



Review

# Protective Role of Melatonin and Its Metabolites in Skin Aging

Georgeta Bocheva <sup>1,\*</sup>, Radomir M. Slominski <sup>2,3</sup>, Zorica Janjetovic <sup>2</sup>, Tae-Kang Kim <sup>2</sup>, Markus Böhm <sup>4</sup>, Kerstin Steinbrink <sup>4</sup>, Russel J. Reiter <sup>5</sup>, Konrad Kleszczyński <sup>4</sup> and Andrzej T. Slominski <sup>2,6,\*</sup>

<sup>1</sup> Department of Pharmacology and Toxicology, Medical University of Sofia, 1431 Sofia, Bulgaria

<sup>2</sup> Department of Dermatology, University of Alabama at Birmingham, Birmingham, AL 35294, USA; radomir.slominski@gmail.com (R.M.S.); zjanjetovic@uabmc.edu (Z.J.); tkkim4567@gmail.com (T.-K.K.)

<sup>3</sup> Graduate Biomedical Sciences Program, University of Alabama at Birmingham, Birmingham, AL 35294, USA

<sup>4</sup> Department of Dermatology, University of Münster, Von-Esmarch-Str. 58, 48149 Münster, Germany; boehmma@ukmuenster.de (M.B.); kerstin.steinbrink@ukmuenster.de (K.S.); konrad.kleszczyński@ukmuenster.de (K.K.)

<sup>5</sup> Department of Cellular and Structural Biology, UT Health Science Center, San Antonio, TX 78229, USA; reiter@uthscsa.edu

<sup>6</sup> Pathology and Laboratory Medicine Service, VA Medical Center, Birmingham, AL 35294, USA

\* Correspondence: bocheva\_georgeta@yahoo.com (G.B.); aslominski@uabmc.edu (A.T.S.)

**Abstract:** The skin, being the largest organ in the human body, is exposed to the environment and suffers from both intrinsic and extrinsic aging factors. The skin aging process is characterized by several clinical features such as wrinkling, loss of elasticity, and rough-textured appearance. This complex process is accompanied with phenotypic and functional changes in cutaneous and immune cells, as well as structural and functional disturbances in extracellular matrix components such as collagens and elastin. Because skin health is considered one of the principal factors representing overall “well-being” and the perception of “health” in humans, several anti-aging strategies have recently been developed. Thus, while the fundamental mechanisms regarding skin aging are known, new substances should be considered for introduction into dermatological treatments. Herein, we describe melatonin and its metabolites as potential “aging neutralizers”. Melatonin, an evolutionarily ancient derivative of serotonin with hormonal properties, is the main neuroendocrine secretory product of the pineal gland. It regulates circadian rhythmicity and also exerts anti-oxidative, anti-inflammatory, immunomodulatory, and anti-tumor capacities. The intention of this review is to summarize changes within skin aging, research advances on the molecular mechanisms leading to these changes, and the impact of the melatonergic anti-oxidative system controlled by melatonin and its metabolites, targeting the prevention or reversal of skin aging.

**Keywords:** melatonin; AFMK; skin aging; photoaging; UV radiation; oxidative stress; anti-aging properties



**Citation:** Bocheva, G.; Slominski, R.M.; Janjetovic, Z.; Kim, T.-K.; Böhm, M.; Steinbrink, K.; Reiter, R.J.; Kleszczyński, K.; Slominski, A.T. Protective Role of Melatonin and Its Metabolites in Skin Aging. *Int. J. Mol. Sci.* **2022**, *23*, 1238. <https://doi.org/10.3390/ijms23031238>

Academic Editor:  
Alessandro Terroni

Received: 21 December 2021

Accepted: 20 January 2022

Published: 22 January 2022

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

The skin is the most complex and multifunctional self-regulating organ. Facing the environment, the cutaneous barrier protects the body from external stressors and is essential for cutaneous and overall body homeostasis [1–4]. Additionally, the skin, together with the hypodermis (subcutaneous fat), is both a source and a target for many hormones and neuromediators [5–15], making it an independent and fully functioning peripheral endocrine organ [5,16]. Important mechanisms of the skin in maintaining homeostasis and protecting the whole body include the regulation of oxidative stress mechanisms and circadian rhythm [17]. The skin has its own peripheral circadian machinery, working either along with the central circadian clock or autonomously [18]. Like other organs, the skin also follows a rhythmicity in the production of bioactive molecules and sebum, and a periodicity in hydration, surface pH, skin temperature, capillary blood flow, etc. [19–21]. To counteract oxidative stress, the skin produces several protective molecules, including

melatonin, vitamin D, and melanin [22–27]. Unfortunately, the endogenous antioxidant capacity of the skin is reduced with age and oxidative damage accumulation during aging, making the aged skin more vulnerable to environmental insults, especially ultraviolet (UV) radiation, air pollutants, and pathogens.

Biological aging is a natural phenomenon accompanied by a progressive loss of functional capacity, physiological integrity, and morphological features of the organism. The chronobiological functioning of the skin influences its aging. Mechanisms underlying the aging process include oxidative stress, mitochondrial dysfunction, disruption of circadian rhythms, inflammation, proteostasis, telomere attrition, genomic instability, epigenetic alterations, and a decreased capacity for tissue repair [28,29]. Circadian clocks are vital to human health through the rhythmic activity of physiological and neuro-endocrine functions. Aging is associated with a decline in circadian rhythm and a dampening of circadian gene expression [30] that can augment oxidative stress through an increased generation and accumulation of reactive oxygen species (ROS) [31]. Melatonin as well as vitamin D can regulate the cutaneous redox state and circadian rhythm [17,32].

The indolic hormone melatonin, which is released by the pineal gland, orchestrates circadian rhythms and sleep promotion [33,34]. There are also extrapineal tissues, like human skin, where it is synthesized [22,35] and works on site as a multifunctional molecule. Cutaneous melatonin production also follows rhythmicity, with the highest levels of cutaneous melatonin in the evening [36]. Melatonin produced in the skin exerts a protective effect against cutaneous damage caused by external factors [37]. Melatonin and its metabolites, including indolic derivatives such as 6-hydroxymelatonin and 2-hydroxymelatonin and kynuric metabolites such as AFMK and AMK, can limit oxidative stress via the scavenging of toxic radicals and inhibition of their generation, especially at the mitochondrial level [22,23,35,38–42]. Additionally, melatonin demonstrates potent antioxidant properties through its capacity to stimulate the production of antioxidant enzymes [43]. Furthermore, melatonin can also ameliorate DNA damage caused by environmental factors [40] and has anti-inflammatory [44] and anti-apoptotic effects [45,46]. This pleotropic regulatory action of melatonin and its metabolites on the skin makes them powerful anti-aging molecules. Because the synthesis of peripheral melatonin decreases with aging, the endogenous cutaneous melatonin production could be amplified with topical application of melatonin, which is considered as an effective photoprotective agent [37,47] and a very promising anti-aging strategy [48].

## 2. Skin Aging

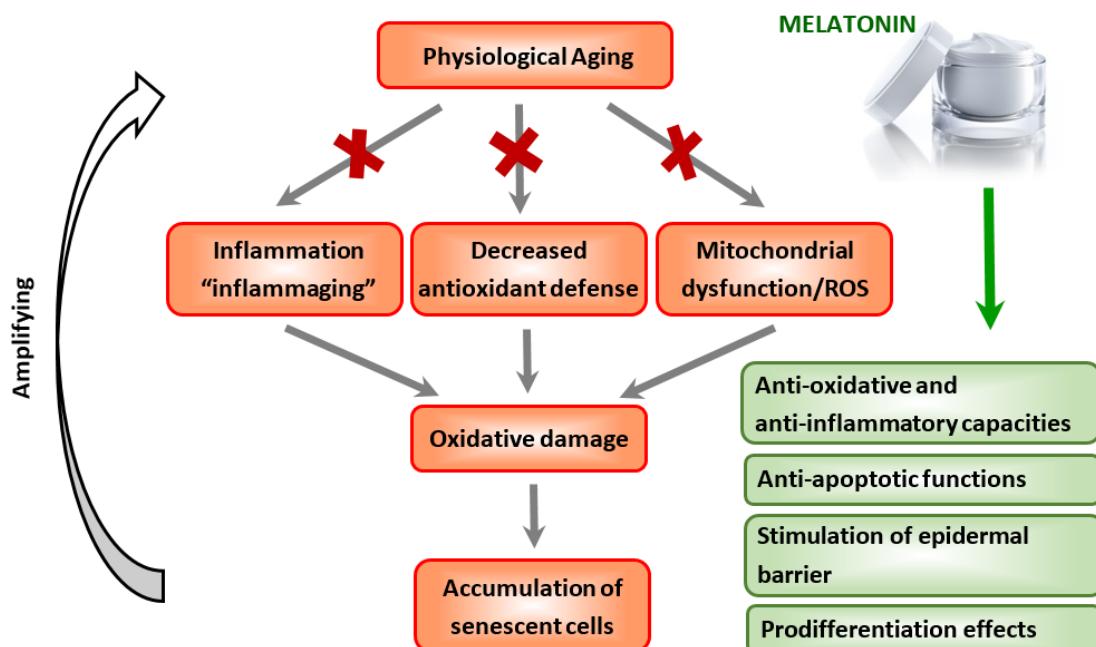
### 2.1. Natural Process of Skin Aging

The aging of the skin is a natural and genetically determined process with progressive morphological and functional alterations, which are influenced by the total exposure to both the environmental and internal factors over the human lifespan [49]. The physiological maturing process results in most of the phenotypic changes of aging observed in all skin areas, including the appearance of fine wrinkles, atrophy with reduced elasticity, and prominent dryness often accompanied by pruritus. However, they vary among different anatomical regions and within different ethnicities [50,51].

Chronological (physiological) skin aging is caused mainly by an imbalanced endocrine circadian rhythmicity, with a hormonal decline and changes in gene expression with advancing age [51–54]. Aging affects proopiomelanocortin (POMC) and POMC-derived peptides, especially melanocortin receptor 1 (MC1R) and MC2R agonists, implicating their role in the general process of skin aging [55]. The single nucleotide polymorphisms (SNPs) of the *MC1R* gene are significantly linked to perceived facial age [56]. Putative functionally relevant SNPs can also affect the other pigmentation-related genes (e.g., *IRF4*, *ASIP*, *BNC2*) [57]. These genetic variations identified in skin color genes contribute to facial pigmented spots during aging through pathways independent of the melanin production [58]. Recently, the association of variants in *IRF4*, *MC1R*, and *SLC45A2* with skin wrinkling was confirmed in more ethnic groups [59]. The same study, using Latin Americans of a

mixed continental ancestry cohort, reported genetic variations in two new candidate genes, *VAV3* and *SLC30A1*, associated with facial skin wrinkling and mole count, respectively [59]. Epigenetic mechanisms are also implicated in the direct regulation of the homeostasis and regeneration of the aged skin [60].

The process of aging involves the excessive senescence of keratinocytes, fibroblasts, and melanocytes over time, with accumulation contributing to decreased cutaneous regenerative potential and skin aging (Figure 1) [61–64]. Senescent skin cells are metabolically active and secrete diverse pro-inflammatory cytokines, chemokines, proteases, and growth factors in a state known as the senescence-associated secretory phenotype (SASP) [65]. This SASP's state plays a role in the functional decline of physiologically aged skin [66,67]. With accelerating age, the immune system also undergoes senescence that can cause dysregulation of immune responses and possible impairment of the cutaneous immunological defense and adaptive capacity [68–70]. Indeed, the main cellular perturbations in the skin inducing senescence are inflammation and oxidative stress.



**Figure 1.** A possible role of melatonin in the prevention and treatment of physiological skin aging. Red crosses ( $\times$ ) indicate protective action of melatonin against inflamaging, oxidative stress, and mitochondrial damage.

In chronological aging, ROS are produced through cellular oxidative metabolism, where mitochondrial dysfunction has an impact. Accumulating evidence supports a strong link between a decline in mitochondrial quality and function and the aging process [71,72]. Mitochondria also undergo aging, characterized by significant increase in ROS generation, a decrease in oxidative capacity and antioxidant defense, and a reduction in oxidative phosphorylation and adenosine triphosphate (ATP) production. This age-related impaired function of mitochondria further enhances mitochondria-mediated apoptosis, which contributes to an increase in the percentage of apoptotic cells [73]. An important target of ROS is mtDNA, in which damage and decline in function result in further enhancement of ROS production [74,75].

## 2.2. Environmentally Induced Skin Aging

Physiological aging is influenced by environmental stressors which can drive the premature aging of the skin. The most prominent external factors are ultraviolet radiation (UV) [76–78] and ambient pollutants [79–82]. Long-term exposure of the skin to these environmental insults stimulates ROS and reactive nitrogen species (RNS) production, and

generates oxidative stress [83,84]. Further, they contribute to premature cutaneous aging, demonstrated by deep-winkle formation, sagging, and pigmentation affecting mainly exposed areas like the skin on the face, neck, head, and hands [85,86]. Chronic exposure can also cause an impairment of the epidermal barrier function [87] and alterations in the skin microbiome [88], leading to significant morbidity [70,89].

UVR is the most widely recognized harmful environmental factor that affects cutaneous biology and contributes to photodamage. The superposition of the solar damage on the physiological aging process leads to chronic inflammation, impaired regenerative capacity, and photoaging, which correlates with enhanced cancer risk [76,90–92]. Both ultraviolet (UV)A (315–400 nm) and UVB (280–315 nm) wavelengths have been shown to contribute to photoaging, either by imbalanced ROS/RNS production or by direct DNA damage [84,91]. Indeed, UVA is considered to play a major role in the skin's aging process. UVA constitutes more than 80% of total daily UV irradiation and penetrates 5–10 times deeper into the reticular dermis, significantly damaging the extracellular matrix (ECM) in comparison with UVB [91]. This UVA effect is based on an increase of the transcription of matrix metalloproteinases (MMPs), especially the collagenolytic enzyme MMP-1 in dermal fibroblasts, causing massive collagen degradation and procollagen inhibition. Loss of balance between the essential tissue-specific inhibitor (TIMP1) of MMPs and MMP-1 can contribute to wrinkle development [93]. Thus, MMP-1 serves as an important regulator in photoaging [94]. Additionally, UVA exposure stimulates the activity of elastase and hyaluronidase and inhibits hyaluronan synthesis, thereby altering the composition of proteoglycans and glycosaminoglycans in the dermis [84,95]. Chronic UVR (mainly UVA exposure) is also indirectly related to photoaging and photocancer due to an excessive generation of ROS and RNS, which can disrupt both the nuclear and mitochondrial DNA [96,97].

UVB can penetrate only through the epidermis but is biologically more active [76,98]. UVB radiation absorbed by DNA and RNA directly induces a formation of cyclobutane pyrimidine dimers (CPDs) and other photoproducts in keratinocytes [99]. Further, DNA photolesions may trigger various typical solar signature mutations in specific genes, including the tumor suppressor gene p53 [100,101]. The induced UVR accumulation of p53 protein in the nucleus in turn activates the transcription of genes responsible for cell cycle arrest, allowing DNA repair. P53 accumulation also results in an induction of apoptosis of the cells with unrepaired DNA damage [102].

The exposure of the skin to environmental air pollutants and their negative impact is of growing concern [103]. Their prolonged exposure can alter the skin homeostasis and has been associated with skin aging and other cutaneous pathologies [49,79,81]. Additionally, air pollutants, persistent organic pollutants, and heavy metals can behave like endocrine-disrupting chemicals (EDCs) [104]. Ozone from the smog and particulate matter (PM) in contact with the skin is capable of stimulating the production of ROS and generates oxidative stress, leading to typical phenotypic features of premature aging, including pigment spots and deep nasolabial folds [105,106]. Moreover, ultrafine particles (<0.1 μm) can penetrate tissues and localize in the mitochondria, resulting in mitochondrial damage from the oxidative processes [107]. Moreover, the chronic photopollution stress of the skin may aggravate UVR-mediated skin aging [108].

