



Great Expectations of COVID-19 Herd Immunity

 Luca T. Giurgea,^a David M. Morens^b

^aLID Clinical Studies Unit, Laboratory of Infectious Diseases, Division of Intramural Research, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Bethesda, Maryland, USA

^bOffice of the Director, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Bethesda, Maryland, USA

ABSTRACT There is a common preconception that reaching an estimated herd immunity threshold through vaccination will end the COVID-19 pandemic. However, the mathematical models underpinning this estimate make numerous assumptions that may not be met in the real world. The protection afforded by vaccines is imperfect, particularly against asymptomatic infection, which can still result in transmission and propagate pandemic viral spread. Immune responses wane and SARS-COV-2 has the capacity to mutate over time to become more infectious and resistant to vaccine elicited immunity. Human behavior and public health restrictions also vary over time and among different populations, impacting the transmissibility of infection. These ever-changing factors modify the number of secondary cases produced by an infected individual, thereby necessitating constant revision of the herd immunity threshold. Even so, vaccination remains a powerful strategy to slow down the pandemic, save lives, and alleviate the burden on limited health care resources.

KEYWORDS coronavirus, epidemiology, herd immunity, immunity, respiratory viruses

The COVID-19 pandemic has had a massive impact on all facets of human life. Despite major public health interventions and development of effective vaccines with unparalleled rapidity, over 290 million cases and 5.4 million deaths have been documented as of January 4th, 2021 (1). As the pandemic continues to abate and resurge globally, individual countries have raced to vaccinate their populations in order to save lives and to relieve strained health care systems, stifled economies, and fatigued populations yearning for return to normalcy. Indeed, immunity elicited by COVID-19 vaccination with BNT162b2, ChAdOx1 nCoV-19, mRNA-1273, Gam-COVID-Vac, Ad26.COV2.S, and CoronaVac vaccines has greatly reduced the incidence of symptomatic disease in individuals (2–8). It is assumed based on public health experience with other infectious diseases that as population immunity rises, as a result of vaccination and natural infection, infections in those who remain susceptible should inevitably decrease.

One approach to disease control has been to immunize populations to reach a (theoretical) “herd immunity threshold,” estimated by some to be approximately 67% (9, 10). The herd immunity threshold is here defined as “the proportion of a population immune to a communicable disease, either from innate immunity, natural infection, or vaccination, that prevents or significantly reduces serial transmission of its infectious agent.” Such thresholds have been predicted mathematically using a transmissibility estimate called the reproductive number (or R_0) in the equation $h = 1 - 1/R_0$ (9, 11). The R_0 is the critical variable in this equation, representing the average number of secondary cases produced by an infected individual in an immunologically susceptible population (11). Initial R_0 estimates varied for COVID-19 within different populations, with most estimates generally ranging from 1.66 to 3.58 (12–15). Social and demographic factors including population density, public health measures, and cultural attitudes and behaviors impact R_0 , inevitably resulting in marked variation of estimates of herd

Editor Jacob Yount, Ohio State University

This is a work of the U.S. Government and is not subject to copyright protection in the United States. Foreign copyrights may apply.

Address correspondence to Luca T. Giurgea, luca.giurgea@nih.gov.

The authors declare no conflict of interest.

Published 25 January 2022

immunity thresholds. Moreover, the herd immunity threshold formula relies on additional assumptions that are unlikely to hold in the case of COVID-19 due to its capacity to mutate, and to the nature of immune responses against nonsystemic respiratory viruses in general, which tend to be incomplete and transient (16–18). Thus, herd immunity threshold estimates should be considered moving targets rather than biologically determined values.

