



Missing sputum samples are common in asthma intervention studies and successful collection at follow-up is related to improvement in clinical outcomes

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Received: 29 Oct 2021
Accepted: 21 Dec 2021

To the Editor:

With only modest agreement between airway and systemic eosinophilia, biomarkers directly assessing the level and type of airway inflammation are becoming increasingly important, both for targeting treatment to the individual patient and for assessing effect [1].

A sputum cell differential count remains the gold standard for airway inflammometry in asthma but missing data are an inherent issue when utilising sputum-based outcome measures as the success rate for induction ranges markedly [2, 3]. It is well known that missing data potentially can lead to substantial bias when inadequately handled [4]. At present, reflections on how to handle missing sputum samples are largely absent in the literature and most studies defer to the use of baseline values to predict outcomes or utilise complete-case analysis despite evidence highlighting multiple imputation as the superior statistical method independent of the missingness of the data [2, 3, 5–8].

With the advances in the feasibility of sputum sampling in a clinical setting, we foresee a marked increase in the utilisation of sputum-based outcome measures, highlighting the necessity to evaluate the missingness of induced sputum [9, 10].

We hypothesised that any patient's ability to produce a sputum sample after receiving medical treatment is not random and that the proportion of missing samples is higher in patients with response to treatment, as a result of resolution of airway inflammation in general and the interleukin-13 driven mucus hypersecretion in particular [11]. Therefore, we pooled data on mannitol-induced sputum from three intervention studies (n=135) with an aim to identify predictors of successful induction prior to and after a medical intervention.

The RECONSTRUCT study was a single-arm intervention study (1600 µg inhaled budesonide once daily for 16 weeks) of steroid-free asthma patients with airway hyperreactivity (AHR) to mannitol (www.clinicaltrials.gov identifier number NCT03034005). The UPSTREAM study was a placebo-controlled intervention study (add-on of anti-thymic stromal lymphopoietin (n=20) or placebo (n=20) for 12 weeks) of predominantly moderate-to-severe asthma patients with AHR to mannitol [12]. The SIGNATURE study was a single-arm intervention study (add-on of 37.5 mg oral prednisolone for 2 weeks) of patients with moderate-to-severe asthma [7].

Successful sputum induction was not required for inclusion in any of the studies and maintenance treatment prior to enrolment was continued unchanged throughout the study period. In all three studies, sputum was collected following a mannitol challenge test in a specimen jar (Petri dish) with the sample quality continuously evaluated by a trained laboratory technician. Samples were processed using the plug selection method, and cut-off values for eosinophilia and neutrophilia were $\geq 3\%$ and $\geq 61\%$, respectively [7, 12].



Shareable abstract (@ERSpublications)

Several factors significantly impact ability to produce a sputum sample after an anti-inflammatory intervention and these authors argue that the widely used complete-case analysis is inappropriate for paired sputum-based outcome measures <https://bit.ly/3qN2pk5>

Cite this article as: Frøssing L, Hvidtfeldt M, Silberbrandt A, et al. Missing sputum samples are common in asthma intervention studies and successful collection at follow-up is related to improvement in clinical outcomes. *ERJ Open Res* 2022; 8: 00612-2021 [DOI: 10.1183/23120541.00612-2021].



Three-quarters (75%, n=101) of patients were able to produce a sufficient sputum sample at baseline, two-thirds (65%, n=88) at follow-up and paired samples were collected in half of the patients (52%, n=70). The success rate for collection of sputum at follow-up was equal in the placebo group and in the patients receiving active treatment, and we found no significant difference in the success rate between baseline and follow-up across all patients, in patients receiving active treatment, or in each study individually.

At baseline, neither demographics, lung function, AHR (provocative dose of mannitol causing a 15% fall in forced expiratory volume in 1 s (FEV₁)) or inflammatory profile (blood eosinophils, exhaled nitric oxide fraction (F_{ENO}), immunoglobulin E and atopy) were significantly related to a successful collection.

In the patients receiving active treatment, success rate at follow-up was significantly higher in patients not receiving inhaled corticosteroids (ICS) at baseline (80% versus 57%; OR 4.3, p=0.006), and significantly lower in patients receiving treatment with a long-acting β_2 -agonist (42% versus 72%; OR 0.27, p=0.003) and long-acting muscarinic antagonist (9% versus 26%; OR 0.29, p=0.02) at baseline (figure 1). Similarly, patients with severe asthma according to European Respiratory Society/American Thoracic Society criteria had a significantly lower success rate compared to those without (54% versus 77%; OR 0.31, p=0.008) [13].

For patients receiving active treatment, those with the paucigranulocytic inflammatory phenotype at baseline had a significantly lower success rate at follow-up (41% versus 59%; OR 0.46, p=0.05). Successful collection at follow-up was not associated with other baseline inflammatory markers (sputum eosinophils, blood eosinophils and F_{ENO}) or reduction in these.