Generally, environmentally induced premature skin aging is mainly driven by oxidative events. The mitochondria can generate about 90% of the intracellular ROS and are thus considered a main source of free radical production [109,110]. In addition to mitochondrial ROS, another important source of free radicals is the nicotinamide adenined-inucleotide phosphate (NADPH) oxidase system, which also plays a key role in triggering of oxidative stress. Because of the oxidative stress, increased levels of highly reactive free radicals promote lipid peroxidation, protein oxidation, genomic and mitochondrial DNA (mtDNA) damage, and depleted enzymatic and non-enzymatic antioxidant defense systems of the skin [111–114]. The accumulation of ROS/RNS dysregulates cell signaling pathways, alters cytokine release, and leads to inflammation. Indeed, the overproduction

of ROS activates mitogen-activated protein kinases (MAPKs) and transcription factors such as nuclear factor- $\kappa$ B (NF- $\kappa$ B), nuclear factor erythroid 2-like (Nrf2), and c-Jun-N-terminal kinase (JNK) [115–117]. Levels of redox-sensitive activator protein-1 (AP-1) and NF- $\kappa$ B are found to be elevated within hours after exposure to low-dose UVB. Both NF- $\kappa$ B and AP-1 contribute to wrinkle formation and inflammation, and play crucial roles in accelerated skin aging. The up-regulation of AP-1 suppresses the transforming growth factor  $\beta$  (TGF- $\beta$ ) receptors, which further blocks procollagen synthesis [118]. Furthermore, activated AP-1 stimulates collagen breakdown by MMPs and triggers the main activator of the inflammatory response, NF- $\kappa$ B. NF- $\kappa$ B pathways are involved in a regulation of tissue homeostasis and aging [119,120]. ROS-triggered activation of NF- $\kappa$ B drives an elevation of proinflammatory cytokines (IL-1, IL-6, and TNF- $\alpha$ ) and MMPs, and decreases TGF- $\beta$  and collagen type I synthesis [119]. Additionally, enhanced NF- $\kappa$ B expression was found in mitochondrial DNA (mtDNA)-depleter mice, confirming that NF- $\kappa$ B signaling is a decisive mechanism contributing to the skin and hair follicle pathologies [114]. Solar-induced inflammation is also associated with deficiency of the aging suppressor hormone klotho [121]. Klotho is a transmembrane protein, and its function is possibly mediated through the toll-like receptor 4 (TLR4)/NF- $\kappa$ B axis signaling pathway [122]. Additionally, klotho can prevent NF- $\kappa$ B translocation, leading to an inhibition of the pro-inflammatory NF- $\kappa$ B pathway.

The endogenous Nrf2 is essential for cutaneous protection from oxidative insults and for regulating the redox balance during skin aging [116,123]. UVA, due to its longer wavelength, reaches the dermal fibroblasts *in vivo*, where it stimulates Nrf2-mediated antioxidant gene expression. Unlike UVA, UVB does not activate Nrf2 in skin cells or even appear to have an inhibitory effect [124,125]. However, vitamin D<sub>3</sub> derivatives, products of UVB action, can activate Nrf2 signaling [125]. Thus, Nrf2 and its downstream signaling play a crucial role in photoprotection [117,126].

Recently, some Sirtuins (SIRTs) have gained attention due to their epigenetic ability to deacetylate histone and nonhistone targets, modulating the expression of genes implicated in the oxidative stress response and apoptosis [127]. The expression of SIRT1 and SIRT6 is found to be significantly reduced in aged human fibroblasts [128]. In addition, UVB irradiation reduces the expression of SIRT1 [129]. Furthermore, down-regulation of SIRT1 leads to an increase in MMPs and NF- $\kappa$ B activity. Thus, the activation of SIRT1 proves to have beneficial impact on both chronological and premature skin aging [127].

### 3. Melatonin and Aging

#### 3.1. An Overview of the Synthesis, Metabolism, and Function of Melatonin

The phylogenetically ancient molecule melatonin (*N*-acetyl-5-methoxytryptamine) is widely distributed in nature [130–132] and can be formed almost in all living organisms, including plants [133–136]. Melatonin was first isolated and identified in bovine pineal gland by the dermatologist Aaron Lerner et al. in 1958 [137]. Lerner, together with his colleagues, was also the first to identify melatonin's chemical structure and its action as a lightening agent in melanophores counteracting the  $\alpha$ -melanocyte stimulating hormone ( $\alpha$ -MSH) [138]. Historically, in mammals, this indolamine was thought to be uniquely released by the pineal gland, playing a major role in the regulation of circadian day-night rhythms and seasonal biorhythms [33,139]. Pineal-released melatonin can be measured at lower concentrations in the blood than in the cerebrospinal fluid (CSF) of the third ventricle of the brain, suggesting its role as a protector of the brain against oxidative stress [140,141]. Later, extrapineal sites of melatonin production were established. Thus, melatonin is also synthesized in numerous peripheral tissues such as the bone marrow, retina, lens, cochlea, lungs, liver, kidney, pancreas, thyroid gland, female reproductive organs, and finally the skin [14,15,22,142–146]. Indeed, the synthesis of melatonin is a multistep process that first starts with hydroxylation of L-tryptophan to 5-hydroxy-tryptophan (5(OH)tryptophan, catalyzed by tryptophan hydroxylase [147–149]. Further 5(OH)tryptophan is decarboxylated to serotonin, which is subsequently transformed to *N*-acetylserotonin (NAS) by the enzyme arylalkylamine *N*-acetyltransferase (AANAT) [150,151]. Furthermore, it has been

found that serotonin can be acetylated to NAS by alternative enzymes including arylamine N-acetyltransferase [152–156]. The last step in the synthesis is a conversion of NAS to melatonin by hydroxyindole-O-methyl transferase (HIOMT) [157].

The levels of melatonin are regulated by its rapid metabolism in the liver or directly at the site of its synthesis in peripheral organs [158]. In the classical hepatic metabolism, CYP450 enzymes (CYP1A1, CYP1A2, and CYP1B1) degrade circulating melatonin to 6-OH-melatonin [159,160]. Melatonin can also be demethylated in the liver to NAS by CYP2C19 or CYP1A, which represents a minor microsomal pathway [161,162]. Through the alternative indolic pathway, melatonin is deacetylated by liver aryl acylamidase to 5-OH-tryptamine, which is further deaminated by monoamine oxidase A [163]. The metabolism of melatonin through kynuric pathway begins with the formation of  $N^1$ -acetyl- $N^2$ -formyl-5-methoxykynuramine (AFMK) in a peroxidase-like reaction. Further AFMK is deformylated to  $N^1$ -acetyl-5-methoxykynuramine (AMK) [164,165]. In mitochondria, an additional route of melatonin metabolism to AFMK by cytochrome C oxidation has also been described [166]. In the skin or skin cells, melatonin is metabolized rapidly through its 6-hydroxylation, through the indolic and kynuric pathway, and through non-enzymatic processes including phototransformation induced by UVB, UVA, and reactive oxygen species [167–169]. The main products of melatonin metabolism in the epidermis are 6-hydroxymelatonin, AFMK, AMK, 5-methoxytryptamine, 5-methoxytryptophol, and 2-hydroxymelatonin. These products accumulate in the epidermis at detectable concentrations [170,171].

The widespread melatonin distribution during evolution has made it as a vital multifunctional hormone, with remarkable essential functions [34,172]. The complex action of melatonin includes its work as a regulator of the circadian clock, a neurotransmitter and hormone, a metabolic modulator, and a modifier of cell response and cytokine release [173–177]. It also regulates the functions of many peripheral organs [174,178] and exerts oncostatic [179–184] and anti-aging capacity [48,185]. Many regulatory effects of melatonin on cardiovascular, endocrine, reproductive, and immune systems are mediated via specific melatonin 1 (MT1) and MT2 membrane receptors [19,186]. Melatonin, by interacting with MT1 and MT2, has been found to limit weight gain [176,187,188]. Melatonin can inhibit adipogenic differentiation and together with vitamin D, exhibits a negative regulation of adipogenesis in adipose-derived stem cells (ADSCs). It was recently found that melatonin significantly inhibited the transcription of specific adipogenesis-orchestrating genes, such as *aP2* and peroxisome proliferator-activated receptor  $\gamma$  (*PPAR- $\gamma$* ), as well as adipocyte-specific genes including lipoprotein lipase (*LPL*) and acyl-CoA thioesterase 2 (*ACOT2*). Moreover, melatonin and vitamin D can modulate ADSCs through the up-regulation of epigenetic regulatory genes like histone deacetylase 1 (HDAC1), SIRT1, and SIRT2 [189].

Melatonin can also inhibit the effects of estrogens [190], and exhibits cardioprotective [191,192] and anticonvulsant activity [193]. MT1 and MT2 are also important for protection of the skin against environmental stressors, aging, and cancerogenesis [179,194]. Moreover, often the melatonin level inversely correlates with an increased risk of cancer development. Of note, the blockage of melatonin receptors can impair the p53-dependent DNA damage response [195]. The antioxidant ability of melatonin relays the indirect receptor-mediated action, likely by the stimulation of antioxidant enzymes, SIRT3, and others [43,196]. Melatonin works also through non-receptor mediated mechanisms such as the direct scavenging of variety of reactive species (both ROS and RNS) to counteract oxidative stress [39,41,130,197–199]. In addition to its high antioxidant potential, receptor-independently, melatonin serves as a mitochondrial protector [200] and anti-inflammatory agent [201]. Some of the protective properties of melatonin are shared with its kynuric metabolites AFMK and AMK [178,202,203].

### 3.2. Protective Role of Melatonin in Systemic Aging

The “free radical theory of aging” has been discussed for over 50 years [204–206]. At the subcellular level, mitochondria are the major source for generation of a highly reactive

and destructive species like peroxynitrite and the hydroxyl radical [207]. Their excessive production, resulting in enhanced mitochondrial oxidative stress and mtDNA mutations, occurs along with human aging and age-related pathologies [208–210]. Some intracellular enzymes outside the mitochondria (e.g., xanthine oxidase, monoamine oxidase, NADPH oxidases) also impact on ROS production with advancing age [211–213]. Disturbances in mitochondrial redox balance promote cellular senescence and thus the mitochondria impairment determines the rate of aging [214]. Recently, it has been thought that most mtDNA mutations are caused by replication errors of mtDNA polymerase [215]. During aging, such defects in mtDNA replication machinery together with a failure of their repair might cause an accumulation of mutations with further mitochondrial dysfunction and augmentation of oxidative damage.

Since free radicals abundantly are generated in mitochondria in aging, molecules that reduce their mitochondrial production or detoxify them may slow the rate of systemic aging. Melatonin is such a molecule, and its role in aging has been on the focus of many scientists in the last 20 years [42,216–218]. It was found that surgical pinealectomy of young rats resulted after time in accelerated oxidative damage in multiple tissues due to circadian disruption, and melatonin-deficient animals aged more rapidly [219].

While dysfunctional mitochondria contribute to the aging process [220], melatonin can maintain optimal mitochondrial physiology [42,221,222]. Melatonin concentrations are found at higher levels in mitochondria than in other cellular organelles, suggesting its significant role as a mitochondrial-targeted molecule involved in mitochondrial processes [42,200]. The multiple beneficial protective actions of this indolic hormone at the mitochondrial level are well documented [223]. Melatonin can limit age-related oxidative stress directly by scavenging ROS/RNS [41,224] and by indirect activation of mitochondria-located superoxide dismutase (SOD2) [225]. Through the stimulation of mitochondria's localized SIRT3, melatonin prompts the deacetylation and activation of SOD2. The activation of antioxidant enzymes involved in the SIRT3/SOD2 signaling pathway by melatonin reduces mitochondrial oxidative damage and cytochrome C release, thus reducing mitochondria-related apoptosis [196,226]. Indeed, melatonin maintains the optimal mitochondrial membrane potential and preserves mitochondrial function not only by quenching free radicals [198] but also by inhibiting the mitochondrial permeability transition pore (MPTP) [227], activating uncoupling proteins (UCPs), and regulating mitochondrial biogenesis and dynamics [228].

Generally, melatonin may act as both pro- and anti-inflammatory molecule in a context-dependent fashion [201,229,230]. In aging, melatonin preferentially exerts anti-inflammatory actions on aging-related low-grade inflammation. Melatonin stimulates SIRT1, and their anti-inflammatory activities overlap during the process of aging [231]. SIRT1, functioning as an epigenetic aging regulator, alleviates the inflammation by down-regulating TLR4, which mediates pro-oxidant effects through the NF- $\kappa$ B signaling pathway [229]. Melatonin, by an inhibition of either TLR-4 and the toll receptor-associated activator of interferon (TRIF), can suppress the release of several pro-inflammatory cytokines like TNF $\alpha$ , IL-1 $\beta$ , IL-6, and IL-8 [232,233].

To summarize, melatonin, with its capacity to mitigate oxidative stress, protect mitochondrial functions, modulate the immune system, reduce inflammation, enhance circadian rhythm amplitudes, and exhibit neuroprotection, beneficially results in retarding the process of aging [174,216,234–240].

#### 4. Melatonin, Its Metabolites and Skin Aging

##### 4.1. Overview of Cutaneous Melatonergic System

Melatonin is synthesized and metabolized in the skin. The ability of the mammalian skin to synthesize melatonin from serotonin through NAS was first published in 1996 [241]. Follow-up studies have provided the evidence that human skin, as well as normal keratinocytes, melanocytes, and melanoma cells, can endogenously produce melatonin [13–15,22,242]. Moreover, the skin cells express the essential enzymes for trans-

forming tryptophan to serotonin and eventually to melatonin, like tryptophan hydroxylase (TPH1—all skin cells; TPH2—melanocytes and dermal fibroblasts) [13,14,23,243], AANAT/serotonin N-acetyltransferase (SNAT) and NAT [154,155], and HIOMT/N-acetylserotonin-methyltransferase (NASM) [13,14]. Cutaneous serotonin can be acetylated to NAS by both AANAT and NAT [13,152,156]. Hair follicles also generate melatonin and express its functional receptors [244]. Recently, the concentrations of melatonin and its metabolites in the human epidermis were quantified by liquid chromatography–mass spectrometry (LC-MS) [170,171]. The level of epidermal melatonin varies depending on race, gender, and age. Kim et al. measured the highest concentrations of melatonin among African Americans and elderly Caucasians. The levels of its kynuric metabolite AFMK were significantly higher in Caucasian males, whereas AMK demonstrated higher concentration in African Americans than in Caucasians [171]. The accumulation of AMK in the epidermis suggests the cutaneous transformation of AFMK to AMK.

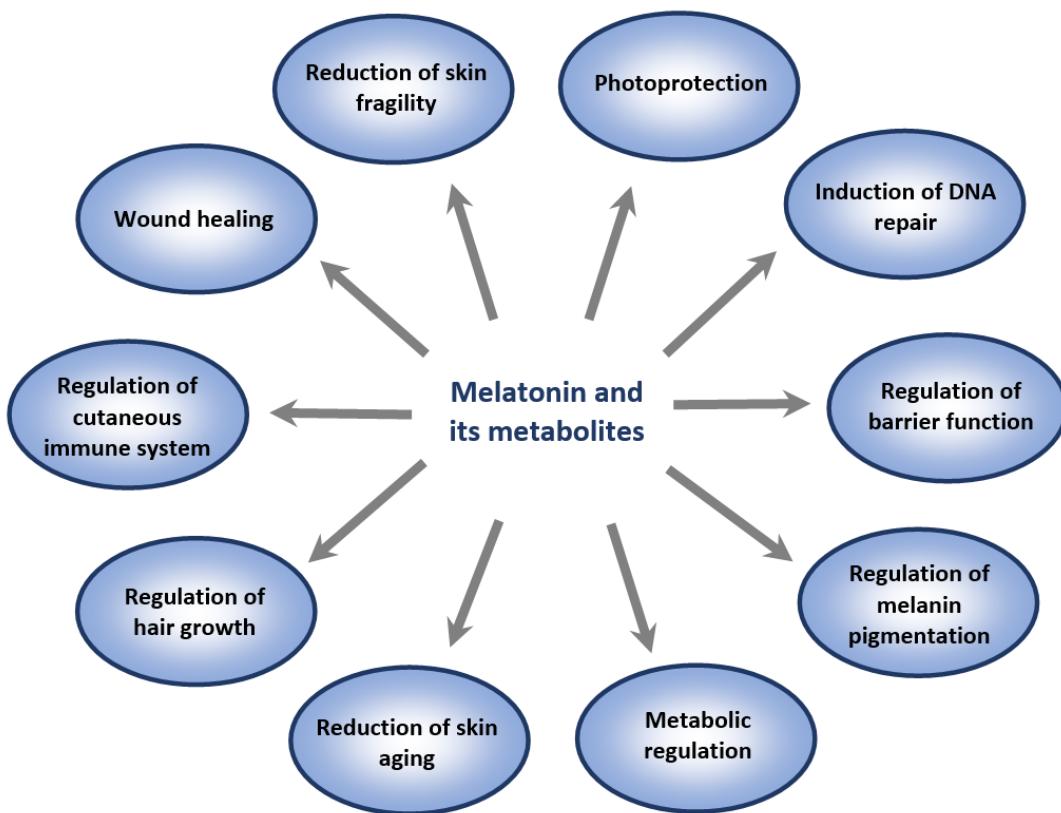
Melatonin in the skin undergoes rapid metabolism in vivo through either the indolic and kynuric pathways, with 6-hydroxymelatonin being a major metabolite [168,169]. Indeed, all metabolites of melatonin, including the final kynuric metabolites AFMK and AMK, are present in the epidermal cells and can potentially affect their mitochondrial functions [35,245]. Exposure of human skin to UVB can induce melatonin metabolism, leading to the generation of antioxidant metabolites AFMK and AMK in human keratinocytes [167,169]. The photo-induced melatonin metabolites further form a very potent anti-oxidative cascade. This cascade has been defined as the melatonergic anti-oxidative system (MAS) of the skin [13,167].