SARS-COV-2, like influenza, is an RNA virus with a high degree of plasticity in its spike protein (S) surface antigen (which elicits protective immune responses) and consequently a potential to rapidly mutate. Host adaptative mutations have been documented in viral variants of concern, including Alpha and Delta (19, 20), which result in higher affinity binding of spike to the human angiotensin-converting enzyme 2 (ACE2) receptor, leading to higher mucosal viral loads and enhanced transmissibility (21, 22). Updated estimates of the reproductive number for variants such as Alpha, Beta, and Gamma have been around 4.7–4.9 (21), and around 5 for the Delta variant (22, 23). Early estimates of Omicron's reproductive number are 4.2 times greater than estimates for Delta (24). Hence, as variants with increasing R_0 values emerge, estimates for the herd immunity threshold will inevitably increase as well.

In addition to host adaptational mutations, viruses have also developed numerous mutations that may allow them to evade host immune responses. Decreased binding of neutralizing antibodies (which are presumed to be correlates of protection) (25) from both convalescent COVID-19 patients and vaccinated individuals, to the Beta, Gamma, and Delta variants has been described (26, 27), and vaccine efficacy has been lower in countries where the Beta and Gamma variants were more prevalent (3, 6). This phenomenon is further exemplified with Omicron, which contains 32 S protein changes contributing to a 27- to 127-fold reduction in neutralization titers relative to wild-type SARS-COV-2. Not surprisingly, it has been spreading explosively in populations with high levels of vaccine- and natural infection-induced immunity (28). While herd immunity threshold formulae assume robust and durable immunity, variants that evade population immunity can change estimates dramatically. Furthermore, waning of detectable anti-S, anti-receptor binding domain (RBD), and neutralizing antibody titers against COVID-19 over time, has occurred in the setting of natural and vaccine-induced immunity, with some half-life estimates ranging from 58 to 106 days (25, 29–31). This is in contrast to immunity against other systemically-infecting respiratory viruses like measles. After recovery from infection, measles immunity is usually lifelong, associated with an estimated antibody half-life as high as 3,014 years (32). Similarly, vaccination with a licensed live measles vaccine provides robust and durable immunity, lasting decades. However, this is not the case with COVID-19, where susceptibility to infection increases with increasing time since vaccination (33–35). Therefore, estimates of herd immunity thresholds must account not only for partial vaccine efficacy but also changes in key parameters associated with a dynamic, mutating virus and with continuously waning immunity.

Asymptomatic infection, which occurs in nearly one third of all COVID-19 cases (36), further complicates the ability to estimate vaccine efficacy and herd immunity thresholds because it perpetuates occult transmission. While an effective vaccine would protect against symptomatic disease, an ideal vaccine would prevent infection entirely, whether symptomatic or not. The effectiveness of COVID-19 vaccines in preventing asymptomatic infection, which may still be associated with transmission, is often lower and not well estimated (37, 38). A retrospective study of health care workers who received the BNT162b2 vaccine determined that its effectiveness in preventing symptomatic disease was 97%, but only 86% in preventing asymptomatic infection (39). Estimates from randomized trials are less optimistic: efficacy against asymptomatic disease of the mRNA-1273 vaccine was only 63% (compared to 93.2% against symptomatic illness), while the efficacy of the ChAdOx1 nCoV-19 vaccine was only 3.8% with the original dosing regimen (3, 40). If the public health goal is to eradicate COVID-19 entirely, the effectiveness of vaccines in preventing both symptomatic and

asymptomatic disease must be considered, since asymptomatic transmission in a pandemic setting can continue indefinitely.

Finally, population heterogeneity must be considered: despite high overall theoretical population immunity, pockets of susceptible individuals can sustain viral circulation, as has been demonstrated with many other viruses including smallpox virus and polioviruses. Although more sophisticated mathematical models have attempted to address imperfect immunity and population heterogeneity (at least from the perspective of social interactions) (41), it is important to consider that persistent circulation of virus in pockets of susceptible hosts can facilitate further mutation and eventually spark additional outbreaks after immunity wanes in the population at large, as seen with vaccine-derived polioviruses (42). In short, mathematical estimates of R_0 and herd immunity thresholds may help guide public health responses but are likely to be poor predictors of epidemic reality and should not replace effective and adaptive public health responses against the evolving pandemic.

