

What factors guide treatment selection in mycosis fungoides and Sezary syndrome?

Youn H. Kim

Department of Dermatology, Stanford University School of Medicine, Palo Alto, CA

Cutaneous T-cell lymphoma (CTCL) comprises a spectrum of T-cell lymphomas with primary skin involvement. Mycosis fungoides (MF) and Sezary syndrome (SS) are the common subtypes of CTCL in which patients present with widely diverse profiles of skin involvement and varying extents of extracutaneous disease. Patients with early-stage disease have an excellent prognosis and are managed primarily with skin-directed therapies; however, those with advanced-stage MF or SS often require multiple lines and recurrent courses of systemic therapies. Many options are available when considering systemic agents, and it is often challenging to know how to prioritize therapies to address a patient's objective disease and quality of life issues. Appreciating the disease heterogeneity and understanding the patient's overall disease profile (eg, skin, lymph nodes, blood, large cell transformation) serve as a useful framework in aligning therapies that can optimally treat active sites of disease. Tissue or blood biomarkers can be integrated into our process of prioritizing therapies and personalizing management in MF or SS. Multidisciplinary management and optimizing supportive care are additional key elements for a favorable outcome. Appropriate patients with high-risk disease should be considered for allogeneic hematopoietic stem cell transplant.

LEARNING OBJECTIVES

- Understand the clinical heterogeneity of patients with MF and SS
- Appreciate the unique quality of life issues associated with cutaneous disease
- Identify the clinical, pathological, and other biomarkers that help prioritize therapies and personalize clinical management in MF and SS

Introduction

Mycosis fungoides (MF) and Sezary syndrome (SS) are the most common subtypes of cutaneous T-cell lymphoma (CTCL), and although the skin is the primary site of involvement, the malignant T cells may expand in the lymph node (LN), visceral, and blood compartments.¹ In the skin lesions of early-stage MF, significant inflammatory infiltrate of immune stimulated profile is observed. However, as the disease progresses the malignant T cells acquire a Th2 phenotype accompanied by a microenvironment skewed toward a more immune-suppressive Th2 cytokine profile.^{2,3} A subset of patients with advanced MF or SS undergo a transformation into large cell disease (LCT) that is often associated with more aggressive biological behavior. Patients with MF and SS subgroups are often distinct clinically and biologically but also share similarities, and clinical features can shift from MF to SS and vice versa in the course of a patient's disease.

The clinical management of MF/SS involves therapies developed specifically for patients with CTCL (eg, total

skin electron beam therapy [TSEBT], extracorporeal photopheresis, bexarotene, vorinostat, romidepsin) and those originally developed to treat other lymphomas but coopted for CTCL (eg, brentuximab vedotin [BV], pralatrexate). More recently, new systemic therapies have emerged, built upon rationally selected targets in MF/SS (eg, mogamulizumab [MOGA], anti-*KIR3DL2* antibody; Table 1).

Although these newer agents represent a major advancement, they have not altered our fundamental treatment strategy. Only a slim minority of patients will ever be cured of their disease, and most patients will receive numerous lines of therapy during their lives. Each failed treatment takes a physical and psychological toll on patients. We have long known that a one-size-fits-all approach cannot work for this heterogeneous disease. As our therapeutic armamentarium continues to expand, we are now challenged with how to best match patients to the treatments most likely to benefit them. Furthermore, it is unclear how best to sequence or combine therapies, and given the lack of curative therapies, the ones with fewer cumulative toxicities and durable

		Global/composite clinical activity of all compartments*	clinical activity o	ıf all					Tolerability
Treatment	N; subtype	ORR/CR % (n/N)	DOR, median	PFS, median	Skin activity	LN activity	Blood activity	LCT included	therapy
Standard therapies Bexarotene' ^{:22} (300mg/m ² =opti- mal dose)	56; MF/SS IIB-IV; SS (17/56)	45% (25/56)/1 CR	42.7 weeks	13.9 weeks	Skin RR=primary end point: IIB, 57% (13/23); III, 32% (6/19); INA, 44% (4/9); IVB, 40% (2/5); SS, 24% (4/17, all doses)	Limited LN data: Of 25 skin OR, 7 w/clinical LAD - 3/7 PR or CR 4/7 PD	Of 17 SS, 8 had partial Sezary data: 4/8 PR 3/8 SD 1/8 PD	Na	Hyperlipidemia
Extracorporeal photopheresis ^{t,24}	51; MF/SS III-IV; SS (39/51)	63% (32/51)/8 CR	22.4 months	N/a	Compartment-specific data not reported but 16% CR w/clearing of blood Sezary disease	ic data not reporte d Sezary disease	d but 16% CR	N/a	None
Vorinostat ⁶³	74; MF/SS IB-IV; SS (30/74)	74; MF/SS IB-IV; 30% (22/74)/1 CR SS (30/74) after 281 days	>185 days	TTP median 148 days	Skin RR=primary end point: IB/IIA, 31% (4/13); IIB-IV, 30% (18/61); skin tumors, 23% (5/22); SS, 33% (10/30)	LN RR 42% (10/24)	14/27 SS w/SC reduction >25%	Yes	Fatigue, dysgeusia, ↓platelets
Romidepsin ⁱ⁴	96; MF/SS IB-IV; SS (37/96)	34% (33/96)/6 CR; SS (B1-2), 32% (12/37)	15 months	TTP median 8 months	Skin RR 40% (38/96)	LN response 35% (13/37) by RECIST	Blood RR 38% (14/37, 2 CR); B2, 46% (6/13, 2 CR)	Yes	Fatigue, dysgeusia, ↓platelets
Pralatrexate ^{\$,37} (optimal CTCL dose 15mg/m² weekly 3/4)	29; CTCL IB-IV; SS (8/29)	45% (13/29)/1 CR	Not reached; 73% cont OR at 6 months	Not reached; 388 days >15mg/m²	Compartment-specific data not reported, response by stage: MF by stage IB 60% (3/5); IIB 67% (4/6); IVA 60% (3/5); SS 25% (2/8)	ic data not reporte	d, response by	Yes	Mucositis
Brentuximab ¹¹	4:8; MF IA-IV; CD30>10% (no SS)	65% (31/48)/ 5 CR; ORR4 LCT 65% (11/17)	15.1 months; MF/ALCL	15.9 months; LCT 15.5 months	Skin RR 77% (37/48); median DOR 20.6 months, MF/ALCL	2 w/LN+, stage IVA: ORR 100% (2/2, 1 CR)	Excluded B2/SS	Yes (17/48)	Z
Mogamulizumab ^{s,15}	186; MF/SS IB-IV; SS (81/186)	28% (52/186)/6 CR; MF 21% (22/105); SS 37% (30/81)	14.1 months; MF 13.1 months; SS 17.3 months months	7.7 months	Skin RR 42% (78/186); median TTR 3.0 months; median DOR 20.6 months	LN RR 17% (21/124); median TTR 3.3 months; median DOR 15.5 months	Blood RR 68% (83/122, 54 CR); median TTR 1.1 months; median DOR 25.5 months	No (LCT excluded)	Rash
Pembrolizumab ⁴¹	24; MF/SS IIB- IV (23/24); SS (15/24)	38% (9/24)/2 CR; SS 27% (4/15); LCT 25% (1/4)	Not reached; median follow-up 58 weeks	Not reached; PFS at 1 year, 65%	Skin RR 38% (9/24); 6/24, >90% skin clearing	N/a	Baseline w/B2, n=6, 17% (1/6)	Yes (4/24)	irAE (colitis, pneu- monitis)