Melatonin and its metabolites are essential for the regulation of many skin functions, including cutaneous pigmentary [13,246], adnexal [244,247,248], barrier [23,40,168], and immune [173] functions. They also protect the skin against external and internal insults (Figure 2), and possess an oncostatic potential in melanoma cells [180,249]. Unlike melatonin, AMK does not inhibit tyrosinase activity and has no significant effect on melanogenesis [170]. Some but not all the phenotypic effects of melatonin are mediated via interaction with membrane bound G-protein-couple MT1 and MT2 receptors. MT1 has widespread localization, mainly in the epidermis (stratum granulosum, stratum spinosum, upper and inner root sheath of hair follicles) [19,22], whereas MT2 is often found in hair follicles and blood vessels, with lower expression or absence in the epidermal cells [13,244]. The expression of MT2 in hair follicles makes them a possible target for hair growth regulation by melatonin [248]. “MT3 receptors” have been also detected in keratinocytes, melanocytes, and fibroblasts; however, their role requires clarification [179]. Nuclear retinoic orphan receptor  $\alpha$  (ROR $\alpha$ ) has been found to be expressed in skin cells but it is not a receptor for melatonin, being identified as a receptor for sterols and secosteroids [250,251]. Melatonin regulation of mitochondrial functions is predominantly receptor-independent and requires high concentrations which can be achieved by an efficient on-site production and/or topical melatonin application.

#### 4.2. Role of Melatonin and Its Metabolites in Attenuation of Photoaging

Although skin has a well-equipped powerful antioxidant system to counteract oxidative stress, chronic exposure to UVR with its excessive ROS production can overcome the endogenous antioxidant defense of the skin, resulting in damage and premature aging in a process known as photoaging. Melatonin is one of the protective molecules biosynthesized at high concentrations in mitochondria of the skin cells to incapacitate ROS by electron donation and RNS by nitrosylation reactions [199,252,253]. Melatonin can prevent the formation of highly reactive free radicals by reducing the superoxide anion radical ( $O_2\bullet^-$ ) in a process referred to as radical avoidance [228,254]. The positional advantage of melatonin increases its ability to immediately scavenge the toxic free radicals formed in abundance in mitochondria, mainly by UVA but also by UVB irradiation [198,245]. Melatonin may additionally stimulate enzymes that are able to degrade the weakly reactive ROS [130,255]. It is important to note that the most harmful species (hydroxyl radicals and peroxy nitrite)

are not degraded by enzymes. They can only be removed by a direct highly efficient scavenger like melatonin [256–258]. The reaction of melatonin with hydroxyl radical initiates the formation of 2-OH-melatonin and 4-OH-melatonin, which are further metabolized to AFMK and by arylamine formamidase or catalase to AMK [196,202]. The effective toxic radical scavenging mediates the reduction of ROS-generated oxidative stress.

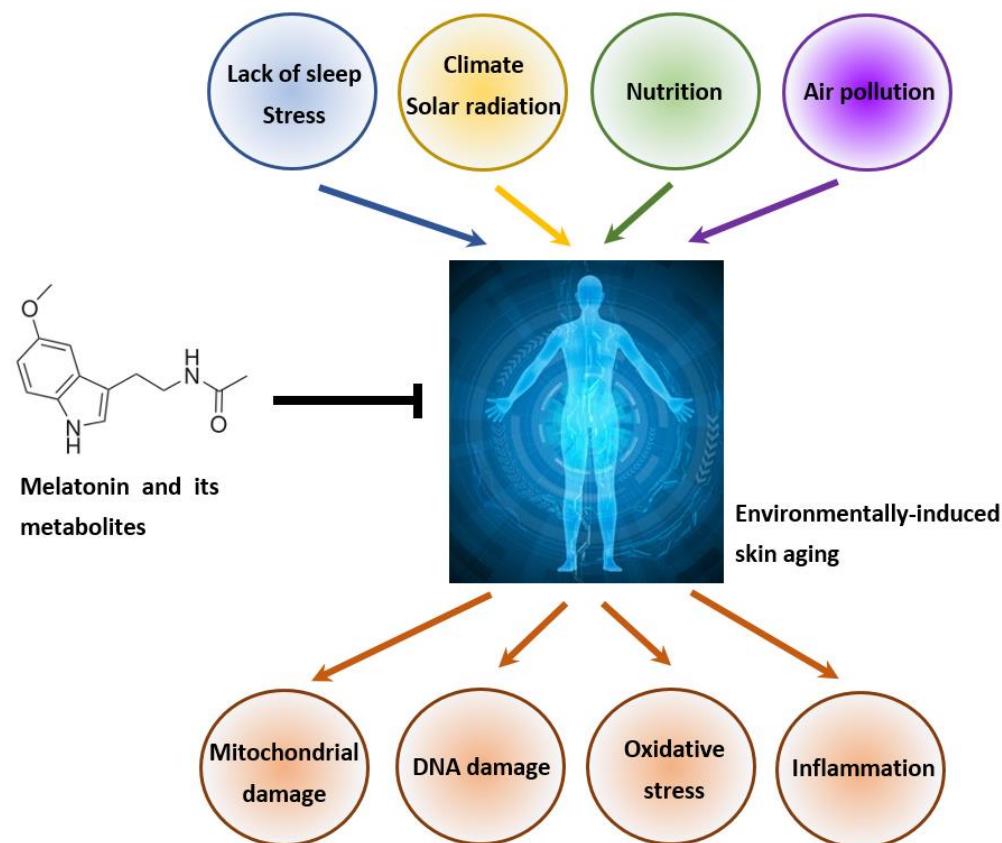


**Figure 2.** Overview of the pleiotropic effects of melatonin and its metabolites as major skin protectants. Melatonin directly or indirectly (via indolic and kynuric metabolites [35,169,245]) reduces deleterious intracellular changes including apoptotic disturbances or oxidative stress, while it maintains mitochondrial homeostasis.

In normal and diabetic human dermal fibroblasts, melatonin can stimulate SOD, catalase (CAT), and glutathione peroxidase (GPx), and promote glutathione (GSH) production [259]. Indeed, through activation of MT1/MT2, melatonin up-regulates the expression of antioxidant genes in irradiated cells [43,245,260].

The molecular mechanism of the indirect antioxidant action of melatonin with regard to the activation of phase-2 antioxidant enzymes has recently been established in UV-exposed human keratinocytes [254] and UVB-treated melanocytes [194]. It was found that melatonin stimulated NRF2 expression and induced its translocation to the nucleus, leading to enhanced gene expression of its target enzymes including  $\gamma$ -glutamylcysteine synthetase ( $\gamma$ -GCS), heme oxygenase-1 (HO-1), and NADPH:quinone dehydrogenase-1 (NQO1) [254]. The up-regulation by the melatonin/NRF2-dependent pathway supports the elevated antioxidant response of both keratinocytes and melanocytes against UVB-induced oxidative stress. [37,47,194]. Moreover, Nrf2 activation protects scalp hair growth against oxidative damage [261]. The ability of melatonin to attenuate UVA/UVB-induced alterations and to prevent the further photodamage has also been demonstrated in fibroblasts (Figure 3) [262,263]. In addition, it was found that melatonin can reduce the number of 8-hydroxy-2'-deoxyguanosine (8-OHdG)-positive cells, a marker of oxidative DNA damage [23,260]. Thus, by being a broad-spectrum antioxidant and amphiphilic molecule, melatonin can penetrate membranes and can also attenuate UVR-induced lipid peroxida-

tion, protein oxidation, and mitochondrial and DNA oxidative damage [23,35,37,41,47,264]. The other protective capability of melatonin is to counteract UVR-induced alterations in the mitochondrial ATP synthesis, plasma membrane potential, and pH in human keratinocytes [46,254,265].



**Figure 3.** The protective role of melatonin and its metabolites against premature skin aging. Cutaneous melatonin can prevent mitochondrial and DNA damage, oxidative stress, and inflammation caused by environmental factors such as stress, solar radiation, poor nutrition, or air pollution.

Importantly, melatonin possesses an advantage when compared with other antioxidants, since melatonin exerts not only a potent antioxidant capacity but most of its metabolites are antioxidants as well [168,202]. Whereas classical antioxidants (vitamins C and E) scavenge a single radical, melatonin's antioxidant cascade detoxifies many toxic radicals. Moreover, accumulating evidence supports the reciprocal interaction between melatonin and NAS in mitochondria that would amplify the detoxification process [169,178,245]. In addition, melatonin activates cytochrome C in mitochondria [159], which possibly mediates the formation of final kynurenic metabolites, which are even better free radical scavengers than melatonin itself [202,203,266]. AFMK and AMK generated non-enzymatically can accumulate in the skin [243]. However, AMK can disappear very quickly through oxidation and interactions with RNS [169].

Melatonin and its derivatives (6-hydroxymelatonin, NAS, AFMK, AMK, and 5-methoxytryptamine) have the capacity to protect keratinocytes and melanocytes against UVB-induced cell damage [23,37,194]. They not only reduce the formation of CPDs and 6–4 pyrimidine-pyrimidone photoproducts, but also induce the repair of DNA damaged by UVB. It has been demonstrated that the topical application of melatonin and AFMK can prevent DNA damage and apoptosis in human and porcine skin *ex vivo* [47]. Furthermore, the pre-incubation of full-thickness skin and normal human keratinocytes with melatonin suppressed the UVB-mediated inflammatory and apoptotic effect, as measured by heat shock protein 70 expression, expression of pro-inflammatory cytokines (IL-1 $\beta$ , IL-6), and

the pro-apoptotic protein caspase-3 [267]. The photoprotective potential of topically administrated melatonin has been shown in many clinical studies. Thus, treatment of the skin with exogenous melatonin before and after sun exposure attenuates UVR-induced erythema and oxidative stress [268]. The effect is greater when the cutaneous application of melatonin cream occurs prior to UVB exposure [269]. Sunscreens supplemented with melatonin could be used to prevent a skin photoaging and photocancerogenesis [270].

One potential anti-wrinkle mechanism of melatonin was studied by Sung-Hoon Kim's group [44]. They found that melatonin, by reducing ROS production, diminished MMP-1 expression and increased collagen XVII expression in HaCaT keratinocytes exposed to UVB. Furthermore, in the same study melatonin was shown to reduce the transepidermal water loss (TEWL) on the skin of hairless mice 8 weeks after UVB irradiation [44]. A clinical study also demonstrated a significant reduction of facial redness and wrinkles, and an improvement in the epidermal barrier function by using a night serum combination of melatonin, vitamin C (lipophilic and non-oxidizable form), and a polyphenol compound (bakuchiol) with retinol-like properties [271]. Additionally, the same night serum containing melatonin has been shown *in vitro* to increase filaggrin levels in keratinocytes, and collagen I and III in dermal fibroblasts, as well as to reduce the formation of apoptotic sunburn cells in UV-exposed skin *ex vivo*. [272].

The above findings confirm the clinical potential of melatonin as a broad-range photoprotector which can have a great impact on an attenuation of the premature skin aging and the improvement of the hallmarks of photoaged skin [14,47,273,274].

#### 4.3. Role of Melatonin and Its Metabolites in the Attenuation of Pollution-Induced Skin Aging

Environmental air pollutants promote mitochondrial dysfunction and oxidative damage due to excessive ROS generation, potentially resulting in prematurely aged skin and skin cancer [107,108]. Melatonin can restore the mitochondrial function and maintain the mitochondrial homeostasis [275]. It can reach the mitochondria by crossing the cell membranes, and it can also be synthesized in the mitochondrion. High concentrations of melatonin in mitochondria (endogenously produced or exogenously applied) can reduce oxidative damage, preserve mitochondrial respiration, limit mitochondria-related apoptosis, increase mitochondrial membrane potential and ATP production, and regulate mitochondrial biogenesis and mitophagy (removal of the damaged mitochondria).

It has been proposed that SIRT1, which can be stimulated by melatonin as well, plays a crucial role against pollutant-related premature skin aging. The up-regulation of SIRT1 could downregulate the MMP-1 and MMP-3 involved in the collagen breakdown, and it could decrease inflammation through inhibition of NF- $\kappa$ B signaling [127].

The use of creams containing melatonin, carnosine, and Helichrysum italicum extract on skin explants exposed to a mixture of polycyclic aromatic hydrocarbons and heavy metals leads to a reduction in skin damage and irritation [276]. The study demonstrated a significant decrease in pollution-activated transcription factor aryl hydrocarbon receptors (AhR) and type I collagen in melatonin-treated explants.

Therefore, a skin care product containing melatonin would be a real "weapon" in the prevention of premature skin aging caused by urban pollutants, heavy metals, and cigarette smoke [277].

#### 4.4. Possible Role of Melatonin in Modifying Natural Process of Skin Aging

The healthy aging of the skin is a complex multifactorial process that can be aggravated by an oxidative environment. With advancing age, the capacity of the skin to produce melatonin, the main direct- and indirect-acting antioxidant, diminishes, thus contributing to a decline in the endogenous protective MAS. The decreased levels of melatonin with age are accompanied with dysregulation of the circadian rhythm. Additionally, an age-dependent decrease in MT1 receptors is found in aged human fibroblasts [278]. The reduction in MT1 receptors along with a reduced melatonin level results in enhanced skin cellular damage and phenotypical signs of aging.

Therefore, the administration of exogenous melatonin would be a good anti-aging strategy. Orally supplemented melatonin appears in rather low levels in the blood due to prominent first-pass degradation in the liver, thus limiting skin access [14]. Topically applied melatonin may penetrate the stratum corneum and form a depot there due to its distinct lipophilic chemical structure [279]. The application of melatonin on the skin is a very good option for retarding the aging process and reducing the hallmarks of skin aging. The cutaneous application of melatonin is efficacious and safe way to improve the clinical signs of aging (wrinkles, TEWL and hydration, skin roughness, sagging, etc.) [186]. Clinically, it is better to apply melatonin at nighttime when the skin permeability is higher and because melatonin can mimic its endogenous production and effects.

With its pleotropic protective function of the skin, melatonin, with its proven beneficial anti-aging properties, could be considered as a therapeutic candidate for retarding skin aging and reversing cutaneous aging signs. Therefore, endogenous intracutaneous melatonin production, together with topically applied exogenous melatonin, is expected to provide the most potent defense system against cutaneous photodamage and multiple other pathological conditions that produce oxidative stress (e.g., in chronic skin inflammation, such as atopic dermatitis) [280]. Additionally, topical melatonin can be used for the treatment of androgenic alopecia in women [281].

## 5. Conclusions and Perspectives

Since the discovery of the strong antioxidant properties that melatonin possesses [137], a massive interest in terms of biological effects of melatonin in human and animal biology has evolved. It was shown that this indoleamine is an important bioregulator as well as a pluripotent and essential protective agent in many cells, tissues, and compartments of unicells, animals, and humans [22,216,282]. Melatonin exerts protective effects on cell physiology and tissue homeostasis, particularly in cutaneous cells exposed to UVR, which induces severe skin damage accompanied by oxidative stress or DNA damage. These intracellular disturbances are significantly counteracted or modulated by melatonin in the context of a complex intracutaneous melatonergic anti-oxidative system with UVR-enhanced or UVR-independent melatonin metabolites. Therefore, endogenous intracutaneous melatonin production, together with topically applied exogenous melatonin or its metabolites, may be expected to represent a promising anti-oxidative defense systems against skin aging. Indeed, more research on appropriate *in vitro*, *ex vivo*, and *in vivo* models must be performed to substantiate the above idea. For example, we need to learn whether melatonin and its derivatives can affect the expression of senescence markers in the skin. It would be fascinating to explore the possibility as to whether cutaneous melatonin production is altered during skin aging. Moreover, it is crucial to know whether the expression of functional MTs in cutaneous cell types is impaired in aged skin, which could eventually limit the anti-aging effects of any topically applied type of melatonin. In summary, the key question is whether melatonin can be exploited therapeutically as a protective agent, as “a skin survival factor” with anti-genotoxic capacities, or as “the neutralizer” of pathological changes including skin aging and cancerogenesis. The efficacy of topically applied melatonin and its derivatives needs further evaluation in future clinical trials. Another important point that needs further investigation is the use of nanotechnologies and nanomaterials for the topical delivery of melatonin and its metabolites for skin rejuvenation or to preserve the young skin phenotype.

**Author Contributions:** Conceptualization and writing—original draft preparation, G.B., K.K., R.M.S. and A.T.S.; supervision, A.T.S.; revision and approval the final version of the manuscript, G.B., R.M.S., T.-K.K., Z.J., M.B., K.S., R.J.R., K.K. and A.T.S. All authors have read and agreed to the published version of the manuscript.

