. doi:10.1038/ni1033
- 108. Tosi P, Kraft R, Luzi P, et al. Involution patterns of the human thymus. I Size of the cortical area as a function of age. Clin Exp Immunol. 1982.
- 109. Constant SL, Bottomly K. INDUCTION OF TH1 AND TH2 CD4 + T CELL RESPONSES: The Alternative Approaches. Annu Rev Immunol. 1997. doi:10.1146/annurev.immunol.15.1.297
- 110. Mosmann TR, Cherwinski H, Bond MW, Giedlin MA, Coffman RL. Two types of murine helper T cell clone. I. Definition according to profiles of lymphokine activities and secreted proteins. J Immunol. 1986.
- 111. Franceschi C, Olivieri F, Marchegiani F, et al. Genes involved in immune response/inflammation, IGF1/insulin pathway and response to oxidative stress play a major role in the genetics of human longevity: The lesson of centenarians. In: Mechanisms of Ageing and Development.; 2005. doi:10.1016/j.mad.2004.08.028
- 112. Zhang W, Brahmakshatriya V, Swain SL. CD4 T cell defects in the aged: Causes, consequences and strategies to circumvent. Exp Gerontol. 2014. doi:10.1016/j.exger.2014.01.002
- 113. Taubman MA, Kawai T. Involvement of T-lymphocytes in periodontal disease and in direct and indirect induction of bone resorption. Crit Rev Oral Biol Med. 2001. doi:10.1177/10454411010120020301
- 114. Gaffen SL, Hajishengallis G. A new inflammatory cytokine on the block: Re-thinking periodontal disease and the Th1/Th2 paradigm in the context of Th17 cells and IL-17. J Dent Res. 2008. doi:10.1177/154405910808700908
- 115. Dutzan N, Kajikawa T, Abusleme L, et al. A dysbiotic microbiome triggers TH17 cells to mediate oral mucosal immunopathology in mice and humans. Sci Transl Med. 2018. doi:10.1126/ scitranslmed.aat0797
- 116. Abusleme L, Moutsopoulos NM. IL-17: overview and role in oral immunity and microbiome. Oral Dis. 2017. doi:10.1111/odi.12598
- 117. Dutzan N, Abusleme L, Bridgeman H, et al. On-going Mechanical Damage from Mastication Drives Homeostatic Th17 Cell Responses at the Oral Barrier. Immunity. 2017. doi:10.1016/ j.immuni.2016.12.010
- 118. Schmitt V, Rink L, Uciechowski P. The Th17/Treg balance is disturbed during aging. Exp Gerontol. 2013;48(12):1379–1386. doi:10.1016/j.exger.2013.09.003 [PubMed: 24055797]
- 119. Foulsham W, Mittal SK, Taketani Y, et al. Aged Mice Exhibit Severe Exacerbations of Dry Eye Disease with an Amplified Memory Th17 Cell Response. Am J Pathol. 2020. doi:10.1016/ j.ajpath.2020.03.016
- 120. Kuilman T, Michaloglou C, Mooi WJ, Peeper DS. The essence of senescence. Genes Dev. 2010. doi:10.1101/gad.1971610