Such conclusions are not surprising considering experience with other respiratory viruses. Serologic studies have demonstrated that the vast majority of the human population has been exposed to seasonal coronaviruses, endemic influenza virus strains, and other respiratory viruses like respiratory syncytial virus, but despite eliciting some degree of protective immunity, the viruses continue to circulate (43, 44) and reinfect individuals months or years after initial infection (17, 18, 45). Vaccine approaches that induce more durable immunity and provide broader protection against future variants, so-called “universal vaccines” (46), may alleviate some of the issues associated with herd immunity against respiratory viruses, but such vaccines are still under development.

SARS-CoV-2, like the descendants of the 1918 influenza pandemic, is undoubtedly here to stay. Indeed, all influenza pandemic strains since 1918 have established endemicity after their initial explosive spread. Fortunately, the presence of some degree of immunity against SARS-CoV-2, even immunity that has waned over time or diminished in the face of viral escape mutations, may still provide protection against severe disease and save lives (47, 48). Current vaccine strategies may be able to slow down SARS-CoV-2 spread and are likely to alleviate the burden that waves of severe cases can inflict on limited health care resources, but they are unlikely to lead to SARS-CoV-2 eradication.

Although herd immunity thresholds should not be thought of as precise biological parameters that can predict SARS-CoV-2 control, they may nevertheless have value in setting public health goals and in reminding us that our current pandemic control tools, when used aggressively, can reduce viral circulation, thereby saving lives and reducing illnesses and social-economic disruption.

ACKNOWLEDGMENTS

This research was supported by the Intramural Research Program of the NIH.

REFERENCES

- World Health Organization. 2021. WHO coronavirus (COVID-19) dashboard. World Health Organization, Geneva, Switzerland.
- Polack FP, Thomas SJ, Kitchin N, Absalon J, Gurtman A, Lockhart S, Perez JL, Perez Marc G, Moreira ED, Zerbini C, Bailey R, Swanson KA, Roychoudhury S, Koury K, Li P, Kalina WV, Cooper D, Frenck RW, Jr, Hammitt LL, Tureci O, Nell H, Schaefer A, Unal S, Tresnan DB, Mather S, Dormitzer PR, Sahin U, Jansen KU, Gruber WC, C4591001 Clinical Trial Group. 2020. Safety and efficacy of the BNT162b2 mRNA Covid-19 vaccine. *N Engl J Med* 383:2603–2615. <https://doi.org/10.1056/NEJMoa2034577>.
- Voysey M, Clemens SAC, Madhi SA, Weckx LY, Folegatti PM, Aley PK, Angus B, Baillie VL, Barnabas SL, Bhorat QE, Bibi S, Briner C, Cicconi P, Collins AM, Colin-Jones R, Cutland CL, Darton TC, Dheda K, Duncan CJA, Emary KRW, Ewer KJ, Fairlie L, Faust SN, Feng S, Ferreira DM, Finn A, Goodman AL, Green CM, Green CA, Heath PT, Hill C, Hill H, Hirsch I, Hodgson SHC, Izu A, Jackson S, Jenkin D, Joe CCD, Kerridge S, Koen A, Kwatra G, Lazarus R, Lawrie AM, Lelliott A, Libri V, Lillie PJ, Mallory R, Mendes AVA, Milan EP, Minassian AM, et al. 2021. Safety and efficacy of the ChAdOx1 nCoV-19 vaccine (AZD1222) against SARS-CoV-2: an interim analysis of four randomised controlled trials in Brazil, South Africa, and the UK. *Lancet* 397:99–111. [https://doi.org/10.1016/S0140-6736\(20\)32661-1](https://doi.org/10.1016/S0140-6736(20)32661-1).
- Baden LR, El Sahly HM, Essink B, Kotloff K, Frey S, Novak R, Diemert D, Spector SA, Rouphael N, Creech CB, McGettigan J, Khetan S, Segall N, Solis J, Brosz A, Fierro C, Schwartz H, Neuzil K, Corey L, Gilbert P, Janes H, Follmann D, Marovich M, Mascola J, Polakowski L, Ledgerwood J, Graham BS, Bennett H, Pajon R, Knightly C, Leav B, Deng W, Zhou H, Han S, Ivarsson M, Miller J, Zaks T, Cove Study Group. 2021. Efficacy and safety of the mRNA-1273 SARS-CoV-2 vaccine. *N Engl J Med* 384:403–416. <https://doi.org/10.1056/NEJMoa2035389>.
- Logunov DY, Dolzhikova IV, Shcheblyakov DV, Tukhvatulin AI, Zubkova OV, Dzharullaeva AS, Kovyrshina AV, Lubenets NL, Grousova DM, Erokhova AS, Botikov AG, Izhaeva FM, Popova O, Ozharovskaya TA, Esmagambetov IB, Favorskaya IA, Zrelkin DI, Voronina DV, Shcherbinin DN, Semikhin AS, Simakova YV, Tokarskaya EA, Egorova DA, Shmarov MM, Nikitenko NA, Gushchin VA, Smolyarchuk EA, Zyryanov SK, Borisevich SV, Naroditsky BS, Gintsburg AL, Gam-COVID-Vac Vaccine Trial Group. 2021. Safety and efficacy of an rAd26 and rAd5 vector-based heterologous