**Funding:** Writing of this review was in part supported by grants as follows: NIH (R01AR073004-01 A1 [A.T.S.], R01AR071189-01A1 [A.T.S.] and R21AI149267-01A1 [A.T.S.]), VA merit award (I101BX004293-01A1 [A.T.S.]), the German Research Foundation (Deutsche Forschungsgemeinschaft [DFG]): KL2900/2-1 [K.K.], TR156/C05-246807620 [K.S.], SFB1009/B11-194468054 [K.S.], SFB1066/B06-213555243 [K.S.] and SFB1450/C06-431460824 [K.S.].

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Acknowledgments:** This review is dedicated to the memory of the Aaron B. Lerner, a discoverer of melatonin. The senior author (ATS) was trained by Aaron B. Lerner.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

- Slominski, A.T.; Zmijewski, M.A.; Skobowiat, C.; Zbytek, B.; Slominski, R.M.; Steketee, J.D. Sensing the environment: Regulation of local and global homeostasis by the skin's neuroendocrine system. *Adv. Anat. Embryol. Cell Biol.* **2012**, *212*, v-115.
- Slominski, A.; Wortsman, J. Neuroendocrinology of the skin. *Endocr. Rev.* **2000**, *21*, 457–487. [[CrossRef](#)] [[PubMed](#)]
- Slominski, A.T.; Zmijewski, M.A.; Zbytek, B.; Tobin, D.J.; Theoharides, T.C.; Rivier, J. Key role of CRF in the skin stress response system. *Endocr. Rev.* **2013**, *34*, 827–884. [[PubMed](#)]
- Slominski, A. Neuroendocrine activity of the melanocyte. *Exp. Dermatol.* **2009**, *18*, 760–763. [[CrossRef](#)]
- Slominski, A.; Zbytek, B.; Nikolakis, G.; Manna, P.R.; Skobowiat, C.; Zmijewski, M.; Li, W.; Janjetovic, Z.; Postlethwaite, A.; Zouboulis, C.C.; et al. Steroidogenesis in the skin: Implications for local immune functions. *J. Steroid Biochem. Mol. Biol.* **2013**, *137*, 107–123. [[CrossRef](#)]
- Slominski, A.; Wortsman, J.; Tuckey, R.C.; Paus, R. Differential expression of HPA axis homolog in the skin. *Mol. Cell. Endocrinol.* **2007**, *265–266*, 143–149. [[CrossRef](#)]
- Slominski, A.; Ermak, G.; Hwang, J.; Chakraborty, A.; Mazurkiewicz, J.E.; Mihm, M. Proopiomelanocortin, corticotropin releasing hormone and corticotropin releasing hormone receptor genes are expressed in human skin. *FEBS Lett.* **1995**, *374*, 113–116. [[CrossRef](#)]
- Slominski, A.; Wortsman, J.; Pisarchik, A.; Zbytek, B.; Linton, E.A.; Mazurkiewicz, J.E.; Wei, E.T. Cutaneous expression of corticotropin-releasing hormone (CRH), urocortin, and CRH receptors. *FASEB J.* **2001**, *15*, 1678–1693. [[CrossRef](#)]
- Slominski, A.; Szczesniewski, A.; Wortsman, J. Liquid chromatography-mass spectrometry detection of corticotropin-releasing hormone and proopiomelanocortin-derived peptides in human skin. *J. Clin. Endocrinol. Metab.* **2000**, *85*, 3582–3588. [[CrossRef](#)]
- Ito, N.; Ito, T.; Betterman, A.; Paus, R. The human hair bulb is a source and target of CRH. *J. Investigig. Dermatol.* **2004**, *122*, 235–237. [[CrossRef](#)]
- Slominski, A.; Wortsman, J.; Luger, T.; Paus, R.; Solomon, S. Corticotropin releasing hormone and proopiomelanocortin involvement in the cutaneous response to stress. *Physiol. Rev.* **2000**, *80*, 979–1020. [[CrossRef](#)] [[PubMed](#)]
- Kono, M.; Nagata, H.; Umemura, S.; Kawana, S.; Osamura, R.Y. In situ expression of corticotropin-releasing hormone (CRH) and proopiomelanocortin (POMC) genes in human skin. *FASEB J.* **2001**, *15*, 2297–2299. [[CrossRef](#)] [[PubMed](#)]
- Slominski, A.; Wortsman, J.; Tobin, D.J. The cutaneous serotoninergic/melatoninergic system: Securing a place under the sun. *FASEB J.* **2005**, *19*, 176–194. [[CrossRef](#)]
- Slominski, A.; Pisarchik, A.; Semak, I.; Sweatman, T.; Wortsman, J.; Szczesniewski, A.; Slugocki, G.; McNulty, J.; Kauser, S.; Tobin, D.J.; et al. Serotoninergic and melatoninergic systems are fully expressed in human skin. *FASEB J.* **2002**, *16*, 896–898. [[CrossRef](#)] [[PubMed](#)]
- Slominski, A.; Pisarchik, A.; Zbytek, B.; Tobin, D.J.; Kauser, S.; Wortsman, J. Functional activity of serotoninergic and melatoninergic systems expressed in the skin. *J. Cell. Physiol.* **2003**, *196*, 144–153. [[CrossRef](#)] [[PubMed](#)]
- Zouboulis, C.C. Human skin: An independent peripheral endocrine organ. *Horm. Res.* **2000**, *54*, 230–242. [[CrossRef](#)]
- Ndiaye, M.A.; Nihal, M.; Wood, G.S.; Ahmad, N. Skin, reactive oxygen species, and circadian clocks. *Antioxid. Redox Signal.* **2014**, *20*, 2982–2996. [[CrossRef](#)]
- Sandu, C.; Dumas, M.; Malan, A.; Sambakhe, D.; Marteau, C.; Nizard, C.; Schnebert, S.; Perrier, E.; Challet, E.; Pévet, P.; et al. Human skin keratinocytes, melanocytes, and fibroblasts contain distinct circadian clock machineries. *Cell. Mol. Life Sci.* **2012**, *69*, 3329–3339. [[CrossRef](#)]
- Slominski, R.M.; Reiter, R.J.; Schlabritz-Loutsevitch, N.; Ostrom, R.S.; Slominski, A.T. Melatonin membrane receptors in peripheral tissues: Distribution and functions. *Mol. Cell. Endocrinol.* **2012**, *351*, 152–166. [[CrossRef](#)]
- Yosipovitch, G.; Xiong, G.L.; Haus, E.; Sackett-Lundeen, L.; Ashkenazi, I.; Maibach, H.I. Time-dependent variations of the skin barrier function in humans: Transepidermal water loss, stratum corneum hydration, skin surface pH, and skin temperature. *J. Investigig. Dermatol.* **1998**, *110*, 20–23. [[CrossRef](#)]
- Matsui, M.S.; Pelle, E.; Dong, K.; Pernodet, N. Biological rhythms in the skin. *Int. J. Mol. Sci.* **2016**, *17*, 801. [[CrossRef](#)] [[PubMed](#)]
- Slominski, A.; Tobin, D.J.; Zmijewski, M.A.; Wortsman, J.; Paus, R. Melatonin in the skin: Synthesis, metabolism and functions. *Trends Endocrinol. Metab.* **2008**, *19*, 17–24. [[CrossRef](#)]

23. Slominski, A.T.; Kleszczyński, K.; Semak, I.; Janjetovic, Z.; Zmijewski, M.A.; Kim, T.K.; Slominski, R.M.; Reiter, R.J.; Fischer, T.W. Local melatonergic system as the protector of skin integrity. *Int. J. Mol. Sci.* **2014**, *15*, 17705–17732. [CrossRef] [PubMed]
24. Slominski, A.T.; Kim, T.K.; Li, W.; Yi, A.K.; Postlethwaite, A.; Tuckey, R.C. The role of CYP11A1 in the production of vitamin D metabolites and their role in the regulation of epidermal functions. *J. Steroid Biochem. Mol. Biol.* **2014**, *144 Pt A*, 28–39. [CrossRef] [PubMed]
25. Bikle, D.; Christakos, S. New aspects of vitamin D metabolism and action—Addressing the skin as source and target. *Nat. Rev. Endocrinol.* **2020**, *16*, 234–252. [CrossRef]
26. Slominski, A.T.; Chaiprasongsuk, A.; Janjetovic, Z.; Kim, T.K.; Stefan, J.; Slominski, R.M.; Hanumanthu, V.S.; Raman, C.; Qayyum, S.; Song, Y.; et al. Photoprotective properties of vitamin D and lumisterol hydroxyderivatives. *Cell Biochem. Biophys.* **2020**, *78*, 165–180. [CrossRef] [PubMed]
27. Brenner, M.; Hearing, V.J. The protective role of melanin against UV damage in human skin. *Photochem. Photobiol.* **2008**, *84*, 539–549. [CrossRef] [PubMed]
28. Melzer, D.; Pilling, L.C.; Ferrucci, L. The genetics of human ageing. *Nat. Rev. Genet.* **2020**, *21*, 88–101. [CrossRef]
29. Campisi, J.; Kapahi, P.; Lithgow, G.J.; Melov, S.; Newman, J.C.; Verdun, E. From discoveries in ageing research to therapeutics for healthy ageing. *Nature* **2019**, *571*, 183–192. [CrossRef]
30. Zanello, S.B.; Jackson, D.M.; Holick, M.F. Expression of the circadian clock genes clock and period1 in human skin. *J. Investig. Dermatol.* **2000**, *115*, 757–760. [CrossRef]
31. Desotelle, J.A.; Wilking, M.J.; Ahmad, N. The circadian control of skin and cutaneous photodamage. *Photochem. Photobiol.* **2012**, *88*, 1037–1047. [CrossRef] [PubMed]
32. Bocheva, G.; Slominski, R.M.; Slominski, A.T. The impact of vitamin D on skin aging. *Int. J. Mol. Sci.* **2021**, *22*, 9097. [CrossRef] [PubMed]
33. Reiter, R.J.; Fraschini, F. Endocrine aspects of the mammalian pineal gland. *Neuroendocrinology* **1969**, *5*, 219–255. [CrossRef]
34. Reiter, R.J.; Tan, D.X.; Fuentes-Broto, L. Melatonin: A multitasking molecule. *Prog. Brain Res.* **2010**, *181*, 127–151.
35. Slominski, A.T.; Hardeland, R.; Zmijewski, M.A.; Slominski, R.M.; Reiter, R.J.; Paus, R. Melatonin: A cutaneous perspective on its production, metabolism, and functions. *J. Investig. Dermatol.* **2018**, *138*, 490–499. [CrossRef]
36. Venegas, C.; Garcia, J.A.; Escames, G.; Ortiz, F.; Lopez, A.; Doerrier, C.; Garcia-Corzo, L.; Lopez, L.C.; Reiter, R.J.; Acuna-Castroviejo, D. Extrapineal melatonin: Analysis of its subcellular distribution and daily fluctuations. *J. Pineal Res.* **2012**, *52*, 217–227. [CrossRef]
37. Janjetovic, Z.; Nahmias, Z.P.; Hanna, S.; Jarrett, S.G.; Kim, T.K.; Reiter, R.J.; Slominski, A.T. Melatonin and its metabolites ameliorate ultraviolet B-induced damage in human epidermal keratinocytes. *J. Pineal Res.* **2014**, *57*, 90–102. [CrossRef]
38. Galano, A. On the direct scavenging activity of melatonin towards hydroxyl and a series of peroxy radicals. *Phys. Chem. Chem. Phys.* **2011**, *13*, 7178–7188. [CrossRef]
39. Tan, D.X.; Chen, L.D.; Poeggeler, B.; Manchester, L.C.; Reiter, R.J. Melatonin: A potent endogenous hydroxyl radical scavenger. *Endocrine* **1993**, *1*, 57–60.
40. Fischer, T.W.; Slominski, A.; Zmijewski, M.A.; Reiter, R.J.; Paus, R. Melatonin as a major skin protectant: From free radical scavenging to DNA damage repair. *Exp. Dermatol.* **2008**, *17*, 713–730. [CrossRef]
41. Reiter, R.J.; Mayo, J.C.; Tan, D.X.; Sainz, R.M.; Alatorre-Jimenez, M.; Qin, L. Melatonin as an antioxidant: Under promises but over delivers. *J. Pineal Res.* **2016**, *61*, 259–278. [CrossRef] [PubMed]
42. Reiter, R.J.; Tan, D.X.; Rosales-Corral, S.; Galano, A.; Zhou, X.J.; Xu, B. Mitochondria: Central organelles for melatonin's antioxidant and anti-aging actions. *Molecules* **2018**, *23*, 509. [CrossRef] [PubMed]
43. Rodriguez, C.; Mayo, J.C.; Sainz, R.M.; Antolín, I.; Herrera, F.; Martín, V.; Reiter, R.J. Regulation of antioxidant enzymes: A significant role for melatonin. *J. Pineal Res.* **2004**, *36*, 1–9. [CrossRef] [PubMed]
44. Park, E.; Lee, H.-J.; Lee, H.; Kim, J.-H.; Hwang, J.; Koo, J.; Kim, S.-H. The anti-wrinkle mechanism of melatonin in UVB treated HaCaT keratinocytes and hairless mice via inhibition of ROS and sonic hedgehog mediated inflammatory proteins. *Int. J. Mol. Sci.* **2018**, *19*, 1995. [CrossRef]
45. Fischer, T.W.; Zbytek, B.; Sayre, R.M.; Apostolov, E.O.; Basnakian, A.G.; Sweatman, T.W.; Wortsman, J.; Elsner, P.; Slominski, A. Melatonin increases survival of HaCaT keratinocytes by suppressing UV-induced apoptosis. *J. Pineal Res.* **2006**, *40*, 18–26. [CrossRef]
46. Fischer, T.W.; Zmijewski, M.A.; Wortsman, J.; Slominski, A. Melatonin maintains mitochondrial membrane potential and attenuates activation of initiator (casp-9) and effector caspases (casp-3/casp-7) and PARP in UVR-exposed HaCaT keratinocytes. *J. Pineal Res.* **2008**, *44*, 397–407. [CrossRef] [PubMed]
47. Skobowiat, C.; Brożyna, A.; Janjetovic, Z.; Jeayeng, S.; Oak, A.S.W.; Kim, T.; Panich, U.; Reiter, R.J.; Slominski, A.T. Melatonin and its derivatives counteract the ultraviolet B radiation-induced damage in human and porcine skin ex vivo. *J. Pineal Res.* **2018**, *65*, 12501. [CrossRef] [PubMed]
48. Milani, M.; Sparavigna, A. Antiaging efficacy of melatonin-based day and night creams: A randomized, split-face, assessor-blinded proof-of-concept trial. *Clin. Cosmet. Investig. Dermatol.* **2018**, *11*, 51–57. [CrossRef]
49. Passeron, T.; Krutmann, J.; Andersen, M.L.; Katta, R.; Zouboulis, C.C. Clinical and biological impact of the exposome on the skin. *J. Eur. Acad. Dermatol. Venereol.* **2020**, *34*, 4–25. [CrossRef]

