




## REVIEW

# Contaminants in the cow's milk we consume? Pasteurization and other technologies in the elimination of contaminants

[version 1; peer review: 3 approved, 1 approved with reservations]

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## Abstract





Cow's milk is currently the most consumed product worldwide. However, due to various direct and indirect contamination sources, different chemical and microbiological contaminants have been found in cow's milk. This review details the main contaminants found in cow's milk, referring to the sources of contamination and their impact on human health. A comparative approach highlights the poor efficacy and effects of the pasteurization process with other methods used in the treatment of cow's milk. Despite pasteurization and related techniques being the most widely applied to date, they have not demonstrated efficacy in eliminating contaminants. New technologies have appeared as alternative treatments to pasteurization. However, in addition to causing physicochemical changes in the raw material, their efficacy is not total in eliminating chemical contaminants, suggesting the need for new research to find a solution that contributes to improving food safety.




## Keywords

human health; chemical contaminant; microbiological contaminant; alternative; technology; food safety

## Open Peer Review

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## 1. Introduction

Milk is a fluid secreted by the female of the mammalian species and fulfills the nutritional requirements of the neonate, for instance: (i) the energetic part (provided by lipids, lactose, and in excess by proteins), essential amino acids, and (ii) amino groups necessary for the biosynthesis of non-essential amino acids (provided by proteins), essential fatty acids, vitamins, inorganic elements, and water.<sup>1</sup>

Global milk production has increased by about 20% in the last decade, from 694 million tons in 2008<sup>2</sup> to 843 million tons in 2018.<sup>3</sup> As a result, bovine milk is the most consumed food product representing about 48% of the total milk consumed globally, the European Union (EU), Australia, and New Zealand being the most important producers, followed by the United States and India.<sup>4</sup>

Collection and processing expose milk to different contaminants, mainly pesticide residues, metals, mycotoxins, hormones, and others reaching the cow through feeding or drug administration by producers.<sup>5</sup> Thus, milk can contain hazardous materials, of either biological or chemical origin.

Although pasteurization has been an efficient antimicrobial method and has contributed to reducing many diseases, several infectious episodes associated with pasteurized milk have continued to occur, mainly when raw milk has an exaggerated population of microorganisms that increase the margin of survival and by post pasteurization contamination.<sup>6</sup> The biggest problem of pathogens in pasteurized milk is that they persist without causing any organoleptic alteration, increasing sanitary risk since the consumer cannot suspect their presence, showing that pasteurization has some drawbacks in treating pathogens.<sup>7</sup>

As population and industrial growth increased, new contaminants appeared, and with this, contamination of cow's milk also increased not only by compounds of biological origin but also by compounds of chemical origin, as mentioned above.<sup>8</sup> However, pasteurization has remained the only established treatment, even though it is only effective for eliminating most biological and non-chemical compounds.<sup>9</sup> In contrast, the literature mentions very few alternative treatments to treat chemical contaminants in cow's milk, leading to a critical analysis of their application to ensure sufficient quality in the milk consumed. Given this evidence, the bibliographic review here aims to identify the different types of contaminants in raw/pasteurized cow's milk and analyze the application of alternative processes for the elimination or degradation of contaminants.

## 2. Contaminants present in cow's milk

There are several hazards of contamination of cow's milk, ranging from biological to chemical compounds. The risk of biological contamination of cow's milk derives mainly from cattle milking due to the exposure of udders to the environment, equipment, storage, dirty pipes, and others.<sup>10</sup> Chemical contamination of cow's milk comes from several sources: application of agrochemicals,<sup>11</sup> use of legal or illegal veterinary products,<sup>12</sup> feed and forages contaminated with natural toxins,<sup>13</sup> or through the improper use of chemicals during milk production, processing and packaging stages.<sup>14</sup>

Figure 1 shows the direct and indirect pathways for contaminants entry into bovine milk.

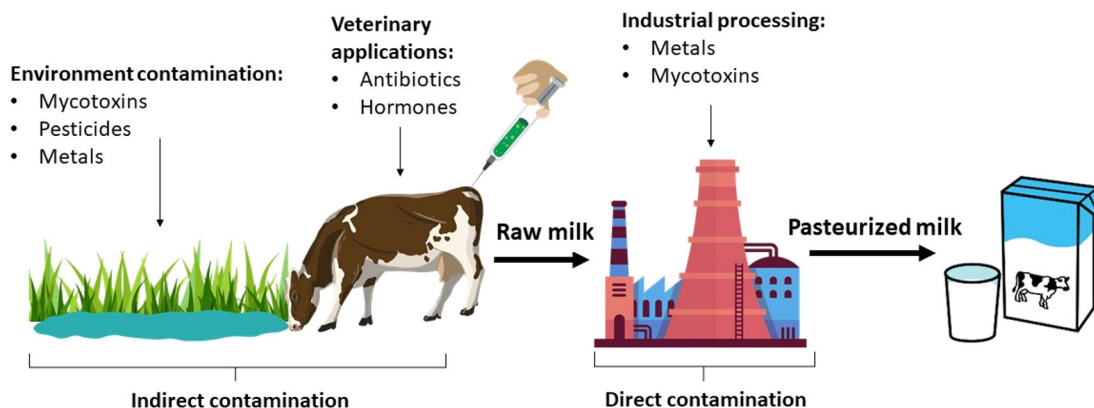
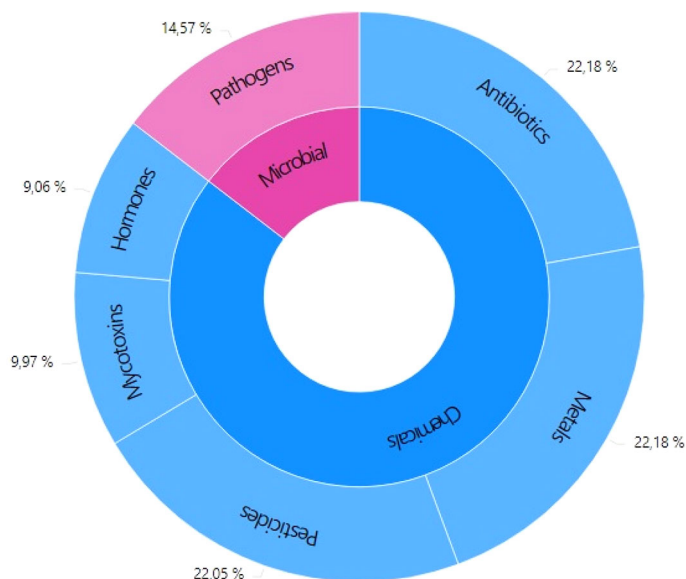


Figure 1. Sources of contamination of bovine milk.



**Figure 2.** Distribution of literature related to contaminants in bovine milk between 2010-2021.

Indirect contamination is associated with the ingestion of contaminants both from the environment and from substances of veterinary use. The most common environmental contaminants are mycotoxins, pesticides, and metals consumed by cattle through feed, forages, and water. In addition, antibiotics and hormones are administered to the cow orally, by injection, or as intramammary infusions to treat diseases, promote animal growth and increase milk production.<sup>5</sup> On the other hand, direct contamination occurs during milk processing from milking, handling, storage and even pasteurization. During the industrialization process, milk comes into contact with metals, residues of cleaning products, mycotoxins, among others.

For better analysis and understanding, the classification of contaminants according to the origin is microbial contaminants and chemical contaminants (Figure 2).

About 14.57% of the literature reports contamination of cow's milk by pathogenic microorganisms. Although the objective of the pasteurization process is the elimination of these microorganisms, there is evidence of their presence in pasteurized milk, which will be presented later. Although pathogenic microorganisms are considered the main hazard that threatens the safety of milk, they do not represent the highest percentage of reported cases. The contaminants that have been more reported in the literature are of chemical origin (Figure 2). Among chemical contaminants, metals, pesticides, and antibiotics stand out. Among chemical contaminants, the most reported are heavy metals (22.18%), pesticides (22.05%), and antibiotics (22.18%); due to bad practices in agriculture and cattle. Although reports of mycotoxins in milk are relatively low (9.97%), they are of great importance due to the increase in reported cases of contamination with Aflatoxin M1 (AFM1). The International Agency for Research on Cancer has classified AFM1 as a carcinogenic substance.<sup>15</sup> This means that the food safety of milk is at risk, as any of these compounds compromise the health of the final consumer. Below is a detailed classification of the different types of contaminants present in both raw and pasteurized milk and the negative effects they have on consumer health.

### 2.1 Microbial contaminants

The presence of several pathogenic microorganisms has been reported in raw and pasteurized cow's milk (Table 1). Microbial contamination of raw milk can be due to diseases such as mastitis, improper handling on production farms, milking equipment, water sources, and feeding of cattle, utensils, and equipment used for milk storage on the farm or during transport.<sup>16</sup> Likewise, poor hygienic practices within the dairy industry can lead to the formation of biofilms on the sprinklers of cooling systems, pipes, cooling tanks, storage, and transport tanks. The contact of pasteurized milk with these surfaces increases the risk of contamination with pathogenic microorganisms, posing a danger to the consumer and the quality of the product.<sup>17</sup>

According to Table 1, Most cases of contamination are recorded in raw milk due to inadequate milking, processing, storage, and transport conditions. On the other hand, although few studies report the presence of microorganisms in

**Table 1. Pathogens in bovine milk reported in literature.**

Pathogens	Type of milk	Reference
<i>Mycobacterium</i>	Raw	18,19
<i>Pseudomonas</i>	Raw	20,21,22,23,24,25,16,26,27,28,29
	Pasteurized	16
<i>Hafnia</i>	Raw	25,23,21
<i>Serratia</i>	Raw	22,25,24,21
<i>Klebsiella</i>	Raw	30
	Pasteurized	30
<i>Citrobacter</i>	Raw	25
<i>Escherichia</i>	Raw	16,30,31
	Pasteurized	16,30
<i>Staphylococcus</i>	Raw	32,27,28,29
<i>Bacillus</i>	Raw	27,28,20,21
<i>Lactococcus</i>	Raw	32,26,21,27,28,20
<i>Corynebacterium</i>	Raw	28
<i>Streptococcus</i>	Raw	27,28
<i>Enterobacter</i>	Raw	27,28,25
<i>Mycoplasma</i>	Raw	27
<i>Enterococcus</i>	Raw	27,28,21,32
<i>Acinetobacter</i>	Raw	27,28,20,29,26,33,32
<i>Sneathia</i>	Raw	27
<i>Kocuria</i>	Raw	27,28,32
<i>Neisseria</i>	Raw	27
<i>Fusobacterium</i>	Raw	27
<i>Macrococcus</i>	Raw	27
<i>Trueperella</i>	Raw	27
<i>Halomonas</i>	Raw	27
<i>Micrococcus</i>	Raw	27
<i>Enhydrobacter</i>	Raw	27
<i>Psychrobacter</i>	Raw	27,28
<i>Campylobacter</i>	Raw	34,31,35
<i>Brachybacterium</i>	Raw	28
<i>Dermacoccus</i>	Raw	28
<i>Leucobacter</i>	Raw	28
<i>Microbacterium</i>	Raw	28,20
<i>Aerococcus</i>	Raw	28
<i>Lactobacillus</i>	Raw	28,33
<i>Ochrobactrum</i>	Raw	28
<i>Pantoea</i>	Raw	28
<i>Paracoccus</i>	Raw	28
<i>Sphingomonas</i>	Raw	28
<i>Deinococcus</i>	Raw	28
<i>Aspergillus</i>	Raw	28

**Table 1.** Continued

Pathogens	Type of milk	Reference
<i>Cladosporium</i>	Raw	28
<i>Eurotium</i>	Raw	28
<i>Penicillium</i>	Raw	28
<i>Wallemia</i>	Raw	28
<i>Listeria</i>	Raw	31,36
<i>Yersinia</i>	Raw	31,36
<i>Salmonella</i>	Raw	30,16
	Pasteurized	30,16
<i>Vibrio</i>	Raw	30
	Pasteurized	30
<i>Stenotrophomonas</i>	Raw	33,32
<i>Chryseobacterium</i>	Raw	33
<i>Paenibacillus</i>	Raw	20,21
<i>Coliforms</i>	Pasteurized	37

pasteurized milk, it is doubtful that it is an efficient process for their elimination. The main types of microorganisms present in milk are bacteria, yeasts, and molds, which represent the different types of microorganisms present in cow's milk. The presence of *Corynebacteria*, *Staphylococcus*, *Streptococcus*, *Bacillus*, and *Micrococcus* species has been evidenced in the teat of dairy cattle.<sup>38,39</sup> These microorganisms have also been identified in cow's milk,<sup>27,28,40,41</sup> demonstrating that during milking, milk can become contaminated by contact with the cow's teat under unhygienic conditions. On the other hand, as a result of mastitis, *Staphylococcus* and *Streptococcus* species have been identified in bovine milk samples,<sup>42,43</sup> with *Staphylococcus aureus* being the main cause of mastitis.<sup>43</sup> The presence of *Enterobacteriaceae*, *Pseudomonas* spp., *Staphylococcus* spp., and lactic acid bacteria has been identified in the equipment used for milking.<sup>17</sup> It is evident that the conditions under which milk is obtained on farms are not the most adequate because these different microorganisms are found in cow's milk.<sup>33,42,44,45</sup>

Consumption of milk contaminated by pathogenic microorganisms such as *Campylobacter*, *Salmonella*, *Yersinia*, *E. coli*, *Listeria*, and *S. aureus* can cause muscle and stomach pain, gastrointestinal diseases with diarrhea, fever, and nausea.<sup>31</sup> These microorganisms are commonly found in the intestinal flora or in the udder of cows, thus facilitating milk contamination.<sup>31</sup> In addition, *Campylobacter* spp. and *E. Coli* O157:H7 are capable of producing Guillain-Barrés syndrome and hemolytic uremic syndrome, respectively.<sup>46</sup>

## 2.2 Chemical contaminants

For a more detailed analysis, the chemical contaminants found in cow's milk have been classified into five groups: pesticides, metals, antibiotics, mycotoxins, and hormones (Table 2).

### 2.2.1 Pesticides

A variety of pesticide residues in detectable amounts in raw milk, pasteurized, and UHT (ultra-high temperature) milk has been reported by several authors. This is due, among other factors, to the lipophilic properties and resistance to biodegradation of these types of contaminants.<sup>8</sup> There are three possible forms in which pesticides can enter the animal's body<sup>172</sup>: (i) through contaminated water, (ii) through the pores of the skin when the animal is sprayed or soaked to treat ectoparasites, and (iii) through contaminated feed and forage, the latter being the main source of entry.

- (i) The presence of organophosphorus pesticide residues (malathion, methyl-parathion, diazinon, ethion) was identified. The average concentrations detected were 0.032-0.78, 0.13, 0.32-0.74, 0.010 µg/L for malathion, methyl-parathion, diazinon and ethion, respectively.<sup>62,173</sup> Fipronil and chlorpyrifos were other pesticides found in water samples supplied to livestock.<sup>174,175</sup> Ashoub & Azam<sup>176</sup> identified DDT (Dichloro diphenyl trichloroethane), aldrin, heptachlor epoxide, lindane, methoxychlor, diazinon, and deltamethrin in water samples from cattle farms. These same compounds have been identified in cattle drinking water and in cow's milk.<sup>11,47,51,54,55,62,65,177-180</sup> This verifies that water contaminated by pesticides and supplied to cattle is one of the main routes of contamination of raw cow's milk.