Table 1. Clinical activity of systemic therapies in CTCL* by disease subtype (clinical stage, MF vs SS), compartments (skin, LN, blood), ±LCT; tolerability

tinued)	
1. (cont	
Table	

		Global/composite cl compartments*	clinical activity of all	of all					Tolerability factors in chronic
Treatment	N; subtype	ORR/CR % (n/N)	DOR, median	PFS, median	Skin activity	LN activity	Blood activity	LCT included	therapy
Liposomal doxorubi- cin ^{5,39} (20mg/m², day 1, day 15, 28-day cycle)	49; MF IIB-IV (no SS)	41% (20/49)/3 CR	6 months	6.2 months; TTP median 7.4 months	skin RR 53% (26/49); best PR/CR	LN±visceral disease had lower RR than skin-only (22% vs 52%)	Excluded SS	Yes	
Gemcitabine ^{s40} (1000 mg/m², day 1, 8, 15)	31; MF/SS; SS (11/33)	65% (20/31)/3 CR; LCT 54% (7/13); SS 73% (8/11)	4.1 months	N/a	Compartment-specific data not reported but case exam- Yes (13/31) ples with activity in LN/blood disease	ompartment-specific data not reportec ples with activity in LN/blood disease	d but case exam- e	Yes (13/31)	
Investigative therapies (w/peer-reviewed papers)									
Anti- <i>KIR3DL2</i> anti- body (lacutamab), phase 1/2 study data (ongoing piv- otal trial in SS) ²⁵	44; SS (35/44); MF IB-IV (8/44)	SS, 43% (15/35); MF, 13% (1/8); LCT no OR	13.8 months; SS 13.8 months; MF 6.9 months	8.2 months; SS 11.7 months, MF 3.9 months	Of SS: skin RR 51% (18/35, 3 CR)	Of SS: LN RR 11% (2/18, 1 CR)	Of SS: blood RR 56% (19/34, 9 CR)	Yes (6/35 SS, 5/8 MF)	Well tolerated
Duvelisib ⁴⁴	19; MF/SS; MF (13/19); SS (5/19)	32% no CR; MF 38% (5/13); LCT 25% (1/4); SS 20% (1/5)	n/a; DOR range 0.7- 10.1 months; TTR 2.4 months	4.5 months	Compartment-specific data not reported	ic data not reporte	σ	Yes (4/19)	≁LFTS
Lenalidomide ⁶⁵	32; MF/SS IB-IV; SS (11/32)	28% (9/32) no CR	10 months; TTR 25mg, 2 months; 10mg, 4 months	8 months	Skin RR 53% (10/19) MF IB-IV	Reports activity, no details	Blood RR 38% (5/13)	N/a	Fatigue, skin flares, transient ≁blood SCs
*Response assessment tools and/or criteria for clinical end points may vary across CTCL trials. Time to response is longer than other listed treatments, 300 mg/m², TTR=15.7 weeks (180 days), >300 mg/m², TTR=8.4 weeks (59 days); poor details on extracutaneous disease sites (4/9 entered as stage IVB had LN+ only): "rate of PD"=39% (12/56) and 32% (12/38) for 300 mg/m² and >300 mg/m²: median TTR for other treatment ~2 cycles.	tools and/or crite nger than other list ad LN+ only); "rate	ria for clinical end poi ted treatments, 300 m of PD"=39% (22/56)	ints may vary acr ng/m², TTR=15.7 and 32% (12/38)	oss CTCL trials. weeks (180 days), for 300 mg/m² ar	>300 mg/m², TTR=8.4 id >300 mg/m², media	• weeks (59 days); p in TTR for other trea	oor details on extr atment ~2 cycles.	acutaneous dise	ase sites (4/9

^{\$}Caution when used with RT, especially TSEBT.

*Median time to response 8 months.

*Caution when used to bridge to allogeneic HSCT (possible risk of severe GVHD). ALCL, anaplastic large cell lymphoma; DOR, duration of response; irAE, immune-related adverse event; LFT, liver function test; n/a, not applicable; OR, odds ratio; PFS, progression-free survival; PN, peripheral neuropathy; RECIST, response evaluation criteria in solid tumors; RT, radiation therapy; SD, stable disease; TTR, time to response; TTP, time to progression.

responses are prioritized. In selected high-risk advanced-stage MF or SS, allogeneic hematopoietic stem cell transplant (HSCT) should be considered as a potential curative therapy.

CLINICAL CASE

A 64-year-old male presented with a 2-year history of progressive skin rash that currently affected >80% of the body surface area with generalized erythroderma associated with severe, debilitating itching. Topical and systemic steroids were minimally helpful. Skin biopsies showed an atypical T-cell infiltrate. Blood flow demonstrated an abnormal T-cell population expressing CD3 and CD4 but lacking CD7 and CD26, a phenotype consistent with Sezary cells (SCs), which represented 68% of lymphocytes with an absolute count of 1250/mm³; T-cell receptor next-generation sequencing (NGS) studies supported a clonal T-cell process in the skin and blood. Whole-body positron emission tomography/computed tomography (PET/CT) imaging showed an approximately 2cm axillary and inguinal lymphadenopathy (LAD) with a maximum standardized uptake value of 3 to 4, consistent with reactive or dermatopathic LNs. A diagnosis of SS was established, presenting with low-burden SCs and clinically reactive LAD and thus clinical stage IVA,. The patient was treated with oral bexarotene with overall global partial response (75% improvement of his skin disease and >50% reduction of his SCs), and supportive care was provided with topical steroids, emollients, and gabapentin for itching. Six months later while continuing maintenance bexarotene, he noted worsening of his skin disease with recurrent erythroderma. A skin biopsy confirmed CTCL without evidence of LCT. His blood Sezary flow now showed >10,000 SCs/mm³. Updated PET/CT findings again supported reactive or dermatopathic LAD with no hepatosplenomegaly.