- 121. Salama R, Sadaie M, Hoare M, Narita M. Cellular senescence and its effector programs. Genes Dev. 2014. doi:10.1101/gad.235184.113
- 122. Collado M, Serrano M. Senescence in tumours: Evidence from mice and humans. Nat Rev Cancer. 2010. doi:10.1038/nrc2772
- 123. Krishnamurthy J, Torrice C, Ramsey MR, et al. Ink4a/Arf expression is a biomarker of aging. J Clin Invest. 2004. doi:10.1172/JCI22475
- 124. Van Deursen JM. The role of senescent cells in ageing. Nature. 2014. doi:10.1038/nature13193
- 125. Prata LGPL, Ovsyannikova IG, Tchkonia T, Kirkland JL. Senescent cell clearance by the immune system: Emerging therapeutic opportunities. Semin Immunol. 2018. doi:10.1016/ j.smim.2019.04.003
- 126. Coppé J-P, Desprez P-Y, Krtolica A, Campisi J. The Senescence-Associated Secretory Phenotype: The Dark Side of Tumor Suppression. Annu Rev Pathol Mech Dis. 2010. doi:10.1146/annurevpathol-121808-102144
- 127. Nelson G, Wordsworth J, Wang C, et al. A senescent cell bystander effect: Senescence-induced senescence. Aging Cell. 2012. doi:10.1111/j.1474-9726.2012.00795.x
- 128. Rodier F, Campisi J. Four faces of cellular senescence. J Cell Biol. 2011. doi:10.1083/ jcb.201009094
- 129. Zampino M, Ferrucci L, Semba RD. Biomarkers in the path from cellular senescence to frailty. Exp Gerontol. 2020. doi:10.1016/j.exger.2019.110750
- 130. Sun S, Cai B, Li Y, et al. HMGB1 and Caveolin-1 related to RPE cell senescence in age-related macular degeneration. Aging (Albany NY). 2019. doi:10.18632/aging.102039
- 131. Birch J, Barnes PJ, Passos JF. Mitochondria, telomeres and cell senescence: Implications for lung ageing and disease. Pharmacol Ther. 2018. doi:10.1016/j.pharmthera.2017.10.005
- 132. Musi N, Valentine JM, Sickora KR, et al. Tau protein aggregation is associated with cellular senescence in the brain. Aging Cell. 2018. doi:10.1111/acel.12840
- 133. Chinta SJ, Woods G, Demaria M, et al. Cellular Senescence Is Induced by the Environmental Neurotoxin Paraquat and Contributes to Neuropathology Linked to Parkinson's Disease. Cell Rep. 2018. doi:10.1016/j.celrep.2017.12.092
- 134. Hou A, Chen P, Tang H, et al. Cellular senescence in osteoarthritis and anti-aging strategies. Mech Ageing Dev. 2018. doi:10.1016/j.mad.2018.08.002
- 135. Minamino T, Miyauchi H, Yoshida T, Ishida Y, Yoshida H, Komuro I. Endothelial cell senescence in human atherosclerosis: Role of telomere in endothelial dysfunction. Circulation. 2002. doi:10.1161/01.CIR.0000013836.85741.17
- Graves D Cytokines That Promote Periodontal Tissue Destruction. J Periodontol. 2008. doi:10.1902/jop.2008.080183
- 137. Aquino-Martinez R, Rowsey JL, Fraser DG, et al. LPS-induced premature osteocyte senescence: Implications in inflammatory alveolar bone loss and periodontal disease pathogenesis. Bone. 2020. doi:10.1016/j.bone.2019.115220
- 138. Wu RX, Bi CS, Yu Y, Zhang LL, Chen FM. Age-related decline in the matrix contents and functional properties of human periodontal ligament stem cell sheets. Acta Biomater. 2015. doi:10.1016/j.actbio.2015.04.024
- 139. Konstantonis D, Papadopoulou A, Makou M, Eliades T, Basdra EK, Kletsas D. Senescent human periodontal ligament fibroblasts after replicative exhaustion or ionizing radiation have a decreased capacity towards osteoblastic differentiation. Biogerontology. 2013. doi:10.1007/ s10522-013-9449-0
- 140. Farr JN, Fraser DG, Wang H, et al. Identification of Senescent Cells in the Bone Microenvironment. J Bone Miner Res. 2016. doi:10.1002/jbmr.2892
- 141. Villaret A, Galitzky J, Decaunes P, et al. Adipose tissue endothelial cells from obese human subjects: Differences among depots in angiogenic, metabolic, and inflammatory gene expression and cellular senescence. Diabetes. 2010. doi:10.2337/db10-0398
- 142. Roos CM, Zhang B, Palmer AK, et al. Chronic senolytic treatment alleviates established vasomotor dysfunction in aged or atherosclerotic mice. Aging Cell. 2016. doi:10.1111/acel.12458

- 143. Dai L, Qureshi AR, Witasp A, Lindholm B, Stenvinkel P. Early Vascular Ageing and Cellular Senescence in Chronic Kidney Disease. Comput Struct Biotechnol J. 2019. doi:10.1016/ j.csbj.2019.06.015
- 144. Baker DJ, Wijshake T, Tchkonia T, et al. Clearance of p16 Ink4a-positive senescent cells delays ageing-associated disorders. Nature. 2011. doi:10.1038/nature10600
- 145. Farr JN, Xu M, Weivoda MM, et al. Targeting cellular senescence prevents age-related bone loss in mice. Nat Med. 2017. doi:10.1038/nm.4385
- 146. Ogrodnik M, Miwa S, Tchkonia T, et al. Cellular senescence drives age-dependent hepatic steatosis. Nat Commun. 2017. doi:10.1038/ncomms15691
- 147. Zhang P, Kishimoto Y, Grammatikakis I, et al. Senolytic therapy alleviates Aβ-associated oligodendrocyte progenitor cell senescence and cognitive deficits in an Alzheimer's disease model. Nat Neurosci. 2019. doi:10.1038/s41593-019-0372-9
- 148. Justice JN, Nambiar AM, Tchkonia T, et al. Senolytics in idiopathic pulmonary fibrosis: Results from a first-in-human, open-label, pilot study. EBioMedicine. 2019. doi:10.1016/ j.ebiom.2018.12.052
- 149. Hickson LTJ, Langhi Prata LGP, Bobart SA, et al. Senolytics decrease senescent cells in humans: Preliminary report from a clinical trial of Dasatinib plus Quercetin in individuals with diabetic kidney disease. EBioMedicine. 2019. doi:10.1016/j.ebiom.2019.08.069
- 150. Stallone G, Infante B, Prisciandaro C, Grandaliano G. MTOR and aging: An old fashioned dress. Int J Mol Sci. 2019. doi:10.3390/ijms20112774
- 151. Laplante M, Sabatini DM. MTOR signaling in growth control and disease. Cell. 2012. doi:10.1016/j.cell.2012.03.017
- 152. Conn CS, Qian SB. mTOR signaling in protein homeostasis: Less is more? Cell Cycle. 2011. doi:10.4161/cc.10.12.15858
- 153. Carroll B, Nelson G, Rabanal-Ruiz Y, et al. Persistent mTORC1 signaling in cell senescence results from defects in amino acid and growth factor sensing. J Cell Biol. 2017. doi:10.1083/ jcb.201610113
- 154. Liu GY, Sabatini DM. mTOR at the nexus of nutrition, growth, ageing and disease. Nat Rev Mol Cell Biol. 2020. doi:10.1038/s41580-019-0199-y
- 155. Kapahi P, Zid BM, Harper T, Koslover D, Sapin V, Benzer S. Regulation of lifespan in Drosophila by modulation of genes in the TOR signaling pathway. Curr Biol. 2004. doi:10.1016/ j.cub.2004.03.059
- 156. Wu JJ, Liu J, Chen EB, et al. Increased mammalian lifespan and a segmental and tissue-specific slowing of aging after genetic reduction of mTOR expression. Cell Rep. 2013. doi:10.1016/ j.celrep.2013.07.030
- 157. Harrison DE, Strong R, Sharp ZD, et al. Rapamycin fed late in life extends lifespan in genetically heterogeneous mice. Nature. 2009. doi:10.1038/nature08221
- 158. Bitto A, Ito TK, Pineda VV., et al. Transient rapamycin treatment can increase lifespan and healthspan in middle-aged mice. Elife. 2016. doi:10.7554/eLife.16351
- 159. An JY, Quarles EK, Mekvanich S, et al. Rapamycin treatment attenuates age-associated periodontitis in mice. GeroScience. 2017. doi:10.1007/s11357-017-9994-6
- 160. An JY, Kerns KA, Ouellette A, et al. Rapamycin rejuvenates oral health in aging mice. Elife. 2020. doi:10.7554/eLife.54318