**Table 2. Chemical contaminants in bovine milk reported in literature.**

Compounds		Type of milk	MRL <sup>a</sup> (µg/kg)	MRL <sup>b</sup> (µg/kg)	Reference
Pesticides	Hexachlorocyclohexane (HCH)	Raw	-	-	11,47,48,49,50,51
		Pasteurized	-	-	52,53,51
		UHT	-	-	47
	Butachlor	Raw	-	-	11
		Pasteurized	-	-	54
	Cyhalothrin	Raw	30	50	11,55,56
		Pasteurized	-	-	54
	Cypermethrin	Raw	100	20	11,56,57
		Pasteurized	-	-	54,58,59
	Fenvalerate	Raw	-	40	11
		Pasteurized	-	-	59
	Deltamethrin	Raw	30	20	11,55,56,57,55
		Pasteurized	-	-	60
	Malathion	Raw	-	-	11,61,62
	Chlorpyrifos	Raw	-	-	11,55,56,61,55,60
		Pasteurized	-	-	54,58,59,60
		UHT	-	-	60
	Carbofuran	Raw	-	-	55,62
	Permethrin	Raw	-	50	56,57
		Pasteurized	-	-	54,58,59
	Profenophos	Raw	-	-	11,60
		Pasteurized	-	-	54,60
		UHT	-	-	60
	Ethion	Raw	-	-	11
		Pasteurized	-	-	54,63
	Dichloro diphenyl trichloroethane (DDT)	Raw	-	-	11,64,65,47,66,48,50,51
		Pasteurized	-	-	64,52,54,67,51
		UHT	-	-	47
	Dicofol	Pasteurized	-	-	59
	Aldrin+Dieldrin	Raw	-	-	64,47,49
Pasteurized		-	-	64,53	
UHT		-	-	47	
Endrin	Raw	-	-	68,69,49	
	Pasteurized	-	-	70,53,54,67	
Fipronil	Raw	-	-	11,65,60	
	Pasteurized	-	-	54,60	
Hexaflumuron	Raw	-	-	65	
Teflubenzuron	Raw	-	-	65	
Diflufenican	Raw	-	-	65	
Piperophos	Raw	-	-	65	
Dimethoate	Raw	-	-	60,62	
	Pasteurized	-	-	60	

Table 2. Continued

Compounds		Type of milk	MRL <sup>a</sup> (µg/kg)	MRL <sup>b</sup> (µg/kg)	Reference
	Atrazine	Pasteurized	-	-	58,59
	Diazinon	Raw	-	20	62,60
		Pasteurized	-	-	58,59,60
		UHT	-	-	60
	Lindane	Raw	-	-	64,51
		Pasteurized	-	-	64,51
	Endosulfane	Raw	-	-	11,65,47,68,48,71,49,51,56
		Pasteurized	-	-	52,70,53,54,67
		UHT	-	-	47
	Hexachlorobenzene	Raw	-	-	72
		Pasteurized	-	-	70,58,59
	Heptachlor epoxide	Raw	-	-	65,47,73,68
		Pasteurized	-	-	59
		UHT	-	-	47,73
	Heptachlor	Raw	-	-	68,69,51
		Pasteurized	-	-	73,52,70,51
		UHT	-	-	73
	Chlordane	Pasteurized	-	-	52,53,67
	Methoxychlor	Raw	-	-	47,69
		Pasteurized	-	-	54
		UHT	-	-	47
	Azoxystrobin	Pasteurized	-	-	74
	Chlorantranilliprole	Pasteurized	-	-	74
	Flubendiamide	Pasteurized	-	-	74
	Imidacloprid	Raw	-	-	55
		Pasteurized	-	-	74
	Lufenuron	Pasteurized	-	-	74
	Metalaxyl	Pasteurized	-	-	74
	Novaluron	Pasteurized	-	-	74
	Uniconazol	Pasteurized	-	-	74
	Monuron	Pasteurized	-	-	75
	Methabenzthiazuron	Pasteurized	-	-	75
	Buturon	Pasteurized	-	-	75
	Linuron	Pasteurized	-	-	75
	Aziprotryne	Pasteurized	-	-	75
	Bitertanol	Pasteurized	-	-	75
	Clofentezine	Pasteurized	-	-	75
	Methyl Parathion	Raw	-	-	62,76
Metals	Cadmium	Raw	-	-	77,78,79,80,81,82,83,84,85,86
		Pasteurized	-	-	87,77,88
		UHT	-	-	89,90



**Table 2.** *Continued*

Compounds		Type of milk	MRL <sup>a</sup> (µg/kg)	MRL <sup>b</sup> (µg/kg)	Reference
Lead	Raw	-	-	77,91,78,79,80,81,82,83,84,92,85,86	
	Pasteurized	-	-	87,77,93,88	
	UHT	-	-	92,94,89,90	
Copper	Raw	-	-	77,79,80,81,82,84,92,89,88,85,86	
	Pasteurized	-	-	87,77,93,88	
	UHT	-	-	92,94,89,90	
Zinc	Raw	-	-	77,95,80,81,82,88,85,96,86	
	Pasteurized	-	-	77,93,88,96	
	UHT	-	-	94,89,90	
Selenium	Raw	-	-	82,85,96,86	
	Pasteurized	-	-	96	
	UHT	-	-	94	
Chromium	Raw	-	-	77,91,88,85,96,86	
	Pasteurized	-	-	77,93,88,96	
	UHT	-	-	90	
Nickel	Raw	-	-	77,91,79,97,88,85,86	
	Pasteurized	-	-	77,93,88	
	UHT	-	-	94,89	
Iron	Raw	-	-	80,82,89,88,85,96,86	
	Pasteurized	-	-	93,88,96	
	UHT	-	-	94,89,90	
Arsenic	Raw	-	-	98,91,83,84,97,88,85,96,86	
	Pasteurized	-	-	88,96	
Magnesium	Raw	-	-	95,82,83,88,85,86	
	Pasteurized	-	-	93,88	
	UHT	-	-	90	
Manganese	Raw	-	-	82,89,88,85,86	
	Pasteurized	-	-	93,88	
	UHT	-	-	89,90	
Aluminum	Raw	-	-	98,91,85,96,86	
	Pasteurized	-	-	96	
Molybdenum	Raw	-	-	98,86	
Mercury	Raw	-	-	91,84,99,97,88,86	
	Pasteurized	-	-	88	
	UHT	-	-	94	
Tin	Raw	-	-	97,85,86	
Cobalt	Raw	-	-	77,79,89,88,86	
	Pasteurized	-	-	77,88	
	UHT	-	-	94,89	
Antibiotics	Oxytetracycline	Raw	100	100	100,101,102,103,104,105
		Pasteurized	-	-	106,58,107,102,108,105,109
		UHT	-	-	102,108

Table 2. Continued

Compounds		Type of milk	MRL <sup>a</sup> (µg/kg)	MRL <sup>b</sup> (µg/kg)	Reference
Lincomycin	Raw	Raw	150	150	100,101,110
	Pasteurized	Pasteurized	-	-	111,112
	UHT	UHT	-	-	111
Quinolone	Raw	Raw	-	-	104,113
	Pasteurized	Pasteurized	-	-	111,114,113
	UHT	UHT	-	-	111,114
Tetracycline	Raw	Raw	100	100	102,103,115,116,104,105,110
	Pasteurized	Pasteurized	-	-	111,114,117,107,102,108,109
	UHT	UHT	-	-	111,117,102,108
Doxycycline	Raw	Raw	-	-	103,104
	Pasteurized	Pasteurized	-	-	106
	UHT	UHT	-	-	108
Penicillin G	Raw	Raw	-	-	101,118,119,120,121,122
	Pasteurized	Pasteurized	-	-	106,109
Trimethoprim	Raw	Raw	-	50	123
Amoxicillin	Raw	Raw	4	4	124,119,120,121,122
	Pasteurized	Pasteurized	-	-	58,109
Cefalexin	Raw	Raw	-	100	120,125
Cephapirin	Raw	Raw	-	60	101,120
Fleroxacin	Raw	Raw	-	-	126
Chlortetracycline	Raw	Raw	100	100	102,104
	Pasteurized	Pasteurized	-	-	102
	UHT	UHT	-	-	102,108
Enrofloxacin	Raw	Raw	-	100	126,127,115,116,128,105,113,129,119,120,122,110
	Pasteurized	Pasteurized	-	-	127,108,105
	UHT	UHT	-	-	108
Ciprofloxacin	Raw	Raw	-	100	126,127,103,129,119,120,122
	Pasteurized	Pasteurized	-	-	127,108
	UHT	UHT	-	-	108
Lomefloxacin	Raw	Raw	-	-	126
Tilmicosin	Pasteurized	Pasteurized	-	50	112,130
Erythromycin A	Pasteurized	Pasteurized	-	40	130
Tylosin	Raw	Raw	100	50	103,116
	Pasteurized	Pasteurized	-	-	112,109
Spiramycin	Pasteurized	Pasteurized	200	200	112
Streptomycin	Raw	Raw	200	200	131,116,128,110
	Pasteurized	Pasteurized	-	-	111
	UHT	UHT	-	-	111
Gentamicin	Raw	Raw	200	100	131,116,128
	Pasteurized	Pasteurized	-	-	109
	UHT	UHT	-	-	108

**Table 2.** *Continued*

Compounds		Type of milk	MRL <sup>a</sup> (µg/kg)	MRL <sup>b</sup> (µg/kg)	Reference
	Gatifloxacin	Raw	-	-	127
		Pasteurized	-	-	127
	Ofloxacin	Raw	-	-	127
		Pasteurized	-	-	127,132
	Norfloxacin	Raw	-	-	127
		Pasteurized	-	-	127,108
		UHT	-	-	108
	Sulfamethoxazole	Raw	-	-	127,103,105,123
		Pasteurized	-	-	127
	Sulfamethazine	Pasteurized	-	-	127,58
		UHT	-	-	114
	Sulfadimethoxine	Raw	-	-	103
		Pasteurized	-	-	58
	Sulfadiazine	Pasteurized	-	-	133
	Sulfathiazole	Pasteurized	-	-	58
	Ceftiofur	Raw	-	-	103
	Sulfonamides	Raw	-	-	116,128,104
		Pasteurized	-	-	114
		UHT	-	-	114
	Cefazolin	Raw	-	50	101,125
Cephoperazone	Raw	-	50	101,119,120,122,125	
Dicloxacillin	Raw	-	30	101,119,120,121,122	
Ampicillin	Raw	-	4	101,120,121	
Cloxacillin	Raw	-	30	101,134,120,121	
Cefacetile	Raw	-	125	101	
Chloramphenicol	Raw	-	-	116,128,104	
Rifaximin	Raw	-	-	101	
Mycotoxins	Aflatoxin M <sub>1</sub>	Raw	0.5	0.05	135,136,137,138,139,140,141,142,143
		Pasteurized	-	-	135,144,137,140,141,142,145,146
		UHT	-	-	137,141,147,148,145,146
	Ochratoxin A	Raw	-	-	135,136,137
		Pasteurized	-	-	135,144,136,149
	α-zearalenol	Raw	-	-	135,137,150
		Pasteurized	-	-	135
	Fumonisin B1	Raw	-	-	137
		Pasteurized	-	-	144,137
	Fumonisin B2	Pasteurized	-	-	144
	β-zearalenol	Raw	-	-	150
		Pasteurized	-	-	144
Zearalenone	Raw	-	-	136,144,137,151,150,135,152,153	
	Pasteurized	-	-	135,154	

Table 2. Continued

Compounds		Type of milk	MRL <sup>a</sup> (µg/kg)	MRL <sup>b</sup> (µg/kg)	Reference
	Aflatoxin B1	Raw	-	-	136,137,140
		Pasteurized	-	-	144,136,137,140
		UHT	-	-	137
	Aflatoxin B2	Raw	-	-	137
		Pasteurized	-	-	137
	Aflatoxin G1	Raw	-	-	137
		Pasteurized	-	-	137
	Aflatoxin G2	Raw	-	-	137
		Pasteurized	-	-	137
	Zearalanol	Raw	-	-	137
	α-zearalenone	Raw	-	-	137
	Cyclopiazonic acid	Pasteurized	-	-	137
	α-zearalanol	Pasteurized	-	-	155
	Deepoxy-deoxynivalenol	Raw	-	-	137
	Deoxynivalenol	Raw	-	-	151,152,150
Aflatoxin M2	Raw	-	-	138	
	Pasteurized	-	-	137	
	UHT	-	-	137,151	
Hormones	Leptin	Pasteurized	-	-	124,156
	Triiodothyronine and Thyroxine	Pasteurized	-	-	156
	Prednisolone	Raw	-	6	124,138
	Relaxin	Pasteurized	-	-	156
	Insulin	Raw	-	-	157
		Pasteurized	-	-	156,157
	Oxytocin	Pasteurized	-	-	156,158,157,159
	Adiponectin	Raw	-	-	160
		Pasteurized	-	-	156
	Estriol	Raw	-	-	161
		UHT	-	-	160,162
	17α-Estradiol	Raw	-	-	161
		Pasteurized	-	-	162
		UHT	-	-	161,162
	17β-Estradiol	Raw	-	-	161,163
		Pasteurized	-	-	164,163,162,165,166
		UHT	-	-	161,162
	Estrone	Raw	-	-	161,167,163
		Pasteurized	-	-	164,163,162
		UHT	-	-	161,162
	Testosterone	Raw	-	-	161,167
Pasteurized		-	-	161	
4-Androstenediol	Raw	-	-	161	

**Table 2.** Continued

Compounds	Type of milk	MRL <sup>a</sup> (µg/kg)	MRL <sup>b</sup> (µg/kg)	Reference
5-Androstenediol	Raw	-	-	161
4-Androstenedione	Raw	-	-	161
Progesterone	Raw	-	-	161,167,168,169
	Pasteurized	-	-	168
	UHT	-	-	161
17 $\alpha$ -Hydroxyprogesterone	Raw	-	-	161
Cortisone	Raw	-	-	161
Cortisol	Raw	-	-	161,170
Corticosterone	Raw	-	-	161
Hydrocortisone	Pasteurized	-	-	171
Insulin-like Growth factor-I	Raw	-	-	157
	Pasteurized	-	-	58,157,171
Pregnenolone	Raw	-	-	167
Androstenedione	Raw	-	-	167
	Pasteurized	-	-	167
Dehydroepiandrosterone	Raw	-	-	167
5- $\alpha$ -Androstane-3,17-dione	Raw	-	-	167
Prolactin	Pasteurized	-	-	156
Growth Hormone	Pasteurized	-	-	156,58

MRLa: Maximum Residue Levels by Codex Alimentarius; MRLb: Maximum Residue Levels by European Union, EU. UHT: ultra-high temperature.

- (ii) According to the analysis of Table 2, Claborn *et al.*<sup>181</sup> report the presence of malathion residues in cow's milk after cattle were sprayed with this pesticide for the treatment of ectoparasites. Malathion was found to be completely secreted from the udder 24 hours after application. In contrast to malathion, lindane was reported not to be completely excreted in milk until seven days after application to the cow's skin.<sup>182</sup> Residues of chlorpyrifos and ethion have been found in cow milk up to 24 and 72 hours after application, respectively.<sup>183</sup> This confirms that skin contaminated with these pesticides is another route of contamination of raw cow's milk.
- (iii) In forage, concentrations of 0.02 mg kg<sup>-1</sup> of DDT residue were reported.<sup>184</sup> The presence of cypermethrin, chlorpyrifos, cyhalothrin, and deltamethrin in forage was reported in a range of mean concentrations between 1.03-6.01 ng g<sup>-1</sup>. In addition to the presence of pesticides in forages, residues of lindane, DDT, fenvalerate, ethion, malathion, profenofos were also reported in feed. The mean concentrations of these varied in the range of 0.63-4.05 ng g<sup>-1</sup>.<sup>175</sup> The presence of deltamethrin in feed was also reported in a concentration range of 41.99-381.30 µg kg<sup>-1</sup>.<sup>185</sup> Another investigation revealed the presence of malathion, dimethoate, methyl-parathion, diazinon in the feed fed to cattle. The range of detected concentrations was between 0.01-80.45 µg L<sup>-1</sup>.<sup>62</sup> All the contaminants reported in forage and feed were also detected in cow's milk.<sup>11,54,55,57,60-62,177,179,186</sup> Thus, like water, pesticide-contaminated forage and feed are a route of contamination as they are directly ingested by cattle and excreted through cow's milk.

Pesticides are one of the most commonly found contaminants, not only in raw cow's milk but also after the pasteurization and UHT process. Their presence in milk, even below the maximum permitted levels, represents a health risk to the consumer. It is related to Hodgkin's disease (HD), non-Hodgkin's lymphoma (NHL), Parkinson's disease, endocrine disruption, respiratory and reproductive disorders, among others.<sup>187</sup>

It is important to note that organochlorine pesticides such as hexachlorocyclohexane, dichloro diphenyl trichloroethane, and endosulfane are still present despite having been banned since the 1970s because of their high persistence in the environment and their harmful effects on human health,<sup>188</sup> are still detected in cow's milk. This indicates that they are still used in agriculture and animal husbandry. With a few exceptions (cyhalothrin, cypermethrin, fenvalerate, deltamethrin,

permethrin, and diazinonella), the vast majority of pesticides found in cow's milk are not regulated by Codex and the EU. This demonstrates the low efficiency of the regulatory controls of these contaminants in the unprocessed and post-processed product, leading to an inefficient safety of this food product.