Recognizing disease heterogeneity within the skin and across extracutaneous compartments

MF and SS are unique in their striking heterogeneity of clinical, biological, and molecular features and the need to individualize management for an optimal outcome. This heterogeneity is further complicated by variable clinical behavior across disease compartments (eg, skin, LN, blood). A meaningful clinical outcome of treatment in MF/SS consists of a durable global (composite of all compartments) objective response and improvement of quality of life (QoL) measures,⁴⁻⁷ and achieving these goals requires individualization of therapy. Traditionally, the clinical stage has been a primary consideration in treatment selection; however, additional guidance is often needed given the heterogeneity within stage groupings. The extent/burden of disease or status of other established prognostic factors (eg, LCT) in the skin or other disease compartments further contributes to treatment selection.⁸

In early-stage MF with patch and/or plaque skin disease, skin-directed treatment is preferred, and improvement of QoL is usually the primary focus of treatment. Primary skin-directed treatments include topical preparations of steroids, mechlorethamine, retinoids, phototherapy (eg, narrowband ultraviolet B), and TSEBT.⁸ TSEBT is very effective at low doses (eg, 12 Gy)

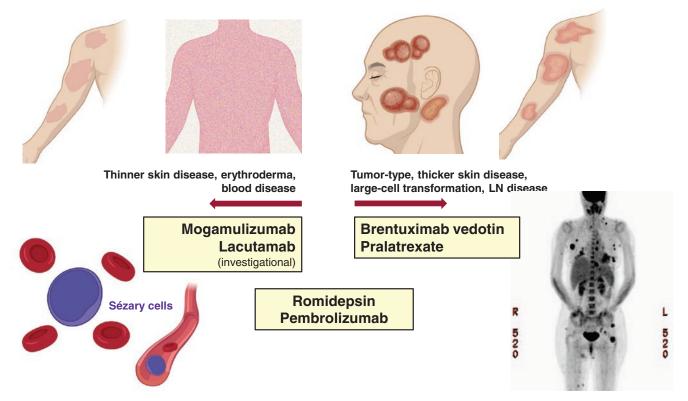


Figure 1. Examples of newer systemic agents and their clinical activity in different patient profiles.

and most appropriate in patients who have widespread thicker or folliculotropic (hair follicle involvement) skin disease.⁹ If skindirected treatment options fail to manage a patient's disease, then escalating to systemic agents is appropriate.

Patients with advanced-stage MF and SS have varying clinical presentations ranging from tumor-type skin disease to generalized erythroderma in which the type and extent of their skin and extracutaneous disease (LN and/or blood disease) drive the therapies that best address a patient's overall disease. Many standard treatment options exist for advanced-stage patients, including systemic biological and targeted therapies or traditional cytotoxic chemotherapy.⁸ The ultimate selection of the primary therapy will be based on the assessment of the skin and the severity of any extracutaneous disease and on aligning therapies that address the most symptomatic and threatening disease.

Systemic treatments often exhibit differential efficacy across skin, blood, and nodal compartments (Table 1 and Figure 1). The response interpretation can be further complicated by the varying response criteria and clinical end points used in earlier clinical trials in CTCL; however, more recent trials have utilized the international consensus criteria that have helped compare results across studies.⁴ There are frequent mixed responses in the skin or across different compartments with disease, and therapies that reduce skin disease may not be effective in clearing the blood or LN disease, and vice versa. For example, in a patient with MF and widespread ulcerating skin tumors (±LCT), we would select treatments with known reliable activity in the skin compartment (eg, TSEBT, BV).9-12 However, in a patient with SS and high-burden Sezary disease, we would opt for therapies that are efficient at reducing the blood disease (eg, MOGA, romidepsin).13,14 Furthermore, some therapies do not have established, meaningful activity in LCT or bulky disease, whereas some therapies have shown activity across all compartments, including LCT disease. Traditional combination chemotherapy regimens in systemic TCLs are often reserved for patients with refractory, advanced disease or when allogeneic HSCT is planned for consolidation.8 Lastly, the potential toxicities and tolerability of chronic therapy must be considered in prioritizing treatments and personalizing management.

Importance of multidisciplinary care and management of QoL issues: essential role of supportive therapy

Ideally, the management of patients with CTCL involves a multidisciplinary approach. It is helpful to establish collaborative colleagues in hematology/medical oncology, radiation oncology, pathology, and dermatology who are interested in cutaneous lymphoma. To address the extensive skin disease, specialized radiation methods such as TSEBT serve as an important therapeutic alternative; however, access to centers with TSEBT can be an issue. Low-dose TSEBT can be combined with treatments that have great efficacy in the blood and/or LN compartments to improve the overall composite response and response duration.^{15,16} Patients may have extensive skin wounds or atypical skin infections or develop skin lesions that clinically and/or pathologically mimic CTCL, including an inflammatory skin reaction to lymphoma therapy (eg, MOGA, pembrolizumab). Thus, a close collaboration across specialties is essential in providing appropriate clinical care and optimal outcome in CTCL.

Intolerable itching is a common QoL in CTCL, especially in SS. The effective management of itching also reduces the risk of sec-

ondary skin infections as patients often have compromised skin barrier function and are commonly colonized with Staphylococcus aureus.¹⁷ Management with an anti-staph supportive regimen has been shown to improve their itching, clear their skin, and lower infection-associated complications.¹⁸ The visible nature of CTCL adds a unique negative life impact because it constantly reminds patients of their disease and often prohibits patients from social interaction, intensifying the emotional burden of their struggles.^{19,20} Thus, in contrast to other indolent lymphomas such as low-grade follicular lymphoma, in which watchful waiting can be acceptable, patients with CTCL are often on constant treatment, and QoL elements may lead to a decision to change therapy without objective disease progression. Optimizing supportive care is essential, and clinical end points such as time to next significant therapy may be useful measures of overall clinical benefit.21

Prioritizing treatment options in SS

In patients with SS, the leukemic subtype of CTCL, clinical management is often stratified by the burden of the blood Sezary disease. Primary systemic therapy for those with a lower Sezary burden (<5,000 SCs/mm³) includes oral bexarotene,²² methotrexate,23 and extracorporeal photopheresis.24 In those with a higher Sezary burden, histone deacetylase inhibitors (eg, romidepsin) or newer therapies, such as MOGA, with more reliable activity in the blood compartment would be appropriate frontline options (Table 1).14 With MOGA, an anti-CCR4 monoclonal antibody in which CCR4 is highly and consistently expressed by SCs, objective blood response was observed in 68% (83/122) of patients with 44% (54/122) achieving complete remission (CR) in the pivotal trial, MAVORIC.¹³ The median time to response in the blood was 1 month, and the median duration of blood response was 26 months. The skin response rate was 42% with a lower nodal response rate of 17% and a global (composite of all compartments) overall response rate of 28%. However, patients with active LCT were excluded in the MOGA pivotal trial. Given the impressive clinical activity in the blood combined with a favorable tolerability profile, MOGA may be a preferred option in high-burden SS without skin LCT or LN disease. Similarly, the anti-KIR3DL2 antibody (lacutamab) currently under clinical development has been shown to be more effective in reducing blood disease compared to nodal disease (blood response rate [RR] 56% vs nodal RR 11%).²⁵ Conversely, BV, which demonstrated reliable efficacy in the skin (including LCT) and the LN disease,¹¹ does not have established activity in Sezary disease and SS and was excluded in the pivotal trial in CTCL. In SS with extensive LN disease (±LCT), romidepsin may be prioritized over MOGA or other options, as romidepsin has demonstrated consistent clinical activity across compartments (Table 1).

