



Published in final edited form as:

J Clin Neurophysiol. 2022 February 01; 39(2): 135–148. doi:10.1097/WNP.0000000000000784.

Biomarkers obtained by transcranial magnetic stimulation in neurodevelopmental disorders

Ali Jannati, M.D., Ph.D.^{1,2,4,*}, Mary A. Ryan, M.Sc.^{1,2}, Harper Lee Kaye, B.Sc.³, Melissa Tsuboyama, M.D.¹, Alexander Rotenberg, M.D., Ph.D.^{1,2,4,*}

¹Neuromodulation Program and Division of Epilepsy and Clinical Neurophysiology, Department of Neurology, Boston Children's Hospital, Harvard Medical School, Boston, MA, USA

²Berenson-Allen Center for Noninvasive Brain Stimulation and Division of Cognitive Neurology, Department of Neurology, Beth Israel Deaconess Medical Center, Harvard Medical School, Boston, MA, USA

³Behavioral Neuroscience Program, Division of Medical Sciences, Boston University School of Medicine, Boston, USA

⁴F.M. Kirby Neurobiology Center, Department of Neurology, Boston Children's Hospital, Harvard Medical School, Boston, MA, USA

Abstract

Transcranial magnetic stimulation (TMS) is a method for focal brain stimulation that is based on the principle of electromagnetic induction where small intracranial electric currents are generated by a powerful fluctuating magnetic field. Over the past three decades, TMS has shown promise in the diagnosis, monitoring, and treatment of neurological and psychiatric disorders in adults. Yet, the use of TMS in children has been more limited. We provide a brief introduction to the TMS technique, common TMS protocols including single-pulse TMS, paired-pulse TMS (ppTMS), paired associative stimulation (PAS), and repetitive TMS (rTMS), and relevant TMS-derived neurophysiological measurements including resting and active motor threshold, cortical silent period, ppTMS measures of intracortical inhibition and facilitation, and plasticity metrics following rTMS. We then discuss the biomarker applications of TMS in a few representative neurodevelopmental disorders including autism spectrum disorder, fragile X syndrome, attention-deficit hyperactivity disorder, Tourette syndrome, and developmental stuttering.

Keywords

transcranial magnetic stimulation; neurodevelopmental disorders; pediatrics; neurology; biomarkers

*Correspondence: Ali Jannati (Ali.Jannati@childrens.harvard.edu) or Alexander Rotenberg (Alexander.Rotenberg@childrens.harvard.edu), Department of Neurology, Boston Children's Hospital, 300 Longwood Ave (Fegan 9), Boston, MA, 02115, USA. Phone: +1-617-667-0307. Fax: +1-617-975-5322.

Conflicts of Interest Statement

A.R. is a founder and advisor for Neuromotion, serves on the medical advisory board or has consulted for Cavion, Epihunter, Gamify, Neural Dynamics, NeuroRex, Roche, Otsuka, and is listed as inventor on a patent related to integration of TMS and EEG. The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

1. Introduction

Transcranial magnetic stimulation (TMS) is a 30-year-old technique for focal, noninvasive, electrical cortical stimulation (see (1) for a comprehensive review). In contrast to many neurostimulation protocols, TMS has robust prospects in the diagnostic and biomarker space. Specifically, TMS-derived biomarkers have been obtained for a range of disease states that include epilepsy (2), migraine (3), and pain (4). Yet, most of these have been studies in adults. Reviews of TMS safety in children (5–11) indicate minimal risk associated with the technique. Accordingly, TMS may serve to further our understanding of the underlying pathophysiology of neurodevelopmental disorders such as autism spectrum disorder or fragile X syndrome, and help identify biomarkers to aid in early diagnosis, monitoring disease progression, and development of novel therapies.

2. Common TMS Protocols and TMS-EMG Measures of Cortical Excitability and Plasticity

TMS is based on Faraday's principle of electromagnetic induction where a powerful fluctuating magnetic field induces a small intracranial electrical current in the brain (12). The induced current depolarizes the nearby neuronal assemblies located underneath the coil and can generate neurophysiological and/or behavioral effects (1). The true biophysical dynamics of TMS and the exact neural elements are activated by TMS remain unclear, and may vary across different brain regions and individuals (13–16). Models of TMS (17–20) raise many of the relevant issues, but need to be tested further on cellular and molecular levels (21,22).

Applying TMS to the primary motor cortex (M1) can selectively activate the contralateral muscles that are controlled by the targeted M1 (23). The extent of this activation can be quantified via motor evoked potential (MEP) amplitudes using skin-surface electromyography (EMG), enabling TMS-derived measures of the cortical excitation and inhibition (6), two opposing forces that influence many cortical functions in the mammalian cerebral cortex (24,25). The extent of inhibition generated in cortical networks is typically proportional to local and/or incoming excitation through the recruitment of interneurons via feedforward and/or feedback excitatory projections (25). Such excitation:inhibition (E:I) balance, in which increases in excitation are accompanied by increases in inhibition, has been observed *in vitro* in several sensory cortical regions (26–28) as well as during spontaneous cortical activity (29–31).

The stimulation intensity in TMS protocols is typically adjusted based on normalized measures such as motor threshold, the intensity that elicits an MEP of 50 μ V on 50% of the pulses applied over the cortical hot spot of a target muscle either at rest (*resting motor threshold*; rMT) (32) or during isometric contraction (*active motor threshold*; aMT).

The pattern of stimulation and outcome measures of TMS modalities can vary based on the number and intensity of the TMS pulses, inter-pulse/inter-train intervals, stimulation

frequency, TMS pulse waveform, and the direction of the induced current in the brain (13,33,34). The three main types of TMS protocols are as follows (Table 1):

a. Single-pulse TMS

Single-pulse TMS (spTMS) typically consists of single pulses delivered at 5–8 s intervals, to avoid cumulative effects of individual pulses over time. A standard TMS pulse is usually 150–300 μ s in duration and can cause depolarization followed by a refractory period, after which neurons recover rapidly. spTMS is used to assess corticospinal integrity and maturity (35–39), for motor mapping (40), and to assess the aftereffects of repetitive TMS (rTMS) protocols (41,42).

The cortical silent period (cSP) is a period of EMG suppression following an MEP elicited by spTMS during the voluntary contraction of a given muscle that lasts between 100–300 ms (43). The early part of the cSP (up to the first 50 ms) is related to activation of spinal inhibitory interneurons by the fibers descending from M1 (44), whereas the later portion is due to the cortical inhibition originating from M1 (45). cSP duration is a reflection of GABA_B- and GABA_A-mediated motor-cortex inhibition (46). For example, several studies have found shortened cSP duration in the epileptic hemisphere in patients with focal motor epilepsy (47–49), which may indicate a disruption in cortical inhibitory mechanisms that results in epileptic EEG activity. Another study found bilaterally lengthened cSP, but less prominently in the epileptic hemisphere, in patients with focal epilepsy (50). The significance of such cSP increase is not certain, but it may reflect a compensatory increase in interictal cortical inhibition to prevent epileptic activity (43). Increased cSP duration has also been observed in patients with idiopathic generalized epilepsy (51,52).

The ipsilateral silent period (iSP), a variant of the cSP, is used to assess the functional integrity of the transcallosal projections between motor cortices in which spTMS to M1 causes a transient suppression of voluntary tonic muscle activity in the ipsilateral hand muscles.

Strength of corticospinal projections are reflected by the shape and slope of an input/output (I/O) curve, also known as the *recruitment curve*, which is obtained by measuring MEP amplitudes elicited by single TMS pulses delivered over a wide range of intensities (53). The I/O curve for the intrinsic hand muscles has a sigmoid shape (54), in which the main outcome measures are the slope of the curve and the intensity at which it reaches a plateau. The slope of the I/O curve is steeper for intrinsic hand muscles that have a lower activation threshold (i.e., a lower rMT) and becomes less steep following administration of sodium- and calcium-channel blockers (e.g., lamotrigine) and GABA_A receptor agonists (e.g., lorazepam) (54). The finding that lorazepam has no effect on rMT but modulates the I/O curve, particularly at highest intensities (54), indicates that the I/O curve provides complementary information relative to rMT about the motor-system activity, including the GABAergic circuitry (13).

b. Paired-pulse TMS

In paired-pulse TMS (ppTMS), a conditioning stimulus (CS) is followed by a test stimulus (TS) after an interstimulus interval (ISI). PpTMS is used to assess intracortical inhibitory

or facilitatory mechanisms, most commonly local to M1. In most ppTMS protocols (except LICI, described below), CS and TS intensities are set at sub-rMT and supra-rMT intensities, respectively. In short-interval intracortical inhibition (SICI) (ISI = 1–6 ms), CS subliminally influences intracortical neurons, which will not resume their full responsiveness until they have had time to recover. Thus, a TS preceded by a CS can only activate a subset of neurons in the target region resulting in lower MEP amplitudes than the ones elicited by the TS alone. While the stimulation sequence utilized in the short-interval intracortical facilitation (SICF) protocol is similar to that in SICI, the two protocols differ in that CS and TS intensities in SICF are supra-rMT and sub-rMT, respectively (55), or both stimuli are close to rMT intensity (56). The TS in SICF is thought to excite directly the initial axon segments of the excitatory interneurons that were depolarized by excitatory post-synaptic potentials elicited by the CS but did not fire an action potential (57,58). In intracortical facilitation (ICF) (ISI = 8–30 ms), CS engages facilitatory intracortical neurons that enhance the output of response to TS, resulting in greater MEP amplitudes compared to those elicited by the TS alone (59). In long-interval intracortical inhibition (LICI) (ISI = 50–200+ ms), both CS and TS are suprathreshold, resulting in inhibition of the TS MEP (60,61). Epidural recordings of the descending corticospinal volley have found LICI likely engages intracortical neurons that inhibit the *I-waves* produced by indirect activation of layer-V pyramidal tract neurons (62). Table 1 details the stimulation parameters of common ppTMS protocols and their likely mechanisms.

Short-interval inhibitory effects are mainly mediated by gamma-aminobutyric acid (GABA) activity, whereas long-interval facilitatory effects are mainly mediated by glutamatergic activity (63–66). Specifically, SICI reflects GABA_A-mediated regional cortical inhibition, while ICF reflects glutamate [*N*-methyl-*D*-aspartate (NMDA) and α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptor types]-mediated excitation, and LICI represents GABA_B-mediated inhibition and (likely) an aggregate inhibitory tone mediated by the GABA_A receptor system (46,67). SICI represents fast inhibitory postsynaptic potentials (IPSPs) in corticospinal neurons mediated by GABA_A receptors, whereas LICI additionally represents slow IPSPs that are mediated, at least in part, by GABA_B receptors. Applying ppTMS to M1 can be used to assess abnormal states of low or high cortical excitability by measuring the cortical E:I ratio, e.g., by measuring the SICI/ICF ratio (68,69), which provides an estimate of the relative strength of local intracortical inhibitory and excitatory activities (38,70), whereas ppTMS, when applied to two cortical regions, can assess inter-regional connectivity and conduction time (71,72).

c. Repetitive TMS

rTMS typically consists of several pulses at 1–20 Hz frequency over long periods of stimulation (up to 30 minutes), at times including stimulation-free intervals (13). rTMS is commonly used to modulate regional cortical activity in neuropsychiatric disorders and, particularly when applied as long trains over consecutive daily visits, can induce long-lasting plastic aftereffects (73,74) by influencing the brain networks associated with the targeted region (1). Typically, low-frequency (< 1 Hz) rTMS with at least 300–900 pulses can induce sustained inhibition of cortical excitability, whereas high-frequency (> 5Hz) or discontinuous rTMS can induce long-lasting facilitation of cortical excitability (41,75). The

inhibitory and facilitatory aftereffects of low-frequency and high-frequency rTMS resemble use-dependent, long-term depression (LTD) and long-term potentiation (LTP) of excitatory (glutamatergic) synaptic strength – however, these aftereffects are in part also mediated by GABAergic mechanisms (76–78).

A form of patterned rTMS known as theta-burst stimulation (TBS) consists of 50 Hz bursts of three TMS pulses repeated every 200 ms (i.e., at 5 Hz), for a total of 600 pulses, in one of two protocols: (1) a 2-s on, 8-s off intermittent TBS (iTBS) for 190 s that can increase MEP amplitude by ~35% for up to 60 min, or (2) a continuous TBS (cTBS) for 40 s that can reduce MEP amplitude by ~25% for up to 50 min (79). Facilitation and suppression of MEPs by iTBS and cTBS are considered indices of LTP- and LTD-like mechanisms of synaptic plasticity, respectively (77,80). The time necessary for post-TBS MEPs to return to pre-TBS baseline and the pattern of post-TBS aftereffects are considered neurophysiologic indices of the mechanisms of cortical plasticity (79,81,82), albeit with considerable inter- and intra-individual variability in responses (83–86). Physiologic and pharmacologic TBS studies in humans indicate the involvement of glutamatergic and GABAergic mediators consistent with LTP and LTD, respectively, and the pattern and time course of their aftereffects (77,78,87,88).

d. Paired Associative Stimulation (PAS)

PAS is an electrophysiological technique that involves repeated pairing of two stimuli, e.g., a peripheral electrical pulse delivered to a nerve of the hand, usually the median nerve (median-nerve stimulation; MNS), that activates the primary sensory cortex (S1) and a TMS pulse over the corresponding hand representation in the contralateral M1 (89). Through this S1-M1 coupling, PAS is able to modulate corticospinal excitability as indicated by the change in MEP amplitudes (89,90). PAS-induced modulation of MEPs reflects the propensity of the nervous system to adapt, i.e., plasticity. This plasticity is presumed to rely on the principles of Hebbian synaptic plasticity (91) such that the modulation of MEP amplitude depends on the interstimulus interval (ISI) between the sensory stimulus and the TMS pulse applied to M1. Specifically, based on a temporally asymmetric Hebbian rule (92), a shorter interval (ISI = 10 ms) produces a decrease in the MEP amplitude, whereas a longer interval (ISI = 25 ms) induces an increase in the MEP amplitude (89,90,92,93). These interventions have been termed PAS10 and PAS25, respectively (93). PAS aftereffects develop rapidly (within 30 min), are long-lasting (at least 60 min), reversible, and cortically generated (89,90,94). PAS25 aftereffects depend on NMDA glutamatergic receptors, and are thought to be mediated by an LTP-like mechanism (89,92). In a variant referred to as PAS_{N20+2}, stimulation consists of 225 pairs (rate, 0.25 Hz) of MNS followed at an interval equal to the individual N20 latency of the median nerve somatosensory-evoked cortical potential plus 2 ms by applying spTMS to the hand area of the contralateral M1 (90,95).