50. Zouboulis, C.C.; Makrantonaki, E.; Nikolakis, G. When the skin is in the center of interest: An aging issue. *Clin. Dermatol.* **2019**, *37*, 296–305. [[CrossRef](#)]
51. Makrantonaki, E.; Bekou, V.; Zouboulis, C.C. Genetics and skin aging. *Dermatoendocrinol.* **2012**, *4*, 280–284. [[CrossRef](#)] [[PubMed](#)]
52. Bocheva, G.; Slominski, R.M.; Slominski, A.T. Neuroendocrine aspects of skin aging. *Int. J. Mol. Sci.* **2019**, *20*, 2798. [[CrossRef](#)] [[PubMed](#)]
53. Hertoghe, T. The “multiple hormone deficiency” theory of aging: Is human senescence caused mainly by multiple hormone deficiencies? *Ann. N. Y. Acad. Sci.* **2005**, *1057*, 448–465. [[CrossRef](#)] [[PubMed](#)]
54. Makrantonaki, E.; Schönknecht, P.; Hossini, A.M.; Kaiser, E.; Katsouli, M.M.; Adjaye, J.; Schröder, J.; Zouboulis, C.C. Skin and brain age together: The role of hormones in the ageing process. *Exp. Gerontol.* **2010**, *45*, 801–813. [[CrossRef](#)]
55. Pain, S.; Dezutter, C.; Reymermier, C.; Vogelgesang, B.; Delay, E.; André, V. Age-related changes in pro-opiomelanocortin (POMC) and related receptors in human epidermis. *Int. J. Cosmet. Sci.* **2010**, *32*, 266–275. [[CrossRef](#)] [[PubMed](#)]
56. Liu, F.; Hamer, M.A.; Deelen, J.; Lall, J.S.; Jacobs, L.; van Heemst, D.; Murray, P.G.; Wollstein, A.; de Craen, A.J.; Uh, H.W.; et al. The MC1R gene and youthful looks. *Curr. Biol.* **2016**, *26*, 1213–1220. [[CrossRef](#)] [[PubMed](#)]
57. Law, M.H.; Medland, S.E.; Zhu, G.; Yazar, S.; Viñuela, A.; Wallace, L.; Shekar, S.N.; Duffy, D.L.; Bataille, V.; Glass, D.; et al. Genome-wide association shows that pigmentation genes play a role in skin aging. *J. Investig. Dermatol.* **2017**, *137*, 1887–1894. [[CrossRef](#)]
58. Jacobs, L.C.; Hamer, M.A.; Gunn, D.A.; Deelen, J.; Lall, J.S.; van Heemst, D.; Uh, H.W.; Hofman, A.; Uitterlinden, A.G.; Griffiths, C.; et al. A Genome-wide association study identifies the skin color genes IRF4, MC1R, ASIP, and BNC2 influencing facial pigmented spots. *J. Investig. Dermatol.* **2015**, *135*, 1735–1742. [[CrossRef](#)]
59. Chen, Y.; André, M.; Adhikari, K.; Blin, M.; Bonfante, B.; Mendoza-Revilla, J.; Fuentes-Guajardo, M.; Palma, S.; Chacón-Duque, J.; Hurtado, M.; et al. A genome-wide association study identifies novel gene associations with facial skin wrinkling and mole count in Latin Americans. *Br. J. Dermatol.* **2021**, *185*, 988–998. [[CrossRef](#)]
60. Orioli, D.; Dellambra, E. Epigenetic regulation of skin cells in natural aging and premature aging diseases. *Cells* **2018**, *7*, 268. [[CrossRef](#)]
61. van Deursen, J.M. The role of senescent cells in ageing. *Nature* **2014**, *509*, 439–446. [[CrossRef](#)] [[PubMed](#)]
62. Wlaschek, M.; Maity, P.; Makrantonaki, E.; Scharffetter-Kochanek, K. Connective tissue and fibroblast senescence in skin aging. *J. Investig. Dermatol.* **2021**, *141*, 985–992. [[CrossRef](#)] [[PubMed](#)]
63. Brun, C.; Jean-Louis, F.; Oddos, T.; Bagot, M.; Bensussan, A.; Michel, L. Phenotypic and functional changes in dermal primary fibroblasts isolated from intrinsically aged human skin. *Exp. Dermatol.* **2016**, *25*, 113–119. [[CrossRef](#)] [[PubMed](#)]
64. Wang, A.S.; Dreesen, O. Biomarkers of cellular senescence and skin aging. *Front. Genet.* **2018**, *9*, 247. [[CrossRef](#)] [[PubMed](#)]
65. Ghosh, K.; Capell, B.C. The senescence-associated secretory phenotype: Critical effector in skin cancer and aging. *J. Investig. Dermatol.* **2016**, *136*, 2133–2139. [[CrossRef](#)]
66. Fitsiou, E.; Pulido, T.; Campisi, J.; Alimirah, F.; Demaria, M. Cellular senescence and the senescence-associated secretory phenotype as drivers of skin photoaging. *J. Investig. Dermatol.* **2021**, *141*, 1119–1126. [[CrossRef](#)]
67. Lee, Y.I.; Choi, S.; Roh, W.S.; Lee, J.H.; Kim, T.G. Cellular senescence and inflammaging in the skin microenvironment. *Int. J. Mol. Sci.* **2021**, *22*, 3849. [[CrossRef](#)]
68. Nguyen, A.V.; Soulka, A.M. The dynamics of the skin’s immune system. *Int. J. Mol. Sci.* **2019**, *20*, 1811. [[CrossRef](#)]
69. De Martinis, M.; Sirufo, M.M.; Ginaldi, L. Allergy and aging: An old/new emerging health issue. *Aging Dis.* **2017**, *8*, 162–175. [[CrossRef](#)]
70. Bocheva, G.S.; Slominski, R.M.; Slominski, A.T. Immunological aspects of skin aging in atopic dermatitis. *Int. J. Mol. Sci.* **2021**, *22*, 5729. [[CrossRef](#)]
71. Bowman, A.; Birch-Machin, M.A. Age-dependent decrease of mitochondrial complex II activity in human skin fibroblasts. *J. Investig. Dermatol.* **2016**, *136*, 912–919. [[CrossRef](#)] [[PubMed](#)]
72. DeBalsi, K.L.; Hoff, K.E.; Copeland, W.C. Role of the mitochondrial DNA replication machinery in mitochondrial DNA mutagenesis, aging and age-related diseases. *Ageing Res. Rev.* **2017**, *33*, 89–104. [[CrossRef](#)]
73. Sun, N.; Youle, R.J.; Finkel, T. The mitochondrial basis of aging. *Mol. Cell.* **2016**, *61*, 654–666. [[CrossRef](#)] [[PubMed](#)]
74. Zhang, R.; Wang, Y.; Ye, K.; Picard, M.; Gu, Z. Independent impacts of aging on mitochondrial DNA quantity and quality in humans. *BMC Genom.* **2017**, *18*, 890. [[CrossRef](#)] [[PubMed](#)]
75. Quan, C.; Cho, M.K.; Perry, D.; Quan, T. Age-associated reduction of cell spreading induces mitochondrial DNA common deletion by oxidative stress in human skin dermal fibroblasts: Implication for human skin connective tissue aging. *J. Biomed. Sci.* **2015**, *28*, 222, 62. [[CrossRef](#)] [[PubMed](#)]
76. Slominski, A.T.; Zmijewski, M.A.; Plonka, P.M.; Szaflarski, J.P.; Paus, R. How UV light touches the brain and endocrine system through skin, and why. *Endocrinology* **2018**, *159*, 1992–2007. [[CrossRef](#)]
77. Rittié, L.; Fisher, G.J. UV-light-induced signal cascades and skin aging. *Ageing Res. Rev.* **2002**, *1*, 705–720. [[CrossRef](#)]
78. Green, A.C.; Wallingford, S.C.; McBride, P. Childhood exposure to ultraviolet radiation and harmful skin effects: Epidemiological evidence. *Prog. Biophys. Mol. Biol.* **2011**, *107*, 349–355. [[CrossRef](#)]
79. Parrado, C.; Mercado-Saenz, S.; Perez-Davo, A.; Gilaberte, Y.; Gonzalez, S.; Juarranz, A. Environmental stressors on skin aging. Mechanistic insights. *Front. Pharmacol.* **2019**, *10*, 759. [[CrossRef](#)]

80. Park, S.Y.; Byun, E.J.; Lee, J.D.; Kim, S.; Kim, H.S. Air pollution, autophagy, and skin aging impact of particulate matter (pm10) on human dermal fibroblasts. *Int. J. Mol. Sci.* **2018**, *19*, 2727. [CrossRef]
81. Damevska, K.; Nikolovska, S.; Kazandjieva, J.; Kotevska, B.; Bocheva, G. Skin and pollution. In *Advances in Integrative Dermatology*; França, K., Lotti, T., Eds.; John Wiley & Sons Ltd.: Hoboken, NJ, USA, 2019; pp. 379–392.
82. Koohgoli, R.; Hudson, L.; Naidoo, K.; Wilkinson, S.; Chavan, B.; Birch-Machin, M.A. Bad air gets under your skin. *Exp. Dermatol.* **2017**, *26*, 384–387. [CrossRef]
83. Naidoo, K.; Birch-Machin, M.A. Oxidative stress and ageing: The influence of environmental pollution, sunlight and diet on skin. *Cosmetics* **2017**, *4*, 4. [CrossRef]
84. Kammerer, A.; Luiten, R.M. Oxidation events and skin aging. *Ageing Res. Rev.* **2015**, *21*, 16–29. [CrossRef]
85. Imokawa, G.; Ishida, K. Biological mechanisms underlying the ultraviolet radiation-induced formation of skin wrinkling and sagging I: Reduced skin elasticity, highly associated with enhanced dermal elastase activity, triggers wrinkling and sagging. *Int. J. Mol. Sci.* **2015**, *16*, 7753–7775. [CrossRef]
86. Hüls, A.; Sugiri, D.; Fuks, K.; Krutmann, J.; Schikowski, T. Lentigine formation in caucasian women—interaction between particulate matter and solar UVR. *J. Investig. Dermatol.* **2018**, *139*, 974–976. [CrossRef] [PubMed]
87. Krutmann, J.; Bouloc, A.; Sore, G.; Bernard, B.A.; Passeron, T. The skin aging exposome. *J. Dermatol. Sci.* **2017**, *85*, 152–161. [CrossRef]
88. Burns, E.M.; Ahmed, H.; Isedeh, P.N.; Kohli, I.; Van Der Pol, W.; Shaheen, A.; Muzaffar, A.F.; Al-Sadek, C.; Foy, T.M.; Abdelgawad, M.S.; et al. Ultraviolet radiation, both UVA and UVB, influences the composition of the skin microbiome. *Exp. Dermatol.* **2019**, *28*, 136–141. [CrossRef]
89. Campisi, J. Aging, cellular senescence, and cancer. *Annu. Rev. Physiol.* **2013**, *75*, 685–705. [CrossRef]
90. Hussein, M.R. Ultraviolet radiation and skin cancer: Molecular mechanisms. *J. Cutan. Pathol.* **2005**, *32*, 191–205. [CrossRef]
91. Battie, C.; Jitsukawa, S.; Bernerd, F.; Del Bino, S.; Marionnet, C.; Verschoore, M. New insights in photoaging, UVA induced damage and skin types. *Exp. Dermatol.* **2014**, *23*, 7–12. [CrossRef]
92. Marionnet, C.; Tricaud, C.; Bernerd, F. Exposure to non-extreme solar UV daylight: Spectral characterization, effects on skin and photoprotection. *Int. J. Mol. Sci.* **2014**, *16*, 68–90. [CrossRef] [PubMed]
93. Pittayapruet, P.; Meephansan, J.; Prapapan, O.; Komine, M.; Ohtsuki, M. Role of matrix metalloproteinases in photoaging and photocarcinogenesis. *Int. J. Mol. Sci.* **2016**, *17*, 868. [CrossRef] [PubMed]
94. Chung, J.H.; Seo, J.Y.; Choi, H.R.; Lee, M.K.; Youn, C.S.; Rhie, G.; Cho, K.H.; Kim, K.H.; Park, K.C.; Eun, H.C. Modulation of skin collagen metabolism in aged and photoaged human skin in vivo. *J. Investig. Dermatol.* **2001**, *117*, 1218–1224. [CrossRef] [PubMed]
95. Tigges, J.; Krutmann, J.; Fritzsche, E.; Haendeler, J.; Schaal, H.; Fischer, J.W.; Kalfalah, F.; Reinke, H.; Reifenberger, G.; Stühler, K.; et al. The hallmarks of fibroblast ageing. *Mech. Ageing Dev.* **2014**, *138*, 26–44. [CrossRef]
96. Birch-Machin, M.A.; Bowman, A. Oxidative stress and ageing. *Br. J. Dermatol.* **2016**, *175*, 26–29. [CrossRef]
97. Baier, J.; Maisch, T.; Maier, M.; Landthaler, M.; Bäumler, W. Direct detection of singlet oxygen generated by UVA irradiation in human cells and skin. *J. Investig. Dermatol.* **2007**, *127*, 1498–1506. [CrossRef]
98. Holick, M.F.; Clark, M.B. The photobiogenesis and metabolism of vitamin D. *Fed. Proc.* **1978**, *37*, 2567–2574.
99. Setlow, R.B.; Carrier, W.L. Pyrimidine dimers in ultraviolet-irradiated DNA's. *J. Mol. Biol.* **1966**, *17*, 237–254. [CrossRef]
100. Brash, D.E. UV signature mutations. *Photochem. Photobiol.* **2015**, *91*, 15–26. [CrossRef]
101. Decraene, D.; Agostinis, P.; Pupe, A.; de Haes, P.; Garmyn, M. Acute response of human skin to solar radiation: Regulation and function of the p53 protein. *J. Photochem. Photobiol. B.* **2001**, *63*, 78–83. [CrossRef]
102. El-Domyati, M.B.; Attia, S.; Saleh, F.; Galaria, N.; Ahmad, H.; Gasparro, F.P.; Uitto, J. Expression of p53 in normal sun-exposed and protected skin (type IV-V) in different decades of age. *Acta Derm. Venereol.* **2003**, *83*, 98–104. [CrossRef] [PubMed]
103. Mancebo, S.E.; Wang, S.Q. Recognizing the impact of ambient air pollution on skin health. *J. Eur. Acad. Dermatol. Venereol.* **2015**, *29*, 2326–2332. [CrossRef] [PubMed]
104. Mousavi, S.E.; Amini, H.; Heydarpour, P.; Amini Chermahini, F.; Godderis, L. Air pollution, environmental chemicals, and smoking may trigger vitamin D deficiency: Evidence and potential mechanisms. *Environ. Int.* **2019**, *122*, 67–90. [CrossRef] [PubMed]
105. Peng, F.; Xue, C.H.; Hwang, S.K.; Li, W.H.; Chen, Z.; Zhang, J.Z. Exposure to fine particulate matter associated with senile lentigo in Chinese women: A cross-sectional study. *J. Eur. Acad. Dermatol. Venereol.* **2017**, *31*, 355–360. [CrossRef] [PubMed]
106. Burke, K.E. Mechanisms of aging and development—a new understanding of environmental damage to the skin and prevention with topical antioxidants. *Mech. Ageing Dev.* **2018**, *172*, 123–130. [CrossRef]
107. Piao, M.J.; Kang, K.A.; Zhen, A.X.; Fernando, P.D.S.M.; Ahn, M.J.; Koh, Y.S.; Kang, H.K.; Yi, J.M.; Choi, Y.H.; Hyun, J.W. Particulate matter 2.5 mediates cutaneous cellular injury by inducing mitochondria-associated endoplasmic reticulum stress: Protective effects of ginsenoside Rb1. *Antioxidants* **2019**, *8*, 383. [CrossRef]
108. Soeur, J.; Belaïdi, J.P.; Chollet, C.; Denat, L.; Dimitrov, A.; Jones, C.; Perez, P.; Zanini, M.; Zobiri, O.; Mezzache, S.; et al. Photopollution stress in skin: Traces of pollutants (PAH and particulate matter) impair redox homeostasis in keratinocytes exposed to UVA1. *J. Dermatol. Sci.* **2017**, *86*, 162–169. [CrossRef]
109. Kandola, K.; Bowman, A.; Birch-Machin, M.A. Oxidative stress—a key emerging impact factor in health, ageing, lifestyle and aesthetics. *Int. J. Cosmet. Sci.* **2015**, *37*, 1–8. [CrossRef]