Length of therapy and maintenance strategies

To address the chronic course of MF/SS, newer systemic therapies were developed in which clinical studies allowed patients to continue treatment without a fixed duration until intolerance or disease progression occurred. However, for therapies that have dose-cumulative toxicities such as the peripheral neuropathy associated with BV treatment, we should consider a fixed treatment course (<6-8 cycles) and re-treat with BV later when needed or use reduced doses of BV.^{10,26.27} MOGA can be continued long-term, but in one-third of patients, it may lead to a rash (median time to rash, 105 days),^{13,28-30} and the severity of the rash may affect the length of treatment with MOGA. Despite the occurrence of MOGA, patients can be re-treated with MOGA. For chronic therapy in CTCL, treatment options or strategies that minimize immune suppression will be preferred given the high-risk of recurrent skin infections.

After initial disease reduction, a maintenance strategy of a sustainable treatment regimen can be considered. This may involve increasing the treatment interval of intravenous therapies or transitioning to an oral agent.³¹ Utilizing a less intense maintenance dose regimen of therapies may lower treatment-related toxicities and provide a longer overall clinical benefit. Integrating skin-directed therapy is not only essential in combination with systemic agents but necessary when the systemic disease is cleared, and patients then have primarily skin-limited disease.

CLINICAL CASE (CONTINUED)

Given the higher burden of blood Sezary disease and lack of LCT in the skin, the patient was treated with MOGA and after 2 cycles experienced >90% skin improvement and reduction of itching. His blood Sezary disease cleared promptly without measurable abnormal circulating T cells (B0) by flow cytometry. After 4 months of continued great response with continued blood CR (B0), MOGA was discontinued, and the patient continued topical therapy as needed for limited residual skin disease. The patient's blood Sezary remained clear and his skin near CR off MOGA, but after several months, he experienced disease progression in the skin with rapidly worsening, widespread thick plaques and nodular, tumor-type disease. Skin biopsy showed LCT with 10-40% of neoplastic T cells expressing CD30 by immunohistochemistry. PET/CT now showed more notable LAD, and a core needle sampling showed involvement with T-cell lymphoma (CD30 expression of 30%). Given the significant CD30 expression in the skin and LN compartments, BV was selected as the next therapy.

Management of LCT in MF and SS

LCT in MF or SS is recognized as one of the strongest independent adverse prognostic indicators,^{32,33} and thus knowledge of LCT is a key factor in treatment selection (Table 1). However, notable inter-rater (pathologist to pathologist) variability is observed in the interpretation of LCT as the criteria is an arbitrary cutoff value (>25% of malignant T cells are large cells) by histopathologic evaluation.³⁴ Accordingly, a subset of patients designated as LCT may have a more indolent course; therefore, appropriate clinical judgment is essential for optimal management. In contrast to systemic lymphomas in which anthracycline-based therapy and autologous transplants are considered in the setting of transformed disease, LCT in MF/SS is managed differently, with greater individualized tailoring for the extent and biology of the transformed disease.

CD30 expression is not a requirement for LCT criteria, but the median CD30 expression is higher in the lesional tissue of patients with LCT compared to those without LCT,³⁵ and thus BV is often a highly effective treatment in MF with LCT.^{10,12,36} In the phase 3 trial of BV in CD30⁺ CTCL, the primary end point of an objective response rate lasting >4 months (ORR4) was higher in patients with LCT than without LCT (65% vs 39%).^{11,12} Pralatrexate and romidepsin, each as a single agent, have also demonstrated clinical activity in MF or SS with LCT,^{14,37,38} though the data in these single agents are not as robust as in the pivotal trial of BV. MOGA would not be prioritized in patients with LCT given the lack of established data.¹³ Older agents such as liposomal doxorubicin or gemcitabine have greater toxicity profiles but can be used in MF/SS with LCT in refractory settings or when newer agents are not available.^{39,40} A recent clinical trial of pembrolizumab in patients with relapsed/refractory MF or SS showed promising clinical activity in patients with LCT and can be considered in relapsed or refractory settings.⁴¹

Biomarker-guided selection and integration of therapies in MF/SS-CTCL

Currently, numerous available and investigational therapies in MF/SS-CTCL can target the cell surface molecules of malignant T cells, disrupted cellular pathways, and/or the tumor microenvironment (TME), including the immune milieu (Table 1). Reliable biomarkers that enrich the patient subsets that can benefit with specific therapies would be ideal, and such biomarker information may help select and prioritize therapies among a list of options.

Of the targets on the malignant cell surface in CTCL, CD30 has the most robust data, with a CD30 targeting agent, BV. With the use of more sensitive tools for CD30 detection (eg, multispectral imaging), samples with nondetectable CD30 expression by routine immunohistochemistry (IHC) and light microscopy demonstrated cell-surface CD30 molecules.¹⁰ In MF/SS, the inter- and intrapatient (interlesional) CD30 expression levels (by routine IHC) can be highly variable in the skin, and BV has demonstrated significant clinical activity in patients with a spectrum of CD30 expression in 2 independent studies in MF/SS.^{10,36,42} Patients with >5% CD30_{max} (maximum CD30 level from multiple skin samples) had more reliable overall clinical response compared with those with CD30 ____ <5%; however, meaningful activity was observed across all CD30 levels. Further, in the pivotal study of BV in CD30⁺ CTCL (ALCANZA), patients with multiple skin biopsies showed a notable variability of CD30 levels, ranging from nondetectable to >50% expression.^{11,12} Thus, BV is a treatment option even in those with low/negligible levels of CD30 expression, especially in refractory settings.