3. TMS biomarkers in neurodevelopmental disorders

a. Autism Spectrum Disorder (ASD)

ASD is a lifelong developmental syndrome that affects ~1 of 59 children (96) and is characterized by difficulties with interpersonal relationships and communication,

and presence of restrictive and repetitive behaviors, interests, or activities (97). There are currently no FDA-approved treatments for core ASD symptoms. Development of novel therapeutics will require reliable biomarkers and improved understanding of ASD pathophysiology (98).

Among many mechanistic ASD models are those reliant on imbalance of cortical excitation and inhibition (99–101). However, most spTMS and ppTMS measures of cortical excitability such as rMT, MEP amplitude, LICI, or ICF show no clear difference between patients with ASD and neurotypical (NT) individuals (81,102–104). In the 2013 study by Enticott and colleagues (103), SICI in cognitively normal patients with ASD was shortened compared to their NT counterparts, and the GABA_Aergic activity, as indexed by SICI, was associated with the extent of developmental delay in language acquisition in patients with ASD. This suggests that some of the heterogeneity in ASD symptomatology may be associated with interindividual variability of GABA_Aergic activity. Two other studies, however, found SICI in ASD patients to be normal as compared to NT controls (105,106).

Pedapati and colleagues (107) recently reported findings from the largest sample of ASD youth to undergo ppTMS to date (n=59), and found normal SICI and cSP in ASD youth compared to typically developing children. However, enhanced SICI (greater inhibition) and prolonged cSP were highly correlated with clinical history and standardized measures reflecting academic struggles in reading, spelling, and/or math. Moreover, across all ASD youth, ICF was inversely correlated with worse inattention, and lack of ICF was associated with inattention and executive dysfunction (107).

In contrast to spTMS and ppTMS measures, Oberman et al. (81) identified a reliable distinction between the ASD and NT groups using TBS. The investigators used cTBS and iTBS paradigms to evaluate LTD- and LTP-like plasticity, respectively, in 20 cognitively normal adults with ASD and found greater and longer-lasting modulation of M1 reactivity following both TBS paradigms compared to age-, gender-, and IQ-matched NT controls. These results were confirmed in a separate cohort of 15 adults with ASD (81) as well as in a follow-up study by Oberman and colleagues on 10 adults with ASD (108). The longer-lasting iTBS aftereffects in the ASD group are consistent with the hypothesis that a lack of inhibitory tone would lead to a greater propensity for LTP. Similarly, the longer-lasting cTBS aftereffects among subjects with ASD are consistent with the Bienenstock, Cooper, and Munro (BCM) model (109) that predicts an already-potentiated synapse will have a lower threshold for LTD, and, thus, will exhibit a stronger LTD-like response than a depressed or neutral synapse (110). The greater potential for both LTP and LTD, coupled with the finding that measures of baseline cortical excitability such as rMT and baseline MEP amplitude were comparable between the ASD and NT groups (81), indicated an unstable state of synaptic plasticity in ASD, which results in exaggerated, *hyperplastic*, response to TBS. In a subsequent study of children and adolescents with ASD, Oberman and colleagues (111) found a positive relationship between age and the extent of cTBS-induced modulation, suggesting a maturational trajectory for LTD-like plasticity during childhood and adolescence in ASD.

Recently, we extended those findings by comparing M1 cTBS aftereffects between 11 children and adolescents with ASD and 18 of their age-, gender-, and IQ-matched controls (112). We found cTBS aftereffects differentiated between the ASD and control groups due to more-facilitatory cTBS aftereffects in the ASD group relative to the control group. Notably, the difference in cTBS responses between ASD and control groups remained after participants with ADHD were excluded from the ASD group. We also found an age-related increase in the maximum cTBS-induced suppression of MEPs in the ASD group only, suggesting a dysmaturity in the LTD-like plasticity in ASD youth (112).

Based on the involvement of GABAergic synaptic transmission in cTBS aftereffects (78,113), the more-facilitatory responses to cTBS in the ASD group than in the TD group lends further support to the notion of GABAergic dysfunction in ASD (114–116). Thus, quantification of M1 cTBS aftereffects may serve as a physiologic biomarker for children and adolescents with ASD, which can improve the classification of clinical endophenotype and understanding of ASD pathophysiology, and be used to assess target engagement and monitor response to experimental pharmacologic or neuromodulatory therapies for ASD (98,117).

However, the findings of hyper-plasticity on ASD are not consistent with the results of one study that compared PAS plasticity measures between ASD and control groups; Jung and colleagues (105) found that, unlike healthy controls, patients with ASD did not show the expected PAS25-induced facilitation of MEPs. The authors found similar results with PAS_{N20+2} in a small subgroup of ASD patients. The results suggested reduced LTP-like plasticity and deficits in sensorimotor integration in ASD (105).

Notably, as the majority of the participants in the aforementioned studies (summarized in Table 2) are children or adolescents with ASD who are cognitively and linguistically normal, it remains unclear whether these results extend to adults or children with ASD who have cognitive or language deficits.

b. Fragile X Syndrome (FXS)

FXS is the most common cause of inherited intellectual disability (118), with a prevalence estimated at 1.4 and 0.9 per 10,000 males and females, respectively (119), and the most common genetic cause of autism, with 30% of children with FXS diagnosed with autism and 2–5% of autistic children having FXS (120). FXS is caused by the expansion of the trinucleotide sequence CGG located in the 5' untranslated region (UTR) of the X-linked *FMR1* gene that results in hypermethylation and consequent silencing of the *FMR1* gene. The silencing of the *FMR1* gene prevents the encoding of the fragile X mental retardation protein (FMRP) and forms the basis for the FXS phenotype (121). FMRP is an inhibitory regulator of translation of metabotropic glutamate receptor 5 (mGluR5), which is involved in formation of new synaptic connections, stabilizing LTD, and inducing LTP. Absence of FMRP results in excess mGluR5 activity and subsequent abnormalities in synaptic plasticity, as observed in animal FXS models (122,123).

rTMS protocols in subjects with FXS have also demonstrated evidence for abnormal neural plasticity. In a study by Oberman and colleagues (124), FXS patients showed a lack of

response to cTBS, an enhanced response to iTBS, and a complete blocking of iTBS response on the subsequent day, suggesting abnormalities in plasticity and *metaplasticity*. Metaplasticity refers to a change in the state of the synapse that alters its ability to make subsequent plastic changes, involves many of the same mechanisms as conventional LTP and LTD, and likely serves as a homeostatic function to maintain plasticity within a normal working range (124).

In a follow-up study, Oberman and colleagues (108) found that six adults with FXS showed abnormal TBS-induced plasticity and metaplasticity; the duration of the cTBS response in the FXS group in the first visit was significantly shorter than that in the control group, and they showed paradoxical facilitation of MEPs following cTBS in their second visit. The results from both ASD and FXS subjects provide evidence for aberrant TMS-induced plasticity and metaplasticity at the level of cortical circuits, which are consistent with the theories proposed to explain these processes at the synaptic level, e.g., the BCM model that predicts a lower-than-normal LTD threshold, and thus, a stronger LTD-like response than a depressed or neutral synapse (109).

c. Attention-Deficit Hyperactivity Disorder (ADHD)

ADHD is one of the most common neurodevelopmental disorders affecting 2–7.5% of school-aged children, with core symptoms of inattention, hyperactivity, and impulsivity (125). The pathophysiology of ADHD remains unclear, and there is currently no widely accepted biomarker or diagnostic test for ADHD. Thus, an ADHD diagnosis is typically based on parent- and teacher-reported behavioral rating scales combined with a physician's clinical impression. An influential theory of ADHD pathophysiology posits that executive dysfunction in ADHD is due to deficient inhibitory control (126,127), the neural substrate of which involves basal ganglia-thalamocortical circuits (128). Impulse-control deficit and hyperactivity in ADHD have been associated with dysfunction of frontostriatal circuits including underactivity of ventrolateral prefrontal cortex, dorsolateral prefrontal cortex, and anterior cingulate cortex. The negative association between inhibitory control and hyperactivity in ADHD is paralleled by abnormally reduced SICI (129), an inverse correlation between SICI and symptom severity in most patients with ADHD (127,130), and, interestingly, paradoxical facilitation following SICI in a few cases (127). SICI is normalized following administration of methylphenidate (131) indicating that SICI may function as an index of therapeutic response to methylphenidate, and, therefore, may play a role in dosing of stimulants, and future drug development for treatment of children with ADHD. Future studies on the effect of other stimulants on SICI are needed to investigate its utility for drug selection in patients with ADHD.

Prolonged latency and duration of the ipsilateral silent period (iSP) found in ADHD suggests deficient transcallosal-mediated inhibition in ADHD (132). Interestingly, the iSP duration is correlated with the degree of hyperactivity and restlessness in ADHD, and is also normalized with a single dose of methylphenidate. This indicates the prominent role of abnormal cortical excitability, rather than structural abnormalities of the corpus callosum, in the pathophysiology of ADHD. This view is supported by the finding that iSP duration in ADHD is inversely correlated with the magnitude of SICI (133,134). In contrast to children

with ADHD, the iSP in adults with ADHD is abnormally shortened but has a normal onset-latency (135). The difference in iSP between children and adults with ADHD could be due to developmental differences in inhibitory intracortical circuits (125).

The findings that SICI is correlated to hyperactivity severity in ADHD, and methylphenidate normalizes SICI while reducing hyperactivity, indicate that SICI could function as an index of therapeutic response to methylphenidate, and, therefore, guide drug selection, dosing of stimulants, and future drug development for treatment of children with ADHD.

Pedapati and colleagues (107) found ICF was significantly reduced in ASD children with ADHD comorbidity (ASD+) compared to ASD children without ADHD comorbidity (ASD-) and typically developing children. This reduction in ICF, together with the reported functional abnormalities in the temporal lobe and amygdala in ASD- and ASD+ youth (136), suggests that ICF may represent the combined effects of several aberrant cortical and subcortical circuits specifically relevant to the presence of ADHD symptoms in the ASD population (107).

d. Tourette Syndrome (TS)

Tourette syndrome (TS) is a chronic, childhood-onset neuropsychiatric disorder characterized by more than 1 year of motor and vocal tics. Tics are repetitive, stereotypical but nonrhythmic, involuntary, or compulsive movements that occur at irregular intervals, usually between purposeful movements. TS affects 1–3% of Western school-aged children, with the age of onset ranging from 2 to 21 years (137). Chronic tic disorders, characterized by either motor or vocal tics, but not both, occur in 5% of school-aged children (137). Common comorbidities include ADHD, obsessive-compulsive disorder (OCD), learning disabilities, conduct disorders, self-injury, and sleep disturbances (138,139). The pathophysiology of tics in TS likely involves dysfunctional integration of information with motor output from many sources via basal ganglia-thalamocortical circuits that results in incomplete suppression of unwanted behaviors (140).

Adults with TS have normal rMT and aMT (141,142) but, according to Orth et al. (143), recruit significantly fewer corticospinal neurons at higher stimulation intensities as indexed by shallower I/O curve slopes compared to healthy controls. This pattern of results suggest that while the most excitable connections (i.e., those recruited at rMT) are in a comparable state in TS patients and healthy controls (141–144), the difference between the most and the least excitable corticospinal neurons is greater in TS than in healthy controls (143). In a subsequent study by Heise et al. (144), no such difference in the I/O curve slopes was found, perhaps due to the less-severe symptoms of TS patients compared to the cohort studied by Orth et al. (143).

Several ppTMS studies have found abnormally reduced SICI at rest in TS patients (141–144). While TS patients have normal thresholds for SICI (the stimulation intensity needed to produce SICI), their recruitment of inhibition at suprathreshold intensities is abnormally reduced (142,143). The reduced SICI in TS patients correlates with greater motor tic severity, especially in patients who are not under treatment with dopamine antagonists and those patients who have comorbid ADHD (145,146). Interestingly, the ability to normalize

SICI during movement preparation is associated with fewer tics and lower tic severity (144), suggesting that control over tic movements depends on the ability to reduce the excitability of motor neurons in line with increased intracortical inhibition (140).