### 2.2.2 Metals

Although metals are found in the environment either naturally or due to industrial and/or agricultural activities, there are several routes by which they reach the milk. Namely, ingestion of contaminated food, fodder, and/or contaminated drinking water. In the soil, they are absorbed by many crop plant species, which, when ingested by animals, are transferred to the lactating glands and finally excreted in milk.<sup>172</sup> Equipment used in the dairy industry is another source of contamination directly to milk with metals such as chromium and nickel.<sup>189</sup> Heavy metals such as cadmium, lead, mercury, and arsenic reach milk by indirect contact through feed consumed by cattle.<sup>189</sup> Although the literature does not report the presence of metals in water or fodder destined for cattle, as well as in pesticides, these can be another of the main routes of contamination.

Several heavy metals have been reported in the literature to be found in raw cow's milk. The metals least found in studies of raw cow milk are tin and molybdenum. These elements are not abundant in nature, and their presence in fodder or water for animal consumption will depend on soil characteristics, while the most reported are lead, cadmium, copper, and zinc, due to environmental pollution produced by man mainly in industrial activities.<sup>79,190</sup> Minerals such as Fe, Cu, and Zn are necessary for various biological functions. However, high concentrations of these minerals have negative effects on human health.<sup>96</sup> Lead is one of the non-essential metals classified as carcinogenic to humans by the International Agency for Research on Cancer.<sup>191</sup> Cadmium is associated with the formation of human lung, kidney, breast, prostate, urinary tract cancer because it affects cell proliferation, differentiation, and other cellular activities.<sup>192</sup>

None of the heavy metals reported in the literature consulted have established maximum residue limits (MRLs) by Codex<sup>193</sup> and the EU.<sup>194</sup> However, these contaminants are known to represent a high risk to human health. Stricter control measures should be adopted in the dairy industry, considering that cow's milk is one of the most consumed products by humans worldwide.

### 2.2.3 Antibiotics

Antibiotics are used in livestock activities in three basic ways: therapeutic, prophylactic, and growth promoters. About 80% of dairy cattle are subjected to antibiotic treatments on at least one occasion throughout their lives, mostly used as growth promoters and for the treatment of various diseases such as mastitis, arthritis, respiratory diseases, gastrointestinal diseases, and bacterial infections.<sup>195</sup> Cows eliminate antibiotics and their metabolites through milk, depending on the dose and route of application, level of milk production, type and degree of mammary disease, and time between treatment and milking. On the other hand, oral, intramuscular, or intravenous administration is less important from the point of view of milk hygiene than intramammary application. However, intramammary antibiotics are easy to apply and generally cheaper, so they are preferred in dairy farms.

The most common disease in dairy cows is mastitis, whose treatment includes the wide use of tetracyclines,  $\beta$ -lactams, oxytetracycline, difloxacin, among others, being the  $\beta$ -lactams of greater application.<sup>8</sup> Within the latter group, the most employed are penicillin, ampicillin, and amoxicillin.<sup>196</sup> According to the literature, the presence of antibiotics in milk has been evidenced, highlighting tetracycline, oxytetracycline, penicillin, and amoxicillin.<sup>103,124,197,198</sup> While other antibiotics less reported in milk were rifamixin, gatifloxacin, spiramycin, and lomeflaxacin, with no indication in the studies of the purpose of their application in cattle.<sup>101,112,126,127</sup>

The consumption of contaminated milk with antibiotic residues is an emerging public health problem worldwide. Therefore, it is important to control the presence of antibiotic residues in food to avoid the appearance of resistance to these antibiotics in humans. The presence of antibiotics at concentrations even below the MRL in milk can cause undesirable effects on human health such as ototoxicity and nephrotoxicity,<sup>199</sup> endocrine disruption,<sup>200</sup> hypersensitivity, and especially bacterial resistance.<sup>130</sup> According to the literature consulted, 43 antibiotics present in cow's milk have been identified, of which 18 are not regulated by Codex<sup>193</sup> and EU standards.<sup>194</sup>

Considering that the use of antibiotics in cattle generates residues in milk, their excessive use should be avoided, and the elimination times before milking should be respected in order to avoid the presence of these contaminants.

## 2.2.4 Mycotoxins

The quality of food products is commonly affected by toxin contamination, of which 60 to 80 % are caused by mycotoxins.<sup>201</sup> This means a risk for human health and great economic losses in the industrial sector.

Mycotoxins are natural contaminants produced by *Aspergillus*, *Penicillium*, and *Fusarium* fungi,<sup>154</sup> the most prominent being AFM1, which results from the metabolism of aflatoxin B1 in the liver of contaminated animals.<sup>15,143</sup> In the 1960s, the first reported case of aflatoxin contamination was reported for the first time, beginning the concern for this type of contaminant. Even during this decade, high consumption of feed contaminated by this mycotoxin was reported, which led to indirect contamination of cow's milk for consumption, compromising the safety of this product.<sup>202</sup> Therefore, it is considered that the main routes of entry of mycotoxins into milk are contaminated crops and feed ingested by cows.<sup>136</sup>

It is known that approximately 0.3-6.2% of AFB1 (Aflatoxin B1) present in animal feed is converted to AFM1.<sup>15</sup> This mycotoxin is neither degraded nor removed by industrial food processes such as pasteurization and sterilization, nor by the cooking of feed.<sup>203</sup> This represents a difficult problem to deal with at the industrial level due to the stability of mycotoxins in general to thermal, physical, and chemical treatments.<sup>204</sup>

AFM1 mycotoxin is the only regulated by Codex<sup>193</sup> and EU<sup>194</sup> and the most reported in cow's milk according to the literature. However, other abundant mycotoxins have been identified in this food product, such as ochratoxin A and zearalenone. The fungi of the genus *Aspergillus* and *Penicillium* produce Ochratoxin A, while fungi of the genus *Fusarium* produces zearalenone, commonly found in cattle feed.<sup>138</sup> On the other hand, aflatoxin G2, aflatoxin G1, aflatoxin B2, and zearalanol show a lower incidence in cow's milk. The literature on the effects on human health associated with the ingestion of mycotoxin-contaminated milk is scarce or almost non-existent, unlike AFM1. Therefore, studies on this type of contaminants should be expanded.

## 2.2.5 Hormones

The use of hormones in the livestock industry increases production yields and medical treatments. Their fat-soluble characteristics favor their high persistence and presence in cow's milk due to the high-fat contents.<sup>156</sup> Therefore, the supply of hormones to cattle represents a form of direct contamination that, like other contaminants, is excreted through milk. However, the European Union banned the use of hormones through the Directive 96/22/EC, and enforcement is regulated by Directive 96/23/EC.<sup>165</sup>

Prednisolone in combination with amoxicillin and clavulanic acid is used to treat mastitis in cows' udders,<sup>205</sup> being an access route of this contaminant to milk. The 17 $\beta$ -estradiol and progesterone, with the highest presence in cow milk, are sex hormones widely used to induce lactation, improve fertility and synchronize the estrous cycle.<sup>8,168</sup> The hormones least found in studies in milk were testosterone, somatostatin, and cortisone. The presence of estrogens in cow's milk has been linked to diseases such as breast cancer<sup>206</sup> and conditions in the gastrointestinal tract.<sup>156</sup> Other diseases associated with the presence of hormones in cow's milk have included acne, prostate cancer, uterine cancer, and male reproductive disorders.<sup>167</sup>

Table 2 shows that several hormones are frequently present in cow's milk, with prednisolone being the only one regulated by the EU.<sup>194</sup> This indicates that regulations should be established for different hormones considering that they are the chemical compounds mostly used to increase milk production yield to preserve quality and consumer safety.

## 3. Pasteurization process in cow's milk

The principles and name of pasteurization come from the studies of the French scientist Louis Pasteur. His interest in milk and other food products was due to their putrefaction, which he later attributed to the growth of undesirable microorganisms.<sup>207</sup> Several pathogenic microorganisms are found in raw milk: *Pseudomonas*, *Enterobacter*, *Bacillus*, *Clostridium*, *Microbacterium*, and *Micrococcus*. Pathogenic microorganisms in cow's milk have been linked to infectious diseases such as campylobacteriosis, salmonellosis, yersiniosis, listeriosis, tuberculosis, brucellosis, staphylococcal enterotoxin intoxication, streptococcal infections, and *Escherichia coli* O157: H7 infection.<sup>208</sup>

It was not until the end of the 1880s that heat treatment began to be used to commercialize milk. This arose with the main objective of inactivating *Mycobacterium tuberculosis*, the cause of tuberculosis in humans associated with the consumption of raw milk. Thus, pasteurization became a process universally employed by developed countries after World War II. However, there is evidence that not all pathogenic microorganisms can be eliminated during pasteurization, such as *Staphylococcus aureus*, *micrococci*, *Streptococcus spp*, and *Bacillus*.<sup>209</sup> Which calls into question the efficiency of this process.

The US Food and Drug Administration (FDA) establishes a maximum limit for bacteria in raw cow's milk of 100,000 cfu ml<sup>-1</sup> and 20,000 cfu ml<sup>-1</sup> for pasteurized milk.<sup>209</sup>

Pasteurization is a technology classified on the basis of operating temperatures and exposure times as follows: LTLT, HTST, and UHT. Low-temperature long-time pasteurization (LTLT) uses a minimum temperature of 62.8°C and a minimum time of 30 min. High-temperature short-time pasteurization (HTST) uses a minimum temperature of 71.1°C, a minimum time of 15 seconds, and ultra-high temperature pasteurization (UHT) works at a minimum of 135°C and during a minimum time of 1 second.<sup>210</sup> Pasteurized milk under UHT conditions can be stored for several months without refrigeration.<sup>211</sup> Whereas the shelf life of pasteurized milk ranges from 10 to 20 days when kept under refrigerated conditions below 6.1°C.<sup>212</sup>

It has been shown that the application of pasteurization denatures proteins with bacteriostatic capacity, as is the case of lactoferrin. This is a glycoprotein that binds iron, and its complete denaturation has been evidenced losing its inhibitory capacity on *Escherichia coli* under UHT conditions.<sup>213</sup> For this reason, it is suggested that heat treatment should be applied below 75°C to avoid denaturation of proteins with bacteriostatic capacity and at the same time cause inactivation of pathogenic microorganisms.<sup>213</sup>

On the other hand, the HTST process degrades up to 20% of the vitamins (B1, B6, B12, and C) present in milk.<sup>214</sup> This evidence shows that, although pasteurization and UHT have been widely used to eliminate pathogenic microorganisms, it is not entirely efficient for this purpose. There are even losses of milk mineralization, varying its nutritional composition.

The presence of microbial contaminants in different samples of pasteurized milk shows that, although pasteurization aims to eliminate microorganisms present in milk, it is not totally effective. Moreover, with the appearance of other contaminants, the quality of milk no longer depends only on the presence of microorganisms. It is, therefore, necessary to study other methods of decontamination to ensure the safety and health of consumers.

#### 4. Alternative methods for the treatment of cow's milk

International regulations require maximum limits for microbial and chemical contaminants to ensure the quality of drinking milk. Pasteurization is a technology widely used in the dairy industry. However, it is exclusive for the elimination of microbial contaminants. The literature mentions alternatives for eliminating specific microbial and chemical contaminants (Table 3).

Supercritical carbon dioxide has been used as an inactivating agent for *E. coli*, where the greatest reduction in the content of microorganisms was observed during a residence time of 20 minutes, achieving almost complete inactivation after 70 minutes.<sup>215</sup> Complete inactivation of coliforms, molds, and yeasts was achieved, while a maximum reduction of aerobic bacteria of 4.96 log was obtained using high-pressure carbon dioxide.<sup>221</sup> Using a thin-film UV-C (Ultraviolet-C) reactor with flow-guiding elements allowed a 4.58 log and 3.19 log reduction for *E. coli* and *L. innocua*, respectively.<sup>216</sup> Makarapong *et al.*<sup>218</sup> employed a UV-C reactor for the inactivation of aerobic bacteria achieving a 4.60 log and 4.70 log reduction at 48W and 39W, respectively. UV-C lamp wattage did not significantly influence the fat concentration in the milk, which means that it is necessary to improve the method to guarantee an effective reduction of these microorganisms if milk transport time exceeds two hours without cooling. It was verified that *L. monocytogenes* was completely inactivated in milk with ozone for 15 minutes. However, nutritional values were affected.<sup>219</sup> Exposure of milk to Nd:YAG laser did not alter the physicochemical properties of milk, but the percentage of reduction was low for *E. coli* (30%), *Salmonella* sp (25%), yeasts (47%), and *Lactobacillus* sp (30%).<sup>223</sup> The combination of ultrasound with hydrogen peroxide and an active lactoperoxidase system was able to guarantee the microbial quality of milk as it was able to completely inactivate *Staphylococcus aureus*, *Listeria monocytogenes*, *Lactobacillus plantarum*, *Lactobacillus pentosus*, *Salmonella Typhimurium*, *Escherichia coli*, and *Pseudomonas fluorescens* at 10 minutes at an amplitude of 125 µm.<sup>220</sup> The application of ultrasound in combination with variations in temperature, time, and constant pressure (manothermosonication) achieved minimal reductions of up to 1.6 log CFU/ml for *E. coli* and *P. fluorescens* and 1.05 log CFU/ml for *S. aureus*. Further studies are needed to ensure effective inactivation using manothermosonication.<sup>225</sup> The application of high pressures (400-600MPa) effectively inactivated (5 log CFU/ml) *E. coli*, *Salmonella* and *L. monocytogenes*, *Enterobacteriaceae*, lactic acid bacteria, and *Pseudomonas* spp.<sup>222</sup> One of the most widely used methods for the inactivation of microorganisms in cow's milk is pulsed electric fields (PEF). This method was applied for the inactivation of *E. coli* and *L. innocua*, achieving a reduction of 2 log CFU/ml.<sup>217</sup> It was found that combining this method with preheating at 50°C achieved a 5-6 log CFU/ml reduction of *Pseudomonas aeruginosa* and a total reduction of *E. coli*, *S. aureus*, and *L. innocua*.<sup>224</sup>

Biosorption methods employing the use of microorganisms prove to be efficient in the removal of pesticides, metals, and mycotoxins. Biosorption with lactic acid bacteria managed to eliminate organophosphate pesticides from cow's milk,



**Table 3. Alternative methods to pasteurization for removal of contaminants in bovine milk.**

Contaminant	Process	Reference	
Pathogens	<i>Escherichia coli</i>	Inactivation with supercritical carbon dioxide technology	215
	<i>Escherichia coli</i> and <i>Listeria innocua</i>	Inactivation using a UV-C thin film reactor	216
		Inactivation by pulsed electric fields	217
	Aerobic bacteria	Reduction by UV-C irradiation	218
	<i>Listeria monocytogenes</i>	Inactivation by ozonation	219
	<i>Staphylococcus aureus</i> , <i>Listeria monocytogenes</i> , <i>Lactobacillus plantarum</i> , <i>Lactobacillus pentosus</i> , <i>Salmonella Typhimurium</i> , <i>Escherichia coli</i> , <i>Pseudomonas fluorescens</i>	Inactivation by combinations of ultrasound, hydrogen peroxide, and active lactoperoxidase system	220
	Aerobic bacteria, coliforms, yeasts, and molds	Inactivation by carbon dioxide at high pressure	221
	<i>Escherichia coli</i> , <i>Salmonella</i> , <i>Listeria monocytogenes</i> , <i>Enterobacteriaceae</i> , <i>lactic acid bacteria</i> , and <i>Pseudomonas</i> spp.	Inactivation by high-pressure processing	222
	<i>Escherichia coli</i> , <i>Salmonella</i> , yeasts, and <i>lactobacillus</i> spp.	Inactivation by ND-YAG laser	223
	<i>Pseudomonas aeruginosa</i> , <i>Escherichia coli</i> , <i>Staphylococcus aureus</i> , and <i>Listeria innocua</i>	Inactivation by pulsed electric fields	224
<i>Escherichia coli</i> , <i>Staphylococcus aureus</i> , and <i>Pseudomonas fluorescens</i>	Inactivation by manothermosonication	225	
Pesticides	Organophosphates (chlorpyrifos, diazinon, fenitrothion, malathion, methyl parathion)	Degradation by lactic acid bacteria	226
	Methyl parathion	High-intensity ultrasound	227
	Dimethoate, fenthion, malathion, methyl parathion, monocrotophos, phorate, and trichlorfon	Degradation by <i>lactobacillus</i> spp. bacteria at 42°C	228
Metals	Pb <sup>2+</sup> and Hg <sup>2+</sup>	Adsorption with pluronic p123 diacrylate hydrogels	229
	Lead	Biosorption with <i>Saccharomyces cerevisiae</i>	230
		Biosorption with <i>Lactobacillus acidophilus</i> ATCC 4356	231
	Mercury	Biosorption with <i>Lactobacillus acidophilus</i> ATCC 4356	232
		Biosorption with <i>Saccharomyces cerevisiae</i>	233
	Copper	Adsorption using imac hp resin	92
	Cadmium	Biosorption with <i>Saccharomyces cerevisiae</i>	234,235
Biosorption with <i>Lactobacillus acidophilus</i> ATCC 4356		231	
Antibiotics	Amoxicillin, doxycycline, ciprofloxacin, and sulfadiazine	Ozonization	236
	Chlortetracycline and cefazolin	Electrochemical method	237
	Tetracycline	Electrochemical method	238
		Adsorption with molecularly imprinted polymer	239