- prime-boost COVID-19 vaccine: an interim analysis of a randomised controlled phase 3 trial in Russia. *Lancet* 397:671–681. [https://doi.org/10.1016/S0140-6736\(21\)00234-8](https://doi.org/10.1016/S0140-6736(21)00234-8).
6. Sadoff J, Gray G, Vandebosch A, Cardenas V, Shukarev G, Grinsztejn B, Goepfert PA, Truyers C, Fennema H, Spiessens B, Offergeld K, Scheper G, Taylor KL, Robb ML, Treanor J, Barouch DH, Stoddard J, Ryser MF, Marovich MA, Neuzil KM, Corey L, Cauwenberghs N, Tanner T, Hardt K, Ruiz-Guinazu J, Le Gars M, Schuitemaker H, Van Hoof J, Struyf F, Douoguih M, ENSEMBLE Study Group. 2021. Safety and efficacy of single-dose Ad26.COV2.S vaccine against Covid-19. *N Engl J Med* 384:2187–2201. <https://doi.org/10.1056/NEJMoa2101544>.
 7. Piliishvili T, Gierke R, Fleming-Dutra KE, Farrar JL, Mohr NM, Talan DA, Krishnadasan A, Harland KK, Smithline HA, Hou PC, Lee LC, Lim SC, Moran GJ, Krebs E, Steele MT, Beiser DG, Faine B, Haran JP, Nandi U, Schrading WA, Chinnock B, Henning DJ, Lovecchio F, Lee J, Barter D, Brackney M, Fridkin SK, Marceaux-Galli K, Lim S, Phipps EC, Dumyati G, Pierce R, Markus TM, Anderson DJ, Debes AK, Lin MY, Mayer J, Kwon JH, Saffar N, Fischer M, Singleton R, Chea N, Magill SS, Verani JR, Schrag SJ. 2021. Effectiveness of mRNA Covid-19 vaccine among U.S. Health Care Personnel. *N Engl J Med* 385:e90. <https://doi.org/10.1056/NEJMoa2106599>.
 8. Jara A, Undurraga EA, Gonzalez C, Paredes F, Fontecilla T, Jara G, Pizarro A, Acevedo J, Leo K, Leon F, Sans C, Leighton P, Suarez P, Garcia-Escorza H, Araos R. 2021. Effectiveness of an inactivated SARS-CoV-2 vaccine in Chile. *N Engl J Med* 385:875–884. <https://doi.org/10.1056/NEJMoa2107715>.
 9. Fontanet A, Cauchemez S. 2020. COVID-19 herd immunity: where are we? *Nat Rev Immunol* 20:583–584. <https://doi.org/10.1038/s41577-020-00451-5>.
 10. Randolph HE, Barreiro LB. 2020. Herd immunity: understanding COVID-19. *Immunity* 52:737–741. <https://doi.org/10.1016/j.immuni.2020.04.012>.
 11. Anderson RM, May RM. 1985. Vaccination and herd immunity to infectious diseases. *Nature* 318:323–329. <https://doi.org/10.1038/318323a0>.
 12. Li Q, Guan X, Wu P, Wang X, Zhou L, Tong Y, Ren R, Leung KSM, Lau EHY, Wong JY, Xing X, Xiang N, Wu Y, Li C, Chen Q, Li D, Liu T, Zhao J, Liu M, Tu W, Chen C, Jin L, Yang R, Wang Q, Zhou S, Wang R, Liu H, Luo Y, Liu Y, Shao G, Li H, Tao Z, Yang Y, Deng Z, Liu B, Ma Z, Zhang Y, Shi G, Lam TTY, Wu JT, Gao GF, Cowling BJ, Yang B, Leung GM, Feng Z. 2020. Early transmission dynamics in Wuhan, China, of novel coronavirus-infected pneumonia. *N Engl J Med* 382:1199–1207. <https://doi.org/10.1056/NEJMoa2001316>.