One study (147) identified greater ICF in patients with TS and comorbid ADHD compared to healthy controls, individuals with TS alone, or individuals with TS and comorbid OCD. This result suggested that comorbid conditions like ADHD may also independently affect TMS measures of cortical excitability or inhibition. Other studies, however, did not find such a difference in ICF between TS and control groups (141,146,148), a discrepancy that may be due to the well-established interindividual variability in ICF (149).

cSP duration in TS patients has been found to be abnormally shortened (141,142) or normal when cSP measurements are corrected for differences in MEP amplitude between TS and control groups (143). These results indicate that the relationship between corticospinal excitability and the intracortical inhibitory mechanism reflected in the cSP, possibly via corticospinal motor-neuron recurrent collaterals (150), is comparable in TS patients and controls (140).

There is a reduction in MEP amplitude elicited by suprathreshold spTMS in individuals with TS during the period immediately preceding volitional movements compared to the expected increase in MEP amplitude in control subjects during the same timeframe (151,152). Ganos and colleagues found that voluntary tic suppression in adults with TS also reduced corticospinal excitability, as indexed by reduced MEP amplitude following spTMS and a shallower I/O curve, the extent of which was associated with the ability to control tics (153). These findings suggest that cortical excitability is reduced in the period immediately preceding volitional movement or that individuals with TS have impaired ability to modulate motor cortical excitability prior to tics. However, unlike adults with TS (141,142), rMT was found to be higher in adolescents and young adults compared to age-matched controls, with the differences being more pronounced in the younger age groups (154). MEP variability is also larger in TS patients, although the degree of variability decreases with age both in individuals with and without TS. Similar to results seen in adults with TS, adolescents (<18 years old) with TS also exhibited reduced motor excitability during the period immediately prior to volitional movements. rMT depends upon recruitment of corticospinal neurons that project to a target muscle, and increases in motor excitability and decreases in MEP variability result from consistent firing patterns of a given group of motor cortical neurons recruited during movement preparation. Therefore, this age-related difference in rMT and MEP variability may be related to a delay in the formation of cortical-cortical and corticospinal motor networks in TS that leads to a reduced number of neurons recruited by a TMS pulse or inconsistent firing of recruited motor cortical neurons (154).

Plasticity induced by TBS protocols has also been studied in TS patients, specifically to test whether abnormal thalamo-cortical motor inputs in TS influence M1 plasticity by eliciting aberrant activity in cortical layers responsible for the measured TBS aftereffects (155–157). Wu and Gilbert (156) found TS patients showed greater iTBS-induced facilitation of MEPs at 1 and 10 minutes post-iTBS than healthy controls. In contrast, Suppa and colleagues (157) found that TS patients, with and without psychiatric comorbidity, showed reduced

responses to both iTBS and cTBS compared to healthy controls. The authors suggested that the lack of correlations between tics and neurophysiological measures could be due to the involvement of nonprimary motor cortices, basal ganglia and cerebellum, in addition to M1 and brainstem, in the generation of tics (157). It has been suggested that alterations of synaptic plasticity in TS may arise from metaplasticity effects occurring as a consequence of tics (155,156).

TMS studies have also investigated LTP-like plasticity response induced by the PAS protocol in TS patients. Brand and colleagues (158) found that while most healthy controls showed the expected LTP-like response following PAS_{N20+2}, TS patients were more likely to show a paradoxical, LTD-like response, the extent of which was correlated with less-severe urges and fewer tics. Their results also suggested that aberrant PAS-induced facilitation of MEPs in TS could be related to reduced long-term consolidation of motor skills in a rotary pursuit task (158).

e. Developmental Stuttering (DS)

DS is a disruption in the normal speech rhythm characterized by repetitions, prolongations, and tense pauses manifesting speech blocks (159). Secondary, associated symptoms of DS include movements/spasms, most commonly in facial muscles (160,161). The prevalence of DS, with various degrees of severity, is ~5% in children and ~1% in adults (162).

In the first TMS study in DS, Sommer and colleagues (163) investigated the abnormalities in intracortical inhibition and facilitation of hand-muscle MEPs at 1–30 ms ISIs. The rMT and aMT were both abnormally high in DS patients, indicating lower corticospinal excitability in DS. Consistent with this finding, Busan and colleagues (164) found lower MEP recruitment in DS patients, especially in DS males, at higher intensities (e.g., 150% of rMT), indicating abnormally low cortical excitability in DS. Interestingly, cSP durations were negatively correlated with stuttering severity in males, perhaps due to a compensatory mechanism by intracortical circuits (164). Another piece of evidence for lower corticospinal excitability was obtained by Alm and colleagues (165) who found higher rMT in the left hemisphere of DS patients, both relative to their right hemisphere and to the left hemisphere of fluent speakers, indicating lower excitability of corticospinal pathways in DS originating from the left M1. The authors interpreted this finding as reflecting difficulty in the initiation of movement in DS patients (165).

rTMS studies have also revealed abnormalities in motor-control networks in DS. Neef and colleagues (166) used subthreshold 1Hz rTMS to investigate the roles of right and left premotor cortices on motor output in DS. They found performance in a timed motor task was more influenced by applying rTMS to the right premotor cortex in DS patients but to the left premotor cortex in fluent speakers. This result suggested a possible compensatory mechanism by motor circuits related to the control of timed movements in the right hemisphere of DS patients due to dysfunction of the corresponding circuits in the left hemisphere (166).

TMS has also been used to investigate the motor representation of speech muscles in DS. In the first TMS study of corticobulbar pathways in DS, Neef and colleagues (167) found

a steeper I/O curve as well as reduced SICI (at 2ms ISI) and ICF among DS patients. These results suggested alterations in intracortical networks in DS, possibly mediated by altered GABAergic activity. More recently, Neef and colleagues (168) examined the change in MEP amplitudes elicited by spTMS over M1 during transition between a fixed labiodental articulatory configuration and immediately following articulatory configurations in adults with DS and healthy controls. Interestingly, adults with DS exhibited a lack of left-hemisphere facilitation during the transition phase, the extent of which was negatively correlated with objective measurements of stuttering severity (168). These results suggested a deficit in controlling speech motor plans in DS, likely in the left M1. Consistent with these results, Busan and colleagues (169) found an abnormally prolonged cSP in the left M1 representation of tongue, as well as significantly higher aMT in the left hemisphere of DS patients relative to their right hemisphere, the extent of which correlated with stuttering severity (169). Abnormalities of TMS measures in DS, especially in the left hemisphere, including reduced corticospinal and corticobulbar excitability, as well as reduced ICF and prolonged cSP can indicate a decreased tonic excitation of afferent inputs to M1 (170), possibly as a result of the white-matter abnormalities reported in DS, involving the left superior longitudinal fasciculus and the white-matter fibers between the left frontal and premotor cortices (171,172). Table 2 details the common TMS-based biomarkers in the disorders included in this review.

4. Adoption of TMS-based biomarkers in the clinic

Several TMS devices have received FDA approval for treatment of medication-resistant major depressive disorder (173–179). Additional devices have also been FDA-approved for acute and prophylactic treatment of migraine headache (180), presurgical motor and language mapping (181,182), and adjunct treatment of obsessive compulsive disorder (183). While TMS-derived biomarkers can be generated with these devices in the clinical setting, there remain a few barriers to wide adoption of their clinical use. We briefly summarize these below.

a. Education

Compared to other clinical protocols, TMS is relatively new, and thus, does not receive adequate coverage in medical educational curricula (184–186). Notably, while “rigorous training” for all physicians administering TMS for clinical purposes is advised by consensus statements (187), licensing and training requirements for clinicians to use TMS are largely absent. Intensive educational courses have been developed that partially address the need for organized TMS training programs (186), but the more-common incomplete training in TMS techniques among neurologists and psychiatrists remains one of the greatest barriers to the widespread adoption of TMS in the clinic.

b. Safety concerns

When application guidelines (32,188) are followed, TMS has an excellent safety profile in adults (188–190) and in children and adolescents (8–11). Still, concerns about an extremely rare, but potentially serious, side effect of rTMS, i.e., seizures, might dampen the motivation of physicians to use TMS in the clinic. However, while rare instances of rTMS-induced

seizure have been reported in adults (188), there has been no reported case of seizure induced by spTMS and ppTMS, among patients without epilepsy, which form the basis of the majority of TMS-based biomarkers. A measure of TMS safety in the pediatric population is provided in a recent study, in which no seizures or any other serious adverse events were reported after near 3 million rTMS stimulations in 119 children and adolescents, of whom 59 participants had perinatal stroke or mild traumatic brain injury and another 43 participants had major depression or Tourette's syndrome (10). These results underscore that TMS protocols, including rTMS, are safe and well-tolerated in children and adolescents, and can be considered minimal risk (8,10). Improving knowledge about the TMS safety profile among physicians can ease their potential safety concerns and set the stage for the more widespread use of TMS techniques in the clinic.

c. Proof-of-principle and confirmatory evidence

A fundamental obstacle to the more-common use of TMS-based biomarkers in the clinic is lack of adequate proof-of-principle studies that establish feasibility and provide preliminary evidence for TMS-derived biomarker utility in clinical pediatric populations. A lack of large confirmatory studies that can accommodate the variability of TMS responses observed in adult participants (83–86,191,192) similarly raises questions of feasibility and TMS-derived biomarker utility. Such studies, ideally followed by FDA approval of the relevant applications, are needed before widespread and evidence-based use of TMS biomarkers for pediatric populations can take place in the clinic.

d. Individualized TMS measures

The vast majority of the TMS measures discussed in the present review represent differences at the group level. At the individual level, responses to TMS protocols show large inter- and intra-individual variability, even in healthy populations (83–86,191,192). Such variability indicates the need for multivariate modeling of TMS measures that yields optimally adjusted TMS “scores” that incorporate relevant demographic, genetic, neurophysiological, neuroimaging, and state-dependent factors. These composite scores can assist clinical decision-making by placing more precisely an individual patient's baseline and post-intervention neurophysiological responses within his/her clinical cohort. Depending on the TMS protocol, obtaining a reliable TMS measure for optimal clinical use may involve considering age (36,193,194), gender (195–197), genetic polymorphisms (198–202), the TMS device, pulse waveform, and induced current direction (33,34,194), stimulation intensity and baseline neurophysiological measures, e.g., rMT, aMT, and baseline MEP amplitude (83,85,194,203), the target muscle (194,204), the time of day (84,194,205), use of neuronavigation (206) and robotic arms (207), amount and quality of sleep the night before the TMS visit (208,209), blood glucose level and caffeine intake before and during the TMS visit (210–212), intensity and duration of physical activity before each visit (213,214), phase of the menstrual cycle (215,216), and the use of closed-loop systems that deliver TMS pulses timed to real-time, EEG indices of brain states (217,218).

The current state-of-the-art individualized methods for TMS target localization or prediction of response to rTMS treatment rely on measuring the baseline or induced changes in resting-state functional connectivity between relevant brain regions at the level of the Human

Connectome (219–221) or the individual patient (220,222–225), or the local/distributed changes in cortical excitability as measured by TMS-evoked EEG potentials (TEPs) (224). It is thus likely that the next generation of individualized TMS biomarkers for “precision medicine” will also need to leverage individual patient data obtained from neuroimaging modalities, whether structural (to determine cortical thickness, scalp-to-cortex distance, sulcal/gyral geometry, or diffusion weighted imaging), functional (task- or resting-state fMRI to identify target regions or networks, or to determine functional connectivity between relevant regions), or neurophysiological (e.g., TEPs).

5. Conclusion

Neurophysiological measures obtained with noninvasive stimulation of the motor cortex can provide practical and feasible biomarkers for neurodevelopmental disorders. Such TMS-derived metrics of cortical excitability, E:I imbalance, and plasticity in the motor cortex can help elucidate the underlying pathophysiology of a range of neurodevelopmental disorders.

Several established single-pulse, paired-pulse, and repetitive TMS protocols are available that can be utilized to assess disease severity, measure and predict therapeutic response to existing therapeutic approaches in neurodevelopmental disorders, and enable novel measures of target engagement in the treatment of those disorders. Due to the substantial overlap in clinical presentations of disorders such as ASD, ADHD, and Tourette’s syndrome, it is important to consider the commonalities in TMS measures across such disorders in order to avoid potential confounding due to comorbidities. As more-comprehensive TMS evidence from larger populations of patients with such disorders becomes available, it becomes important to leverage the contrasting TMS findings in subgroups of these disorders, e.g., ASD with ADHD and ASD without ADHD, to enable more-refined classification and monitoring of patients in such subgroups based on the most-relevant TMS measure or combination of different TMS measures. Considering demographic, neurophysiological, genetic, and neuroimaging data may enable more-granular TMS biomarkers, approaching individualized measures.

Funding

This study was primarily funded by the National Institutes of Health (NIH R01 MH100186). A.J. was further supported by a postdoctoral fellowship from the Canadian Institutes of Health Research (CIHR 41791). A.R. was further supported by the NIH (R01 NS088583), The Boston Children’s Hospital Translational Research Program, Autism Speaks, Massachusetts Life Sciences, The Assimon Family, Brainsway, CRE Medical, Eisai, Neuroelectrics, Roche, Sage Therapeutics, and Takeda Medical. The content is solely the responsibility of the authors and does not necessarily represent the official views of Harvard Catalyst, Harvard University and its affiliated academic health care centers, the National Institutes of Health, or any of the other listed granting agencies.

