

Cardiac stem cells: Current knowledge and future prospects

Radwa A Mehanna, Marwa M Essawy, Mona A Barkat, Ashraf K Awaad, Eman H Thabet, Heba A Hamed, Hagar Elkafrawy, Nehal A Khalil, Abeer Sallam, Marwa A Kholief, Samar S Ibrahim, Ghada M Mourad

ORCID number: Radwa Ali Mehanna 0000-0001-9048-4464; Marwa M Essawy 0000-0002-4781-4293; Mona A Barkat 0000-0003-3850-0193; Ashraf K Awaad 0000-0001-8285-0188; Eman H Thabet 0000-0003-3490-3825; Heba A Hamed 0000-0001-8080-4620; Hagar Elkafrawy 0000-0002-1591-1689; Nehal A Khalil 0000-0001-9380-7425; Abeer Sallam 0000-0002-4690-6348; Marwa A Kholief 0000-0001-8776-5257; Samar S Ibrahim 0000-0002-5547-9667; Ghada M Mourad 0000-0002-6145-3076.

Author contributions: Mehanna RA designed, supervised and revised the manuscript, collected and analyzed the data, and shared in figures' creation; Mourad GM supervised the work and revised the paper; all other authors contributed to data collection, writing the manuscript, and creating figures and tables.

Conflict-of-interest statement: Dr. Mehanna has nothing to disclose.

Supported by Science and Technology Development Fund, No. 28932; and Cardiovascular Research, Education, Prevention Foundation, CVREP - Dr. Wael Al Mahmeed Grant.

Country/Territory of origin: Egypt

Specialty type: Cardiac and cardiovascular systems

Radwa A Mehanna, Eman H Thabet, Abeer Sallam, Medical Physiology Department/Center of Excellence for Research in Regenerative Medicine and Applications, Faculty of Medicine, Alexandria University, Alexandria 21500, Egypt

Marwa M Essawy, Oral Pathology Department, Faculty of Dentistry/Center of Excellence for Research in Regenerative Medicine and Applications, Faculty of Medicine, Alexandria University, Alexandria 21500, Egypt

Mona A Barkat, Human Anatomy and Embryology Department/Center of Excellence for Research in Regenerative Medicine and Applications, Faculty of Medicine, Alexandria University, Alexandria 21500, Egypt

Ashraf K Awaad, Samar S Ibrahim, Center of Excellence for Research in Regenerative Medicine and Applications, Faculty of Medicine, Alexandria University, Alexandria 21500, Egypt

Heba A Hamed, Ghada M Mourad, Histology and Cell Biology Department/Center of Excellence for Research in Regenerative Medicine and Applications, Faculty of Medicine, Alexandria University, Alexandria 21500, Egypt

Hagar Elkafrawy, Nehal A Khalil, Medical Biochemistry Department/Center of Excellence for Research in Regenerative Medicine and Applications, Faculty of Medicine, Alexandria University, Alexandria 21500, Egypt

Marwa A Kholief, Forensic Medicine and Clinical toxicology Department/Center of Excellence for Research in Regenerative Medicine and Applications, Faculty of Medicine, Alexandria University, Alexandria 21500, Egypt

Corresponding author: Radwa A Mehanna, MD, PhD, Academic Research, Professor, Executive President, Medical Physiology Department/Center of Excellence for Research in Regenerative Medicine and Applications, Faculty of Medicine, Alexandria University, Al Khartoum Square, Azareeta, Alexandria 21500, Egypt. radwa.mehanna@alexmed.edu.eg

Abstract

Regenerative medicine is the field concerned with the repair and restoration of the integrity of damaged human tissues as well as whole organs. Since the inception of the field several decades ago, regenerative medicine therapies, namely stem cells, have received significant attention in preclinical studies and clinical trials. Apart from their known potential for differentiation into the various body cells, stem cells enhance the organ's intrinsic regenerative capacity by altering its environment, whether by exogenous injection or introducing their products that

Provenance and peer review:

Invited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's scientific quality classification

Grade A (Excellent): 0
 Grade B (Very good): B, B
 Grade C (Good): 0
 Grade D (Fair): 0
 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Received: February 26, 2021

Peer-review started: February 26, 2021

First decision: June 24, 2021

Revised: July 2, 2021

Accepted: January 6, 2022

Article in press: January 6, 2022

Published online: January 26, 2022

P-Reviewer: Jin C, Matsushita S

S-Editor: Gong ZM

L-Editor: A

P-Editor: Gong ZM



modulate endogenous stem cell function and fate for the sake of regeneration. Recently, research in cardiology has highlighted the evidence for the existence of cardiac stem and progenitor cells (CSCs/CPCs). The global burden of cardiovascular diseases' morbidity and mortality has demanded an in-depth understanding of the biology of CSCs/CPCs aiming at improving the outcome for an innovative therapeutic strategy. This review will discuss the nature of each of the CSCs/CPCs, their environment, their interplay with other cells, and their metabolism. In addition, important issues are tackled concerning the potency of CSCs/CPCs in relation to their secretome for mediating the ability to influence other cells. Moreover, the review will throw the light on the clinical trials and the preclinical studies using CSCs/CPCs and combined therapy for cardiac regeneration. Finally, the novel role of nanotechnology in cardiac regeneration will be explored.

Key Words: Cardiac stem and progenitor cells; Cardiac stem cells' secretome; Cardiac stem cells' niche and metabolism; Nanotechnology; Clinical trials; Combined therapy

©The Author(s) 2022. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: With the growing evidence for the existence of regenerating cardiac stem and progenitor cells, studies to evaluate their therapeutic potential have received increasing attention. Although pre-clinical research and clinical trials have demonstrated promising results, yet the latter were often inconsistent in many aspects thus imposing the need for deeper exploration of the molecular biology and relevant pathways regulating cardiogenesis and cardiac muscle repair. This review gives an insight into cardiac stem and progenitor cells regarding their embryological origin, populations, niche, secretome, and metabolism. It overviews the current preclinical research, including medical nanotechnology, and the clinical trials generally applied for cardiac regeneration.

Citation: Mehanna RA, Essawy MM, Barkat MA, Awaad AK, Thabet EH, Hamed HA, Elkafrawy H, Khalil NA, Sallam A, Kholief MA, Ibrahim SS, Mourad GM. Cardiac stem cells: Current knowledge and future prospects. *World J Stem Cells* 2022; 14(1): 1-40

URL: <https://www.wjgnet.com/1948-0210/full/v14/i1/1.htm>

DOI: <https://dx.doi.org/10.4252/wjsc.v14.i1.1>

INTRODUCTION

Cardiovascular diseases are the leading cause of death globally, as stated by the latest report 2019 for the World Health Organization, with 17.9 million deaths per year, accounting for 31% of all deaths worldwide.

The heart is one of the least proliferative organs in the human body, and its minimal regenerative capacity has been dogma for decades. Such dogma has been led by the belief that the heart cannot regenerate from ischemic damage. The absence of primary tumors in the heart has further supported the notion of low proliferation. In an alleged post-mitotic organ, it has been debatable whether cardiac cells repair through activation of resident cardiac stem cells (CSCs) and cardiac progenitor cells (CPCs) or by the proliferation of pre-existing cardiomyocytes (CMs). In 2009, Bergmann *et al*[1] were the first to refute that notion and have reported that the heart can in fact self-renew. Based on the results obtained from their carbon-14-labelled DNA study to track CMs, Bergmann *et al*[1] stated that about 50% of CMs renew over the lifespan of an adult. Hsieh *et al*[2] provided further evidence for the origin of newly generated CMs from progenitor cells in an alpha myosin heavy chain (MHC) transgenic model. They estimated that approximately 15% of CMs can regenerate in adult hearts following ischemic damage. With progression of research, lineage tracing of regenerated cardiac tissue confirmed that the newly regenerated CMs develop from a non-CM and possibly from stem cells (SCs)[2].

Further studies have revealed various CSC/CPC candidates that are morphologically and functionally distinct from each other yet act in a complementary fashion and contribute to the regeneration process. This complex cell aggregation is known as the CSC niche that has been a challenge to characterize and locate anatomically[3].

SC applications have been under intensive research interest since the early 20th century. Many types have been isolated, starting from the embryonic, amniotic, and cord blood mesenchymal stem cells (MSCs) and passing through the adult SCs till the induced pluripotent SCs (iPSCs). Adult MSCs are undifferentiated cells with the same potentials as progenitor cells regarding the ability to differentiate into all three germ layer cells[4]. Exogenous MSCs from various sources, including bone marrow, adipose tissue, umbilical cord, placenta, and amniotic fluid[5], have shown promising results in the treatment of cardiovascular diseases. However, the outcome of CSC therapy has shown superior results in experimental studies but to a lesser extent in human clinical trials[6]. The applications of SC therapy for cardiovascular regeneration still hold a plethora of queries to be answered as well as commandment of the molecular and signaling features for CSCs in order to standardize this therapy. Among the aspects that need optimization are the types of SCs and supporting cells to be used, the number of cells, the route of injection, the frequency, and best timing for transplantation. Standardization requires an advanced understanding of the full biological features of CSCs.

SC therapy in cardiac regeneration has dual beneficiary actions. Primarily, the transplanted exogenous SCs would directly differentiate into CMs. Concomitantly, SCs activate the endogenous progenitors through their rich secretome of extracellular vesicles, immunomodulatory and growth factors, protein, and nucleic acid families[7]. These paracrine factors act to activate resident SCs and enhance vascularization to potentiate cardiac repair.

This review aims to provide insight into CSCs/CPCs regarding their embryological origin, populations, niche, metabolism, secretome, and therapeutic potentials. Also discussed is the interplay of nanotechnology with SCs in several aspects, including differentiation, tracking, imaging, and assisted therapy, showing the prospects and limitations of nanoparticle (NP)-based cardiac therapy. Finally, preclinical trials and ongoing, completed, and future clinical trials using CSCs and combined therapy are shown to delineate the potential applications in treating cardiac disease.

EMBRYOLOGICAL ORIGIN OF CPCs

The heart is formed of a wide range of cell types originating from the mesodermal precursor cells. They include CMs and endocardial cells forming the inner layer, while epicardial-derived cells (EPDCs) and smooth muscle cells (SMCs) are found on the external layer. Differentiation of the mesodermal cells is initiated by the T-box transcriptional factors Brachyury (Bry) and Eomes. Bry⁺ cells differentiate into insulin gene enhancer protein islet-1 (ISL1) and T-box transcription factor 5 (TBX5) expressing cells, while Eomes induce expression of mesoderm posterior 1 (MESP1). MESP1⁺ cells are identified before the first heart field (FHF) and the second heart field (SHF) separations, so MESP1 serves as an indicator of early CPCs for both heart fields[8]. Chemokine receptor type 4 (CXCR4), fetal liver kinase 1 (FLK-1), and platelet derived growth factor receptor A are other surface markers that coincide with MESP1 and are used in combination to isolate CPCs[9,10].

In addition, a novel cell surface marker known as G protein-coupled receptor lysophosphatidic acid receptor 4 is specific to CPCs and determines its functional significance. Interestingly, its transient expression peaks in cardiac progenitors after 3 to 7 d of human (h)PSCs differentiation toward cardiac lineage, then it declines. *In vivo*, lysophosphatidic acid receptor 4 shows high expression in the initial stages of embryonic heart development and decreases throughout development[11].

The FHF cells are the firstly differentiated myocardial cells that are derived from cells in the anterior lateral plate mesoderm; they give rise to the left ventricle, partially some of the right ventricle population, sinoatrial node, atrioventricular node, and both atria[12]. Meanwhile, the SHF cells originate from the pharyngeal mesoderm to the posterior side of the heart and further divide into anterior and posterior SHF. They contribute to the right ventricle, atria, and the cardiac outflow tract (OFT) formation. Addition of the SHF-derived CMs to the ventricles depend on myocyte enhancer factor 2C (MEF2C). It has been found that MEF2C null mice die at 9.5-d post conception with severe heart defects due to failure of heart looping[13]. In OFT formation, two waves of SHF progenitors and their derivatives have been identified, making a differential

contribution to the aorta and pulmonary artery. The early wave of cells is favorably directed to the aorta, while the second wave of cells contributes to the pulmonary artery. Phosphoinositide-dependent kinase-1 critically regulates the second wave of cells, and its deletion results in pulmonary stenosis[14]. The epicardium of the heart is formed of a transient proepicardial organ. Proepicardium is formed from homeobox protein NKx2.5 (NKx2.5) and ISL1⁺ cells. After epicardial formation, subepicardial mesenchymal space is formed by epithelial to mesenchymal cell transformation of the epicardial cells[15] (Figure 1).

The differentiation in the posterior SHF is regulated by *Hoxb1* gene. Stimulation of *Hoxb1* in embryonic stem cells (ESCs) halts cardiac differentiation, while *Hoxb1*-deficiency shows premature cardiac differentiation in embryos. Moreover, an atrioventricular septal defect develops as a result of ectopic differentiation in the posterior SHF of embryos deficient in *Hoxb1* and its paralog *Hoxa1*[16].

Multiple signaling pathways have essential roles in cardiogenesis with a sequential arrangement. The transforming growth factor- β (TGF- β) superfamily, retinoic acid, Hedgehog, Notch, Wnt, and fibroblast growth factors (FGFs) pathways comprise the chief signaling pathways involved in cardiac development. These pathways, along with transcription factors and epigenetic regulators, regulate cardiac progenitors' specification, proliferation, and differentiation into the different cardiac cell lineages [17].

SIGNALING PATHWAYS DURING CARIOGENESIS

TGF- β superfamily

The TGF- β superfamily members consist of over 30 structurally associated polypeptide growth factors including nodal and bone morphogenetic proteins (BMP)[18].

Nodal signaling is vital for the formation of sinoatrial node. Nodal inhibition during the cardiac mesoderm differentiation stage downregulates *PITX2c*, a transcription factor recognized to inhibit the formation of the sinoatrial in the left atrium during cardiac development[19]. Moreover, nodal signaling is dispensable for initiation of heart looping; however, it regulates asymmetries that result in a helical shape at the heart tube poles[20].

BMP signaling, as a member of TGF- β , has an important role in the different stages of heart development including the OFT formation, endocardium, and lastly the epicardium. The cardiac neural crest cells have a crucial role in normal cardiovascular development. They give rise to the vascular smooth muscle of the pharyngeal arch arteries, OFT septation, valvulogenesis, and development of the cardiac conduction system[21] (Figure 1). The role of BMP in OFT septation mainly depends on their gradient signaling, which arranges neural crest cell aggregation along the OFT; this Dullard-mediated tuning of BMP signaling ensures the fine timed zipper-like closure of the OFT by the neural crest cells[22]. Furthermore, the BMP signaling promotes the development of endocardial cells (ECs) from hPSC-derived cardiovascular progenitors [23]. It is also integrated with Notch signaling for influencing the proepicardium formation, where overexpression of Notch intracellular receptor in the endothelium enhances BMP expression and increases the number of phospho-Smad1/5⁺ cells for enhancing the formation of the proepicardium[24].

Retinoic acid, hedgehog, and Notch signaling pathways

Retinoic acid signaling plays a role in heart development. It is a key factor for efficient lateral mesoderm differentiation into atrial-like cells in a confined time frame. The structural, electrophysiological, and metabolic maturation of CMs are significantly influenced by retinoic acid[25]. However, it is reported that retinoic acid receptor agonists transiently enhance the proliferation of human CPCs at the expense of terminal cardiac differentiation[26].

The downregulation of the retinoic acid responsive gene, ripply transcriptional repressor 3 (*RIPPLY3*), within the SHF progenitors by histone deacetylase 1 is required during OFT formation[27].

Hedgehog signaling has a role in OFT morphogenesis. Lipoprotein-related protein 2 (*LRP2*) is a member of the LDL receptor gene family, a class of multifunctional endocytic receptors that play crucial roles in embryonic development. *LRP2* is expressed in the anterior SHF cardiac progenitor niche, which leads to the elongation of the OFT during separation into aorta and pulmonary trunk. Loss of *LRP2* in mutant mice results in depleting a pool of sonic hedgehog-dependent progenitor cells in the anterior SHF as they migrate into the OFT myocardium due to premature differen-

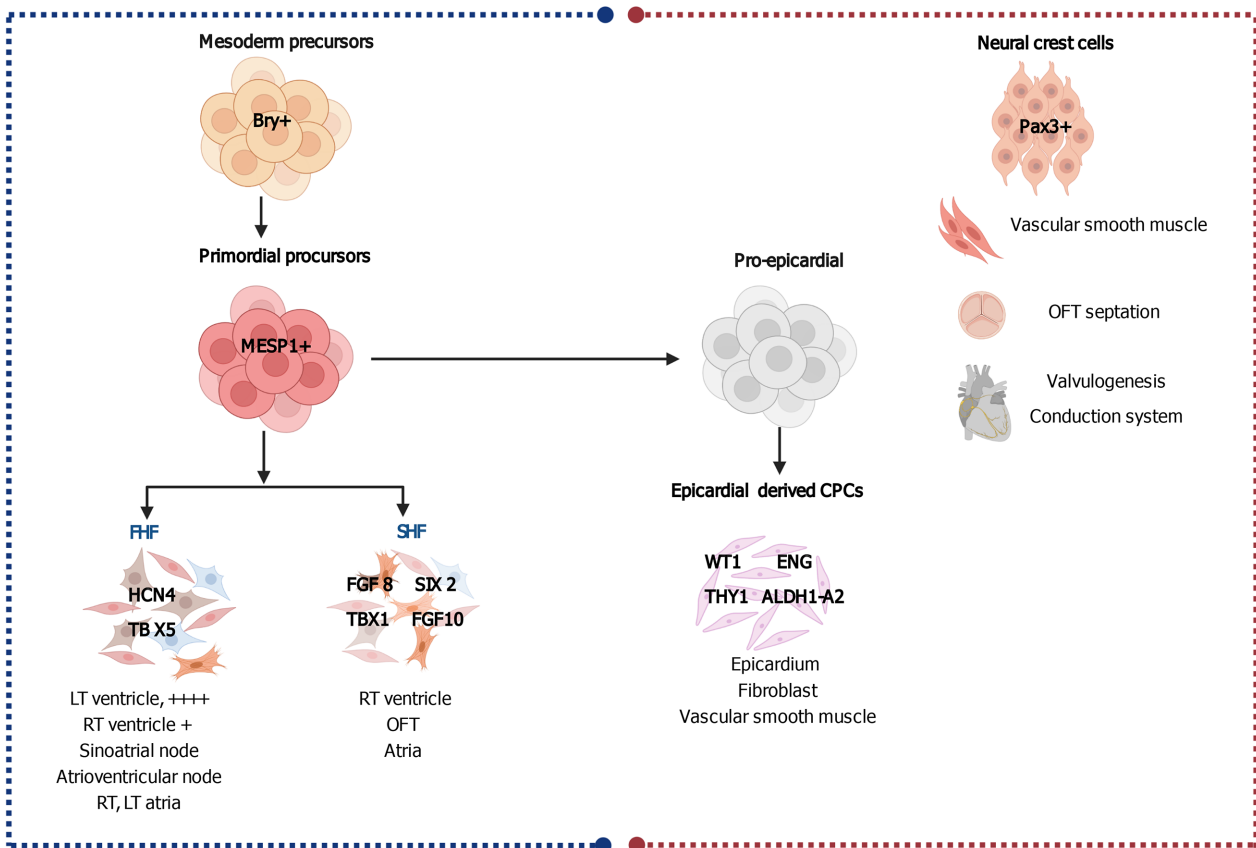


Figure 1 Embryonic cardiac progenitors, Brachyury-positive mesoderm precursors and Pax3⁺ neural crest cells. Brachyury (Bry⁺) mesoderm precursors give rise to the mesoderm posterior 1⁺ primordial precursors, which are the origin of the first heart field, second heart field, and proepicardial progenitors, each population of which is responsible for the development of different parts in the heart. Pax3⁺ neural crest cells are responsible for the development of vascular smooth muscle, outflow tract, valves and the conductive system. Progenitors are tagged with their specific markers. Created with BioRender.com. CPC: Cardiac progenitor cell; LT: Left; RT: Right; FHF: First heart field; SHF: Second heart field; OFT: Outflow tract.

tiation into CMs. This depletion results in aberrant shortening of the OFT[28].

Four Notch receptors (Notch1–Notch4) and five structurally similar Notch ligands [Delta-like (DLL) 1, DLL3, DLL4, Jagged1, and Jagged2] have been detected in mammals[29]. Activation of Notch signaling enhances CM differentiation from human PSCs. However, the CMs derived from Notch-induced cardiac mesoderm are developmentally immature[30]. *In vivo*, the Notch pathway plays a significant role in CPC biology. An arterial-specific Notch ligand known as DLL4 is expressed by SHF progenitors at critical time-points in SHF biology. The DLL4-mediated Notch signaling is a crucial requirement for maintaining an adequate SHF progenitor pool, in a way that *DLL4* knockout results in decreased proliferation and increased apoptosis. Reduced SHF progenitor pool leads to an underdeveloped OFT and right ventricle[31].

Wnt pathway

The Wnt signaling pathway has an essential role in many developmental stages of embryogenesis. The Wnt family consists of 19 distinct Wnt proteins and other 10 types of Frizzled receptors. On the basis of their primary functions, the Wnt and Frizzled receptors are divided into two major classes, which are the canonical and non-canonical Wnt pathways[32]. Accumulating evidence suggests a role for the dynamic balance between canonical and non-canonical Wnt signaling in cardiac formation and differentiation. Wnt/ β -catenin signaling is required for proper mesoderm formation and proliferation of CMs but needs to be low for terminal differentiation and cardiac specification. In contrast, for cardiac specification in murine and human ESCs, non-canonical β -catenin independent Wnt signaling is essential, while the non-canonical Wnt signaling is necessary for terminal differentiation later in development[33].

The activation of non-canonical Wnt is non-catenin-independent, and the downstream proteins involve several kinases, including protein kinase C, calcium/calmodulin-dependent kinase, and Jun N terminal kinase (JNK). Wnt11 enhances angiogenesis and improves cardiac function through non-canonical Wnt-protein kinase C-Jun N terminal kinase dependent pathways in myocardial infarction (MI)

[34]. In hypoxia, Wnt11 expression preserves the integrity of mitochondrial membrane and facilitates the release of insulin growth factor-1 (IGF-1) and vascular endothelial growth factor (VEGF), thus protecting CMs against hypoxia[35]. Canonical dependent Wnt signaling, Wnt 3 Ligand, favors the pacemaker lineage, while its suppression promotes the chamber CM lineage[36].

TRANSCRIPTOME AND REGENERATIVE CAPACITY OF SUB-POPULATIONS

The regenerative capacity of most organs is contingent on the adult SC populations that exist in their niches and are activated by injury. Adult SC populations vary greatly in their molecular marker expression profile and hence in their possible role in regenerative medicine. The transcriptome is a representation of the gene read-outs, the cellular state, and is imperative for studying all genetic disease and biological processes. The genome-wide profiling using novel sequencing technology has made transcriptome research accessible.

c-KIT⁺ CPCs

Receptor tyrosine kinase (RTK) c-KIT (also referred to as SC factor receptor or CD117)-expressing CPCs are mainly located in the atria and the ventricular apex, comprising most of the ventricular and atrial myocardium[37]. c-KIT⁺ cells also express the cardiac transcription factors NKx2.5, GATA binding protein 4 (GATA4), and MEF2C but are negative for the hematopoietic markers CD45, CD3, CD34, CD19, CD16, CD20, CD14, and CD56[38,39]. SC factor ligand attaches to the c-KIT receptor and activates the phosphoinositide 3-kinase/protein kinase B (PI3K/AKT) and p38 mitogen-activated protein kinase (MAPK) signaling pathways[40]. Both PI3K/AKT and MAPK pathways control various CPCs functions like self-renewal, proliferation, migration, and survival [41]. During embryonic development and the early post-natal time, c-KIT⁺ CPCs contribute to the generation of new CMs. Such capacity declines in the adult heart with only a few new CMs originating from CPCs[42]. In a rat MI model, the c-KIT⁺ CPCs have migrated through the collagen type I and type III matrices into the infarcted area. The transplanted CPCs have shown overexpressed matrix metalloproteinases (MMPs; MMP2, MMP9, and MMP14) that degrade extracellular matrix (ECM), concluding that c-KIT⁺ CPCs hold an invasive capacity[43]. Transplanted CPCs (c-KIT⁺ CPCs and cardiospheres) also show an endogenous proliferative potential *in vivo* and additionally activate endogenous CPCs[44].

SCA-1⁺ CPCs

Stem cell antigen 1 (SCA-1) expressing CPC population exists predominantly in the atrium, intra-atrial septum, and atrium-ventricular boundary and dispersed inside the epicardial layer of adult hearts[45]. SCA-1 is a cell surface protein of the lymphocyte antigen-6 (*Ly6*) gene family, which has roles in cell survival, proliferation, and differentiation[46]. A population of SCA-1⁺ cells from murine adult myocardium hold a telomerase activity comparable to that of a neonatal heart. This SCA-1⁺ population is different from hematopoietic SCs as they lack CD45, CD34, c-KIT, LIM domain only 2, GATA2, VEGF receptor 1, and T-cell acute lymphoblastic leukemia 1/SC leukemia proteins. SCA-1⁺ cells are also distinct from endothelial progenitor cells and express cardiac lineage transcriptional factors such as GATA4, MEF2C, and translation elongation factor 1 yet lack transcripts for cardiomyocytic structural genes such as *BMP1r1* and α -, β -MHC[47,48]. Although this population exhibits the endothelial marker CD31, it is suggested to be due to the contaminating endothelial CD31⁺/SCA-1⁺ cells. *In vitro* studies have revealed that 5-azacytidine (5-aza), a demethylating agent, pushed SCA-1⁺ cells to differentiate into CMs[48,49]. Further studies have isolated SCA-1⁺ cells that lack CD31 and CD45 markers, referring to them as lineage negative (Lin⁻). The SCA-1⁺/Lin⁻ cells display a mesenchymal cell-surface profile (CD34⁻, CD29⁺, CD90⁺, CD105⁺, and CD44⁺) and are able to differentiate, to a certain extent, into CMs and endothelial and smooth muscle-like cells[50,51].

Human SCA-1⁺-like cells also express early cardiac transcription factors (GATA4, MEF2C, insulin gene enhancer protein ISL-1, and Nkx-2.5) and can differentiate into contractile CMs[52]. Although a human ortholog of the SCA-1 protein has not been yet identified, an anti-mouse SCA-1 antibody is used to isolate SCA-1⁺-like cells from the adult human heart.

