Pulmonary Infection Caused by an Unusual, Slowly Growing Nontuberculous Mycobacterium

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Mycobacterium triplex, a recently described slowly growing nontuberculous mycobacterium, was isolated from a Finnish patient with pulmonary mycobacteriosis. The disease was successfully treated with antimycobacterial drugs. The strain isolated, which was similar to the type strain but differed slightly from the species description, was regarded as a variant of *M. triplex* sensu stricto. According to present knowledge this variant of the species has never been isolated before.

Recent advances in diagnostic methodology for mycobacteria other than *Mycobacterium tuberculosis* (MOTT), and new possibilities introduced for drug therapy of the diseases they induce, have renewed interest in mycobacterial research. The recognition that most patients with AIDS are at high risk of complications and/or terminal illness caused by MOTT has further stimulated research efforts. The principles of diagnosis and therapy of diseases caused by MOTT have also recently been updated by the American Thoracic Society (1). These guidelines include recommendations for specific drug regimens which recognize the major impact of the newer chemotherapeutic agents.

Improved diagnostic means allow enhanced detection and more-accurate species identification of mycobacteria isolated from clinical specimens, including the classification of species not recognized earlier. In 1996, Floyd et al. (5) characterized a new species of slow-growing MOTT, which they designated *Mycobacterium triplex*. The primary characterization of this novel species was based on conventional testing, but the conclusive genetic evidence relied on 16S rRNA analyses. Although the species is uncommon, it was found to be geographically widespread in the United States (5). There is one very recent report of *M. triplex* in Europe (4).

We isolated *M. triplex* from a Finnish patient suffering from pulmonary mycobacteriosis. *M. triplex* was the only mycobacterium isolated from the specimens. As illustrated in this paper, the strain found in this patient differed slightly from the earlier description of the species. To our knowledge, this is the first report of a pulmonary disease associated with *M. triplex*.

Case report. A 67-year-old, human immunodeficiency virusnegative dentist presented in hospital with hemoptysis in 1995. On chest radiographs, thin fibrotic changes were detected in both lower fields, but no cavitary changes were detected. Highresolution computed tomography (HRCT), performed in December 1995, revealed multifocal bronchiectases in both lungs and multiple small nodules in the middle and lower lobes of the right lung and the lower lobe of the left lung, indicating mycobacteriosis (1). Bronchoscopy showed nonspecific inflammatory mucosal changes on the right side. Smears of sputum and bronchoalveolar lavage specimens were negative for acidfast bacilli, but cultures of sputum samples taken on three successive days grew a slow-growing mycobacterium. It was identified using conventional tests, gas-liquid chromatographic (GLC) analysis of cellular fatty acid and alcohol composition, and 16S rRNA gene sequencing as described earlier in detail (6, 7, 11). The isolates met the identification criteria for M. triplex (5). Since this was the only organism growing in the three cultures, we believe that the isolation of *M. triplex* represents infection rather than colonization. Moreover, the patient has bronchiectases throughout both lungs, which would predispose her to this kind of lung infection. Treatment was initiated using a standard antituberculous regimen for 2 months, to which the lesions responded only partially. Treatment was continued with rifampin, ciprofloxacin, ethambutol, and clarithromycin for a total of 10 months, with the exception of a 3-week period in March and April 1996 due to a temporary rise in liver enzymes. This regimen proved to be well tolerated. The bacteriological and clinical responses were good. An HRCT scan in October 1996, after 10 months of treatment, showed a good radiological response (Fig. 1). The patient was regarded as cured at follow-up, after taking antituberculosis medication for 18 months.

The growth and biochemical characteristics of the patient's isolates were most closely compatible with *M. triplex* (Table 1) (5). By in vitro testing, performed using the proportion method in Löwenstein-Jensen medium, the strain was susceptible to rifampin, streptomycin, and clarithromycin and moderately sensitive to ciprofloxacin, but it had decreased susceptibility to ethambutol, isoniazid, and pyrazinamide. The isolates failed to hybridize with the commercially available genetic probe for *Mycobacterium avium* complex (Accuprobe; Gen-Probe Inc., San Diego, Calif.). Two of the patient's isolates were analyzed by GLC for cellular fatty acid methyl esters and alcohols, prepared by acid methanolysis, and analyzed and identified as described earlier in detail (10). These two isolates had an identical GLC profile, which closely resembled that of the *M. triplex* type strain (ATCC 70071). However, repeated analyses

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FIG. 1. HRCT scans before (A) and after (B) treatment.

of the fatty acid composition verified minor but constantly detectable differences from the *M. triplex* type strain, including a higher relative amount of tetracosanoate (24:0) and a low relative amount of tuberculostearic acid (10-Me-18:0) (Table

1). The fatty acid profile of the *M. triplex* type strain was found to be indistinguishable from that of *M. simiae* (3), a slow-growing photochromogenic species. Pigment production easily separates these two species. They both have GLC profile is

 TABLE 1. Biochemical tests and markers in the GLC profile of fatty acids and mycolic acid cleavage products found useful in differentiation of *M. triplex* and the isolate described

	Result ^a					
Characteristic	Patient isolates	M. triplex	M. tuberculosi			
Biochemical characteristics						
Growth at						
25°C	+	V	_			
37°C	+	+	+			
45°C	_	V	_			
Pigment production	_	_	_			
Niacin	_	_	+			
Arylsulfatase						
3 days	_	_	_			
14 days	(Weak +)	V	_			
Semiquantitative catalase	÷	+	_			
Nitrate reductase	+	+	+			
Urease	+	+	+			
Tween 80 hydrolysis	-	_	_			
M. avium complex DNA probe	_	_	_			
Marker peaks in GLC fatty acid profile ^b						
14:0	7.2	7.8	1.6			
10-Me-18:0	7.2	12.6	19.4			
24:0	8.9	5.9	3.2			
26:0	8.7	13.6	9.1			

 a Biochemical tests were performed by the procedure of Floyd et al. (5). +, positive; -, negative; V, variable.

