

Optimization of Brigatinib as New Wild-Type Sparing Inhibitors of EGFR^{T790M/C797S} MutantsShan Li,[▽] Tao Zhang,[▽] Su-Jie Zhu,[▽] Chong Lei, Mengzhen Lai, Lijie Peng, Linjiang Tong, Zilu Pang, Xiaoyun Lu, Jian Ding, Xiaomei Ren,^{*} Cai-Hong Yun,^{*} Hua Xie,^{*} and Ke Ding^{*}Cite This: *ACS Med. Chem. Lett.* 2022, 13, 196–202

Read Online

ACCESS |



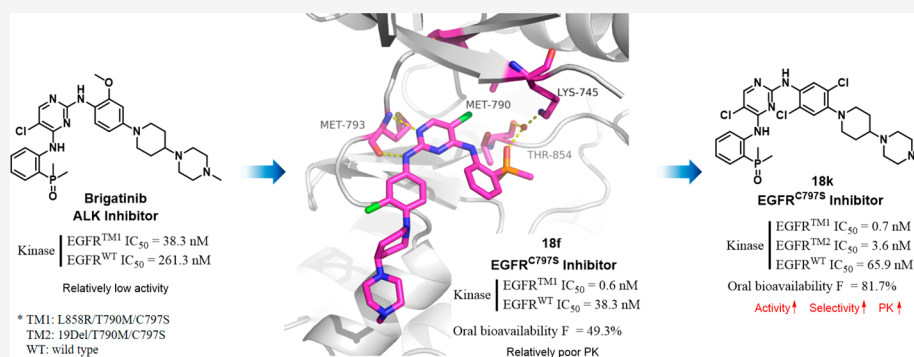
Metrics & More



Article Recommendations



Supporting Information



ABSTRACT: A series of brigatinib derivatives were designed and synthesized as new potent and selective EGFR^{T790M/C797S} inhibitors. One of the most potent and selective compounds **18k** strongly suppressed the EGFR^{L858R/T790M/C797S} and EGFR^{19Del/T790M/C797S} kinases with IC₅₀ values of 0.7 and 3.6 nM, respectively, which were over 54-fold more potent than the lead compound. **18k** also demonstrated promising EGFR^{T790M/C797S} mutant selectivity, and was 94-fold less potent against the wild type EGFR. A cocrystal structure of EGFR^{T790M/C797S} with a close derivative **18f** was solved to provide insight on the inhibitor's binding mode. Moreover, compound **18k** was orally bioavailable and demonstrated highly desirable PK properties, making it a promising lead compound for further structural optimization.

KEYWORDS: Epidermal Growth Factor Receptor (EGFR), Resistant Mutation, C797S, Antitumor

Epidermal growth factor receptor (EGFR) is a well validated target for non-small cell lung cancer (NSCLC) drug discovery.¹ Five small molecular EGFR inhibitors (i.e., gefitinib (1),² erlotinib (2),³ afatinib (3),⁴ dacomitinib (4),⁵ and osimertinib (AZD9291, 5),^{6,7} Figure 1) have been approved by the U.S. Food and Drug Administration (FDA). The first generation drugs 1 and 2 clinically apply to the treatment of NSCLC patients with EGFR activating mutations (e.g., exon 19 deletion (19 Del) and a Leu⁸⁵⁸ → Arg⁸⁵⁸ point mutation (L858R)). Irreversible compounds 3 and 4 belong to the second generation EGFR inhibitor drugs which achieve significant clinical benefit for metastatic and advanced NSCLC patients harboring “gain of function” mutated EGFRs. Although drugs 3 and 4 exhibit strong suppression on the resistant Thr⁷⁹⁰ → Met⁷⁹⁰ (T790M) mutants in preclinical models, the low clinical maximal tolerated dose (MTD) attributing to the strong wild-type EGFR (EGFR^{WT}) inhibition limited their application in resistant patients with the EGFR^{T790M} mutant. Wild-type sparing EGFR^{T790M} inhibitor **5** is the first FDA approved third generation drug for resistant NSCLC patients with EGFR^{T790M} mutations, and it was later approved as a first-line treatment for metastatic NSCLC

patients with positive EGFR mutations. Following the success of **5**, two structurally close derivatives, i.e., almonertinib (6)⁸ and furmonertinib (7),⁹ were approved by National Medical Products Administration of China (NMPA) in 2020 and 2021, respectively. Several other third generation EGFR inhibitors, e.g., lazertinib, rezivertinib, and abivertinib, are in late stages of clinical investigation.¹⁰ We have also developed ASK120067 (8) as a highly potent and selective wild-type sparing EGFR^{T790M} inhibitor. A New Drug Application (NDA) was filed recently in China based on its highly promising efficacy in advanced and multiple brain metastatic NSCLC patients harboring EGFR^{T790M} mutations.¹¹

Structurally, all of the third generation EGFR inhibitors possess an acrylamide warhead to covalently react with the