MESP1⁺ CPCs

MESP1 expressing cells mainly contribute to the mesoderm and to the myocardium of the heart tube during development[53]. Transient expression of MESP1 seems to accelerate and enhance the appearance of cardiac progenitor. However, homologous disruption of the *MESP1* gene has resulted in aberrant cardiac morphogenesis. *MESP1* interacts with the promoter area of main cardiac transcription factors, including heart and neural crest derivatives expressed 2, Nkx2-5, myocardin, and GATA4[54]. These factors induce fibroblasts to express a full battery of cardiac genes, form sarcomeres, develop CM-like electrical activity, and in a few cases elicit beating activity[55]. Several studies have shown that the addition of MESP1 could enhance the efficacy of direct reprogramming of fibroblasts into CMs[56,57]. The transdifferentiation of fibroblasts to CMs *via MESP1* suggests that *MESP1* chiefly modulates the gene regulatory network for cardiogenesis[52].

KDR⁺ CPCs

Kinase insert domain receptor (KDR), also known as Flk-1, is one of the earliest discovered cardiogenic progenitor cell markers acting during the early stages of cardiac development in human[58]. Nelson *et al*[59] have reported that Flk-1 has a distinctive transcriptome that has been evident at day 6, immediately after gastrulation but prior to the expression of the cardiac transcription factors. KDR⁺ population lack the pluripotent octamer-binding transcription factor 4, sex determining region Y-Box transcription factor (SOX) 2, and endoderm SOX17 markers. On the other hand, KDR⁺ CPCs have shown a noteworthy upregulation in SOX7, a vasculogenic transcription factor, overlapping with the emergence of primordial cardiac transcription factors GATA4, myocardin, and NKx2.5. Moreover, KDR subpopulations that overexpress SOX7 are associated with a vascular phenotype rather than a cardiogenic phenotype. These outcomes offer insights for refining the therapeutic regenerative interventions.

CPCs from the first and second heart fields

The FHF cells express hyperpolarization activated cyclic nucleotide gated potassium channel 4 and TBX5, while SHF progenitors express TBX1, FGF 8, FGF10, and sine oculis homeobox2 (Figure 1). Cells from the SHF exhibit high proliferative and migratory capacities and are mostly responsible for the elongation and winding of the heart tube. Moreover, SHF cells differentiate to CMs, SMCs, fibroblasts, and endothelial cells (ECs) along their journey in the heart tube to form the right ventricle, right ventricular OFT, and most of the atria[60,61]. However, FHF cells hold less proliferative and migratory potentials and differentiate predominantly to CMs that form the left ventricle and small parts of the atria[62]. The cells of the cardiac crescent, theoretically the progeny of FHF CPCs, are terminally differentiated cells expressing the markers of CMs, such as actin alpha cardiac muscle 1 and myosin light chain 7[63, 64], hence they are unlikely to be multipotent progenitors. Therefore, it is difficult to identify FHF before Nkx2.5 and TBX5 expressions. Conversely, multipotent SHF CPCs were validated with a clonal tracing experiment and identified by ISL1 expression[65]. However, ISL1 expression is not specific for SHF and has been proposed to represent only the developmental stages[66]. Tampakakis *et al*[67] generated ESCs by using hyperpolarization activated cyclic nucleotide gated potassium channel 4-green fluorescent protein and TBX1-Cre; Rosa-red fluorescent protein reporters of the FHF and the SHF respectively, and also by using live immunostaining of the cell membrane CXCR4, a SHF marker and the reporters. The ESC-derived progenitor cells have shown functional properties and transcriptome similar to their *in vivo* equivalents. Thus, chamber-specific cardiac cells have been generated for modelling of heart diseases *in vitro*.

Epicardium-derived CPCs

The EPDCs are important as a signaling source for heart development, cardiac regeneration, and post-MI heart repair. Throughout the development of the heart in mice, EPDCs aid in the formation of various cardiac cell types and secrete paracrine factors for myocardial maturation[68]. In the adult heart, EPDCs are normally dormant and become stimulated following myocardial injury. Transcriptional analysis of the EPDCs derived from human (h) iPSCs cells have revealed several markers of EPDCs including Wilm's tumor protein 1, endoglin, thymus cell antigen 1, and aldehyde dehydrogenase 1 family member A2[69] (Figure 1). Following MI in mice, EPDCs undergo an epithelial-to-mesenchymal transition, with overexpression of Wilms tumor protein 1, and differentiate mainly into SMCs/fibroblasts[70,71]. EPDC-secreted paracrine factors include VEGF-A, FGF2, and PDGF-C, which support the growth of

blood vessels, protect the myocardium, and recover cardiac functions in an acute MI-mouse model[70].

Side population-derived CPCs

Side population (SP) cells have been detected in the heart and other various tissues and hold enhanced stem and progenitor cell activity[72]. SP cells, when stained *in vitro*, hold the ability to flush out the DNA Hoechst dye from their nuclei[73]. Gene expression profiling of SP cells after MI has revealed a downregulation of Wnt-related signals coupled with increased SP cell proliferation. This has been validated *in vitro* by treatment of isolated SP cells with canonical Wnt agonists or recombinant Wnt, where the proliferation of SP cells has been repressed with partial arresting the G1 cell cycle phase[74]. Consistent with this observation, delivery of secreted Frizzled-related proteins (SFRP; the Wnt antagonist) improves post-MI remodeling[75,76].

SP cells can be identified by surface marker adenosine triphosphate (ATP) binding cassette subfamily G member 2 (ABCG2), also referred to as the breast cancer resistance protein1[77]. ABCG2⁺ cells have been also observed in the adult heart and can differentiate *in vitro* into CMs[78]. When SP cells have been injected into the injured hearts of rats, they have been recruited to the injured regions, where they differentiate into CMs, ECs, and SMCs, suggesting that they may be endogenous SP cells[79]. However, ABCG2-CreER based genetic lineage tracing has demonstrated that ABCG2⁺ cells could only differentiate into the multiple cardiac cell lineages during the embryonic stages but not in adulthood[80,81]. The combination of ABCG2⁺ cells with pre-existing CMs is more likely to stimulate CM proliferation rather than differentiation into CMs directly[82]. Therefore, genetic fate mapping investigations have disproved the SP cells property of the adult endogenous ABCG2⁺ SP and their *in vivo* renewing myogenic ability[83].

Cardiosphere-derived CPCs

Cardiospheres contain a combination of stromal, mesenchymal, and progenitor cells that are isolated from cultures of human heart biopsy[39,84]. They represent a niche-like environment, with cardiac-committed cells in the center and supporting cells in the periphery of the spherical cluster[85]. The cardiosphere-derived cells (CDCs) were originally isolated from mouse heart explants and human ventricular biopsies based on their ability to form three-dimensional (3D) spheroids in suspension cultures[86]. CDCs have grabbed much attention due to their proliferation and differentiation abilities by inherent stimulation of cardio-specific differentiation factors [GATA4, MEF2C, Nkx2.5, heart and neural crest derivatives expressed 2, and cardiac troponin T (TNNT2)] using a clustered regularly interspaced short palindromic repeat/dead Cas9 (CRISPR/dCas9) assisted transcriptional enhancement system[87,88]. Sano *et al*[89] have postulated that the CRISPR/dCas9 system may provide a proficient method of modifying *TNNT2* gene activation in SCs. Consequently, CRISPR/dCas9 can improve the therapeutic outcomes of patients with ischemic heart disease by enhancing the transplanted CDCs differentiation capacity within the ischemic myocardium. Heart tissue is usually obtained by endomyocardial biopsy or during open cardiac surgery and grown in explants to form CDCs. CDCs have shown a superior myogenic differentiation potential, angiogenesis, and paracrine factor secretion as compared to other cell types. In heart failure animal models, the injected CDCs potentially differentiated into CMs and vascular cells. Additionally, CDCs have diminished unfavorable remodeling and infarct size, and hence improve cardiac function[90]. Accordingly, cardiospheres and CDCs may be some of the most promising sources of CPCs for cardiac repair.

CSC niche

The niche in the heart integrates several heterogeneous cell types, including CSCs, progenitors, fibroblasts, SMCs, CMs, capillaries, and supporting telocytes (TCs)[91], together with the junctions and cementing ECM that hold the niche together. Such architectural arrangement is essential for protection against external damaging stimuli and for preserving the stemness of the CSCs (Figure 2). Without the niche microenvironment, CSCs lose their stemness and initiate differentiation eventually, leading to the exhaustion of the CSC pool. Similarly, *in vitro* studies require feeder layers and cytokines supplements in the culture media to ensure that SCs remain in their undifferentiated state[37].

In vitro studies have recapitulated the niche theory using cardiospheres, which are 20–150 μm spheres (Figure 2) of cells generated from the explant outgrowth of heart tissues[92,93]. Cardiospheres consist of CSCs in the core and cells committed to the cardiac lineage such as myofibroblasts, while vascular SMCs and ECs form the outer

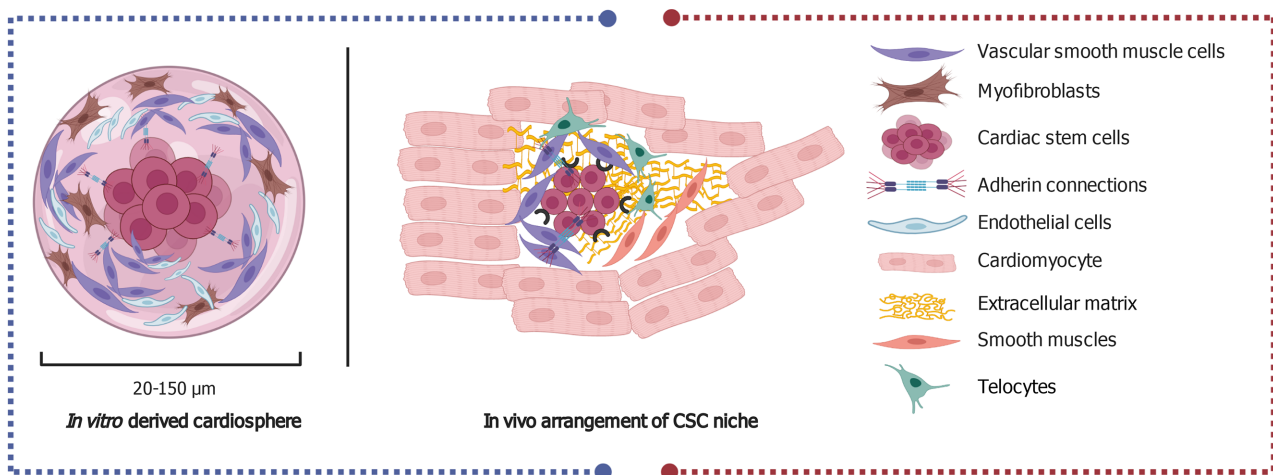


Figure 2 *In vivo* arrangement of the central cardiac stem cells and the surrounding cells that comprise the niche (right side) and the *in vitro* derived cardio spheres (left side). The key delineates the types of cells identified in the niche and cardio spheres. Created with BioRender. CSC: Cardiac stem cell.

layer of the spheres. The 3D structure of cardiospheres protects the interiorly located CSCs from oxidative stress as well as maintain their stemness and function[84].

Accurate anatomical identification of CSCs *in vivo* remains a challenge due to the lack of basal-apical anatomical orientation as seen in epithelial organs such as the intestines[94]. Moreover, the heart does not comprise a specific compartment, where cells form a well-defined lining as seen in the bone marrow osteoblasts[95]. The adult heart epicardial lining anatomically contains several classes of niches, which are not limited to the sub epicardium[96] but dispersed throughout the myocardium, more in the atria and apex away from hemodynamic stress[97]. Some niches have been described in the atrio-ventricular junction of adult mouse and rat hearts[98] and interestingly in the human hearts[99]. The young mouse heart has been studied morphometrically to identify the location of CSCs niche and has been defined as a randomly positioned ellipsoid structure consisting of cellular and extracellular components. Within the niches, undifferentiated CSCs are usually assembled together with early committed cells that express c-KIT on surface, Nkx2.5 in the nucleus, and the contractile protein α -sarcomeric actin in the cytoplasmic[97].

CSCs niche consists of clusters of c-kit⁺, MDR1⁺, and Sca-1⁺ cells[98] but lack the expression of the transcription factors and cytoplasmic or membrane proteins of cardiac cells[99,100]. Cardiac c-kit⁺/CD45⁻ cells comprise about 1% of the CSC niche [97], are self-renewing clonogenic, and possess a cardiac multilineage differentiation potential comprise[101].

Within the niche, gap junctions (connexins) and (cadherins) connect SCs to their supporting cells, myocytes/fibroblasts. Conversely, ECs and SMCs do not act as supporting cells. Hence, the communication between CSCs with CMs and fibroblasts has been investigated by using *in vitro* assays[102]. The transmission of dyes *via* gap junctions between CSCs and CMs or fibroblasts was demonstrated previously and verified the functional coupling of these three cell populations[97]. In addition, micro ribonucleic acid (miRNA-499) translocates from CMs to CSCs comprising to the initiation of lineage specification and formation of myocytes[103].

Identification of SC niches is contingent upon the fulfillment of explicit criteria, including the recognition and determination of the affixing of SCs to their supporting cells as well as assuring the existence of an ancestor-progeny association[104]. Chemical and physical signals modulate the behavior of SCs within the niche. Amongst these signals are cytokines, cell surface adhesion molecules, shear forces, oxygen tension, innervation, and ions that serve as major determinants of SCs function [97]. Cell-to-cell signaling mediates the fate of SCs within the niches to promote self-renewal and favors their migration and differentiation. The fine-tuned crosstalk between SCs and their supporting cells regulates the state of the niche regarding quiescence or activity[105].

CSC niches, similar to the bone marrow, characteristically live in low oxygen tension, which favors a quiescent primitive state for SCs[106]. The longstanding perpetuation of the CSC niche requires a hypoxic environment, while physiological normoxia could be required for active cardiomyogenesis[107]. Hypoxic c-KIT⁺ CSCs

within niches have been found throughout the myocardium, especially at the atria and apex. Throughout all ages, bundles of CSCs with low oxygen content coexist with normoxic CSCs niches. Hypoxic CSCs, especially in the atria, are quiescent cells undergoing cell cycle arrest and cannot divide. Normoxic CSCs are pushed into intense proliferation and differentiation with continuous telomere erosion, resulting finally in dysfunctional aged CMs[108]. Additionally, Nkx2.5 and GATA4 expressions are only restricted to the normoxic CSC niche. A balance between the hypoxic and normoxic niche is essential for the preservation of the CSC compartment and for the maintenance of myocardial homeostasis during the organ lifespan. Some factors such as aging cause an imbalance by expanding the hypoxic quiescent CSCs so that less pools of cycling CSCs maintain cell turnover[100]. Hypoxic cardiac niches are abundant in the epicardium and subepicardium in an adult mouse heart, which also fosters a metabolically distinctive population of glycolytic progenitor cells[109].

The pool of CSCs seems to be heterogeneous, incorporating quiescent and actively proliferating cells, migratory and adherent cells, uncommitted and early committed cells, with young and senescent cells. Additional surface epitopes remain to be disclosed to classify pools of CSCs holding specific properties. Surface Notch1 expression distinguishes multipotent CSCs that are poised for lineage commitment, while c-Met and ephrin type-A receptor 2 receptors reveal cells with particular migratory potential out of the niche area. A specific compartment of CSCs, expressing IGF-1 receptor, can be stimulated to regenerate damaged myocardium, while those expressing IGF-2 receptor hold higher probability for senescence and apoptosis. Although this arrangement of cells seems to equip properly the CSC with homeostasis regulation, it does not effectively protect against aging or ischemic injury of the heart [100].

CSCs RELATIONSHIP WITH OTHER CELLS

Circulatory angiogenic cells

Circulatory angiogenic cells (CACs) are endothelial progenitor cells involved in vasculogenesis, angiogenesis, and stimulating myocardial repair, mainly through paracrine action. Latham *et al*[110] demonstrated that conditioned medium from CAC-CSC co-cultures exhibited greatly mobilized CACs, with induction of tubule formation in human umbilical vein endothelial cells, mainly through the upregulation of the angiogenic factors angiogenin, stromal cell-derived factor 1 (SDF-1 α), and VEGF. Moreover, administration of CACs and CSCs in infarcted hearts of non-obese/severe combined immunodeficient mice restored substantially the left ventricular ejection fraction (LVEF), with reduction of scar formation as revealed by echocardiography. Successful yet modest SMCs, ECs, and CM differentiation has been also reported.

Saphenous vein-derived pericytes

Pericytes (also called Rouget cells, mural cells, or perivascular mesenchymal precursor cells) are mesodermal cells that border the endothelial lining. They are highly proliferative cells and express neural/glial antigen 2, SOX-2, PDGFR- β , CD34, and several mesenchymal markers such as CD105, CD90, and CD44. It was previously reported that the transplantation of saphenous vein-derived pericytes (SVPs) into an ischemic limb of an immunodeficient mice restored the local circulatory network *via* angiogenesis[111]. Moreover, treatment with SVP reduced fibrotic scar, CM death, and vascular permeability in a mouse model of MI *via* miRNA-132 facilitated angiogenesis [112]. Avolio *et al*[113] were the first to describe the relationship between SVP and the endogenous CSCs. Combined CSC and SVP transplantation in the infarcted myocardium of severe combined immunodeficient/Beige-immunodeficient mice showed similar results to treatment with CSCs or SVP cells *per se*, regarding scar size and ventricular function, indicating that SVPs alone are as potent as CSCs.

TCs

TCs represent a recently described cell population in the stromal spaces located in many organs, including the heart. They are broadly dispersed throughout the heart and comprise a network in the three cardiac layers, heart valves, and in CSC niches. TCs have been documented also in primary culture from heart tissues[114,115]. The ratio of cardiac TCs (0.5%-1%) exceeds that of CSCs. Although they still represent a minute portion of human cardiac interstitial cells, their extremely long and extensive telopodes allow them to occupy more surface area, forming a 3D platform probably that extends to support other cells[116]. The telopodes act as tracks for the sliding of

precursor cells towards mature CMs and their integration into heart architecture[91]. TCs form a tandem with CSCs/CPCs in niches, where they communicate through direct physical contact by atypical junctions or indirect paracrine signaling[115].

TC-CSC co-culturing have suggested that TCs and CSCs act synergistically to control the level of secreted proteins, as shown by the increased levels of monocyte chemoattractant protein 1 (MCP-1), macrophage inflammatory protein1 α and 2 (MIP-1 α and MIP-2), and interleukin (IL)-13. Whereas, the level of IL-2 decreased compared to the monoculture of CSCs or TCs. IL-6 found in TC culture is behind the upregulation of these chemokines. Chemokines elucidated the role of TCs in directing the formation of CMs. Within the context, MIP-1 α and MCP-1 play roles in the formation of SMCs in the airway. Additionally, MCP-1 is also involved in mouse skeletal muscle regeneration by recruiting macrophages. The enhancement of MCP-1 secretion serves as an activator of another cell population, primarily macrophages, which are generally involved in such processes[117].

IL-6 also activates downstream signaling pathways and contributes to cardioprotection and vessel formation in the heart through activation of gp130/signal transducer and activator of transcription 3. The Gp130/signal transducer and activator of transcription 3 is essential for the commitment of cardiac SCA-1⁺ cells into endothelial lineage[118].

Furthermore, IL-6 targets *VEGF* and hepatocyte growth factor (*HGF*) genes. *VEGF* has a mitogenic effect on CMs[119]. It is known to mobilize bone marrow-derived mesenchymal stem cells (BM-MSCs) into the peripheral blood in MI patients[120]. *HGF* and its receptor (c-Met) are also involved in cardiogenesis, as it is expressed early during cardiac development[121]. The level of *HGF* mRNA is normally low in the heart, but it is upregulated for at least 14 d after ischemic insult in rats, enhancing CMs survival under ischemic conditions[122,123]. Moreover, it has the potential to generate an adhesive micro-environment for SCs, as demonstrated in a study of transplantation of *HGF* transfected BM-MSCs in the infarcted myocardium[124]. *HGF* is also a powerful angiogenic agent, conducting its mitogenic and morphogenic effects through the expression of its specific receptor in various types of cells, including myocytes. Moreover, *HGF* exerts antifibrotic and antiapoptotic effects on the myocardium[125, 126].

Transcriptomic analysis also has disclosed that TCs express pro-angiogenic miRNAs including let-7e, miRNA-21, miRNA-27b, miRNA-126, miRNA-130, miRNA-143, miRNA-503, and miRNA-100[127]. The TCs and CSCs interact *in vitro* forming atypical junctions, such as puncta adherentia and stromal synapses. The puncta adherentia consists of cadherin-catenin clusters. It controls the symmetry of division by facilitating the proper positioning of centrosomes. Therefore, an increased number of CSCs has been reported to be encountered in the presence of cardiac TCs[128,129].

CSCs SECRETOME

The paracrine potential of CSCs/CPCs has been recently under focus. CSC-derived cytokines and growth factors include epidermal growth factor (EGF), *HGF*, IGF-1, IGF-2, IL-6, IL-1 α , and TGF- β 1[130,131]. Exosomes appear to harbor relevant reparative signals, which mechanistically underlie the beneficial effects of CSCs transplantation [132].

Structurally, exosomes are lipid bilayer nano-sized organelles, 20-150 nm in diameter, secreted from all cell types, and function as intercellular communicators. Exosomes are highly heterogenic in content, and this stems from the unique packaging process that occurs inside progenitor and SCs. Exosomes carry lipids, proteins, and nucleic acids, with an abundance of miRNAs that hold profound post-transcriptional gene regulatory effects[133].

Protein content of exosomes

Amongst the distinctive protein content of cardiac exosomes are the chaperone proteins heat shock protein (HSP) 70 and HSP60. The HSP70 and HSP60, which under normal conditions assist in protein folding processes and deter misfolding and protein aggregation under pathological states induced by stress, also play major roles in apoptosis[134]. Circulating exosomes from healthy individuals have been found to activate cardioprotective pathways in CMs *via* HSP70 through extracellular signal-regulated kinase 1/2 and HSP27 phosphorylation[135].

The exosome protein cargo of CPCs is distinct from BM-MSCs, fibroblasts, and other sources as it contains ample amounts of the pregnancy-associated plasma protein-A

(PAPP-A). PAPP-A is present on the surface of human exosomes and interacts with IGF binding proteins (IGFBPs) to release IGF-1[136]. The cardioprotective role of CPCs-exosomes has been proven experimentally in *in vitro* ischemia/reperfusion and MI models and on CMs apoptosis to surpass that of BM-MS-C-exosomes owing to their rich content of PAPP-A[137].

Exosomes' surface and intra-vesicular markers

Like all exosomes, mouse CPCs-derived exosomes are positive for the surface markers CD63, CD81, and CD9, TSG-101, and Alix, however, they express a high-level of GATA4-responsive-miRNA-451. MiRNA-451 has been shown to inhibit CM apoptosis in an acute mouse myocardial ischemia-reperfusion model through inhibition of the caspases 3/7. The expression of miRNA-21 in the mouse CPCs-exosomes additionally justifies their CM protection against oxidative stress and antiapoptotic effects *via* inhibition of programmed cell death protein 4 (PDCD4)[138]. Human CPCs-exosomes are enriched with miRNA-210, miRNA-132, and miRNA-146a-3p, which account for the diminished CM apoptosis, enhanced angiogenesis, and improved LVEF[139]. MiRNA-146a-5p is the most highly upregulated miRNA in human CPCs-exosomes and targets genes involved in inflammatory and cell death pathways[137].

The CDCs contain CD34⁺ stromal cells of cardiac origin and are multipotent and clonogenic but not self-renewing[140]. CDCs secrete exosomes that induce cardiomyogenesis and angiogenesis, regulate the immune response, downgrade fibrosis, and improve the overall cardiac function[141,142]. Moreover, CDCs homogeneously express CD105 but not CD45 or other hematopoietic markers. They also exhibit a high expression of miRNA-126[143]. Circulating miRNA-126 may participate in cardiac repair during acute MI and has been demonstrated to be downregulated in heart damage[144].

Exosome secretion and function

While exosomes are constitutively secreted, changes in the surrounding microenvironment, such as hypoxia, can induce modifications in CPCs- and CM- derived extracellular vesicles. Hypoxic CMs secrete large extracellular vesicles containing long noncoding RNA neat 1 (LNCRNA NEAT1), which is transcriptionally regulated under basal conditions by p53, while during hypoxia it is regulated by the hypoxia inducible factor 2A. An uptake of the hypoxic CM-derived extracellular vesicles by fibroblasts can prompt the expression of profibrotic genes[145]. Oxidative stress may also induce the release of cardiac CPCs exosomes, which in turn inhibit apoptosis when taken up by H9C2 (rat cardiomyoblast cell line)[132]. Furthermore, oxidative stress stimulates secretion of miRNA-21 rich exosomes, which could inhibit H9C2 apoptosis by targeting PDCD4 and hence can be accounted as a new method to treat ischemia-reperfusion[138].

Intercellular communication *via* exosomes occurs as part of various biological processes, including immune modulation, vasculogenesis, transport of genetic materials, and pathological conditions such as inflammation, apoptosis, and fibrosis, which can lead to cardiovascular disease when altered[146]. Hence, isolation and analysis of cardiac exosomes contents, mainly miRNA and proteins, could offer diagnostic information for several cardiovascular diseases[147] (Figure 3).

Functionally, exosomes mediate several intra-cardiac inter-cellular communications such as:

CPC-CM crosstalk through factors, such as miRNA-146a and PAPP-A, which activate extracellular signal-regulated kinases 1/2 pathway and inhibit apoptosis[139].

CPC-macrophage (M1) crosstalk *via* miRNA-181b and Y-RNA fragment transforms M1 to M2 macrophages with attenuated proinflammatory cytokines and increased IL-10[148,149] (Figure 4).

CPC-fibroblast interaction *via* exosomes primes the fibroblasts and increases expression of VEGF and SDF-1. Experimental injection of fibroblasts primed with CPCs-exosomes into the myocardium of a MI model proved to reduce infarct size and improve cardiac function. In addition, cardiosphere-isolated exosomes have been used to prime inert fibroblasts, leading to an intensification of their angiogenic, cardiomyogenic, antifibrotic, and collective regenerative effects[150] (Figure 4).

CPC-self regulatory mechanisms: Exosomes derived from CPCs may play critical roles in maintaining the self-renewal state of CPCs themselves and balance their differentiation, *i.e.* preserve their stemness[151] (Figure 4). The CPC-derived exosomes activate the endogenous CPCs by transferring signal molecules directly within their niche[152].