**Table 3.** Continued

	Contaminant	Process	Reference
	Ciprofloxacin	Adsorption with BiPO <sub>4</sub> @ fluorescent photocatalytic graphene oxide-based magnetic molecular imprinted polymer	<sup>240</sup>
	Amoxicillin, ciprofloxacin, doxycycline	Decomposition by gamma irradiation	<sup>241</sup>
Mycotoxins	Aflatoxin M1	Adsorption with molecularly imprinted polymer coated on the surface of the stainless-steel plate	<sup>242</sup>
		Removal using <i>Saccharomyces cerevisiae</i> and <i>Lactobacillus helveticus</i>	<sup>243</sup>
		Adsorption with clay minerals (kaolin and bentonite)	<sup>244</sup>
		Elimination by a combination of yeast and probiotic bacteria species	<sup>245,246</sup>
		Biofilm elimination of <i>Lactobacillus rhamnosus</i> gg	<sup>247</sup>
		Adsorption with clay minerals (kaolin and bentonite)	<sup>248</sup>

UV-C: Ultraviolet-C (200-280 nm); Nd:YAG: neodymium-doped yttrium aluminum garnet; Pb: lead; Hg: mercury; ATCC: American Type Culture Collection; BiPO<sub>4</sub>: Bismuth phosphate (III).

being more effective for chlorpyrifos, fenitrothion, and malathion, whose degradation constants were greater than 0.018 h<sup>-1</sup>. On the other hand, diazinon and methyl parathion were more resistant when applying of the different strains of lactic acid bacteria separately and in combination. The degradation rate constants were correlated with the measurement of phosphatase activity, and it was found that the lower the phosphatase activity, the lower the degradation constant.<sup>226</sup> The same method was applied for this group of contaminants finding that dimethoate and methyl parathion were the most stable with the lowest degradation rate constants (0.0165-0.0184 and 0.0213 h<sup>-1</sup>, being more efficient for the removal of malathion with higher degradation rate constants (0.0218-0.0420 h<sup>-1</sup>).<sup>228</sup> Although the application of lactic acid bacteria was shown to be an effective method for removing diazinon, dimethoate, and methyl parathion in cow's milk it was not very selective since it cannot eliminate all the organophosphates studied.

Biosorption with *Saccharomyces Cerevisiae* allowed the removal of 70% of lead, mercury, and cadmium metals.<sup>230,233-235</sup> The removal percentage was higher when *Lactobacillus Acidophilus* was used, eliminating 80, 75, and 72%, respectively.<sup>231,232</sup> The use of *Saccharomyces cerevisiae* and *Lactobacillus helveticus* removed AFM1 from milk by an as yet unknown binding mechanism.<sup>243</sup> A combination of probiotic bacteria with yeast species managed to remove 90.88% of AFM1 within 72 hours.<sup>245</sup> This percentage of removal was higher than that obtained in another study (19-61%).<sup>246</sup> By applying a biofilm of *Lactobacillus rhamnosus*, an AFM1 removal of 60.74% was achieved. Despite that, the method is not a viable alternative for application because a reduction in the percentage of fat and total dry matter was observed.<sup>247</sup>

Biosorption methods employing microorganisms (*Lactobacillus acidophilus* and *Saccharomyces cerevisiae*) are efficient for removing heavy metals in cow's milk (lead, mercury, copper, and cadmium). However, they require a minimum fermentation period of 4 days. When using lactic acid bacteria to degrade organophosphorus pesticides, a minimum fermentation period of 24 hours is required. These times would represent economic losses for the industry, and given the existing world demand for milk, it would be almost impossible to apply them on a large scale.

Adsorption methods prove to be efficient for removing metals, antibiotics, and mycotoxins. By adsorption with diacrylate Pluronic P123 (P123-DA) hydrogels removed about 85.3% and 81.9% of Pb<sup>2+</sup>, and Hg<sup>2+</sup> ions, respectively.<sup>229</sup> Resins have been another adsorbent used in the adsorption of heavy metals in cow's milk. IMAC HP resin was described for the removal of copper ions (76.89%).<sup>92</sup> Tetracycline, oxytetracycline, chlortetracycline, and doxycycline have been removed by adsorption on a molecularly imprinted polymer, achieving 81.83, 95.47, 96.44, and 93.25% removal, respectively.<sup>239</sup> A photocatalytic-fluorescent polymer, produced from graphene oxide and bismuth phosphate with molecular magnetic imprinting, allowed ciprofloxacin's complete degradation.<sup>240</sup> Bodbodak *et al.*,<sup>242</sup> developed a molecularly imprinted polymer coated on the surface of a stainless-steel plate as an adsorbent material for the decontamination of AFM1 in cow's milk. This method was able to remove 87.3 to 96.2% of AFM1 without causing a change in the physicochemical

properties of the milk. Adsorption with kaolin and natural calcium bentonite clay for adsorption was able to remove AFM1 by 86.1-93.3% and 93.7-97.7%, respectively. It was observed that no change in the nutritional properties of milk would occur.<sup>244</sup> Despite this, few studies have been reported in cow's milk. Therefore, there are not enough to consider its application at the industrial level.

Other methods less reported in the literature were also applied for the removal of pesticides and antibiotics. The ultrasonic treatment proved to be effective for the degradation of 97.10% of methyl parathion. However, this method is limited by the generation of degradation products with toxic effects.<sup>227</sup> For the elimination of antibiotics in cow's milk, methods such as ozonation have been applied, with about 95% degradation for amoxicillin, doxycycline, ciprofloxacin, and sulfadiazine.<sup>236</sup> Electrochemical oxidation applied for the removal of small concentrations of chlortetracycline, cefazolin,<sup>237</sup> and oxytetracycline<sup>238</sup> was also described. Gamma radiation was also found to be effective for the removal of amoxicillin, ciprofloxacin, and doxycycline by 90% in cow's milk samples.<sup>241</sup> However, of all the antibiotics detected in cow's milk, they have only been tested for the elimination of amoxicillin, doxycycline, ciprofloxacin, sulfadiazine, chlortetracycline, cefazolin, y tetracycline. More studies are needed to validate the application of these methods for the decontamination of cow's milk.

It has not been demonstrated that a single method is capable of eliminating different groups of contaminants, as is the case of pasteurization for microbial contaminants. Despite the wide use of hormones in the cattle industry and their consequent generation of traces in cow's milk, no removal methods have been reported for them. The alternative methods studied to date have been applied on an industrial scale, and many of them alter the nutritional properties of milk. The fact that most of these chemical contaminants are not regulated by standards does not oblige the dairy industry to use alternative methods to pasteurization. Nor is it economically viable to use a different method for the elimination of each contaminant present in milk. However, to guarantee the safety of milk, it is essential to study processes that complement pasteurization and can eliminate pathogenic microorganisms and chemical contaminants.

## 5. Conclusions and future prospects

The presence of contaminants in raw cow's milk (many of them banned) is an indication that they are currently used illegally in both agriculture and animal husbandry. Although the presence of contaminant residues in milk represents a health risk to the consumer, there are no MRLs established for all of them. In addition, pasteurization processes are not efficient for the degradation or elimination of the different contaminants addressed.

Although, the literature exposes alternative methods for removing various contaminants in milk, they are still not sufficient nor applied on an industrial scale. Instead, they have been applied individually or in very small families of contaminants. There are no evidence or results concerning the interactions between them or with intermediate products formed on cow's milk, nor changes in the organoleptic properties. A particular case is hormones, which although they are a direct source of contamination, with evidence of their presence in raw, pasteurized, and UHT milk, the literature does not report specific elimination methods for these types of contaminants.

However, alternative methods have proven to be efficient in degrading several contaminants present in milk. Based on this hypothesis, it is suggested to deepen the application of these methods, including the study of interactions between different families of contaminants, application of new materials, or modification of existing ones. Studies on toxicity or changes in organoleptic properties. In this sense, the field of nano-biotechnology, nano-fibers, nano-membranes, biochar, MOF's (metal-organic framework), among others, could play a relevant role, guaranteeing the safety of the milk consumed, and consequently, a better quality of life for consumers.

## Data availability

No data are associated with this article.

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## References

- O'Mahony JA, Fox PF: **Milk: An Overview**. *Milk Proteins*. Singh H, Boland M, Thompson A, editors. San Diego: Academic Press; 2014; p. 19–73.  
[Publisher Full Text](#)
- FAO: *Food Outlook. Trade and Market Division of FAO*. Food and Agriculture Organization of the United Nations; 2010 [cited 2020 Sep, 29].  
[Reference Source](#)
- FAO: *Food Outlook-Biannual Report on Global Food Markets*. 2019 [cited 2020 Sep, 27].  
[Reference Source](#)
- Fox PF: **Milk | Bovine Milk**. *Encyclopedia of Dairy Sciences*. Fuquay JW, editor. San Diego: Academic Press; 2011; p. 478–483.  
[Publisher Full Text](#)
- Jensen RG: **Contaminants in Bovine Milk**. *Handbook of Milk Composition*. Jensen RG, editor. San Diego: Academic Press; 1995; p. 887–901.  
[Publisher Full Text](#)
- Oliveira GB, et al.: **Psychrotrophic bacteria in milk: How much do we really know?**. *Braz. J. Microbiol.* 2015; **46**: 313–321.  
[PubMed Abstract](#) | [Publisher Full Text](#)
- Alegbeleye OO, et al.: **Hazards of a 'healthy' trend? An appraisal of the risks of raw milk consumption and the potential of novel treatment technologies to serve as alternatives to pasteurization**. *Trends Food Sci. Technol.* 2018; **82**: 148–166.  
[Publisher Full Text](#)
- Fischer WJ, et al.: **Contaminants of Milk and Dairy Products: Contamination Resulting from Farm and Dairy Practices**. *Reference Module in Food Science*. Elsevier; 2016.  
[Publisher Full Text](#)
- Ryser ET: **Pasteurization of Liquid Milk Products: Principles, Public Health Aspects**. *Reference Module in Food Science*. Elsevier; 2016.  
[Publisher Full Text](#)
- Tamime AY: *Milk processing and quality management*. John Wiley & Sons; 2009.  
[Publisher Full Text](#)
- Bedi JS, et al.: **Pesticide Residues in Bovine Milk in Punjab, India: Spatial Variation and Risk Assessment to Human Health**. *Arch. Environ. Contam. Toxicol.* 2015; **69**(2): 230–240.  
[PubMed Abstract](#) | [Publisher Full Text](#)
- Kantiani L, et al.: **Fully automated analysis of beta-lactams in bovine milk by online solid phase extraction-liquid chromatography-electrospray-tandem mass spectrometry**. *Anal. Chem.* 2009; **81**(11): 4285–4295.  
[PubMed Abstract](#) | [Publisher Full Text](#)
- Hoogenboom LA, et al.: **Carry-over of pyrrolizidine alkaloids from feed to milk in dairy cows**. *Food Addit. Contam. Part A Chem. Anal. Control Expo. Risk Assess.* 2011; **28**(3): 359–372.  
[PubMed Abstract](#) | [Publisher Full Text](#)
- Schettler T: **Human exposure to phthalates via consumer products**. *Int. J. Androl.* 2006; **29**(1): 134–139.  
[Publisher Full Text](#)
- Mollayusefian I, et al.: **The concentration of aflatoxin M1 in raw and pasteurized milk: A worldwide systematic review and meta-analysis**. *Trends Food Sci. Technol.* 2021; **115**: 22–30.  
[Publisher Full Text](#)
- Yadav J, et al.: **Comparative evaluation of pathogenic bacterial incidence in raw and pasteurized milk**. *Int. J. Eng. Sci. Invention.* 2014; **3**(5): 11–20.
- Weber M, et al.: **Bacterial community composition of biofilms in milking machines of two dairy farms assessed by a combination of culture-dependent and-independent methods**. *PLoS One.* 2019; **14**(9): 1–21.  
[Publisher Full Text](#)
- Franco MM, et al.: **Occurrence of mycobacteria in bovine milk samples from both individual and collective bulk tanks at farms and informal markets in the southeast region of Sao Paulo, Brazil**. *BMC Vet. Res.* 2013; **9**(1): 1–8.
- Zumárraga MJ, et al.: **Detection of Mycobacterium bovis-infected dairy herds using PCR in bulk tank milk samples**. *Foodborne Pathog. Dis.* 2012; **9**(2): 132–137.  
[PubMed Abstract](#) | [Publisher Full Text](#)
- Vithanage NR, et al.: **Biodiversity of culturable psychrotrophic microbiota in raw milk attributable to refrigeration conditions, seasonality and their spoilage potential**. *Int. Dairy J.* 2016; **57**: 80–90.  
[Publisher Full Text](#)
- de Oliveira Pinto C, et al.: **Identificação de bactérias psicrotróficas proteolíticas isoladas de leite cru refrigerado e caracterização do seu potencial deteriorador**. *Revista do Instituto de Laticínios Cândido Tostes.* 2015; **70**(2): 105–116.  
[Publisher Full Text](#)
- Lo R, et al.: **Culture-independent bacterial community profiling of carbon dioxide treated raw milk**. *Int. J. Food Microbiol.* 2016; **233**: 81–89.  
[PubMed Abstract](#) | [Publisher Full Text](#)
- Vithanage NR, et al.: **Comparison of identification systems for psychrotrophic bacteria isolated from raw bovine milk**. *Int. J. Food Microbiol.* 2014; **189**: 26–38.  
[PubMed Abstract](#) | [Publisher Full Text](#)
- Machado SG, et al.: **Pseudomonas spp. and Serratia liquefaciens as Predominant Spoilers in Cold Raw Milk**. *J. Food Sci.* 2015; **80**(8): M1842–M1849.  
[PubMed Abstract](#) | [Publisher Full Text](#)
- Decimo M, et al.: **Characterization of Gram-negative psychrotrophic bacteria isolated from Italian bulk tank milk**. *J. Food Sci.* 2014; **79**(10): M2081–M2090.  
[PubMed Abstract](#) | [Publisher Full Text](#)
- von Neubeck M, et al.: **Biodiversity of refrigerated raw milk microbiota and their enzymatic spoilage potential**. *Int. J. Food Microbiol.* 2015; **211**: 57–65.  
[PubMed Abstract](#) | [Publisher Full Text](#)
- Kable ME, et al.: **The Core and Seasonal Microbiota of Raw Bovine Milk in Tanker Trucks and the Impact of Transfer to a Milk Processing Facility**. *MBio.* 2016; **7**(4).  
[PubMed Abstract](#) | [Publisher Full Text](#)
- Vacheyrou M, et al.: **Cultivable microbial communities in raw cow milk and potential transfers from stables of sixteen French farms**. *Int. J. Food Microbiol.* 2011; **146**(3): 253–262.  
[PubMed Abstract](#) | [Publisher Full Text](#)
- Raats D, et al.: **Molecular analysis of bacterial communities in raw cow milk and the impact of refrigeration on its structure and dynamics**. *Food Microbiol.* 2011; **28**(3): 465–471.  
[PubMed Abstract](#) | [Publisher Full Text](#)
- Marjan S, et al.: **Drug-resistant bacterial pathogens in milk and some milk products**. *Nutr. Food Sci.* 2014; **44**(3): 241–248.  
[Publisher Full Text](#)
- Artursson K, et al.: **Foodborne pathogens in unpasteurized milk in Sweden**. *Int. J. Food Microbiol.* 2018; **284**: 120–127.  
[PubMed Abstract](#) | [Publisher Full Text](#)
- Mallet A, et al.: **Quantitative and qualitative microbial analysis of raw milk reveals substantial diversity influenced by herd management practices**. *Int. Dairy J.* 2012; **27**(1–2): 13–21.  
[Publisher Full Text](#)
- Boubendir A, et al.: **Changes in bacterial populations in refrigerated raw milk collected from a semi-arid area of Algeria**. *Ann. Microbiol.* 2015; **66**(2): 777–783.
- Christidis T, et al.: **Campylobacter spp. Prevalence and Levels in Raw Milk: A Systematic Review and Meta-Analysis**. *J. Food Prot.* 2016; **79**(10): 1775–1783.  
[PubMed Abstract](#) | [Publisher Full Text](#)
- Bianchini V, et al.: **Prevalence in bulk tank milk and epidemiology of Campylobacter jejuni in dairy herds in Northern Italy**. *Appl. Environ. Microbiol.* 2014; **80**(6): 1832–1837.  
[PubMed Abstract](#) | [Publisher Full Text](#)
- Ruusunen M, et al.: **Pathogenic bacteria in Finnish bulk tank milk**. *Foodborne Pathog. Dis.* 2013; **10**(2): 99–106.  
[PubMed Abstract](#) | [Publisher Full Text](#)
- Silva R, et al.: **Pasteurized milk: efficiency of pasteurization and its microbiological conditions in Brazil**. *Foodborne Pathog. Dis.* 2010; **7**(2): 217–219.  
[PubMed Abstract](#) | [Publisher Full Text](#)
- Braem G, et al.: **Culture-independent exploration of the teat apex microbiota of dairy cows reveals a wide bacterial species diversity**. *Vet. Microbiol.* 2012; **157**(3–4): 383–390.  
[PubMed Abstract](#) | [Publisher Full Text](#)
- White DG, et al.: **Isolation and identification of coagulase-negative Staphylococcus species from bovine body sites and streak canals of nulliparous heifers**. *J. Dairy Sci.* 1989; **72**(7): 1886–1892.  
[PubMed Abstract](#) | [Publisher Full Text](#)
- Ercolini D, et al.: **Molecular identification of mesophilic and psychrotrophic bacteria from raw cow's milk**. *Food Microbiol.* 2009; **26**(2): 228–231.  
[PubMed Abstract](#) | [Publisher Full Text](#)

