

Recognition of Two Groups of Methicillin-Resistant *Staphylococcus aureus* Strains Based on Epidemiology, Antimicrobial Susceptibility, Hypervariable-Region Type, and Ribotype in Finland

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Epidemiological evidence suggests that some methicillin-resistant *Staphylococcus aureus* (MRSA) strains are more prone to dissemination than others. We studied 72 MRSA strains, collected through nationwide MRSA surveillance in 1992 through 1999 and known to be either (i) sporadic, (ii) local outbreak strains spread within one hospital, or (iii) epidemic strains spread among hospitals, by antimicrobial susceptibility testing, hybridization of the *mec* hypervariable region (HVR), and ribotyping. Our results show that two main groups can be identified among these strains. The first group includes mainly nonepidemic, nonmultiresistant MRSA strains showing a specific *mec* HVR hybridization pattern, A, in combination with a variety of ribotypes. The other group includes multiresistant strains with *mec* HVR hybridization pattern B or C in association with closely related ribotype a or b. Sixty-four percent (9 of 14) of Finnish epidemic MRSA strains belong to the latter group. These findings support the existence of differences in epidemic potential among MRSA strains.

Certain methicillin-resistant *Staphylococcus aureus* (MRSA) strains have efficiently disseminated both locally and internationally. The factors affecting dissemination of resistant organisms include hospital size and type, antimicrobial policies of hospitals, frequency of patient transfer between wards and hospitals, certain risk factors predisposing patients to MRSA colonization and infection, and infection control measures (1, 2, 4, 25). Based on epidemiological evidence, it has also been suggested that some MRSA strains have properties that enable them to disseminate particularly well (19).

The search for genetic properties linked to epidemic spread can be initiated by comparing the characteristics of known epidemic and sporadic strains. The genetic background of MRSA strains is determined by the *mec* DNA and the original chromosome of methicillin-susceptible *S. aureus* to which the *mec* DNA has integrated. Although the genomes of the susceptible and the resistant strains have diversified since this initial integration, certain genotyping methods, such as ribotyping, can be used to delineate the genetic background of the MRSA strains (10). Ribotyping with concomitant analysis of the *mec* DNA thus provides information on the potential differences in the genomic background or on the *mec* DNA of epidemic and sporadic strains.

The 20- to 50-kb *mec* DNA includes the *mecA* gene, coding for penicillin-binding protein 2a, which is primarily responsible for methicillin resistance, and the complete or truncated regulatory region *mecRI-mecI* (14, 20). The *mecA* downstream sequences contain series of direct repeat units (*dru* region), an open reading frame whose product has amino acid similarity to the N terminus of *Escherichia coli* glycerophosphoryl diester

phosphodiesterase (UppQ), and insertion sequence IS431 (16, 21). The genomic distance between *mecA* and IS431 is highly variable, and this region is called the *mec* hypervariable region (HVR) (11, 16, 21). This region and the insertion sequences and transposons within the *mec* DNA can serve as hot spots, which collect antibiotic resistance genes or other markers into the MRSA genome (24).

In this study, we searched for molecular traits linked to epidemic spread using a subset of 72 MRSA isolates from the Finnish MRSA strain collection. This population-based collection represents all Finnish MRSA strains typed since 1992 and includes both epidemic and sporadic strains.

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MATERIALS AND METHODS

Since 1992, all clinical microbiology laboratories in Finland have sent MRSA isolates to the Hospital Infection Laboratory at the National Public Health Institute (KTL) for typing. In 1995, MRSA became a notifiable laboratory finding in Finland. Most laboratories report the findings electronically to the National Infectious Disease Register at KTL.