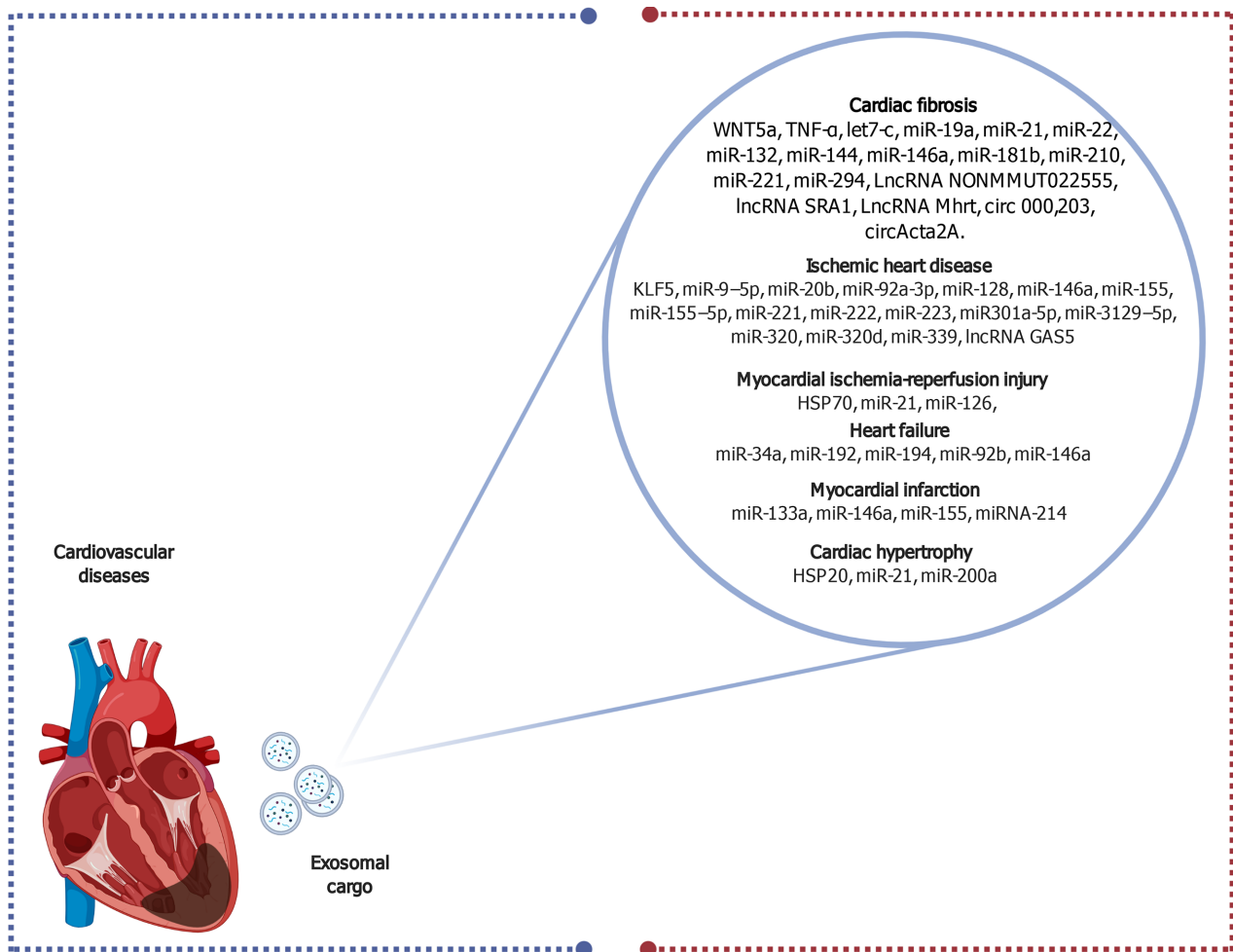


Figure 3 Schematic diagram elucidating the diverse exosomal contents that serve as biomarkers for several cardiovascular diseases.

Created with BioRender.com. HSP: Heat shock protein; lncRNA: Long non-coding RNA; miR: MicroRNA.

CPC-derived exosomes release various RNA species in the extracellular space, modulating endogenous SC plasticity and tissue regeneration through their cytoprotective, immunomodulatory, pro-angiogenic, and anti-apoptotic actions[153].

Fibroblasts and pericytes interact after transdifferentiating to myofibroblasts and deposit ECM causing cardiac fibrosis. These fibrotic changes are usually induced by cardiac damage and lead to scar formation. Exosomes serve as messengers for cell-to-cell communication during cardiac fibrosis[154]. Molecular mechanisms of cardiac fibrosis are primarily related to TGF- β pathways, IL-11 signaling pathway, nuclear factor- κ β pathway, and Wnt pathways[155]. Accordingly, the bioactive substances targeted at these pathways could hypothetically be applied in the treatment of cardiac fibrosis. Wnt3a, being highly expressed in exosomes, could activate the Wnt/ β -catenin pathway in cardiac fibroblasts by restricting GSK3 β activation[156]. Moreover, tumor necrosis factor α contained in exosomes can be transferred between cardiac myocytes. In general activation/inhibition of the exosomes conveying remodeling substance secretion or uptake can control the myocardial remodeling and repair following MI [154,157].

The highlighted complex cell-to-cell communication from endogenous or exogenous CSCs provides an optimal microenvironment for resident CPC proliferation and differentiation (Figure 4), rendering the environment receptive to transplanted CPCs. This adaptation is promoted through activation of pro-survival kinases, leading to the induction of a glycolytic switch in recipient CPCs[158].

Therapeutic efficiency of CPCs/CDCs exosomes

Data from experimental models suggest that the exosomal component of the CPC secretome can fully recapitulate the effects of cellular therapy on ischemic and non-ischemic heart models[140]. In an ischemia-reperfusion injury rat model, Ciullo and partners[159] have shown that the systemic injection of exosomes (genetically

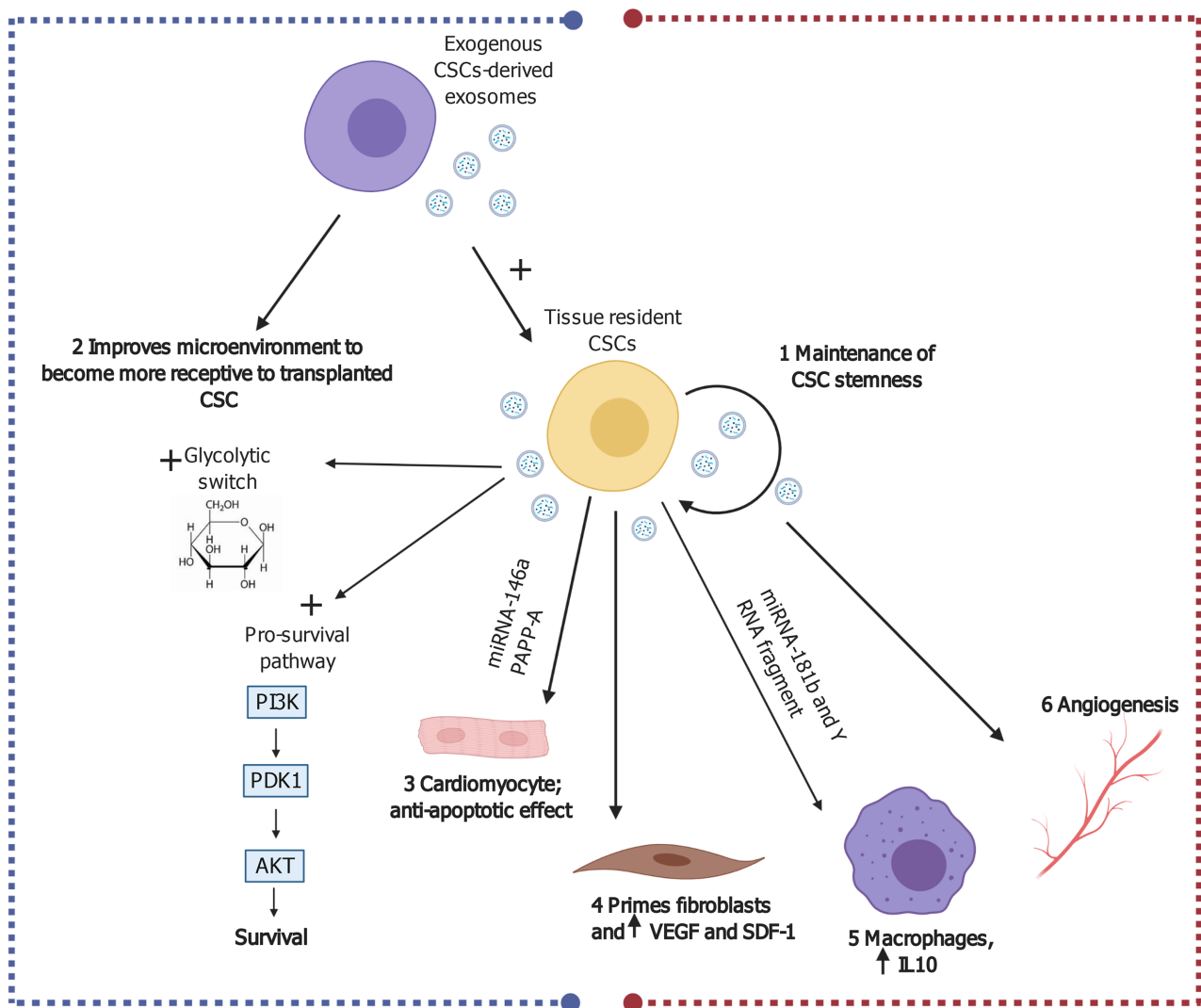


Figure 4 Possible cardiac reparative effects of cardiac stem cell/cardiosphere-derived cell-derived exosomes in myocardial ischemia and ischemia/reperfusion injury. Created with BioRender.com. CSC: Cardiac stem cell; IL: Interleukin; IR: Ischemia/reperfusion; miRNA: MicroRNA; PI3K: Phosphoinositide 3-kinase; SDF-1: Stromal cell-derived factor 1; VEGF: Vascular endothelial growth factor.

manipulated to overexpress CXCR4–ExoCXCR4) improve cardiac function. Additionally, expression of hypoxia-inducible factor 1 (HIF-1) in the infarcted myocardium is upregulated through the stimulation of SDF-1 α . The latter is one of the CXC chemokine family overexpressed in heart post-MI that readily attaches to the CXCR4 receptor and acts as a potent chemoattractant for CXCR4 expressing circulating progenitor cells. The ExoCXCR4 are more bioactive in the infarcted zone than naturally occurring exosomes injected *via* tail-vein, confirming their superior homing and cardioprotective properties in the damaged heart.

Gallet *et al*[160] postulated the safety and efficiency of CDC-derived exosomes in acute and chronic myocardial injury animal models. Within the context of experimental research to validate the paracrine hypothesis for CDCs-derived exosomes, it has been proven that human CDC-exosomes can recapitulate CDC therapy and boost cardiac function post-MI in pig models. Intramyocardial injection of human CDC-exosomes has resulted in higher exosome retention and efficacy as compared to intracoronary injection, with great reduction of scar size and increased ejection fraction. This indicates that the route of administration is imperative for full functional capacity of the exosomes. Subsequently, the researchers have devised a randomized preclinical study by means of a NOGA-guided intramyocardial exosome injection. Decreased collagen content in the infarct and border zone and increased neovascularization and Ki67⁺ CMs are indicative of the reparative functions of CDC-exosomes. Notably, human CDC-exosomes have shown a lack of an immune reaction, as seen by the lack of inflammatory reactions or CM necrosis in pig models. These observations strongly support the view that CDC-exosomes are ready to be tested in clinical trials.

Similar promising outcomes were observed in a Duchenne muscular dystrophy model (mdx), in which intramyocardial injection of CDC-exosomes efficiently recapitulated the effects of CDC injection on cardiac function, leading to recovery of movement. Administration of CPC-derived exosomes has resulted in transient restoration of partial expression of full-length dystrophin in mdx mice[161]. Further studies assessed the therapeutic potential of CPC-exosomes in a doxorubicin cardiotoxicity model and non-ischemic heart disease[162]. In addition, two concluded phase I clinical trials in patients with heart failure and revealed the capacity of CDCs to enhance cardiac function by reducing ventricular remodeling and scar formation. Despite receiving a single injection at the beginning of the study, the improvement in cardiac function was noted after the 1-year follow-up. This finding consequently leads to the proposition that transplanted CDCs mainly have imposed their actions at the site of injury by secreting paracrine factors including exosomes. In other words, CDC-exosomes achieved a biphasic beneficiary regenerative effect involving acute cardio protection coupled with long-term stimulation of endogenous cardiac repair[163].

METABOLISM OF CSCs

While the fetal heart obtains most of its ATP supply *via* glycolysis[164], the adult heart relies mainly on fatty acid oxidation to fulfill the contracting myocardium high energy demand[164,165]. The loss of the regenerative phenotype is related to the oxidative metabolism of glucose and fatty acids[166,167] and is mediated by various physiological changes including increased workload and the demand for growth, which cannot be solely met by glycolysis[168,169], as well as postnatal increase in both circulating levels of free fatty acids and blood oxygen levels[164,165]. Studies have shown the involvement of the HIF-1 signaling pathway[170], peroxisome proliferator-activated receptor α (PPAR α)[171], and peroxisome proliferator-activated receptor γ coactivator-1 (PGC-1) in the switch toward oxidative metabolism[172], which is accompanied by dramatic increase in the number of mitochondria in CMs[173].

Notably, similar metabolic reprogramming occurs during differentiation from cardiac SCs to CMs[167]. Studies reported that after differentiation into CMs, there is an increase in the mitochondrial number and activity[174], increased oxidative metabolism[175], and increased respiratory capacity resulting in an increased adenosine diphosphate:ATP ratio[173] after differentiation into CMs.

The fact of the various metabolic changes that accompany the transition from glycolysis to fatty acids oxidation affect cardiac cell maturation[164,167] has mandated the consideration of substrate composition in cardiac differentiation protocols[167].

A study by Malandraki-Miller *et al*[176] investigated the effect of fatty acid supplementation, which mimics the metabolic switch from glucose to fatty acid oxidation, on adult cardiac progenitors. The study used radiolabeled substrate consumption for metabolic flux to investigate the role of the PPAR α /PGC-1 axis during metabolic maturation. Oleic acid stimulated the PPAR α pathway, enhanced the maturation of the cardiac progenitor, and increased the expression of MHC and connexin after differentiation. Moreover, total glycolytic metabolism, mitochondrial membrane potential, the expression of glucose, and fatty acid transporter increased. The recorded results contributed greatly in highlighting the role of fatty acids and PPAR α in CPC differentiation.

Another study by Correia *et al*[177] has linked substrate utilization and functional maturation of CMs *via* studying the effect of the metabolic shift from glucose to galactose and fatty acid-containing medium in the maturation of hPSCs-derived CMs (hPSCs-CMs). The shift accelerated hPSC-CM maturation into adult-like CMs with higher oxidative metabolism, mature transcriptional signatures, higher myofibril density, improved calcium influx, and enhanced contractility. Galactose improved total oxidative capacity with reduction of fatty acid oxidation, thereby protecting the cells from lipotoxicity.

In CDCs, oxidative metabolism and cell differentiation reciprocally affect each other. *In vitro* cultures for CDCs revealed a PPAR α agonist that triggers fatty acid oxidation. Metabolic changes have been characterized as the CDC differentiated towards a cardiac phenotype. Addition of a PPAR α agonist at the onset of differentiation has induced a switch towards oxidative metabolism, as shown by changes in gene expression with decreasing glycolytic flux and increasing oxidation of glucose and palmitate. Undifferentiated CDCs have generated high levels of ATP from glycolysis and from oxidation of acetoacetate. Upon differentiation, oxidative metabolism of glucose and fatty acids is upregulated with decreased oxidation of

acetoacetate, a metabolic phenotype similar to that of the adult heart[178].

Taken together, the metabolic hallmarks of differentiated CMs vary from their undifferentiated SCs. Energy substrate metabolism during cardiac development and differentiation shows gradual decrease in the contribution of glycolysis to ATP synthesis with simultaneous increase in fatty acid-dependent mitochondrial respiration[179].

Common methods for the investigation of substrate metabolism include the measurement of metabolic fluxes using radio-labeled substrates, such as D-U-14C-glucose[180,181] as well as measurement of mitochondrial oxygen consumption rate and extracellular acidification rate using the XF Extracellular Flux Analyzer (Seahorse Bioscience, North Billerica, MA, United States)[182,183].

Recently, a detailed protocol for metabolic characterization of hiPSCs-CMs has been developed. The hiPSCs are obtained from adult somatic cells *via* novel cell reprogramming approaches, followed by differentiation to CMs. The novel *in vitro* cardiac cellular model provided new insights into studying cardiac disease mechanisms and therapeutic potentials. The characterization protocol measures small metabolites and combines gas- and liquid-chromatography-mass spectrometry metabolic profiling, lactate/pyruvate, and glucose uptake assays as important tools[184]. Integration between the implemented assays has provided complementary metabolic characteristics besides the already established electrophysiological and imaging techniques, such as monitoring ion channel activities[185], measurement of action potentials, changes in Ca²⁺ fluxes[186], and mitochondria viability and apoptosis[187].

An alternative pathway for glucose metabolism in CMs involves the entry of glucose-6-phosphate (G6P) in the pentose phosphate pathway, with resultant generation of reduced nicotinamide adenine dinucleotide phosphate (NADPH)[188]. Reduced NADPH helps to regenerate reduced glutathione and thus acts protectively against reactive oxygen species induced cell injury.

The cardioprotective role of the pentose/G6P/NADPH/glutathione pathway has been emphasized by Jain *et al*[189] who demonstrated that G6P dehydrogenase (G6PD) lacking mice have more severe heart damage induced by the myocardial ischemia reperfusion injury in Langendorff-perfused hearts as compared with wild-type mice.

Moreover, Katare *et al*[190] studied this pathway in CPCs isolated from hearts of diabetic mice. They reported that both G6PD and transketolase activities were markedly reduced in diabetes mellitus, which resulted in apoptosis of CMs. Interestingly, they have also reported that apoptosis was induced under high glucose conditions *via* inhibition of the pentose phosphate pathway, which mediates prosurvival signaling pathways.

Cellular metabolic transcriptome profile is an important determinant of many critical cell functions such as survival, growth, differentiation, and reprogramming. With the fast-track advancements in CSCs research, in-depth and thorough metabolic transcriptome analyses on CSCs are needed. It has been also suggested that metabolic genes can be targeted to manipulate the differentiation of ESCs into specific CM phenotypes or to modulate the maturation grade of CMs derived from ESCs[179].

As mentioned earlier in the review, the energy demand of the contracting myocardium of an adult heart is met mainly through fatty acid oxidation, which explains the fact that genes required for fatty acid metabolism are upregulated in the differentiated CMs. These genes include acetyl-CoA acyltransferase 2 (*ACAA2*), NADH dehydrogenase ubiquinone 1, α/β subcomplex 1 (*NDUFAB1*), protein kinase AMP-activated α -2 catalytic subunit (*PRKAA2*), and ECII enoyl-CoA delta isomerase 1 (*DCI*). In addition, other genes involved in glucose metabolism are also upregulated in α -MHC⁺ CMs, including protein phosphatase 1 regulatory subunit 3C (*PPP1R3C*), glycogen phosphorylase, muscle associated (*PYGM*), enolase 3 (*ENO3*), phosphoglycerate mutase 2 (*PGAM2*), amylo- α -1,6-glucosidase 4- α -glucanotransferase (*AGL*), 6-phosphofructokinase muscle (*PFKM*), and malate dehydrogenase 1 (*MDH1*)[191]. This is interpreted by the fact that adult cardiac cells are metabolically flexible, being capable of oxidizing other energy sources, such as glucose, lactate, amino acids, and ketone bodies for the production of ATP and non-ATP-producing intermediate metabolites with high biological significance[169]. Another example of CMs' metabolic plasticity is shown by *HIF-1* expression, which is important for their metabolic adaptation to hypoxic and ischemic conditions[192].

NANOTECHNOLOGY AND CSCs

Since Richard Feynman laid down the foundation of nanotechnology 1959[193],

remarkable developments have been witnessed attributed to the novel properties possessed by the materials at the nanoscale, which differ from their bulk forms. A panel of NPs, ranging from soft to inorganic, are used in nanomedicine, depending on their unique property matching the field of interest[194-197] (Figure 5).

The interplay of nanotechnology with SCs has gained increasing interest, whether for differentiating, tracking, imaging, or for therapeutic purposes. Accumulating evidence presents that the small size and bioactive characteristics of NPs could influence SC function. Several engineering techniques have been developed to obtain nano-fibrous scaffolds that facilitate controlling SC proliferation, migration, and differentiation. A diversity of SCs, including ESCs, skeletal myoblasts, BM-MSCs, and CSCs, have been tested to repair acutely or chronically damaged myocardium. However, the optimal cell type, the efficient cell number, the appropriate route for cell delivery, and the ideal time point for cell delivery after MI are still unanswered questions. The biodistribution of SCs and the specific mechanism by which therapeutic cells improve cardiac function remains under investigation[198]. Using NPs could solve some of these obstacles either by gene delivery to SCs, enhancing the retention of SCs, facilitating SCs' proangiogenic effect, or mimicking the ECM[199].

In view of this, we have summarized the impacts of NPs on SCs, especially CSCs, from differentiating, therapeutic, and tracking viewpoints (Figure 5, Table 1).

NP-assisted CSC differentiation

Nanotechnology has revolutionized the tissue engineering field and altered the landscape of scaffolds syntheses. In typical tissue engineering, a bio-mimicked scaffold provides adhesive surfaces for the seeded cells, where the SCs deposit their proteins to make the engineered-scaffold more biocompatible. However, improper vascularization, lack of functional cells, the low mechanical strength of engineered cells, immunological incompatibility with the host, and nutrient constraints are the main limitations encountered in tissue engineering. Therefore, synthesis of a biomimetic scaffold at the nanoscale, in a minimum of one dimension, would offer a more effective microenvironment needed for cell growth. Nano-tissue engineering provides the scaffold with a simple substrate for SC adhesion and active agents for their proliferation[200].

In consideration for CSCs, nano- and microstructured electrospun matrices have been used as non-woven scaffolds for the construction of cardiac tissue from primary CMs. Among different nanostructured poly (D, L-lactic-co-glycolic acid; PLGA) membranes, the poly (L-lactide; PLLA) scaffolds superiorly developed mature contractile machinery (sarcomeres). Functional studies (excitability) of CMs tested by optical imaging of electrical activity have confirmed the superior response on PLLA scaffolds compared with other ones[201].

An *in vitro* and *in vivo* study conducted by Liu *et al*[202] using porous ECM-mimicking nanofibrous PLLA scaffolds (porous NF PLLA) demonstrated cardiac tissue formation from CPCs. The scaffold has facilitated the *in vitro* differentiation of isolated mouse ESCs into CPCs. Thereafter, the transplanted NF PLLA/CPCs integrated successfully with the host tissue, with superior expression of cardiac committed markers cardiac troponin T, smooth muscle MHC, and CD31.

The inductive and therapeutic properties of biodegradable PLGA nanofibers have been tested *in vitro* and *in vivo*. Different hiPSCs-CMs have been seeded on aligned PLGA nanofibers to differentiate into high-quality cardiac tissue-like constructs, where cardiac biomarkers and cardiac functions have been upregulated. When utilized *in vivo* for treating MI, the cardiac tissue-like constructs have shown more robust results than the two-dimensional conventional control in improving the ejection fraction, the fractional shortening, and left ventricular end-systolic diameter[203].

Recently, an injectable ECM hydrogel loaded with gold (Au)/Laponite (Lap) nanocomposite has been tested on the biological activity of resident CSCs. The electroactive Au/Lap-ECM hydrogel improved cell biocompatibility and phenotypes maturation of cardiac-specific markers (SAC, cardiac troponin 1, and Cx43)[204].

NP-assisted CSC therapy

More than 90 years ago, the Nobel laureate German immunologist Paul Ehrlich proposed the term "magic bullets" to describe the artificial biochemical agents that would transport and release drugs at the desired sites only[205]. Since then, drug delivery research has witnessed notable growth due to NPs utilization as "controlled release reservoirs" for drug delivery in order to combat many diseases[206].