 13. Sy KTL, White LF, Nichols BE. 2021. Population density and basic reproductive number of COVID-19 across United States counties. *PLoS One* 16:e0249271. <https://doi.org/10.1371/journal.pone.0249271>.
 14. Locatelli I, Trachsel B, Rousson V. 2021. Estimating the basic reproduction number for COVID-19 in Western Europe. *PLoS One* 16:e0248731. <https://doi.org/10.1371/journal.pone.0248731>.
 15. Zhao S, Lin Q, Ran J, Musa SS, Yang G, Wang W, Lou Y, Gao D, Yang L, He D, Wang MH. 2020. Preliminary estimation of the basic reproduction number of novel coronavirus (2019-nCoV) in China, from 2019 to 2020: A data-driven analysis in the early phase of the outbreak. *Int J Infect Dis* 92:214–217. <https://doi.org/10.1016/j.ijid.2020.01.050>.
 16. Macnaughton MR. 1982. Occurrence and frequency of coronavirus infections in humans as determined by enzyme-linked immunosorbent assay. *Infect Immun* 38:419–423. <https://doi.org/10.1128/iai.38.2.419-423.1982>.
 17. Petrie JG, Bazzi LA, McDermott AB, Follmann D, Esposito D, Hatcher C, Mateja A, Narpala SR, O'Connell SE, Martin ET, Monto AS. 2021. Coronavirus occurrence in the HIVE cohort of Michigan households: reinfection frequency and serologic responses to seasonal and SARS coronaviruses. *J Infect Dis* 224:49–59. <https://doi.org/10.1093/infdis/jiab161>.
 18. Memoli MJ, Han A, Walters KA, Czajkowski L, Reed S, Athota R, Angela Rosas L, Cervantes-Medina A, Park JK, Morens DM, Kash JC, Taubenberger JK. 2020. Influenza A reinfection in sequential human challenge: implications for protective immunity and “universal” vaccine development. *Clin Infect Dis* 70:748–753. <https://doi.org/10.1093/cid/ciz281>.
 19. Ramanathan M, Ferguson ID, Miao W, Khavari PA. 2021. SARS-CoV-2 B.1.1.7 and B.1.351 spike variants bind human ACE2 with increased affinity. *Lancet Infect Dis* 21:1070. [https://doi.org/10.1016/S1473-3099\(21\)00262-0](https://doi.org/10.1016/S1473-3099(21)00262-0).
 20. Kim S, Liu Y, Lei Z, Dicker J, Cao Y, Zhang XF, Im W. 2021. Differential interactions between human ACE2 and spike RBD of SARS-CoV-2 variants of concern. *bioRxiv* <https://doi.org/10.1101/2021.07.23.453598>.
 21. Lippi G, Henry BM. 2021. How will emerging SARS-CoV-2 variants impact herd immunity? *Ann Transl Med* 9:585–585. <https://doi.org/10.21037/atm-21-893>.