Therapies such as MOGA can target the malignant cells as well as TME, as the target molecule *CCR4* is expressed not only by the malignant T cells but also by the regulatory T cells. *CCR4* is a chemokine receptor associated with skin trafficking, but unlike CD30, the molecule is frequently and consistently expressed in the lesional skin (median *CCR4*, 80%; range of 1%-100% by IHC) and may not be useful as a biomarker in CTCL.¹³ However, some patients with MF/SS have negligible *CCR4* expression, and further studies will be needed to establish the role of MOGA in these patients. Further, assessment for *CCR4* expression is not routinely available, so a standardized protocol for *CCR4* assessment and a better understanding of its correlation with clinical outcome needs to be established.⁴³ Although both *CCR4* and *KIR3DL2* are both commonly expressed in skin disease with LCT,

neither MOGA nor anti-*KIR3DL2* antibody (lacutamab) has established activity data in patients with LCT.^{25,44}

Newer biomarker platforms (eg, actionable NGS panels, TME profiling) may help guide treatment selection and prioritize therapies in selected settings.^{45,46} The knowledge of Th2 skewing with disease progression has led to the selection of immune therapies that shift the immune profile from Th2 to Th1, including interferons, IL-12, and toll-like receptor agonists. Agents known to deplete regulatory T cells in the TME, such as MOGA or CD25-targeting agents,⁴⁷ and to activate macrophages (increase the "eat-me" signal), such as agents targeting the CD47-SIRPa axis (eg, SIRPaFc, magrolimab),^{48,49} may be considered as single agents or part of combination regimens. Patients with genomic data that demonstrate pathogenic alteration in the T-cell immune regulatory axis may benefit from immune therapies that include checkpoint inhibitors.⁴¹

In patients with refractory or high-risk disease, using NGS platforms to evaluate for additional prognostic clues or actionable pathogenic genetic variants can help prioritize therapies. For example, *JAK* inhibitors may be considered in patients with relapsed/refractory disease with a pathogenic mutation of *JAK1/3*.⁵⁰ Moreover, we can extrapolate genomic biomarker data from other T-cell lymphomas. For example, *CCR4* gain of function mutations have been associated with superior clinical outcomes in adult T-cell leukemia/lymphoma patients treated with MOGA.⁵¹ Thus, if such potential pathogenic variants of *CCR4* are noted by NGS data, perhaps MOGA can be prioritized over other therapies in high-burden Sezary patients or in refractory disease.

As previously discussed, LCT is a very important clinical and pathologic factor to consider when selecting therapy. An NGS study by Ungewickell et al reported that LCT is associated with a high mutation burden and recurrent *PLCG1* alterations.⁵² Furthermore, recent studies exploring the programmed cell death 1 (*PD-1*) and programmed cell death 1 ligand 1 (*PD-L1*) axis show a possible link between *PD-L1* structural variants and LCT,⁴⁵ supporting the hypothesis that patients with LCT may have more TME immune alterations that allow tumor growth. These patients with *PD-L1* alteration have experienced clinical benefit with anti-*PD-1* antibody such as pembrolizumab.^{41,53}

CLINICAL CASE (Continued)

The patient was treated with BV, with great clinical response in the skin and flattening of his thicker, tumor-type skin disease that showed LCT by pathology. PET/CT obtained after 3 cycles of BV showed clinical CR of LN disease. The blood compartment remained clear of measurable disease by flow cytometry. The patient had clinical features consistent with a high-risk prognostic profile, including a history of high blood SC burden, LN disease, and LCT.³³ At the initiation of BV, allogeneic HSCT was discussed as a potential curative therapy in this high-risk setting. The patient's sibling was identified as a 10/10 matched donor. The patient was 67 years old at this time point, and he agreed to move forth with allogeneic HSCT using TSEBT and a total lymphoid irradiation/antithymocyte globulin nonmyeloablative regimen.

Reference	N	Intervention	Efficacy	Transplant-related toxicities
Hosing et al ⁵⁵	47	6% ablative 94% RIC/NMA	ORR = n/a, 2-year PFS 40%, 2-year OS 67% 4-year PFS 26%, 4-year OS 51%	1-year NRM 10% 2-year NRM 17% Gr 2–4 aGVHD 40%; cGVHD 28%
Duarte et al ⁵⁶	60	33% ablative 67% RIC/NMA	ORR = n/a, 2-year PFS 34%, 2-year OS 54% 5-year PFS 32%, 5-year OS 46% 7-year PFS 30%, 7-year OS 44%	1-year NRM 20% 7-year NRM 22%
Paralkar et al57	12	17% ablative 83% RIC	ORR = 67%, 2-year PFS 23%, 2-year OS 56%	1-year NRM 25%
De Masson et al ⁵⁸	37	32% ablative 68% RIC/NMA	ORR = n/a, 2-year PFS 31%, 2-year OS 57%	1-year NRM 18% 2-year NRM 18% Gr 2–4 aGVHD 76%; cGVHD 44%
Lechowicz et al ⁵⁹	129	36% ablative 64% RIC/NMA	ORR = n/a, 1-year PFS 31%, 1-year OS 54% 5-year PFS 17%, 5-year OS 32%	1-year NRM 19% 5-year NRM 22% Gr 2–4 aGVHD 41%; cGVHD 439
Isufi et al ⁶⁰	16 (n = 23; 16 MF/SS; 7 G/D TCL)	RIC except 2 haploidentical	CR rate = 56% OS 75% (12 of 16 patients) w/median fol- low-up 5.5 years	100-day NRM 12% Gr 2–4 aGVHD 50%; cGVHD 56%
Weng et al ⁶¹	35	100% NMA	ORR (CR) = 80% (57%), 2-year PFS 60%, 2-year OS 68% 5-year PFS 41%, 3-year OS 62%, 5-year OS 56%	1-year NRM 3% 2-year NRM 14% Gr 2–4 aGVHD 16%; cGVHD 32%

Table 2. Allogeneic HSCT in MF/SS-CTCL

aGVHD, acute GVHD; cGVHD, chronic GVHD; Gr, grade; NMA, non-myeloablative; RIC, reduced-intensity conditioning.