In cardiovascular diseases, NP-based drug delivery targeting CSCs would be a successful therapeutic regimen. In an *in vivo* study of induced MI, self-assembling peptide nanofibers tethered with NF-IGF-1 positively influenced CPCs in female

Table 1 Main outcomes of the studies investigated the impact of nanotechnology in cardiac stem cell-based studies

Nanotechnology field	Types of nanoparticles	Type of cardiac disease/stem cells	Type of research	Outcomes	Ref.
Tissue engineering	Semi-crystalline PLLA nanostructured membranes among several PLGA membranes	Non-diseased/primary CMs	<i>In vitro</i> : electrospun matrices were used as scaffolds for generating cardiac tissue constructs	Nanostructured non-woven PLLA scaffolds provide flexibility and guidance for CMs growth and can be successfully applied to obtain structurally and functionally competent cardiac tissue constructs.	[201]
Tissue engineering	ECM-mimicking nanofibrous PLLA scaffolds with porous structure (porous NF PLLA) of high interconnection for cardiac tissue formation	Non-diseased/mouse ESCs CPCs	<i>In vitro</i> : CPCs with porous NF PLLA <i>In vivo</i> : male athymic nude mice	<i>In vitro</i> : porous NF PLLA scaffolds facilitate cell attachment, extension, and differentiation. <i>In vivo</i> : subcutaneous implantation of cell/scaffold supports survival of grafted cells and differentiation to CMs, SMCs, ECs lineages.	[202]
Tissue engineering/therapeutic	Biodegradable ANF	MI/hiPSCs-CMs	<i>In vitro</i> : hiPSCs (253G1) <i>In vivo</i> : nude rat	<i>In vitro</i> : multilayered, elongated, organized CMs at high density along ANF, with up regulation of genes of sarcomere structures (<i>ACTN2</i> , <i>TNNT2</i> , <i>TNNI3</i>), cardiac maturation (<i>MYH7</i>), ventricular structures (<i>MYL2</i> , <i>HAND2</i>). <i>In vivo</i> : CTLCs improve MI functionally due to transplantation of organized functional CMs.	[203]
Tissue engineering	Electroactive Au-Lap NPs loaded myocardial ECM	Non diseased/Resident CSCs	<i>In vitro</i> : rat CMs from 2-d old neonatal rats	Combination of electrically active nano-formulations and biologically active ECM boost the expression of cardiac-specific proteins (SAC, cTnl, Cx43).	[204]
Therapeutic	Self-assembling peptide nanofibers tethered with insulin-like growth factor-1 (NF-IGF-1)	MI/CPCs	<i>In vitro</i> : clonogenic CPCs <i>In vivo</i> : female Fischer 344 rats	<i>In vitro</i> : NF-IGF-1 promote CPCs division (↑BrdU) and protect them from death signal (↓TdT). <i>In vivo</i> : CPCs-NF-IGF-1 enhance postinfarction ventricular remodeling, attenuate chamber dilation, and improve cardiac performance.	[207]
Therapeutic	Transplantation of self-assembling nanopeptides: Cell-PM complex	MI/cSCA-1 ⁺ cardiac progenitors Other stem cells BM, SM, AMC	<i>In vivo</i> : Wild-type mice (C57Bl/6j); Adult GFP transgenic mice	cSCA-1/PM attenuates ventricular enlargement, restore cardiac function, with high capillary density (↑vWF) and conductive vessels (↑αSMA, ↑VEGF). ↓TUNEL ⁺ CMs in the infarct area of cSCA-1/PM.	[208]
Therapeutic	CMMP contained control-released stem cell factors in its polymeric core and cloaked with hCSC membrane fragments on the surface	MI/Human CSCs	<i>In situ</i> : characterization <i>In vitro</i> : NRCM <i>In vivo</i> : male SCID Beige mice	<i>In situ</i> : CMMPs express hCSC surface markers. <i>In vitro</i> : CMMPs promote NRCM contractility and proliferation. <i>In vivo</i> : CMMPs preserve viable myocardium,	[209]

Therapeutic and drug delivery tool	Statin PLGA nanoparticles	MI/hAdSCs	<i>In vivo</i> : male nude mice (BALB/c nu/nu)	augment cardiac functions, with safety profile. A small number of intravenously administered SimNP-loaded AdSCs improve cardiac function following MI, stimulating endogenous cardiac regeneration in the infarcted myocardium.	[244]
Tracking of treatment	Colloidal nanoparticles containing europium loaded on collagen matrix	MI/Lewis rat BM-MSCs	<i>In vivo</i> : female Fischer rat	Collagen matrix enhance transplanted MSC retention and reduce migration of the cells into remote organs as tracked by the radioactive NPs.	[211]
Tracking and magnetic targeting of treatment	Superparamagnetic iron microspheres	MI/Rat CDCs	<i>In vitro</i> : rat CDCs <i>In vivo</i> : female WKY rats	<i>In vitro</i> : \downarrow caspase 3 ⁺ , \downarrow TUNEL ⁺ . <i>In vivo</i> : enhanced cell engraftment, with attenuated left ventricular remodeling and increased ejection fraction. \uparrow GFP ⁺ , \uparrow Ki67 ⁺ CMs, and \uparrow GFP ⁻ /c-KIT ⁺ cells.	[212]
Imaging and therapeutic by magnetic targeting	Ferumoxytol (FDA-approved SPIONs) nanoparticles linked by heparin sulfate and protamine sulfate	MI/Human and ratCDCs	<i>In vitro</i> : hCDCs and rCDCs <i>In vivo</i> : female WKY rats	<i>In vitro</i> : \downarrow TUNEL ⁺ , \downarrow ROS and \uparrow CCK-8, \uparrow Ki67. <i>In vivo</i> : augmentation of acute cell retention and attenuation of left ventricular remodeling, 3 wk after treatment by MRI, fluorescence imaging, qPCR.	[213]
Imaging and tracking for differentiation	Potassium niobate harmonic nanoparticles stabilized by polyethylene glycol	Non-diseased/ESC-derived CMs	<i>In vitro</i> : mouse ESC (CGR8 cell line)	Monitoring at high acquisition speed the rhythmic contractions of ESC-derived CMs beating within 3D cluster.	[245]
Therapeutic by magnetic guidance of NPs	Iron oxide nanoparticle-incorporated nanovesicles (exosome memetic nanovesicles); (IONP-NVs)	MI/MSCs	<i>In vitro</i> : rat CM, rat CFs, macrophage, HUVECs. <i>In vivo</i> : Fischer 344 rats	<i>In vitro</i> : under hypoxia IONP-MSCs exert Antiapoptotic effect on CMs: \downarrow caspase 3 ⁺ , \uparrow Cx43, \uparrow PI3K. Antifibrotic effect on CFs: \uparrow Cx43, \downarrow TGF β 1, \downarrow α Acta2, \downarrow MMP2, \downarrow MMP9. Anti-inflammatory effect on macrophage. Proangiogenic effect on HUVECs: \uparrow tube formation, \uparrow EC migration. <i>In vivo</i> : magnetic guidance increases IONP-MSCs retention within the infarcted heart, with early shift from inflammatory stage to reparative stage.	[217]

AMC: Adipose tissue-derived mesenchymal cell; ANF: Aligned PLGA nanofibers; Au-Lap NP: Gold and laponite nanoparticle; BrdU: Bromodeoxyuridine cell proliferation assay; CCK-8: Cell counting kit-8; CF: Cardiac fibroblast; CM: Cardiomyocyte; CMMP: Cell-mimicking microparticle; CPC: Cardiac progenitor cell; CTLC: Cardiac tissue-like construct; cTnl: Cardiac troponin 1; FDA: Food and Drug Administration; ECM: Extracellular matrix; ESC: Embryonic stem cell; hAdSC: Human adipose-derived stem cell; HUVEC: Human umbilical cord vein endothelial cell; IONP: Iron oxide nanoparticle; MRI: Magnetic resonance imaging; NF: Nanofibrous; MI: Myocardial infarction; MMP: Matrix metalloproteinase; NRCM: Neonatal rat cardiomyocyte; PLGA: Poly D,L-lactic-co-glycolic acid; PLLA: Poly (L-lactic acid); PM: Puramatrix™; qPCR: Quantitative polymerase chain reaction; SM: Skeletal myoblast; SPION: Superparamagnetic iron oxide NP; TdT: Terminal deoxynucleotidyl transferase apoptotic assay; TUNEL: Terminal deoxynucleotidyl transferase dUTP nick end labeling apoptotic assay; vWF: von-Willebrand factor; WKY: Wistar-Kyoto; α SMA: Alpha smooth muscle actin; 3-D: Three-dimensional.

Fischer rats. The local injection of CPCs loaded on NF, with the prolonged release of

Other self-assembling nanopeptides loaded with cell-Puramatrix™ complex have revealed promising results in treating MI, especially when targeting clonal SCA-1⁺ CPCs. The infarct area of SCA-1⁺/PM became smaller than that of other tested SCs. Moreover, SCA-1⁺/PM have secreted VEGF, enhancing their differentiation potentiality into CMs and vascular SMCs[208].

Interestingly, a synthetic cell-mimicking microparticle (CMMP) has recapitulated CSC function with a safe immunological profile. The core of the CMMP is a PLGA containing human-derived CSC (hCSC) secretome, while the surface has been cloaked with SC membrane fragments. These CD105⁺ and CD90⁺ CMMPs have shown synchronized movement with adjacent beating CMs *in vitro*. When injected in the MI mouse model, they have shown prolonged retention without eliciting the T-cell immunoreaction that transplanted hCSCs provoked[209].

NP-assisted CSC imaging and tracking

Combining the therapeutic effect of SC-based therapy with concomitant *in vitro* or *in vivo* visualization of the SCL is a strategy helped by the auto-luminescent NPs. Furthermore, delivery of the SCs to the infarcted cardiac area could be guided by a magnetic field depending on the unique magnetic properties of iron oxide NPs[210].

Isotopic colloidal NPs tagged with europium have successfully tracked the retention of BM-MSCs in the infarcted area. The NP-labeled MSCs loaded on a collagen matrix have shown reduced relocation of MSCs to remote organs. However, delivering the NP-MSCs *via* collagen has failed to improve cardiac function[211]. Further retention and magnetic targeting of CDCs have been retrieved using superparamagnetic microspheres (SPM). Quantitative polymerase chain reaction and optical imaging have confirmed the magnetic targeting and the increased cardiac retention of transplanted cells, with reduced lung migration in a rat model of ischemia/reperfusion injury. Moreover, the prolonged survival of SPM-labelled CDCs by cell counting kit-8 and Western blot has proved the safety profile of SPM[212].

The success of magnetic cell delivery in various preclinical studies potentiates the translation into clinical ones encouraged with the Food and Drug Administration-approved superparamagnetic iron oxide NP ferumoxytol. A thorough investigation of ferumoxytol-labeled (FHP) human and rat CDCs offered the potential for rapid clinical translation of the magnetically targeted cell delivery to an ischemic heart. The *in vitro* study proved that FHP nanocomplex is not toxic to hCDCs, where a panel of cytotoxicity assays have revealed prolonged survival, potentiated differentiation, and genetic stability of FHP-hCDCs. Furthermore, *in vivo* tracking of FHP-rCDCs by magnetic resonance imaging (MRI), fluorescence imaging, and quantitative polymerase chain reaction have shown that magnetic targeting increased cardiac retention without eliciting cardiac inflammation or causing iron overload. The histological assessment revealed enhancement of angiogenesis and cell engraftment in the hearts of the magnetic targeting group[213].

NP-based cardiac therapy limitations and prospects

Despite the promising results of NP-assisted SC interventions, the reported nanotoxicity is considered a major obstacle for clinical translation of these preclinical trials [214]. Besides, most of the *in vitro* and *in vivo* NP-based SC trials focused on the short-term effect of the NP interventions. The long-term safety profile of the injected NPs/CSCs or NPs/scaffolds with host interactions needs large scale investigations. These shortcomings have directed the search for natural cell-derived immune compatible nanostructures.

Exosomes attracted the attention as therapeutic cellular-derived NPs. However, the small quantity of these exosomes secreted from SCs is considered the main limitation to its therapeutic implementation. A novel exosome-mimetic extracellular nanovesicles (NVs) have bypassed this obstacle[215]. The large-scaled mechanically synthesized NVs from ESCs, by Jo and his colleagues[216], have conserved both RNA and protein profiles of the ESCs. Furthermore, treatment of MSCs with NVs has promoted cellular proliferation, which has been comparable with or even superior to the positive MSCs control treated with silica nanobeads that are well known for their ability to stimulate proliferation *via* activation of the MAPK pathway.

Recently, NVs derived from iron oxide NPs (IONPs) incorporated MSCs (IONP-MSCs) have co-cultured with different primary cell lines to investigate their physical and biological characteristics. IONPs-MSCs have revealed cardioprotective effects *via* PI3K/AKT activation. Under *in vitro* hypoxic conditions, IONPs-MSCs have upregulated Cx43, an electrical coupling molecule, whose reduction is responsible for arrhythmia and cardiac cell death in hypoxia. Furthermore, the NVs inhibited cardiac fibrosis by inhibiting the differentiation of cardiac fibroblasts into cardiac myofibro-

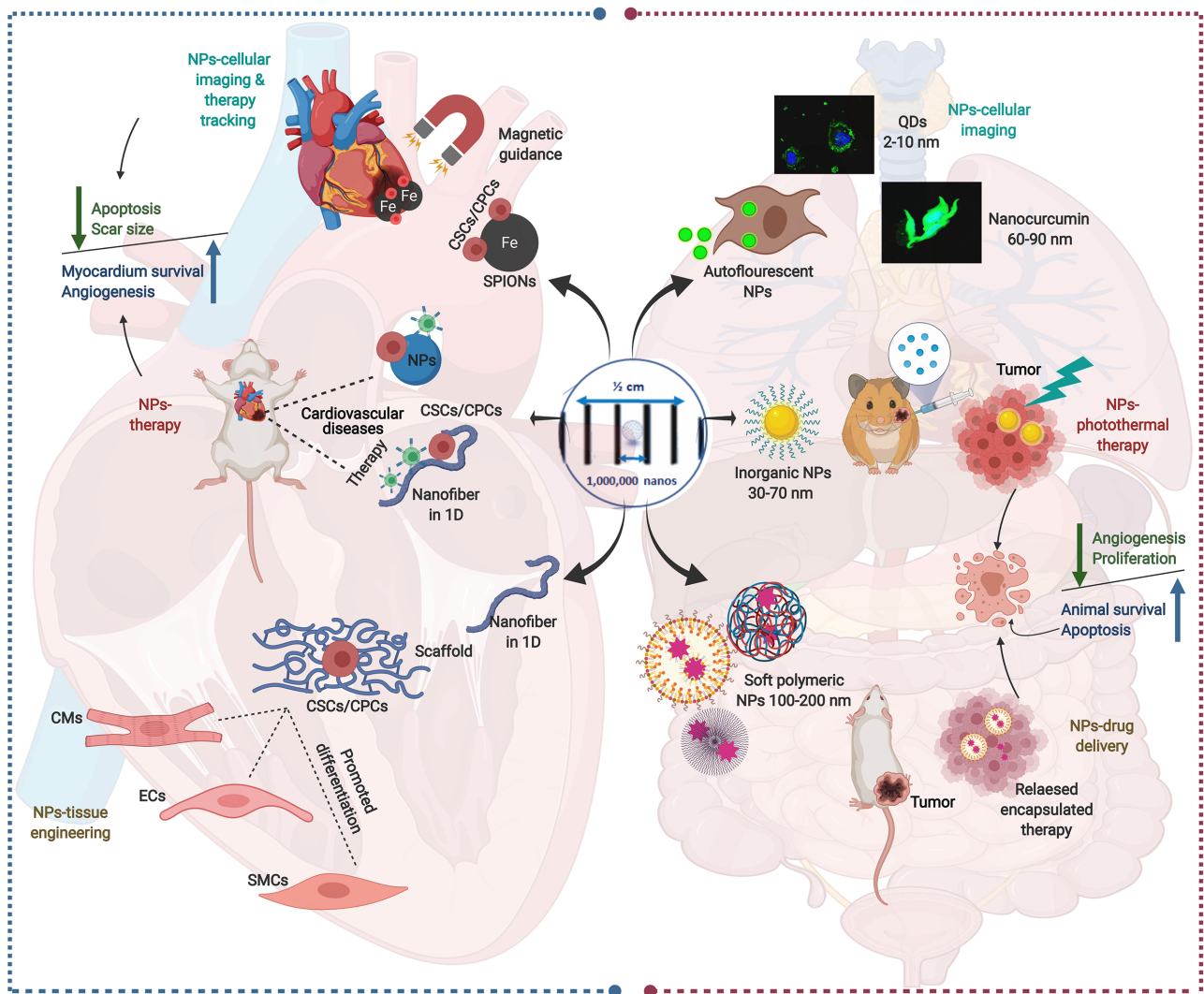


Figure 5 Schematic presentation of multiple nanoparticles-based paradigms in nanomedicine (right side) and their mirror images in nanoparticle-assisted cardiac stem cell interventions (left side). In the right side, the auto-luminescence of quantum dots and some polymeric natural nanoparticles (NPs) as nanocurcumin qualifies them to be used primarily in tracking and diagnostic imaging. Meanwhile, inorganic NPs have been extensively investigated in cancer treatment as photothermal therapy due to the plasmonic resonance of their outer electron. For drug delivery purposes, soft NPs, such as liposomes, polymericelles, and dendrimers, are used due to their flexibility and biophysical interaction with components of the cell membrane, which enable them to penetrate biological membranes. In the left side, the impact of different NPs on cardiac stem cell (CSC)-based therapy are shown. The superparamagnetic iron oxide NPs are used for tracking and imaging of the CSCs-therapy, while nanofibers are used either as a dual delivery system for CSCs and therapy. Moreover, nanofiber scaffold holding the CSCs is used in tissue engineering to assist CSCs' differentiation. The confocal images of NPs-cellular imaging are reproduced with permission [196]. Created with BioRender.com. NPs: Nanoparticles; SMCs: Smooth muscle cells; CMs: Cardiomyocytes; ECs: Endocardial cells; CSCs: Cardiac stem cells; CPCs: Cardiac progenitor cells; SPION: Superparamagnetic iron oxide NP.

blasts after hypoxia. Magnetically targeting an induced MI with IONPs-MSCs have attenuated apoptosis, reduced inflammation, and increased blood vessel density, with increased retention of IONPs-MSCs in the infarcted myocardium, improving left ventricular remodeling[217].

Designer exosomes is another hope, boosting the exosomal theragnostic potential independently of its low yielding. Kojima and coworkers[218] genetically engineered the producer cells with the three genes responsible for potentiating exosome production: Six-Transmembrane Epithelial Antigen of Prostate 3 for exosome biogenesis, syndecan-4 for budding of endosomal membranes, and the fragment of L-aspartate oxidase for cellular metabolism. These exosome production boosters have been further genetically upgraded to RNA packaging device and cytosolic delivery helper. Thereafter, they have used this collectively EXOTic device to deliver cargo messenger RNA to the mice brain, attenuating neurotoxicity in neuroinflammatory disease by enhancing cell-to-cell communication without the need for exosomes concentration. These results demonstrate the usefulness of designer exosomes for therapeutic RNA delivery that can be applied to CPC/CDC exosomes to enhance their efficacy.

CLINICAL TRIALS

The rationale of clinical studies testing CSC therapy on human subjects has been based on the conclusions from the promising results of experimental studies utilizing SCs [219,220]. Clinical trials using CSCs, CPCs, and CDCs have been and are still targeting variable cardiac diseases, namely ischemic heart diseases, non-ischemic cardiomyopathy, heart failure, congenital heart diseases, and pulmonary atrial hypertension (Figure 6, Tables 2 and 3).

Ischemic heart diseases

There have been numerous ischemic heart diseases (IHD) trials. Cardiosphere-derived autologous stem cells to reverse ventricular dysfunction (CAUDEUS) and allogeneic heart stem cells to achieve myocardial regeneration (ALLSTAR) are two complementary studies in which CAUDEUS is the phase I and ALLSTAR is the phase II of clinical trial of intracoronary delivery of autologous CDCs in patients with IHD and left ventricular dysfunction. Firstly, CAUDEUS, with 6 mo and 12-mo follow-up of patients has shown significant reduction in the scar size (-7.7%, -11.1% respectively), increased viable myocardium (+13 g, +22.6 g respectively), and improvement of the regional function of infarcted myocardium[221]. Then, ALLSTAR results showed attenuation of the post infarct cardiac remodeling and improvement in the left ventricular diastolic volume, but it has failed to achieve the results of CAUDEUS regarding the reduction of scar size; hence ALLSTAR was dismissed[222].

The clinical trial transplantation of human embryonic stem cell-derived progenitors in severe heart failure (ESCORT) was developed to assess the regenerative effects of human ESC-derived CPCs. At the beginning of coronary artery bypass grafting, patients received a fibrin gel implanted with the hESC derived CPCs. The main conclusion from this trial demonstrated the technical feasibility of producing a clinically operational product. This product is hESCs derived CPCs that can be safely transplanted to patients with severe ischemic left ventricle (LV) dysfunction. It also supports their short- and medium-term safety after transplantation in patients with severe post-infarction LV dysfunction[223]. Direct engraftment of CPCs into cardiac tissue along with the paracrine effects of CPCs are the main milieu of cardiac regeneration[224,225]. Additionally, fibrin gel has been used as a delivering synthetic biomaterial to improve the long term cell engraftment in the ischemic environment [226].

Safety and efficacy evaluation of intracoronary infusion of allogeneic human cardiac stem cells in patients with AMI (CAREMI), a phase I/II placebo-controlled clinical trial, has been designed to evaluate the safety, practicability, and efficiency of intracoronary transport of allogeneic adult CSC in patients with large ST segment elevation secondary to myocardial infarction, LV dysfunction at risk of developing heart failure. CAREMI has intended to interfere with allogeneic cells immediately after the initial ischemic insult (dodging the aggressive part of the first 5 d) and before myocardial scar starts to form (within 7 d after percutaneous infusion). CAREMI trial is documented proof that allogeneic CSCs intracoronary infusion early after acute MI is safe with rational efficacy outcomes and is promising hope for future clinical trials with Allo-CSCs[227].

Non-ischemic cardiomyopathy

Halt cardiomyopathy progression in Duchenne (HOPE) and dilated cardiomyopathy intervention with allogeneic myocardially-regenerative cells (DYNAMIC) studies aim to assess safety and to discover the usefulness of CDCs in patients with non-ischemic cardiomyopathy. HOPE was targeted specifically to explore efficacy of CDCs in patients with advanced stages of Duchenne muscular dystrophy. The results have shown significant and sustained improvements in cardiac structure, function, and significant reduction in cardiac scarring as compared with the control group[228]. Meanwhile, the DYNAMIC study does not have any published data so far.

Heart failure

In autologous human cardiac-derived stem cell to treat ischemic cardiomyopathy (ALCADIA), intramyocardial delivery of CDCs with controlled releases of basic FGF in a biodegradable gelatin hydrogel sheet to patients with ischemic heart failure was done. This was at the time of coronary artery bypass grafting. After 6 mo, it was reported that small improvements in regional but not global function, *i.e.* LVEF (+ 9%-12%), had been noted as well as decreased scar sizes 3.3% at 6-mo follow-up *vs* baseline[229]. A randomized, double-blinded phase II clinical trial started in 2015 to

Table 2 Completed clinical trials reporting on the use of cardiac derived stem cells in cardiovascular disorders

Study name, NCT Number	Start year	Study phase	Cardio-vascular disease	Patients number	Type of cardiac stem cells/origin	Route of delivery /count of cells	Timing of cell delivery	Follow up times	Imaging techniques	Outcomes	Ref.
CADUCEUS, 00893360	2009	I	IHD	17	Autologous CDCs/endomyocardial biopsies	IC/12.5-25 × 10 ⁶	1.5-3 m post STEMI	6 mo	MRI	Significant reduction in infarct size Significant growth in viable mass global LVEF remains unchanged LVEF and volumes.	[221]
ALLSTAR, 01458405	2017	I/II	IHD	90	Allogenic CDCs/endomyocardial biopsies	IC/25 × 10 ⁶	< 5 d post MI	12 mo	MRI	No effect on scar size Attenuation of post infarct cardiac remodeling Improvement in LV end diastolic volume	[222]
ALCADIA, 00981006	2010	I	CHF/IHD/D	6	Autologous hCSCs/endomyocardial biopsies + bFGF on gelatin hydrogel sheet	IM/0.5 × 10 ⁶ /kg and 200 µg of bFGF	At CABG	12 mo	Not mentioned	Decreased scar size	[229]
ESCORT, 020579000	2013	I	IHD	6	ESCs-derived ISL1 ⁺ CSCs	Epicardial patch 5-10 × 10 ⁶ CSCs embedded in a fibrin patch	At CABG	18 mo	CT PET scan Echo	Symptomatically improved patients with an increased systolic motion of the cell-treated segments. The protocol generated a highly purified population of cardiovascular progenitors. One patient died of heart failure after 22 mo	[223]
CAREMI, 02439398.	2014	I/II	AMI	55	Allogeneic hCSCs/right atrial appendage	IC/35 × 10 ⁶ cells	5 to 7 d after successful reperfusion of AMI by PCI or 8 d from symptoms onset	1 wk, 1, 2, 3, 4, 5, 6, 9 and 12 mo	MRI ECG	Allogeneic CSCs intracoronary infusion early after AMI is safe and anticipates reasonable efficacy outcomes	[227]
CONCERT-HF, 02501811.	2015	II	HF	125	Autologous c-KIT ⁺ CPCs + MSCs/right ventricular endocardial biopsy + Bone marrow aspiration	Trans endocardial/150 × 10 ⁶ MSCs and 5 × 10 ⁶ CPCs	14 wk after cell harvest	6 and 12 mo	MRI Treadmill Questionnaire	Increased LVEF Decrease in infarct size with LV end systolic volume reduction. Strong safety profile	[230]
HOPE, 02485938	2015	I\II	CM secondary to DMD	25	Allogeneic CDCs	IC/75 × 10 ⁶	Not specified	12 mo	MRI Questionnaire	Significant scar reduction improvement in inferior wall systolic thickening compared to the usual care group.	[228]

DYNAMIC, 02293603	2014	I	Idiopathic dilated CM	42	Allogeneic CDC/not specified	IC/Stepwise dose escalation	Not specified	6, 12 mo	ECG Cardiac enzymes	CDCs are generally safe and well-tolerated Not published	-
TICAP, 01273857	2011	I	HLHS	7	Autologous CDCs/right atrial appendage	IC/0.3 × 10 ⁶ cells/kg	1 m after cardiac surgery	36 mo	Echo	Safety of the procedure Increased right ventricle ejection fraction Improved somatic growth Reduced heart failure status	[233]
Perseus, 01829750	2013	II	HLHS	34	Autologous CDCs/not specified	IC/0.3 × 10 ⁶ cells/kg	4 to 9 wk after surgery	3 and 12 mo follow- up	EchoMRIQuestionnaire	Significant improve of ventricular function Improved somatic growth, and quality of life Reduced heart failure status and cardiac fibrosis compared with baseline	[231]

AMI: Acute myocardial infarction; bFGF: Basic fibroblast growth factor; CABG: Coronary artery by bass graft; CHF: Congestive heart failure; CM: Cardiomyopathy; CSC: Cardiac stem cell; DMD: Duchenne muscle dystrophy; hCSC: Human-derived cardiac stem cell; HLHS: Hypoplastic left heart syndrome; IC: Intracoronary; IHD: Ischemic heart disease; IHF: Ischemic heart failure; IM: Intramyocardial; MRI: Magnetic resonance imaging; NCT: National clinical trial; PCI: Percutaneous infusion; STEMI: ST segment elevation after MI; VD: Ventricular dysfunction.

study the efficacy of combinatorial therapy in treatment of cardiac ischemic disorders. Combination of mesenchymal and c-KIT⁺ CSCs as regenerative therapy for heart failure (CONCERT-HF) utilized autologous endomyocardial CSCs combined with MSCs for heart failure patients. After trans-endocardial injection of this combined therapy, the LVEF increased to 89%, with about 4.5% decrease in infarct size and LV end systolic volume. This is the first clinical trial to assess the benefits of cell combination therapy in humans, which addresses the therapeutic potential for cardiac ischemic patients[230]. In addition, according to the ClinicalTrials.gov Identifier (NCT02503280), there is an expected clinical study (TAC-HFT-II) that might start in 2025 to study the safety and effectiveness of trans-endocardial injection of combination of MSCs and CSCs in patients with post-MI HF. Moreover, another ongoing randomized, double blind, placebo-controlled phase II clinical trial for regression of fibrosis and reversal of diastolic dysfunction in heart failure with preserved ejection fraction patients treated with allogeneic CDCs (Regress-HFpEF, NCT02941705) was conducted to determine whether treatment with intracoronary allogeneic CDCs will affect clinical functional status of HFpEF patients, regarding regression of fibrosis and reversal of diastolic dysfunction.

Table 3 Ongoing and future expected clinical trials

Study name, NCT number	Year of study start	Study phase	Cardiovascular disease	Number of patients	Type of cardiac cells	Route of delivery
Regress-HFpEF, 02941705	2017	II	Symptomatic hypertensive heart disease induced HFpEF	40	Allogeneic CDCs	IC
APOLLON trial, 02781922	2016	III	HLHS	40	Autologous CSCs	IC
CHILD, 03406884	2019	I	HLHS	32	Autologous c-KIT ⁺ cells	IM
ALPHA, 03145298	2017	I	PAH	26	Allogeneic CDC	IV
TAC-HFT-II, 02503280	2025	I/II	IHF	0 enrollment until now	Autologous combination of MSCs and CSCs	Trans endocardial Injection

CSC: Cardiac stem cell; HFpEF: Heart failure with preserved ejection fraction; HLHS: Hypoplastic left heart syndrome; IC: Intracardiac; IHF: Ischemic heart failure; IM: Intramyocardial; IV: Intravenous; NCT: National clinical trial; PAH: Pulmonary atrial hypertension.