Received: October 10, 2021

Accepted: January 5, 2022

Published: January 7, 2022



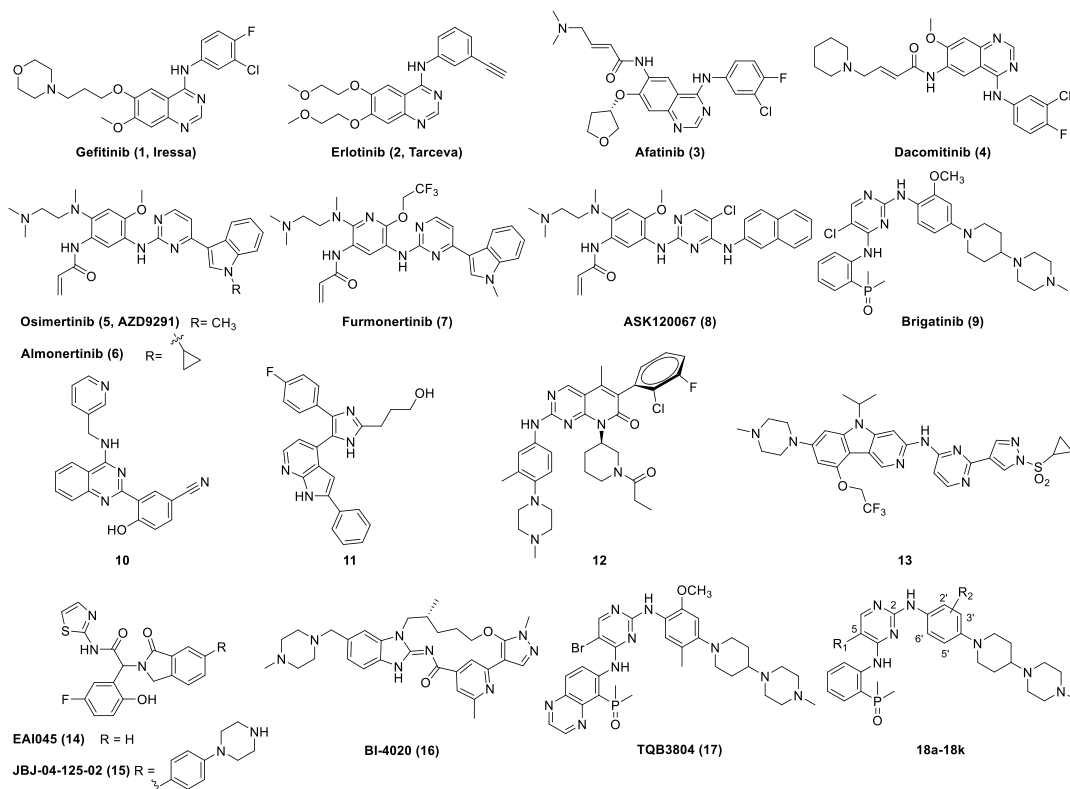


Figure 1. Chemical structures of approved or literature reported EGFR inhibitors.

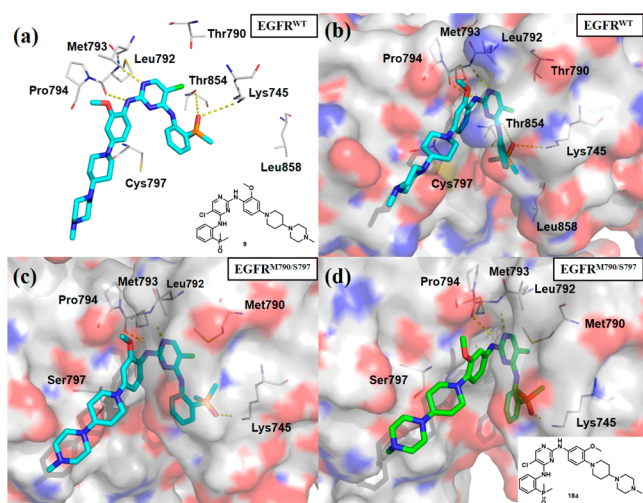
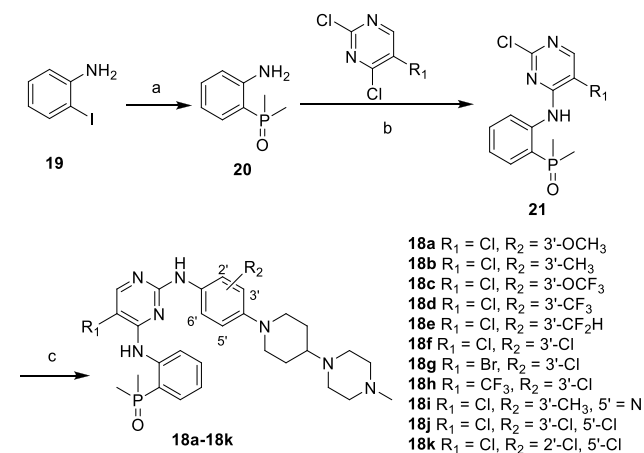


Figure 2. X-ray crystal structure of brigatinib (9) with EGFR^{Thr790/Cys797} (EGFR^{WT}) kinase domain (PDB 7AEM) shown in cyan and gray in stick and line representation (a) or in surface representation (b). Preliminary computational docking complex of 9 (c) and 18a (d) with EGFR^{Met790/Ser797} (PDB SZTO). Hydrogen bonds are indicated by yellow hatched lines to key amino acids.