References

1. Valero-Cabré A, Amengual JL, Stengel C, Pascual-Leone A, Coubard OA. Transcranial magnetic stimulation in basic and clinical neuroscience: A comprehensive review of fundamental principles and novel insights. *Neurosci Biobehav Rev.* 2017 Dec;83:381–404. [PubMed: 29032089]
2. Tsuboyama M, Kaye HL, Rotenberg A. Biomarkers Obtained by Transcranial Magnetic Stimulation of the Motor Cortex in Epilepsy. *Front Integr Neurosci.* 2019 Oct 30;13:57. [PubMed: 31736722]

3. Brighina F, Raieli V, Messina LM, Santangelo G, Puma D, Drago F, et al. Non-invasive Brain Stimulation in Pediatric Migraine: A Perspective From Evidence in Adult Migraine. *Front Neurol*. 2019 Apr 12;10:364. [PubMed: 31031695]
4. Zaghi S, Thiele B, Pimentel D, Pimentel T, Fregni F. Assessment and treatment of pain with non-invasive cortical stimulation. *Restorative neurology and neuroscience*. 2011;29(6):439–51. [PubMed: 22124038]
5. Croarkin PE, Wall CA, Lee J. Applications of transcranial magnetic stimulation (TMS) in child and adolescent psychiatry. *Int Rev Psychiatry*. 2011 Oct;23(5):445–53. [PubMed: 22200134]
6. Frye RE, Rotenberg A, Ousley M, Pascual-Leone A. Transcranial Magnetic Stimulation in Child Neurology: Current and Future Directions. *Journal of Child Neurology*. 2008 Jan;23(1):79–96. [PubMed: 18056688]
7. Garvey MA, Gilbert DL. Transcranial magnetic stimulation in children. *European Journal of Paediatric Neurology*. 2004 Jan;8(1):7–19. [PubMed: 15023371]
8. Gilbert DL, Garvey MA, Bansal AS, Lipps T, Zhang J, Wassermann EM. Should transcranial magnetic stimulation research in children be considered minimal risk? *Clinical Neurophysiology*. 2004;115(8):1730–9. [PubMed: 15261851]
9. Hong YH, Wu SW, Pedapati EV, Horn PS, Huddleston DA, Laue CS, et al. Safety and tolerability of theta burst stimulation vs. single and paired pulse transcranial magnetic stimulation: a comparative study of 165 pediatric subjects. *Frontiers in human neuroscience*. 2015;9(29).
10. Zewdie E, Ciecchanski P, Kuo H, Giuffre A, Kahl C, King R, et al. Safety and tolerability of transcranial magnetic and direct current stimulation in children: Prospective single center evidence from 3.5 million stimulations. *Brain Stimulation*. 2020;13(3):565–75. [PubMed: 32289678]
11. Krishnan C, Santos L, Peterson MD, Ehinger M. Safety of Noninvasive Brain Stimulation in Children and Adolescents. *Brain Stimulation*. 2015;8(1):76–87. [PubMed: 25499471]
12. Barker AT. Transcranial Magnetic Stimulation - past, present and future. *Brain Stimulation*. 2017 Mar;10(2):540.
13. Rotenberg A, Horvath JC, Pascual-Leone A, editors. *Transcranial magnetic stimulation*. New York: Humana Press : Springer; 2014.
14. Wagner T, Valero-Cabre A, Pascual-Leone A. Noninvasive human brain stimulation. *Annu Rev Biomed Eng*. 2007;9:527–65. [PubMed: 17444810]
15. Amassian VE, Eberle L, Maccabee PJ, Cracco RQ. Modelling magnetic coil excitation of human cerebral cortex with a peripheral nerve immersed in a brain-shaped volume conductor: the significance of fiber bending in excitation. *Electroencephalogr Clin Neurophysiol*. 1992 Oct;85(5):291–301. [PubMed: 1385089]
16. Maccabee PJ, Amassian VE, Eberle LP, Cracco RQ. Magnetic coil stimulation of straight and bent amphibian and mammalian peripheral nerve in vitro: locus of excitation. *J Physiol (Lond)*. 1993 Jan;460:201–19. [PubMed: 8487192]
17. Neggers SFW, Petrov PI, Mandija S, Sommer IEC, van den Berg NAT. Understanding the biophysical effects of transcranial magnetic stimulation on brain tissue: the bridge between brain stimulation and cognition. *Prog Brain Res*. 2015;222:229–59. [PubMed: 26541383]
18. Wilson MT, Fulcher BD, Fung PK, Robinson PA, Fornito A, Rogasch NC. Biophysical modeling of neural plasticity induced by transcranial magnetic stimulation. *Clin Neurophysiol*. 2018;129(6):1230–41. [PubMed: 29674089]
19. Rotem A, Moses E. Magnetic stimulation of one-dimensional neuronal cultures. *Biophys J*. 2008 Jun;94(12):5065–78. [PubMed: 18326634]
20. Opitz A, Windhoff M, Heidemann RM, Turner R, Thielscher A. How the brain tissue shapes the electric field induced by transcranial magnetic stimulation. *Neuroimage*. 2011 Oct 1;58(3):849–59. [PubMed: 21749927]
21. Vlachos A, Müller-Dahlhaus F, Roskopp J, Lenz M, Ziemann U, Deller T. Repetitive magnetic stimulation induces functional and structural plasticity of excitatory postsynapses in mouse organotypic hippocampal slice cultures. *J Neurosci*. 2012 Nov 28;32(48):17514–23. [PubMed: 23197741]
22. Funke K. Transcranial Magnetic Stimulation of Rodents. In: *Handbook of Behavioral Neuroscience*. Elsevier; 2018. p. 365–87.

23. Amassian VE, Stewart M, Quirk GJ, Rosenthal JL. Physiological basis of motor effects of a transient stimulus to cerebral cortex. *Neurosurgery*. 1987 Jan;20(1):74–93. [PubMed: 3543727]
24. Haider B, McCormick DA. Rapid neocortical dynamics: cellular and network mechanisms. *Neuron*. 2009 Apr 30;62(2):171–89. [PubMed: 19409263]
25. Isaacson JS, Scanziani M. How inhibition shapes cortical activity. *Neuron*. 2011 Oct 20;72(2):231–43. [PubMed: 22017986]
26. Anderson JS, Carandini M, Ferster D. Orientation tuning of input conductance, excitation, and inhibition in cat primary visual cortex. *J Neurophysiol*. 2000 Aug;84(2):909–26. [PubMed: 10938316]
27. Wehr M, Zador AM. Balanced inhibition underlies tuning and sharpens spike timing in auditory cortex. *Nature*. 2003 Nov 27;426(6965):442–6. [PubMed: 14647382]
28. Wilentz WB, Contreras D. Synaptic responses to whisker deflections in rat barrel cortex as a function of cortical layer and stimulus intensity. *J Neurosci*. 2004 Apr 21;24(16):3985–98. [PubMed: 15102914]
29. Atallah BV, Scanziani M. Instantaneous modulation of gamma oscillation frequency by balancing excitation with inhibition. *Neuron*. 2009 May 28;62(4):566–77. [PubMed: 19477157]
30. Haider B, Duque A, Hasenstaub AR, McCormick DA. Neocortical network activity in vivo is generated through a dynamic balance of excitation and inhibition. *J Neurosci*. 2006 Apr 26;26(17):4535–45. [PubMed: 16641233]
31. Okun M, Lampl I. Instantaneous correlation of excitation and inhibition during ongoing and sensory-evoked activities. *Nat Neurosci*. 2008 May;11(5):535–7. [PubMed: 18376400]
32. Rossini PM, Burke D, Chen R, Cohen LG, Daskalakis Z, Di Iorio R, et al. Non-invasive electrical and magnetic stimulation of the brain, spinal cord, roots and peripheral nerves: Basic principles and procedures for routine clinical and research application. An updated report from an I.F.C.N. Committee. *Clin Neurophysiol*. 2015 Jun;126(6):1071–107. [PubMed: 25797650]
33. Di Lazzaro V, Rothwell JC. Corticospinal activity evoked and modulated by non-invasive stimulation of the intact human motor cortex: Corticospinal activity and the human motor cortex. *The Journal of Physiology*. 2014 Oct 1;592(19):4115–28. [PubMed: 25172954]
34. Davila-Pérez P, Jannati A, Fried PJ, Cudeiro Mazaira J, Pascual-Leone A. The Effects of Waveform and Current Direction on the Efficacy and Test–Retest Reliability of Transcranial Magnetic Stimulation. *Neuroscience*. 2018 Nov;393:97–109. [PubMed: 30300705]
35. Kaye HL, Gersner R, Boes AD, Pascual-Leone A, Rotenberg A. Persistent uncrossed corticospinal connections in patients with intractable focal epilepsy. *Epilepsy & Behavior*. 2017 Oct;75:66–71. [PubMed: 28830029]
36. Papadelis C, Kaye H, Shore B, Snyder B, Grant PE, Rotenberg A. Maturation of Corticospinal Tracts in Children With Hemiplegic Cerebral Palsy Assessed by Diffusion Tensor Imaging and Transcranial Magnetic Stimulation. *Front Hum Neurosci*. 2019 Jul 24;13:254. [PubMed: 31396066]
37. Kaye HL, Peters JM, Gersner R, Chamberland M, Sansevere A, Rotenberg A. Neurophysiological evidence of preserved connectivity in tuber tissue. *Epilepsy Behav Case Rep*. 2017;7:64–8. [PubMed: 28616385]
38. Chen R, Cros D, Curra A, Di Lazzaro V, Lefaucheur J-P, Magistris MR, et al. The clinical diagnostic utility of transcranial magnetic stimulation: report of an IFCN committee. *Clin Neurophysiol*. 2008 Mar;119(3):504–32. [PubMed: 18063409]
39. Lamy J-C, Wargon I, Mazevet D, Ghanim Z, Pradat-Diehl P, Katz R. Impaired efficacy of spinal presynaptic mechanisms in spastic stroke patients. *Brain*. 2009 Mar;132(3):734–48. [PubMed: 19036767]
40. Levy WJ, Amassian VE, Schmid UD, Jungreis C. Mapping of motor cortex gyral sites non-invasively by transcranial magnetic stimulation in normal subjects and patients. *Electroencephalogr Clin Neurophysiol Suppl*. 1991;43:51–75. [PubMed: 1773777]
41. Maeda F, Keenan JP, Tormos JM, Topka H, Pascual-Leone A. Modulation of corticospinal excitability by repetitive transcranial magnetic stimulation. *Clin Neurophysiol*. 2000 May;111(5):800–5. [PubMed: 10802449]

42. Pascual-Leone A, Valls-Solé J, Wassermann EM, Hallett M. Responses to rapid-rate transcranial magnetic stimulation of the human motor cortex. *Brain*. 1994;117(4):847–58. [PubMed: 7922470]
43. Wolters A, Ziemann U, Benecke R. The cortical silent period. In: *The Oxford handbook of transcranial stimulation*. Oxford University Press; 2008. p. 91–102.
44. Person R, Kozhina G. Investigation of the silent period by a poststimulus histogram method. *Neurophysiology*. 1978;10(2):123–9.
45. Wolters A, Ziemann U, Benecke R. The cortical silent period. Epstein CM, Wassermann EM, Ziemann U, editors. Vol. 1. Oxford University Press; 2012.
46. Rotenberg A. Measures of cortical excitability by transcranial magnetic stimulation. In: Pearl PL, editor. *Inherited Metabolic Epilepsies*. New York, NY: Demos; 2018. p. 201–6.
47. Inghilleri M, Mattia D, Berardelli A, Manfredi M. Asymmetry of cortical excitability revealed by transcranial stimulation in a patient with focal motor epilepsy and cortical myoclonus. *Electroencephalogr Clin Neurophysiol*. 1998 Feb;109(1):70–2. [PubMed: 11003066]
48. Cicinelli P, Mattia D, Spanedda F, Traversa R, Marciani MG, Pasqualetti P, et al. Transcranial magnetic stimulation reveals an interhemispheric asymmetry of cortical inhibition in focal epilepsy. *Neuroreport*. 2000 Mar 20;11(4):701–7. [PubMed: 10757504]
49. Hamer HM, Reis J, Mueller H-H, Knake S, Overhof M, Oertel WH, et al. Motor cortex excitability in focal epilepsies not including the primary motor area--a TMS study. *Brain*. 2005 Apr;128(Pt 4):811–8. [PubMed: 15728658]
50. Cincotta M, Borgheresi A, Lori S, Fabbri M, Zaccara G. Interictal inhibitory mechanisms in patients with cryptogenic motor cortex epilepsy: a study of the silent period following transcranial magnetic stimulation. *Electroencephalogr Clin Neurophysiol*. 1998 Jul;107(1):1–7. [PubMed: 9743265]
51. Tataroglu C, Ozkiziltan S, Baklan B. Motor cortical thresholds and cortical silent periods in epilepsy. *Seizure*. 2004 Oct;13(7):481–5. [PubMed: 15324826]
52. Macdonell RA, King MA, Newton MR, Curatolo JM, Reutens DC, Berkovic SF. Prolonged cortical silent period after transcranial magnetic stimulation in generalized epilepsy. *Neurology*. 2001 Aug 28;57(4):706–8. [PubMed: 11524485]
53. Devanne H, Lavoie BA, Capaday C. Input-output properties and gain changes in the human corticospinal pathway. *Exp Brain Res*. 1997 Apr;114(2):329–38. [PubMed: 9166922]
54. Boroojerdi B, Battaglia F, Muellbacher W, Cohen LG. Mechanisms influencing stimulus-response properties of the human corticospinal system. *Clin Neurophysiol*. 2001 May;112(5):931–7. [PubMed: 11336911]
55. Ziemann U, Tergau F, Wassermann EM, Wischer S, Hildebrandt J, Paulus W. Demonstration of facilitatory I wave interaction in the human motor cortex by paired transcranial magnetic stimulation. *J Physiol (Lond)*. 1998 Aug 15;511 (Pt 1):181–90. [PubMed: 9679173]
56. Tokimura H, Ridding MC, Tokimura Y, Amassian VE, Rothwell JC. Short latency facilitation between pairs of threshold magnetic stimuli applied to human motor cortex. *Electroencephalogr Clin Neurophysiol*. 1996 Aug;101(4):263–72. [PubMed: 8761035]
57. Hanajima R, Ugawa Y, Terao Y, Enomoto H, Shiio Y, Mochizuki H, et al. Mechanisms of intracortical I-wave facilitation elicited with paired-pulse magnetic stimulation in humans. *J Physiol (Lond)*. 2002 Jan 1;538(Pt 1):253–61. [PubMed: 11773332]
58. Ili TV, Meintzschel F, Cleff U, Ruge D, Kessler KR, Ziemann U. Short-interval paired-pulse inhibition and facilitation of human motor cortex: the dimension of stimulus intensity. *J Physiol (Lond)*. 2002 15;545(1):153–67. [PubMed: 12433957]
59. Moliadze V, Giannikopoulos D, Eysel UT, Funke K. Paired-pulse transcranial magnetic stimulation protocol applied to visual cortex of anaesthetized cat: effects on visually evoked single-unit activity: ppTMS effects on visually evoked activity. *The Journal of Physiology*. 2005 Aug;566(3):955–65. [PubMed: 15919717]
60. Claus D, Weis M, Jahnke U, Plewe A, Brunhölzl C. Corticospinal conduction studied with magnetic double stimulation in the intact human. *J Neurol Sci*. 1992 Sep;111(2):180–8. [PubMed: 1431984]