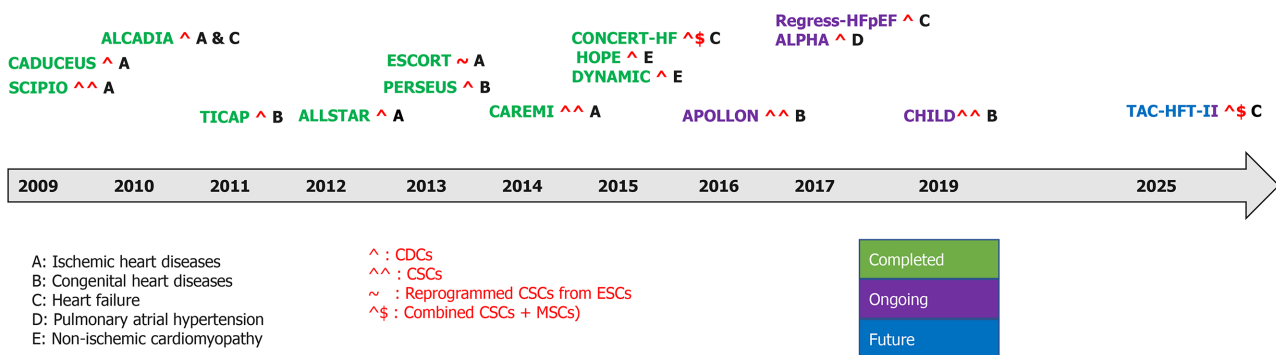


Figure 6 Timeline of completed, ongoing, and future clinical trials in cardiovascular diseases. CDCs: Cardiosphere-derived cells; CSCs: Cardiac stem cells; ESCs: Embryonic stem cells; MSCs: Mesenchymal stem cells.

Congenital heart diseases

There are reports on phase I, II, and III clinical trials assessing the use of autologous CDCs in pediatric patients. One of the early clinical trials in children is the transcatheter infusion of CPCs in patients with single ventricle physiology (TICAP) trial. It has confirmed the practicality of intracoronary delivery of CDCs in post palliative single ventricle physiology patients. In this study, autologous CDCs have been isolated, expanded, and then finally administered *via* intracoronary delivery 4-5 wk after the stage II/III palliative surgery. TICAP has proved that the methodology is safe and achievable for improving cardiac function after 18 mo. The safety of the CDC therapy has been also analyzed at 36 mo post-transplantation. Neither tumor formation nor arrhythmias have been reported, and the initial observed benefits have enhanced, with attenuation of ventricular stiffness and improvement of ventriculoarterial coupling[231].

Following the TICAP trial, the cardiac progenitor cell infusion to treat univentricular heart disease (PERSEUS) trial, a phase II randomized controlled trial has been planned to follow-up on the TICAP trial. CDCs have been given *via* the coronary arteries after stage II or III palliative operations[232]. The main result is the change in EF from baseline to 3 mo that has been measured by cardiac MRI and echocardiography. The CDC treated group showed greater improvement in right ventricle (RV) function at 3 mo than controls (+6.4% *vs* +1.3%) and at 1 year continued to exhibit augmented RV function. Follow-up analysis at 36 mo demonstrated an improvement in RVEF among patients receiving CDCs (+8.0% *vs* +2.2%) even at this late time point. Furthermore, the therapy is currently being tested in a phase III trial cardiac stem/progenitor cell infusion in univentricular physiology (APOLLON). The phase III trial is an extension of the TICAP and PERSEUS trials exploiting autologous CDCs administered by intracoronary injection in single ventricle patients, for which results are still awaiting[233]. In addition, there is another ongoing phase I clinical trial (CHILD, NCT03406884) for examining the safety and efficacy of intramyocardial

injection of c-KIT CSCs injection in patients with hypoplastic left heart syndrome.

Pulmonary atrial hypertension

Pulmonary arterial hypertension (PAH) is a progressive condition for which there is no cure. As reported in PAH archives and with the pharmacologic developments in the current treatment scope, survival remains poor. Preclinical studies suggest that the transplantation of allogeneic CDCs could reduce unfavorable PAH related arteriolar remodeling. Until now, there are no completed studies on PAH, though, there is an ongoing phase I clinical trial [(ALPHA), NCT03145298] exploring the potential beneficial effects of central intravenous delivery of allogeneic human CDCs.

THERAPEUTIC POTENTIAL OF COMBINED CELLULAR THERAPY

Despite the great differentiation potential of CSCs and their ability to restore the injured area, it is influenced by many factors, such as oxidative stress and the aging process, which diminish its function. Thus, the administration of mixtures of SCs can exhibit synergistic effects to get the maximum benefit from the transplanted cells in cardiac regeneration. Cell combination therapies are classified into various entities; the most commonly performed is combination of CSCs with MSCs either applied from autologous or allogeneic sources. The most recent combinatorial approaches are CardioChimerias (CCs) and CardioClusters. Merging several cell types harmonized efficiently cardiomyoplasty in preclinical cardiac ischemic models[234,235].

The mechanism behind the effect of MSCs when combined with CSCs has been clarified in a study conducted by Hatzistergos *et al*[236], where transendocardial injection of MSCs derived from allogeneic male bone marrow in a reperfused MI swine model led to myocardial repair by stimulating and increasing the differentiation potential of endogenous CSCs to regenerate myocardium. Remarkably, 2 wk after MSC injection, the MSC-treated hearts showed chimeric collections containing both exogenous immature MSCs and endogenous CSCs with a 20-fold increase in c-KIT⁺ cells, indicating cardiac lineage commitment. In addition, to confirm the origin of CSCs, porcine endomyocardial biopsies have been isolated and plated as explants with or without MSC feeder layers, signifying that the MSCs have stimulated the expansion of c-KIT⁺ CSCs. The MSC co-culture derived CSCs have been more than 90% positive for Nkx2-5, which is considered an embryonic heart phenotype. MSCs have facilitated cardiac repair through stimulating a succession of secondary endogenous responses that triggered considerable amounts of adult CMs and immature CSCs to multiply and repair the injured areas with CMs, vascular SMCs, and ECs of host origin.

From this point of view, a study conducted by Williams *et al*[237] to assess the combination of CSCs and MSCs isolated from a human cardiac tissue and iliac crest, respectively, where 1×10^6 human CSCs and 200×10^6 human MSCs were injected intra-myocardially in the infarcted area in immunosuppressed pigs after 2 wk of MI induction. All cell-based cardiomyoplasty groups showed a decrease in the infarct size 4 wk following cell injection with a 2-fold greater reduction in scar mass with combined SCs in comparison with each therapy alone. The results have been evaluated by cardiac MRI and conductance catheterization hemodynamics, revealing the improvement of cardiac response to MI functionally and structurally with returning of EF values to baseline, where the combined cell therapy enhanced the LV performance during systole and diastole.

In addition to cardiomyogenic repair in acute MI, the combination therapy enhances cardiac performance, both functionally and structurally, in chronic ischemic cardiomyopathies. The hypothesis was adopted by Karantalis *et al*[238]. The autologous CSCs isolated from the septal wall of the right ventricle immediately after MI induction and the autologous MSCs obtained from the tibial cavity were injected trans-endocardially in the same non-immunosuppressed pigs that undergo ischemic injury for 3 mo. The favorable effects of combinational therapy occurred in chronic MI models, where the size of the infarcted area diminished with improvement of EF, diastolic strain, and cardiac output in comparison with single therapy with MSCs.

In agreement with previous studies to explore the effect of allogeneic combination therapy in non-immunosuppressed swine chronic ischemic cardiomyopathy model, Natsumeda *et al*[239] addressed the advantage of such therapy from allogeneic sources to reduce the evoked immune response to administrated CSCs alone. The MSCs inhibited the rejection of the immune system to allo-CSCs, improving the retention of the combined cell therapy in the affected area to maintain the regenerative capacity. Moreover, in comparison with the autologous combination approach, angiogenesis

increased with improvement of perfusion to the previously infarcted area. Furthermore, the added CSCs to allo-MSCs augmented the contractile function of cardiac muscle by enhancing CM proliferation. This denotes that allogeneic combination cell therapy exerts cardiomyogenic regeneration in chronic ischemic conditions without immune rejection from the host.

The enumerated promising findings demonstrate the privileges of combinational therapy to attain the desirable effects of the differentiation potentiality of CSCs with the supporting and paracrine effects of MSCs in cardiac regeneration. However, the effectiveness of this therapy has been compromised by certain factors such as the persistence and long-term cardiogenic repair. Additionally, optimization of the dose of each cell type in the combination mixture is undefined. Therefore, new *in vitro* cell engineering techniques[240-243] have emerged to unify the maximum favorable effects of this therapy and eliminate the limitations concerning cardiac repair.

A novel modality has been aroused by Zang *et al*[240] to enhance the engraftment and angiogenesis properties of c-KIT⁺ CSCs to precondition the cells with exosomes-derived MSCs, resulting in increased proliferation, formation of angiotube, and enhanced migration *in vitro*. Moreover, in a rat MI model after 28 d, the CSCs combined with exosomes decreased the infarct size with neovascularization and increased the capillary density with higher EF relative to treated groups with CSCs only. The beneficial effect of exosomes on CSCs was evaluated by miRNA network analysis that exhibit influences on Wnt, VEGF, and PTEN/PI3K/Akt signaling pathways. These effects have enhanced CSCs potentials in cardiac repair by adding new properties to CSCs such as increased proliferation, differentiation, and angiogenesis to the pre infarcted area with defense against oxidative stress.

Quijada *et al*[241] is the first study conducted to administer a new cardiac hybrid murine MI model created by *in vitro* viral fusion between CPCs and MSCs with further clonal expansion known as CCs in order to improve and boost combinatorial cell delivery approaches. In this study, two variable clones were selected from 18 clones after 1 mo of fluorescent activated cell sorting of fused fluorescent-tagged CPCs (mcherry) and MSCs (enhanced green fluorescent protein) with an inactivated RNA Sendai virus to accomplish artificial cell fusion. It has been found that CCs exhibit augmented proliferation and survival properties in addition to opposing cell death (CC1 and CC2) compared with individual cell and combination therapies in a MI mouse model. The CCs injected into the acutely damaged heart have recurred structural integrity with functional improvement up to 18 wk with reduction of infarct size at 12 wk and earlier correction of EF at 6 wk. Moreover, they are not subjected to cell aging relative to combinational and single cell therapy. The tremendous effect of CCs was presented in enhanced engraftment in the border zone of infarcted area, which denotes the selectivity of this hybrid in addition to the already retained properties of CSCs and MSCs in cardiomyogenic repair. Notably, in comparison with mixed dual cell therapy the preservation of CM size and induction of c-KIT⁺ cells have occurred at 12 wk, with enhanced capillary density maintaining a longstanding cardiac vasculature.

In agreement with reparative potentials of CCs, Firouzi *et al*[242] applied the principle of cell fusion between c-KIT⁺ cardiac interstitial cells (cCICs) and MSCs in a 2:1 ratio with inactivated Sendai virus. The cells were isolated from human cardiac samples removed during left ventricular assisted device implantation, then were allowed for clonal expansion to study the phenotypic features and function as compared to the parent cells. It was been found that the survival of human CCs is prolonged more than parent cells in addition to the increased survival of CMs when co-cultured with serum deprived neonatal rat CMs. Compared to CCs originating from a mouse model, its ability to resist oxidative stress is increased, thus enhancing survival potential and inheriting the same karyotype of the parent cells with diploid DNA content. However, it has not yet been tested in a MI model to assess the degree of retention or the cardiomyogenic repair potentials.

The evolution of next generation applications to improve the efficacy of combinatorial cell therapy in heart failure have been accomplished by Monsanto *et al*[243] who have implemented a 3D structure composed of MSCs in the center surrounded by cCICs, while endothelial progenitor cells (EPCs) inhabited the outer layer. These isolated autologous SCs in a ratio 3:2:1 of EPCs, cCICs, and MSCs, respectively, have been co-cultured to enhance the interactions between the cells before injection intramyocardially in a murine MI model. Moreover, this scaffold has inherited the same efficiency of each cell type. The EPCs protected the cCICs from oxidative stress and cell death with neovascularization to the infarcted area. Thus, the power of cCIC differentiation was increased in addition to the supporting milieu exerted by MSCs. The beneficiary outcomes were evidenced by the improved cardiac function in the

murine model initiated from the first week post MI injection and sustained up to 20 wk, with greater improvement in EF, end systolic, and diastolic volume. Due to the administered distinct cell ratios and the controllable 3D structure size, which minimize cell loss, the CCs were retained for a long time and hence provided maximum effects of cell combination therapies in cardiomyogenic repair[240].

CONCLUSION

CSCs/CPCs carry a great therapeutic potential in different cardiac diseases owing to their numerous cell population, rich secretome and interplay with other cells. Conducted preclinical studies and clinical trials had demonstrated promising results in regards to the improvement of cardiac function. Studies using these cells targeting variable cardiac diseases are still ongoing aiming to discover more of their regenerative power and thus decrease the burden of morbidity and mortality due to cardiac tissue loss.

REFERENCES

- 1 **Bergmann O**, Bhardwaj RD, Bernard S, Zdunek S, Barnabé-Heider F, Walsh S, Zupicich J, Alkass K, Buchholz BA, Druid H, Jovinge S, Frisén J. Evidence for cardiomyocyte renewal in humans. *Science* 2009; **324**: 98-102 [PMID: [19342590](#) DOI: [10.1126/science.1164680](#)]
- 2 **Hsieh PC**, Segers VF, Davis ME, MacGillivray C, Gannon J, Molkentin JD, Robbins J, Lee RT. Evidence from a genetic fate-mapping study that stem cells refresh adult mammalian cardiomyocytes after injury. *Nat Med* 2007; **13**: 970-974 [PMID: [17660827](#) DOI: [10.1038/nm1618](#)]
- 3 **Wang X**. Stem cells in tissues, organoids, and cancers. *Cell Mol Life Sci* 2019; **76**: 4043-4070 [PMID: [31317205](#) DOI: [10.1007/s00018-019-03199-x](#)]
- 4 **Krampera M**, Franchini M, Pizzolo G, Aprili G. Mesenchymal stem cells: from biology to clinical use. *Blood Transfus* 2007; **5**: 120-129 [PMID: [19204764](#) DOI: [10.2450/2007.0029-07](#)]
- 5 **Henning RJ**. Current status of stem cells in cardiac repair. *Future Cardiol* 2018; **14**: 181-192 [PMID: [29405072](#) DOI: [10.2217/fca-2017-0072](#)]
- 6 **Lee CY**, Kim R, Ham O, Lee J, Kim P, Lee S, Oh S, Lee H, Lee M, Kim J, Chang W. Therapeutic Potential of Stem Cells Strategy for Cardiovascular Diseases. *Stem Cells Int* 2016; **2016**: 4285938 [PMID: [27829839](#) DOI: [10.1155/2016/4285938](#)]
- 7 **Madigan M**, Atoui R. Therapeutic Use of Stem Cells for Myocardial Infarction. *Bioengineering (Basel)* 2018; **5** [PMID: [29642402](#) DOI: [10.3390/bioengineering5020028](#)]
- 8 **Swedlund B**, Lescroart F. Cardiopharyngeal Progenitor Specification: Multiple Roads to the Heart and Head Muscles. *Cold Spring Harb Perspect Biol* 2020; **12** [PMID: [31818856](#) DOI: [10.1101/cshperspect.a036731](#)]
- 9 **Faustino RS**, Terzic A. Interactome of a cardiopoietic precursor. *J Cardiovasc Transl Res* 2008; **1**: 120-126 [PMID: [20559906](#) DOI: [10.1007/s12265-008-9019-z](#)]
- 10 **Bondue A**, Tännler S, Chiapparo G, Chabab S, Ramialison M, Paulissen C, Beck B, Harvey R, Blanpain C. Defining the earliest step of cardiovascular progenitor specification during embryonic stem cell differentiation. *J Cell Biol* 2011; **192**: 751-765 [PMID: [21383076](#) DOI: [10.1083/jcb.201007063](#)]
- 11 **Lee JW**, Lee CS, Ryu YR, Lee J, Son H, Cho HJ, Kim HS. Lysophosphatidic Acid Receptor 4 Is Transiently Expressed during Cardiac Differentiation and Critical for Repair of the Damaged Heart. *Mol Ther* 2021; **29**: 1151-1163 [PMID: [33160074](#) DOI: [10.1016/j.ymthe.2020.11.004](#)]
- 12 **Meilhac SM**, Buckingham ME. The deployment of cell lineages that form the mammalian heart. *Nat Rev Cardiol* 2018; **15**: 705-724 [PMID: [30266935](#) DOI: [10.1038/s41569-018-0086-9](#)]
- 13 **Kula-Alwar D**, Marber MS, Hughes SM, Hinitz Y. Mef2c factors are required for early but not late addition of cardiomyocytes to the ventricle. *Dev Biol* 2021; **470**: 95-107 [PMID: [33245870](#) DOI: [10.1016/j.ydbio.2020.11.008](#)]
- 14 **Antimisiaris SG**, Mourtas S, Marazioti A. Exosomes and Exosome-Inspired Vesicles for Targeted Drug Delivery. *Pharmaceutics* 2018; **10** [PMID: [30404188](#) DOI: [10.3390/pharmaceutics10040218](#)]
- 15 **Lamouille S**, Xu J, Derynck R. Molecular mechanisms of epithelial-mesenchymal transition. *Nat Rev Mol Cell Biol* 2014; **15**: 178-196 [PMID: [24556840](#) DOI: [10.1038/nrm3758](#)]
- 16 **Stefanovic S**, Laforest B, Desvignes JP, Lescroart F, Argiro L, Maurel-Zaffran C, Salgado D, Plaidoux E, De Bono C, Pazur K, Théveniau-Ruissy M, Bérout C, Puceat M, Gavalas A, Kelly RG, Zaffran S. Hox-dependent coordination of mouse cardiac progenitor cell patterning and differentiation. *Elife* 2020; **9** [PMID: [32804075](#) DOI: [10.7554/eLife.55124](#)]
- 17 **Aval SF**, Lotfi H, Sheervalilou R, Zarghami N. Tuning of major signaling networks (TGF- β , Wnt, Notch and Hedgehog) by miRNAs in human stem cells commitment to different lineages: Possible clinical application. *Biomed Pharmacother* 2017; **91**: 849-860 [PMID: [28501774](#) DOI: [10.1016/j.biopha.2017.05.020](#)]
- 18 **Tzavlaki K**, Moustakas A. TGF- β Signaling. *Biomolecules* 2020; **10** [PMID: [32210029](#) DOI: [10.3390/biom10020029](#)]

- 10.3390/biom10030487]
- 19 **Li X**, Gao F, Wang X, Liang Q, Bai A, Liu Z, Chen X, Li E, Chen S, Lu C, Qian R, Sun N, Liang P, Xu C. E2A ablation enhances proportion of nodal-like cardiomyocytes in cardiac-specific differentiation of human embryonic stem cells. *EBioMedicine* 2021; **71**: 103575 [PMID: [34488017](#) DOI: [10.1016/j.ebiom.2021.103575](#)]
 - 20 **Desgrange A**, Le Garrec JF, Bernheim S, Bønnelykke TH, Meilhac SM. Transient Nodal Signaling in Left Precursors Coordinates Opposed Asymmetries Shaping the Heart Loop. *Dev Cell* 2020; **55**: 413-431.e6 [PMID: [33171097](#) DOI: [10.1016/j.devcel.2020.10.008](#)]
 - 21 **Phillips HM**, Mahendran P, Singh E, Anderson RH, Chaudhry B, Henderson DJ. Neural crest cells are required for correct positioning of the developing outflow cushions and pattern the arterial valve leaflets. *Cardiovasc Res* 2013; **99**: 452-460 [PMID: [23723064](#) DOI: [10.1093/cvr/cvt132](#)]
 - 22 **Darrigrand JF**, Valente M, Comai G, Martinez P, Petit M, Nishinakamura R, Osorio DS, Renault G, Marchiol C, Ribes V, Cadot B. Dullard-mediated Smad1/5/8 inhibition controls mouse cardiac neural crest cells condensation and outflow tract septation. *Elife* 2020; **9** [PMID: [32105214](#) DOI: [10.7554/eLife.50325](#)]
 - 23 **Morrell NW**, Bloch DB, ten Dijke P, Goumans MJ, Hata A, Smith J, Yu PB, Bloch KD. Targeting BMP signalling in cardiovascular disease and anaemia. *Nat Rev Cardiol* 2016; **13**: 106-120 [PMID: [26461965](#) DOI: [10.1038/nrcardio.2015.156](#)]
 - 24 **Andrés-Delgado L**, Galardi-Castilla M, Münch J, Peralta M, Ernst A, González-Rosa JM, Tessadori F, Santamaria L, Bakkers J, Vermot J, de la Pompa JL, Mercader N. Notch and Bmp signaling pathways act coordinately during the formation of the proepicardium. *Dev Dyn* 2020; **249**: 1455-1469 [PMID: [33103836](#) DOI: [10.1002/dvdy.229](#)]
 - 25 **Miao S**, Zhao D, Wang X, Ni X, Fang X, Yu M, Ye L, Yang J, Wu H, Han X, Qu L, Li L, Lan F, Shen Z, Lei W, Zhao ZA, Hu S. Retinoic acid promotes metabolic maturation of human Embryonic Stem Cell-derived Cardiomyocytes. *Theranostics* 2020; **10**: 9686-9701 [PMID: [32863954](#) DOI: [10.7150/thno.44146](#)]
 - 26 **Drowley L**, McPheat J, Nordqvist A, Peel S, Karlsson U, Martinsson S, Müllers E, Dellsén A, Knight S, Barrett I, Sánchez J, Magnusson B, Greber B, Wang QD, Plowright AT. Discovery of retinoic acid receptor agonists as proliferators of cardiac progenitor cells through a phenotypic screening approach. *Stem Cells Transl Med* 2020; **9**: 47-60 [PMID: [31508905](#) DOI: [10.1002/sctm.19-0069](#)]
 - 27 **Janesick A**, Shiotsugu J, Taketani M, Blumberg B. RIPPLY3 is a retinoic acid-inducible repressor required for setting the borders of the pre-placodal ectoderm. *Development* 2012; **139**: 1213-1224 [PMID: [22354841](#) DOI: [10.1242/dev.071456](#)]
 - 28 **Christ A**, Marzenke M, Willnow TE. LRP2 controls sonic hedgehog-dependent differentiation of cardiac progenitor cells during outflow tract formation. *Hum Mol Genet* 2020; **29**: 3183-3196 [PMID: [32901292](#) DOI: [10.1093/hmg/ddaa200](#)]
 - 29 **Badr AM**, Mahran YF. Notch pathway and its role in cardiovascular system: Review. *Sys Rev Pharm* 2020; **11**: 342-349 [DOI: [10.31838/srp.2020.10.56](#)]
 - 30 **Bardot ES**, Jadhav B, Wickramasinghe N, Rezza A, Rendl M, Sharp AJ, Dubois NC. Notch signaling commits mesoderm to the cardiac lineage. *BioRxiv* 2020; 2020.2002.2020.958348 [DOI: [10.1101/2020.02.20.958348](#)]
 - 31 **De Zoysa P**, Liu J, Toubat O, Choi J, Moon A, Gill PS, Duarte A, Sucov HM, Kumar SR. Delta-like ligand 4-mediated Notch signaling controls proliferation of second heart field progenitor cells by regulating Fgf8 expression. *Development* 2020; **147** [PMID: [32778568](#) DOI: [10.1242/dev.185249](#)]
 - 32 **Gordon MD**, Nusse R. Wnt signaling: multiple pathways, multiple receptors, and multiple transcription factors. *J Biol Chem* 2006; **281**: 22429-22433 [PMID: [16793760](#) DOI: [10.1074/jbc.R600015200](#)]
 - 33 **Guo Y**, Dorn T, Kühl SJ, Linnemann A, Rothe M, Pfister AS, Vainio S, Laugwitz KL, Moretti A, Kühl M. The Wnt inhibitor Dkk1 is required for maintaining the normal cardiac differentiation program in *Xenopus laevis*. *Dev Biol* 2019; **449**: 1-13 [PMID: [30797757](#) DOI: [10.1016/j.ydbio.2019.02.009](#)]
 - 34 **Wang J**, Gong M, Zuo S, Xu J, Paul C, Li H, Liu M, Wang YG, Ashraf M, Xu M. WNT11-Conditioned Medium Promotes Angiogenesis through the Activation of Non-Canonical WNT-PKC-JNK Signaling Pathway. *Genes (Basel)* 2020; **11** [PMID: [33137935](#) DOI: [10.3390/genes11111277](#)]
 - 35 **Li HX**, Lin J, Jiang B, Yang XJ. Wnt11 preserves mitochondrial membrane potential and protects cardiomyocytes against hypoxia through paracrine signaling. *J Cell Biochem* 2020; **121**: 1144-1155 [PMID: [31463993](#) DOI: [10.1002/jcb.29349](#)]
 - 36 **Gao Y**, Pu J. Differentiation and Application of Human Pluripotent Stem Cells Derived Cardiovascular Cells for Treatment of Heart Diseases: Promises and Challenges. *Front Cell Dev Biol* 2021; **9**: 658088 [PMID: [34055788](#) DOI: [10.3389/fcell.2021.658088](#)]
 - 37 **Marino F**, Scalise M, Cianflone E, Mancuso T, Aquila I, Agosti V, Torella M, Paolino D, Mollace V, Nadal-Ginard B, Torella D. Role of c-Kit in Myocardial Regeneration and Aging. *Front Endocrinol (Lausanne)* 2019; **10**: 371 [PMID: [31275242](#) DOI: [10.3389/fendo.2019.00371](#)]
 - 38 **Xu J**, Lian W, Li L, Huang Z. Generation of induced cardiac progenitor cells via somatic reprogramming. *Oncotarget* 2017; **8**: 29442-29457 [PMID: [28199972](#) DOI: [10.18632/oncotarget.15272](#)]
 - 39 **Le T**, Chong J. Cardiac progenitor cells for heart repair. *Cell Death Discov* 2016; **2**: 16052 [PMID: [27551540](#) DOI: [10.1038/cddiscovery.2016.52](#)]