Cys⁷⁹⁷ residue to achieve improved competitiveness with the high concentration ATP (mM level) in cells. A tertiary C797S (Cys⁷⁹⁷ → Ser⁷⁹⁷) mutation would impair the covalent formation between Cys⁷⁹⁷ and the acrylamide warhead and induce the resistance against the third generation drugs in approximately 20% of the pretreated NSCLC patients.¹² Although the patients with *trans* EGFR^{T790M/C797S} mutations in which the T790M and C797S mutations are located on different alleles could be efficiently treated by combinational

Scheme 1. Synthesis of Compounds 18a–18k^a



^aReagents and conditions: (a) dimethylphosphine oxide, Pd(OAc)₂, xantphos, K₃PO₄, DMF, 120 °C, overnight, 82%; (b) R₁ substituted pyrimidine, K₂CO₃, *n*-Bu₄NHSO₄, DMF, 65 °C, overnight, 63–84%; (c) R₂ substituted aniline, 2.5 M HCl in EtOH, 2-methoxyethanol, 120 °C, overnight, 37–75%.

therapies of a first generation drug and 5, the *cis*-EGFR^{T790M/C797S} mutation (mutations are on a same allele) mediated resistance remains an unmet clinical need.^{13,14} Therefore, development of the fourth generation EGFR^{T790M/C797S} inhibitors is highly desirable for overcoming the new mutation mediated resistance.¹⁵

Several classes of EGFR^{T790M/C797S} inhibitors have been reported. Examples include the traditional ATP competitive inhibitor brigatinib (9) and derivatives,¹⁶ 4-hydroxy-3-(4-aminoquinazolin-2-yl)benzimidazole derivative 10,¹⁷ trisubsti-

Table 2. Antiproliferative Activity of 18k in BaF3, PC9, PC9-EGFR^{TM2}, and H1299 Cell Lines^a

IC ₅₀ (μM)	BaF3			PC9	PC9-EGFR ^{TM2}	H1299 ^b
	EGFR ^{TM1}	EGFR ^{TM2}	parental cells			
18k	0.258 ± 0.167	0.141 ± 0.081	1.691 ± 0.253	1.192 ± 0.727	0.603 ± 0.294	2.115 ± 0.302
brigitinib (9)	0.286 ± 0.176	0.155 ± 0.046	4.191 ± 1.614	0.829 ± 0.148	1.110 ± 0.028	1.049 ± 0.160
AZD9291 (5)	2.893 ± 0.601	2.885 ± 0.689	4.427 ± 1.162	0.011 ± 0.005	3.210 ± 0.853	>10.0

^aBaF3-EGFR^{TM1}, BaF3-EGFR^{TM2}, and PC9-EGFR^{TM2} cell lines were built by Jian Ding's laboratory, Shanghai Institute of Materia Medica, Chinese Academy of Sciences. PC9-EGFR^{TM2} cell line was constructed by introducing the T790M/C797S mutant into PC9 (EGFR^{19Del}). Cellular assay was determined using the CCK8 assay. ^bH1299 (EGFR^{WT}) cellular assay was determined using sulforhodamine B (SRB) assay. The data were shown as mean ± SD from at least three independent experiments. TM1, L858R/T790M/C797S; TM2, 19Del/T790M/C797S.

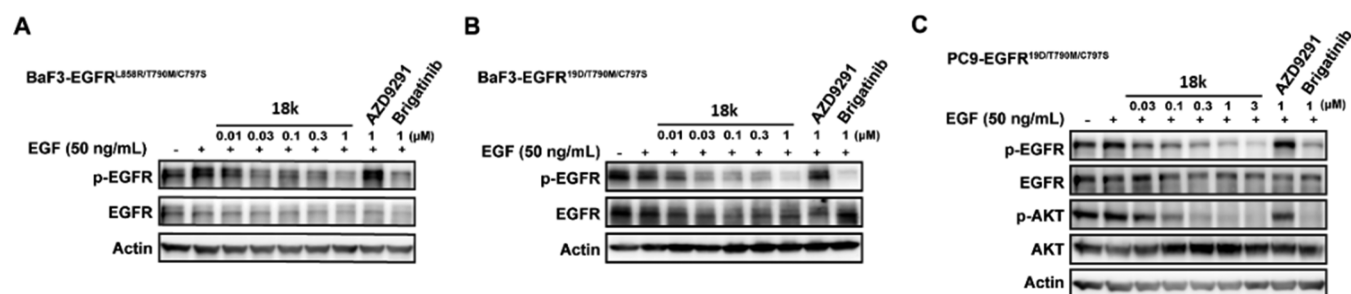


Figure 5. Target inhibition potency of 18k in EGFR^{C797S} mutant BaF3 cells and PC9 cells. Cells were treated with indicated compounds for 2 h, and the activation of EGFR and downstream signaling molecule were evaluated using Western Blot analysis.

Table 3. Mean PK Parameters of 18f and 18k after po or iv Administration in SD Rats^a

parameters	18f		18k	
	po ^b	iv ^c	po ^b	iv ^c
T _{1/2} (h)	2.84 ± 0.11	3.73 ± 1.36	2.16 ± 0.2	2.67 ± 0.45
T _{max} (h)	1.50 ± 0.87		2.67 ± 1.15	
C _{max} (ng/mL)	470 ± 155	345 ± 52	868 ± 181	288 ± 16
AUC _(0-t) (h·ng/mL)	2553 ± 704	862 ± 88	4552 ± 1154	929 ± 91
AUC _(0-∞) (h·ng/mL)	2563 ± 709	898 ± 57	5219 ± 1536	1060 ± 144
CL (mL/min/kg)		9.30 ± 0.57		7.97 ± 1.17
MRT _(0-∞) (h)	4.79 ± 0.49	4.08 ± 1.24	4.37 ± 0.62	3.65 ± 0.56
V _{ss} (mL/kg)		2275 ± 710		1720 ± 40
F (%)	49.3 ± 11.8		81.7 ± 20.7	

^an = Three animals per study. ^bDosed at 3 mg/kg. ^cDosed at 0.5 mg/kg.