 22. Liu Y, Rocklöv J. 2021. The reproductive number of the Delta variant of SARS-CoV-2 is far higher compared to the ancestral SARS-CoV-2 virus. *J Travel Medicine* 28:taab124. <https://doi.org/10.1093/jtm/taab124>.
 23. Del Rio C, Malani PN, Omer SB. 2021. Confronting the Delta variant of SARS-CoV-2, summer 2021. *JAMA* 326:1001. <https://doi.org/10.1001/jama.2021.14811>.
 24. Nishiura H, Ito K, Anzai A, Kobayashi T, Piantham C, Rodríguez-Morales AJ. 2021. Relative reproduction number of SARS-CoV-2 Omicron (B.1.1.529) compared with Delta variant in South Africa. *Jcm* 11:30. <https://doi.org/10.3390/jcm11010030>.
 25. Khoury DS, Cromer D, Reynaldi A, Schlub TE, Wheatley AK, Juno JA, Subbarao K, Kent SJ, Triccas JA, Davenport MP. 2021. Neutralizing antibody levels are highly predictive of immune protection from symptomatic SARS-CoV-2 infection. *Nat Med* 27:1205–1211. <https://doi.org/10.1038/s41591-021-01377-8>.
 26. Liu Y, Liu J, Xia H, Zhang X, Fontes-Garfias CR, Swanson KA, Cai H, Sarkar R, Chen W, Cutler M, Cooper D, Weaver SC, Muik A, Sahin U, Jansen KU, Xie X, Dormitzer PR, Shi PY. 2021. Neutralizing activity of BNT162b2-elicited serum. *N Engl J Med* 384:1466–1468. <https://doi.org/10.1056/NEJMc2102017>.
 27. Planas D, Veyer D, Baidaliuk A, Staropoli I, Guivel-Benhassine F, Rajah MM, Planchais C, Porrot F, Robillard N, Puech J, Prot M, Gallais F, Gantner P, Velay A, Le Guen J, Kassis-Chikhani N, Edriss D, Belec L, Seve A, Courtellemont L, Pere H, Hocqueloux L, Fafi-Kremer S, Prazuck T, Mouquet H, Bruel T, Simon-Loriere E, Rey FA, Schwartz O. 2021. Reduced sensitivity of SARS-CoV-2 variant Delta to antibody neutralization. *Nature* 596:276–280. <https://doi.org/10.1038/s41586-021-03777-9>.
 28. Schmidt F, Muecksch F, Weisblum Y, Da Silva J, Bednarski E, Cho A, Wang Z, Gaebler C, Caskey M, Nussenzweig MC, Hatziioannou T, Bieniasz PD. 2021. Plasma neutralization of the SARS-CoV-2 Omicron variant. *N Engl J Med* <https://doi.org/10.1056/NEJMc2119641>.
 29. Widge AT, Roupael NG, Jackson LA, Anderson EJ, Roberts PC, Makhene M, Chappell JD, Denison MR, Stevens LJ, Pruijssers AJ, McDermott AB, Flach B, Lin BC, Doria-Rose NA, O'Dell S, Schmidt SD, Neuzil KM, Bennett H, Leav B, Makowski M, Albert J, Cross K, Edara VR, Floyd K, Suthar MS, Buchanan W, Luke CJ, Ledgerwood JE, Mascola JV, Graham BS, Beigel JH, mRNA-1273 Study Group. 2021. Durability of responses after SARS-CoV-2 mRNA-1273 vaccination. *N Engl J Med* 384:80–82. <https://doi.org/10.1056/NEJMc2032195>.