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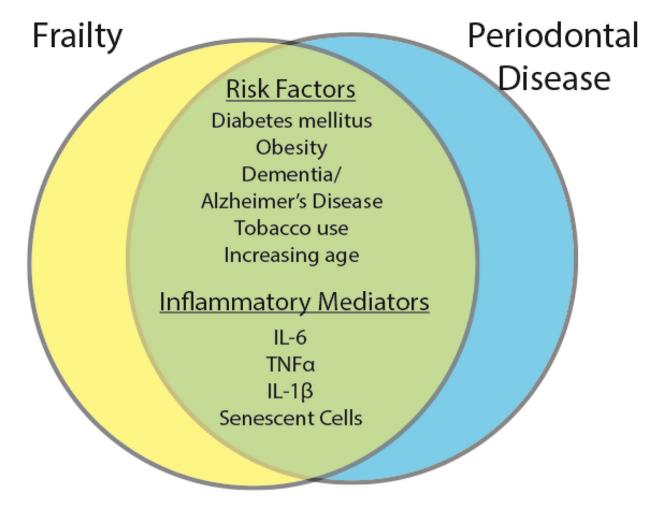


Figure 1:

Shared features of periodontal disease and frailty. Many of the risk factors and comorbid diseases are similarly associated with frailty and periodontal disease. Similarly, systemic and local inflammatory mediators are shared across the two conditions^{48,49,53,136,137}

Table 1:

Therapeutic targeting of age-related changes: Studies reporting a therapeutic benefit by targeting cellular and physiological age-related changes using animal models and human trials.

Target	Disease/Condition	Method	Outcome
Aged infiltrating macrophages (Mouse model)	Fracture healing ⁶⁰	Pharmacologic inhibition of M-CSF	Improved fracture healing in old animals resulting from the inhibition of macrophage infiltration
Aged Macrophages (Mouse model)	Cutaneous Healing ¹⁰³	Transplant of young macrophages into old mice	Acceleration in healing of experimental cutaneous wounds with application of young macrophages in old mice.
Th17 Cells (Mouse model)	Periodontal Disease ¹¹⁵	Pharmacologic inhibition of Th17 cell differentiation	Reduced periodontal disease severity in mice resulting from the reduction of Th17 cell differentiation.
Th17 Cells (Mouse model)	Corneal epithelial disease ¹¹⁹	Antibody treatment	Decreased Th17 cell quantity with antibody treatment resulted in decreased disease severity.
Senescent Cells (Mouse models)	Osteoporosis ¹⁴⁵ Liver disease ¹⁴⁶ Alzheimer's disease ¹⁴⁷	Senolytics	Reduction of senescent cells within tissue resulted in decreased disease severity
Senescent Cells (Human trials)	Idiopathic pulmonary fibrosis ¹⁴⁸ Diabetic kidney disease ¹⁴⁹	Senolytics (Dasatinib + Quercetin)	Decreased disease severity ¹⁴⁸ , and decreased quantity of senescent cells in adipose tissue ¹⁴⁹
mTOR (Mouse models)	Periodontal Disease ^{159,160}	Pharmacologic inhibition of mTOR (Rapamycin)	Decreased disease severity with lifetime of rapamycin administration ¹⁵⁹ . Increased alveolar bone formation with short course of rapamycin in old mice ¹⁶⁰ .