Across all patients, successful collection of sputum at follow-up was significantly more prevalent in patients with improvement in FEV₁ (200 mL and 12%) and AHR at follow-up measured with a mannitol challenge test (79% versus 57% (OR 2.4, p=0.02) and 82% versus 58% (OR 3.3, p=0.01), respectively). In the patients receiving active treatment, this remained significant for mannitol (p=0.01) and showed a strong trend for FEV₁ (p=0.08).

We did not identify any factors affecting the success rate of induction at baseline; however, the likelihood of a successful induction post-intervention decreased with higher maintenance ICS doses at baseline and was, in line with our hypothesis, lower in the absence of airway inflammation (paucigranulocytic sputum). Surprisingly, a clinical response to treatment, with increase in lung function or improvement in AHR, was associated with a higher likelihood of a successful induction post-intervention. Speculatively, this could be explained by relief of airway obstruction that, in turn, allows for mobilisation of distal airway mucus plugs.

Interestingly, the level of maintenance treatment and the anti-inflammatory add-on intervention exerted opposite effects on the success rate. We believe this to be explained by the fact that they reflect different

	All (n=135)	Induction success rate		Signature OCS (n=48)	Reconstruct ICS (n=47)
		Upstream Anti-TSLP (n=20)	Placebo (n=20)		
Baseline	75%	80%	75%	73%	76%
Follow-up	65%	55%	60%	58%	80%
Paired samples [#]	52%	40%	50%	46%	65%

Significant predictors of higher success rate at follow-up

- Absence of maintenance ICS treatment (OR 4.3)
- Decreased AHR at follow-up[¶] (OR 3.3)
- Improvement in FEV₁ at follow-up (2.4)[†]

Significant predictors of lower success rate at follow-up

- High-dose[§] maintenance ICS treatment (OR 0.31)
- Maintenance treatment with a long-acting bronchodilator (LABA: OR 0.27; LAMA: OR 0.29)
- Paucigranulocytic airway inflammation (OR 0.46)

FIGURE 1 Success rate of sputum induction in asthma and predictors of successful induction at follow-up. TSLP: thymic stromal lymphopoietin; OCS: oral corticosteroid; ICS: inhaled corticosteroid; AHR: airway hyperreactivity; FEV₁: forced expiratory volume in 1 s; LABA: long-acting β_2 -agonist; LAMA: long-acting muscarinic antagonist. [#]: proportion of patients with successful sputum at baseline and follow-up; [¶]: normalisation of mannitol challenge test; [†]: 200 mL and 12% improvement; [§]: 1600 μ g budesonide equivalent.

aspects (traits) of disease: maintenance ICS dose is reflective of disease severity (*i.e.* chronicity) whereas response to add-on treatment reflects reversibility of disease, a notion we believe to be supported by the increase in success rate at follow-up in the steroid-naïve patients in the RECONSTRUCT study (figure 1).

Mannitol-induced sputum samples are of good quality, and are comparable with samples induced with hypertonic saline for the analysis of inflammatory cells and soluble markers [14, 15]. Further, the success rate for mannitol and hypertonic saline induction are similar, and the success rates reported in this study are equal to our own prior efforts using saline and to previous reports using mannitol [3, 14].

Still, we note several factors that potentially influence the generalisability of our findings to hypertonic saline-induced samples and we believe future studies confirming our results in hypertonic saline-induced samples are warranted. Wood *et al.* [14] reported a significantly lower total cell count (3.8 *versus* 2.1×10⁶, p=0.003) in samples induced with mannitol compared with hypertonic saline, which we believe may hamper our generalisability as we defined successful induction based on total cell count. Further to this point, Wood *et al.* [14] and Alvarez-Puebla *et al.* [15] have reported significant differences in inflammatory phenotype classification using mannitol and hypertonic saline induction respectively, which, as we found the paucigranulocytic phenotype to be significantly associated with a lower success rate at follow-up, again hampers the generalisability of our results.

Successful induction was defined based on identification of ≥250 cells and we acknowledge that other cut-offs or criteria, such as viability, could have been chosen; however, as no consensus exists, this task is inevitably difficult and biased.

In summary, our findings suggest that the ability to produce repeated sputum is not random, and should be taken into account in the planning and analysis of interventions studies. Based on our findings, we believe that complete-case analysis is inappropriate for paired sputum-based outcome measures. We speculate that imputation better accommodates the missingness of sputum but future studies evaluating different methods for handling missing data are warranted [8].

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Provenance: Submitted article, peer reviewed.

Conflict of interest: L. Frøssing has nothing to disclose. M. Hvidtfeldt has nothing to disclose. A. Silberbrandt has nothing to disclose. A. Sverrild has nothing to disclose. C. Porsbjerg has nothing to disclose.

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