1).²⁹ Beginning with the commercially available 2-iodoaniline (19) and dimethylphosphine oxide, intermediate 20 was obtained via a palladium-catalyzed coupling reaction. Then a two-step, sequential S_NAr reaction was conducted to give the target compounds 18a–18k. Biological evaluation showed that compound 18a indeed exhibited a 16-fold improved EGFR^{L858R/T790M/C797S} (EGFR^{TM1}) inhibitory potency with an IC₅₀ value of 2.4 nM compared with 9 (Table 1). In addition, the switch of the substituent position also resulted in EGFR^{WT} and EGFR^{L858R/T790M} (EGFR^{DM}) potency improvement by factors of 5.1 and 21.2, respectively. Notably, compound 18a exhibited an improved wild-type sparing selectivity with an EGFR^{WT}/EGFR^{TM1} ratio of 21.3, while the corresponding value for 9 was 6.8. Further investigation also revealed that the 3'-position could be substituted by a variety of substituents, e.g., -CH₃ (18b), -OCF₃ (18c), -CF₃ (18d), -CF₂H (18e), and -Cl (18f), to demonstrate wild-type sparing strong inhibition against the resistant EGFR^{TM1} and EGFR^{DM} kinases. Particularly, compounds 18b and 18f exhibited the strongest EGFR^{TM1} inhibition with IC₅₀ values of 0.7 and 0.6 nM, respectively. The compounds could also potentially suppress EGFR^{DM} kinase with IC₅₀ values of 2.7 and 1.8 nM, but their

activities against EGFR^{WT} were 23- and 21-fold less potent, respectively. Additionally, results showed that a replacement of 5-Cl of the pyrimidine motif in 18f by a Br group (18g) barely led to an obvious effect on the activity and selectivity, while the 5-CF₃ substituted analogue (18h) totally abolished the EGFR inhibitory potency. The investigation also suggested the aniline group at the pyrimidine 2-position could be replaced by a pyridinyl moiety (18i) without causing a significant potency change to the EGFR triple mutated kinase (IC₅₀ = 2.1 nM). Further investigation revealed that 3',5'-dichloro analogue 18j was 55-fold less potent than 18f against EGFR^{TM1} triple mutant, while the 2',5'-dichloro compound 18k exhibited identical EGFR^{TM1} inhibition to that of 18f, with an IC₅₀ value of 0.7 nM. Further evaluation suggested that 18k potentially suppressed the EGFR^{19Del/T790M/C797S} mutated kinase, which is a clinically more common resistant mutation, with an IC₅₀ value of 3.6 (±1.3) nM. In addition, 18k was also highly potent against EGFR^{L858R/T790M} with an IC₅₀ value of 0.7 nM, but its potency against EGFR^{WT} was 94.1-fold less potent.

The X-ray cocrystal structure of EGFR^{T790M/C797S} with 18f was solved (PDB 7ER2, Figure 3a). Similar to our prediction, the results showed that the molecule anchored in the hinge

region of the ATP-binding pocket with bidentate hydrogen bond interactions between Met⁷⁹³ and 2-aminopyrimidine motif. Interestingly, structural alignment of EGFR-18f complex and EGFR-brigatinib complex (PDB 7AEM) showed that the backbone amido linkages between Thr⁸⁵⁴ and Asp⁸⁵⁵ pointed in opposite directions (Figure 3b). Therefore, instead of forming a hydrogen bond directly with side chain OH of Thr⁸⁵⁴ as exhibited in Figure 2a, the oxygen atom on the DMPO group of 18f fostered water-molecule-mediated hydrogen bond interactions with the backbone carbonyl of Thr⁸⁵⁴ and additionally the side chain NH₃ of Lys⁷⁴⁵, inducing a stable conformation and improving binding affinity toward the protein. In addition, the 3'-Cl in 18f located in an appropriate cavity with no steric clash with any residues, providing a rational explanation for the improved EGFR inhibitory potency. Additionally, the C5-chlorine atom on the pyrimidine scaffold was directed toward the "gatekeeper" residue Met⁷⁹⁰ with a distance of 3.2 Å, indicating that this position was unable to accommodate large hydrophobic groups such as CF₃, which explains the poor potency of 18h.

Having demonstrated the selectivity of 18k for its direct target EGFR, we next examined its inhibitory activities against a small kinome panel at a concentration of 100 nM. The results in Figure 4 showed that 18k exhibited >30× selectivity in all 28 tyrosine kinases examined (see the Supporting Information for raw inhibition rate). Strikingly, 18k markedly attenuated the inhibitory effect on ALK, the original target of brigatinib, indicating its improved kinase selectivity.

Furthermore, the antiproliferative activity of 18k was evaluated in BaF3 cells stably expressing EGFR^{L858R/T790M/C797S} (BaF3-EGFR^{TM1}) and EGFR^{19Del/T790M/C797S} (BaF3-EGFR^{TM2}) mutants (Table 2). It was shown that 18k exhibited moderate growth inhibition against the cell models with IC₅₀ values of 0.258 and 0.141 μM, respectively, which were comparable to those of drug 9, whereas its cytotoxic effect on BaF3 parental cells was a bit more potent with an IC₅₀ value of 1.69 μM. Compound 18k also exhibited a 0.60 μM IC₅₀ value against the proliferation of self-constructed osimertinib-resistant PC9 NSCLC cells harboring EGFR^{19Del/T790M/C797S} mutation (PC9-EGFR^{TM2}). Selectivity was observed against PC9 (EGFR^{19Del}) and H1299 (EGFR^{WT}) cell lines, which was within the kinase selectivity margin. Further immunoblotting analysis suggested that the compound dose-dependently suppressed the phosphorylation of EGFR in BaF3-EGFR^{TM1}, BaF3-EGFR^{TM2}, as well as PC9-EGFR^{TM2} cells, supporting its cellular inhibitory potency on the resistance related EGFR mutations (Figure 5).

