LETTER TO EDITOR

T_{FH}2 cells associate with enhanced humoral immunity to **SARS-CoV-2 inactivated vaccine in patients with allergic rhinitis**

To the Editor:

We report that allergic rhinitis (AR) patients displayed enhanced humoral immune response to SARS-CoV-2 inactivated vaccine as compared to healthy controls, and highlight a pivotal role of type 2 T follicular helper cells in this response. Vaccination is considered as the best strategy to reduce SARS-CoV-2 infections and prevent severe illness after breakthrough infections.^{1,2} Although SARS-CoV-2 vaccines are highly protective against COVID-19 in healthy individuals, the vaccine efficacy may be compromised in patients with pre-existing conditions, resulting in dysregulated immune systems. Recent data described poorer humoral immunity to COVID-19 vaccines in individuals with kidney transplantation, undertaking dialysis, or cancer, compared to healthy controls.^{1,2} Allergic diseases, including AR, atopic dermatitis and asthma, are caused by the immune system hypersensitivity to innocuous environmental antigens and characterized by skewed type 2 immune responses. 3 Such chronic immunological disorders are highly prevalent and estimated to affect up to 50% of the world population.³ Allergic airway diseases have been revealed to have a potential protective role in COVID-19, possibly because of the reduction of SARS-CoV-2 receptor angiotensin-converting enzyme 2 in airway epithelial cells caused by type 2 cytokines and the abundant infiltration of eosinophils in airways.⁴ However, there are no studies formally investigating whether allergic diseases modulate the humoral response following vaccination against SARS-CoV-2. In the present study, we compare immunological response after two-dose inactivated SARS-CoV-2 immunization between healthy subjects and patients with AR.

We conducted a prospective study to evaluate the potential impact of AR on the immune response after two-dose inactivated SARS-CoV-2 immunization (Figure [1A\)](#page-2-0). The trial was registered at <https://clinicaltrials.gov> (NCT05009134). Twenty-five healthy adults and 32 patients

with AR were recruited at Tongji Hospital (Table [1\)](#page-1-0). AR was diagnosed based on the concordance between typical allergic symptoms and atopic status. 5 Atopic status was assessed by skin-prick testing (SPT) and/or allergenspecific immunoglobulin E (IgE) levels. 6 AR patients had at least 1-year disease history and had never been infected with SARS-CoV-2. The exclusion criteria included (i) the presence of sinusitis, (ii) pregnancy or breastfeeding, (iii) with cardiovascular diseases, severe immunologic diseases, chronic obstructive pulmonary disease, chronic infections, diabetes, tumours, chronic kidney diseases or stroke, and (iv) use of intranasal steroid or antihistamines in the previous 1 week or oral steroids in the previous 3 months before this study. Healthy subjects had negative SPT and specific IgE test and had no history of allergy.⁶ All participants received two doses of inactivated SARS-CoV-2 vaccines (WIBP-CorV, Sinopharm, Wuhan) with 1 month apart. Peripheral blood samples were collected on days 0, 7, 30 (before the second dose), 37 (7 days post the second dose) and 60 (30 days post the second dose) to analyze humoral immune responses to vaccination (Figure [1A\)](#page-2-0). None was lost to follow-up. In addition, a complementary approach was adopted to further investigate the influence of AR on the antibody response to natural SARS-CoV-2 infection in 78 patients recovered from COVID-19 (64 control subjects without AR and 14 AR patients), whose peripheral bloods were collected at 10–12 months post the infection. The Ethics Committee of Tongji Hospital reviewed and approved this trial, and informed consent was provided by every participant. More information regarding subjects and methods is provided in Tables S1–S3.

There was no noticeable difference in the incidence of adverse reactions between healthy subjects and patients with AR (Table S1). Protective antibody responses to WIBP-CorV were assessed by the detection of neutralizing antibodies targeting the receptor-binding domain (RBD) of SARS-CoV-2 spike (S) 1 protein and IgG and IgM against

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Sensitization p

TABLE 1 Demographic characteristics of subjects in the study of inactivated SARS-CoV-2 vaccination

 $Cockroad$ and $2(6.4%)$ Cat dander $\begin{array}{ccc} 5 (15.6\%) \end{array}$ Dog dander and the control of the control Artemisia 0 1 (3.1%) Platanus 0 1 (3.1%) Alternaria 0 3 (9.4%)

VAS symptom score $0(0, 0)$ 5.0 (3.3, 7.0) <.001

Note: For continuous variables, data are expressed as medians and interquartile ranges.

Abbreviations: AR, allergic rhinitis; VAS, visual analogue scale.

the S and nucleocapsid proteins. The inactivated SARS-CoV-2 vaccine elicited robust serological responses, with gradual increase in neutralizing antibody, IgG and IgM following vaccination (Figure [1B\)](#page-2-0). Although neutralizing antibody, IgG and IgM levels were low following the first immunization, the second injection boosted their titres by [∼]35-, 161- and 16-fold (mean) at day 60 relative to day 0 (baseline), respectively (Figure S1A). As reported in COVID-19 infection or vaccination, $\frac{7}{1}$ $\frac{7}{1}$ $\frac{7}{1}$ the levels of neutralizing antibody strongly correlated with IgG levels (Figure S1B).