41. Desmaures N, Bazin F, Gueguen M: **Microbiological composition of raw milk from selected farms in the Camembert region of Normandy.** *J. Appl. Microbiol.* 1997; **83**(1): 53–58.  
[PubMed Abstract](#) | [Publisher Full Text](#)
42. Wilson DJ, Gonzalez RN, Das HH: **Bovine mastitis pathogens in New York and Pennsylvania: prevalence and effects on somatic cell count and milk production.** *J. Dairy Sci.* 1997; **80**(10): 2592–2598.  
[PubMed Abstract](#) | [Publisher Full Text](#)
43. Makovec JA, Ruegg PL: **Results of milk samples submitted for microbiological examination in Wisconsin from 1994 to 2001.** *J. Dairy Sci.* 2003; **86**(11): 3466–3472.  
[PubMed Abstract](#) | [Publisher Full Text](#)
44. Jayarao BM, Wang L: **A study on the prevalence of gram-negative bacteria in bulk tank milk.** *J. Dairy Sci.* 1999; **82**(12): 2620–2624.  
[PubMed Abstract](#) | [Publisher Full Text](#)
45. Ternstrom A, Lindberg AM, Molin G: **Classification of the spoilage flora of raw and pasteurized bovine milk, with special reference to *Pseudomonas* and *Bacillus*.** *J. Appl. Bacteriol.* 1993; **75**(1): 25–34.  
[PubMed Abstract](#) | [Publisher Full Text](#)
46. Claeys WL, et al.: **Raw or heated cow milk consumption: Review of risks and benefits.** *Food Control.* 2013; **31**(1): 251–262.  
[Publisher Full Text](#)
47. Aydin S, et al.: **Organohalogenated pollutants in raw and UHT cow's milk from Turkey: a risk assessment of dietary intake.** *Environ. Sci. Pollut. Res. Int.* 2019; **26**(13): 12788–12797.  
[PubMed Abstract](#) | [Publisher Full Text](#)
48. Kaushik CP, Kaushik A, Sharma HR: **Seasonal trends in organochlorine pesticide residues in raw bovine milk from rural areas of Haryana, India.** *Bull. Environ. Contam. Toxicol.* 2014; **92**(1): 15–22.  
[PubMed Abstract](#) | [Publisher Full Text](#)
49. Gutiérrez R, et al.: **Organochlorine pesticide residues in bovine milk from organic farms in Chiapas, Mexico.** *Bull. Environ. Contam. Toxicol.* 2012; **89**(4): 882–887.  
[PubMed Abstract](#) | [Publisher Full Text](#)
50. Kaushik CP, et al.: **Changing patterns of organochlorine pesticide residues in raw bovine milk from Haryana, India.** *Environ. Monit. Assess.* 2011; **182**(1-4): 467–475.  
[PubMed Abstract](#) | [Publisher Full Text](#)
51. Bošnjir J, et al.: **Organochlorine pesticide residues in cows' milk from Karlovac County, Croatia.** *Acta Aliment.* 2010; **39**(3): 317–326.  
[Publisher Full Text](#)
52. Avancini RM, et al.: **Organochlorine compounds in bovine milk from the state of Mato Grosso do Sul-Brazil.** *Chemosphere.* 2013; **90**(9): 2408–2413.  
[PubMed Abstract](#) | [Publisher Full Text](#)
53. Lans-Ceballos E, Lombana-Gomez M, Pinedo-Hernandez J: **Organochlorine insecticide residues in pasteurized milk distributed in Montería Colombia.** *Rev. Salud Pública. (Bogotá).* 2018; **20**(2): 208–214.  
[PubMed Abstract](#) | [Publisher Full Text](#)
54. Gill JPS, et al.: **Pesticide Residues in Peri-Urban Bovine Milk from India and Risk Assessment: A Multicenter Study.** *Sci. Rep.* 2020; **10**(1): 8054.  
[PubMed Abstract](#) | [Publisher Full Text](#)
55. Shahzadi N, et al.: **Identification of pesticides residues in different samples of milk.** *J. Agroaliment. Process. Technol.* 2013; **19**(2): 167–172.
56. Sajid MW, et al.: **The impact of seasonal variation on organochlorine pesticide residues in buffalo and cow milk of selected dairy farms from Faisalabad region.** *Environ. Monit. Assess.* 2016; **188**(10): 589.  
[PubMed Abstract](#) | [Publisher Full Text](#)
57. ul Hassan A, et al.: **Organochlorine and pyrethroid pesticides analysis in dairy milk samples collected from cotton growing belt of Punjab, Pakistan.** *Pak. J. Agric. Sci.* 2014; **51**(2): 331–335.
58. Welsh JA, et al.: **Production-related contaminants (pesticides, antibiotics and hormones) in organic and conventionally produced milk samples sold in the USA.** *Public Health Nutr.* 2019; **22**(16): 2972–2980.  
[PubMed Abstract](#) | [Publisher Full Text](#)
59. Chen X, et al.: **Method for the quantification of current use and persistent pesticides in cow milk, human milk and baby formula using gas chromatography tandem mass spectrometry.** *J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.* 2014; **970**: 121–130.  
[PubMed Abstract](#) | [Publisher Full Text](#)
60. Jayasinghe J, et al.: **Pesticide residues in cow milk and dairy products from the major milk producing area of Sri Lanka.** *AGROFOR Int. J.* 2019; **4**: 83–90.  
[Publisher Full Text](#)
61. Nath A, et al.: **Carcinogenic pesticides residue detection in cow milk and water samples from Patna, India.** *Current Trends in Biotechnology and Chemical Research.* 2013; **3**(1): 1–7.
62. Fagnani R, et al.: **Organophosphorus and carbamate residues in milk and feedstuff supplied to dairy cattle.** *Pesqui. Vet. Bras.* 2011; **31**(7): 598–602.  
[Publisher Full Text](#)
63. Salas JH, et al.: **Organophosphorus pesticide residues in Mexican commercial pasteurized milk.** *J. Agric. Food Chem.* 2003; **51**(15): 4468–4471.  
[PubMed Abstract](#) | [Publisher Full Text](#)
64. Kampire E, et al.: **Organochlorine pesticide in fresh and pasteurized cow's milk from Kampala markets.** *Chemosphere.* 2011; **84**(7): 923–927.  
[PubMed Abstract](#) | [Publisher Full Text](#)
65. Tian H: **Determination of chloramphenicol, enrofloxacin and 29 pesticides residues in bovine milk by liquid chromatography-tandem mass spectrometry.** *Chemosphere.* 2011; **83**(3): 349–355.  
[PubMed Abstract](#) | [Publisher Full Text](#)
66. Kuba J, et al.: **Comparison of DDT and its metabolites concentrations in cow milk from agricultural and industrial areas.** *J. Environ. Sci. Health B.* 2015; **50**(1): 1–7.  
[PubMed Abstract](#) | [Publisher Full Text](#)
67. Castilla Y, Mercado I, González G: **Determinación y cuantificación de los niveles de compuestos organoclorados en leche pasteurizada.** *Producción + Limpia.* 2012; **7**(1): 19–31.
68. Hernández M, Vidal JV, Marrugo JL: **Organochlorine pesticides in cows' milk supplemented with cotton waste in San Pedro, Colombia.** *Revista de salud pública (Bogotá, Colombia).* 2010; **12**(6): 982–989.
69. Bulut S, et al.: **Organochlorine pesticide (OCP) residues in cow's, buffalo's, and sheep's milk from Afyonkarahisar region, Turkey.** *Environ. Monit. Assess.* 2011; **181**(1-4): 555–562.  
[PubMed Abstract](#) | [Publisher Full Text](#)
70. Luzardo OP, et al.: **Polychlorobiphenyls and organochlorine pesticides in conventional and organic brands of milk: occurrence and dietary intake in the population of the Canary Islands (Spain).** *Chemosphere.* 2012; **88**(3): 307–315.  
[PubMed Abstract](#) | [Publisher Full Text](#)
71. Karabasanavar NS, Singh SP: **Occurrence of endosulphan residues in dairy milk in plains of Uttarakhand, India.** *Int. J. Dairy Technol.* 2013; **66**(2): 189–193.  
[Publisher Full Text](#)
72. Kim DG, et al.: **Monitoring of environmental contaminants in raw bovine milk and estimates of dietary intakes of children in South Korea.** *Chemosphere.* 2013; **93**(3): 561–566.  
[PubMed Abstract](#) | [Publisher Full Text](#)
73. Özdemir C, et al.: **Determination of organochlorine pesticide residues in pasteurized and sterilized milk using QuEChERS sample preparation followed by gas chromatography-mass spectrometry.** *J. Food Process. Preserv.* 2019; **43**(11): e14173.  
[Publisher Full Text](#)
74. Bommuraj V, et al.: **Human pharmaceutical and pesticide residues in Israeli dairy milk in association with dietary risk assessment.** *Food Addit. Contam. Part B Surveill.* 2020; **13**(4): 233–243.  
[PubMed Abstract](#) | [Publisher Full Text](#)
75. Rejczak T, Tuzimski T: **QuEChERS-based extraction with dispersive solid phase extraction clean-up using PSA and ZrO<sub>2</sub>-based sorbents for determination of pesticides in bovine milk samples by HPLC-DAD.** *Food Chem.* 2017; **217**: 225–233.  
[PubMed Abstract](#) | [Publisher Full Text](#)
76. Melgar MJ, Santaefemia M, Garcia MA: **Organophosphorus pesticide residues in raw milk and infant formulas from Spanish northwest.** *J. Environ. Sci. Health B.* 2010; **45**(7): 595–600.  
[PubMed Abstract](#) | [Publisher Full Text](#)
77. Ștefănescu L, Stezar CI, Groza IȘ: **The influence of environmental contamination on heavy metals and organochlorine compounds levels in milk.** *Environ. Eng. Manag. J.* 2011; **10**(1): 37–42.
78. Norouzirad R, et al.: **Lead and cadmium levels in raw bovine milk and dietary risk assessment in areas near petroleum extraction industries.** *Sci. Total Environ.* 2018; **635**: 308–314.  
[PubMed Abstract](#) | [Publisher Full Text](#)
79. Ismail A, et al.: **Estimated daily intake and health risk of heavy metals by consumption of milk.** *Food Addit. Contam. Part B Surveill.* 2015; **8**(4): 260–265.  
[PubMed Abstract](#) | [Publisher Full Text](#)
80. Meshref AMS, Moselhy WA, Hassan NE-HY: **Heavy metals and trace elements levels in milk and milk products.** *J. Food Meas. Charact.* 2014; **8**(4): 381–388.  
[Publisher Full Text](#)