Allogeneic HSCT as a path to establish durable remission in high-risk MF/SS

Patients with advanced-stage MF and SS have a median survival of <5 years, are often treated with multiple sequential systemic therapies, and eventually become refractory to available agents.³³ High-dose chemotherapy followed by autologous HSCT has not shown a durable benefit,⁵⁴ but allogeneic HSCT that relies on a graft vs lymphoma effect has led to long-term remissions (Table 2).⁵⁵⁻⁶¹

There is some controversy regarding which subset of patients should move forth for allogeneic transplant in MF/SS and when in their disease course it is appropriate. Allogeneic HSCT should be considered for appropriate patients with stage IIB through IV disease who are refractory to multiple lines of systemic therapy, demonstrate declining duration of clinical benefit, or present with a very high-risk profile. A large international study of prognostic factors in advanced-stage MF and SS (retro-CLIPI) identified 3 risk groups with distinct prognostic outcomes based on the number of key adverse prognostic parameters, including extracutaneous disease and LCT.³³ Patients in the high-risk group with a 5-year overall survival (OS) of 28% were clearly appropriate for allogeneic HSCT. A significant proportion of patients in the intermediate-risk (5-year OS, 44%) group may be considered for allogeneic HSCT. The 5-year OS with allogeneic HSCT in the published studies ranged from 32% to 56%, 55-62 likely supporting an improved survival benefit provided that most patients who received allogeneic HSCT included those with intermediate- to high-risk retro-CLIPI profiles. The optimal timing for allogeneic HSCT is when the disease is well controlled (CR or near CR of all disease compartments) and before the disease has progressed to a state in which the chance of response or survival with allogeneic HSCT is low. The skin compartment is a frequent site of posttransplant disease relapse, and thus TSEBT is often an important and necessary component of the preparatory strategy in MF/SS. The median age of patients with advanced MF or SS at diagnosis is 60 to 65 years, so an allogeneic regimen with fewer transplant-related toxicities has also been explored.⁶¹ For each patient, a decision to transplant requires careful consideration of the long-term benefits and significant risks, including the potential for graft-versus-host disease (GVHD) and disease relapse after transplant.

In patients with aggressive MF or SS, there remains an unmet need for additional safe and effective therapies that can bridge patients to allogeneic HSCT. And especially for those who are not eligible for allogeneic HSCT or have no optimal donors, new and novel immune therapies and approaches are needed. As these innovative therapies are made available, it is essential to integrate them to manage patients successfully and to continue exploring biomarkers that can help us guide our selection or prioritization of treatment options.

Conclusions and future directions

The clinical, pathological, and biological features in MF and SS are highly heterogeneous, with significant inter- and intra-patient variability that requires a personalized approach to management. Available and investigative therapies often have differential activity across disease compartments, and thus in addition to clinical stage and the presence of LCT, compartmental activity is an important element that guides treatment selection. In appropriate high-risk settings, patients should be considered for allogeneic HSCT. The unique QoL issues bring unique challenges in managing patients with CTCL.

Ultimately, in this highly heterogeneous disease group we hope to establish biomarkers and strategies that enrich (better align subsets) for durable clinical response and address resistance/escape mechanisms while minimizing toxicities. More personalized and tolerable immunotherapies may be a pathway to achieve a more durable or sustained clinical response. Advances in biomarkerguided investigations in CTCL will enable us to apply evidencebased approaches in prioritizing therapies and help develop combination strategies that may lead to the clinical synergy of combined agents.

Conflict-of-interest disclosure

Youn H. Kim: Advisory board, Research funding; Innate: Research funding; Corvus: Research funding; Galderma: Advisory board, Research funding; CRISPR therapeutics: Research funding; Secura Bio: Advisory board; Trillium: Research funding.

Off-label drug use

Youn H. Kim: none.

Correspondence

Youn H. Kim, Stanford University School of Medicine, 780 Welch Rd, CJ220D, C. J. Huang Bldg, Palo Alto, CA 94304; e-mail: younkim@stanford.edu.

References

- Willemze R, Cerroni L, Kempf W, et al. The 2018 update of the WHO-EORTC classification for primary cutaneous lymphomas. *Blood*. 2019;133(16):1703-1714.
- Kim EJ, Hess S, Richardson SK, et al. Immunopathogenesis and therapy of cutaneous T cell lymphoma. J Clin Invest. 2005;115(4):798-812.
- Bobrowicz M, Fassnacht C, Ignatova D, Chang YT, Dimitriou F, Guenova E. Pathogenesis and therapy of primary cutaneous T-cell lymphoma: Collegium Internationale Allergologicum (CIA) update 2020. Int Arch Allergy Immunol. 2020;181(10):733-745.
- Olsen EA, Whittaker S, Kim YH, et al. Clinical end points and response criteria in mycosis fungoides and Sezary syndrome: a consensus statement of the International Society for Cutaneous Lymphomas, the United States Cutaneous Lymphoma Consortium, and the Cutaneous Lymphoma Task Force of the European Organisation for Research and Treatment of Cancer. J Clin Oncol. 2011;29(18):2598-2607.
- Kim YH, Demierre MF, Kim EJ, et al. Clinically meaningful reduction in pruritus in patients with cutaneous T-cell lymphoma treated with romidepsin. *Leuk Lymphoma*. 2013;54(2):284-289.
- Dummer R, Prince HM, Whittaker S, et al. Patient-reported quality of life in patients with relapsed/refractory cutaneous T-cell lymphoma: results from the randomised phase III ALCANZA study. *Eur J Cancer*. 2020;133(July):120-130.
- Porcu P, Hudgens S, Horwitz S, et al. Quality of life effect of the anti-CCR4 monoclonal antibody mogamulizumab versus vorinostat in patients with cutaneous T-cell lymphoma. *Clin Lymphoma Myeloma Leuk*. 2021;21(2):97-105.
- Mehta-Shah N, Horwitz SM, Ansell S, et al. NCCN guidelines insights: primary cutaneous lymphomas, version 2.2020. J Natl Compr Canc Netw. 2020;18(5):522-536.
- Hoppe RT, Harrison C, Tavallaee M, et al. Low-dose total skin electron beam therapy as an effective modality to reduce disease burden in patients with mycosis fungoides: results of a pooled analysis from 3 phase-II clinical trials. J Am Acad Dermatol. 2015;72(2):286-292.
- Kim YH, Tavallaee M, Sundram U, et al. Phase II investigator-initiated study of brentuximab vedotin in mycosis fungoides and Sézary syndrome with variable CD30 expression level: a multi-institution collaborative project. *J Clin Oncol.* 2015;33(32):3750-3758.