Routine MRSA typing scheme in 1992 through 1999. All MRSA isolates referred to KTL were typed by phages and tested for antimicrobial susceptibility. Phage typing was performed with the universal set of phages (18) at 1× and 100× routine test dilutions, both with and without heat treatment of the bacteria (5). The antimicrobial susceptibilities were tested by the disk diffusion method according to guidelines recommended by the National Committee for Clinical Laboratory Standards (15). The antimicrobials tested were oxacillin, ampicillin, penicillin, cephalixin, cefuroxime, gentamicin, tobramycin, erythromycin, clindamycin, chloramphenicol, ciprofloxacin, rifampin, fusidic acid, and mupirocin. MICs of oxacillin were determined by the E-test according to the manufacturer's instructions (AB Biodisk, Solna, Sweden). Additional studies using pulsed-field gel electrophoresis (PFGE) and, occasionally, ribotyping were performed if the isolates met any of the following criteria: (i) they were known to be linked to a suspected epidemic based on epidemiological information obtained from the

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TABLE 1. MRSA strains isolated in Finland in 1992 through 1999

Yr	No. of MRSA strains reported to the National Infectious Disease Register	Incidence ^a	No. of MRSA strains referred to the Hospital Infection Laboratory ^c	No. of sporadic strains (%)	No. of local outbreak strains (%)	No. of epidemic strains (%)
1992	NA ^b	NA	140	30 (21)	2 (1)	108 (77)
1993	NA	NA	128	41 (32)	30 (23)	57 (45)
1994	NA	NA	225	76 (34)	12 (5)	137 (61)
1995	89	0.17	137	47 (35)	6 (4)	84 (61)
1996	108	0.21	118	55 (46)	2 (2)	61 (52)
1997	120	0.23	143	93 (65)	8 (5)	42 (30)
1998	189	0.37	196	120 (61)	11 (6)	65 (33)
1999	211	0.41	212	76 (36)	38 (18)	98 (46)
Total	NA	NA	1,299	538 (41)	109 (8)	652 (50)

^a Incidence per population of 10,000 based on MRSA cases reported to the National Infectious Disease Register.

^b NA, not applicable.

^c Mean, 162 per year.

submitting hospital, (ii) they possessed an antimicrobial susceptibility pattern and a phage type similar to those for a strain identified earlier in the laboratory, and (iii) they could not be typed by phages. PFGE and ribotyping were performed as described earlier (22). Briefly, for PFGE, genomic DNA prepared in agarose blocks was digested with *Sma*I restriction endonuclease, and the chromosomal fragments were separated with a CHEF DR III (Bio-Rad, Richmond, Calif.) for 24 h, with initial and final switching times of 10 and 60 s, respectively. PFGE patterns were interpreted as different if more than three band differences occurred. Ribotyping was performed by digesting genomic DNA with *Eco*RI restriction endonuclease. DNA fragments were separated by electrophoresis, transferred to a nylon membrane, and hybridized with a probe for the *rm* operon of *E. coli* (3).

Strain definitions. By combining the typing data and epidemiological information from hospitals, all strains were classified as either sporadic, local outbreak, or epidemic strains. A sporadic MRSA strain was defined as a strain isolated from one person only and displaying an individual antimicrobial susceptibility pattern and/or phage type. For non-phage-typeable strains, the strain relatedness was ascertained by PFGE. MRSA isolates sharing the same typing patterns and isolated from two or more persons in the same hospital were defined as representative of a locally spread outbreak strain. MRSA isolates sharing the same typing patterns and isolated from two or more persons and at two or more hospitals were defined as representative of an epidemic MRSA strain.

Selection of isolates for analysis of genomic background. From the MRSA collection, built up by strains sent for typing since 1992, 72 isolates were selected for more detailed analysis of genomic background. These isolates included all 47 sporadic isolates from one representative year, namely, 1995, as well as one representative of each MRSA strain which has caused epidemics ($n = 14$) or local outbreaks ($n = 11$) from 1992 to 1998. Eighteen of these strains have been described previously (22).