 30. Wheatley AK, Juno JA, Wang JJ, Selva KJ, Reynaldi A, Tan HX, Lee WS, Wragg KM, Kelly HG, Esterbauer R, Davis SK, Kent HE, Mordant FL, Schlub TE, Gordon DL, Khoury DS, Subbarao K, Cromer D, Gordon TP, Chung AW, Davenport MP, Kent SJ. 2021. Evolution of immune responses to SARS-CoV-2 in mild-moderate COVID-19. *Nat Commun* 12:1162. <https://doi.org/10.1038/s41467-021-21444-5>.
 31. Dan JM, Mateus J, Kato Y, Hastie KM, Yu ED, Faliti CE, Grifoni A, Ramirez SI, Haupt S, Frazier A, Nakao C, Rayaprolu V, Rawlings SA, Peters B, Krammer F, Simon V, Saphire EO, Smith DM, Weiskopf D, Sette A, Crotty S. 2021. Immunological memory to SARS-CoV-2 assessed for up to 8 months after infection. *Science* 371:eabf4063. <https://doi.org/10.1126/science.abf4063>.
 32. Amanna IJ, Carlson NE, Slifka MK. 2007. Duration of humoral immunity to common viral and vaccine antigens. *N Engl J Med* 357:1903–1915. <https://doi.org/10.1056/NEJMoa066092>.
 33. Goldberg Y, Mandel M, Bar-On YM, Bodenheimer O, Freedman L, Haas EJ, Milo R, Alroy-Preis S, Ash N, Huppert A. 2021. Waning immunity after the BNT162b2 vaccine in Israel. *N Engl J Med* 385:e85. <https://doi.org/10.1056/NEJMoa2114228>.
 34. Baden LR, El Sahly HM, Essink B, Follmann D, Neuzil KM, August A, Clouting H, Fortier G, Deng W, Han S, Zhao X, Leav B, Talarico C, Girard B, Paila YD, Tomassini JE, Schödel F, Pajon R, Zhou H, Das R, Miller J. 2021. Phase 3 trial of mRNA-1273 during the Delta-variant surge. *N Engl J Med* 385:2485–2487. <https://doi.org/10.1056/NEJMc2115597>.
 35. Mizrahi B, Lotan R, Kalkstein N, Peretz A, Perez G, Ben-Tov A, Chodick G, Gazit S, Patalon T. 2021. Correlation of SARS-CoV-2 breakthrough infections to time-from-vaccine: preliminary study. *medRxiv* <https://doi.org/10.1101/2021.07.29.21261317>.
 36. Oran DP, Topol EJ. 2021. The proportion of SARS-CoV-2 infections that are asymptomatic: a systematic review. *Ann Intern Med* 174:655–662. <https://doi.org/10.7326/M20-6976>.
 37. Madewell ZJ, Yang Y, Longini IM, Jr, Halloran ME, Dean NE. 2020. Household transmission of SARS-CoV-2: a systematic review and meta-analysis. *JAMA Netw Open* 3:e2031756. <https://doi.org/10.1001/jamanetworkopen.2020.31756>.
 38. Wilmes P, Zimmer J, Schulz J, Glod F, Veiber L, Mombaerts L, Rodrigues B, Aalto A, Pastore J, Snoeck CJ, Ollert M, Fagherazzi G, Mossong J, Goncalves J, Skupin A, Nehrbass U. 2021. SARS-CoV-2 transmission risk from asymptomatic carriers: results from a mass screening programme in