The pharmacokinetic (PK) parameters of 18f and 18k were also assessed in Sprague–Dawley (SD) rats (Table 3). Both compounds exhibited good PK properties with high oral bioavailability, decent exposure, and acceptable half-life. For instance, 18k exhibited an oral bioavailability value of 81.7%, an AUC (area under curve) value of 5219 h·ng/mL, and a T_{1/2} value of 2.67 h, which were optimal for oral administration for the following in vivo efficacy investigation.

In summary, a straightforward structure optimization of brigatinib was conducted to yield compound 18k as a new potent and selective EGFR^{T790M/C797S} inhibitor. The compound strongly suppressed the EGFR^{L858R/T790M/C797S} and EGFR^{19Del/T790M/C797S} kinases with sub-nanomolar IC₅₀ values but was approximately 94-fold less potent against the wild type EGFR. A cocrystal structure of EGFR^{T790M/C797S} with a close derivative 18f was also solved to provide understanding of the binding mode of the inhibitor. Moreover, compound 18k

exhibited cellular activities in all three cell models, BaF3-EGFR^{TM1}, BaF3-EGFR^{TM2}, and PC9-EGFR^{TM2}, comparable to that of brigatinib. Compound 18k is orally bioavailable and demonstrates highly desirable PK properties, making it a promising lead compound for further structural optimization.

■ ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acsmmedchemlett.1c00555>.

Chemistry, biological assay, X-ray crystallography data, and copies of NMR spectra and HPLC purity data (PDF)

■ AUTHOR INFORMATION

Corresponding Authors

Xiaomei Ren – International Cooperative Laboratory of Traditional Chinese Medicine Modernization and Innovative Drug Development, Ministry of Education (MOE) of China, Guangzhou City Key Laboratory of Precision Chemical Drug Development, School of Pharmacy, Jinan University, Guangzhou 511436, China; Email: ren_xiaomei@jnu.edu.cn

Cai-Hong Yun – Department of Biochemistry and Biophysics, Institute of Systems Biomedicine, School of Basic Medical Sciences, Peking University Health Science Center, Beijing 100191, China; Email: yunch@bjmu.edu.cn

Hua Xie – Division of Antitumor Pharmacology, State Key Laboratory of Drug Research, Shanghai Institute of Materia Medica, Chinese Academy of Sciences, Shanghai 201203, China; Zhongshan Institute for Drug Discovery, Shanghai Institute of Materia Medica, Chinese Academy of Sciences, Zhongshan 528400, China; Email: hxie@simm.ac.cn

Ke Ding – International Cooperative Laboratory of Traditional Chinese Medicine Modernization and Innovative Drug Development, Ministry of Education (MOE) of China, Guangzhou City Key Laboratory of Precision Chemical Drug Development, School of Pharmacy, Jinan University, Guangzhou 511436, China; State Key Laboratory of Bioorganic Chemistry and Natural Products, Shanghai Institute of Organic Chemistry, Chinese Academy of Sciences, Shanghai 200032, China; orcid.org/0000-0001-9016-812X; Email: dingke@jnu.edu.cn

Authors

Shan Li – International Cooperative Laboratory of Traditional Chinese Medicine Modernization and Innovative Drug Development, Ministry of Education (MOE) of China, Guangzhou City Key Laboratory of Precision Chemical Drug Development, School of Pharmacy, Jinan University, Guangzhou 511436, China; orcid.org/0000-0001-5269-5631

Tao Zhang – Division of Antitumor Pharmacology, State Key Laboratory of Drug Research, Shanghai Institute of Materia Medica, Chinese Academy of Sciences, Shanghai 201203, China

Su-Jie Zhu – Institute of Translational Medicine, The Affiliated Hospital of Qingdao University, College of Medicine, Qingdao University, Qingdao 266021, China

Chong Lei – State Key Laboratory of Bioorganic Chemistry and Natural Products, Shanghai Institute of Organic

Chemistry, Chinese Academy of Sciences, Shanghai 200032, China

Mengzhen Lai – Division of Antitumor Pharmacology, State Key Laboratory of Drug Research, Shanghai Institute of Materia Medica, Chinese Academy of Sciences, Shanghai 201203, China

Lijie Peng – International Cooperative Laboratory of Traditional Chinese Medicine Modernization and Innovative Drug Development, Ministry of Education (MOE) of China, Guangzhou City Key Laboratory of Precision Chemical Drug Development, School of Pharmacy, Jinan University, Guangzhou 511436, China

Linjiang Tong – Division of Antitumor Pharmacology, State Key Laboratory of Drug Research, Shanghai Institute of Materia Medica, Chinese Academy of Sciences, Shanghai 201203, China

Zilu Pang – Division of Antitumor Pharmacology, State Key Laboratory of Drug Research, Shanghai Institute of Materia Medica, Chinese Academy of Sciences, Shanghai 201203, China

Xiaoyun Lu – International Cooperative Laboratory of Traditional Chinese Medicine Modernization and Innovative Drug Development, Ministry of Education (MOE) of China, Guangzhou City Key Laboratory of Precision Chemical Drug Development, School of Pharmacy, Jinan University, Guangzhou 511436, China; orcid.org/0000-0001-7931-6873

Jian Ding – Division of Antitumor Pharmacology, State Key Laboratory of Drug Research, Shanghai Institute of Materia Medica, Chinese Academy of Sciences, Shanghai 201203, China

Complete contact information is available at:

<https://pubs.acs.org/10.1021/acsmchemlett.1c00555>

Author Contributions

[†]S.L., T.Z., and S.-J.Z. contributed equally to this work.