- 40 **Wandzioch E**, Edling CE, Palmer RH, Carlsson L, Hallberg B. Activation of the MAP kinase pathway by c-Kit is PI-3 kinase dependent in hematopoietic progenitor/stem cell lines. *Blood* 2004; **104**: 51-57 [PMID: 14996702 DOI: 10.1182/blood-2003-07-2554]
- 41 **Kuang D**, Zhao X, Xiao G, Ni J, Feng Y, Wu R, Wang G. Stem cell factor/c-kit signaling mediated cardiac stem cell migration *via* activation of p38 MAPK. *Basic Res Cardiol* 2008; **103**: 265-273 [PMID: 18087667 DOI: 10.1007/s00395-007-0690-z]
- 42 **van Berlo JH**, Kanisicak O, Maillet M, Vagnozzi RJ, Karch J, Lin SC, Middleton RC, Marbán E, Molkentin JD. c-kit+ cells minimally contribute cardiomyocytes to the heart. *Nature* 2014; **509**: 337-341 [PMID: 24805242 DOI: 10.1038/nature13309]
- 43 **Rota M**, Padin-Iruegas ME, Misao Y, De Angelis A, Maestroni S, Ferreira-Martins J, Fiumana E, Rastaldo R, Arcarese ML, Mitchell TS, Boni A, Bolli R, Urbanek K, Hosoda T, Anversa P, Leri A, Kajstura J. Local activation or implantation of cardiac progenitor cells rescues scarred infarcted myocardium improving cardiac function. *Circ Res* 2008; **103**: 107-116 [PMID: 18556576 DOI: 10.1161/CIRCRESAHA.108.178525]
- 44 **Malliaras K**, Zhang Y, Seinfeld J, Galang G, Tseliou E, Cheng K, Sun B, Aminzadeh M, Marbán E. Cardiomyocyte proliferation and progenitor cell recruitment underlie therapeutic regeneration after myocardial infarction in the adult mouse heart. *EMBO Mol Med* 2013; **5**: 191-209 [PMID: 23255322 DOI: 10.1002/emmm.201201737]
- 45 **van Vliet P**, Rocco M, Smits AM, van Oorschot AA, Metz CH, van Veen TA, Sluijter JP, Doevendans PA, Goumans MJ. Progenitor cells isolated from the human heart: a potential cell source for regenerative therapy. *Neth Heart J* 2008; **16**: 163-169 [PMID: 18566670 DOI: 10.1007/BF03086138]
- 46 **Holmes C**, Stanford WL. Concise review: stem cell antigen-1: expression, function, and enigma. *Stem Cells* 2007; **25**: 1339-1347 [PMID: 17379763 DOI: 10.1634/stemcells.2006-0644]
- 47 **Välimäki MJ**, Leigh RS, Kinnunen SM, March AR, de Sande AH, Kinnunen M, Varjosalo M, Heinäniemi M, Kaynak BL, Ruskoaho H. GATA-targeted compounds modulate cardiac subtype cell differentiation in dual reporter stem cell line. *Stem Cell Res Ther* 2021; **12**: 190 [PMID: 33736688 DOI: 10.1186/s13287-021-02259-z]
- 48 **Valente M**, Nascimento DS, Cumano A, Pinto-do-Ó P. Sca-1+ cardiac progenitor cells and heart-making: a critical synopsis. *Stem Cells Dev* 2014; **23**: 2263-2273 [PMID: 24926741 DOI: 10.1089/scd.2014.0197]
- 49 **Tsuji-Takayama K**, Inoue T, Ijiri Y, Otani T, Motoda R, Nakamura S, Orita K. Demethylating agent, 5-azacytidine, reverses differentiation of embryonic stem cells. *Biochem Biophys Res Commun* 2004; **323**: 86-90 [PMID: 15351705 DOI: 10.1016/j.bbrc.2004.08.052]
- 50 **Barreto S**, Hamel L, Schiatti T, Yang Y, George V. Cardiac Progenitor Cells from Stem Cells: Learning from Genetics and Biomaterials. *Cells* 2019; **8**: 1536 [PMID: 31795206 DOI: 10.3390/cells8121536]
- 51 **Takamiya M**, Haider KH, Ashraf M. Identification and characterization of a novel multipotent sub-population of Sca-1+ cardiac progenitor cells for myocardial regeneration. *PLoS One* 2011; **6**: e25265 [PMID: 21980409 DOI: 10.1371/journal.pone.0025265]
- 52 **Smits AM**, van Vliet P, Metz CH, Korfage T, Sluijter JP, Doevendans PA, Goumans MJ. Human cardiomyocyte progenitor cells differentiate into functional mature cardiomyocytes: an in vitro model for studying human cardiac physiology and pathophysiology. *Nat Protoc* 2009; **4**: 232-243 [PMID: 19197267 DOI: 10.1038/nprot.2008.229]
- 53 **Saga Y**, Kitajima S, Miyagawa-Tomita S. Mesp1 expression is the earliest sign of cardiovascular development. *Trends Cardiovasc Med* 2000; **10**: 345-352 [PMID: 11369261 DOI: 10.1016/s1050-1738(01)00069-x]
- 54 **Bondue A**, Lapouge G, Paulissen C, Semeraro C, Iacovino M, Kyba M, Blanpain C. Mesp1 acts as a master regulator of multipotent cardiovascular progenitor specification. *Cell Stem Cell* 2008; **3**: 69-84 [PMID: 18593560 DOI: 10.1016/j.stem.2008.06.009]
- 55 **Rockey DC**, Bell PD, Hill JA. Fibrosis--a common pathway to organ injury and failure. *N Engl J Med* 2015; **372**: 1138-1149 [PMID: 25785971 DOI: 10.1056/NEJMr1300575]
- 56 **Yamakawa H**, Ieda M. Cardiac regeneration by direct reprogramming in this decade and beyond. *Inflamm Regen* 2021; **41**: 20 [PMID: 34193320 DOI: 10.1186/s41232-021-00168-5]
- 57 **Fu JD**, Stone NR, Liu L, Spencer CI, Qian L, Hayashi Y, Delgado-Olguin P, Ding S, Bruneau BG, Srivastava D. Direct reprogramming of human fibroblasts toward a cardiomyocyte-like state. *Stem Cell Reports* 2013; **1**: 235-247 [PMID: 24319660 DOI: 10.1016/j.stemcr.2013.07.005]
- 58 **Yang L**, Soonpaa MH, Adler ED, Roepke TK, Kattman SJ, Kennedy M, Henckaerts E, Bonham K, Abbott GW, Linden RM, Field LJ, Keller GM. Human cardiovascular progenitor cells develop from a KDR+ embryonic-stem-cell-derived population. *Nature* 2008; **453**: 524-528 [PMID: 18432194 DOI: 10.1038/nature06894]
- 59 **Nelson TJ**, Chiriac A, Faustino RS, Crespo-Diaz RJ, Behfar A, Terzic A. Lineage specification of Flk-1+ progenitors is associated with divergent Sox7 expression in cardiopoiesis. *Differentiation* 2009; **77**: 248-255 [PMID: 19272523 DOI: 10.1016/j.diff.2008.10.012]
- 60 **Galdos FX**, Guo Y, Paige SL, VanDusen NJ, Wu SM, Pu WT. Cardiac Regeneration: Lessons From Development. *Circ Res* 2017; **120**: 941-959 [PMID: 28302741 DOI: 10.1161/CIRCRESAHA.116.309040]
- 61 **Kelly RG**, Buckingham ME, Moorman AF. Heart fields and cardiac morphogenesis. *Cold Spring Harb Perspect Med* 2014; **4** [PMID: 25274757 DOI: 10.1101/cshperspect.a015750]

- 62 **Zhou Z**, Wang J, Guo C, Chang W, Zhuang J, Zhu P, Li X. Temporally Distinct Six2-Positive Second Heart Field Progenitors Regulate Mammalian Heart Development and Disease. *Cell Rep* 2017; **18**: 1019-1032 [PMID: 28122228 DOI: 10.1016/j.celrep.2017.01.002]
- 63 **Rana MS**, Christoffels VM, Moorman AF. A molecular and genetic outline of cardiac morphogenesis. *Acta Physiol (Oxf)* 2013; **207**: 588-615 [PMID: 23297764 DOI: 10.1111/apha.12061]
- 64 **Xin M**, Olson EN, Bassel-Duby R. Mending broken hearts: cardiac development as a basis for adult heart regeneration and repair. *Nat Rev Mol Cell Biol* 2013; **14**: 529-541 [PMID: 23839576 DOI: 10.1038/nrm3619]
- 65 **Li Y**, Tian S, Lei I, Liu L, Ma P, Wang Z. Transplantation of multipotent Isl1+ cardiac progenitor cells preserves infarcted heart function in mice. *Am J Transl Res* 2017; **9**: 1530-1542 [PMID: 28386378]
- 66 **Zhou B**, von Gise A, Ma Q, Rivera-Feliciano J, Pu WT. Nkx2-5- and Isl1-expressing cardiac progenitors contribute to proepicardium. *Biochem Biophys Res Commun* 2008; **375**: 450-453 [PMID: 18722343 DOI: 10.1016/j.bbrc.2008.08.044]
- 67 **Tampakakis E**, Miyamoto M, Kwon C. In Vitro Generation of Heart Field-specific Cardiac Progenitor Cells. *J Vis Exp* 2019 [PMID: 31329174 DOI: 10.3791/59826]
- 68 **Streef TJ**, Smits AM. Epicardial Contribution to the Developing and Injured Heart: Exploring the Cellular Composition of the Epicardium. *Front Cardiovasc Med* 2021; **8**: 750243 [PMID: 34631842 DOI: 10.3389/fcvm.2021.750243]
- 69 **Synnergren J**, Drowley L, Plowright AT, Brolén G, Goumans MJ, Gittenberger-de Groot AC, Sartipy P, Wang QD. Comparative transcriptomic analysis identifies genes differentially expressed in human epicardial progenitors and hiPSC-derived cardiac progenitors. *Physiol Genomics* 2016; **48**: 771-784 [PMID: 27591124 DOI: 10.1152/physiolgenomics.00064.2016]
- 70 **Zhou B**, Honor LB, He H, Ma Q, Oh JH, Butterfield C, Lin RZ, Melero-Martin JM, Dolmatova E, Duffy HS, Gise Av, Zhou P, Hu YW, Wang G, Zhang B, Wang L, Hall JL, Moses MA, McGowan FX, Pu WT. Adult mouse epicardium modulates myocardial injury by secreting paracrine factors. *J Clin Invest* 2011; **121**: 1894-1904 [PMID: 21505261 DOI: 10.1172/JCI45529]
- 71 **Zangi L**, Lui KO, von Gise A, Ma Q, Ebina W, Ptaszek LM, Später D, Xu H, Tabebordbar M, Gorbatov R, Sena B, Nahrendorf M, Briscoe DM, Li RA, Wagers AJ, Rossi DJ, Pu WT, Chien KR. Modified mRNA directs the fate of heart progenitor cells and induces vascular regeneration after myocardial infarction. *Nat Biotechnol* 2013; **31**: 898-907 [PMID: 24013197 DOI: 10.1038/nbt.2682]
- 72 **Sandstedt J**, Jonsson M, Kajic K, Sandstedt M, Lindahl A, Dellgren G, Jeppsson A, Asp J. Left atrium of the human adult heart contains a population of side population cells. *Basic Res Cardiol* 2012; **107**: 255 [PMID: 22361742 DOI: 10.1007/s00395-012-0255-7]
- 73 **Hierlihy AM**, Seale P, Lobe CG, Rudnicki MA, Megeney LA. The post-natal heart contains a myocardial stem cell population. *FEBS Lett* 2002; **530**: 239-243 [PMID: 12387899 DOI: 10.1016/s0014-5793(02)03477-4]
- 74 **Oikonomopoulos A**, Sereti KI, Conyers F, Bauer M, Liao A, Guan J, Crapps D, Han JK, Dong H, Bayomy AF, Fine GC, Westerman K, Biechele TL, Moon RT, Force T, Liao R. Wnt signaling exerts an antiproliferative effect on adult cardiac progenitor cells through IGFBP3. *Circ Res* 2011; **109**: 1363-1374 [PMID: 22034491 DOI: 10.1161/CIRCRESAHA.111.250282]
- 75 **Barandon L**, Casassus F, Leroux L, Moreau C, Allières C, Lamazière JM, Dufourcq P, Couffignal T, Duplâa C. Secreted frizzled-related protein-1 improves postinfarction scar formation through a modulation of inflammatory response. *Arterioscler Thromb Vasc Biol* 2011; **31**: e80-e87 [PMID: 21836067 DOI: 10.1161/ATVBAHA.111.232280]
- 76 **Fu WB**, Wang WE, Zeng CY. Wnt signaling pathways in myocardial infarction and the therapeutic effects of Wnt pathway inhibitors. *Acta Pharmacol Sin* 2019; **40**: 9-12 [PMID: 30002488 DOI: 10.1038/s41401-018-0060-4]
- 77 **Zhou S**, Schuetz JD, Bunting KD, Colapietro AM, Sampath J, Morris JJ, Lagutina I, Grosveld GC, Osawa M, Nakauchi H, Sorrentino BP. The ABC transporter Bcrp1/ABCG2 is expressed in a wide variety of stem cells and is a molecular determinant of the side-population phenotype. *Nat Med* 2001; **7**: 1028-1034 [PMID: 11533706 DOI: 10.1038/nm0901-1028]
- 78 **Higashikuni Y**, Sainz J, Nakamura K, Takaoka M, Enomoto S, Iwata H, Sahara M, Tanaka K, Koibuchi N, Ito S, Kusuha H, Sugiyama Y, Hirata Y, Nagai R, Sata M. The ATP-binding cassette transporter BCRP1/ABCG2 plays a pivotal role in cardiac repair after myocardial infarction via modulation of microvascular endothelial cell survival and function. *Arterioscler Thromb Vasc Biol* 2010; **30**: 2128-2135 [PMID: 20829509 DOI: 10.1161/ATVBAHA.110.211755]
- 79 **Oyama T**, Nagai T, Wada H, Naito AT, Matsuura K, Iwanaga K, Takahashi T, Goto M, Mikami Y, Yasuda N, Akazawa H, Uezumi A, Takeda S, Komuro I. Cardiac side population cells have a potential to migrate and differentiate into cardiomyocytes in vitro and in vivo. *J Cell Biol* 2007; **176**: 329-341 [PMID: 17261849 DOI: 10.1083/jcb.200603014]
- 80 **Maher TJ**, Ren Y, Li Q, Braunlin E, Garry MG, Sorrentino BP, Martin CM. ATP-binding cassette transporter Abcg2 lineage contributes to the cardiac vasculature after oxidative stress. *Am J Physiol Heart Circ Physiol* 2014; **306**: H1610-H1618 [PMID: 24727496 DOI: 10.1152/ajpheart.00638.2013]
- 81 **Doyle MJ**, Maher TJ, Li Q, Garry MG, Sorrentino BP, Martin CM. Abcg2-Labeled Cells Contribute to Different Cell Populations in the Embryonic and Adult Heart. *Stem Cells Dev* 2016; **25**: 277-284 [PMID: 26573225 DOI: 10.1089/scd.2015.0272]
- 82 **Yellamilli A**, Ren Y, McElmurry RT, Lambert JP, Gross P, Mohsin S, Houser SR, Elrod JW, Tolar

- J, Garry DJ, van Berlo JH. Abcg2-expressing side population cells contribute to cardiomyocyte renewal through fusion. *FASEB J* 2020; **34**: 5642-5657 [PMID: 32100368 DOI: 10.1096/fj.201902105R]
- 83 **Yellamilli A**, van Berlo JH. The Role of Cardiac Side Population Cells in Cardiac Regeneration. *Front Cell Dev Biol* 2016; **4**: 102 [PMID: 27679798 DOI: 10.3389/fcell.2016.00102]
- 84 **Cho HJ**, Lee HJ, Chung YJ, Kim JY, Cho HJ, Yang HM, Kwon YW, Lee HY, Oh BH, Park YB, Kim HS. Generation of human secondary cardiospheres as a potent cell processing strategy for cell-based cardiac repair. *Biomaterials* 2013; **34**: 651-661 [PMID: 23103158 DOI: 10.1016/j.biomaterials.2012.10.011]
- 85 **Li TS**, Cheng K, Lee ST, Matsushita S, Davis D, Malliaras K, Zhang Y, Matsushita N, Smith RR, Marbán E. Cardiospheres recapitulate a niche-like microenvironment rich in stemness and cell-matrix interactions, rationalizing their enhanced functional potency for myocardial repair. *Stem Cells* 2010; **28**: 2088-2098 [PMID: 20882531 DOI: 10.1002/stem.532]
- 86 **Cheng K**, Malliaras K, Smith RR, Shen D, Sun B, Blusztajn A, Xie Y, Ibrahim A, Aminzadeh MA, Liu W, Li TS, De Robertis MA, Marbán L, Czer LSC, Trento A, Marbán E. Human cardiosphere-derived cells from advanced heart failure patients exhibit augmented functional potency in myocardial repair. *JACC Heart Fail* 2014; **2**: 49-61 [PMID: 24511463 DOI: 10.1016/j.jchf.2013.08.008]
- 87 **Vincentz JW**, Barnes RM, Firulli BA, Conway SJ, Firulli AB. Cooperative interaction of Nkx2.5 and Mef2c transcription factors during heart development. *Dev Dyn* 2008; **237**: 3809-3819 [PMID: 19035347 DOI: 10.1002/dvdy.21803]
- 88 **Carroll KJ**, Makarewich CA, McAnally J, Anderson DM, Zentilin L, Liu N, Giacca M, Bassel-Duby R, Olson EN. A mouse model for adult cardiac-specific gene deletion with CRISPR/Cas9. *Proc Natl Acad Sci U S A* 2016; **113**: 338-343 [PMID: 26719419 DOI: 10.1073/pnas.1523918113]
- 89 **Sano T**, Ito T, Ishigami S, Bandaru S, Sano S. Intrinsic activation of cardiosphere-derived cells enhances myocardial repair. *J Thorac Cardiovasc Surg* 2020 [PMID: 32682583 DOI: 10.1016/j.jtcvs.2020.05.040]
- 90 **Hare JM**, Fishman JE, Gerstenblith G, DiFede Velazquez DL, Zambrano JP, Suncion VY, Tracy M, Ghersin E, Johnston PV, Brinker JA, Breton E, Davis-Sproul J, Schulman IH, Byrnes J, Mendizabal AM, Lowery MH, Rouy D, Altman P, Wong Po Foo C, Ruiz P, Amador A, Da Silva J, McNiece IK, Heldman AW, George R, Lardo A. Comparison of allogeneic vs autologous bone marrow-derived mesenchymal stem cells delivered by transendocardial injection in patients with ischemic cardiomyopathy: the POSEIDON randomized trial. *JAMA* 2012; **308**: 2369-2379 [PMID: 23117550 DOI: 10.1001/jama.2012.25321]
- 91 **Sukhacheva TV**, Nizyaeva NV, Samsonova MV, Chernyaev AL, Shchegolev AI, Serov RA. Telocytes in the Myocardium of Children with Congenital Heart Disease Tetralogy of Fallot. *Bull Exp Biol Med* 2020; **169**: 137-146 [PMID: 32488790 DOI: 10.1007/s10517-020-04840-7]
- 92 **Tan JJ**, Carr CA, Stuckey DJ, Ellison GM, Messina E, Giacomello A, Clarke K. Isolation and expansion of cardiosphere-derived stem cells. *Curr Protoc Stem Cell Biol* 2011; **16**: 2C-3.1 [DOI: 10.1002/9780470151808.sc02c03s16]
- 93 **Chen L**, Pan Y, Zhang L, Wang Y, Weintraub N, Tang Y. Two-step protocol for isolation and culture of cardiospheres. *Methods Mol Biol* 2013; **1036**: 75-80 [PMID: 23807787 DOI: 10.1007/978-1-62703-511-8_6]
- 94 **Barker N**. Adult intestinal stem cells: critical drivers of epithelial homeostasis and regeneration. *Nat Rev Mol Cell Biol* 2014; **15**: 19-33 [PMID: 24326621 DOI: 10.1038/nrm3721]
- 95 **Bianco P**. Bone and the hematopoietic niche: a tale of two stem cells. *Blood* 2011; **117**: 5281-5288 [PMID: 21406722 DOI: 10.1182/blood-2011-01-315069]
- 96 **Huson DH**, Bryant D. Application of phylogenetic networks in evolutionary studies. *Mol Biol Evol* 2006; **23**: 254-267 [PMID: 16221896 DOI: 10.1093/molbev/msj030]
- 97 **Urbanek K**, Cesselli D, Rota M, Nascimbene A, De Angelis A, Hosoda T, Bearzi C, Boni A, Bolli R, Kajstura J, Anversa P, Leri A. Stem cell niches in the adult mouse heart. *Proc Natl Acad Sci USA* 2006; **103**: 9226-9231 [PMID: 16754876 DOI: 10.1073/pnas.0600635103]
- 98 **Vukusic K**, Asp J, Henriksson HB, Brisby H, Lindahl A, Sandstedt J. Physical exercise affects slow cycling cells in the rat heart and reveals a new potential niche area in the atrioventricular junction. *J Mol Histol* 2015; **46**: 387-398 [PMID: 26047663 DOI: 10.1007/s10735-015-9626-3]
- 99 **Bearzi C**, Rota M, Hosoda T, Tillmanns J, Nascimbene A, De Angelis A, Yasuzawa-Amano S, Trofimova I, Siggins RW, Lecapitaine N, Cascapera S, Beltrami AP, D'alessandro DA, Zias E, Quaini F, Urbanek K, Michler RE, Bolli R, Kajstura J, Leri A, Anversa P. Human cardiac stem cells. *Proc Natl Acad Sci USA* 2007; **104**: 14068-14073 [PMID: 17709737 DOI: 10.1073/pnas.0706760104]
- 100 **Leri A**, Rota M, Hosoda T, Goichberg P, Anversa P. Cardiac stem cell niches. *Stem Cell Res* 2014; **13**: 631-646 [PMID: 25267073 DOI: 10.1016/j.scr.2014.09.001]
- 101 **Vicinanza C**, Aquila I, Scalise M, Cristiano F, Marino F, Cianflone E, Mancuso T, Marotta P, Sacco W, Lewis FC, Couch L, Shone V, Gritti G, Torella A, Smith AJ, Terracciano CM, Britti D, Veltri P, Indolfi C, Nadal-Ginard B, Ellison-Hughes GM, Torella D. Adult cardiac stem cells are multipotent and robustly myogenic: c-kit expression is necessary but not sufficient for their identification. *Cell Death Differ* 2017; **24**: 2101-2116 [PMID: 28800128 DOI: 10.1038/cdd.2017.130]
- 102 **Leri A**, Kajstura J, Anversa P. Cardiac Stem Cells and Mechanisms of Myocardial Regeneration. *Physiol Rev* 2005; **85**: 1373-1416 [PMID: 16183916 DOI: 10.1152/physrev.00013.2005]

- 103 **Scalise M**, Marino F, Salerno L, Mancuso T, Cappetta D, Barone A, Parrotta EI, Torella A, Palumbo D, Veltri P, De Angelis A, Berrino L, Rossi F, Weisz A, Rota M, Urbanek K, Nadal-Ginard B, Torella D, Cianflone E. In vitro CSC-derived cardiomyocytes exhibit the typical microRNA-mRNA blueprint of endogenous cardiomyocytes. *Commun Biol* 2021; **4**: 1146 [PMID: [34593953](#) DOI: [10.1038/s42003-021-02677-y](#)]
- 104 **Scadden DT**. Nice neighborhood: emerging concepts of the stem cell niche. *Cell* 2014; **157**: 41-50 [PMID: [24679525](#) DOI: [10.1016/j.cell.2014.02.013](#)]
- 105 **Donnelly H**, Salmeron-Sanchez M, Dalby MJ. Designing stem cell niches for differentiation and self-renewal. *J R Soc Interface* 2018; **15** [PMID: [30158185](#) DOI: [10.1098/rsif.2018.0388](#)]
- 106 **Simsek T**, Kocabas F, Zheng J, Deberardinis RJ, Mahmoud AI, Olson EN, Schneider JW, Zhang CC, Sadek HA. The distinct metabolic profile of hematopoietic stem cells reflects their location in a hypoxic niche. *Cell Stem Cell* 2010; **7**: 380-390 [PMID: [20804973](#) DOI: [10.1016/j.stem.2010.07.011](#)]
- 107 **Kimura W**, Sadek HA. The cardiac hypoxic niche: emerging role of hypoxic microenvironment in cardiac progenitors. *Cardiovasc Diagn Ther* 2012; **2**: 278-289 [PMID: [24282728](#) DOI: [10.3978/j.issn.2223-3652.2012.12.02](#)]
- 108 **Mas-Bargues C**, Sanz-Ros J, Román-Domínguez A, Inglés M, Gimeno-Mallench L, El Alami M, Viña-Almunia J, Gambini J, Viña J, Borrás C. Relevance of Oxygen Concentration in Stem Cell Culture for Regenerative Medicine. *Int J Mol Sci* 2019; **20**: 1195 [PMID: [30857245](#) DOI: [10.3390/ijms20051195](#)]
- 109 **Kocabas F**, Mahmoud AI, Susic D, Porrello ER, Chen R, Garcia JA, DeBerardinis RJ, Sadek HA. The hypoxic epicardial and subepicardial microenvironment. *J Cardiovasc Transl Res* 2012; **5**: 654-665 [PMID: [22566269](#) DOI: [10.1007/s12265-012-9366-7](#)]
- 110 **Latham N**, Ye B, Jackson R, Lam BK, Kuraitis D, Ruel M, Suuronen EJ, Stewart DJ, Davis DR. Human blood and cardiac stem cells synergize to enhance cardiac repair when cotransplanted into ischemic myocardium. *Circulation* 2013; **128**: S105-S112 [PMID: [24030393](#) DOI: [10.1161/CIRCULATIONAHA.112.000374](#)]
- 111 **Teng YC**, Porfirio-Sousa AL, Ribeiro GM, Arend MC, da Silva Meirelles L, Chen ES, Rosa DS, Han SW. Analyses of the pericyte transcriptome in ischemic skeletal muscles. *Stem Cell Res Ther* 2021; **12**: 183 [PMID: [33726849](#) DOI: [10.1186/s13287-021-02247-3](#)]
- 112 **Silvestre JS**, Smadja DM, Lévy BI. Postischemic revascularization: from cellular and molecular mechanisms to clinical applications. *Physiol Rev* 2013; **93**: 1743-1802 [PMID: [24137021](#) DOI: [10.1152/physrev.00006.2013](#)]
- 113 **Avolio E**, Meloni M, Spencer HL, Riu F, Katare R, Mangialardi G, Oikawa A, Rodriguez-Arabaolaza I, Dang Z, Mitchell K, Reni C, Alvino VV, Rowlinson J, Livi U, Cesselli D, Angelini G, Emanueli C, Beltrami AP, Madeddu P. Combined intramyocardial delivery of human pericytes and cardiac stem cells additively improves the healing of mouse infarcted hearts through stimulation of vascular and muscular repair. *Circ Res* 2015; **116**: e81-e94 [PMID: [25801898](#) DOI: [10.1161/CIRCRESAHA.115.306146](#)]
- 114 **Li YY**, Zhang S, Li YG, Wang Y. Isolation, culture, purification and ultrastructural investigation of cardiac telocytes. *Mol Med Rep* 2016; **14**: 1194-1200 [PMID: [27314536](#) DOI: [10.3892/mmr.2016.5386](#)]
- 115 **Li H**, Zhang Y, Zhang W, Zhong X, Zhao G, Ge J. Paracrine Endocrinology of cardiac telocytes in culture. *Cardiol Plus* 2017; **2**: 8 [DOI: [10.4103/2470-7511.248218](#)]
- 116 **Sukhacheva TV**, Nizyaeva NV, Samsonova MV, Cherniaev AL, Burov AA, Iurova MV, Shchegolev AI, Serov RA, Sukhikh GT. Morpho-functional changes of cardiac telocytes in isolated atrial amyloidosis in patients with atrial fibrillation. *Sci Rep* 2021; **11**: 3563 [PMID: [33574429](#) DOI: [10.1038/s41598-021-82554-0](#)]
- 117 **Albulescu R**, Tanase C, Codrici E, Popescu DI, Cretoiu SM, Popescu LM. The secretome of myocardial telocytes modulates the activity of cardiac stem cells. *J Cell Mol Med* 2015; **19**: 1783-1794 [PMID: [26176909](#) DOI: [10.1111/jcmm.12624](#)]
- 118 **Mohri T**, Fujio Y, Obana M, Iwakura T, Matsuda K, Maeda M, Azuma J. Signals through glycoprotein 130 regulate the endothelial differentiation of cardiac stem cells. *Arterioscler Thromb Vasc Biol* 2009; **29**: 754-760 [PMID: [19213943](#) DOI: [10.1161/ATVBAHA.108.174870](#)]
- 119 **Laguens R**, Cabeza Meckert P, Vera Janavel G, Del Valle H, Lascano E, Negroni J, Werba P, Cuniberti L, Martínez V, Melo C, Papouchado M, Ojeda R, Criscuolo M, Crottogini A. Entrance in mitosis of adult cardiomyocytes in ischemic pig hearts after plasmid-mediated rhVEGF165 gene transfer. *Gene Ther* 2002; **9**: 1676-1681 [PMID: [12457281](#) DOI: [10.1038/sj.gt.3301844](#)]
- 120 **Sid-Otmane C**, Perrault LP, Ly HQ. Mesenchymal stem cell mediates cardiac repair through autocrine, paracrine and endocrine axes. *J Transl Med* 2020; **18**: 336 [PMID: [32873307](#) DOI: [10.1186/s12967-020-02504-8](#)]
- 121 **Gatti S**, Leo C, Gallo S, Sala V, Bucci E, Natale M, Cantarella D, Medico E, Crepaldi T. Gene expression profiling of HGF/Met activation in neonatal mouse heart. *Transgenic Res* 2013; **22**: 579-593 [PMID: [23224784](#) DOI: [10.1007/s11248-012-9667-2](#)]
- 122 **Funatsu T**, Sawa Y, Ohtake S, Takahashi T, Matsumiya G, Matsuura N, Nakamura T, Matsuda H. Therapeutic angiogenesis in the ischemic canine heart induced by myocardial injection of naked complementary DNA plasmid encoding hepatocyte growth factor. *J Thorac Cardiovasc Surg* 2002; **124**: 1099-1105 [PMID: [12447174](#) DOI: [10.1067/mtc.2002.123809](#)]
- 123 **Zhang J**, Wang LL, Du W, Yu YC, Ju WZ, Man YL, Li XR, Chen Y, Wang ZD, Gu WJ, Zhang FX,