81. Maas S, *et al.*: Trace metals in raw cows' milk and assessment of transfer to Comté cheese. *Food Chem.* 2011; **129**(1): 7–12. [Publisher Full Text](#)
82. Pilarczyk R, *et al.*: Concentrations of toxic heavy metals and trace elements in raw milk of Simmental and Holstein-Friesian cows from organic farm. *Environ. Monit. Assess.* 2013; **185**(10): 8383–8392. [PubMed Abstract](#) | [Publisher Full Text](#)
83. Singh M, *et al.*: Assessment of contamination of milk and milk products with heavy metals. *Int. J. Dairy Sci.* 2020; **72**(6): 608–615. [Publisher Full Text](#)
84. Bilandžić N, *et al.*: Trace element levels in raw milk from northern and southern regions of Croatia. *Food Chem.* 2011; **127**(1): 63–66. [Publisher Full Text](#)
85. Chen L, *et al.*: Analysis of 17 elements in cow, goat, buffalo, yak, and camel milk by inductively coupled plasma mass spectrometry (ICP-MS). *RSC Adv.* 2020; **10**(12): 6736–6742. [Publisher Full Text](#)
86. Gabryszuk M, *et al.*: Content of mineral elements in milk and hair of cows from organic farms. *J. Elem.* 2010; **15**(2): 259–267.
87. Suturovic Z, *et al.*: Determination of heavy metals in milk and fermented milk products by potentiometric stripping analysis with constant inverse current in the analytical step. *Food Chem.* 2014; **155**: 120–125. [PubMed Abstract](#) | [Publisher Full Text](#)
88. Choudhury T, *et al.*: Evaluation of elemental, microbial and biochemical status of raw and pasteurized cow's milk. *Int. Food Res. J.* 2018; **25**(4): 1682–1690.
89. Akhtar S, *et al.*: Minerals and heavy metals in raw and ultra heat treated commercial milks in Pakistan. *International Journal of Food and Allied Sciences.* 2015; **1**: 18. [Publisher Full Text](#)
90. Santos C, *et al.*: Determination of the concentrations of essential and toxic metals in UHT milk produced in Mato Grosso State, Brazil. *Int. Food Res. J.* 2015; **22**(3): 981–986.
91. Qu XY, *et al.*: Analysis and Risk Assessment of Seven Toxic Element Residues in Raw Bovine Milk in China. *Biol. Trace Elem. Res.* 2018; **183**(1): 92–101. [PubMed Abstract](#) | [Publisher Full Text](#)
92. Abdelfatah E, *et al.*: Heavy metal residues and health risk assessment in raw milk and dairy products with a trail for removal of copper residues. *Benha Veterinary Medical Journal.* 2019; **36**: 51–64. [Publisher Full Text](#)
93. Soares VA, *et al.*: Determination of nutritional and toxic elements in pasteurized bovine milk from Vale do Paraiba region (Brazil). *Food Control.* 2010; **21**(1): 45–49. [Publisher Full Text](#)
94. Ribeiro Sant'Ana MA, de Carvalho TC, da Silva IF: Concentration of heavy metals in UHT dairy milk available in the markets of Sao Luis, Brazil, and potential health risk to children. *Food Chem.* 2021; **346**: 128961. [PubMed Abstract](#) | [Publisher Full Text](#)
95. do Nascimento IR, *et al.*: Determination of the mineral composition of fresh bovine milk from the milk-producing areas located in the State of Sergipe in Brazil and evaluation employing exploratory analysis. *Microchem. J.* 2010; **96**(1): 37–41. [Publisher Full Text](#)
96. Totan FE, Filazi A: Determination of some element levels in various kinds of cow's milk processed in different ways. *Environ. Monit. Assess.* 2020; **192**(2): 1–10.
97. Arianejad M, *et al.*: Levels of Some Heavy Metals in Raw Cow's Milk from Selected Milk Production Sites in Iran: Is There any Health Concern?. *Health Promot. Perspect.* 2015; **5**(3): 176–182. [PubMed Abstract](#) | [Publisher Full Text](#)
98. Gonzalez-Montana JR, *et al.*: Some toxic metals (Al, As, Mo, Hg) from cow's milk raised in a possibly contaminated area by different sources. *Environ. Sci. Pollut. Res. Int.* 2019; **26**(28): 28909–28918. [PubMed Abstract](#) | [Publisher Full Text](#)
99. Najarnezhad V, Akbarabadi M: Heavy metals in raw cow and ewe milk from north-east Iran. *Food Addit. Contam. Part B Surveill.* 2013; **6**(3): 158–162. [PubMed Abstract](#) | [Publisher Full Text](#)
100. Chiesa LM, *et al.*: Analysis of antibiotic residues in raw bovine milk and their impact toward food safety and on milk starter cultures in cheese-making process. *LWT.* 2020; **131**: 109783. [Publisher Full Text](#)
101. Moretti S, *et al.*: Multiclass method for the determination of 62 antibiotics in milk. *J. Mass Spectrom.* 2016; **51**(9): 792–804. [PubMed Abstract](#) | [Publisher Full Text](#)
102. Mesgari Abbasi M, *et al.*: Simultaneous Determination of Tetracyclines Residues in Bovine Milk Samples by Solid Phase Extraction and HPLC-FL Method. *Adv. Pharm. Bull.* 2011; **1**(1): 34–39. [PubMed Abstract](#) | [Publisher Full Text](#)
103. Alija G, Hajrulai-Musliu Z, Uzunov R: Development and validation of confirmatory LC-MS/MS method for multi-residue analysis of antibiotic drugs in bovine milk. *SN Appl. Sci.* 2020; **2**(9). [Publisher Full Text](#)
104. Dimitrieska-Stojkovic E, *et al.*: Screening of veterinary drug residues in milk from individual farms in Macedonia. *Maced. Vet. Rev.* 2011; **34**(1): 5–13.
105. Moudgil P, *et al.*: Analysis of antibiotic residues in raw and commercial milk in Punjab, India vis-à-vis human health risk assessment. *J. Food Saf.* 2019; **39**(4): 1–8.
106. Florez DHA, Dutra FVA, Borges KB: Magnetic solid phase extraction employing a novel restricted access material based on mesoporous polyaniline coated with hydrophilic monomers and casein for determination of antibiotics in milk samples. *Microchem. J.* 2019; **150**: 104145. [Publisher Full Text](#)
107. Spisso BF, *et al.*: Pilot survey of commercial pasteurized milk consumed in the metropolitan area of Rio de Janeiro, Brazil, for tetracyclines residues, including the 4-epimers of oxytetracycline, tetracycline and chlortetracycline. *Food Addit. Contam. Part B Surveill.* 2010; **3**(4): 220–227. [Publisher Full Text](#)
108. Novaes SFD, *et al.*: Residues of veterinary drugs in milk in Brazil. *Ciência Rural.* 2017; **47**(8): p. 1–7. [Publisher Full Text](#)
109. Zhang WQ, *et al.*: Analysis of Veterinary Drug Residues in Pasteurized Milk Samples in Chinese Milk Bars. *J. Food Prot.* 2020; **83**: 204–210. [PubMed Abstract](#) | [Publisher Full Text](#)
110. Du B, *et al.*: Evaluation of an ELISA-based visualization microarray chip technique for the detection of veterinary antibiotics in milk. *Food Control.* 2019; **106**: 106713. [Publisher Full Text](#)
111. Du B, *et al.*: Presence of tetracyclines, quinolones, lincomycin and streptomycin in milk. *Food Control.* 2019; **100**: 171–175. [Publisher Full Text](#)
112. Juan C, *et al.*: Determination of macrolide and lincosamide antibiotics by pressurised liquid extraction and liquid chromatography-tandem mass spectrometry in meat and milk. *Food Control.* 2010; **21**(12): 1703–1709. [Publisher Full Text](#)
113. Mohammed HA, *et al.*: Rapid tests for detection of enrofloxacin residues in liquid milk. *Benha Veterinary Medical Journal.* 2016; **30**(1): 97–103. [Publisher Full Text](#)
114. Zhang YD, *et al.*: Occurrence of tetracyclines, sulfonamides, sulfamethazine and quinolones in pasteurized milk and UHT milk in China's market. *Food Control.* 2014; **36**(1): 238–242. [Publisher Full Text](#)
115. Kumarswamy N, *et al.*: Detection of antibiotic residues in raw cow milk in Thrissur, India. *Pharm. Innov.* 2018; **7**(8): 452–454.
116. Bilandžić N, *et al.*: Veterinary drug residues determination in raw milk in Croatia. *Food Control.* 2011; **22**(12): 1941–1948. [Publisher Full Text](#)
117. Urapen R, Masawat P: Novel method for the determination of tetracycline antibiotics in bovine milk based on digital-image-based colorimetry. *Int. Dairy J.* 2015; **44**: 1–5. [Publisher Full Text](#)
118. Jank L, *et al.*: beta-lactam antibiotics residues analysis in bovine milk by LC-ESI-MS/MS: a simple and fast liquid-liquid extraction method. *Food Addit. Contam. Part A Chem. Anal. Control Expo. Risk Assess.* 2012; **29**(4): 497–507. [PubMed Abstract](#) | [Publisher Full Text](#)
119. Dorival-García N, *et al.*: Simultaneous determination of quinolone and  $\beta$ -lactam residues in raw cow milk samples using ultrasound-assisted extraction and dispersive-SPE prior to UHPLC-MS/MS analysis. *Food Control.* 2016; **60**: 382–393. [Publisher Full Text](#)
120. Junza A, *et al.*: Comparative study of the LC-MS/MS and UPLC-MS/MS for the multi-residue analysis of quinolones, penicillins and cephalosporins in cow milk, and validation according to the regulation 2002/657/EC. *J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.* 2011; **879**(25): 2601–2610. [Publisher Full Text](#)
121. Rama A, *et al.*: Assessment of antibacterial drug residues in milk for consumption in Kosovo. *J. Food Drug Anal.* 2017; **25**(3):

- 525–532.  
[PubMed Abstract](#) | [Publisher Full Text](#)
122. Junza A, *et al.*: **Multiclass method for the determination of quinolones and  $\beta$ -lactams, in raw cow milk using dispersive liquid-liquid microextraction and ultra high performance liquid chromatography-tandem mass spectrometry.** *J. Chromatogr. A.* 2014; **1356**: 10–22.  
[PubMed Abstract](#) | [Publisher Full Text](#)
123. Dinali LAF, *et al.*: **Efficient development of a magnetic molecularly imprinted polymer for selective determination of trimethoprim and sulfamethoxazole in milk.** *Microchem. J.* 2020; **154**: 104648.  
[Publisher Full Text](#)
124. Li H, *et al.*: **Simultaneous determination of amoxicillin and prednisolone in bovine milk using ultra-high performance liquid chromatography tandem mass spectrometry.** *J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.* 2012; **900**: 59–63.  
[PubMed Abstract](#) | [Publisher Full Text](#)
125. Liu X, *et al.*: **Solid phase extraction using magnetic core mesoporous shell microspheres with C18-modified interior pore-walls for residue analysis of cephalosporins in milk by LC-MS/MS.** *Food Chem.* 2014; **150**: 206–212.  
[PubMed Abstract](#) | [Publisher Full Text](#)
126. Kantiani L, Farre M, Barcelo D: **Rapid residue analysis of fluoroquinolones in raw bovine milk by online solid phase extraction followed by liquid chromatography coupled to tandem mass spectrometry.** *J. Chromatogr. A.* 2011; **1218**(50): 9019–9027.  
[PubMed Abstract](#) | [Publisher Full Text](#)
127. Meng Z, *et al.*: **Residues investigation of fluoroquinolones and sulphonamides and their metabolites in bovine milk by quantification and confirmation using ultra-performance liquid chromatography-tandem mass spectrometry.** *Food Chem.* 2015; **174**: 597–605.  
[PubMed Abstract](#) | [Publisher Full Text](#)
128. Bilandžić N, *et al.*: **Concentrations of veterinary drug residues in milk from individual farms in Croatia.** *Mljekarstvo.* 2011; **61**(3): 260–267.
129. Navratilova P, *et al.*: **Fluoroquinolone residues in raw cow's milk.** *Czech J. Food Sci.* 2011; **29**(6): 641–646.  
[Publisher Full Text](#)
130. Zhou W, *et al.*: **Simultaneous determination of 16 macrolide antibiotics and 4 metabolites in milk by using Quick, Easy, Cheap, Effective, Rugged, and Safe extraction (QuEChERS) and high performance liquid chromatography tandem mass spectrometry.** *J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.* 2017; **1061-1062**: 411–420.  
[PubMed Abstract](#) | [Publisher Full Text](#)
131. Zeina K, Fawwak S, Abi P: **Quantification of antibiotic residues and determination of antimicrobial resistance profiles of microorganisms isolated from bovine milk in Lebanon.** *Food Nutr. Sci.* 2013; **04**(7): 1–9.  
[Publisher Full Text](#)
132. Hou XL, *et al.*: **Development and validation of an ultra high performance liquid chromatography tandem mass spectrometry method for simultaneous determination of sulfonamides, quinolones and benzimidazoles in bovine milk.** *J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.* 2014; **962**: 20–29.  
[PubMed Abstract](#) | [Publisher Full Text](#)
133. Wang Y, *et al.*: **Rapid Determination of Trace Sulfonamides in Milk by Graphene Oxide-Based Magnetic Solid Phase Extraction Coupled with HPLC-MS/MS.** *Food Anal. Methods.* 2016; **9**(9): 2521–2530.  
[Publisher Full Text](#)
134. Jank L, *et al.*: **High-throughput method for the determination of residues of beta-lactam antibiotics in bovine milk by LC-MS/MS.** *Food Addit. Contam. Part A Chem. Anal. Control Expo. Risk Assess.* 2015; **32**(12): 1992–2001.
135. Huang LC, *et al.*: **Simultaneous determination of aflatoxin M1, ochratoxin A, zearalenone and alpha-zearalenol in milk by UHPLC-MS/MS.** *Food Chem.* 2014; **146**: 242–249.  
[PubMed Abstract](#) | [Publisher Full Text](#)
136. Wang X, Li P: **Rapid screening of mycotoxins in liquid milk and milk powder by automated size-exclusion SPE-UPLC-MS/MS and quantification of matrix effects over the whole chromatographic run.** *Food Chem.* 2015; **173**: 897–904.  
[PubMed Abstract](#) | [Publisher Full Text](#)
137. Benkerroum N: **Mycotoxins in dairy products: A review.** *Int. Dairy J.* 2016; **62**: 63–75.  
[Publisher Full Text](#)
138. Mao J, *et al.*: **Multi-mycotoxins analysis in raw milk by ultra high performance liquid chromatography coupled to quadrupole orbitrap mass spectrometry.** *Food Control.* 2018; **84**: 305–311.  
[Publisher Full Text](#)
139. Puga-Torres B, *et al.*: **Determination of Aflatoxin M1 in Raw Milk from Different Provinces of Ecuador.** *Toxins (Basel).* 2020; **12**(8): 498.  
[PubMed Abstract](#) | [Publisher Full Text](#)
140. Keller L, *et al.*: **Incidence of Mycotoxins (AFB1 and AFM1) in Feeds and Dairy Farms from Rio de Janeiro State, Brazil.** *Vet. Med.* 2016; **1**: 29–35.
141. Kos J, *et al.*: **Occurrence and estimation of aflatoxin M1 exposure in milk in Serbia.** *Food Control.* 2014; **38**: 41–46.  
[Publisher Full Text](#)
142. Tomašević I, *et al.*: **Two year survey on the occurrence and seasonal variation of aflatoxin M1 in milk and milk products in Serbia.** *Food Control.* 2015; **56**: 64–70.  
[Publisher Full Text](#)
143. Ruangwises N, Ruangwises S: **Aflatoxin M(1) contamination in raw milk within the central region of Thailand.** *Bull. Environ. Contam. Toxicol.* 2010; **85**(2): 195–198.  
[PubMed Abstract](#) | [Publisher Full Text](#)
144. Zhao Z, *et al.*: **Multi-mycotoxin analysis of animal feed and animal-derived food using LC-MS/MS system with timed and highly selective reaction monitoring.** *Anal. Bioanal. Chem.* 2015; **407**(24): 7359–7368.  
[PubMed Abstract](#) | [Publisher Full Text](#)
145. Duarte SC, *et al.*: **Aflatoxin M1 in marketed milk in Portugal: Assessment of human and animal exposure.** *Food Control.* 2013; **30**(2): 411–417.  
[Publisher Full Text](#)
146. Fallah AA: **Assessment of aflatoxin M1 contamination in pasteurized and UHT milk marketed in central part of Iran.** *Food Chem. Toxicol.* 2010; **48**(3): 988–991.  
[PubMed Abstract](#) | [Publisher Full Text](#)
147. Cano-Sancho G, *et al.*: **Occurrence of aflatoxin M(1) and exposure assessment in Catalonia (Spain).** *Rev. Iberoam. Micol.* 2010; **27**(3): 130–135.  
[PubMed Abstract](#) | [Publisher Full Text](#)
148. Silva MV, *et al.*: **Occurrence and estimative of aflatoxin M1 intake in UHT cow milk in Paraná State, Brazil.** *Food Control.* 2015; **53**: 222–225.  
[Publisher Full Text](#)
149. Pattono D, Gallo PF, Civera T: **Detection and quantification of ochratoxin A in milk produced in organic farms.** *Food Chem.* 2011; **127**(1): 374–377.  
[Publisher Full Text](#)
150. Winkler J, *et al.*: **Development of a multi-toxin method for investigating the carryover of zearalenone, deoxynivalenol and their metabolites into milk of dairy cows.** *Food additives & contaminants. Part A.* 2015; **32**(3): 371–380.
151. Pleadin J, *et al.*: **Presence of Fusarium mycotoxins in feedstuffs and cow milk sampled from Croatian farms during 2015.** *Mljekarstvo/Dairy.* 2017; **67**(2): 102–111.  
[Publisher Full Text](#)
152. Signorini ML, *et al.*: **Exposure assessment of mycotoxins in cow's milk in Argentina.** *Food Chem. Toxicol.* 2012; **50**(2): 250–257.  
[PubMed Abstract](#) | [Publisher Full Text](#)
153. Mahmoudi R: **Occurrence of Zearalenone in raw animal origin food produced in North-West of Iran.** *J. Food Qual. Hazards Control.* 2014; **1**(1): 25–28.
154. Jiang K, *et al.*: **Reduced graphene oxide and gold nanoparticle composite-based solid-phase extraction coupled with ultra-high-performance liquid chromatography-tandem mass spectrometry for the determination of 9 mycotoxins in milk.** *Food Chem.* 2018; **264**: 218–225.  
[PubMed Abstract](#) | [Publisher Full Text](#)
155. Xia X, *et al.*: **Ultra-high-pressure liquid chromatography-tandem mass spectrometry for the analysis of six resorcylic acid lactones in bovine milk.** *J. Chromatogr. A.* 2009; **1216**(12): 2587–2591.  
[PubMed Abstract](#) | [Publisher Full Text](#)
156. Baumrucker CR, Macrina AL: **Hormones and Regulatory Factors in Bovine Milk.** *Reference Module in Food Science.* Elsevier; 2020.
157. Ollikainen P, Muuronen K: **Determination of insulin-like growth factor-1 and bovine insulin in raw milk and its casein and whey fractions after microfiltration and ultrafiltration.** *Int. Dairy J.* 2013; **28**(2): 83–87.  
[Publisher Full Text](#)
158. Mishra M, Ali S, Das M: **A New Extraction Method for the Determination of Oxytocin in Milk by Enzyme Immune Assay or High-Performance Liquid Chromatography: Validation by**