Analysis of the HVR *mec* determinant. Genomic variation within the *mec* determinant was analyzed by hybridization analysis. The genomic DNA from MRSA strains was digested with *Eco*RI and *Bgl*II restriction enzymes and hybridized separately with two different probes prepared from plasmid pBBB30 (21). HVR probe I is a 3.6-kb fragment that recognizes the HVR of the *mec* determinant, starting within the *mecA* gene up to IS431 *mec*. HVR probe II is a 1.5-kb fragment digested with *Hind*III from the 3.6-kb fragment. This probe recognizes the *dru* and an *E. coli* *ugpQ*-like sequence (*orf145*) within the *mec* determinant. The extraction of genomic DNA, digestion with restriction enzymes, gel electrophoresis with digoxigenin-labeled molecular weight marker III (Roche Molecular Biochemicals) as a migration control, blotting on to a nylon membrane, and labeling reactions were performed as previously described (22). The hybridization was performed according to instructions provided by the nylon membrane manufacturer (Boehringer Mannheim). Briefly, after 2 h of prehybridization in 20 ml of hybridization buffer (0.75 M NaCl, 75 mM sodium citrate, 1% blocking reagent, 0.1% *N*-lauroylsarcosine, 0.02% sodium dodecyl sulfate) at 60°C, about 80 ng of probe (HVR I or HVR II) was added to 20 ml of hybridization buffer, boiled for 10 min, and hybridized with DNA fragments for 20 to 24 h at 60°C.

Ribotyping. Ribotyping with *Eco*RI was used to detect the phylogenetic relationship within the MRSA strains. *Mlu*I-digested *Citrobacter koseri* was used as the migration control (22). A difference of one band in the ribopattern, as detected by visual analysis, was considered to represent a new type. An arbitrary identification letter was given to each ribotype. Computer-assisted analysis of the ribotypes was performed by BioNumerics, version 1.0 (Applied Maths, Kortrijk, Belgium), using the Dice coefficient with a position tolerance setting of 0.9%, optimization setting of 0.8%, and unweighted pair-group method using arithmetic averages.

Statistical analysis. Statistical associations were tested by the chi-square method using Yates' correction.

RESULTS

MRSA strains. Since 1995, the annual number of MRSA isolations reported to the National Infectious Disease Register has varied from 89 to 211, with an incidence per population of 10,000 of 0.17 to 0.41 (Table 1). During the period from 1992 through 1999, a total of 1,299 MRSA strains were referred to the Hospital Infection Laboratory (Table 1), i.e., 118 to 225 MRSA strains per year. According to the definitions described above, 538 (41%) of the isolates were sporadic, 109 (8%) were local outbreak strains, and 652 (50%) were epidemic strains.

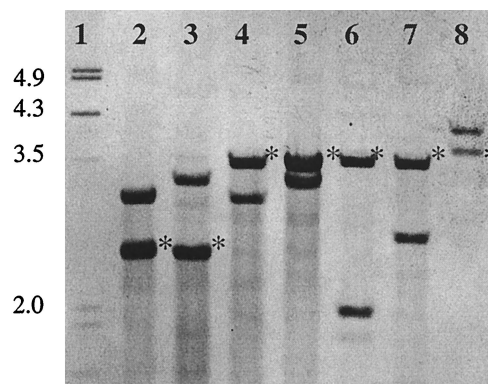


FIG. 1. HVR probe I hybridization patterns. Lanes 1, molecular size marker (fragment sizes are indicated as kilobase pairs); 2 to 8, HVR types B, D, A, C, E, F, and G. Asterisks indicate the fragments recognized by HVR probe II (the *dru-orf145* fragments).

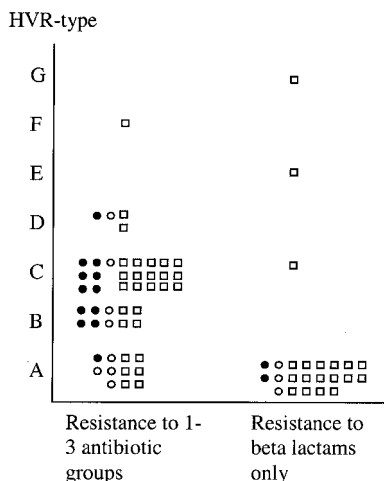


FIG. 2. HVR type in relation to antibiotic resistance patterns and epidemic spread. ●, epidemic strain; ○, local outbreak strain; □, sporadic strain.

The annual percentage of epidemic strains ranged between 30 and 77%.

The observation that epidemic strains represent half of all MRSA isolates in Finland led us to hypothesize that these strains may harbor characteristics favorable for epidemic spread. To test this hypothesis, we selected 72 MRSA strains for further analysis. These included all sporadic isolates from 1995 ($n = 47$) and one representative isolate of each epidemic ($n = 14$) and outbreak ($n = 11$) strain identified between 1992 and 1998.