- Wang H, Wu CT, Cao KJ. Hepatocyte growth factor modification enhances the anti-arrhythmic properties of human bone marrow-derived mesenchymal stem cells. *PLoS One* 2014; **9**: e111246 [PMID: 25360679 DOI: 10.1371/journal.pone.0111246]
- 124 **Duan HF**, Wu CT, Wu DL, Lu Y, Liu HJ, Ha XQ, Zhang QW, Wang H, Jia XX, Wang LS. Treatment of myocardial ischemia with bone marrow-derived mesenchymal stem cells overexpressing hepatocyte growth factor. *Mol Ther* 2003; **8**: 467-474 [PMID: 12946320 DOI: 10.1016/s1525-0016(03)00186-2]
- 125 **Nakamura T**, Mizuno S, Matsumoto K, Sawa Y, Matsuda H, Nakamura T. Myocardial protection from ischemia/reperfusion injury by endogenous and exogenous HGF. *J Clin Invest* 2000; **106**: 1511-1519 [PMID: 11120758 DOI: 10.1172/jci10226]
- 126 **Shirakawa Y**, Sawa Y, Takewa Y, Tatsumi E, Kaneda Y, Taenaka Y, Matsuda H. Gene transfection with human hepatocyte growth factor complementary DNA plasmids attenuates cardiac remodeling after acute myocardial infarction in goat hearts implanted with ventricular assist devices. *J Thorac Cardiovasc Surg* 2005; **130**: 624-632 [PMID: 16153905 DOI: 10.1016/j.jtcvs.2004.02.045]
- 127 **Liao Z**, Chen Y, Duan C, Zhu K, Huang R, Zhao H, Hintze M, Pu Q, Yuan Z, Lv L, Chen H, Lai B, Feng S, Qi X, Cai D. Cardiac telocytes inhibit cardiac microvascular endothelial cell apoptosis through exosomal miRNA-21-5p-targeted *cdip1* silencing to improve angiogenesis following myocardial infarction. *Theranostics* 2021; **11**: 268-291 [PMID: 33391474 DOI: 10.7150/thno.47021]
- 128 **Gherghiceanu M**, Popescu LM. Heterocellular communication in the heart: electron tomography of telocyte-myocyte junctions. *J Cell Mol Med* 2011; **15**: 1005-1011 [PMID: 21426485 DOI: 10.1111/j.1582-4934.2011.01299.x]
- 129 **Ceausu Z**, Socea B, Dimitriu MCT, Predescu D, Constantin VD, Bacalbaşa N, Cirstoveanu C, Costache M, Ceausu M. Dormant cardiac stem cells: A promising tool in cardiac regeneration. *Exp Ther Med* 2020; **20**: 3452-3457 [PMID: 32905130 DOI: 10.3892/etm.2020.9015]
- 130 **Wang L**, Gu H, Turrentine M, Wang M. Estradiol treatment promotes cardiac stem cell (CSC)-derived growth factors, thus improving CSC-mediated cardioprotection after acute ischemia/reperfusion. *Surgery* 2014; **156**: 243-252 [PMID: 24957669 DOI: 10.1016/j.surg.2014.04.002]
- 131 **Sadat S**, Gehmert S, Song YH, Yen Y, Bai X, Gaiser S, Klein H, Alt E. The cardioprotective effect of mesenchymal stem cells is mediated by IGF-I and VEGF. *Biochem Biophys Res Commun* 2007; **363**: 674-679 [PMID: 17904522 DOI: 10.1016/j.bbrc.2007.09.058]
- 132 **Liu Q**, Piao H, Wang Y, Zheng D, Wang W. Circulating exosomes in cardiovascular disease: Novel carriers of biological information. *Biomed Pharmacother* 2021; **135**: 111148 [PMID: 33412387 DOI: 10.1016/j.biopha.2020.111148]
- 133 Gould SJ. Exosomes and Microvesicles. In: Encyclopedia of Biological Chemistry: Second Edition. Elsevier Inc. 2013: 262-264
- 134 **Hartl FU**. Molecular chaperones in cellular protein folding. *Nature* 1996; **381**: 571-579 [PMID: 8637592 DOI: 10.1038/381571a0]
- 135 **Boulanger CM**, Loyer X, Rautou PE, Amabile N. Extracellular vesicles in coronary artery disease. *Nat Rev Cardiol* 2017; **14**: 259-272 [PMID: 28150804 DOI: 10.1038/nrcardio.2017.7]
- 136 **Xiao J**, Pan Y, Li XH, Yang XY, Feng YL, Tan HH, Jiang L, Feng J, Yu XY. Cardiac progenitor cell-derived exosomes prevent cardiomyocytes apoptosis through exosomal miR-21 by targeting PDCD4. *Cell Death Dis* 2016; **7**: e2277 [PMID: 27336721 DOI: 10.1038/cddis.2016.181]
- 137 **Wu Q**, Wang J, Tan WLW, Jiang Y, Wang S, Li Q, Yu X, Tan J, Liu S, Zhang P, Tiang Z, Chen Z, Foo RS, Yang HT. Extracellular vesicles from human embryonic stem cell-derived cardiovascular progenitor cells promote cardiac infarct healing through reducing cardiomyocyte death and promoting angiogenesis. *Cell Death Dis* 2020; **11**: 354 [PMID: 32393784 DOI: 10.1038/s41419-020-2508-y]
- 138 **Oxvig C**. The role of PAPP-A in the IGF system: location, location, location. *J Cell Commun Signal* 2015; **9**: 177-187 [PMID: 25617049 DOI: 10.1007/s12079-015-0259-9]
- 139 **Andriolo G**, Provasi E, Lo Cicero V, Brambilla A, Soncin S, Torre T, Milano G, Biemmi V, Vassalli G, Turchetto L, Barile L, Radrizzani M. Exosomes From Human Cardiac Progenitor Cells for Therapeutic Applications: Development of a GMP-Grade Manufacturing Method. *Front Physiol* 2018; **9**: 1169 [PMID: 30197601 DOI: 10.3389/fphys.2018.01169]
- 140 **White AJ**, Smith RR, Matsushita S, Chakravarty T, Czer LS, Burton K, Schwarz ER, Davis DR, Wang Q, Reinsmoen NL, Forrester JS, Marbán E, Makkar R. Intrinsic cardiac origin of human cardiosphere-derived cells. *Eur Heart J* 2013; **34**: 68-75 [PMID: 21659438 DOI: 10.1093/eurheartj/ehr172]
- 141 **Davis DR**, Zhang Y, Smith RR, Cheng K, Terrovitis J, Malliaras K, Li TS, White A, Makkar R, Marbán E. Validation of the cardiosphere method to culture cardiac progenitor cells from myocardial tissue. *PLoS One* 2009; **4**: e7195 [PMID: 19779618 DOI: 10.1371/journal.pone.0007195]
- 142 **Marbán E**. The Secret Life of Exosomes: What Bees Can Teach Us About Next-Generation Therapeutics. *J Am Coll Cardiol* 2018; **71**: 193-200 [PMID: 29325643 DOI: 10.1016/j.jacc.2017.11.013]
- 143 **Yu H**, Lu K, Zhu J, Wang J. Stem cell therapy for ischemic heart diseases. *Br Med Bull* 2017; **121**: 135-154 [PMID: 28164211 DOI: 10.1093/bmb/ldw059]
- 144 **Zhang L**, Ding H, Zhang Y, Wang Y, Zhu W, Li P. Circulating MicroRNAs: Biogenesis and Clinical Significance in Acute Myocardial Infarction. *Front Physiol* 2020; **11**: 1088 [PMID: 33013463 DOI: 10.3389/fphys.2020.01088]

- 145 **Kenneweg F**, Bang C, Xiao K, Boulanger CM, Loyer X, Mazlan S, Schroen B, Hermans-Beijnsberger S, Foinquinos A, Hirt MN, Eschenhagen T, Funcke S, Stojanovic S, Genschel C, Schimmel K, Just A, Pfanne A, Scherf K, Dehmel S, Raemon-Buettner SM, Fiedler J, Thum T. Long Noncoding RNA-Enriched Vesicles Secreted by Hypoxic Cardiomyocytes Drive Cardiac Fibrosis. *Mol Ther Nucleic Acids* 2019; **18**: 363-374 [PMID: 31634682 DOI: 10.1016/j.omtn.2019.09.003]
- 146 **Rezaie J**, Rahbarghazi R, Pezeshki M, Mazhar M, Yekani F, Khaksar M, Shokrollahi E, Amini H, Hashemzadeh S, Sokullu SE, Tokac M. Cardioprotective role of extracellular vesicles: A highlight on exosome beneficial effects in cardiovascular diseases. *J Cell Physiol* 2019; **234**: 21732-21745 [PMID: 31140622 DOI: 10.1002/jcp.28894]
- 147 **Soda N**, Rehm BHA, Sonar P, Nguyen NT, Shiddiky MJA. Advanced liquid biopsy technologies for circulating biomarker detection. *J Mater Chem B* 2019; **7**: 6670-6704 [PMID: 31646316 DOI: 10.1039/c9tb01490j]
- 148 **Viola M**, de Jager SCA, Sluijter JPG. Targeting Inflammation after Myocardial Infarction: A Therapeutic Opportunity for Extracellular Vesicles? *Int J Mol Sci* 2021; **22**: 7831 [PMID: 34360595 DOI: 10.3390/ijms22157831]
- 149 **Cambier L**, de Couto G, Ibrahim A, Echavez AK, Valle J, Liu W, Kreke M, Smith RR, Marbán L, Marbán E. Y RNA fragment in extracellular vesicles confers cardioprotection *via* modulation of IL-10 expression and secretion. *EMBO Mol Med* 2017; **9**: 337-352 [PMID: 28167565 DOI: 10.15252/emmm.201606924]
- 150 **Tseliou E**, Fouad J, Reich H, Slipczuk L, de Couto G, Aminzadeh M, Middleton R, Valle J, Weixin L, Marbán E. Fibroblasts Rendered Antifibrotic, Antiapoptotic, and Angiogenic by Priming With Cardiosphere-Derived Extracellular Membrane Vesicles. *J Am Coll Cardiol* 2015; **66**: 599-611 [PMID: 26248985 DOI: 10.1016/j.jacc.2015.05.068]
- 151 **Balbi C**, Costa A, Barile L, Bollini S. Message in a Bottle: Upgrading Cardiac Repair into Rejuvenation. *Cells* 2020; **9** [PMID: 32183455 DOI: 10.3390/cells9030724]
- 152 **Wu R**, Hu X, Wang J. Concise Review: Optimized Strategies for Stem Cell-Based Therapy in Myocardial Repair: Clinical Translatability and Potential Limitation. *Stem Cells* 2018; **36**: 482-500 [PMID: 29330880 DOI: 10.1002/stem.2778]
- 153 **Barile L**, Milano G, Vassalli G. Beneficial effects of exosomes secreted by cardiac-derived progenitor cells and other cell types in myocardial ischemia. *Stem Cell Investig* 2017; **4**: 93 [PMID: 29270419 DOI: 10.21037/sci.2017.11.06]
- 154 **Rogers RG**, Ciullo A, Marbán E, Ibrahim AG. Extracellular Vesicles as Therapeutic Agents for Cardiac Fibrosis. *Front Physiol* 2020; **11**: 479 [PMID: 32528309 DOI: 10.3389/fphys.2020.00479]
- 155 **Tikhomirov R**, Donnell BR, Catapano F, Faggian G, Gorelik J, Martelli F, Emanuelli C. Exosomes: From Potential Culprits to New Therapeutic Promise in the Setting of Cardiac Fibrosis. *Cells* 2020; **9** [PMID: 32131460 DOI: 10.3390/cells9030592]
- 156 **Yuan Y**, Du W, Liu J, Ma W, Zhang L, Du Z, Cai B. Stem Cell-Derived Exosome in Cardiovascular Diseases: Macro Roles of Micro Particles. *Front Pharmacol* 2018; **9**: 547 [PMID: 29904347 DOI: 10.3389/fphar.2018.00547]
- 157 **Prathipati P**, Nandi SS, Mishra PK. Stem Cell-Derived Exosomes, Autophagy, Extracellular Matrix Turnover, and miRNAs in Cardiac Regeneration during Stem Cell Therapy. *Stem Cell Rev Rep* 2017; **13**: 79-91 [PMID: 27807762 DOI: 10.1007/s12015-016-9696-y]
- 158 **Chimenti I**, Forte E, Angelini F, Giacomello A, Messina E. From ontogenesis to regeneration: learning how to instruct adult cardiac progenitor cells. *Prog Mol Biol Transl Sci* 2012; **111**: 109-137 [PMID: 22917228 DOI: 10.1016/B978-0-12-398459-3.00005-8]
- 159 **Ciullo A**, Biemmi V, Milano G, Bolis S, Cervio E, Fertig ET, Gherghiceanu M, Moccetti T, Camici GG, Vassalli G, Barile L. Exosomal Expression of CXCR4 Targets Cardioprotective Vesicles to Myocardial Infarction and Improves Outcome after Systemic Administration. *Int J Mol Sci* 2019; **20** [PMID: 30678240 DOI: 10.3390/ijms20030468]
- 160 **Gallet R**, Dawkins J, Valle J, Simsolo E, de Couto G, Middleton R, Tseliou E, Luthringer D, Kreke M, Smith RR, Marbán L, Ghaleh B, Marbán E. Exosomes secreted by cardiosphere-derived cells reduce scarring, attenuate adverse remodelling, and improve function in acute and chronic porcine myocardial infarction. *Eur Heart J* 2017; **38**: 201-211 [PMID: 28158410 DOI: 10.1093/eurheartj/ehw240]
- 161 **Makkar RR**, Smith RR, Cheng K, Malliaras K, Thomson LE, Berman D, Czer LS, Marbán L, Mendizabal A, Johnston PV, Russell SD, Schuleri KH, Lardo AC, Gerstenblith G, Marbán E. Intracoronary cardiosphere-derived cells for heart regeneration after myocardial infarction (CADUCEUS): a prospective, randomised phase 1 trial. *Lancet* 2012; **379**: 895-904 [PMID: 22336189 DOI: 10.1016/S0140-6736(12)60195-0]
- 162 **Ghafouri-Fard S**, Niazi V, Hussen BM, Omrani MD, Taheri M, Basiri A. The Emerging Role of Exosomes in the Treatment of Human Disorders With a Special Focus on Mesenchymal Stem Cells-Derived Exosomes. *Front Cell Dev Biol* 2021; **9**: 653296 [PMID: 34307345 DOI: 10.3389/fcell.2021.653296]
- 163 **Milano G**, Biemmi V, Lazzarini E, Balbi C, Ciullo A, Bolis S, Ameri P, Di Silvestre D, Mauri P, Barile L, Vassalli G. Intravenous administration of cardiac progenitor cell-derived exosomes protects against doxorubicin/trastuzumab-induced cardiac toxicity. *Cardiovasc Res* 2020; **116**: 383-392 [PMID: 31098627 DOI: 10.1093/cvr/cvz108]
- 164 **Lopaschuk GD**, Jaswal JS. Energy metabolic phenotype of the cardiomyocyte during development, differentiation, and postnatal maturation. *J Cardiovasc Pharmacol* 2010; **56**: 130-140 [PMID:

20505524 DOI: [10.1097/FJC.0b013e3181e74a14](https://doi.org/10.1097/FJC.0b013e3181e74a14)]

- 165 **Breckenridge RA**, Piotrowska I, Ng KE, Ragan TJ, West JA, Kotecha S, Towers N, Bennett M, Kiensberger PC, Smolenski RT, Siddall HK, Offer JL, Mocanu MM, Yelon DM, Dyck JR, Griffin JL, Abramov AY, Gould AP, Mohun TJ. Hypoxic regulation of hand1 controls the fetal-neonatal switch in cardiac metabolism. *PLoS Biol* 2013; **11**: e1001666 [PMID: [24086110](https://pubmed.ncbi.nlm.nih.gov/24086110/) DOI: [10.1371/journal.pbio.1001666](https://doi.org/10.1371/journal.pbio.1001666)]
- 166 **Puente BN**, Kimura W, Muralidhar SA, Moon J, Amatruda JF, Phelps KL, Grinsfelder D, Rothermel BA, Chen R, Garcia JA, Santos CX, Thet S, Mori E, Kinter MT, Rindler PM, Zacchigna S, Mukherjee S, Chen DJ, Mahmoud AI, Giacca M, Rabinovitch PS, Aroumougame A, Shah AM, Szweda LI, Sadek HA. The oxygen-rich postnatal environment induces cardiomyocyte cell-cycle arrest through DNA damage response. *Cell* 2014; **157**: 565-579 [PMID: [24766806](https://pubmed.ncbi.nlm.nih.gov/24766806/) DOI: [10.1016/j.cell.2014.03.032](https://doi.org/10.1016/j.cell.2014.03.032)]
- 167 **Malandraki-Miller S**, Lopez CA, Al-Siddiqi H, Carr CA. Changing Metabolism in Differentiating Cardiac Progenitor Cells-Can Stem Cells Become Metabolically Flexible Cardiomyocytes? *Front Cardiovasc Med* 2018; **5**: 119 [PMID: [30283788](https://pubmed.ncbi.nlm.nih.gov/30283788/) DOI: [10.3389/fcvm.2018.00119](https://doi.org/10.3389/fcvm.2018.00119)]
- 168 **Lai L**, Leone TC, Zechner C, Schaeffer PJ, Kelly SM, Flanagan DP, Medeiros DM, Kovacs A, Kelly DP. Transcriptional coactivators PGC-1alpha and PGC-1beta control overlapping programs required for perinatal maturation of the heart. *Genes Dev* 2008; **22**: 1948-1961 [PMID: [18628400](https://pubmed.ncbi.nlm.nih.gov/18628400/) DOI: [10.1101/gad.1661708](https://doi.org/10.1101/gad.1661708)]
- 169 **Karwi QG**, Uddin GM, Ho KL, Lopaschuk GD. Loss of Metabolic Flexibility in the Failing Heart. *Front Cardiovasc Med* 2018; **5**: 68 [PMID: [29928647](https://pubmed.ncbi.nlm.nih.gov/29928647/) DOI: [10.3389/fcvm.2018.00068](https://doi.org/10.3389/fcvm.2018.00068)]
- 170 **Menendez-Montes I**, Escobar B, Palacios B, Gómez MJ, Izquierdo-García JL, Flores L, Jiménez-Borreguero LJ, Aragonés J, Ruiz-Cabello J, Torres M, Martín-Puig S. Myocardial VHL-HIF Signaling Controls an Embryonic Metabolic Switch Essential for Cardiac Maturation. *Dev Cell* 2016; **39**: 724-739 [PMID: [27997827](https://pubmed.ncbi.nlm.nih.gov/27997827/) DOI: [10.1016/j.devcel.2016.11.012](https://doi.org/10.1016/j.devcel.2016.11.012)]
- 171 **Steinmetz M**, Quentin T, Poppe A, Paul T, Jux C. Changes in expression levels of genes involved in fatty acid metabolism: upregulation of all three members of the PPAR family (alpha, gamma, delta) and the newly described adiponectin receptor 2, but not adiponectin receptor 1 during neonatal cardiac development of the rat. *Basic Res Cardiol* 2005; **100**: 263-269 [PMID: [15754086](https://pubmed.ncbi.nlm.nih.gov/15754086/) DOI: [10.1007/s00395-005-0520-0](https://doi.org/10.1007/s00395-005-0520-0)]
- 172 **Lehman JJ**, Barger PM, Kovacs A, Saffitz JE, Medeiros DM, Kelly DP. Peroxisome proliferator-activated receptor gamma coactivator-1 promotes cardiac mitochondrial biogenesis. *J Clin Invest* 2000; **106**: 847-856 [PMID: [11018072](https://pubmed.ncbi.nlm.nih.gov/11018072/) DOI: [10.1172/JCI10268](https://doi.org/10.1172/JCI10268)]
- 173 **Chung S**, Dzeja PP, Faustino RS, Perez-Terzic C, Behfar A, Terzic A. Mitochondrial oxidative metabolism is required for the cardiac differentiation of stem cells. *Nat Clin Pract Cardiovasc Med* 2007; **4** Suppl 1: S60-S67 [PMID: [17230217](https://pubmed.ncbi.nlm.nih.gov/17230217/) DOI: [10.1038/ncpcardio0766](https://doi.org/10.1038/ncpcardio0766)]
- 174 **Dai DF**, Danoviz ME, Wiczler B, Laflamme MA, Tian R. Mitochondrial Maturation in Human Pluripotent Stem Cell Derived Cardiomyocytes. *Stem Cells Int* 2017; **2017**: 5153625 [PMID: [28421116](https://pubmed.ncbi.nlm.nih.gov/28421116/) DOI: [10.1155/2017/5153625](https://doi.org/10.1155/2017/5153625)]
- 175 **Birket MJ**, Casini S, Kosmidis G, Elliott DA, Gerencser AA, Baartscheer A, Schumacher C, Mastroberardino PG, Elefanty AG, Stanley EG, Mummery CL. PGC-1 α and reactive oxygen species regulate human embryonic stem cell-derived cardiomyocyte function. *Stem Cell Reports* 2013; **1**: 560-574 [PMID: [24371810](https://pubmed.ncbi.nlm.nih.gov/24371810/) DOI: [10.1016/j.stemcr.2013.11.008](https://doi.org/10.1016/j.stemcr.2013.11.008)]
- 176 **Malandraki-Miller S**, Lopez CA, Alonaizan R, Purnama U, Perbellini F, Pakzad K, Carr CA. Metabolic flux analyses to assess the differentiation of adult cardiac progenitors after fatty acid supplementation. *Stem Cell Res* 2019; **38**: 101458 [PMID: [31102832](https://pubmed.ncbi.nlm.nih.gov/31102832/) DOI: [10.1016/j.scr.2019.101458](https://doi.org/10.1016/j.scr.2019.101458)]
- 177 **Correia C**, Koshkin A, Duarte P, Hu D, Teixeira A, Domian I, Serra M, Alves PM. Distinct carbon sources affect structural and functional maturation of cardiomyocytes derived from human pluripotent stem cells. *Sci Rep* 2017; **7**: 8590 [PMID: [28819274](https://pubmed.ncbi.nlm.nih.gov/28819274/) DOI: [10.1038/s41598-017-08713-4](https://doi.org/10.1038/s41598-017-08713-4)]
- 178 **Pakzad KK**, Tan JJ, Anderson S, Board M, Clarke K, Carr CA. Metabolic maturation of differentiating cardiosphere-derived cells. *Stem Cell Res* 2021; **54**: 102422 [PMID: [34118565](https://pubmed.ncbi.nlm.nih.gov/34118565/) DOI: [10.1016/j.scr.2021.102422](https://doi.org/10.1016/j.scr.2021.102422)]
- 179 **Lopaschuk GD**, Ussher JR, Folmes CD, Jaswal JS, Stanley WC. Myocardial fatty acid metabolism in health and disease. *Physiol Rev* 2010; **90**: 207-258 [PMID: [20086077](https://pubmed.ncbi.nlm.nih.gov/20086077/) DOI: [10.1152/physrev.00015.2009](https://doi.org/10.1152/physrev.00015.2009)]
- 180 **Schnelle M**, Chong M, Zoccarato A, Elkenani M, Sawyer GJ, Hasenfuss G, Ludwig C, Shah AM. In vivo [^{13}C]glucose labeling to assess heart metabolism in murine models of pressure and volume overload. *Am J Physiol Heart Circ Physiol* 2020; **319**: H422-H431 [PMID: [32648823](https://pubmed.ncbi.nlm.nih.gov/32648823/) DOI: [10.1152/ajpheart.00219.2020](https://doi.org/10.1152/ajpheart.00219.2020)]
- 181 **Allen DK**, Young JD. Tracing metabolic flux through time and space with isotope labeling experiments. *Curr Opin Biotechnol* 2020; **64**: 92-100 [PMID: [31864070](https://pubmed.ncbi.nlm.nih.gov/31864070/) DOI: [10.1016/j.copbio.2019.11.003](https://doi.org/10.1016/j.copbio.2019.11.003)]
- 182 **Mookerjee SA**, Goncalves RLS, Gerencser AA, Nicholls DG, Brand MD. The contributions of respiration and glycolysis to extracellular acid production. *Biochim Biophys Acta* 2015; **1847**: 171-181 [DOI: [10.1016/j.bbabo.2014.10.005](https://doi.org/10.1016/j.bbabo.2014.10.005)]
- 183 **Plitzko B**, Loesgen S. Measurement of oxygen consumption rate (OCR) and extracellular acidification rate (ECAR) in culture cells for assessment of the energy metabolism. *Bio Protoc* 2018; **8**: e2850 [PMID: [34285967](https://pubmed.ncbi.nlm.nih.gov/34285967/) DOI: [10.21769/BioProtoc.2850](https://doi.org/10.21769/BioProtoc.2850)]