- Liquid Chromatography–Mass Spectrometry.** *Food Anal. Methods.* 2012; **6**(5): 1308–1319.
159. Mishra M, Ali S, Das M: **Analysis of oxytocin in milk samples and intake pattern in different age groups of Indian population.** *Toxicol. Mech. Methods.* 2014; **24**(5): 342–346.  
[PubMed Abstract](#) | [Publisher Full Text](#)
160. Singh S, et al.: **Circulating and milk adiponectin change differently during energy deficiency at different stages of lactation in dairy cows.** *J. Dairy Sci.* 2014; **97**(3): 1535–1542.  
[PubMed Abstract](#) | [Publisher Full Text](#)
161. Goyon A, et al.: **Determination of steroid hormones in bovine milk by LC-MS/MS and their levels in Swiss Holstein cow milk.** *Food Addit. Contam. Part A Chem. Anal. Control Expo. Risk Assess.* 2016; **33**(5): 804–816.  
[PubMed Abstract](#) | [Publisher Full Text](#)
162. Chen C, et al.: **A preliminary risk assessment of potential exposure to naturally occurring estrogens from Beijing (China) market milk products.** *Food Chem. Toxicol.* 2014; **71**: 74–80.  
[PubMed Abstract](#) | [Publisher Full Text](#)
163. Azzouz A, et al.: **Simultaneous determination of 20 pharmacologically active substances in cow's milk, goat's milk, and human breast milk by gas chromatography-mass spectrometry.** *J. Agric. Food Chem.* 2011; **59**(9): 5125–5132.  
[PubMed Abstract](#) | [Publisher Full Text](#)
164. Socas-Rodríguez B, et al.: **Multiclass analytical method for the determination of natural/synthetic steroid hormones, phytoestrogens, and mycoestrogens in milk and yogurt.** *Anal. Bioanal. Chem.* 2017; **409**(18): 4467–4477.  
[PubMed Abstract](#) | [Publisher Full Text](#)
165. Kaklamano G, Theodoridis G: **Rapid multi-method for the determination of growth promoters in bovine milk by liquid chromatography-tandem mass spectrometry.** *J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.* 2013; **930**: 22–29.  
[PubMed Abstract](#) | [Publisher Full Text](#)
166. Florez DHÁ, de Oliveira HL, Borges KB: **Polythiophene as highly efficient sorbent for microextraction in packed sorbent for determination of steroids from bovine milk samples.** *Microchem. J.* 2020; **153**: 104521.  
[Publisher Full Text](#)
167. Regal P, Cepeda A, Fente C: **Development of an LC-MS/MS method to quantify sex hormones in bovine milk and influence of pregnancy in their levels.** *Food Addit. Contam. Part A Chem. Anal. Control Expo. Risk Assess.* 2012; **29**(5): 770–779.  
[PubMed Abstract](#) | [Publisher Full Text](#)
168. Trapiella-Alfonso L, et al.: **Development of a quantum dot-based fluorescent immunoassay for progesterone determination in bovine milk.** *Biosens. Bioelectron.* 2011; **26**(12): 4753–4759.  
[PubMed Abstract](#) | [Publisher Full Text](#)
169. Tan X-t, et al.: **Analysis of 13 kinds of steroid hormones in raw milk using modified QuEChERS method combined with UPLC-QTOF-MS.** *J. Integr. Agric.* 2016; **15**(9): 2163–2174.  
[Publisher Full Text](#)
170. Gellrich K, et al.: **Cortisol levels in skimmed milk during the first 22 weeks of lactation and response to short-term metabolic stress and lameness in dairy cows.** *J. Anim. Sci. Biotechnol.* 2015; **6**(1): 31.  
[PubMed Abstract](#) | [Publisher Full Text](#)
171. Ma L, et al.: **Multiresidue analysis of glucocorticoids in milk by LC-MS/MS with low-temperature purification and dispersive solid-phase extraction.** *J. Sep. Sci.* 2017; **40**(13): 2759–2768.  
[PubMed Abstract](#) | [Publisher Full Text](#)
172. Nag SK: **Contaminants in milk: routes of contamination, analytical techniques and methods of control.** *Improving the Safety and Quality of Milk.* Griffiths MW, editor. Woodhead Publishing; 2010; p. 146–178.  
[Publisher Full Text](#)
173. Ahmadi F, et al.: **Determination of organophosphorus pesticides in water samples by single drop microextraction and gas chromatography-flame photometric detector.** *J. Chromatogr. A.* 2006; **1101**(1–2): 307–312.  
[PubMed Abstract](#) | [Publisher Full Text](#)
174. Pathirana K, et al.: **Pesticide contaminated crop residues and water usage for dairy cattle rearing in Walapane DS division, Sri Lanka.** *Int. J. Innov. Res. Comput. Sci. Technol.* 2015; **2**(6).
175. Bedi J, et al.: **Pesticide residues in milk and their relationship with pesticide contamination of feedstuffs supplied to dairy cattle in Punjab (India).** *J. Anim. Feed Sci.* 2018; **27**(1): 18–25.  
[Publisher Full Text](#)
176. Ashoub M, Azam A: **Seasonal variations on the levels of some pesticide residues in dairy farms.** *Benha Veterinary Medical Journal.* 2016; **30**(1): 312–322.  
[Publisher Full Text](#)
177. Srivastava S, Narvi S, Prasad S: **Organochlorines and organophosphates in bovine milk samples in Allahabad region.** *Int. J. Environ. Res.* 2008; **2**(2): 165–168.
178. Waliszewski SM, et al.: **Detection of some organochlorine pesticides in cow's milk.** *Food Addit. Contam.* 1996; **13**(2): 231–235.  
[Publisher Full Text](#)
179. Losada A, et al.: **Organochlorine pesticide residues in bovine milk from León (Spain).** *Sci. Total Environ.* 1996; **181**(2): 133–135.  
[Publisher Full Text](#)
180. John PJ, Bakore N, Bhatnagar P: **Assessment of organochlorine pesticide residue levels in dairy milk and buffalo milk from Jaipur City, Rajasthan, India.** *Environ. Int.* 2001; **26**(4): 231–236.  
[PubMed Abstract](#) | [Publisher Full Text](#)
181. Claborn HV, et al.: **Pesticide Residues, Malathion in Milk and Fat from Sprayed Cattle.** *J. Agric. Food Chem.* 1956; **4**(11): 941–942.  
[Publisher Full Text](#)
182. Bushland RC, et al.: **Contamination of meat and milk by chlorinated hydrocarbon insecticides used for livestock pest control.** *J. Econ. Entomol.* 1950; **43**(5): 649–652.  
[Publisher Full Text](#)
183. Rodrigues FM, et al.: **Development of a headspace solid-phase microextraction/gas chromatography-mass spectrometry method for determination of organophosphorus pesticide residues in cow milk.** *Microchem. J.* 2011; **98**(1): 56–61.  
[Publisher Full Text](#)
184. Willett LB, et al.: **Mechanisms of movement of organochlorine pesticides from soils to cows via forages.** *J. Dairy Sci.* 1993; **76**(6): 1635–1644.  
[PubMed Abstract](#) | [Publisher Full Text](#)
185. El Bahgy HE, Elbarbary HA, Ibrahim SS: **Open Access Estimation of deltamethrin residues in cow's and goat's environment and trials to reduce its level in milk.** *Veterinary world.* 2018; **11**(5): 606–611.  
[PubMed Abstract](#) | [Publisher Full Text](#)
186. Heck MC, et al.: **Estimation of children exposure to organochlorine compounds through milk in Rio Grande do Sul, Brazil.** *Food Chem.* 2007; **102**(1): 288–294.  
[Publisher Full Text](#)
187. Sabarwal A, Kumar K, Singh RP: **Hazardous effects of chemical pesticides on human health-Cancer and other associated disorders.** *Environ. Toxicol. Pharmacol.* 2018; **63**: 103–114.  
[PubMed Abstract](#) | [Publisher Full Text](#)
188. Karasali H, Maragou N: **Pesticides and Herbicides: Types of Pesticide.** *Encyclopedia of Food and Health.* 2016; 319–325.  
[Publisher Full Text](#)
189. Fischer WJ, et al.: **CONTAMINANTS OF MILK AND DAIRY PRODUCTS|Environmental Contaminants.** *Encyclopedia of Dairy Sciences.* Roginski H, editor. Oxford: Elsevier; 2002; p. 525–533.  
[Publisher Full Text](#)
190. Licata P, et al.: **Levels of "toxic" and "essential" metals in samples of bovine milk from various dairy farms in Calabria, Italy.** *Environ. Int.* 2004; **30**(1): 1–6.  
[PubMed Abstract](#) | [Publisher Full Text](#)
191. IARC W: **World Cancer Report 2008.** World Health Organization; 2008; vol. 29.
192. Zhou X, et al.: **Relationships between Pb, As, Cr, and Cd in individual cows' milk and milk composition and heavy metal contents in water, silage, and soil.** *Environ. Pollut.* 2019; **255**(Pt 2): 113322.  
[PubMed Abstract](#) | [Publisher Full Text](#)
193. FAO: **Maximum residue limits (MRLs) and risk management recommendations (RMRs) for residues of veterinary drugs in foods.** *CX/MRL 2–2018.* London: FAO; 2018.
194. Europea, C: **Reglamentos UE N 37/2010 de la Comisión de 22 de diciembre de 2009.** Diario Oficial de La Unión Europea; 2010.
195. Bacanlı M, Başaran N: **Importance of antibiotic residues in animal food.** *Food Chem. Toxicol.* 2019; **125**: 462–466.  
[PubMed Abstract](#) | [Publisher Full Text](#)
196. De Oliveira AP, et al.: **Antimicrobial susceptibility of Staphylococcus aureus isolated from bovine mastitis in Europe and the United States.** *J. Dairy Sci.* 2000; **83**(4): 855–862.  
[PubMed Abstract](#) | [Publisher Full Text](#)
197. Ghidini S, et al.: **Residues of beta-lactam antibiotics in bovine milk: confirmatory analysis by liquid chromatography tandem mass spectrometry after microbial assay screening.** *Food Addit. Contam.* 2003; **20**(6): 528–534.  
[PubMed Abstract](#) | [Publisher Full Text](#)
198. Navratilova P, et al.: **Occurrence of tetracycline, chlortetracycline, and oxytetracycline residues in raw cow's milk.** *Czech J. Food Sci.* 2009; **27**(5): 379–385.  
[Publisher Full Text](#)



199. Shi Q, *et al.*: **Utilization of a lateral flow colloidal gold immunoassay strip based on surface-enhanced Raman spectroscopy for ultrasensitive detection of antibiotics in milk.** *Spectrochim. Acta A Mol. Biomol. Spectrosc.* 2018; **197**: 107–113. [PubMed Abstract](#) | [Publisher Full Text](#)
200. Marrugo-Padilla A, Méndez-Cuadro D, Rodríguez-Cavallo E: **Combined tetracycline and pyrethroid residues increases protein carbonylation in bovine milk.** *Int. Dairy J.* 2020; **107**: 104708. [Publisher Full Text](#)
201. Eskola M, *et al.*: **Worldwide contamination of food-crops with mycotoxins: Validity of the widely cited 'FAO estimate' of 25%.** *Crit. Rev. Food Sci. Nutr.* 2020; **60**(16): 2773–2789. [PubMed Abstract](#) | [Publisher Full Text](#)
202. Becker-Algeri TA, *et al.*: **Mycotoxins in Bovine Milk and Dairy Products: A Review.** *J. Food Sci.* 2016; **81**(3): R544–R552. [PubMed Abstract](#) | [Publisher Full Text](#)
203. Sweeney MJ, Dobson AD: **Mycotoxin production by *Aspergillus*, *Fusarium* and *Penicillium* species.** *Int. J. Food Microbiol.* 1998; **43**(3): 141–158. [Publisher Full Text](#)
204. Marin S, *et al.*: **Mycotoxins: occurrence, toxicology, and exposure assessment.** *Food Chem. Toxicol.* 2013; **60**: 218–237. [Publisher Full Text](#)
205. Liu Y, *et al.*: **Simultaneous detection and comparative pharmacokinetics of amoxicillin, clavulanic acid and prednisolone in cows' milk by UPLC-MS/MS.** *J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.* 2016; **1008**: 74–80. [PubMed Abstract](#) | [Publisher Full Text](#)
206. Ganmaa D, Sato A: **The possible role of female sex hormones in milk from pregnant cows in the development of breast, ovarian and corpus uteri cancers.** *Med. Hypotheses.* 2005; **65**(6): 1028–1037. [PubMed Abstract](#) | [Publisher Full Text](#)
207. Wilbey RA: **HEAT TREATMENT OF FOODS | Principles of Pasteurization.** *Encyclopedia of Food Microbiology.* Batt CA, Tortorello ML, editors. Oxford: Academic Press; 2014; p. 169–174. [Publisher Full Text](#)
208. Steele JH: **History, trends, and extent of pasteurization.** *J. Am. Vet. Med. Assoc.* 2000; **217**(2): 175–178. [PubMed Abstract](#) | [Publisher Full Text](#)
209. Özer B, Yaman H: **MILK AND MILK PRODUCTS | Microbiology of Liquid Milk.** *Encyclopedia of Food Microbiology.* Batt CA, Tortorello ML, editors. Oxford: Academic Press; 2014; p. 721–727. [Publisher Full Text](#)
210. Ryser ET: **Liquid Milk Products | Pasteurization of Liquid Milk Products: Principles, Public Health Aspects.** *Encyclopedia of Dairy Sciences.* Fuquay JW, editor. San Diego: Academic Press; 2011; p. 310–315. [Publisher Full Text](#)
211. Lorenzen PC, *et al.*: **A survey of the quality of extended shelf life (ESL) milk in relation to HTST and UHT milk.** *Int. J. Dairy Technol.* 2011; **64**(2): 166–178. [Publisher Full Text](#)
212. Sarkar S: **Microbiological considerations: pasteurized milk.** *Int. J. Dairy Sci.* 2015; **10**(5): 206–218. [Publisher Full Text](#)
213. Xiong L, *et al.*: **Effect of heat treatment on bacteriostatic activity and protein profile of bovine whey proteins.** *Food Res. Int.* 2020; **127**: 108688. [PubMed Abstract](#) | [Publisher Full Text](#)
214. Meunier-Goddik L, Sandra S: **Liquid Milk Products: Pasteurized Milk.** *Reference Module in Food Science.* Elsevier; 2016. [Publisher Full Text](#)
215. Ceni G, *et al.*: **Continuous inactivation of alkaline phosphatase and *Escherichia coli* in milk using compressed carbon dioxide as inactivating agent.** *J. CO2 Util.* 2016; **13**: 24–28. [Publisher Full Text](#)
216. Barut Gok S, *et al.*: **Inactivation of *E. coli* and *L. innocua* in milk by a thin film UV-C reactor modified with flow guiding elements (FGE).** *Int. J. Food Microbiol.* 2021; **343**: 109105. [PubMed Abstract](#) | [Publisher Full Text](#)
217. Sharma P, *et al.*: **Reduction of bacterial counts and inactivation of enzymes in bovine whole milk using pulsed electric fields.** *Int. Dairy J.* 2014; **39**(1): 146–156. [Publisher Full Text](#)
218. Makarapong D, *et al.*: **Development of an innovative apparatus using UV-C for controlling the number of microorganisms in raw milk after milking.** *Int. J. Dairy Technol.* 2020; **73**(1): 301–305. [Publisher Full Text](#)
219. Sheelamary M, Muthukumar M: **Effectiveness of ozone in inactivating *Listeria monocytogenes* from milk samples.** *World Journal of Young Researchers.* 2011; **1**(3): 40–44.
220. Shamila-Syuhada AK, *et al.*: **Inactivation of microbiota and selected spoilage and pathogenic bacteria in milk by combinations of ultrasound, hydrogen peroxide, and active lactoperoxidase system.** *Int. Dairy J.* 2016; **61**: 120–125. [Publisher Full Text](#)
221. Hongmei L, *et al.*: **Inactivation of microorganisms naturally present in raw bovine milk by high-pressure carbon dioxide.** *Int. J. Food Sci. Technol.* 2014; **49**(3): 696–702. [Publisher Full Text](#)
222. Stratakos AC, *et al.*: **Effect of high pressure processing on the safety, shelf life and quality of raw milk.** *Innovative Food Sci. Emerg. Technol.* 2019; **52**: 325–333. [Publisher Full Text](#)
223. Yasmin N, *et al.*: **Inactivation of foodborne pathogens on food packaging and in cow milk by exposure to a Nd:YAG laser.** *Can. J. Phys.* 2017; **95**(7): 662–669. [Publisher Full Text](#)
224. Sharma P, *et al.*: **Bacterial inactivation in whole milk using pulsed electric field processing.** *Int. Dairy J.* 2014; **35**(1): 49–56. [Publisher Full Text](#)
225. Cregenzán-Alberti O, *et al.*: **Suitability of cCRSM as a tool to predict inactivation and its kinetics for *Escherichia coli*, *Staphylococcus aureus* and *Pseudomonas fluorescens* in homogenized milk treated by manothermosonication (MTS).** *Food Control.* 2014; **39**: 41–48. [Publisher Full Text](#)
226. Zhang YH, *et al.*: **Enhanced degradation of five organophosphorus pesticides in skimmed milk by lactic acid bacteria and its potential relationship with phosphatase production.** *Food Chem.* 2014; **164**: 173–178. [PubMed Abstract](#) | [Publisher Full Text](#)
227. Yuan S, *et al.*: **Degradation of parathion methyl in bovine milk by high-intensity ultrasound: Degradation kinetics, products and their corresponding toxicity.** *Food Chem.* 2020; **327**: 127103. [PubMed Abstract](#) | [Publisher Full Text](#)
228. Zhao X-H, Wang J: **A brief study on the degradation kinetics of seven organophosphorus pesticides in skimmed milk cultured with *Lactobacillus* spp. at 42°C.** *Food Chem.* 2012; **131**(1): 300–304. [Publisher Full Text](#)
229. Shen C, *et al.*: **Pb(2+) and Hg(2+) removal from polluted milk by di-acrylated Pluronic P123 hydrogels.** *Food Chem.* 2018; **258**: 331–336. [PubMed Abstract](#) | [Publisher Full Text](#)
230. Massoud R, *et al.*: **Lead bioremoval from milk by *Saccharomyces cerevisiae*.** *Biocatal. Agric. Biotechnol.* 2019; **22**: 101437. [Publisher Full Text](#)
231. Massoud R, *et al.*: **Lead and cadmium biosorption from milk by *Lactobacillus acidophilus* ATCC 4356.** *Food Sci. Nutr.* 2020; **8**(10): 5284–5291. [PubMed Abstract](#) | [Publisher Full Text](#)
232. Massoud R, *et al.*: **Mercury Biodecontamination from Milk by using *L. acidophilus* ATCC 4356.** *J. Pure Appl. Microbiol.* 2020; **14**(4): 2313–2321. [Publisher Full Text](#)
233. Massoud R, *et al.*: **Mercury biosorption process by using *Saccharomyces cerevisiae* in milk.** *J. Food Process. Preserv.* 2021; **45**(1): 1–9.
234. Massoud R, *et al.*: **Cadmium Bioremoval by *Saccharomyces cerevisiae* in Milk.** *Journal of Medical Microbiology and Infectious Diseases.* 2020; **8**(1): 29–33. [Publisher Full Text](#)
235. Massoud R, *et al.*: **The Biosorption Capacity of *Saccharomyces Cerevisiae* for Cadmium in Milk.** *Dairy.* 2020; **1**(2): 169–176. [Publisher Full Text](#)
236. Alsager OA, *et al.*: **Removal of antibiotics from water and waste milk by ozonation: kinetics, byproducts, and antimicrobial activity.** *Ecotoxicol. Environ. Saf.* 2018; **158**: 114–122. [PubMed Abstract](#) | [Publisher Full Text](#)
237. Kitazono Y, *et al.*: **Antibiotic removal from waste milk by electrochemical process: degradation characteristics in concentrated organic solution.** *J. Mater. Cycles Waste Manag.* 2016; **19**(3): 1261–1269. [Publisher Full Text](#)
238. Kitazono Y, *et al.*: **Selective degradation of tetracycline antibiotics present in raw milk by electrochemical method.** *J. Hazard. Mater.* 2012; **243**: 112–116. [PubMed Abstract](#) | [Publisher Full Text](#)
239. Aguilar JFF, *et al.*: **Selective removal of tetracycline residue in milk samples using a molecularly imprinted polymer.** *J. Polym. Res.* 2020; **27**(7). [Publisher Full Text](#)