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

The authors appreciate the financial support from National Natural Science Foundation of China (21807045, 82173650, 81820108029, 81874284, 22037003, 22077050, and 81903638), Guangdong Province (2021A1515011239 and 2018B030337001), the National Science & Technology Major Project “Key New Drug Creation and Manufacturing Program”, China (2019ZX09301157-004), and the “Personalized Medicines-Molecular Signature-based Drug Discovery and Development”, Strategic Priority Research Program of the Chinese Academy of Sciences (XDA12020112).

ABBREVIATIONS

EGFR, epidermal growth factor receptor; NSCLC, nonsmall cell lung cancer; 19Del, Exon 19 deletion; MTD, maximal tolerated dose; WT, wild type; NMPA, National Medical Products Administration of China; FDA, U.S. Food and Drug Administration; NDA, New Drug Application; DMPO, dimethylphosphine oxide group; TM1, L858R/T790M/C797S; TM2, 19Del/T790M/C797S; DM, L858R/T790M; PK, pharmacokinetic; AUC, area under curve; SD rats, Sprague–Dawley rats; ALK, anaplastic lymphoma kinase; CCK8, cell counting kit 8; SRB, sulforhodamine.

REFERENCES

- (1) Hynes, N. E.; Lane, H. A. ERBB receptors and cancer: the complexity of targeted inhibitors. *Nat. Rev. Cancer* **2005**, *5* (5), 341–354.
- (2) Paez, J. G.; Janne, P. A.; Lee, J. C.; Tracy, S.; Greulich, H.; Gabriel, S.; Herman, P.; Kaye, F. J.; Lindeman, N.; Boggon, T. J.; Naoki, K.; Sasaki, H.; Fujii, Y.; Eck, M. J.; Sellers, W. R.; Johnson, B. E.; Meyerson, M. EGFR mutations in lung cancer: correlation with clinical response to gefitinib therapy. *Science* **2004**, *304* (5676), 1497–1500.
- (3) Dowell, J.; Minna, J. D.; Kirkpatrick, P. Erlotinib hydrochloride. *Nat. Rev. Drug Discov* **2005**, *4* (1), 13–14.
- (4) Dungo, R. T.; Keating, G. M. Afatinib: first global approval. *Drugs* **2013**, *73* (13), 1503–1515.
- (5) Wu, Y. L.; Cheng, Y.; Zhou, X.; Lee, K. H.; Nakagawa, K.; Niho, S.; Tsuji, F.; Linke, R.; Rosell, R.; Corral, J.; Migliorino, M. R.; Pluzanski, A.; Sbar, E. I.; Wang, T.; White, J. L.; Nadanaciva, S.; Sandin, R.; Mok, T. S. Dacomitinib versus gefitinib as first-line treatment for patients with EGFR-mutation-positive non-small-cell lung cancer (ARCHER 1050): a randomised, open-label, phase 3 trial. *Lancet Oncol* **2017**, *18* (11), 1454–1466.
- (6) Finlay, M. R.; Anderton, M.; Ashton, S.; Ballard, P.; Bethel, P. A.; Box, M. R.; Bradbury, R. H.; Brown, S. J.; Butterworth, S.; Campbell, A.; Chorley, C.; Colclough, N.; Cross, D. A.; Currie, G. S.; Grist, M.; Hassall, L.; Hill, G. B.; James, D.; James, M.; Kemmitt, P.; Klinowska, T.; Lamont, G.; Lamont, S. G.; Martin, N.; McFarland, H. L.; Mellor, M. J.; Orme, J. P.; Perkins, D.; Perkins, P.; Richmond, G.; Smith, P.; Ward, R. A.; Waring, M. J.; Whittaker, D.; Wells, S.; Wrigley, G. L. Discovery of a potent and selective EGFR inhibitor (AZD9291) of both sensitizing and T790M resistance mutations that spares the wild type form of the receptor. *J. Med. Chem.* **2014**, *57* (20), 8249–8267.
- (7) Cross, D. A.; Ashton, S. E.; Ghiorghiu, S.; Eberlein, C.; Nebhan, C. A.; Spitzler, P. J.; Orme, J. P.; Finlay, M. R.; Ward, R. A.; Mellor, M. J.; Hughes, G.; Rahi, A.; Jacobs, V. N.; Red Brewer, M.; Ichihara, E.; Sun, J.; Jin, H.; Ballard, P.; Al-Kadhimi, K.; Rowlinson, R.; Klinowska, T.; Richmond, G. H.; Cantarini, M.; Kim, D. W.; Ranson, M. R.; Pao, W. AZD9291, an irreversible EGFR TKI, overcomes T790M-mediated resistance to EGFR inhibitors in lung cancer. *Cancer Discovery* **2014**, *4* (9), 1046–1061.
- (8) Yang, J. C.; Camidge, D. R.; Yang, C. T.; Zhou, J.; Guo, R.; Chiu, C. H.; Chang, G. C.; Shiah, H. S.; Chen, Y.; Wang, C. C.; Berz, D.; Su, W. C.; Yang, N.; Wang, Z.; Fang, J.; Chen, J.; Nikolinakos, P.; Lu, Y.; Pan, H.; Maniam, A.; Bazhenova, L.; Shirai, K.; Jahanzeb, M.; Willis, M.; Masood, N.; Chowhan, N.; Hsia, T. C.; Jian, H.; Lu, S. Safety, Efficacy, and Pharmacokinetics of Almonertinib (HS-10296) in Pretreated Patients With EGFR-Mutated Advanced NSCLC: A Multicenter, Open-label, Phase 1 Trial. *J. Thorac Oncol* **2020**, *15* (12), 1907–1918.
- (9) Shi, Y.; Hu, X.; Zhang, S.; Lv, D.; Wu, L.; Yu, Q.; Zhang, Y.; Liu, L.; Wang, X.; Cheng, Y.; Ma, Z.; Niu, H.; Wang, D.; Feng, J.; Huang, C.; Liu, C.; Zhao, H.; Li, J.; Zhang, X.; Jiang, Y.; Gu, C. Efficacy, safety, and genetic analysis of furmonertinib (AST2818) in patients with EGFR T790M mutated non-small-cell lung cancer: a phase 2b, multicentre, single-arm, open-label study. *Lancet Respir Med.* **2021**, *9* (8), 829–839.
- (10) Nagasaka, M.; Zhu, V. W.; Lim, S. M.; Greco, M.; Wu, F.; Ou, S. I. Beyond Osimertinib: The Development of Third-Generation EGFR Tyrosine Kinase Inhibitors For Advanced EGFR+ NSCLC. *J. Thorac Oncol* **2021**, *16* (5), 740–763.
- (11) Zhang, T.; Qu, R.; Chan, S.; Lai, M.; Tong, L.; Feng, F.; Chen, H.; Song, T.; Song, P.; Bai, G.; Liu, Y.; Wang, Y.; Li, Y.; Su, Y.; Shen, Y.; Sun, Y.; Chen, Y.; Geng, M.; Ding, K.; Ding, J.; Xie, H. Discovery of a novel third-generation EGFR inhibitor and identification of a potential combination strategy to overcome resistance. *Mol. Cancer* **2020**, *19* (1), 90.
- (12) Thress, K. S.; Paweletz, C. P.; Felip, E.; Cho, B. C.; Stetson, D.; Dougherty, B.; Lai, Z.; Markovets, A.; Vivancos, A.; Kuang, Y.; Ercan, D.; Matthews, S. E.; Cantarini, M.; Barrett, J. C.; Janne, P. A.; Oxnard, G. R. Acquired EGFR C797S mutation mediates resistance to