61. Valls-Sole J, Pascual-Leone A, Wassermann EM, Hallett M. Human motor evoked responses to paired transcranial magnetic stimuli. *Electroencephalogr Clin Neurophysiol.* 1992 Dec;85(6):355–64. [PubMed: 1282453]
62. Nakamura H, Kitagawa H, Kawaguchi Y, Tsuji H. Intracortical facilitation and inhibition after transcranial magnetic stimulation in conscious humans. *J Physiol (Lond).* 1997 Feb 1;498 (Pt 3):817–23. [PubMed: 9051592]
63. Di Lazzaro V, Pilato F, Dileone M, Ranieri F, Ricci V, Profice P, et al. GABA_A receptor subtype specific enhancement of inhibition in human motor cortex: GABA_A receptor subtype-specific enhancement of SICI. *The Journal of Physiology.* 2006 Sep 15;575(3):721–6. [PubMed: 16809358]
64. Lang N, Sueske E, Hasan A, Paulus W, Tergau F. Pregabalin Exerts Oppositional Effects on Different Inhibitory Circuits in Human Motor Cortex: A Double-blind, Placebo-controlled Transcranial Magnetic Stimulation Study. *Epilepsia.* 2006 May;47(5):813–9. [PubMed: 16686645]
65. Sohn YH, Kaelin-Lang A, Jung HY, Hallett M. Effect of levetiracetam on human corticospinal excitability. *Neurology.* 2001 Sep 11;57(5):858–63. [PubMed: 11552017]
66. Ziemann U, Tam A, Büttefisch C, Cohen LG. Dual modulating effects of amphetamine on neuronal excitability and stimulation-induced plasticity in human motor cortex. *Clin Neurophysiol.* 2002 Aug;113(8):1308–15. [PubMed: 12140012]
67. Hsieh T-H, Dhamne SC, Chen J-JJ, Pascual-Leone A, Jensen FE, Rotenberg A. A new measure of cortical inhibition by mechanomyography and paired-pulse transcranial magnetic stimulation in unanesthetized rats. *Journal of Neurophysiology.* 2012 Feb;107(3):966–72. [PubMed: 22013238]
68. Walther M, Berweck S, Schessl J, Linder-Lucht M, Fietzek UM, Glocker FX, et al. Maturation of inhibitory and excitatory motor cortex pathways in children. *Brain Dev.* 2009 Aug;31(7):562–7. [PubMed: 19329268]
69. Daskalakis ZJ, Möller B, Christensen BK, Fitzgerald PB, Gunraj C, Chen R. The effects of repetitive transcranial magnetic stimulation on cortical inhibition in healthy human subjects. *Exp Brain Res.* 2006 Oct;174(3):403–12. [PubMed: 16683138]
70. Chapter Ziemann U. 23 Pharmacology of TMS. In: *Supplements to Clinical Neurophysiology.* Elsevier; 2003. p. 226–31.
71. Bäumer T, Bock F, Koch G, Lange R, Rothwell JC, Siebner HR, et al. Magnetic stimulation of human premotor or motor cortex produces interhemispheric facilitation through distinct pathways: Interhemispheric facilitation through motor and premotor pathways. *The Journal of Physiology.* 2006 May;572(3):857–68. [PubMed: 16497712]
72. Pascual-Leone A, Walsh V. Fast Backprojections from the Motion to the Primary Visual Area Necessary for Visual Awareness. *Science.* 2001;292(5516):510–2. [PubMed: 11313497]
73. Pascual-Leone A, Rubio B, Pallardó F, Catalá MD. Rapid-rate transcranial magnetic stimulation of left dorsolateral prefrontal cortex in drug-resistant depression. *Lancet.* 1996 Jul 27;348(9022):233–7. [PubMed: 8684201]
74. Pascual-Leone A, Gomez-Tortosa E, Grafman J, Alway D, Nichelli P, Hallett M. Induction of visual extinction by rapid-rate transcranial magnetic stimulation of parietal lobe. *Neurology.* 1994 Mar;44(3 Pt 1):494–8. [PubMed: 8145921]
75. Valero-Cabré A, Payne BR, Pascual-Leone A. Opposite impact on 14C-2-deoxyglucose brain metabolism following patterns of high and low frequency repetitive transcranial magnetic stimulation in the posterior parietal cortex. *Exp Brain Res.* 2007 Jan 22;176(4):603–15. [PubMed: 16972076]
76. Fitzgerald PB, Fountain S, Daskalakis ZJ. A comprehensive review of the effects of rTMS on motor cortical excitability and inhibition. *Clinical Neurophysiology.* 2006;117(12):2584–96. [PubMed: 16890483]
77. Huang Y-Z, Edwards MJ, Rounis E, Bhatia KP, Rothwell JC. Theta Burst Stimulation of the Human Motor Cortex. *Neuron.* 2005 Jan 1;45(2):201–6. [PubMed: 15664172]
78. Stagg CJ, Wylezinska M, Matthews PM, Johansen-Berg H, Jezzard P, Rothwell JC, et al. Neurochemical Effects of Theta Burst Stimulation as Assessed by Magnetic Resonance Spectroscopy. *Journal of Neurophysiology.* 2009 Jun 1;101(6):2872–7. [PubMed: 19339458]

79. Wischnewski M, Schutter DJLG. Efficacy and Time Course of Theta Burst Stimulation in Healthy Humans. *Brain Stimulation*. 2015 Jul 8;8(4):685–92. [PubMed: 26014214]
80. Huerta PT, Volpe BT. Transcranial magnetic stimulation, synaptic plasticity and network oscillations. *Journal of NeuroEngineering and Rehabilitation*. 2009;6(1):7. [PubMed: 19254380]
81. Oberman LM, Eldaief M, Fecteau S, Ifert-Miller F, Tormos JM, Pascual-Leone A. Abnormal modulation of corticospinal excitability in adults with Asperger’s syndrome. *European Journal of Neuroscience*. 2012 Sep;36(6):2782–8.
82. Pascual-Leone A, Freitas C, Oberman L, Horvath JC, Halko M, Eldaief M, et al. Characterizing Brain Cortical Plasticity and Network Dynamics Across the Age-Span in Health and Disease with TMS-EEG and TMS-fMRI. *Brain Topography*. 2011 Oct;24(3–4):302–15. [PubMed: 21842407]
83. Jannati A, Block G, Oberman LM, Rotenberg A, Pascual-Leone A. Interindividual variability in response to continuous theta-burst stimulation in healthy adults. *Clinical Neurophysiology*. 2017 Nov;128(11):2268–78. [PubMed: 29028501]
84. Jannati A, Fried PJ, Block G, Oberman LM, Rotenberg A, Pascual-Leone A. Test–retest reliability of the effects of continuous theta-burst stimulation. *Front Neurosci*. 2019 May 17;13:447. [PubMed: 31156361]
85. Fried PJ, Jannati A, Davila-Pérez P, Pascual-Leone A. Reproducibility of Single-Pulse, Paired-Pulse, and Intermittent Theta-Burst TMS Measures in Healthy Aging, Type-2 Diabetes, and Alzheimer’s Disease. *Frontiers in Aging Neuroscience*. 2017 Aug 21;9(263).
86. López-Alonso V, Cheeran B, Río-Rodríguez D, Fernández-Del-Olmo M. Inter-individual variability in response to non-invasive brain stimulation paradigms. *Brain Stimul*. 2014 Jun;7(3):372–80. [PubMed: 24630849]
87. Cárdenas-Morales L, Nowak DA, Kammer T, Wolf RC, Schönfeldt-Lecuona C. Mechanisms and applications of theta-burst rTMS on the human motor cortex. *Brain Topogr*. 2010 Jan;22(4):294–306. [PubMed: 19288184]
88. Huang Y-Z, Chen R-S, Rothwell JC, Wen H-Y. The after-effect of human theta burst stimulation is NMDA receptor dependent. *Clinical Neurophysiology*. 2007 May;118(5):1028–32. [PubMed: 17368094]
89. Stefan K, Kunesch E, Cohen LG, Benecke R, Classen J. Induction of plasticity in the human motor cortex by paired associative stimulation. *Brain*. 2000 Mar 1;123(3):572–84. [PubMed: 10686179]
90. Müller-Dahlhaus JFM, Orekhov Y, Liu Y, Ziemann U. Interindividual variability and age-dependency of motor cortical plasticity induced by paired associative stimulation. *Experimental Brain Research*. 2008 May;187(3):467–75. [PubMed: 18320180]
91. Hebb DO. *The organization of behavior: A neuropsychological theory*. Psychology Press; 2005.
92. Wolters A, Sandbrink F, Schlottmann A, Kunesch E, Stefan K, Cohen LG, et al. A Temporally Asymmetric Hebbian Rule Governing Plasticity in the Human Motor Cortex. *Journal of Neurophysiology*. 2003 Jan 15;89(5):2339–45. [PubMed: 12612033]
93. Classen J, Wolters A, Stefan K, Wycislo M, Sandbrink F, Schmidt A, et al. Chapter 59 Paired associative stimulation. In: *Supplements to Clinical Neurophysiology*. Elsevier; 2004. p. 563–9.
94. Di Lazzaro V, Dileone M, Profice P, Pilato F, Oliviero A, Mazzone P, et al. LTD-like plasticity induced by paired associative stimulation: direct evidence in humans. *Experimental Brain Research*. 2009 Apr;194(4):661–4. [PubMed: 19319509]
95. Ziemann U, Ili TV, Ili TV, Pauli C, Meintzschel F, Ruge D. Learning modifies subsequent induction of long-term potentiation-like and long-term depression-like plasticity in human motor cortex. *J Neurosci*. 2004 Feb 18;24(7):1666–72. [PubMed: 14973238]
96. Christensen DL, Braun KVN, Baio J, Bilder D, Charles J, Constantino JN, et al. Prevalence and Characteristics of Autism Spectrum Disorder Among Children Aged 8 Years — Autism and Developmental Disabilities Monitoring Network, 11 Sites, United States, 2012. *MMWR Surveill Summ*. 2018 Nov 16;65(13):1–23. [PubMed: 30439868]
97. American Psychiatric Association. *Diagnostic and statistical manual of mental disorders (DSM-5®)*. American Psychiatric Pub; 2013.
98. Cole EJ, Enticott PG, Oberman LM, Gwynette MF, Casanova MF, Jackson SLJ, et al. The Potential of Repetitive Transcranial Magnetic Stimulation for Autism Spectrum Disorder: A Consensus Statement. *Biol Psychiatry*. 2019 Feb 15;85(4):e21–2. [PubMed: 30103951]