Of a particular interest, serological responses in AR patients were stronger than those in healthy controls, showing higher levels of neutralizing antibody at days 30, 37 and 60, and anit-SARS-CoV-2 IgG on days 37 and 60, and anit-SARS-CoV-2 IgM on day 60 (Figure [1C;](#page-2-0) Figure S1C). Higher frequencies of CD19+CD27+CD38++ circulating plasmablasts were found in patients with AR at day 60, although there was no noticeable difference in CD3–CD19⁺ total B cells and its subsets (IgD–CD27– double negative, IgD⁻CD27⁺ switched memory, IgD⁺CD27⁻ naive and $IgD^{+}CD27^{+}$ nonswitched memory B cells) between healthy subjects and AR patients (Figures S2 and S3). To explore the immune memory after vaccination, we also analyzed memory B-cell response specific to RBD in the peripheral blood (Figure S2). Flow cytometric analysis revealed higher frequencies of RBD-specific memory B cells, again, in AR patients compared to healthy subjects at day 60 (Figure [1D\)](#page-2-0). As the negative controls, RBDspecific memory B cells were undetectable in samples collected before vaccination (Figure [1D\)](#page-2-0). These results suggest AR patients demonstrated enhanced immune responses to inactive SARS-CoV-2 vaccine than control subjects.

T follicular helper (T_{FH}) cells essentially support B cells for the generation of neutralizing antibodies and longlived humoral immunity after vaccination.⁸ We assessed vaccination-induced T_{FH} activation by examining circulating $CXCR5+ICOS^{high}PD-1^{high} T_{FH} (ICOS^{high}PD-1^{high})$ cT_{FH}) cells (Figure S4).⁶ In both groups, inactivated SARS-CoV-2 vaccination induced a modest expansion of $ICOS^{high}PD-1^{high} cT_{FH} cells, 7 days after the first immu$ nization and a drastic expansion 7 days after the second immunization (day 37), while there was no significant difference in the frequencies of ICOShighPD-1high cT_{FH} cells at all the time points between the two groups (Figure [2A\)](#page-3-0). To reduce the impact introduced by individual variation, we normalized the frequencies of ICOS^{high}PD-1^{high} cT_{FH} cells to the baseline at day 0 and detected higher frequencies of ICOShighPD-1high cT_{FH} cells at days 7, 37 and 60 in AR patients than control subjects (Figure [2B\)](#page-3-0). In contrast, total CD4⁺ T cells, CXCR5⁺ total cT_{FH} cells, T_{REG} cells and T_{FR} cells were not affected by vaccination and remained comparable between two groups (Figure S5A,B). Therefore, both AR patients and control subjects mounted potent T_{FH} activation by two-dose of WIBP-CorV vaccination, with a stronger signature by the former group.

As both T_{FH} activation and the production neutralizing antibody were strongly elicited after the second dose of vaccine, we focused on the memory response after the second dose in order to understand the relationship between

FIGURE 1 Enhanced antibody responses in patients with allergic rhinitis (AR) following inactivated SARS-CoV-2 vaccination. (A) Healthy subjects ($n = 25$) and patients with AR ($n = 32$) were enrolled and received inactivated SARS-CoV-2 vaccine at days 0 and 30. Peripheral blood was collected at days 0 (baseline), 7, 30, 37 and 60. (B) Plasma neutralizing antibodies against the RBD of the SARS-CoV-2 S1 protein and IgG and IgM against the SARS-CoV-2 S and N proteins were measured by chemiluminescent immunoassay. Data are analyzed by two-sided paired-sample *t*-test. (C) Fold changes of SARS-CoV-2 neutralizing antibody, IgG and IgM in control subjects and AR patients by normalizing to the baseline levels. (D) Frequencies of circulating RBD-specific memory B cells at day 60 were analyzed by flow cytometry. Numbers indicate the frequency of cells in the gated region. For (C) and (D), data are presented as median and interquartile range and analyzed by Mann–Whitney *U*-test

 T_{FH} function and protective antibody response. In patients with AR, the fold changes of ICOS^{high}PD-1^{high} cT_{FH} at day 37 positively correlated with the fold changes of IgG and IgM at day 60 but, intriguingly, not the fold changes of neutralizing antibody, which are more important in evaluating protective humoral immunity (Figure [2C;](#page-3-0) Figure S5C). Based on CXCR3 and CCR6 expression, cT_{FH} cells can be classified into three major subsets, referred as $T_{FH}1$, T_{FH} 2 and T_{FH} 17 cells.⁶ T_{FH} 2 and T_{FH} 17 cells are reported to support chronic humoral immunity, such as autoantibody production in autoimmune diseases, whereas $T_{FH}1$

cells can support antibody response in infection and vaccine with relatively short duration.⁷ With ICOS^{high}PD-1^{high} cT_{FH} cells, the T_{FH}1 and T_{FH}2, but not T_{FH}17 subsets, were strongly induced by the vaccination (Figure [2D,E;](#page-3-0) Figure S6A). Importantly, we noted a marked increase in the frequencies of $T_{FH}2$ cells among either ICOS^{high}PD-1^{high} cT_{FH} cells or total CD4⁺ T cells in AR patients compared with healthy subjects (Figure S6A). As expected, the fold changes of $T_{FH}2$ cells to baseline were also elevated in AR patients at days 7 and 37 (Figure [2E\)](#page-3-0). Furthermore, only $T_{\rm FH}$ 2 activation, not $T_{\rm FH}$ 1 and $T_{\rm FH}$ 17 activation, was found