- Prince HM, Kim YH, Horwitz SM, et al; ALCANZA Study Group. Brentuximab vedotin or physician's choice in CD30-positive cutaneous T-cell lymphoma (ALCANZA): an international, open-label, randomised, phase 3, multicentre trial. Lancet. 2017;390(10094):555-566.
- Kim YH, Prince HM, Whittaker S, et al. Response to brentuximab vedotin versus physician's choice by CD30 expression and large cell transformation status in patients with mycosis fungoides: an ALCANZA sub-analysis. *Eur J Cancer.* 2021;148(May):411-421.
- Kim YH, Bagot M, Pinter-Brown L, et al; MAVORIC Investigators. Mogamulizumab versus vorinostat in previously treated cutaneous T-cell lymphoma (MAVORIC): an international, open-label, randomised, controlled phase 3 trial. Lancet Oncol. 2018;19(9):1192-1204.
- Whittaker SJ, Demierre MF, Kim EJ, et al. Final results from a multicenter, international, pivotal study of romidepsin in refractory cutaneous T-cell lymphoma. J Clin Oncol. 2010;28(29):4485-4491.
- Jothishankar B, Almazan T, Kim Y, et al. Romidepsin and total skin electron beam therapy in advanced stage mycosis fungoides and Sézary syndrome. *Br J Haematol.* 2019;186(2):377-379.
- Fong S, Hong EK, Khodadoust MS, et al. Low-dose total skin electron beam therapy combined with mogamulizumab for refractory mycosis fungoides and Sézary syndrome. Adv Radiat Oncol. 2021;6(3):100629.
- 17. Ahern K, Gilmore ES, Poligone B. Pruritus in cutaneous T-cell lymphoma: a review. J Am Acad Dermatol. 2012;67(4):760-768.
- Lindahl LM, Willerslev-Olsen A, Gjerdrum LMR, et al. Antibiotics inhibit tumor and disease activity in cutaneous T-cell lymphoma. *Blood*. 2019;134(13):1072-1083.
- Demierre MF, Kim YH, Zackheim HS. Prognosis, clinical outcomes and quality of life issues in cutaneous T-cell lymphoma. *Hematol Oncol Clin North Am*. 2003;17(6):1485-1507.
- Jonak C, Porkert S, Oerlemans S, et al. Health-related quality of life in cutaneous lymphomas: past, present and future. Acta Derm Venereol. 2019;99(7):640-646.
- Campbell BA, Scarisbrick JJ, Kim YH, Wilcox RA, McCormack C, Prince HM. Time to next treatment as a meaningful endpoint for trials of primary cutaneous lymphoma. *Cancers (Basel)*. 2020;12(8):2311.
- 22. Duvic M, Hymes K, Heald P, et al; Bexarotene Worldwide Study Group. Bexarotene is effective and safe for treatment of refractory advanced-stage cutaneous T-cell lymphoma: multinational phase II-III trial results. J Clin Oncol. 2001;19(9):2456-2471.
- Zackheim HS, Epstein EH Jr. Low-dose methotrexate for the Sézary syndrome. J Am Acad Dermatol. 1989;21(suppl 4, pt 1):757-762.
- 24. Quaglino P, Knobler R, Fierro MT, et al. Extracorporeal photopheresis for the treatment of erythrodermic cutaneous T-cell lymphoma: a single center clinical experience with long-term follow-up data and a brief overview of the literature. *Int J Dermatol.* 2013;52(11):1308-1318.
- Bagot M, Porcu P, Marie-Cardine A, et al. IPH4102, a first-in-class anti-KIR3DL2 monoclonal antibody, in patients with relapsed or refractory cutaneous T-cell lymphoma: an international, first-in-human, open-label, phase 1 trial. Lancet Oncol. 2019;20(8):1160-1170.
- Corbin ZA, Nguyen-Lin A, Li S, et al. Characterization of the peripheral neuropathy associated with brentuximab vedotin treatment of mycosis fungoides and Sézary syndrome. J Neurooncol. 2017;132(3):439-446.
- Khan N, Noor S, Geller S, et al. A phase II trial of reduced dose brentuximab vedotin for cutaneous T-cell lymphoma. Paper presented at: International Conference on Malignant Lymphoma; June 2021; Lugano, Switzerland.
- Bagot M, Dalle S, Sokol L, et al. Long-term clinical benefit to anti-CCR4 mogamulizumab: results from the phase 3 MAVORIC study in previously treated cutaneous T-cell lymphoma. *Blood*. 2018;132(suppl 1):2901.
- 29. Musiek A, Whittaker S, Horwitz SM, et al. Characterization and outcomes in patients with mogamulizumab-associated skin reactions in the MAVORIC trial. *Blood.* 2020;136(suppl 1):23-24.
- Hirotsu KE, Neal TM, Khodadoust MS, et al. Clinical characterization of mogamulizumab-associated rash during treatment of mycosis fungoides or Sézary syndrome. JAMA Dermatol. 2021;157(6):700-707.
- Martinez-Escala ME, Kuzel TM, Kaplan JB, et al. Durable responses with maintenance dose-sparing regimens of romidepsin in cutaneous T-cell lymphoma. JAMA Oncol. 2016;2(6):790-793.
- Arulogun SO, Prince HM, Ng J, et al. Long-term outcomes of patients with advanced-stage cutaneous T-cell lymphoma and large cell transformation. *Blood.* 2008;112(8):3082-3087.
- 33. Scarisbrick JJ, Prince HM, Vermeer MH, et al. Cutaneous Lymphoma International Consortium study of outcome in advanced stages of mycosis fungoides and Sézary syndrome: effect of specific prognostic markers on survival

and development of a prognostic model. J Clin Oncol. 2015;33(32):3766-3773.