99. Gogolla N, Leblanc JJ, Quast KB, Südhof TC, Fagiolini M, Hensch TK. Common circuit defect of excitatory-inhibitory balance in mouse models of autism. *J Neurodev Disord*. 2009 Jun;1(2):172–81. [PubMed: 20664807]
100. Nelson SB, Valakh V. Excitatory/Inhibitory Balance and Circuit Homeostasis in Autism Spectrum Disorders. *Neuron*. 2015 Aug 19;87(4):684–98. [PubMed: 26291155]
101. Rubenstein JLR, Merzenich MM. Model of autism: increased ratio of excitation/inhibition in key neural systems. *Genes Brain Behav*. 2003 Oct;2(5):255–67. [PubMed: 14606691]
102. Théoret H, Halligan E, Kobayashi M, Fregni F, Tager-Flusberg H, Pascual-Leone A. Impaired motor facilitation during action observation in individuals with autism spectrum disorder. *Curr Biol*. 2005 Feb 8;15(3):R84–85. [PubMed: 15694294]
103. Enticott PG, Kennedy HA, Rinehart NJ, Tonge BJ, Bradshaw JL, Fitzgerald PB. GABAergic activity in autism spectrum disorders: an investigation of cortical inhibition via transcranial magnetic stimulation. *Neuropharmacology*. 2013;68:202–9. [PubMed: 22727823]
104. Enticott PG, Kennedy HA, Rinehart NJ, Tonge BJ, Bradshaw JL, Taffe JR, et al. Mirror neuron activity associated with social impairments but not age in autism spectrum disorder. *Biol Psychiatry*. 2012 Mar 1;71(5):427–33. [PubMed: 21974786]
105. Jung NH, Janzarik WG, Delvendahl I, Münchau A, Biscaldi M, Mainberger F, et al. Impaired induction of long-term potentiation-like plasticity in patients with high-functioning autism and Asperger syndrome: Long-term Potentiation-like Plasticity in ASD. *Developmental Medicine & Child Neurology*. 2013 Jan;55(1):83–9. [PubMed: 23157428]
106. Pedapati EV, Gilbert DL, Erickson CA, Horn PS, Shaffer RC, Wink LK, et al. Abnormal Cortical Plasticity in Youth with Autism Spectrum Disorder: A Transcranial Magnetic Stimulation Case-Control Pilot Study. *J Child Adolesc Psychopharmacol*. 2016;26(7):625–31. [PubMed: 27007257]
107. Pedapati EV, Mooney LN, Wu SW, Erickson CA, Sweeney JA, Shaffer RC, et al. Motor cortex facilitation: a marker of attention deficit hyperactivity disorder co-occurrence in autism spectrum disorder. *Transl Psychiatry*. 2019 13;9(1):298. [PubMed: 31723120]
108. Oberman LM, Ifert-Miller F, Najib U, Bashir S, Gonzalez-Heydrich J, Picker J, et al. Abnormal Mechanisms of Plasticity and Metaplasticity in Autism Spectrum Disorders and Fragile X Syndrome. *Journal of Child and Adolescent Psychopharmacology*. 2016 Sep;26(7):617–24. [PubMed: 27218148]
109. Bienenstock EL, Cooper LN, Munro PW. Theory for the development of neuron selectivity: orientation specificity and binocular interaction in visual cortex. *J Neurosci*. 1982 Jan;2(1):32–48. [PubMed: 7054394]
110. Oberman LM, Rotenberg A, Pascual-Leone A. Aberrant brain plasticity in autism spectrum disorders. In: *Cognitive Plasticity in Neurologic Disorders*. 2015. p. 176–96.
111. Oberman LM, Pascual-Leone A, Rotenberg A. Modulation of corticospinal excitability by transcranial magnetic stimulation in children and adolescents with autism spectrum disorder. *Frontiers in Human Neuroscience*. 2014 Jan 1;8(5):627. [PubMed: 25165441]
112. Jannati A, Block G, Ryan MA, Kaye HL, Kayarian FB, Bashir S, et al. Continuous theta-burst stimulation in children with high-functioning autism spectrum disorder and typically developing children. *Front Integr Neurosci*. 2020 Mar 13;14:13. [PubMed: 32231523]
113. Trippe J, Mix A, Aydin-Abidin S, Funke K, Benali A. Theta burst and conventional low-frequency rTMS differentially affect GABAergic neurotransmission in the rat cortex. *Experimental Brain Research*. 2009 Dec;199(3–4):411–21. [PubMed: 19701632]
114. Ben-Ari Y, Khalilov I, Kahle KT, Cherubini E. The GABA excitatory/inhibitory shift in brain maturation and neurological disorders. *Neuroscientist*. 2012 Oct;18(5):467–86. [PubMed: 22547529]
115. Coghlan S, Horder J, Inkster B, Mendez MA, Murphy DG, Nutt DJ. GABA system dysfunction in autism and related disorders: from synapse to symptoms. *Neurosci Biobehav Rev*. 2012 Oct;36(9):2044–55. [PubMed: 22841562]
116. LeBlanc JJ, Fagiolini M. Autism: A “Critical Period” Disorder? Neural Plasticity [Internet]. 2011 [cited 2019 Jun 11];2011(921680). Available from: <http://www.hindawi.com/journals/np/2011/921680/>

117. Lemonnier E, Villeneuve N, Sonie S, Serret S, Rosier A, Roue M, et al. Effects of bumetanide on neurobehavioral function in children and adolescents with autism spectrum disorders. *Transl Psychiatry*. 2017 14;7(3):e1056. [PubMed: 28291262]
118. Hersh JH, Saul RA, Committee on Genetics. Health supervision for children with fragile X syndrome. *Pediatrics*. 2011 May;127(5):994–1006. [PubMed: 21518720]
119. Hunter J, Rivero-Arias O, Angelov A, Kim E, Fotheringham I, Leal J. Epidemiology of fragile X syndrome: a systematic review and meta-analysis. *Am J Med Genet A*. 2014 Jul;164A(7):1648–58. [PubMed: 24700618]
120. Kaufmann WE, Cortell R, Kau ASM, Bukelis I, Tierney E, Gray RM, et al. Autism spectrum disorder in fragile X syndrome: communication, social interaction, and specific behaviors. *Am J Med Genet A*. 2004 Sep 1;129A(3):225–34. [PubMed: 15326621]
121. Penagarikano O, Mulle JG, Warren ST. The pathophysiology of fragile x syndrome. *Annu Rev Genomics Hum Genet*. 2007;8:109–29. [PubMed: 17477822]
122. Huber KM, Gallagher SM, Warren ST, Bear MF. Altered synaptic plasticity in a mouse model of fragile X mental retardation. *Proc Natl Acad Sci USA*. 2002 May 28;99(11):7746–50. [PubMed: 12032354]
123. Simonyi A, Schachtman TR, Christoffersen GRJ. The role of metabotropic glutamate receptor 5 in learning and memory processes. *Drug News Perspect*. 2005 Aug;18(6):353–61. [PubMed: 16247513]
124. Oberman L, Ifert-Miller F, Najib U, Bashir S, Woollacott I, Gonzalez-Heydrich J, et al. Transcranial magnetic stimulation provides means to assess cortical plasticity and excitability in humans with fragile X syndrome and autism spectrum disorder. *Frontiers in Synaptic Neuroscience*. 2010;2:26. [PubMed: 21423512]
125. Rubio B, Boes AD, Laganieri S, Rotenberg A, Jeurissen D, Pascual-Leone A. Noninvasive Brain Stimulation in Pediatric Attention-Deficit Hyperactivity Disorder (ADHD): A Review. *J Child Neurol*. 2015 Dec 10;
126. Sonuga-Barke EJS. Causal Models of Attention-Deficit/Hyperactivity Disorder: From Common Simple Deficits to Multiple Developmental Pathways. *Biological Psychiatry*. 2005 Jun;57(11):1231–8. [PubMed: 15949993]
127. Gilbert DL, Isaacs KM, Augusta M, MacNeil LK, Mostofsky SH. Motor cortex inhibition: A marker of ADHD behavior and motor development in children. *Neurology*. 2011 Feb 15;76(7):615–21. [PubMed: 21321335]
128. Alexander GE, Crutcher MD, DeLong MR. Chapter 6 Basal ganglia-thalamocortical circuits: Parallel substrates for motor, oculomotor, “prefrontal” and “limbic” functions. In: *Progress in Brain Research*. Elsevier; 1991. p. 119–46.
129. Gilbert DL, Huddleston DA, Wu SW, Pedapati EV, Horn PS, Hirabayashi K, et al. Motor cortex inhibition and modulation in children with ADHD. *Neurology*. 2019;93(6):e599–610. [PubMed: 31315973]
130. Dutra T, Baltar A, Monte-Silva K. Motor cortex excitability in attention-deficit hyperactivity disorder (ADHD): A systematic review and meta-analysis. *Research in developmental disabilities*. 2016;56:1–9. [PubMed: 27240241]
131. Buchmann J, Gierow W, Weber S, Hoepfner J, Klauer T, Benecke R, et al. Restoration of Disturbed Intracortical Motor Inhibition and Facilitation in Attention Deficit Hyperactivity Disorder Children by Methylphenidate. *Biological Psychiatry*. 2007 Nov;62(9):963–9. [PubMed: 17719015]
132. Buchmann J, Wolters A, Haessler F, Bohne S, Nordbeck R, Kunesch E. Disturbed transcallosally mediated motor inhibition in children with attention deficit hyperactivity disorder (ADHD). *Clin Neurophysiol*. 2003 Nov;114(11):2036–42. [PubMed: 14580601]
133. Bruxel EM, Akutagava-Martins GC, Salatino-Oliveira A, Contini V, Kieling C, Hutz MH, et al. ADHD pharmacogenetics across the life cycle: New findings and perspectives. *Am J Med Genet*. 2014 Jun;165(4):263–82.
134. Chen R, Tam A, Bütefisch C, Corwell B, Ziemann U, Rothwell JC, et al. Intracortical inhibition and facilitation in different representations of the human motor cortex. *J Neurophysiol*. 1998 Dec;80(6):2870–81. [PubMed: 9862891]

135. Trompetto C, Buccolieri A, Marchese R, Marinelli L, Michelozzi G, Abbruzzese G. Impairment of transcallosal inhibition in patients with corticobasal degeneration. *Clin Neurophysiol.* 2003 Nov;114(11):2181–7. [PubMed: 14580617]
136. Di Martino A, Zuo X-N, Kelly C, Grzadzinski R, Mennes M, Schvarcz A, et al. Shared and distinct intrinsic functional network centrality in autism and attention-deficit/hyperactivity disorder. *Biol Psychiatry.* 2013 Oct 15;74(8):623–32. [PubMed: 23541632]
137. Robertson MM. The prevalence and epidemiology of Gilles de la Tourette syndrome. *Journal of Psychosomatic Research.* 2008 Nov;65(5):461–72. [PubMed: 18940377]
138. Robertson MM, Cavanna AE, Eapen V. Gilles de la Tourette syndrome and disruptive behavior disorders: prevalence, associations, and explanation of the relationships. *J Neuropsychiatry Clin Neurosci.* 2015;27(1):33–41. [PubMed: 25162416]
139. Du J-C, Chiu T-F, Lee K-M, Wu H-L, Yang Y-C, Hsu S-Y, et al. Tourette syndrome in children: an updated review. *Pediatr Neonatol.* 2010 Oct;51(5):255–64. [PubMed: 20951354]
140. Orth M, Münchau A. Transcranial magnetic stimulation studies of sensorimotor networks in Tourette syndrome. *Behav Neurol.* 2013;27(1):57–64. [PubMed: 23187144]
141. Ziemann U, Paulus W, Rothenberger A. Decreased motor inhibition in Tourette's disorder: evidence from transcranial magnetic stimulation. *Am J Psychiatry.* 1997 Sep;154(9):1277–84. [PubMed: 9286189]
142. Orth M, Amann B, Robertson MM, Rothwell JC. Excitability of motor cortex inhibitory circuits in Tourette syndrome before and after single dose nicotine. *Brain.* 2005 Jun;128(Pt 6):1292–300. [PubMed: 15774505]
143. Orth M, Münchau A, Rothwell JC. Corticospinal system excitability at rest is associated with tic severity in tourette syndrome. *Biol Psychiatry.* 2008 Aug 1;64(3):248–51. [PubMed: 18243162]
144. Heise K-F, Steven B, Liuzzi G, Thomalla G, Jonas M, Muller-Vahl K, et al. Altered modulation of intracortical excitability during movement preparation in Gilles de la Tourette syndrome. *Brain.* 2010 Feb 1;133(2):580–90. [PubMed: 20008030]
145. Gilbert DL, Bansal AS, Sethuraman G, Sallee FR, Zhang J, Lipps T, et al. Association of cortical disinhibition with tic, ADHD, and OCD severity in Tourette syndrome. *Mov Disord.* 2004 Apr;19(4):416–25. [PubMed: 15077239]
146. Gilbert DL, Sallee FR, Zhang J, Lipps TD, Wassermann EM. Transcranial magnetic stimulation-evoked cortical inhibition: a consistent marker of attention-deficit/hyperactivity disorder scores in tourette syndrome. *Biol Psychiatry.* 2005 Jun 15;57(12):1597–600. [PubMed: 15953499]
147. Orth M, Rothwell JC. Motor cortex excitability and comorbidity in Gilles de la Tourette syndrome. *J Neurol Neurosurg Psychiatry.* 2009 Jan;80(1):29–34. [PubMed: 18931001]
148. Greenberg BD, Ziemann U, Corá-Locatelli G, Harmon A, Murphy DL, Keel JC, et al. Altered cortical excitability in obsessive-compulsive disorder. *Neurology.* 2000 Jan 11;54(1):142–7. [PubMed: 10636140]
149. Orth M, Snijders AH, Rothwell JC. The variability of intracortical inhibition and facilitation. *Clin Neurophysiol.* 2003 Dec;114(12):2362–9. [PubMed: 14652096]
150. Orth M, Rothwell JC. The cortical silent period: intrinsic variability and relation to the waveform of the transcranial magnetic stimulation pulse. *Clin Neurophysiol.* 2004 May;115(5):1076–82. [PubMed: 15066533]
151. Jackson SR, Parkinson A, Manfredi V, Millon G, Hollis C, Jackson GM. Motor excitability is reduced prior to voluntary movements in children and adolescents with Tourette syndrome. *J Neuropsychol.* 2013 Mar;7(1):29–44. [PubMed: 22804795]
152. Draper A, Jude L, Jackson GM, Jackson SR. Motor excitability during movement preparation in Tourette syndrome. *J Neuropsychol.* 2015 Mar;9(1):33–44. [PubMed: 24283505]
153. Ganos C, Rocchi L, Latorre A, Hockey L, Palmer C, Joyce EM, et al. Motor cortical excitability during voluntary inhibition of involuntary tic movements. *Mov Disord.* 2018;33(11):1804–9. [PubMed: 30379360]
154. Pépés SE, Draper A, Jackson GM, Jackson SR. Effects of age on motor excitability measures from children and adolescents with Tourette syndrome. *Dev Cogn Neurosci.* 2016;19:78–86. [PubMed: 26934638]