^b 14:0, tetradecanoate; 18:0, octadecanoate; 10-Me-18:0, tuberculostearate; 24:0, tetracosanoate; 26:0, hexacosanoate. Results are mean area percentages for the patient's isolates (eight analyses), for the *M. triplex* type strain (three analyses), and for 20 clinical *M. tuberculosis* strains.

grossly similar to that of *M. tuberculosis*. As shown in Table 1, the high relative content of tetradecanoate (14:0) in both the type strain of *M. triplex* and the patient isolates (7%) differentiated them from *M. tuberculosis* (<2%). Others have verified the same difference between *M. tuberculosis* and *M. simiae* (3). Overall, it was difficult to identify the species of the isolated strains using conventional techniques.

Sequencing of the 16S rRNA gene is widely used for species identification of mycobacteria. The complete 16S rRNA gene was amplified, and both strands of the amplified DNA were sequenced as described elsewhere (6). A BLAST sequence similarity search (EBI, Hinxton, United Kingdom) of the 16S sequence obtained showed a 99.5% similarity to *M. triplex* (Fig. 2A). The internal transcribed spacer (ITS) sequences have been shown to be useful phylogenetic markers for species differentiation and identification (8, 9). The ITS region of the patient isolate was amplified and sequenced as described previously (10). Comparison of the sequences showed 8 differences in the 16S rRNA gene and 14 differences in the ITS region between the patient isolate and the *M. triplex* type strain (Fig. 2A and B, respectively) (5, 8).

The result of 16S rRNA sequencing, with 99.5% sequence similarity with *M. triplex*, indicated that the isolate was very closely related to *M. triplex*. We found the sequencing of the ITS region necessary because of the eight differences in the 16S rRNA gene. The result of the ITS sequencing confirmed the other results obtained in this study, indicating that this organism was an *M. triplex*-like mycobacterium. According to currently available data, the patient's strain is regarded as a variant of *M. triplex*. Further analyses are needed to evaluate

Α

<i>M. triplex</i> taxon 47839 Patient isolate	37 ACGGAAAGGC CTCTTCGGAG	GTACTCGAGT	155 GACGCATGTC	CTGTGGTGC	A AAGC	
M. tríplex taxon 47839 Patient isolate	218 TGGTGGGGTG ACGGCCTACC T	1064 CTCATGTTGC	CAGCGGGTAA	TGCCGGGGG	AC TCGTGAG	;
В						
M. tuberculosis taxon 1773	AAGGAGCACC ACGA-AAACG	CCCCAACTGG TG	GGGCGTAG GCC	CGTGAGGG (TTCTTGTCT	59
M. simiae taxon 1784 M. triplex taxon 47839		.TT		<i></i> .	C	60 60
Patient isolate	GA	.TT	TGA		C	60
M. tuberculosis taxon 1773	GTAGTGGGCG AGAGCCGGGT	GCATGACAAC A-	AAGTTGGC CAG	CAACACA (TGTTGGGTC	118
M. simiae taxon 1784	AG	CAG	GCAA.C	3	.AC.	118
M. triplex taxon 47839 Patient isolate	AG A G.GT	CAGG	GCAA.C(.CAA.C(3 3		118 118
M. tuberculosis taxon 1773	CTGAGGCAAC ACTCGGA	CTTGTT CC	AGGTGTTG TCC	CACCGCC 1	TGGTGGTGG	171
M. simiae taxon 1784	A	CGGTTGA AG	T		• • • • • • • • • •	175
M. triplex taxon 47839 Patient isolate		AGGTCGA TG	ИГ ИГ	TAT		175
M. tuberculosis taxon 1773	GGTGTGGTGT TTGAGAACTG	GATAGTGGTT GC	GAGCGCGA GC	ATC-AA-T (GATACGCTG	229
M. simiae taxon 1784	T.T			TA.	A.CGTC.	235
M. triplex taxon 47839 Patient isolate	C.T AT.T	A		T.GAC		235
M. tuberculosis taxon 1773	CCGGCTAGCG GTGGCGTGTT	CTTT GT	GCAA-TTA TTO	C-TTTGGT 1	TTTGTGT	279
M. simiae taxon 1784	ATA	.GT	.TTT	.T		288
M. triplex taxon 47839	CAAGT.GCCA	.GTCAAAAA	.TT	.T		290
PALIEUL ISOLATE						403

FIG. 2. Comparison of ribosomal gene sequences. Dots indicate nucleotide identity, and dashes indicate deletions. (A) Differences in the 16S rRNA gene between the isolate (AJ276890) and *M. triplex* (U57362). Numbering is according to the reference sequence. (B) Alignment of ITS region sequences of selected species including the patient isolate. Reference sequence accession numbers are as follows: *M. triplex*, Y14189; *M. simiae*, AB026694; *M. tuberculosis*, AB026698.

the relevance of the differences detected and to answer the open question of whether the strain should be regarded as a subspecies of *M. triplex* or as another as yet unclassified but closely related species.

Several recent reports describe infections caused by unusual *Mycobacterium* species (2, 12). The development of molecular biological methods has allowed more-accurate species identification and the study of species diversity. The clinical case reported here indicated that the *M. triplex* variant described had to be regarded as a potential human pathogen. The results also demonstrated that the regimen often used to treat pulmonary mycobacteriosis caused by *M. avium* is also sufficient for the treatment of *M. triplex* infections.

Nucleotide sequence accession number. The 16S and ITS sequences determined in this study have been deposited in the EMBL nucleotide sequence databank under accession number AJ276890.

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