- 184 **House A**, Fatica E, Shah R, Stergar J, Pearce R, Sandlers Y. A protocol for metabolic characterization of human induced pluripotent stem cell-derived cardiomyocytes (iPS-CM). *MethodsX* 2020; **7**: 100572 [PMID: 32140435 DOI: 10.1016/j.mex.2019.05.028]
- 185 **Rajamohan D**, Kalra S, Duc Hoang M, George V, Staniforth A, Russell H, Yang X, Denning C. Automated Electrophysiological and Pharmacological Evaluation of Human Pluripotent Stem Cell-Derived Cardiomyocytes. *Stem Cells Dev* 2016; **25**: 439-452 [PMID: 26906236 DOI: 10.1089/scd.2015.0253]
- 186 **Sirenko O**, Crittenden C, Callamaras N, Hesley J, Chen YW, Funes C, Rusyn I, Anson B, Cromwell EF. Multiparameter in vitro assessment of compound effects on cardiomyocyte physiology using iPSC cells. *J Biomol Screen* 2013; **18**: 39-53 [PMID: 22972846 DOI: 10.1177/1087057112457590]
- 187 **Mioulane M**, Foldes G, Ali NN, Schneider MD, Harding SE. Development of high content imaging methods for cell death detection in human pluripotent stem cell-derived cardiomyocytes. *J Cardiovasc Transl Res* 2012; **5**: 593-604 [PMID: 22896035 DOI: 10.1007/s12265-012-9396-1]
- 188 **Gupte SA**. Glucose-6-phosphate dehydrogenase: a novel therapeutic target in cardiovascular diseases. *Curr Opin Investig Drugs* 2008; **9**: 993-1000 [PMID: 18729006]
- 189 **Jain M**, Cui L, Brenner DA, Wang B, Handy DE, Leopold JA, Loscalzo J, Apstein CS, Liao R. Increased myocardial dysfunction after ischemia-reperfusion in mice lacking glucose-6-phosphate dehydrogenase. *Circulation* 2004; **109**: 898-903 [PMID: 14757696 DOI: 10.1161/01.CIR.0000112605.43318.CA]
- 190 **Katara R**, Oikawa A, Cesselli D, Beltrami AP, Avolio E, Muthukrishnan D, Munasinghe PE, Angelini G, Emanuelli C, Madeddu P. Boosting the pentose phosphate pathway restores cardiac progenitor cell availability in diabetes. *Cardiovasc Res* 2013; **97**: 55-65 [PMID: 22997160 DOI: 10.1093/cvr/cvs291]
- 191 **Doss MX**, Gaspar JA, Winkler J, Hescheler J, Schulz H, Sachinidis A. Specific gene signatures and pathways in mesodermal cells and their derivatives derived from embryonic stem cells. *Stem Cell Rev Rep* 2012; **8**: 43-54 [PMID: 21519850 DOI: 10.1007/s12015-011-9263-5]
- 192 **Semenza GL**. Hypoxia-inducible factor 1: regulator of mitochondrial metabolism and mediator of ischemic preconditioning. *Biochim Biophys Acta* 2011; **1813**: 1263-1268 [PMID: 20732359 DOI: 10.1016/j.bbamer.2010.08.006]
- 193 **Bayda S**, Adeel M, Tuccinardi T, Cordani M, Rizzolio F. The History of Nanoscience and Nanotechnology: From Chemical-Physical Applications to Nanomedicine. *Molecules* 2019; **25** [PMID: 31892180 DOI: 10.3390/molecules25010112]
- 194 **Wang M**, Thanou M. Targeting nanoparticles to cancer. *Pharmacol Res* 2010; **62**: 90-99 [PMID: 20380880 DOI: 10.1016/j.phrs.2010.03.005]
- 195 **Affi MM**, El Sheikh SM, Abdelsalam MM, Ramadan H, Omar TA, El Tantawi M, Abdel-Razek KM, Mohamed M. Therapeutic efficacy of plasmonic photothermal nanoparticles in hamster buccal pouch carcinoma. *Oral Surg Oral Med Oral Pathol Oral Radiol* 2013; **115**: 743-751 [PMID: 23454046 DOI: 10.1016/j.oooo.2012.11.020]
- 196 **Mohamed MM**, Raslan HS, Ramadan OR, Rafik ST, Awaad AK, Essawy MM. Biocompatible Luminescent Nanosized Curcumin: Verified Parameters Affecting Stability and Bioavailability. *Int J Dentistry Oral Sci* 2020; **7**: 1000-1006 [DOI: 10.19070/2377-8075-20000238]
- 197 **Essawy MM**, El-Sheikh SM, Raslan HS, Ramadan HS, Kang B, Talaat IM, Affi MM. Function of gold nanoparticles in oral cancer beyond drug delivery: Implications in cell apoptosis. *Oral Dis* 2021; **27**: 251-265 [PMID: 32657515 DOI: 10.1111/odi.13551]
- 198 **Chavakis E**, Urbich C, Dimmeler S. Homing and engraftment of progenitor cells: a prerequisite for cell therapy. *J Mol Cell Cardiol* 2008; **45**: 514-522 [PMID: 18304573 DOI: 10.1016/j.yjmcc.2008.01.004]
- 199 **Sun Y**, Lu Y, Yin L, Liu Z. The Roles of Nanoparticles in Stem Cell-Based Therapy for Cardiovascular Disease. *Front Bioeng Biotechnol* 2020; **8**: 947 [PMID: 32923434 DOI: 10.3389/fbioe.2020.00947]
- 200 **Kingsley JD**, Ranjan S, Dasgupta N, Saha P. Nanotechnology for tissue engineering: need, techniques and applications. *J Pharm Res* 2013; **7**: 200-204 [DOI: 10.1016/j.jopr.2013.02.021]
- 201 **Zong X**, Bien H, Chung CY, Yin L, Fang D, Hsiao BS, Chu B, Entcheva E. Electrospun fine-textured scaffolds for heart tissue constructs. *Biomaterials* 2005; **26**: 5330-5338 [PMID: 15814131 DOI: 10.1016/j.biomaterials.2005.01.052]
- 202 **Liu Q**, Tian S, Zhao C, Chen X, Lei I, Wang Z, Ma PX. Porous nanofibrous poly(L-lactic acid) scaffolds supporting cardiovascular progenitor cells for cardiac tissue engineering. *Acta Biomater* 2015; **26**: 105-114 [PMID: 26283164 DOI: 10.1016/j.actbio.2015.08.017]
- 203 **Li J**, Minami I, Shiozaki M, Yu L, Yajima S, Miyagawa S, Shiba Y, Morone N, Fukushima S, Yoshioka M, Li S, Qiao J, Li X, Wang L, Kotera H, Nakatsuji N, Sawa Y, Chen Y, Liu L. Human Pluripotent Stem Cell-Derived Cardiac Tissue-like Constructs for Repairing the Infarcted Myocardium. *Stem Cell Reports* 2017; **9**: 1546-1559 [PMID: 29107590 DOI: 10.1016/j.stemcr.2017.09.007]
- 204 **Zhang Y**, Fan W, Wang K, Wei H, Zhang R, Wu Y. Novel preparation of Au nanoparticles loaded Laponite nanoparticles/ECM injectable hydrogel on cardiac differentiation of resident cardiac stem cells to cardiomyocytes. *J Photochem Photobiol B* 2019; **192**: 49-54 [PMID: 30682654 DOI: 10.1016/j.jphotobiol.2018.12.022]
- 205 **Strebhardt K**, Ullrich A. Paul Ehrlich's magic bullet concept: 100 years of progress. *Nat Rev Cancer* 2008; **8**: 473-480 [PMID: 18469827 DOI: 10.1038/nrc2394]

- 206 **Naahidi S**, Jafari M, Edalat F, Raymond K, Khademhosseini A, Chen P. Biocompatibility of engineered nanoparticles for drug delivery. *J Control Release* 2013; **166**: 182-194 [PMID: 23262199 DOI: 10.1016/j.jconrel.2012.12.013]
- 207 **Padin-Iruegas ME**, Misao Y, Davis ME, Segers VF, Esposito G, Tokunou T, Urbanek K, Hosoda T, Rota M, Anversa P, Leri A, Lee RT, Kajstura J. Cardiac progenitor cells and biotinylated insulin-like growth factor-1 nanofibers improve endogenous and exogenous myocardial regeneration after infarction. *Circulation* 2009; **120**: 876-887 [PMID: 19704095 DOI: 10.1161/CIRCULATIONAHA.109.852285]
- 208 **Tokunaga M**, Liu ML, Nagai T, Iwanaga K, Matsuura K, Takahashi T, Kanda M, Kondo N, Wang P, Naito AT, Komuro I. Implantation of cardiac progenitor cells using self-assembling peptide improves cardiac function after myocardial infarction. *J Mol Cell Cardiol* 2010; **49**: 972-983 [PMID: 20869968 DOI: 10.1016/j.yjmcc.2010.09.015]
- 209 **Tang J**, Shen D, Caranasos TG, Wang Z, Vandergriff AC, Allen TA, Hensley MT, Dinh PU, Cores J, Li TS, Zhang J, Kan Q, Cheng K. Therapeutic microparticles functionalized with biomimetic cardiac stem cell membranes and secretome. *Nat Commun* 2017; **8**: 13724 [PMID: 28045024 DOI: 10.1038/ncomms13724]
- 210 **Mazrouaa AM**, Mohamed MG, Fekry M. Physical and magnetic properties of iron oxide nanoparticles with a different molar ratio of ferrous and ferric. *Egypt J Petrol* 2019; **28**: 165-171 [DOI: 10.1016/j.ejpe.2019.02.002]
- 211 **Dai W**, Hale SL, Kay GL, Jyrala AJ, Kloner RA. Delivering stem cells to the heart in a collagen matrix reduces relocation of cells to other organs as assessed by nanoparticle technology. *Regen Med* 2009; **4**: 387-395 [DOI: 10.2217/rme.09.2]
- 212 **Cheng K**, Malliaras K, Li TS, Sun B, Houde C, Galang G, Smith J, Matsushita N, Marbán E. Magnetic enhancement of cell retention, engraftment, and functional benefit after intracoronary delivery of cardiac-derived stem cells in a rat model of ischemia/reperfusion. *Cell Transplant* 2012; **21**: 1121-1135 [PMID: 22405128 DOI: 10.3727/096368911X627381]
- 213 **Vandergriff AC**, Hensley TM, Henry ET, Shen D, Anthony S, Zhang J, Cheng K. Magnetic targeting of cardiosphere-derived stem cells with ferumoxytol nanoparticles for treating rats with myocardial infarction. *Biomaterials* 2014; **35**: 8528-8539 [PMID: 25043570 DOI: 10.1016/j.biomaterials.2014.06.031]
- 214 **Sukhanova A**, Bozrova S, Sokolov P, Berestovoy M, Karaulov A, Nabiev I. Dependence of Nanoparticle Toxicity on Their Physical and Chemical Properties. *Nanoscale Res Lett* 2018; **13**: 44 [PMID: 29417375 DOI: 10.1186/s11671-018-2457-x]
- 215 **Oh K**, Kim SR, Kim DK, Seo MW, Lee C, Lee HM, Oh JE, Choi EY, Lee DS, Gho YS, Park KS. In Vivo Differentiation of Therapeutic Insulin-Producing Cells from Bone Marrow Cells via Extracellular Vesicle-Mimetic Nanovesicles. *ACS Nano* 2015; **9**: 11718-11727 [PMID: 26513554 DOI: 10.1021/acsnano.5b02997]
- 216 **Jo W**, Jeong D, Kim J, Park J. Self-Renewal of Bone Marrow Stem Cells by Nanovesicles Engineered from Embryonic Stem Cells. *Adv Healthc Mater* 2016; **5**: 3148-3156 [PMID: 27860451 DOI: 10.1002/adhm.201600810]
- 217 **Lee JR**, Park BW, Kim J, Choo YW, Kim HY, Yoon JK, Kim H, Hwang JW, Kang M, Kwon SP, Song SY, Ko IO, Park JA, Ban K, Hyeon T, Park HJ, Kim BS. Nanovesicles derived from iron oxide nanoparticles-incorporated mesenchymal stem cells for cardiac repair. *Sci Adv* 2020; **6**: eaaz0952 [PMID: 32494669 DOI: 10.1126/sciadv.aaz0952]
- 218 **Kojima R**, Bojar D, Rizzi G, Hamri GC, El-Baba MD, Saxena P, Ausländer S, Tan KR, Fussenegger M. Designer exosomes produced by implanted cells intracerebrally deliver therapeutic cargo for Parkinson's disease treatment. *Nat Commun* 2018; **9**: 1305 [PMID: 29610454 DOI: 10.1038/s41467-018-03733-8]
- 219 **von Harsdorf R**, Poole-Wilson PA, Dietz R. Regenerative capacity of the myocardium: implications for treatment of heart failure. *Lancet* 2004; **363**: 1306-1313 [PMID: 15094278 DOI: 10.1016/S0140-6736(04)16006-6]
- 220 **Laflamme MA**, Zbinden S, Epstein SE, Murry CE. Cell-based therapy for myocardial ischemia and infarction: pathophysiological mechanisms. *Annu Rev Pathol* 2007; **2**: 307-339 [PMID: 18039102 DOI: 10.1146/annurev.pathol.2.010506.092038]
- 221 **Malliaras K**, Makkar RR, Smith RR, Cheng K, Wu E, Bonow RO, Marbán L, Mendizabal A, Cingolani E, Johnston PV, Gerstenblith G, Schuleri KH, Lardo AC, Marbán E. Intracoronary cardiosphere-derived cells after myocardial infarction: evidence of therapeutic regeneration in the final 1-year results of the CADUCEUS trial (CARDiosphere-Derived aUtologous stem CElls to reverse ventricular dysfunction). *J Am Coll Cardiol* 2014; **63**: 110-122 [PMID: 24036024 DOI: 10.1016/j.jacc.2013.08.724]
- 222 **Makkar RR**, Kereiakes DJ, Aguirre F, Kowalchuk G, Chakravarty T, Malliaras K, Francis GS, Povsic TJ, Schatz R, Traverse JH, Pogoda JM, Smith RR, Marbán L, Ascheim DD, Ostovaneh MR, Lima JAC, DeMaria A, Marbán E, Henry TD. Intracoronary ALLogeneic heart Stem cells to Achieve myocardial Regeneration (ALLSTAR): a randomized, placebo-controlled, double-blinded trial. *Eur Heart J* 2020; **41**: 3451-3458 [PMID: 32749459 DOI: 10.1093/eurheartj/ehaa541]
- 223 **Menasché P**, Vanneau V, Hagege A, Bel A, Cholley B, Parouchev A, Cacciapuoti I, Al-Daccak R, Benhamouda N, Blons H, Agbulut O, Tosca L, Trouvin JH, Fabreguettes JR, Bellamy V, Charron D, Tartour E, Tachdjian G, Desnos M, Larghero J. Transplantation of Human Embryonic Stem Cell-Derived Cardiovascular Progenitors for Severe Ischemic Left Ventricular Dysfunction. *J Am Coll*

- Cardiol* 2018; **71**: 429-438 [PMID: 29389360 DOI: 10.1016/j.jacc.2017.11.047]
- 224 **Mirotsov M**, Jayawardena TM, Schmeckpeper J, Gnecci M, Dzau VJ. Paracrine mechanisms of stem cell reparative and regenerative actions in the heart. *J Mol Cell Cardiol* 2011; **50**: 280-289 [PMID: 20727900 DOI: 10.1016/j.yjmcc.2010.08.005]
- 225 **Lalit PA**, Salick MR, Nelson DO, Squirrell JM, Shafer CM, Patel NG, Saeed I, Schmuck EG, Markandeya YS, Wong R, Lea MR, Eliceiri KW, Hacker TA, Crone WC, Kyba M, Garry DJ, Stewart R, Thomson JA, Downs KM, Lyons GE, Kamp TJ. Lineage Reprogramming of Fibroblasts into Proliferative Induced Cardiac Progenitor Cells by Defined Factors. *Cell Stem Cell* 2016; **18**: 354-367 [PMID: 26877223 DOI: 10.1016/j.stem.2015.12.001]
- 226 **Witman N**, Sahara M. Cardiac Progenitor Cells in Basic Biology and Regenerative Medicine. *Stem Cells Int* 2018; **2018**: 8283648 [PMID: 29535783 DOI: 10.1155/2018/8283648]
- 227 **Sanz-Ruiz R**, Casado Plasencia A, Borlado LR, Fernández-Santos ME, Al-Daccak R, Claus P, Palacios I, Sádaba R, Charron D, Bogaert J, Mulet M, Yotti R, Gilaberte I, Bernad A, Bermejo J, Janssens S, Fernández-Avilés F. Rationale and Design of a Clinical Trial to Evaluate the Safety and Efficacy of Intracoronary Infusion of Allogeneic Human Cardiac Stem Cells in Patients With Acute Myocardial Infarction and Left Ventricular Dysfunction: The Randomized Multicenter Double-Blind Controlled CAREMI Trial (Cardiac Stem Cells in Patients With Acute Myocardial Infarction). *Circ Res* 2017; **121**: 71-80 [PMID: 28533209 DOI: 10.1161/CIRCRESAHA.117.310651]
- 228 **Victor RG**. Significant Improvements Reported in Duchenne Muscular Dystrophy Patients Treated with Capricor's Investigational Cell Therapy. Capricor Therapeutics, Inc. 2017. [cited 26 February 2021] Available from: <http://www.pnewswire.com/news-releases/significant-improvements-reported-in-duchenne-muscular-dystrophy-patients-treated-with-capricors-investigational-cell-therapy-300556679.html>
- 229 **Takehara N**, Tsutsumi Y, Tateishi K, Ogata T, Tanaka H, Ueyama T, Takahashi T, Takamatsu T, Fukushima M, Komeda M, Yamagishi M, Yaku H, Tabata Y, Matsubara H, Oh H. Controlled delivery of basic fibroblast growth factor promotes human cardiosphere-derived cell engraftment to enhance cardiac repair for chronic myocardial infarction. *J Am Coll Cardiol* 2008; **52**: 1858-1865 [PMID: 19038683 DOI: 10.1016/j.jacc.2008.06.052]
- 230 **Bolli R**, Hare JM, March KL, Pepine CJ, Willerson JT, Perin EC, Yang PC, Henry TD, Traverse JH, Mitrani RD, Khan A, Hernandez-Schulman I, Taylor DA, DiFede DL, Lima JAC, Chugh A, Loughran J, Vojvodic RW, Sayre SL, Bettencourt J, Cohen M, Moyé L, Ebert RF, Simari RD; Cardiovascular Cell Therapy Research Network (CCTRN). Rationale and Design of the CONCERT-HF Trial (Combination of Mesenchymal and c-kit⁺ Cardiac Stem Cells As Regenerative Therapy for Heart Failure). *Circ Res* 2018; **122**: 1703-1715 [PMID: 29703749 DOI: 10.1161/circresaha.118.312978]
- 231 **Ishigami S**, Ohtsuki S, Tarui S, Ousaka D, Eitoku T, Kondo M, Okuyama M, Kobayashi J, Baba K, Arai S, Kawabata T, Yoshizumi K, Tateishi A, Kuroko Y, Iwasaki T, Sato S, Kasahara S, Sano S, Oh H. Intracoronary autologous cardiac progenitor cell transfer in patients with hypoplastic left heart syndrome: the TICAP prospective phase 1 controlled trial. *Circ Res* 2015; **116**: 653-664 [PMID: 25403163 DOI: 10.1161/CIRCRESAHA.116.304671]
- 232 **Ishigami S**, Ohtsuki S, Eitoku T, Ousaka D, Kondo M, Kurita Y, Hirai K, Fukushima Y, Baba K, Goto T, Horio N, Kobayashi J, Kuroko Y, Kotani Y, Arai S, Iwasaki T, Sato S, Kasahara S, Sano S, Oh H. Intracoronary Cardiac Progenitor Cells in Single Ventricle Physiology: The PERSEUS (Cardiac Progenitor Cell Infusion to Treat Univentricular Heart Disease) Randomized Phase 2 Trial. *Circ Res* 2017; **120**: 1162-1173 [PMID: 28052915 DOI: 10.1161/CIRCRESAHA.116.310253]
- 233 **Tarui S**, Ishigami S, Ousaka D, Kasahara S, Ohtsuki S, Sano S, Oh H. Transcoronary infusion of cardiac progenitor cells in hypoplastic left heart syndrome: Three-year follow-up of the Transcoronary Infusion of Cardiac Progenitor Cells in Patients With Single-Ventricle Physiology (TICAP) trial. *J Thorac Cardiovasc Surg* 2015; **150**: 1198-1207, 1208.e1 [PMID: 26232942 DOI: 10.1016/j.jtcvs.2015.06.076]
- 234 **Bergmann O**, Zdunek S, Felker A, Salehpour M, Alkass K, Bernard S, Sjöström SL, Szewczykowska M, Jackowska T, Dos Remedios C, Malm T, Andrä M, Jashari R, Nyengaard JR, Possnert G, Jovinge S, Druid H, Frisén J. Dynamics of Cell Generation and Turnover in the Human Heart. *Cell* 2015; **161**: 1566-1575 [PMID: 26073943 DOI: 10.1016/j.cell.2015.05.026]
- 235 **Doppler SA**, Deutsch MA, Lange R, Krane M. Cardiac regeneration: current therapies-future concepts. *J Thorac Dis* 2013; **5**: 683-697 [PMID: 24255783 DOI: 10.3978/j.issn.2072-1439.2013.08.71]
- 236 **Hatzistergos KE**, Quevedo H, Oskouei BN, Hu Q, Feigenbaum GS, Margitich IS, Mazhari R, Boyle AJ, Zambrano JP, Rodriguez JE, Dulce R, Pattany PM, Valdes D, Revilla C, Heldman AW, McNiece I, Hare JM. Bone marrow mesenchymal stem cells stimulate cardiac stem cell proliferation and differentiation. *Circ Res* 2010; **107**: 913-922 [PMID: 20671238 DOI: 10.1161/CIRCRESAHA.110.222703]
- 237 **Williams AR**, Hatzistergos KE, Addicott B, McCall F, Carvalho D, Suncion V, Morales AR, Da Silva J, Sussman MA, Heldman AW, Hare JM. Enhanced effect of combining human cardiac stem cells and bone marrow mesenchymal stem cells to reduce infarct size and to restore cardiac function after myocardial infarction. *Circulation* 2013; **127**: 213-223 [PMID: 23224061 DOI: 10.1161/CIRCULATIONAHA.112.131110]
- 238 **Karantalis V**, Suncion-Loescher VY, Bagno L, Golpanian S, Wolf A, Sanina C, Premer C, Kanelidis AJ, McCall F, Wang B, Balkan W, Rodriguez J, Rosado M, Morales A, Hatzistergos K,

- Natsumeda M, Margitich I, Schulman IH, Gomes SA, Mushtaq M, DiFede DL, Fishman JE, Pattany P, Zambrano JP, Heldman AW, Hare JM. Synergistic Effects of Combined Cell Therapy for Chronic Ischemic Cardiomyopathy. *J Am Coll Cardiol* 2015; **66**: 1990-1999 [PMID: 26516002 DOI: 10.1016/j.jacc.2015.08.879]
- 239 **Natsumeda M**, Florea V, Rieger AC, Tompkins BA, Banerjee MN, Golpanian S, Fritsch J, Landin AM, Kashikar ND, Karantalis V, Loescher VY, Hatzistergos KE, Bagno L, Sanina C, Mushtaq M, Rodriguez J, Rosado M, Wolf A, Collon K, Vincent L, Kanelidis AJ, Schulman IH, Mitrani R, Heldman AW, Balkan W, Hare JM. A Combination of Allogeneic Stem Cells Promotes Cardiac Regeneration. *J Am Coll Cardiol* 2017; **70**: 2504-2515 [PMID: 29145950 DOI: 10.1016/j.jacc.2017.09.036]
- 240 **Zhang Z**, Yang J, Yan W, Li Y, Shen Z, Asahara T. Pretreatment of Cardiac Stem Cells With Exosomes Derived From Mesenchymal Stem Cells Enhances Myocardial Repair. *J Am Heart Assoc* 2016; **5** [PMID: 26811168 DOI: 10.1161/jaha.115.002856]
- 241 **Quijada P**, Salunga HT, Hariharan N, Cubillo JD, El-Sayed FG, Moshref M, Bala KM, Emathingier JM, De La Torre A, Ormachea L, Alvarez R Jr, Gude NA, Sussman MA. Cardiac Stem Cell Hybrids Enhance Myocardial Repair. *Circ Res* 2015; **117**: 695-706 [PMID: 26228030 DOI: 10.1161/CIRCRESAHA.115.306838]
- 242 **Firouzi F**, Sinha Choudhury S, Broughton K, Salazar A, Bailey B, Sussman MA. Human CardioChimeras: Creation of a Novel "Next-Generation" Cardiac Cell. *J Am Heart Assoc* 2020; **9**: e013452 [PMID: 31902324 DOI: 10.1161/JAHA.119.013452]
- 243 **Monsanto MM**, Wang BJ, Ehrenberg ZR, Echeagaray O, White KS, Alvarez R Jr, Fisher K, Sengphanith S, Muliono A, Gude NA, Sussman MA. Enhancing myocardial repair with CardioClusters. *Nat Commun* 2020; **11**: 3955 [PMID: 32769998 DOI: 10.1038/s41467-020-17742-z]
- 244 **Yokoyama R**, Ii M, Masuda M, Tabata Y, Hoshiga M, Ishizaka N, Asahi M. Cardiac Regeneration by Statin-Polymer Nanoparticle-Loaded Adipose-Derived Stem Cell Therapy in Myocardial Infarction. *Stem Cells Transl Med* 2019; **8**: 1055-1067 [PMID: 31157513 DOI: 10.1002/sctm.18-0244]
- 245 **Magouroux T**, Extermann J, Hoffmann P, Mugnier Y, Le Dantec R, Jaconi ME, Kasparian C, Ciepielewski D, Bonacina L, Wolf JP. High-speed tracking of murine cardiac stem cells by harmonic nanodoublers. *Small* 2012; **8**: 2752-2756 [PMID: 22859385 DOI: 10.1002/sml.201200366]



Published by **Baishideng Publishing Group Inc**
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA
Telephone: +1-925-3991568
E-mail: bpgoffice@wjgnet.com
Help Desk: <https://www.f6publishing.com/helpdesk>
<https://www.wjgnet.com>