155. Suppa A, Belvisi D, Bologna M, Marsili L, Berardelli I, Moretti G, et al. Abnormal cortical and brain stem plasticity in Gilles de la Tourette syndrome. *Mov Disord*. 2011 Aug 1;26(9):1703–10. [PubMed: 21442662]
156. Wu SW, Gilbert DL. Altered neurophysiologic response to intermittent theta burst stimulation in Tourette syndrome. *Brain Stimul*. 2012 Jul;5(3):315–9. [PubMed: 22037119]
157. Suppa A, Marsili L, Di Stasio F, Berardelli I, Roselli V, Pasquini M, et al. Cortical and brainstem plasticity in Tourette syndrome and obsessive-compulsive disorder. *Mov Disord*. 2014 Oct;29(12):1523–31. [PubMed: 24996148]
158. Brandt VC, Niessen E, Ganos C, Kahl U, Bäumer T, Münchau A. Altered synaptic plasticity in Tourette's syndrome and its relationship to motor skill learning. *PLoS ONE*. 2014;9(5):e98417. [PubMed: 24878665]
159. Bloodstein O. *A handbook on stuttering*. 5th ed. San Diego, Calif: Singular Pub. Group; 1995. 586 p.
160. Mulligan HF, Anderson TJ, Jones RD, Williams MJ, Donaldson IM. Tics and developmental stuttering. *Parkinsonism Relat Disord*. 2003 Jun;9(5):281–9. [PubMed: 12781595]
161. Riva-Posse P, Busto-Marolt L, Scheitschnaider A, Martinez-Echenique L, Cammarota A, Merello M. Phenomenology of abnormal movements in stuttering. *Parkinsonism Relat Disord*. 2008;14(5):415–9. [PubMed: 18316236]
162. Yairi E, Ambrose NG. *Early childhood stuttering for clinicians by clinicians*. Austin, Tex: PRO-ED; 2005. 521 p. (For clinicians by clinicians).
163. Sommer M, Wischer S, Tergau F, Paulus W. Normal intracortical excitability in developmental stuttering. *Mov Disord*. 2003 Jul;18(7):826–30. [PubMed: 12815664]
164. Busan P, D'Ausilio A, Borelli M, Monti F, Pelamatti G, Pizzolato G, et al. Motor excitability evaluation in developmental stuttering: a transcranial magnetic stimulation study. *Cortex*. 2013 Mar;49(3):781–92. [PubMed: 22225881]
165. Alm PA, Karlsson R, Sundberg M, Axelson HW. Hemispheric Lateralization of Motor Thresholds in Relation to Stuttering. Holmes NP, editor. *PLoS ONE*. 2013 Oct 11;8(10):e76824. [PubMed: 24146930]
166. Neef NE, Jung K, Rothkegel H, Pollok B, von Gudenberg AW, Paulus W, et al. Right-shift for non-speech motor processing in adults who stutter. *Cortex*. 2011 Sep;47(8):945–54. [PubMed: 20822768]
167. Neef NE, Paulus W, Neef A, von Gudenberg AW, Sommer M. Reduced intracortical inhibition and facilitation in the primary motor tongue representation of adults who stutter. *Clin Neurophysiol*. 2011 Sep;122(9):1802–11. [PubMed: 21377925]
168. Neef NE, Hoang TNL, Neef A, Paulus W, Sommer M. Speech dynamics are coded in the left motor cortex in fluent speakers but not in adults who stutter. *Brain*. 2015 Mar;138(3):712–25. [PubMed: 25595146]
169. Busan P, Del Ben G, Bernardini S, Ntarelli G, Bencich M, Monti F, et al. Altered Modulation of Silent Period in Tongue Motor Cortex of Persistent Developmental Stuttering in Relation to Stuttering Severity. Avenanti A, editor. *PLoS ONE*. 2016 Oct 6;11(10):e0163959. [PubMed: 27711148]
170. Busan P, Battaglini PP, Sommer M. Transcranial magnetic stimulation in developmental stuttering: Relations with previous neurophysiological research and future perspectives. *Clin Neurophysiol*. 2017;128(6):952–64. [PubMed: 28431323]
171. Chang S-E, Horwitz B, Ostuni J, Reynolds R, Ludlow CL. Evidence of Left Inferior Frontal–Premotor Structural and Functional Connectivity Deficits in Adults Who Stutter. *Cerebral Cortex*. 2011 Nov;21(11):2507–18. [PubMed: 21471556]
172. Chang S-E, Erickson KI, Ambrose NG, Hasegawa-Johnson MA, Ludlow CL. Brain anatomy differences in childhood stuttering. *Neuroimage*. 2008 Feb 1;39(3):1333–44. [PubMed: 18023366]
173. Brainsway Deep TMS System [Internet]. Food and Drug Administration Web site. 2013 [cited 2016 Feb 1]. Available from: https://www.accessdata.fda.gov/cdrh_docs/pdf12/K122288.pdf
174. Magstim Rapid2 Therapy System [Internet]. Food and Drug Administration Web site. 2015 [cited 2016 Feb 1]. Available from: https://www.accessdata.fda.gov/cdrh_docs/pdf14/K143531.pdf

175. Magstim Horizon TMS Therapy System with Navigation [Internet]. Food and Drug Administration Web site. 2019 [cited 2019 Jun 15]. Available from: https://www.accessdata.fda.gov/cdrh_docs/pdf18/K183376.pdf
176. MagVenture MagVita TMS Therapy System w/Theta Burst Stimulation [Internet]. Food and Drug Administration Web site. 2018 [cited 2019 Jun 15]. Available from: https://www.accessdata.fda.gov/cdrh_docs/pdf17/K173620.pdf
177. TeleEMG Neurosoft TMS [Internet]. Food and Drug Administration Web site. 2016 [cited 2019 Jun 15]. Available from: https://www.accessdata.fda.gov/cdrh_docs/pdf16/K160309.pdf
178. Mag & More Apollo TMS Therapy System [Internet]. Food and Drug Administration Web site. 2018 [cited 2019 Jun 15]. Available from: https://www.accessdata.fda.gov/cdrh_docs/pdf18/K180313.pdf
179. Nexstim Navigated Brain Therapy (NBT) System 2 [Internet]. Food and Drug Administration Web site. 2018 [cited 2019 Jun 15]. Available from: https://www.accessdata.fda.gov/cdrh_docs/pdf18/K182700.pdf
180. eNeura Spring TMS [Internet]. Food and Drug Administration Web site. 2016 [cited 2019 Jun 15]. Available from: https://www.accessdata.fda.gov/cdrh_docs/pdf16/K162797.pdf
181. Nexstim eXimia Navigated Brain Stimulation System [Internet]. Food and Drug Administration Web site. 2009 [cited 2016 Feb 1]. Available from: https://www.accessdata.fda.gov/cdrh_docs/pdf9/K091457.pdf
182. Nexstim Navigational Brain Stimulation (NBS) System 4, and Nexstim NBS System 4 with NEXSPEECH® [Internet]. Food and Drug Administration Web site. 2011 [cited 2016 Feb 1]. Available from: https://www.accessdata.fda.gov/cdrh_docs/pdf11/K112881.pdf
183. Brainsway Deep Transcranial Magnetic Stimulation System [Internet]. Food and Drug Administration Web site. 2017 [cited 2019 Jun 15]. Available from: https://www.accessdata.fda.gov/cdrh_docs/reviews/DEN170078.pdf
184. Peruzzotti-Jametti L, Bacigaluppi M, Sandrone S, Cambiaghi M. Emerging subspecialties in Neurology: transcranial stimulation. *Neurology*. 2013 Jan 22;80(4):e33–35. [PubMed: 23339212]
185. Williams NR, Taylor JJ, Snipes JM, Short EB, Kantor EM, George MS. Interventional psychiatry: how should psychiatric educators incorporate neuromodulation into training? *Acad Psychiatry*. 2014 Apr;38(2):168–76. [PubMed: 24554501]
186. Boes AD, Kelly MS, Trapp NT, Stern AP, Press DZ, Pascual-Leone A. Noninvasive Brain Stimulation: Challenges and Opportunities for a New Clinical Specialty. *J Neuropsychiatry Clin Neurosci*. 2018;30(3):173–9. [PubMed: 29685065]
187. Lefaucheur J-P, André-Obadia N, Antal A, Ayache SS, Baeken C, Benninger DH, et al. Evidence-based guidelines on the therapeutic use of repetitive transcranial magnetic stimulation (rTMS). *CLINICAL NEUROPHYSIOLOGY*. 2014 Nov 1;125(11):2150–206. [PubMed: 25034472]
188. Rossi S, Hallett M, Rossini PM, Pascual-Leone A. Safety, ethical considerations, and application guidelines for the use of transcranial magnetic stimulation in clinical practice and research. *Clin Neurophysiol*. 2009 Dec;120(12):2008–39. [PubMed: 19833552]
189. Loo CK, McFarquhar TF, Mitchell PB. A review of the safety of repetitive transcranial magnetic stimulation as a clinical treatment for depression. *Int J Neuropsychopharmacol*. 2008 Feb;11(1):131–47. [PubMed: 17880752]
190. Janicak PG, Dokucu ME. Transcranial magnetic stimulation for the treatment of major depression. *Neuropsychiatr Dis Treat*. 2015;11:1549–60. [PubMed: 26170668]
191. Maeda F, Keenan JP, Tormos JM, Topka H, Pascual-Leone A. Interindividual variability of the modulatory effects of repetitive transcranial magnetic stimulation on cortical excitability. *Experimental Brain Research*. 2000 Aug 4;133(4):425–30. [PubMed: 10985677]
192. Maeda F, Gangitano M, Thall M, Pascual-Leone A. Inter- and intra-individual variability of paired-pulse curves with transcranial magnetic stimulation (TMS). *Clinical Neurophysiology*. 2002;113(3):376–82. [PubMed: 11897538]
193. Freitas C, Perez J, Knobel M, Tormos JM, Oberman L, Eldaief M, et al. Changes in Cortical Plasticity Across the Lifespan. *Frontiers in Aging Neuroscience*. 2011;3.

194. Corp DT, Bereznicki HGK, Clark GM, Youssef GJ, Fried PJ, Jannati A, et al. Large-scale analysis of interindividual variability in theta-burst stimulation data: Results from the “Big TMS Data Collaboration.” *Brain Stimul.* 2020 Aug 3;
195. Huber TJ, Schneider U, Rollnik J. Gender differences in the effect of repetitive transcranial magnetic stimulation in schizophrenia. *Psychiatry research.* 2003;120(1):103–5. [PubMed: 14500119]
196. De Gennaro L, Bertini M, Pauri F, Cristiani R, Curcio G, Ferrara M, et al. Callosal effects of transcranial magnetic stimulation (TMS): the influence of gender and stimulus parameters. *Neuroscience research.* 2004;48(2):129–37. [PubMed: 14741387]
197. Cahn SD, Herzog AG, Pascual-Leone A. Paired-pulse transcranial magnetic stimulation: effects of hemispheric laterality, gender, and handedness in normal controls. *Journal of clinical neurophysiology.* 2003;20(5):371–4. [PubMed: 14701998]
198. Cheeran BJ, Talelli P, Mori F, Koch G, Suppa A, Edwards M, et al. A common polymorphism in the brain-derived neurotrophic factor gene (BDNF) modulates human cortical plasticity and the response to rTMS. *The Journal of Physiology.* 2008 Dec 1;586(23):5717–25. [PubMed: 18845611]
199. Antal A, Chaieb L, Moliadze V, Monte-Silva K, Poreisz C, Thirugnanasambandam N, et al. Brain-derived neurotrophic factor (BDNF) gene polymorphisms shape cortical plasticity in humans. *Brain Stimulation.* 2010 Oct;3(4):230–7. [PubMed: 20965453]
200. Chang WH, Bang OY, Shin Y-I, Lee A, Pascual-Leone A, Kim Y-H. BDNF Polymorphism and Differential rTMS Effects on Motor Recovery of Stroke Patients. *Brain Stimulation.* 2014 Jul;7(4):553–8. [PubMed: 24767962]
201. Di Lazzaro V, Pellegrino G, Di Pino G, Corbetta M, Ranieri F, Brunelli N, et al. Val66Met BDNF Gene Polymorphism Influences Human Motor Cortex Plasticity in Acute Stroke. *Brain Stimulation.* 2015 Jan;8(1):92–6. [PubMed: 25241287]
202. Peña-Gomez C, Solé-Padullés C, Clemente IC, Junqué C, Bargalló N, Bosch B, et al. APOE Status Modulates the Changes in Network Connectivity Induced by Brain Stimulation in Non-Demented Elders. *Theoret H, editor. PLoS ONE.* 2012 Dec 19;7(12):e51833. [PubMed: 23284783]
203. Corp DT, Bereznicki HG, Clark GM, Youssef GJ, Fried PJ, Jannati A, et al. Large-scale analysis of interindividual variability in single and paired-pulse TMS data: Results from the ‘Big TMS Data Collaboration.’ *Brain Stimulation.* In Press;
204. Morris TP, Davila-Pérez P, Jannati A, Menardi A, Pascual-Leone A, Fried PJ. Aftereffects of Intermittent Theta-Burst Stimulation in Adjacent, Non-Target Muscles. *Neuroscience.* 2019 Aug 31;418:157–165. [PubMed: 31476358]
205. Cohen DA, Pascual-Leone A, Press DZ, Robertson EM. Off-line learning of motor skill memory: A double dissociation of goal and movement. *Proceedings of the National Academy of Sciences.* 2005 Dec 13;102(50):18237–41.
206. Julkunen P, Säisänen L, Danner N, Niskanen E, Hukkanen T, Mervaala E, et al. Comparison of navigated and non-navigated transcranial magnetic stimulation for motor cortex mapping, motor threshold and motor evoked potentials. *NeuroImage.* 2009 Feb;44(3):790–5. [PubMed: 18976714]
207. Foucher J, Lorgouilloux K, Turek J, Pham B-T, Elowe J, Bayle B, et al. Robotic assistance in coil positioning improves reliability and comfort. In: 3rd Annual Conference of the German Society for Brain Stimulation - Modulating Emotions. Berlin, Germany; 2012.
208. Civardi C, Boccagni C, Vicentini R, Bolamperti L, Tarletti R, Varrasi C, et al. Cortical excitability and sleep deprivation: a transcranial magnetic stimulation study. *J Neurol Neurosurg Psychiatry.* 2001 Dec 1;71(6):809. [PubMed: 11723210]
209. Kreuzer P, Langguth B, Popp R, Raster R, Busch V, Frank E, et al. Reduced intra-cortical inhibition after sleep deprivation: A transcranial magnetic stimulation study. *Neuroscience Letters.* 2011 Apr 15;493(3):63–6. [PubMed: 21352891]
210. Specterman M, Bhuiya A, Kuppaswamy A, Strutton P, Catley M, Davey N. The effect of an energy drink containing glucose and caffeine on human corticospinal excitability. *Physiology & Behavior.* 2005 Jan 17;83(5):723–8. [PubMed: 15639157]