AZD9291 in non-small cell lung cancer harboring EGFR T790M. *Nat. Med.* **2015**, *21* (6), 560–562.

(13) Song, H. N.; Jung, K. S.; Yoo, K. H.; Cho, J.; Lee, J. Y.; Lim, S. H.; Kim, H. S.; Sun, J. M.; Lee, S. H.; Ahn, J. S.; Park, K.; Choi, Y. L.; Park, W.; Ahn, M. J. Acquired C797S Mutation upon Treatment with a T790M-Specific Third-Generation EGFR Inhibitor (HM61713) in Non-Small Cell Lung Cancer. *J. Thorac. Oncol.* **2016**, *11* (4), E45–E47.

(14) Niederst, M. J.; Hu, H.; Mulvey, H. E.; Lockerman, E. L.; Garcia, A. R.; Piotrowska, Z.; Sequist, L. V.; Engelman, J. A. The Allelic Context of the C797S Mutation Acquired upon Treatment with Third-Generation EGFR Inhibitors Impacts Sensitivity to Subsequent Treatment Strategies. *Clin. Cancer Res.* **2015**, *21* (17), 3924–3933.

(15) Ayeni, D.; Politi, K.; Goldberg, S. B. Emerging Agents and New Mutations in EGFR-Mutant Lung Cancer. *Clin. Cancer Res.* **2015**, *21* (17), 3818–3820.

(16) Uchibori, K.; Inase, N.; Araki, M.; Kamada, M.; Sato, S.; Okuno, Y.; Fujita, N.; Katayama, R. Brigatinib combined with anti-EGFR antibody overcomes osimertinib resistance in EGFR-mutated non-small-cell lung cancer. *Nat. Commun.* **2017**, *8*, 14768.

(17) Park, H.; Jung, H. Y.; Mah, S.; Hong, S. Discovery of EGF Receptor Inhibitors That Are Selective for the d746–750/T790M/C797S Mutant through Structure-Based de Novo Design. *Angew. Chem. Int. Edit* **2017**, *56* (26), 7634–7638.

(18) Gunther, M.; Juchum, M.; Kelter, G.; Fiebig, H.; Laufer, S. Lung Cancer: EGFR Inhibitors with Low Nanomolar Activity against a Therapy-Resistant L858R/T790M/C797S Mutant. *Angew. Chem. Int. Edit* **2016**, *55* (36), 10890–10894.

(19) Shen, J.; Zhang, T.; Zhu, S. J.; Sun, M.; Tong, L.; Lai, M.; Zhang, R.; Xu, W.; Wu, R.; Ding, J.; Yun, C. H.; Xie, H.; Lu, X.; Ding, K. Structure-Based Design of 5-Methylpyrimidopyridone Derivatives as New Wild-Type Sparing Inhibitors of the Epidermal Growth Factor Receptor Triple Mutant (EGFR(L858R/T790M/C797S)). *J. Med. Chem.* **2019**, *62* (15), 7302–7308.

(20) Kashima, K.; Kawauchi, H.; Tanimura, H.; Tachibana, Y.; Chiba, T.; Torizawa, T.; Sakamoto, H. CH7233163 overcomes osimertinib resistant EGFR-Del19/T790M/C797S mutation. *Mol. Cancer Ther* **2020**, *19* (11), 2288–2297.

(21) Jia, Y.; Yun, C. H.; Park, E.; Ercan, D.; Manuia, M.; Juarez, J.; Xu, C.; Rhee, K.; Chen, T.; Zhang, H.; Palakurthi, S.; Jang, J.; Lelais, G.; DiDonato, M.; Bursulaya, B.; Michellys, P. Y.; Epple, R.; Marsilje, T. H.; McNeill, M.; Lu, W.; Harris, J.; Bender, S.; Wong, K. K.; Janne, P. A.; Eck, M. J. Overcoming EGFR(T790M) and EGFR(C797S) resistance with mutant-selective allosteric inhibitors. *Nature* **2016**, *534* (7605), 129–132.