- 34. Gru AA, Kim J, Pulitzer M, et al. The use of central pathology review with digital slide scanning in advanced-stage mycosis fungoides and Sézary syndrome: a multi-institutional and international pathology study. Am J Surg Pathol. 2018;42(6):726-734.
- Edinger JT, Clark BZ, Pucevich BE, Geskin LJ, Swerdlow SH. CD30 expression and proliferative fraction in nontransformed mycosis fungoides. *Am J Surg Pathol.* 2009;33(12):1860-1868.
- 36. Duvic M, Tetzlaff MT, Gangar P, Clos AL, Sui D, Talpur R. Results of a phase II trial of brentuximab vedotin for CD30+ cutaneous T-cell lymphoma and lymphomatoid papulosis. J Clin Oncol. 2015;33(32):3759-3765.
- Horwitz SM, Kim YH, Foss F, et al. Identification of an active, well-tolerated dose of pralatrexate in patients with relapsed or refractory cutaneous T-cell lymphoma. *Blood.* 2012;119(18):4115-4122.
- O'Connor OA, Pro B, Pinter-Brown L, et al. Pralatrexate in patients with relapsed or refractory peripheral T-cell lymphoma: results from the pivotal PROPEL study. J Clin Oncol. 2011;29(9):1182-1189.
- Dummer R, Quaglino P, Becker JC, et al. Prospective international multicenter phase II trial of intravenous pegylated liposomal doxorubicin monochemotherapy in patients with stage IIB, IVA, or IVB advanced mycosis fungoides: final results from EORTC 21012. J Clin Oncol. 2012;30(33):4091-4097.
- 40. Duvic M, Talpur R, Wen S, Kurzrock R, David CL, Apisarnthanarax N. Phase II evaluation of gemcitabine monotherapy for cutaneous T-cell lymphoma. *Clin Lymphoma Myeloma*. 2006;7(1):51-58.
- Khodadoust MS, Rook AH, Porcu P, et al. Pembrolizumab in relapsed and refractory mycosis fungoides and Sézary syndrome: a multicenter phase II study. J Clin Oncol. 2020;38(1):20-28.
- Rahbar Z, Li S, Tavallaee M, Novoa RA, Kim J, Kim YH. Variability in the expression of immunohistochemical markers: implications for biomarker interpretation in cutaneous T-cell lymphoma. J Invest Dermatol. 2018;138(5): 1204-1206.
- Fujii K, Sakamoto Y, Masaki A, et al. Immunohistochemistry for CCR4 C-terminus predicts CCR4 mutations and mogamulizumab efficacy in adultT-cell leukemia/ lymphoma. J Pathol Clin Res. 2021;7(1):52-60.
- 44. Duvic M, Pinter-Brown LC, Foss FM, et al. Phase ¹/₂ study of mogamulizumab, a defucosylated anti-CCR4 antibody, in previously treated patients with cutaneous T-cell lymphoma. *Blood*. 2015;125(12):1883-1889.
- Beygi S, Fernandez-Pol S, Duran G, et al. Pembrolizumab in mycosis fungoides with PD-L1 structural variants. *Blood Adv.* 2021;5(3):771-774.
- Houlahan KE, Curtis C. A tumor "personality" test to guide therapeutic decision making. Cancer Cell. 2021;39(6):747-749.
- Kawai H, Ando K, Maruyama D, et al. Phase II study of E7777 in Japanese patients with relapsed/refractory peripheral and cutaneous T-cell lymphoma. *Cancer Sci.* 2021;112(6):2426-2435.
- Folkes AS, Feng M, Zain JM, Abdulla F, Rosen ST, Querfeld C. Targeting CD47 as a cancer therapeutic strategy: the cutaneous T-cell lymphoma experience. Curr Opin Oncol. 2018;30(5):332-337.
- Johnson LDS, Banerjee S, Kruglov O, et al. Targeting CD47 in Sezary syndrome with SIRPalphaFc. *Blood Adv*. 2019;3(7):1145-1153.
- 50. Gomez-Arteaga A, Margolskee E, Wei MT, van Besien K, Inghirami G, Horwitz S. Combined use of tofacitinib (pan-JAK inhibitor) and ruxolitinib (a JAK1/2 inhibitor) for refractory T-cell prolymphocytic leukemia (T-PLL) with a JAK3 mutation. *Leuk Lymphoma*. 2019;60(7):1626-1631.
- Sakamoto Y, Ishida T, Masaki A, et al. CCR4 mutations associated with superior outcome of adult T-cell leukemia/lymphoma under mogamulizumab treatment. *Blood*. 2018;132(7):758-761.
- Ungewickell A, Bhaduri A, Rios E, et al. Genomic analysis of mycosis fungoides and Sézary syndrome identifies recurrent alterations in TNFR2. Nat Genet. 2015;47(9):1056-1060.
- 53. Lesokhin AM, Ansell SM, Armand P, et al. Nivolumab in patients with relapsed or refractory hematologic malignancy: preliminary results of a phase Ib study. J Clin Oncol. 2016;34(23):2698-2704.
- 54. Wu PA, Kim YH, Lavori PW, Hoppe RT, Stockerl-Goldstein KE. A metaanalysis of patients receiving allogeneic or autologous hematopoietic stem cell transplant in mycosis fungoides and Sézary syndrome. *Biol Blood Marrow Transplant*. 2009;15(8):982-990.
- Hosing C, Bassett R, Dabaja B, et al. Allogeneic stem-cell transplantation in patients with cutaneous lymphoma: updated results from a single institution. Ann Oncol. 2015;26(12):2490-2495.
- 56. Duarte RF, Boumendil A, Onida F, et al. Long-term outcome of allogeneic hematopoietic cell transplantation for patients with mycosis fungoides

and Sézary syndrome: a European Society for Blood and Marrow Transplantation Lymphoma Working Party extended analysis. *J Clin Oncol.* 2014;32(29):3347-3348.

- Paralkar VR, Nasta SD, Morrissey K, et al. Allogeneic hematopoietic SCT for primary cutaneous T cell lymphomas. Bone Marrow Transplant. 2012;47(7):940-945.
- 58. de Masson A, Beylot-Barry M, Bouaziz JD, et al; French Study Group on Cutaneous Lymphomas and Société Française de Greffe de Moëlle et Thérapie Cellulaire. Allogeneic stem cell transplantation for advanced cutaneous T-cell lymphomas: a study from the French Society of Bone Marrow Transplantation and French Study Group on Cutaneous Lymphomas. *Haematologica*. 2014;99(3):527-534.
- Lechowicz MJ, Lazarus HM, Carreras J, et al. Allogeneic hematopoietic cell transplantation for mycosis fungoides and Sezary syndrome. *Bone Marrow Transplant*. 2014;49(11):1360-1365.
- 60. Isufi I, Seropian S, Gowda L, et al. Outcomes for allogeneic stem cell transplantation in refractory mycosis fungoides and primary cutaneous gamma delta T cell lymphomas. *Leuk Lymphoma*. 2020;61(12):2955-2961.
- Weng WK, Arai S, Rezvani A, et al. Nonmyeloablative allogeneic transplantation achieves clinical and molecular remission in cutaneous T-cell lymphoma. *Blood Adv.* 2020;4(18):4474-4482.

- 62. Mehta-Shah N, Kommalapati A, Teja S, et al. Successful treatment of mature T-cell lymphoma with allogeneic stem cell transplantation: the largest multicenter retrospective analysis. Paper presented at: 62nd American Society of Hematology Annual Meeting; 5-8 December 2020; virtual meeting.
- 63. Olsen EA, Kim YH, Kuzel TM, et al. Phase IIb multicenter trial of vorinostat in patients with persistent, progressive, or treatment refractory cutaneous T-cell lymphoma. J Clin Oncol. 2007;25(21):3109-3115.
- 64. Horwitz SM, Koch R, Porcu P, et al. Activity of the PI3K-δ_iγ inhibitor duvelisib in a phase 1 trial and preclinical models of T-cell lymphoma. *Blood*. 2018;131(8):888-898.
- 65. Querfeld C, Rosen ST, Guitart J, et al. Results of an open-label multicenter phase 2 trial of lenalidomide monotherapy in refractory mycosis fungoides and Sezary syndrome. *Blood*. 2014;123(8):1159-1166.

© 2021 by The American Society of Hematology DOI 10.1182/hematology.2021000263