110. Naidoo, K.; Hanna, R.; Birch-Machin, M.A. What is the role of mitochondrial dysfunction in skin photoaging? *Exp. Dermatol.* **2018**, *27*, 124–128. [[CrossRef](#)]
111. Mullenders, L.H.F. Solar UV damage to cellular DNA: From mechanisms to biological effects. *Photochem. Photobiol. Sci.* **2018**, *17*, 1842–1852. [[CrossRef](#)]
112. Kudryavtseva, A.V.; Krasnov, G.S.; Dmitriev, A.A.; Alekseev, B.Y.; Kardymon, O.L.; Sadritdinova, A.F.; Fedorova, M.S.; Pokrovsky, A.V.; Melnikova, N.V.; Kaprin, A.D.; et al. Mitochondrial dysfunction and oxidative stress in aging and cancer. *Oncotarget* **2016**, *7*, 44879–44905. [[CrossRef](#)] [[PubMed](#)]
113. Singh, B.; Schoeb, T.R.; Bajpai, P.; Slominski, A.; Singh, K.K. Reversing wrinkled skin and hair loss in mice by restoring mitochondrial function. *Cell Death Dis.* **2018**, *9*, 735. [[CrossRef](#)] [[PubMed](#)]
114. Birch-Machin, M.A.; Russell, E.V.; Latimer, J.A. Mitochondrial DNA damage as a biomarker for ultraviolet radiation exposure and oxidative stress. *Br. J. Dermatol.* **2013**, *169*, 9–14. [[CrossRef](#)] [[PubMed](#)]
115. Chung, J.H.; Kang, S.; Varani, J.; Lin, J.; Fisher, G.J.; Voorhees, J.J. Decreased extracellular-signal-regulated kinase and increased stress-activated MAP kinase activities in aged human skin in vivo. *J. Investigig. Dermatol.* **2000**, *115*, 177–182. [[CrossRef](#)] [[PubMed](#)]
116. Schäfer, M.; Werner, S. Nrf2—A regulator of keratinocyte redox signaling. *Free Radic. Biol. Med.* **2015**, *88*, 243–252. [[CrossRef](#)]
117. Chaiprasongsuk, A.; Lohakul, J.; Soonthrapa, K.; Sampattavanich, S.; Akarasereenont, P.; Panich, U. Activation of Nrf2 reduces UVA-mediated MMP-1 upregulation via MAPK/AP-1 signaling cascades: The photoprotective effects of sulforaphane and hispidulin. *J. Pharmacol. Exp. Ther.* **2017**, *360*, 388–398. [[CrossRef](#)]
118. Quan, T.; He, T.; Kang, S.; Voorhees, J.J.; Fisher, G.J. Solar ultraviolet irradiation reduces collagen in photoaged human skin by blocking transforming growth factor-beta type II receptor/Smad signaling. *Am. J. Pathol.* **2004**, *165*, 741–751. [[CrossRef](#)]
119. Haustead, D.J.; Stevenson, A.; Saxena, V.; Marriage, F.; Firth, M.; Silla, R.; Martin, L.; Adcroft, K.F.; Rea, S.; Day, P.J.; et al. Transcriptome analysis of human ageing in male skin shows mid-life period of variability and central role of NF-κB. *Sci. Rep.* **2016**, *6*, 26846. [[CrossRef](#)]
120. Ansary, T.M.; Hossain, M.R.; Kamiya, K.; Komine, M.; Ohtsuki, M. Inflammatory molecules associated with ultraviolet radiation-mediated skin aging. *Int. J. Mol. Sci.* **2021**, *22*, 3974. [[CrossRef](#)]
121. Zhang, B.; Xu, J.; Quan, Z.; Qian, M.; Liu, W.; Zheng, W.; Yin, F.; Du, J.; Zhi, Y.; Song, N. Klotho protein protects human keratinocytes from UVB-induced damage possibly by reducing expression and nuclear translocation of NF-κB. *Med. Sci. Monit.* **2018**, *24*, 8583–8591. [[CrossRef](#)]
122. Bi, F.; Liu, W.; Wu, Z.; Ji, C.; Chang, C. Antiaging factor klotho retards the progress of intervertebral disc degeneration through the toll-like receptor 4-NF-κB pathway. *Int. J. Cell. Biol.* **2020**, *2020*, 8319516. [[CrossRef](#)]
123. Battino, M.; Giampieri, F.; Pistollato, F.; Sureda, A.; de Oliveira, M.R.; Pittalà, V.; Fallarino, F.; Nabavi, S.F.; Atanasov, A.G.; Nabavi, S.M. Nrf2 as regulator of innate immunity: A molecular Swiss army knife! *Biotechnol. Adv.* **2018**, *36*, 358–370. [[CrossRef](#)]
124. Hirota, A.; Kawachi, Y.; Yamamoto, M.; Koga, T.; Hamada, K.; Otsuka, F. Acceleration of UVB-induced photoageing in nrf2 gene-deficient mice. *Exp. Dermatol.* **2011**, *20*, 664–668. [[CrossRef](#)]
125. Chaiprasongsuk, A.; Janjetovic, Z.; Kim, T.K.; Jarrett, S.G.; D’Orazio, J.A.; Holick, M.F.; Tang, E.K.Y.; Tuckey, R.C.; Panich, U.; Li, W.; et al. Protective effects of novel derivatives of vitamin D<sub>3</sub> and lumisterol against UVB-induced damage in human keratinocytes involve activation of Nrf2 and p53 defense mechanisms. *Redox Biol.* **2019**, *24*, 101206. [[CrossRef](#)]
126. Rojo de la Vega, M.; Krajisnik, A.; Zhang, D.D.; Wondrak, G.T. Targeting NRF2 for improved skin barrier function and photoprotection: Focus on the achiole-derived apocarotenoid bixin. *Nutrients* **2017**, *9*, 1371. [[CrossRef](#)]
127. Bielach-Bazyluk, A.; Zbroch, E.; Mysliwiec, H.; Rydzewska-Rosolowska, A.; Kakareko, K.; Flisiak, I.; Hryszko, T. Sirtuin 1 and skin: Implications in intrinsic and extrinsic aging—A systematic review. *Cells* **2021**, *10*, 813. [[CrossRef](#)]
128. Kalfalah, F.; Sobek, S.; Bornholz, B.; Götz-Rösch, C.; Tigges, J.; Fritzsche, E.; Krutmann, J.; Köhrer, K.; Deenen, R.; Ohse, S.; et al. Inadequate mito-biogenesis in primary dermal fibroblasts from old humans is associated with impairment of PGC1A-independent stimulation. *Exp. Gerontol.* **2014**, *56*, 59–68. [[CrossRef](#)]
129. Ding, S.; Chen, J.; Zeng, Q.; Lu, J.; Tan, L.; Guo, A.; Kang, J.; Yang, S.; Xiang, Y.; Zuo, C.; et al. Chronic sun exposure is associated with distinct histone acetylation changes in human skin. *Br. J. Dermatol.* **2018**, *179*, 110–117. [[CrossRef](#)]
130. Manchester, L.C.; Coto-Montes, A.; Boga, J.A.; Andersen, L.P.; Zhou, Z.; Galano, A.; Vriend, J.; Tan, D.X.; Reiter, R.J. Melatonin: An ancient molecule that makes oxygen metabolically tolerable. *J. Pineal Res.* **2015**, *59*, 403–419. [[CrossRef](#)]
131. Zhao, D.; Yu, Y.; Shen, Y.; Liu, Q.; Zhao, Z.; Sharma, R.; Reiter, R.J. Melatonin synthesis and function: Evolutionary history in animals and plants. *Front. Endocrinol.* **2019**, *10*, 249. [[CrossRef](#)]
132. Reiter, R.J.; Tan, D.X.; Burkhardt, S.; Manchester, L.C. Melatonin in plants. *Nutr. Rev.* **2001**, *59*, 286–290. [[CrossRef](#)] [[PubMed](#)]
133. Back, K.; Tan, D.X.; Reiter, R.J. Melatonin biosynthesis in plants: Multiple pathways catalyze tryptophan to melatonin in the cytoplasm or chloroplasts. *J. Pineal Res.* **2016**, *61*, 426–437. [[CrossRef](#)] [[PubMed](#)]
134. Tan, D.X.; Zheng, X.; Kong, J.; Manchester, L.C.; Hardeland, R.; Kim, S.J.; Xu, X.; Reiter, R.J. Fundamental issues related to the origin of melatonin and melatonin isomers during evolution: Relation to their biological functions. *Int. J. Mol. Sci.* **2014**, *15*, 15858–15890. [[CrossRef](#)] [[PubMed](#)]
135. Hardeland, R.; Fuhrberg, B. Ubiquitous melatonin—presence and effects in unicells, plants and animals. *Trends Comp. Biochem. Physiol.* **1996**, *2*, 25–45.
136. Paredes, S.D.; Korkmaz, A.; Manchester, L.C.; Tan, D.X.; Reiter, R.J. Phytomelatonin: A review. *J. Exp. Bot.* **2009**, *60*, 57–69. [[CrossRef](#)]

137. Lerner, A.B.; Case, J.D.; Takahashi, Y.; Lee, T.H.; Mori, W. Isolation of melatonin, the pineal gland factor that lightens melanocytes. *J. Am. Chem. Soc.* **1958**, *80*, 2587. [[CrossRef](#)]
138. Lerner, A.B.; Lee, T.H.; Wright, M.R.; McGuire, J.S. The mechanism of action of the melanocyte stimulating hormones. *Eur. J. Endocrinol.* **1960**, *34*, S73. [[CrossRef](#)]
139. Reiter, R.J. Melatonin: The chemical expression of darkness. *Mol. Cell. Endocrinol.* **1991**, *79*, C153–C158. [[CrossRef](#)]
140. Pablos, M.I.; Reiter, R.J.; Ortiz, G.G.; Guerrero, J.M.; Agapito, M.T.; Chuang, J.I.; Sewerynek, E. Rhythms of glutathione peroxidase and glutathione reductase in brain of chick and their inhibition by light. *Neurochem. Int.* **1998**, *32*, 69–75. [[CrossRef](#)]
141. Reiter, R.J.; Tan, D.X.; Rosales-Corral, S.; Manchester, L.C. The universal nature, unequal distribution and antioxidant functions of melatonin and its derivatives. *Mini-Rev. Med. Chem.* **2013**, *13*, 373–384.
142. Acuña-Castroviejo, D.; Escames, G.; Venegas, C.; Díaz-Casado, M.E.; Lima-Cabello, E.; López, L.C.; Rosales-Corral, S.; Tan, D.X.; Reiter, R.J. Extrpineal melatonin: Sources, regulation, and potential functions. *Cell. Mol. Life Sci.* **2014**, *71*, 2997–3025. [[CrossRef](#)] [[PubMed](#)]
143. Tan, D.X.; Manchester, L.C.; Reiter, R.J.; Qi, W.B.; Zhang, M.; Weintraub, S.T.; Cabrera, J.; Sainz, R.M.; Mayo, J.C. Identification of highly elevated levels of melatonin in bone marrow: Its origin and significance. *Biochim. Biophys. Acta* **1999**, *1472*, 206–214. [[CrossRef](#)]
144. Iuvone, P.M.; Tosini, G.; Pozdnyev, N.; Haque, R.; Klein, D.C.; Chaurasia, S.S. Circadian clocks, clock networks, arylalkylamine N-acetyltransferase, and melatonin in the retina. *Prog. Retin. Eye Res.* **2005**, *24*, 433–456. [[CrossRef](#)] [[PubMed](#)]
145. Naranjo, M.C.; Guerrero, J.M.; Rubio, A.; Lardone, P.J.; Carrillo-Vico, A.; Carrascosa-Salmoral, M.P.; Jiménez-Jorge, S.; Arellano, M.V.; Leal-Noval, S.R.; Leal, M.; et al. Melatonin biosynthesis in the thymus of humans and rats. *Cell. Mol. Life Sci.* **2007**, *64*, 781–790. [[CrossRef](#)] [[PubMed](#)]
146. Wang, J.; Zhu, T.; Ma, X.; Wang, Y.; Liu, J.; Li, G.; Liu, Y.; Ji, P.; Zhang, Z.; Zhang, L.; et al. Melatonergic systems of AANAT, melatonin, and its receptor MT2 in the corpus luteum are essential for reproductive success in mammals. *Biol. Reprod.* **2021**, *104*, 430–444. [[CrossRef](#)]
147. Kema, I.P.; de Vries, E.G.; Muskiet, F.A. Clinical chemistry of serotonin and metabolites. *J. Chromatogr. B Biomed. Sci. Appl.* **2000**, *747*, 33–48. [[CrossRef](#)]
148. Mockus, S.M.; Vrana, K.E. Advances in the molecular characterization of tryptophan hydroxylase. *J. Mol. Neurosci.* **1998**, *10*, 163–179. [[CrossRef](#)]
149. Tidemand, K.D.; Peters, G.H.; Harris, P.; Stensgaard, E.; Christensen, H.E.M. Isoform-specific substrate inhibition mechanism of human tryptophan hydroxylase. *Biochemistry* **2017**, *56*, 6155–6164. [[CrossRef](#)]
150. McIsaac, W.M.; Page, I.H. The metabolism of serotonin (5-hydroxytryptamine). *J. Biol. Chem.* **1959**, *234*, 858–864. [[CrossRef](#)]
151. Lovenberg, W.; Jequier, E.; Sjoerdsma, A. Tryptophan hydroxylation: Measurement in pineal gland, brainstem, and carcinoid tumor. *Science* **1967**, *155*, 217–219. [[CrossRef](#)]
152. Gaudet, S.J.; Slominski, A.; Etminan, M.; Pruski, D.; Paus, R.; Namboodiri, M. Identification and characterization of two isozymic forms of arylamine N-acetyltransferase in Syrian hamster skin. *J. Investig. Dermatol.* **1993**, *101*, 660–665. [[CrossRef](#)] [[PubMed](#)]
153. Liu, T.; Borjigin, J. N-acetyltransferase is not the rate-limiting enzyme of melatonin synthesis at night. *J. Pineal Res.* **2005**, *39*, 91–96. [[CrossRef](#)] [[PubMed](#)]
154. Slominski, A.; Pisarchik, A.; Semak, I.; Sweatman, T.; Wortsman, J. Characterization of the serotonergic system in the C57BL/6 mouse skin. *Eur. J. Biochem.* **2003**, *270*, 3335–3344. [[CrossRef](#)] [[PubMed](#)]
155. Slominski, A.; Pisarchik, A.; Semak, I.; Sweatman, T.; Szczesniewski, A.; Wortsman, J. Serotonergic system in hamster skin. *J. Investig. Dermatol.* **2002**, *119*, 934–942. [[CrossRef](#)] [[PubMed](#)]
156. Semak, I.; Korik, E.; Naumova, M.; Wortsman, J.; Slominski, A. Serotonin metabolism in rat skin: Characterization by liquid chromatography-mass spectrometry. *Arch. Biochem. Biophys.* **2004**, *421*, 61–66. [[CrossRef](#)] [[PubMed](#)]
157. Axelrod, J.; Weissbach, H. Enzymatic O-methylation of N-acetylserotonin to melatonin. *Science* **1960**, *131*, 1312. [[CrossRef](#)]
158. Mannino, G.; Pernici, C.; Serio, G.; Gentile, C.; Berte, C.M. Melatonin and phytomelatonin: Chemistry, biosynthesis, metabolism, distribution and bioactivity in plants and animals-An overview. *Int. J. Mol. Sci.* **2021**, *22*, 9996. [[CrossRef](#)]
159. Ma, X.; Idle, J.R.; Krausz, K.W.; Gonzalez, F.J. Metabolism of melatonin by human cytochromes p450. *Drug Metab. Dispos.* **2005**, *33*, 489–494. [[CrossRef](#)]
160. Reiter, R.J. Pineal melatonin: Cell biology of its synthesis and of its physiological interactions. *Endocr. Rev.* **1991**, *12*, 151–180. [[CrossRef](#)]
161. Facciolá, G.; Hidestrand, M.; von Bahr, C.; Tybring, G. Cytochrome P450 isoforms involved in melatonin metabolism in human liver microsomes. *Eur. J. Clin. Pharmacol.* **2001**, *56*, 881–888. [[CrossRef](#)]
162. Semak, I.; Korik, E.; Antonova, M.; Wortsman, J.; Slominski, A. Metabolism of melatonin by cytochrome P450s in rat liver mitochondria and microsomes. *J. Pineal Res.* **2008**, *45*, 515–523. [[CrossRef](#)] [[PubMed](#)]
163. Rogawski, M.A.; Roth, R.H.; Aghajanian, G.K. Melatonin: Deacetylation to 5-methoxytryptamine by liver but not brain aryl acylamidase. *J. Neurochem.* **1979**, *32*, 1219–1226. [[CrossRef](#)] [[PubMed](#)]
164. Hirata, F.; Hayaishi, O.; Tokuyama, T.; Seno, S. In vitro and in vivo formation of two new metabolites of melatonin. *J. Biol. Chem.* **1974**, *249*, 1311–1313. [[CrossRef](#)]
165. Hardeland, R.; Reiter, R.J.; Poeggeler, B.; Tan, D.X. The significance of the metabolism of the neurohormone melatonin: Antioxidative protection and formation of bioactive substances. *Neurosci. Biobehav. Rev.* **1993**, *17*, 347–357. [[CrossRef](#)]