- Luxembourg. *Lancet Reg Health Eur* 4:100056. <https://doi.org/10.1016/j.lanepe.2021.100056>.
39. Angel Y, Spitzer A, Henig O, Saiag E, Sprecher E, Padova H, Ben-Ami R. 2021. Association between vaccination with BNT162b2 and incidence of symptomatic and asymptomatic SARS-CoV-2 infections among health care workers. *JAMA* 325:2457. <https://doi.org/10.1001/jama.2021.7152>.
 40. El Sahly HM, Baden LR, Essink B, Doblecki-Lewis S, Martin JM, Anderson EJ, Campbell TB, Clark J, Jackson LA, Fichtenbaum CJ, Zervos M, Rankin B, Eder F, Feldman G, Kennelly C, Han-Conrad L, Levin M, Neuzil KM, Corey L, Gilbert P, Janes H, Follmann D, Marovich M, Polakowski L, Mascola JR, Ledgerwood JE, Graham BS, August A, Clouting H, Deng W, Han S, Leav B, Manzo D, Pajon R, Schodel F, Tomassini JE, Zhou H, Miller J, COVE Study Group. 2021. Efficacy of the mRNA-1273 SARS-CoV-2 vaccine at completion of blinded phase. *N Engl J Med* 385:1774–1785. <https://doi.org/10.1056/NEJMoa2113017>.
 41. Britton T, Ball F, Trapman P. 2020. A mathematical model reveals the influence of population heterogeneity on herd immunity to SARS-CoV-2. *Science* 369:846–849. <https://doi.org/10.1126/science.abc6810>.
 42. Burns CC, Diop OM, Sutter RW, Kew OM. 2014. Vaccine-derived polioviruses. *The J Infectious Diseases* 210:S283–S293. <https://doi.org/10.1093/infdis/jiu295>.
 43. Gorse GJ, Patel GB, Vitale JN, O'Connor TZ. 2010. Prevalence of antibodies to four human coronaviruses is lower in nasal secretions than in serum. *Clin Vaccine Immunol* 17:1875–1880. <https://doi.org/10.1128/CVI.00278-10>.
 44. Bodewes R, de Mutsert G, van der Klis FR, Ventresca M, Wilks S, Smith DJ, Koopmans M, Fouchier RA, Osterhaus AD, Rimmelzwaan GF. 2011. Prevalence of antibodies against seasonal influenza A and B viruses in children in Netherlands. *Clin Vaccine Immunol* 18:469–476. <https://doi.org/10.1128/CVI.00396-10>.
 45. Edridge AWD, Kaczorowska J, Hoste ACR, Bakker M, Klein M, Loens K, Jebbink MF, Matser A, Kinsella CM, Rueda P, Ieven M, Goossens H, Prins M, Sastre P, Deijs M, van der Hoek L. 2020. Seasonal coronavirus protective immunity is short-lasting. *Nat Med* 26:1691–1693. <https://doi.org/10.1038/s41591-020-1083-1>.
 46. Giurgea LT, Han A, Memoli MJ. 2020. Universal coronavirus vaccines: the time to start is now. *NPJ Vaccines* 5:43. <https://doi.org/10.1038/s41541-020-0198-1>.
 47. Abu-Raddad LJ, Chemaitelly H, Butt AA, National Study Group for C-V. 2021. Effectiveness of the BNT162b2 Covid-19 vaccine against the B.1.1.7 and B.1.351 variants. *N Engl J Med* 385:187–189. <https://doi.org/10.1056/NEJMc2104974>.
 48. Andrews N, Tessier E, Stowe J, Gower C, Kirsebom F, Simmons R, Gallagher E, Chand M, Brown K, Ladhani SN, Ramsay M, Bernal JL. 2021. Vaccine effectiveness and duration of protection of Comirnaty, Vaxzevria and Spikevax against mild and severe COVID-19 in the UK. *medRxiv* <https://doi.org/10.1101/2021.09.15.21263583>.