(22) To, C.; Jang, J.; Chen, T.; Park, E.; Mushajiang, M.; De Clercq, D. J. H.; Xu, M.; Wang, S.; Cameron, M. D.; Heppner, D. E.; Shin, B. H.; Gero, T. W.; Yang, A.; Dahlberg, S. E.; Wong, K. K.; Eck, M. J.; Gray, N. S.; Janne, P. A. Single and Dual Targeting of Mutant EGFR with an Allosteric Inhibitor. *Cancer Discov* **2019**, *9* (7), 926–943.

(23) Engelhardt, H.; Bose, D.; Petronczki, M.; Scharn, D.; Bader, G.; Baum, A.; Bergner, A.; Chong, E.; Dobel, S.; Egger, G.; Engelhardt, C.; Ettmayer, P.; Fuchs, J. E.; Gerstberger, T.; Gonnella, N.; Grimm, A.; Grondal, E.; Haddad, N.; Hopfgartner, B.; Kousek, R.; Krawiec, M.; Kriz, M.; Lamarre, L.; Leung, J.; Mayer, M.; Patel, N. D.; Simov, B. P.; Reeves, J. T.; Schnitzer, R.; Schrenk, A.; Sharps, B.; Solca, F.; Stadtmuller, H.; Tan, Z.; Wunberg, T.; Zoephel, A.; McConnell, D. B. Start Selective and Rigidify: The Discovery Path toward a Next Generation of EGFR Tyrosine Kinase Inhibitors. *J. Med. Chem.* **2019**, *62* (22), 10272–10293.

(24) Liu, X. L.; Zhang, X. Q.; Yang, L.; Tian, X.; Dong, T. T.; Ding, C. Z.; Hu, L. H.; Wu, L. Y.; Zhao, L. L.; Mao, J.; Ji, Q. S.; Yan, S. Y.; Zhu, Z. Z.; Xia, Y. F.; Chan, C. C.; Chen, S. H. Preclinical evaluation of TQB3804, a potent EGFR C797S inhibitor. *Cancer Res.* **2019**, *79* (13), 1320.

(25) Dong, R. F.; Zhu, M. L.; Liu, M. M.; Xu, Y. T.; Yuan, L. L.; Bian, J.; Xia, Y. Z.; Kong, L. Y. EGFR mutation mediates resistance to

EGFR tyrosine kinase inhibitors in NSCLC: From molecular mechanisms to clinical research. *Pharmacol. Res.* **2021**, *167*, 105583.

(26) Finlay, M. R. V.; Barton, P.; Bickerton, S.; Bista, M.; Colclough, N.; Cross, D. A. E.; Evans, L.; Floc'h, N.; Gregson, C.; Guerot, C. M.; Hargreaves, D.; Kang, X.; Lenz, E. M.; Li, X.; Liu, Y.; Lorthioir, O.; Martin, M. J.; McKerrecher, D.; McWhirter, C.; O'Neill, D.; Orme, J. P.; Mosallanejad, A.; Rahi, A.; Smith, P. D.; Talbot, V.; Ward, R. A.; Wrigley, G.; Wylot, M.; Xue, L.; Yao, T.; Ye, Y.; Zhao, X. Potent and Selective Inhibitors of the Epidermal Growth Factor Receptor to Overcome C797S-Mediated Resistance. *J. Med. Chem.* **2021**, *64* (18), 13704–13718.

(27) Zhou, W.; Ercan, D.; Chen, L.; Yun, C. H.; Li, D.; Capelletti, M.; Cortot, A. B.; Chirieac, L.; Iacob, R. E.; Padera, R.; Engen, J. R.; Wong, K. K.; Eck, M. J.; Gray, N. S.; Janne, P. A. Novel mutant-selective EGFR kinase inhibitors against EGFR T790M. *Nature* **2009**, *462* (7276), 1070–1074.

(28) Xu, S.; Xu, T.; Zhang, L.; Zhang, Z.; Luo, J.; Liu, Y.; Lu, X.; Tu, Z.; Ren, X.; Ding, K. Design, synthesis, and biological evaluation of 2-oxo-3,4-dihydropyrimido[4,5-d]pyrimidinyl derivatives as new irreversible epidermal growth factor receptor inhibitors with improved pharmacokinetic properties. *J. Med. Chem.* **2013**, *56* (21), 8803–8813.

(29) Huang, W. S.; Liu, S.; Zou, D.; Thomas, M.; Wang, Y.; Zhou, T.; Romero, J.; Kohlmann, A.; Li, F.; Qi, J.; Cai, L.; Dwight, T. A.; Xu, Y.; Xu, R.; Dodd, R.; Toms, A.; Parillon, L.; Lu, X.; Anjum, R.; Zhang, S.; Wang, F.; Keats, J.; Wardwell, S. D.; Ning, Y.; Xu, Q.; Moran, L. E.; Mohemmad, Q. K.; Jang, H. G.; Clackson, T.; Narasimhan, N. I.; Rivera, V. M.; Zhu, X.; Dalgarno, D.; Shakespeare, W. C. Discovery of Brigatinib (AP26113), a Phosphine Oxide-Containing, Potent, Orally Active Inhibitor of Anaplastic Lymphoma Kinase. *J. Med. Chem.* **2016**, *59* (10), 4948–4964.