166. Semak, I.; Naumova, M.; Korik, E.; Terekhovich, V.; Wortsman, J.; Slominski, A. A novel metabolic pathway of melatonin: Oxidation by cytochrome C. *Biochemistry* **2005**, *44*, 9300–9307. [CrossRef] [PubMed]
167. Fischer, T.W.; Sweatman, T.W.; Semak, I.; Sayre, R.M.; Wortsman, J.; Slominski, A. Constitutive and UV-induced metabolism of melatonin in keratinocytes and cell-free systems. *FASEB J.* **2006**, *20*, 1564–1566. [CrossRef]
168. Kim, T.K.; Kleszczyński, K.; Janjetovic, Z.; Sweatman, T.; Lin, Z.; Li, W.; Reiter, R.J.; Fischer, T.W.; Slominski, A.T. Metabolism of melatonin and biological activity of intermediates of melatonergic pathway in human skin cells. *FASEB J.* **2013**, *27*, 2742–2755. [CrossRef]
169. Slominski, A.T.; Semak, I.; Fischer, T.W.; Kim, T.K.; Kleszczyński, K.; Hardeland, R.; Reiter, R.J. Metabolism of melatonin in the skin: Why is it important? *Exp. Dermatol.* **2017**, *26*, 563–568. [CrossRef]
170. Kim, T.K.; Lin, Z.; Li, W.; Reiter, R.J.; Slominski, A.T. N<sup>1</sup>-Acetyl-5-Methoxykynuramine (AMK) is produced in the human epidermis and shows antiproliferative effects. *Endocrinology* **2015**, *156*, 1630–1636. [CrossRef]
171. Kim, T.K.; Lin, Z.; Tidwell, W.J.; Li, W.; Slominski, A.T. Melatonin and its metabolites accumulate in the human epidermis in vivo and inhibit proliferation and tyrosinase activity in epidermal melanocytes in vitro. *Mol. Cell. Endocrinol.* **2015**, *404*, 1–8. [CrossRef]
172. Reiter, R.J. Functional pleiotropy of the neurohormone melatonin: Antioxidant protection and neuroendocrine regulation. *Front. Neuroendocrinol.* **1995**, *16*, 383–415. [CrossRef] [PubMed]
173. Carrillo-Vico, A.; Guerrero, J.M.; Lardone, P.J.; Reiter, R.J. A review of the multiple actions of melatonin on the immune system. *Endocrine* **2005**, *27*, 189–200. [CrossRef]
174. Celinski, K.; Konturek, S.J.; Konturek, P.C.; Brzozowski, T.; Cichoz-Lach, H.; Slomka, M.; Malgorzata, P.; Bielanski, W.; Reiter, R.J. Melatonin or L-tryptophan accelerates healing of gastroduodenal ulcers in patients treated with omeprazole. *J. Pineal Res.* **2011**, *50*, 389–394. [CrossRef] [PubMed]
175. Korkmaz, A.; Topal, T.; Tan, D.X.; Reiter, R.J. Role of melatonin in metabolic regulation. *Rev. Endocr. Metab. Disord.* **2009**, *10*, 261–270. [CrossRef] [PubMed]
176. Tan, D.X.; Manchester, L.C.; Fuentes-Broto, L.; Paredes, S.D.; Reiter, R.J. Significance and application of melatonin in the regulation of brown adipose tissue metabolism: Relation to human obesity. *Obes. Rev.* **2011**, *12*, 167–188. [CrossRef]
177. Tan, D.X.; Manchester, L.C.; Terron, M.P.; Flores, L.J.; Reiter, R.J. One molecule, many derivatives: A never-ending interaction of melatonin with reactive oxygen and nitrogen species? *J. Pineal Res.* **2007**, *42*, 28–42. [CrossRef]
178. Slominski, A.; Fischer, T.W.; Zmijewski, M.A.; Wortsman, J.; Semak, I.; Zbytek, B.; Slominski, R.M.; Tobin, D.J. On the role of melatonin in skin physiology and pathology. *Endocrine* **2005**, *27*, 137–148. [CrossRef]
179. Fischer, T.W.; Zmijewski, M.A.; Zbytek, B.; Sweatman, T.W.; Slominski, R.M.; Wortsman, J.; Slominski, A. Oncostatic effects of the indole melatonin and expression of its cytosolic and nuclear receptors in cultured human melanoma cell lines. *Int. J. Oncol.* **2006**, *29*, 665–672. [CrossRef]
180. González-González, A.; Mediavilla, M.D.; Sánchez-Barceló, E.J. Melatonin: A molecule for reducing breast cancer risk. *Molecules* **2018**, *23*, 336. [CrossRef]
181. Kleszczyński, K.; Bilska, B.; Stegemann, A.; Flis, D.J.; Ziolkowski, W.; Pyza, E.; Luger, T.A.; Reiter, R.J.; Böhm, M.; Slominski, A.T. Melatonin and its metabolites ameliorate UVR-induced mitochondrial oxidative stress in human MNT-1 melanoma cells. *Int. J. Mol. Sci.* **2018**, *19*, 3786. [CrossRef]
182. Kleszczyński, K.; Kim, T.K.; Bilska, B.; Sarna, M.; Mokrzynski, K.; Stegemann, A.; Pyza, E.; Reiter, R.J.; Steinbrink, K.; Böhm, M.; et al. Melatonin exerts oncostatic capacity and decreases melanogenesis in human MNT-1 melanoma cells. *J. Pineal Res.* **2019**, *67*, e12610. [CrossRef] [PubMed]
183. Li, Y.; Li, S.; Zhou, Y.; Meng, X.; Zhang, J.J.; Xu, D.P.; Li, H.B. Melatonin for the prevention and treatment of cancer. *Oncotarget* **2017**, *8*, 39896–39921. [CrossRef]
184. Mao, L.; Yuan, L.; Slakey, L.M.; Jones, F.E.; Burow, M.E.; Hill, S.M. Inhibition of breast cancer cell invasion by melatonin is mediated through regulation of the p38 mitogen-activated protein kinase signaling pathway. *Breast Cancer Res.* **2010**, *12*, R107. [CrossRef]
185. Majidinia, M.; Reiter, R.J.; Shakouri, S.K.; Yousefi, B. The role of melatonin, a multitasking molecule, in retarding the processes of ageing. *Ageing Res. Rev.* **2018**, *47*, 198–213. [CrossRef] [PubMed]
186. Jockers, R.; Delagrange, P.; Dubocovich, M.L.; Markus, R.P.; Renault, N.; Tosini, G.; Cecon, E.; Zlotos, D.P. Update on melatonin receptors: IUPHAR Review 20. *Br. J. Pharmacol.* **2016**, *173*, 2702–2725. [CrossRef] [PubMed]
187. Agil, A.; Navarro-Alarcon, M.; Ali, F.A.Z.; Albrakati, A.; Salagre, D.; Campoy, C.; Elmahallawy, E.K. Melatonin enhances the mitochondrial functionality of brown adipose tissue in obese-diabetic rats. *Antioxidants* **2021**, *10*, 1482. [CrossRef] [PubMed]
188. Liu, Z.; Gan, L.; Luo, D.; Sun, C. Melatonin promotes circadian rhythm-induced proliferation through Clock/histone deacetylase 3/c-Myc interaction in mouse adipose tissue. *J. Pineal Res.* **2017**, *62*, e12383. [CrossRef]
189. Santaniello, S.; Cruciani, S.; Basoli, V.; Balzano, F.; Bellu, E.; Garroni, G.; Ginesu, G.C.; Cossu, M.L.; Facchin, F.; Delitala, A.P.; et al. Melatonin and Vitamin D Orchestrate Adipose Derived Stem Cell Fate by Modulating Epigenetic Regulatory Genes. *Int. J. Med. Sci.* **2018**, *15*, 1631–1639. [CrossRef]
190. Cos, S.; González, A.; Martínez-Campa, C.; Mediavilla, M.D.; Alonso-González, C.; Sánchez-Barceló, E.J. Melatonin as a selective estrogen enzyme modulator. *Curr. Cancer Drug Targets* **2008**, *8*, 691–702. [CrossRef]
191. Tan, D.X.; Manchester, L.C.; Reiter, R.J.; Qi, W.; Kim, S.J.; El-Sokkary, G.H. Ischemia/reperfusion-induced arrhythmias in the isolated rat heart: Prevention by melatonin. *J. Pineal Res.* **1998**, *25*, 184–191. [CrossRef]

192. Tengattini, S.; Reiter, R.J.; Tan, D.X.; Terron, M.P.; Rodella, L.F.; Rezzani, R. Cardiovascular diseases: Protective effects of melatonin. *J. Pineal Res.* **2008**, *44*, 16–25. [CrossRef] [PubMed]
193. Tan, D.X.; Manchester, L.C.; Reiter, R.J.; Qi, W.; Kim, S.J.; El-Sokkary, G.H. Melatonin protects hippocampal neurons in vivo against kainic acid-induced damage in mice. *J. Neurosci. Res.* **1998**, *54*, 382–389. [CrossRef]
194. Janjetovic, Z.; Jarrett, S.G.; Lee, E.F.; Duprey, C.; Reiter, R.J.; Slominski, A.T. Melatonin and its metabolites protect human melanocytes against UVB-induced damage: Involvement of NRF2-mediated pathways. *Sci. Rep.* **2017**, *7*, 1274. [CrossRef]
195. Santoro, R.; Mori, F.; Marani, M.; Grasso, G.; Cambria, M.A.; Blandino, G.; Muti, P.; Strano, S. Blockage of melatonin receptors impairs p53-mediated prevention of DNA damage accumulation. *Carcinogenesis* **2013**, *34*, 1051–1061. [CrossRef] [PubMed]
196. Reiter, R.J.; Tan, D.X.; Rosales-Corral, S.; Galano, A.; Jou, M.J.; Acuna-Castroviejo, D. Melatonin mitigates mitochondrial meltdown: Interactions with SIRT3. *Int. J. Mol. Sci.* **2018**, *19*, 2439. [CrossRef] [PubMed]
197. Hardeland, R. Antioxidant protection by melatonin: Multiplicity of mechanisms from radical detoxification to radical avoidance. *Endocrine* **2005**, *27*, 119–130. [CrossRef]
198. Reiter, R.J.; Rosales-Corral, S.; Tan, D.X.; Jou, M.J.; Galano, A.; Xu, B. Melatonin as a mitochondria-targeted antioxidant: One of evolution’s best ideas. *Cell. Mol. Life Sci.* **2017**, *74*, 3863–3881. [CrossRef] [PubMed]
199. Reiter, R.J.; Tan, D.X.; Manchester, L.C.; Qi, W. Biochemical reactivity of melatonin with reactive oxygen and nitrogen species: A review of the evidence. *Cell Biochem. Biophys.* **2001**, *34*, 237–256. [CrossRef]
200. Tan, D.X.; Manchester, L.C.; Qin, L.; Reiter, R.J. Melatonin: A mitochondrial targeting molecule involving mitochondrial protection and dynamics. *Int. J. Mol. Sci.* **2016**, *17*, 2124. [CrossRef]
201. Hardeland, R. Melatonin and inflammation-story of a double-edged blade. *J. Pineal Res.* **2018**, *65*, e12525. [CrossRef]
202. Galano, A.; Tan, D.X.; Reiter, R.J. On the free radical scavenging activities of melatonin’s metabolites, AFMK and AMK. *J. Pineal Res.* **2013**, *54*, 245–257. [CrossRef] [PubMed]
203. Tan, D.X.; Manchester, L.C.; Burkhardt, S.; Sainz, R.M.; Mayo, J.C.; Kohen, R.; Shohami, E.; Huo, Y.S.; Hardeland, R.; Reiter, R.J. N<sup>1</sup>-acetyl-N<sup>2</sup>-formyl-5-methoxykynuramine, a biogenic amine and melatonin metabolite, functions as a potent antioxidant. *FASEB J.* **2001**, *15*, 2294–2296. [CrossRef] [PubMed]
204. Bokov, A.; Chaudhuri, A.; Richardson, A. The role of oxidative damage and stress in aging. *Mech. Ageing Dev.* **2004**, *125*, 811–826. [CrossRef] [PubMed]
205. Harman, D. Aging: A theory based on free radical and radiation chemistry. *J. Gerontol.* **1956**, *11*, 298–300. [CrossRef]
206. Harman, D. The free radical theory of aging. *Antioxid. Redox Signal.* **2003**, *5*, 557–561. [CrossRef] [PubMed]
207. Alexeyev, M.F. Is there more to aging than mitochondrial DNA and reactive oxygen species? *FEBS J.* **2009**, *276*, 5768–5787. [CrossRef] [PubMed]
208. Hardeland, R.; Coto-Montes, A. New vistas on oxidative damage and aging. *Open Biol.* **2010**, *3*, 39–52. [CrossRef]
209. Lee, H.C.; Wei, Y.H. Mitochondria and aging. *Adv. Exp. Med. Biol.* **2012**, *942*, 311–327.
210. Stefanatos, R.; Sanz, A. The role of mitochondrial ROS in the aging brain. *FEBS Lett.* **2018**, *592*, 743–758. [CrossRef]
211. Krause, K.H. Aging: A revisited theory based on free radicals generated by NOX family NADPH oxidases. *Exp. Gerontol.* **2007**, *42*, 256–262. [CrossRef]
212. Alper, G.; Girgin, F.K.; Ozgönül, M.; Mentes, G.; Ersöz, B. MAO inhibitors and oxidant stress in aging brain tissue. *Eur. Neuropsychopharmacol.* **1999**, *9*, 247–252. [CrossRef]
213. Vida, C.; Corpas, I.; De la Fuente, M.; González, E.M. Age-related changes in xanthine oxidase activity and lipid peroxidation, as well as in the correlation between both parameters, in plasma and several organs from female mice. *J. Physiol. Biochem.* **2011**, *67*, 551–558. [CrossRef] [PubMed]
214. Legakis, J.E.; Koepke, J.I.; Jedeszko, C.; Barlaskar, F.; Terlecky, L.J.; Edwards, H.J.; Walton, P.A.; Terlecky, S.R. Peroxisome senescence in human fibroblasts. *Mol. Biol. Cell.* **2002**, *13*, 4243–4255. [CrossRef] [PubMed]
215. Kauppinen, T.E.S.; Kauppinen, J.H.K.; Larsson, N.G. Mammalian mitochondria and aging: An update. *Cell Metab.* **2017**, *25*, 57–71. [CrossRef]
216. Hardeland, R. Melatonin and the theories of aging: A critical appraisal of melatonin’s role in antiaging mechanisms. *J. Pineal Res.* **2013**, *55*, 325–356. [CrossRef]
217. Poeggeler, B. Melatonin, aging, and age-related diseases: Perspectives for prevention, intervention, and therapy. *Endocrine* **2005**, *27*, 201–212. [CrossRef]
218. Reiter, R.J.; Tan, D.X.; Poeggeler, B.; Menendez-Pelaez, A.; Chen, L.D.; Saarela, S. Melatonin as a free radical scavenger: Implications for aging and age-related diseases. *Ann. N Y Acad. Sci.* **1994**, *719*, 1–12. [CrossRef] [PubMed]
219. Reiter, R.J.; Tan, D.X.; Kim, S.J.; Manchester, L.C.; Qi, W.; Garcia, J.J.; Cabrera, J.C.; El-Sokkary, G.; Rouvier-Garay, V. Augmentation of indices of oxidative damage in life-long melatonin-deficient rats. *Mech. Ageing Dev.* **1999**, *110*, 157–173. [CrossRef]
220. Cedikova, M.; Pitule, P.; Kripnerova, M.; Markov, M.; Kuncova, J. Multiple roles of mitochondria in aging processes. *Physiol. Res.* **2016**, *65*, S519–S531. [CrossRef]
221. Mocayar Marón, F.J.; Ferder, L.; Reiter, R.J.; Manucha, W. Daily and seasonal mitochondrial protection: Unraveling common possible mechanisms involving vitamin D and melatonin. *J. Steroid Biochem. Mol. Biol.* **2020**, *199*, 105595. [CrossRef]
222. Reiter, R.J.; Paredes, S.D.; Korkmaz, A.; Jou, M.J.; Tan, D.X. Melatonin combats molecular terrorism at the mitochondrial level. *Interdiscip. Toxicol.* **2008**, *1*, 137–149. [CrossRef] [PubMed]