211. Cerqueira V, de Mendonça A, Minez A, Dias AR, de Carvalho M. Does caffeine modify corticomotor excitability? *Neurophysiologie Clinique/Clinical Neurophysiology*. 2006 Jul;36(4):219–26. [PubMed: 17095411]
212. Badawy RAB, Vogrin SJ, Lai A, Cook MJ. Cortical excitability changes correlate with fluctuations in glucose levels in patients with epilepsy. *Epilepsy & Behavior*. 2013 Jun;27(3):455–60. [PubMed: 23603690]
213. Samii A, Wassermann EM, Hallett M. Post-exercise depression of motor evoked potentials as a function of exercise duration. *Electroencephalography and Clinical Neurophysiology/Electromyography and Motor Control*. 1997 Oct;105(5):352–6. [PubMed: 9362999]
214. Lentz M, Nielsen JF. Post-exercise facilitation and depression of M wave and motor evoked potentials in healthy subjects. *Clin Neurophysiol*. 2002 Jul;113(7):1092–8. [PubMed: 12088705]
215. Smith MJ, Keel JC, Greenberg BD, Adams LF, Schmidt PJ, Rubinow DA, et al. Menstrual cycle effects on cortical excitability. *Neurology*. 1999;53(9):2069–2069. [PubMed: 10599783]
216. Hattemer K, Knake S, Reis J, Rochon J, Oertel WH, Rosenow F, et al. Excitability of the motor cortex during ovulatory and anovulatory cycles: a transcranial magnetic stimulation study. *Clinical Endocrinology*. 2007;66(3):387–93. [PubMed: 17302873]
217. Zrenner C, Belardinelli P, Müller-Dahlhaus F, Ziemann U. Closed-Loop Neuroscience and Non-Invasive Brain Stimulation: A Tale of Two Loops. *Frontiers in Cellular Neuroscience* [Internet]. 2016 Apr 7 [cited 2019 Mar 14];10. Available from: <http://journal.frontiersin.org/Article/10.3389/fncel.2016.00092/abstract>
218. Zrenner C, Desideri D, Belardinelli P, Ziemann U. Real-time EEG-defined excitability states determine efficacy of TMS-induced plasticity in human motor cortex. *Brain Stimulation*. 2018 Mar;11(2):374–89. [PubMed: 29191438]
219. Fox MD. Mapping Symptoms to Brain Networks with the Human Connectome. *N Engl J Med*. 2018 06;379(23):2237–45. [PubMed: 30575457]
220. Weigand A, Horn A, Caballero R, Cooke D, Stern AP, Taylor SF, et al. Prospective validation that subgenual connectivity predicts antidepressant efficacy of transcranial magnetic stimulation sites. *Biological psychiatry*. 2017;
221. Horn A, Fox MD. Opportunities of connectomic neuromodulation. *NeuroImage*. 2020 Nov;221:117180. [PubMed: 32702488]
222. Cash RFH, Cocchi L, Anderson R, Rogachov A, Kucyi A, Barnett AJ, et al. A multivariate neuroimaging biomarker of individual outcome to transcranial magnetic stimulation in depression. *Hum Brain Mapp*. 2019 01;40(16):4618–29. [PubMed: 31332903]
223. Cash RFH, Zalesky A, Thomson RH, Tian Y, Cocchi L, Fitzgerald PB. Subgenual Functional Connectivity Predicts Antidepressant Treatment Response to Transcranial Magnetic Stimulation: Independent Validation and Evaluation of Personalization. *Biol Psychiatry*. 2019 15;86(2):e5–7. [PubMed: 30670304]
224. Eshel N, Keller CJ, Wu W, Jiang J, Mills-Finnerty C, Huemer J, et al. Global connectivity and local excitability changes underlie antidepressant effects of repetitive transcranial magnetic stimulation. *Neuropsychopharmacology*. 2020;45(6):1018–25. [PubMed: 32053828]
225. Brady RO, Gonsalvez I, Lee I, Öngür D, Seidman LJ, Schmahmann JD, et al. Cerebellar-Prefrontal Network Connectivity and Negative Symptoms in Schizophrenia. *Am J Psychiatry*. 2019 01;176(7):512–20. [PubMed: 30696271]

Table 1. Neurophysiological measures derived from transcranial magnetic stimulation-electromyography (TMS-EMG).

TMS-EMG measure	Protocol	Likely mechanism(s)
Resting motor threshold (rMT)	spTMS: measure of minimum stimulus intensity necessary for a motor response at rest (recorded either by visual inspection or EMG) on 5 / 10 trials	Cortical motor neuron voltage-gated sodium channel-mediated membrane excitability
Active motor threshold (aMT)	spTMS: measure of minimum stimulus strength necessary for 200 μ V MEP during isometric contraction on 5 / 10 trials	Cortical motor neuron voltage-gated sodium channel-mediated membrane excitability + spinal contributions
MEP amplitude	spTMS: average peak-to-peak amplitude of MEP over 15–30 single pulses at 5–8 s ITI	Cortical motor neuron voltage-gated sodium channel-mediated membrane excitability
Cortical silent period (cSP)	spTMS: measure of EMG suppression following MEP during voluntary contraction of a contralateral muscle	GABA _B -mediated and GABA _A -mediated motor cortex inhibition
Ipsilateral silent period (ISP)	spTMS: measure of EMG suppression following MEP during voluntary contraction of an ipsilateral muscle	GABA _B -mediated and GABA _A -mediated motor cortex inhibition
Input / Output (I/O) curve	spTMS: measure of slope and plateau of the sigmoid curve of MEP amplitudes obtained at a wide range of stimulus intensities	Cortical motor neuron sodium- and calcium channel-mediated membrane excitability and GABA _A -mediated motor cortex inhibition
Short-interval intracortical inhibition (SICI)	ppTMS: CS = 50–90% rMT < 100% aMT TS = 1 mV 100–120% rMT ISI = 1–6 ms	GABA _A -mediated regional cortical inhibition
Long-interval intracortical inhibition (LICI)	ppTMS: CS = 100–130% rMT TS 1 mV TS = 1 mV 120% rMT ISI = 50–200+ ms	GABA _B -mediated inhibition and (likely) GABA _A -mediated network inhibition
Intracortical facilitation (ICF)	ppTMS: CS = 90+ % rMT > 80% aMT TS = 0.5–1.5 mV ISI = 8–30 ms	Glutamate (NMDA and AMPA receptor types)-mediated excitation
Short-interval intracortical facilitation (SICF)	ppTMS: CS = 100–130% RMT TS 1 mV TS = < rMT 90% rMT ISI = 1–5 ms	Glutamate (non-NMDA receptor type)-mediated excitation
Intermittent theta-burst stimulation (iTBS)	rTMS: 50 Hz bursts of three TMS pulses repeated every 200 ms in a 2-s on, 8-s off pattern for 190 s (a total of 600 pulses)	Glutamate- and GABA-mediated mechanisms
Continuous theta-burst stimulation (cTBS)	rTMS: 50 Hz bursts of three TMS pulses repeated every 200 ms for 40 s (a total of 600 pulses)	Glutamate- and GABA-mediated mechanisms
Paired associative stimulation (PAS)	spTMS + MNS: 90–200+ pairs of median nerve stimulation (MNS) at the wrist and spTMS to contralateral M1 at 7–10 ITI; ISI = 25 ms (PAS ₂₅) 10 ms (PAS ₁₀) N20 latency of MNS-evoked SSEP + 2 ms (PAS _{N20+2})	Glutamate (NMDA receptor type)-mediated excitation

AMPA, α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid; CS, conditioning stimulus; EMG, electromyography; GABA, γ -aminobutyric acid; ICF, intracortical facilitation; ISI, interstimulus interval; ITI, intertrial interval; M1, primary motor cortex; MEP, motor evoked potential; NMDA; *N*-methyl-*D*-aspartate; ppTMS, paired-pulse TMS; rTMS, repetitive TMS; SICF, short-interval intracortical

Author Manuscript Author Manuscript Author Manuscript Author Manuscript Author Manuscript
facilitation; SICI, short-interval intracortical inhibition; SSEP, somatosensory evoked potential; spTMS, single-pulse TMS; TS, test stimulus; TS ## mV stimulator intensity required to elicit MEPs with peak-to-peak amplitudes of ## mV.

Table 2.

TMS biomarkers in neurodevelopmental disorders.

Disorder	MT	MEP amplitude	cSP	I/O curve slope	SICI	ICF	LICI	TBS	PAS	Clinical correlates	Comments
Autism spectrum disorder	-	-	-	-	↓, -	↓, -	-	↑ cTBS/ ↓TBS duration in adults and children; ↓cTBS- induced facilitation in children	↓	<ul style="list-style-type: none"> • greater SICI and prolonged cSP negatively correlated w/ academic performance; • ICF inversely correlated w/ inattention and executive dysfunction • reduced SICI correlated w/ developmental delay in language acquisition 	<ul style="list-style-type: none"> • positive correlation between age and cTBS-induced modulation or maximum cTBS-induced inhibition
Fragile X syndrome					-		-	↓ cTBS response; ↑ iTBS response; ↓ cTBS duration; ↓cTBS- induced facilitation on 2nd visit		<ul style="list-style-type: none"> • complete blocking of iTBS response on 2nd visit • normal SICI and LICI could be due to unaffected X chromosome / non-optimal ISI 	
Attention-deficit hyperactivity disorder	↑, -		- , ↓		↓, ↓ SICI-induced facilitation in a few cases	↓, ↑, -	↓			<ul style="list-style-type: none"> • SICI inversely correlated w/ symptom severity • ISP duration correlated w/ hyperactivity and restlessness • SICI index of response to MPH 	<ul style="list-style-type: none"> • SICI inversely correlated w/ ISP duration • ↑ ISP latency in children • ↑ ISP duration in children • ↓ ISP latency in adults • ICF greater in ASD children w/ ADHD than in ASD children w/o ADHD

Disorder	MT	MEP amplitude	cSP	I/O curve slope	SICI	ICF	LICI	TBS	PAS	Clinical correlates	Comments
Tourette syndrome	- , ↑	↓ during movement preparation and voluntary tic inhibition	↓ , -	↓ , -	↓	↑ , -	↑ , ↓ iTBS response ↓ cTBS response	↓ PAS-induced inhibition		<ul style="list-style-type: none"> • reduced SICI positively correlated w/ motor tic severity • SICI normalization during movement preparation correlated w/ fewer tics and lower tic severity • PAS-induced inhibition correlated with less severe urges and fewer tics 	<ul style="list-style-type: none"> • ↑ MEP variability • ↓ I/O curve slope during voluntary tic inhibition
Developmental stuttering	↑		↑ for tongue	↓ for hand; ↑ for tongue	↓ for tongue	↓ for tongue				<ul style="list-style-type: none"> • cSP duration negatively correlated w/ stuttering severity in males • extent of higher aMT for tongue in left M1 relative to right M1 correlated with stuttering severity 	<ul style="list-style-type: none"> • aMT for tongue in left M1 higher than in right M1

ADHD, attention-deficit hyperactivity disorder; aMT, active motor threshold; ASD, autism spectrum disorder; cSP, cortical silent period; cTBS, continuous theta-burst stimulation; ICF, intracortical facilitation; I/O, input/output; ISI, inter-stimulus interval; iSP, ipsilateral silent period; iTBS, intermittent theta-burst stimulation; LICI, long-interval intracortical inhibition; M1, primary motor cortex; MEP, motor evoked potential; MPH, methylphenidate; MT, motor threshold; rTMS, neuronavigated transcranial magnetic stimulation; PAS, paired associative stimulation; SICI, short-interval intracortical facilitation; SICI, short-interval intracortical inhibition; TBS, theta-burst stimulation; w/, with; w/o, without; ↑ increase; ↓ decrease; - no change; ⊥ paradoxical; blank cells, not tested; conflicting results.