FIGURE 2 Increased circulating T_{FH}2 cells associate with protective antibodies in patients with allergic rhinitis (AR) following vaccination. (A) Frequencies of circulating ICOShighPD-1high T_{FH} cells were analyzed by flow cytometry. (B) Changes of ICOShighPD-1high T_{FH} cells at the indicated time points were normalized to the baseline levels and represented as fold changes. (C) Correlation of fold changes of ICOShighPD-1high T_{FH} cells at day 37 with fold changes of SARS-CoV-2 neutralizing antibody at day 60 relative to those at baseline in patients with AR. (D and E) Frequencies of circulating $ICOS^{high}PD-1^{high}$ T_{FH} cell subsets were analyzed by flow cytometry. Changes of $ICOS^{high}PD-1^{high} T_{FH}$ cell subsets at the indicated time points were normalized to the baseline levels and represented as fold changes. (F) Correlations of fold changes of ICOShigh PD-1high T_{FH} cell subsets at day 37 with fold changes of SARS-CoV-2 neutralizing antibody at day 60 in patients with AR. (G) Plasma SARS-CoV-2 neutralizing antibody and IgG levels in recovered COVID-19 patients at 10–12 months after infection ($n = 64$ for controls without AR and $n = 14$ for those with AR). Numbers indicate the frequency of cells in the gated region. For (A), (B), (E) and (G), data are presented as median and interquartile range and analyzed by Mann–Whitney *U*-test. For (C) and (F), data are analyzed by Spearman's rank correlation

to be positively correlated with neutralizing antibodies in patients with AR (Figure [2F\)](#page-3-0), despite positive correlations between $T_{FH}1$ or $T_{FH}17$ activation and IgG and IgM in AR patients (Figure S6B). Therefore, such results suggest that $T_{FH}2$ subset might play a prominent role in promoting protective humoral response to two-dose inactivated vaccination against SARS-CoV-2.

To verify the influence of AR on the humoral response to SARS-CoV-2, we additionally recruited 78 recovered COVID-19 patients, including 64 without AR and 14 with AR, who showed no difference in demographic characteristics (Table S2). Similar to the humoral response elicited by inactivated SARS-CoV-2 vaccine, patients with preexisting AR displayed higher levels of neutralizing antibody 10–12 months after infection than control subjects, while the difference of SARS-CoV-2-specific IgG did not reach statistical difference (Figure [2G\)](#page-3-0).

Our study of AR patients for the antibody response to SARS-CoV-2 induced by inactivated vaccine or infection demonstrates that the protective humoral immune response was enhanced rather than compromised in AR patients. The prospective design in the vaccination study allowed us to measure both neutralizing antibody and RBD-specific memory B cells, both supporting the above conclusion. As viral infection and vaccination, especially with influenza, induce a strong T_H1 immune response, the generation and function of the $T_{FH}1$ subset associated with T_H1 response has been extensively investigated in viral infection and vaccination. Notably, COVID-19 infection induces both T_H1 and T_H2 responses.⁷ Despite the $T_{FH}1$ subset was variously activated by inactivated SARS-CoV-2 vaccine, our study revealed an unappreciated role of the $T_{FH}2$ subset following vaccination. We have previously reported an increased $T_{\rm FH}$ 2 cells in patients with AR[.6](#page-5-0) Our new results demonstrate that the activation of $T_{FH}2$ rather than $T_{FH}1$ subset correlated with the induction of neutralizing antibodies in AR patients. Indeed, a recent study showed that IL-4, a signature cytokine from $T_{FH}2$ cells, plays a critical role in supporting broadly protective antibody response in viral infection.⁹ Although differing in intensity, similar humoral immune responses have been elicited by different SARS-CoV-2 vaccines.¹⁰ Whether AR patients have comparable immune responses to other SARS-CoV-2 vaccines, such as BNT162b2 mRNA vaccine, deserves further investigations.

In summary, our study provides the necessary evidence to justify the recommendation of SARS-CoV-2 vaccination to patients with AR and potentially other allergic diseases. Although the precise molecular mechanisms underlying the enhanced protective humoral immunity to COVID-19 vaccine in patients with AR remain to be elucidated, the function of $T_{FH}2$ cells in regulating anti-SARS-CoV-2 humoral immune response should be a key area deserving a future investigation. In addition, it is also critical to understand whether type 2 response-modifying treatments, such as allergen immunotherapy and biologics, will impair the efficacy of SARS-CoV-2 vaccination.

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CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

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