240. Kumar S, *et al.*: **Photocatalytic, fluorescent BiPO<sub>4</sub>@Graphene oxide based magnetic molecularly imprinted polymer for detection, removal and degradation of ciprofloxacin.** *Mater. Sci. Eng. C Mater. Biol. Appl.* 2020; **111**: 110777.  
[PubMed Abstract](#) | [Publisher Full Text](#)
241. Alsager OA, Alnajrani MN, Alhazzaa O: **Decomposition of antibiotics by gamma irradiation: Kinetics, antimicrobial activity, and real application in food matrices.** *Chem. Eng. J.* 2018; **338**: 548–556.  
[Publisher Full Text](#)
242. Bodbodak S, *et al.*: **Selective decontamination of aflatoxin M1 in milk by molecularly imprinted polymer coated on the surface of stainless steel plate.** *Int. J. Dairy Technol.* 2018; **71**(4): 868–878.  
[Publisher Full Text](#)
243. Ismail A, *et al.*: **Effect of different microbial concentrations on binding of aflatoxin M 1 and stability testing.** *Food Control.* 2017; **73**: 492–496.  
[Publisher Full Text](#)
244. Moussa AI, *et al.*: **Efficacy of Kaolin and Bentonite Clay to Reduce Aflatoxin M1 Content in Contaminated Milk and Effects on Milk Quality.** *Pak. Vet. J.* 2020; **40**(2): 181–186.  
[Publisher Full Text](#)
245. Abdelmotilib N, *et al.*: **Aflatoxin M1 Reduction in Milk by a Novel Combination of Probiotic Bacterial and Yeast Strains.** *European J. Nutr. Food Saf.* 2018; **8**: 83–99.  
[Publisher Full Text](#)
246. Martinez MP, *et al.*: **Probiotic bacteria and yeasts adsorb aflatoxin M1 in milk and degrade it to less toxic AFM1-metabolites.** *Toxicon.* 2019; **172**: 1–7.  
[PubMed Abstract](#) | [Publisher Full Text](#)
247. Assaf JC, *et al.*: **A novel method for elimination of aflatoxin M1 in milk using Lactobacillus rhamnosus GG biofilm.** *Int. J. Dairy Technol.* 2019; **72**(2): 248–256.  
[Publisher Full Text](#)
248. Carraro A, *et al.*: **Clay minerals as adsorbents of aflatoxin M1 from contaminated milk and effects on milk quality.** *Appl. Clay Sci.* 2014; **88-89**: 92–99.  
[Publisher Full Text](#)

# Open Peer Review

Current Peer Review Status:    

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## Version 1

Reviewer Report 09 February 2022

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**Lourdes Casas-Cardoso** 

Department of Chemical Engineering and Food Technology, Faculty of Science, International Campus of Excellence in Agrifood, ceiA3, University of Cadiz, Puerto Real, Spain

The review topic: "Contaminants in cow's milk, pasteurization and alternative technologies in the removal of these contaminants" is discussed extensively in the context of the paper.

Table 1 shows the pathogens in cow's milk reported in the literature and Table 2 summarizes the chemical contaminants in cow's milk reported in the literature. All statements are adequately supported by citations.

Pasteurization is a technology used in the industry, however it is exclusive for the elimination of microbial contaminants. Table 3 describes alternative methods to pasteurization: supercritical carbon dioxide, inactivation by pulsed electric fields, inactivation by ozonation. Alternative methods have proven to be efficient in degrading several contaminants; however, they are still not sufficient nor applied on an industrial scale.

The conclusions are appropriate. It is suggested to continue research on alternative methods.

**Is the topic of the review discussed comprehensively in the context of the current literature?**

Yes

**Are all factual statements correct and adequately supported by citations?**

Yes

**Is the review written in accessible language?**

Yes

**Are the conclusions drawn appropriate in the context of the current research literature?**

Yes

**Competing Interests:** No competing interests were disclosed.

**Reviewer Expertise:** natural products, supercritical fluid extraction, countercurrent extraction column, supercritical impregnation, active compounds

**I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.**

Reviewer Report 09 February 2022

<https://doi.org/10.5256/f1000research.120201.r121007>

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**Qiansheng Huang** 

Center for Excellence in Regional Atmospheric Environment, Key Lab of Urban Environment and Health, Institute of Urban Environment, Chinese Academy of Sciences, Xiamen, China

**This review attempts to summarize the knowledge generated with respect to the various types of contamination that cow's milk has and the effect they have on human health. As a central axis, it focuses on strategies to eliminate these contaminants so that their consumption is safer over the population in general. The treated theme is significant, with a real involvement in food security. I consider that the manuscript deserves to be published, not before taking into account the comments detailed below:**

**COMMENT 1:** This idea "they have not demonstrated efficacy in elimination contaminants" is repetitive in the summary.

**COMMENT 2:** The first paragraph of the introduction is not really a paragraph but a single long sentence. Please correct those errors throughout the document for readability.

**COMMENT 3:** Second paragraph of the introduction, last line: please be specific when "United States" e.g. United States of America.

**COMMENT 4:** The last sentence of the first paragraph of section 2, this must be better organized, the phrase is very long and can lead to confusion. Additionally, use the punctuation signs correctly.

**COMMENT 5:** In the fourth paragraph of section 1, reference is made to pasteurization is a method considered for the elimination of "non-chemical compounds". Specify what types and metabolites resulting from the application of this process.

**COMMENT 6:** The following phrase, "For better analysis and understanding, the classification of

contaminants according to the origin is microbial contaminants and chemical contaminants (Figure 2)" must be incorporated into a paragraph, it is not correct to leave it alone.

**COMMENT 7:** In the paragraph before point 2.1, two contiguous phrases initiate in this same way: "Among chemical contaminants.....". Please correct that.

**COMMENT 8:** This phrase "The main types of microorganisms present in milk are bacteria, yeasts, and molds, which represent the different types of microorganisms present in cow's milk." It is redundant. Please improve it.

**COMMENT 9:** When a bibliographic reference is made within the text, indicate the year after appointing the authors. Apply in all cases.

**COMMENT 10:** In reference to Figure 1: specify contaminants that come from milk containers (each type).

**COMMENT 11:** Last paragraph of section "2.", third line: indicate the reasons why there may be remanence of microbes even after pasteurization, considering that the process (pasteurization) has been carried out correctly.

**COMMENT 12:** SECTION 2.1: Here the "non-pathogenic" organisms should be included and those that cause alteration of milk, whose result is harmful to consumer. Improve the format of the tables.

**COMMENT 13:** SECTION 2.2.1, Literal "ii". What mean "According to the analysis of Table 2, Claborn et al.181 report.....". What type of analysis was made with "Table 2"?, Who made it?

**COMMENT 14:** SECTION 2.2.1, Literal "II". In reference to "This confirms that skin contaminated with these pesticides is another route of contamination of raw cow's milk." The evidence presented is causal, the most appropriate term, in this case, would be "evidence suggests...". "

**COMMENT 15:** SECTION 2.2.1, Literal "II". "In forage..." is indicated. Specify the conditions of the sample and the possible source of contamination, allowing each case to understand more dynamically. Apply in all cases.

**COMMENT 16:** Throughout the text, there are many redundant ideas in the same phrase or paragraph, please correct that.

**COMMENT 17:** SECTION 2.2.1: Include contaminants such as PFAAs and other organic compounds.

**COMMENT 18:** SECTION 2.2.2: Is not informative and redundant in front of other sections of the document. Improve it by including figures or tables with broader data.

**COMMENT 19:** Linking words and phrases like "such as...." are used excessively. Please vary the expressions used in the text. Apply in all cases.

**COMMENT 20:** SECTION 2.2.3, first sentence. Be more specific in the statement.

**COMMENT 21:** SECTION 2.2.3, Second paragraph, first sentence. Here is specified about mastitis, although this pathology has already been named previously. Any specification of some pathology must be made it the first time in which it is named. A similar case occurs with the pasteurization reference on page 16.

**COMMENT 22:** The document has good information, but this is shown a little messy. The writing and sequential logical structure of the manuscript are the main problems. Also, it is necessary to improve the format of the tables. Improve the resolution of Figure 2.

**COMMENT 23:** Specify the harmful effects of each type of contaminant. Specify the accumulation of every contaminant depending on the milk class (which type of cow produces). Specify the difference, in relation to the presence of contaminants, according to the fat content of the milk (whole, half- skimmed, and skimmed).

**COMMENT 24:** SECTION 2.2.5: Include the explanation about the hormones of the cows depending on the life stage of them, and its effect on the consumer.

**COMMENT 25:** SECTION 3: It is messy, it does not have a "friendly" order for the reader. Address the actual effect that the pasteurization process has on the structure of each type of contaminant (chemical contaminants).

**COMMENT 26:** TABLE 3: Describe the metabolites produced by alternative methods for the elimination of chemical contaminants.

**COMMENT 27:** Describe the effect of each type of contaminant removal method on the nutritional profile of milk.

**Is the topic of the review discussed comprehensively in the context of the current literature?**

Yes

**Are all factual statements correct and adequately supported by citations?**

Yes

**Is the review written in accessible language?**

Yes

**Are the conclusions drawn appropriate in the context of the current research literature?**

Yes

**Competing Interests:** No competing interests were disclosed.

**Reviewer Expertise:** public health

**I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have**

**significant reservations, as outlined above.**

Reviewer Report 07 February 2022

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### **Esmeralda García Díaz**

Centro de Química, Instituto de Ciencias, Benemérita Universidad Autónoma de Puebla, Puebla, Mexico

The manuscript entitled: "Contaminants in the cow's milk we consume? Pasteurization and other technologies in the elimination of contaminants" is valuable and presents an exhaustive review of the different types of contaminants in raw/pasteurized cow's milk and analyze the application of alternative processes for the elimination or degradation of contaminants. It provides relevant information about the sources of contamination and the health implications of ingesting these contaminants through milk, adequately supported by citations. It presents a wide variety of contaminants with their details regarding the contamination of milk. Information about alternative treatments to remove contaminants is also relevant and abundant. However, the tables need to be improved to present relevant information, which the authors can surely provide. Please see the following comments regarding the tables:

#### **COMMENTS**

in Figure 1, the "processing and packaging stages" label is missing. Plasticizers that are used in containers, such as BPA need to be included.

Robert Frankowski, Tomasz Grześkowiak, Beata Czarczyńska-Goślińska & Agnieszka Zgoła-Grześkowiak (2022) Occurrence and dietary risk of bisphenols and parabens in raw and processed cow's milk, *Food Additives & Contaminants: Part A*, 39:1, 116-129, DOI: 10.1080/19440049.2021.1986234

A better title for figure 2 would be "Incidence of contaminants in bovine..."

Page 4, sentence "For better analysis and understanding, the classification of contaminants according to the origin is microbial contaminants and chemical contaminants (Figure 2)", needs to be rewritten, since the word "contaminants" is repeated several times. I suggest: Figure 2 presents the classification of cow's milk contaminants and their microbial or chemical origin.

Table 1 is long but contains relatively little information for its size. The column "Type of milk" is repetitive - separate horizontally by type of milk and add a column with the main identification method used in each case.

Table 2 is also long and contain repetitive information, with columns labelled as MRL being practically empty. it would be more useful to put the concentration interval reported in the referred works. MRL data can be mentioned in text.

Throughout the document, round percentage values to make analysis easy.

Page 14,

In the sentence "Therefore, it is important to control the presence of antibiotic residues in food to avoid the appearance of resistance to these antibiotics in humans".

Who becomes resistant, humans or microorganisms? Its unclear in the sentence with the expression "appearance of resistance".

In sentence "Considering that the use of antibiotics in cattle generates residues in milk, their excessive use should be avoided, and the elimination times before milking should be respected in order to avoid the presence of these contaminants".

Please, reference the "elimination times before milking" to support the establishment of this time in some reported work.

Page 16

The sentence "The literature mentions alternatives for eliminating specific microbial and chemical contaminants", change 'mentions' to 'reports' instead.

This sentence is confusing: "UV-C lamp wattage did not significantly influence the fat concentration in the milk, which means that it is necessary to improve the method to guarantee an effective reduction of these microorganisms if milk transport time exceeds two hours without cooling".

It's not clear the relationship between lamp wattage, fat concentration, microorganisms and time of cooling.

My conclusion: accept after minor revision.

## References

1. Frankowski R, Grześkowiak T, Czarczyńska-Goślińska B, Zgoła-Grześkowiak A: Occurrence and dietary risk of bisphenols and parabens in raw and processed cow's milk. *Food Addit Contam Part A Chem Anal Control Expo Risk Assess.* 2022; **39** (1): 116-129 [PubMed Abstract](#) | [Publisher Full Text](#)

**Is the topic of the review discussed comprehensively in the context of the current literature?**

Yes

**Are all factual statements correct and adequately supported by citations?**

Yes

**Is the review written in accessible language?**

Yes

**Are the conclusions drawn appropriate in the context of the current research literature?**

Yes

**Competing Interests:** No competing interests were disclosed.

**Reviewer Expertise:** Adsorption and photocatalysis of emerging pollutants of



water. Chromatography of compounds of ambient and biological interest

**I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.**

Reviewer Report 02 February 2022

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**Pawel Konieczynski** 

Department of Analytical Chemistry, Medical University of Gdansk, Gdansk, Poland

In my opinion the article entitled: "Contaminants in the cow's milk we consume? Pasteurization and other technologies in the elimination of contaminants" is valuable and covers an important problem from the point of view of studies of food safety, especially milk. It was written based on wide literature screening (about 250 references) and presents the data and conclusions in a clear and comprehensive way. However, I have two comments:

1. Why didn't the authors discuss the contaminants in milk (metals especially) in the context of norms of PTWI or ADI?
2. Please add "Metals and metalloids" in the heading of Table 2 (page 8) since selenium is included in it.

My conclusion: accept after minor revision

**Is the topic of the review discussed comprehensively in the context of the current literature?**

Yes

**Are all factual statements correct and adequately supported by citations?**

Yes

**Is the review written in accessible language?**

Yes

**Are the conclusions drawn appropriate in the context of the current research literature?**

Yes

**Competing Interests:** No competing interests were disclosed.

**Reviewer Expertise:** analytical chemistry

**I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.**

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