Antibiotic susceptibility. Of the 47 sporadic strains, 14 (30%) were multiresistant (i.e., resistant to more than three antibiotic groups in addition to beta-lactams), 14 (30%) were resistant to beta-lactams and one to three other antibiotic groups, and 19 (40%) were resistant to beta-lactams only. The corresponding figures for epidemic strains were 11 (79%), 1

(7%), and 2 (14%), respectively. The corresponding figures for local outbreak strains were 5 (45%), 3 (27%), and 3 (27%), respectively. Together, these results indicate that multiresistance was more common among epidemic strains (11 of 14) than among nonepidemic MRSA strains (19 of 58) ($P < 0.005$).

Analysis of the HVR *mec* determinant. Hybridization with HVR probe I differentiated the 72 strains into 7 types: A, B, C, D, E, F, and G (Fig. 1), each of which comprised two bands. HVR probe II recognized the other of these two bands showing the position of *dru-orf145* and flanking sequences with three different migration profiles. HVR types A, C, E, and F showed a *dru-orf145* band of approximately 3.5 kb, HVR types B and D showed a band of approximately 2.5 kb, and HVR type G showed a band of more than 3.5 kb.

Most sporadic strains were either HVR type A (22 of 47 [47%]) or type C (16 of 47 [34%]) (Fig. 2). The epidemic strains were mostly HVR type B (4 of 14 [29%]) or type C (6 of 14 [43%]). Three epidemic strains were HVR type A, and one was type D. Of the 11 local outbreak strains, 7 (64%) were HVR type A and 4 were type B, C, or D.

Ribotyping. Ribotyping with *EcoRI* yielded 18 different ribotypes (Fig. 3). The most prevalent types were a and b (35 [49%] of 72 strains). Of the sporadic strains, 21 (45%) showed either ribotype a or ribotype b. The corresponding figures for epidemic and local outbreak strains were 9 (64%) and 5 (45%), respectively. Ribotypes a and b differed from each other by only one band. Computer-assisted analysis of ribotypes showed that a >92% similarity level corresponded to a one- to two-band difference and an 87 to 90% similarity level corresponded to a three- to four-band difference (Fig. 3).

Linkage of antibiotic susceptibility to genotype and epidemiological background. In combined analysis, the MRSA strains clustered into two main groups (Tables 2 and 3 and Fig. 2). Thirty-two of 48 strains resistant to more than one antibiotic group, the majority of which were multiresistant, associated with ribotype a or b (32 of all 35 strains with ribotype a or

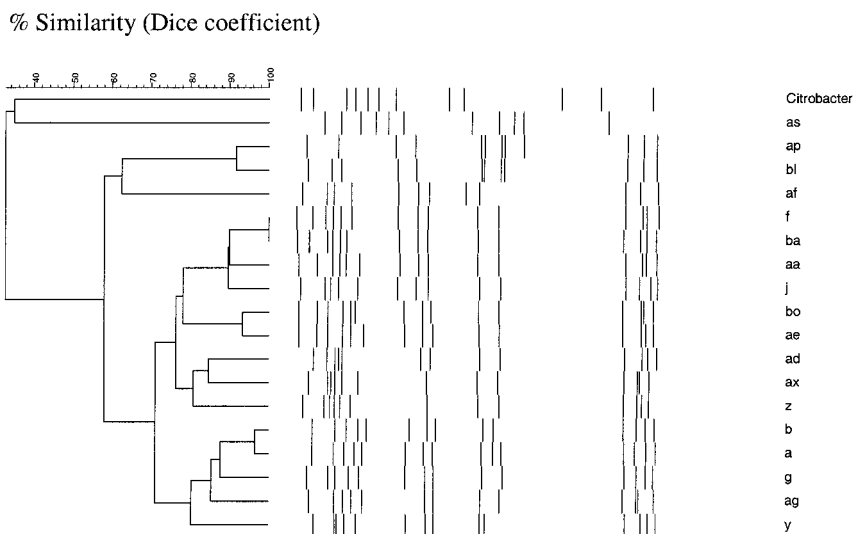


FIG. 3. Dendrogram of relatedness of the ribotypes.

diagnostic tests for detection of MRSA strains with epidemic-spreading capacity.

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