223. Acuña Castroviejo, D.; López, L.C.; Escames, G.; López, A.; García, J.A.; Reiter, R.J. Melatonin-mitochondria interplay in health and disease. *Curr. Top Med. Chem.* **2011**, *11*, 221–240. [CrossRef] [PubMed]
224. Galano, A.; Tan, D.X.; Reiter, R.J. Melatonin as a natural ally against oxidative stress: A physicochemical examination. *J. Pineal Res.* **2011**, *51*, 1–16. [CrossRef] [PubMed]
225. Reiter, R.J.; Rosales-Corral, S.; Zhou, X.; Tan, D.X. Role of SIRT3/SOD2 signaling in mediating the antioxidant actions of melatonin in mitochondria. *Curr. Trends Endocrinol.* **2017**, *9*, 45–49.
226. Mayo, J.C.; Sainz, R.M.; González Menéndez, P.; Cepas, V.; Tan, D.X.; Reiter, R.J. Melatonin and sirtuins: A “not-so unexpected” relationship. *J. Pineal Res.* **2017**, *62*, e12391. [CrossRef] [PubMed]
227. Paradies, G.; Paradies, V.; Ruggiero, F.M.; Petrosillo, G. Changes in the mitochondrial permeability transition pore in aging and age-associated diseases. *Mech. Ageing Dev.* **2013**, *134*, 1–9. [CrossRef]
228. Hardeland, R. Melatonin and the electron transport chain. *Cell. Mol. Life Sci.* **2017**, *74*, 3883–3896. [CrossRef]
229. Hardeland, R. Aging, melatonin, and the pro- and anti-inflammatory networks. *Int. J. Mol. Sci.* **2019**, *20*, 1223. [CrossRef]
230. Hardeland, R. Opposite effects of melatonin in different systems and under different conditions. *Curr. Top. Biochem. Res.* **2016**, *17*, 57–69.
231. Hardeland, R. Brain inflammaging: Roles of melatonin, circadian clocks and sirtuins. *J. Clin. Cell. Immunol.* **2018**, *9*, 543. [CrossRef]
232. Tamtaji, O.R.; Mobini, M.; Reiter, R.J.; Azami, A.; Gholami, M.S.; Asemi, Z. Melatonin, a toll-like receptor inhibitor: Current status and future perspectives. *J. Cell. Physiol.* **2019**, *234*, 7788–7795. [CrossRef] [PubMed]
233. Xia, M.Z.; Liang, Y.L.; Wang, H.; Chen, X.; Huang, Y.Y.; Zhang, Z.H.; Chen, Y.H.; Zhang, C.; Zhao, M.; Xu, D.X.; et al. Melatonin modulates TLR4-mediated inflammatory genes through MyD88- and TRIF-dependent signaling pathways in lipopolysaccharide-stimulated RAW264.7 cells. *J. Pineal Res.* **2012**, *53*, 325–334. [CrossRef]
234. Cardinali, D.P.; Hardeland, R. Inflammaging, metabolic syndrome and melatonin: A call for treatment studies. *Neuroendocrinology* **2017**, *104*, 382–397. [CrossRef] [PubMed]
235. Dai, Y.; Peng, L.; Liu, Y.; Xu, Y.; Qiao, J. Melatonin binds with high affinity and specificity to beta-amyloid: LC-MS provides insight into Alzheimer’s disease treatment. *FEBS Open Bio.* **2021**, *11*, 2800–2806. [CrossRef] [PubMed]
236. Hardeland, R.; Cardinali, D.P.; Brown, G.M.; Pandi-Perumal, S.R. Melatonin and brain inflammaging. *Prog. Neurobiol.* **2015**, *127*, 46–63. [CrossRef]
237. Lin, L.; Huang, Q.X.; Yang, S.S.; Chu, J.; Wang, J.Z.; Tian, Q. Melatonin in Alzheimer’s disease. *Int. J. Mol. Sci.* **2013**, *14*, 14575–14593. [CrossRef]
238. Pandi-Perumal, S.R.; BaHammam, A.S.; Brown, G.M.; Spence, D.W.; Bharti, V.K.; Kaur, C.; Hardeland, R.; Cardinali, D.P. Melatonin antioxidative defense: Therapeutic implications for aging and neurodegenerative processes. *Neurotox. Res.* **2013**, *23*, 267–300. [CrossRef]
239. Rosales-Corral, S.A.; Acuña-Castroviejo, D.; Coto-Montes, A.; Boga, J.A.; Manchester, L.C.; Fuentes-Broto, L.; Korkmaz, A.; Ma, S.; Tan, D.X.; Reiter, R.J. Alzheimer’s disease: Pathological mechanisms and the beneficial role of melatonin. *J. Pineal Res.* **2012**, *52*, 167–202. [CrossRef]
240. Srinivasan, V.; Pandi-Perumal, S.R.; Cardinali, D.P.; Poeggeler, B.; Hardeland, R. Melatonin in Alzheimer’s disease and other neurodegenerative disorders. *Behav. Brain Funct.* **2006**, *2*, 15. [CrossRef]
241. Slominski, A.; Baker, J.; Rosano, T.G.; Guisti, L.W.; Ermak, G.; Grande, M.; Gaudet, S.J. Metabolism of serotonin to N-acetylserotonin, melatonin, and 5-methoxytryptamine in hamster skin culture. *J. Biol. Chem.* **1996**, *271*, 12281–12286. [CrossRef] [PubMed]
242. Slominski, A.; Semak, I.; Pisarchik, A.; Sweatman, T.; Szczesniewski, A.; Wortsman, J. Conversion of L-tryptophan to serotonin and melatonin in human melanoma cells. *FEBS Lett.* **2002**, *511*, 102–106. [CrossRef]
243. Slominski, A.; Pisarchik, A.; Johansson, O.; Jing, C.; Semak, I.; Slugocki, G.; Wortsman, J. Tryptophan hydroxylase expression in human skin cells. *Biochim. Biophys. Acta* **2003**, *1639*, 80–86. [CrossRef]
244. Kobayashi, H.; Kromminga, A.; Dunlop, T.W.; Tychsen, B.; Conrad, F.; Suzuki, N.; Memezawa, A.; Bettermann, A.; Aiba, S.; Carlberg, C.; et al. A role of melatonin in neuroectodermal-mesodermal interactions: The hair follicle synthesizes melatonin and expresses functional melatonin receptors. *FASEB J.* **2005**, *19*, 1710–1712. [CrossRef]
245. Slominski, A.T.; Zmijewski, M.A.; Semak, I.; Kim, T.K.; Janjetovic, Z.; Slominski, R.M.; Zmijewski, J.W. Melatonin, mitochondria, and the skin. *Cell Mol. Life Sci.* **2017**, *74*, 3913–3925. [CrossRef]
246. Slominski, A.; Tobin, D.J.; Shibahara, S.; Wortsman, J. Melanin pigmentation in mammalian skin and its hormonal regulation. *Physiol. Rev.* **2004**, *84*, 1155–1228. [CrossRef]
247. Fischer, T.W.; Slominski, A.; Tobin, D.J.; Paus, R. Melatonin and the hair follicle. *J. Pineal Res.* **2008**, *44*, 1–15. [CrossRef]
248. Fischer, T.W.; Burmeister, G.; Schmidt, H.W.; Elsner, P. Melatonin increases anagen hair rate in women with androgenetic alopecia or diffuse alopecia: Results of a pilot randomized controlled trial. *Br. J. Dermatol.* **2004**, *150*, 341–345. [CrossRef]
249. Slominski, A.; Pruski, D. Melatonin inhibits proliferation and melanogenesis in rodent melanoma cells. *Exp. Cell Res.* **1993**, *206*, 189–194. [CrossRef]
250. Slominski, A.T.; Zmijewski, M.A.; Jetten, A.M. ROR $\alpha$  is not a receptor for melatonin (response to DOI 10.1002/bies.201600018). *BioEssays* **2016**, *38*, 1193–1194. [CrossRef]

251. Slominski, A.T.; Kim, T.K.; Takeda, Y.; Janjetovic, Z.; Brozyna, A.A.; Skobowiat, C.; Wang, J.; Postlethwaite, A.; Li, W.; Tuckey, R.C.; et al. ROR $\alpha$  and ROR $\gamma$  are expressed in human skin and serve as receptors for endogenously produced non-calcemic 20-hydroxy- and 20,23-dihydroxyvitamin D. *FASEB J.* **2014**, *28*, 2775–2789. [[CrossRef](#)]
252. Fischer, T.W.; Elsner, P. The antioxidative potential of melatonin in the skin. *Curr. Probl. Dermatol.* **2001**, *29*, 165–174. [[PubMed](#)]
253. Hardeland, R. Melatonin, its metabolites and their interference with reactive nitrogen compounds. *Molecules* **2021**, *26*, 4105. [[CrossRef](#)] [[PubMed](#)]
254. Kleszczyński, K.; Zillikens, D.; Fischer, T.W. Melatonin enhances mitochondrial ATP synthesis, reduces reactive oxygen species formation, and mediates translocation of the nuclear erythroid 2-related factor 2 resulting in activation of phase-2 antioxidant enzymes ( $\gamma$ -GCS, HO-1, NQO1) in ultraviolet radiation-treated normal human epidermal keratinocytes (NHEK). *J. Pineal Res.* **2016**, *61*, 187–197.
255. Kerem, H.; Akdemir, O.; Ates, U.; Uyanikgil, Y.; Demirel Sezer, E.; Bilkay, U.; Turgut, M.; Sozmen, E.; Songur, E. The effect of melatonin on a dorsal skin flap model. *J. Investigig. Surg.* **2014**, *27*, 57–64. [[CrossRef](#)] [[PubMed](#)]
256. Reiter, R.J.; Tan, D.X.; Osuna, C.; Gitto, E. Actions of melatonin in the reduction of oxidative stress. A review. *J Biomed. Sci.* **2000**, *7*, 444–458. [[CrossRef](#)] [[PubMed](#)]
257. Tan, D.X.; Manchester, L.C.; Reiter, R.J.; Plummer, B.F.; Limson, J.; Weintraub, S.T.; Qi, W. Melatonin directly scavenges hydrogen peroxide: A potentially new metabolic pathway of melatonin biotransformation. *Free Radic. Biol. Med.* **2000**, *29*, 1177–1185. [[CrossRef](#)]
258. Poeggeler, B.; Reiter, R.J.; Tan, D.X.; Chen, L.D.; Manchester, L.C. Melatonin, hydroxyl radical-mediated oxidative damage, and aging: A hypothesis. *J. Pineal Res.* **1993**, *14*, 151–168. [[CrossRef](#)]
259. Kilańczyk, E.; Bryszewska, M. The effect of melatonin on antioxidant enzymes in human diabetic skin fibroblasts. *Cell Mol. Biol. Lett.* **2003**, *8*, 333–336.
260. Fischer, T.W.; Kleszczyński, K.; Hardkop, L.H.; Kruse, N.; Zillikens, D. Melatonin enhances antioxidative enzyme gene expression (CAT, GPx, SOD), prevents their UVR-induced depletion, and protects against the formation of DNA damage (8-hydroxy-2'-deoxyguanosine) in ex vivo human skin. *J. Pineal Res.* **2013**, *54*, 303–312. [[CrossRef](#)]
261. Haslam, I.S.; Jadkauskaite, L.; Szabó, I.L.; Staeghe, S.; Hesebeck-Brinckmann, J.; Jenkins, G.; Bhogal, R.K.; Lim, F.L.; Farjo, N.; Farjo, B.; et al. Oxidative damage control in a human (Mini-) organ: Nrf2 activation protects against oxidative stress-induced hair growth inhibition. *J. Investigig. Dermatol.* **2017**, *137*, 295–304. [[CrossRef](#)]
262. Rezzani, R.; Rodella, L.F.; Favero, G.; Damiani, G.; Paganelli, C.; Reiter, R.J. Attenuation of ultraviolet A-induced alterations in NIH3T3 dermal fibroblasts by melatonin. *Br. J. Dermatol.* **2014**, *170*, 382–391. [[CrossRef](#)] [[PubMed](#)]
263. Ryoo, Y.W.; Suh, S.I.; Mun, K.C.; Kim, B.C.; Lee, K.S. The effects of the melatonin on ultraviolet-B irradiated cultured dermal fibroblasts. *J. Dermatol. Sci.* **2001**, *27*, 162–169. [[CrossRef](#)]
264. Sliwinski, T.; Rozej, W.; Morawiec-Bajda, A.; Morawiec, Z.; Reiter, R.; Blasiak, J. Protective action of melatonin against oxidative DNA damage: Chemical inactivation versus base-excision repair. *Mutat. Res.* **2007**, *634*, 220–227. [[CrossRef](#)] [[PubMed](#)]
265. Kleszczyński, K.; Tukaj, S.; Kruse, N.; Zillikens, D.; Fischer, T.W. Melatonin prevents ultraviolet radiation-induced alterations in plasma membrane potential and intracellular pH in human keratinocytes. *J. Pineal Res.* **2013**, *54*, 89–99. [[CrossRef](#)] [[PubMed](#)]
266. Schaefer, M.; Hardeland, R. The melatonin metabolite N-acetyl-5-methoxykynuramine is a potent singlet oxygen scavenger. *J. Pineal Res.* **2009**, *46*, 49–52. [[CrossRef](#)] [[PubMed](#)]
267. Kleszczyński, K.; Zwicker, S.; Tukaj, S.; Kasperkiewicz, M.; Zillikens, D.; Wolf, R.; Fischer, T.W. Melatonin compensates silencing of heat shock protein 70 and suppresses ultraviolet radiation-induced inflammation in human skin ex vivo and cultured keratinocytes. *J. Pineal Res.* **2015**, *58*, 117–126. [[CrossRef](#)] [[PubMed](#)]
268. Scheuer, C. Melatonin for prevention of erythema and oxidative stress in response to ultraviolet radiation. *Dan. Med. J.* **2017**, *64*, B5358.
269. Scheuer, C.; Pommergaard, H.C.; Rosenberg, J.; Gögenur, I. Dose dependent sun protective effect of topical melatonin: A randomized, placebo-controlled, double-blind study. *J. Dermatol. Sci.* **2016**, *84*, 178–185. [[CrossRef](#)]
270. Sierra, A.F.; Ramírez, M.L.; Campmany, A.C.; Martínez, A.R.; Náveros, B.C. In vivo and in vitro evaluation of the use of a newly developed melatonin loaded emulsion combined with UV filters as a protective agent against skin irradiation. *J. Dermatol. Sci.* **2013**, *69*, 202–214. [[CrossRef](#)]
271. Goldberg, D.J.; Robinson, D.M.; Granger, C. Clinical evidence of the efficacy and safety of a new 3-in-1 anti-aging topical night serum-in-oil containing melatonin, bakuchiol, and ascorbyl tetraisopalmitate: 103 females treated from 28 to 84 days. *J. Cosmet. Dermatol.* **2019**, *18*, 806–814. [[CrossRef](#)]
272. Narda, M.; Brown, A.; Muscatelli-Groux, B.; Grimaud, J.A.; Granger, C. Epidermal and dermal hallmarks of photoaging are prevented by treatment with night serum containing melatonin, bakuchiol, and ascorbyl tetraisopalmitate: In vitro and ex vivo studies. *Dermatol. Ther.* **2020**, *10*, 191–202. [[CrossRef](#)] [[PubMed](#)]
273. Kleszczyński, K.; Hardkop, L.H.; Fischer, T.W. Differential effects of melatonin as a broad range UV-damage preventive dermatendoctrine regulator. *Dermato-Endocrinology* **2011**, *3*, 27–31. [[CrossRef](#)] [[PubMed](#)]
274. Rusanova, I.; Martínez-Ruiz, L.; Florido, J.; Rodríguez-Santana, C.; Guerra-Librero, A.; Acuña-Castroviejo, D.; Escames, G. Protective effects of melatonin on the skin: Future perspectives. *Int. J. Mol. Sci.* **2019**, *20*, 4948. [[CrossRef](#)]

275. Bilksa, B.; Schedel, F.; Piotrowska, A.; Stefan, J.; Zmijewski, M.; Pyza, E.; Reiter, R.J.; Steinbrink, K.; Slominski, A.T.; Tulic, M.K.; et al. Mitochondrial function is controlled by melatonin and its metabolites in vitro in human melanoma cells. *J. Pineal Res.* **2021**, *70*, e12728. [[CrossRef](#)]
276. Granger, C.; Brown, A.; Aladren, S.; Narda, M. Night cream containing melatonin, carnosine and helichrysum italicum extract helps reduce skin reactivity and signs of photodamage: Ex vivo and clinical studies. *Dermatol. Ther.* **2020**, *10*, 1315–1329. [[CrossRef](#)]
277. Sagan, D.; Stepniak, J.; Gesing, A.; Lewinski, A.; Karbownik-Lewinska, M. Melatonin reverses the enhanced oxidative damage to membrane lipids and improves skin biophysical characteristics in former-smokers—A study in postmenopausal women. *Ann. Agric. Environ. Med.* **2017**, *24*, 659–666. [[CrossRef](#)]
278. Dong, K.; Goyarts, E.; Rella, A.; Pelle, E.; Wong, Y.H.; Pernodet, N. Age associated decrease of MT-1 melatonin receptor in human dermal skin fibroblasts impairs protection against UV-induced DNA damage. *Int. J. Mol. Sci.* **2020**, *21*, 326. [[CrossRef](#)] [[PubMed](#)]
279. Fischer, T.W.; Greif, C.; Fluhr, J.W.; Wigger-Alberti, W.; Elsner, P. Percutaneous penetration of topically applied melatonin in a cream and an alcoholic solution. *Skin Pharmacol. Physiol.* **2004**, *17*, 190–194. [[CrossRef](#)]
280. Hirokazu Tsukahara, H.; Shibata, R.; Ohshima, Y.; Todoroki, Y.; Sato, S.; Ohta, N.; Hiraoka, M.; Yoshida, A.; Nishima, S.; Mayumi, M. Oxidative stress and altered antioxidant defenses in children with acute exacerbation of atopic dermatitis. *Life Sci.* **2003**, *72*, 2509–2516. [[CrossRef](#)]
281. Hatem, S.; Nasr, M.; Moftah, N.H.; Ragai, M.H.; Geneidi, A.S.; Elkhesen, S.A. Clinical cosmeceutical repurposing of melatonin in androgenic alopecia using nanostructured lipid carriers prepared with antioxidant oils. *Expert Opin. Drug Deliv.* **2018**, *15*, 927–935. [[CrossRef](#)]
282. Reiter, R.J.; Paredes, S.D.; Manchester, L.C.; Tan, D.X. Reducing oxidative/nitrosative stress: A newly-discovered genre for melatonin. *Crit. Rev. Biochem. Mol. Biol.* **2009**, *44*, 175–200. [[CrossRef](#)] [[PubMed](#)]